

Appendix A

A.1 Carbonated Hydroxyapatite (CHA)

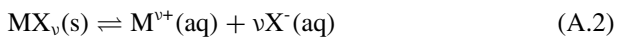
For carbonated hydroxyapatite (CHA), with the formula: $\text{Ca}_{10}(\text{PO}_4)_6(\text{CO}_3)_x(\text{OH})_{2-2x}$, a fraction of $x = 0.4$ was chosen to accommodate the reduction in carbonate content experienced during heat treatment by bone, which usually has $\text{CO}_3 = (4\dots6)\%$. The chosen value, $x = 0.4$, represents a $\text{CO}_3 \approx 2.4\%$ and a $\text{CO}_2 \approx 1.7\%$, with the latter value needed to calculate the K_{sp} value according to Ito et al. [1].

The CHA molar mass can be expressed as a function of pure hydroxyapatite molar mass, M_{HA} , and the carbonate content, x , as:

$$\begin{aligned} M_{\text{CHA}} &= 10M_{\text{Ca}} + 6(M_{\text{P}} + 4M_{\text{O}}) + x(M_{\text{C}} + 3M_{\text{O}}) + (2 - 2x)(M_{\text{H}} + M_{\text{O}}) \\ &= M_{\text{HA}} + x(M_{\text{O}} + M_{\text{C}} - 2M_{\text{H}}) \\ &= 1004.6162 + x(25.994) \\ &= 1004.6162 + 0.4(25.994) \\ &= 1015.010 \qquad \qquad \qquad \text{g mol}^{-1} \quad (\text{A.1}) \end{aligned}$$

A.2 Solubility Product (K_{sp}) and Equilibrium Calcium Concentration ($[\text{Ca}^{2+}]_{\text{eq}}$)

The general equilibrium equation for the dissolution of a sparingly soluble ionic compound MX_v is given by the following expression from [2]:



The solubility product (K_{sp}) can be defined as:

$$K_{\text{sp}} = [M^+][X^-]^\nu \quad (\text{A.3})$$

with $[M^+]$, the concentration of M^+ ions in water and $[X^-]$, the concentration of X^- ions in water.

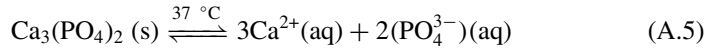
A generalised version can be expressed as:

$$K_{\text{sp}} = \prod_{i=1}^n C_i^{m_i} |_{\text{eq}} \quad (\text{A.4})$$

with $C_i |_{\text{eq}}$ the concentration of constituents ions at equilibrium and m_i , their respective coefficients in the ionic compound.

A.2.1 Tricalcium Phosphate (TCP)

Tricalcium phosphate (TCP), whose chemical formula is $\text{Ca}_3(\text{PO}_4)_2$, has the following dissolution reaction:



Assuming stoichiometric dissolution: $[\text{PO}_4^{3-}]_{\text{eq}} = \frac{2}{3}[\text{Ca}^{2+}]_{\text{eq}}$ and the solubility product can be expressed as:

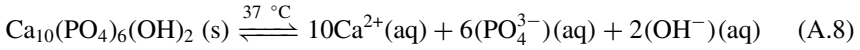
$$\begin{aligned} K_{\text{sp}} &= ([\text{Ca}^{2+}]_{\text{eq}})^3 ([\text{PO}_4^{3-}]_{\text{eq}})^2 \\ &= ([\text{Ca}^{2+}]_{\text{eq}})^3 \left(\frac{2}{3}\right)^2 ([\text{Ca}^{2+}]_{\text{eq}})^2 \\ &= \frac{4}{9} ([\text{Ca}^{2+}]_{\text{eq}})^5 \quad (\text{mol}^5 \text{ dm}^{-15}) \quad (\text{A.6}) \end{aligned}$$

Thus, the equilibrium calcium concentration is:

$$\begin{aligned} [\text{Ca}^{2+}]_{\text{eq}} &= \left(\frac{9}{4} K_{\text{sp}}\right)^{\frac{1}{5}} \\ &= \left(\frac{9}{4} 10^{-(-\log_{10} K_{\text{sp}})}\right)^{\frac{1}{5}} \quad (\text{mol dm}^{-3}) \quad (\text{A.7}) \end{aligned}$$

A.2.2 Pure Hydroxyapatite (HA_p)

Pure hydroxyapatite, whose chemical formula is $\text{Ca}_{10}(\text{PO}_4)_6(\text{OH})_2$, has the following dissolution reaction:



Assuming stoichiometric dissolution: $[\text{PO}_4^{3-}]_{\text{eq}} = \frac{3}{5}[\text{Ca}^{2+}]_{\text{eq}}$ and $[\text{OH}^-]_{\text{eq}} = \frac{1}{5}[\text{Ca}^{2+}]_{\text{eq}}$, and K_{sp} can be expressed as:

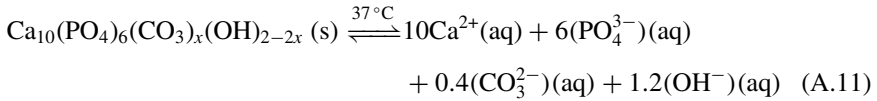
$$\begin{aligned} K_{\text{sp}} &= ([\text{Ca}^{2+}]_{\text{eq}})^{10} ([\text{PO}_4^{3-}]_{\text{eq}})^6 ([\text{OH}^-]_{\text{eq}})^2 \\ &= ([\text{Ca}^{2+}]_{\text{eq}})^{10} \left(\frac{3}{5}\right)^6 ([\text{Ca}^{2+}]_{\text{eq}})^6 \left(\frac{1}{5}\right)^2 ([\text{Ca}^{2+}]_{\text{eq}})^2 \\ &= \frac{3^6}{5^8} ([\text{Ca}^{2+}]_{\text{eq}})^{18} \quad (\text{mol}^{18} \text{ dm}^{-54}) \quad (\text{A.9}) \end{aligned}$$

Thus, the equilibrium calcium concentration is:

$$\begin{aligned} [\text{Ca}^{2+}]_{\text{eq}} &= \left(\frac{5^8}{3^6} K_{\text{sp}}\right)^{\frac{1}{18}} \\ &= \left(\frac{5^8}{3^6} 10^{-(-\log_{10} K_{\text{sp}})}\right)^{\frac{1}{18}} \quad (\text{mol dm}^{-3}) \quad (\text{A.10}) \end{aligned}$$

A.2.3 Carbonated Hydroxyapatite (CHA)

Carbonated hydroxyapatite (CHA), with the formula: $\text{Ca}_{10}(\text{PO}_4)_6(\text{CO}_3)_x(\text{OH})_{2-2x}$ with $x = 0.4$, has the following dissolution reaction:



Assuming stoichiometric dissolution: $[\text{PO}_4^{3-}]_{\text{eq}} = \frac{3}{5}[\text{Ca}^{2+}]_{\text{eq}}$, $[\text{CO}_3^{2-}]_{\text{eq}} = \frac{x}{10}[\text{Ca}^{2+}]_{\text{eq}}$ and $[\text{OH}^-]_{\text{eq}} = \frac{1-x}{5}[\text{Ca}^{2+}]_{\text{eq}}$, and K_{sp} can be expressed as:

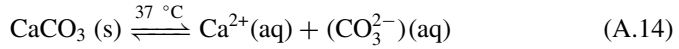
$$\begin{aligned} K_{\text{sp}} &= ([\text{Ca}^{2+}]_{\text{eq}})^{10} ([\text{PO}_4^{3-}]_{\text{eq}})^6 ([\text{CO}_3^{2-}]_{\text{eq}})^x ([\text{OH}^-]_{\text{eq}})^{(2-2x)} \\ &= ([\text{Ca}^{2+}]_{\text{eq}})^{10} \left(\frac{3}{5}\right)^6 ([\text{Ca}^{2+}]_{\text{eq}})^6 \left(\frac{x}{5}\right)^x ([\text{Ca}^{2+}]_{\text{eq}})^x \left(\frac{1-x}{5}\right)^{(2-2x)} ([\text{Ca}^{2+}]_{\text{eq}})^{(2-2x)} \\ &= \frac{3^6(2.5x)^x(1-x)^{(2-2x)}}{5^8} ([\text{Ca}^{2+}]_{\text{eq}})^{(18-x)} \quad (\text{mol}^{18-x} \text{ dm}^{-(54-1.2x)}) \quad (\text{A.12}) \end{aligned}$$

Thus, the equilibrium calcium concentration is:

$$\begin{aligned}
[Ca^{2+}]_{eq} &= \left(\frac{5^8}{3^6(2.5x)^x(1-x)^{(2-2x)}} K_{sp} \right)^{\frac{1}{18-x}} \\
&= \left(\frac{5^8}{3^6(2.5x)^x(1-x)^{(2-2x)}} 10^{-(-\log_{10} K_{sp})} \right)^{\frac{1}{18-x}} \\
&= \left(\frac{5^8}{3^6 1^{0.4} 0.6^{1.2}} 10^{-(-\log_{10} K_{sp})} \right)^{\frac{1}{17.6}} \quad (\text{mol dm}^{-3}) \quad (\text{A.13})
\end{aligned}$$

A.2.4 Calcium Carbonate (CC)

Calcium carbonate, whose chemical formula is CaCO_3 , has the following dissolution reaction:



Assuming stoichiometric dissolution: $[Ca^{2+}]_{eq} = [CO_3^{2-}]_{eq}$, and K_{sp} can be expressed as:

$$\begin{aligned}
K_{sp} &= [Ca^{2+}]_{eq}[CO_3^{2-}]_{eq} \\
&= ([Ca^{2+}]_{eq})^2 \quad (\text{mol}^2 \text{ dm}^{-6}) \quad (\text{A.15})
\end{aligned}$$

Thus, the equilibrium calcium concentration is:

$$\begin{aligned}
[Ca^{2+}]_{eq} &= (K_{sp})^{\frac{1}{2}} \\
&= \left(10^{-(-\log_{10} K_{sp})} \right)^{\frac{1}{2}} \quad (\text{mol dm}^{-3}) \quad (\text{A.16})
\end{aligned}$$

A.3 Polymer Dissociation Constant (pK_a) for Copolymers and Blends

The dissociation constant K_a of a copolymer or polymer blend can be computed using the K_a values of their individual components, usually reported in literature, as shown for a two-components case:

$$K_{a1} = \frac{[H^+][R_1-COO^-]}{[R_1-COOH]} \quad (\text{A.17})$$

$$K_{a2} = \frac{[H^+][R_2-COO^-]}{[R_2-COOH]} \quad (\text{A.18})$$

The overall K_a value can be expressed as:

$$K_a = \frac{[H^+][R-COO^-]}{[R-COOH]} = \frac{[H^+]([R_1-COO^-] + [R_2-COO^-])}{[R_1-COOH] + [R_2-COOH]} \quad (\text{A.19})$$

Assuming random polymerisation, the possibility of finding a specific monomer at the end of a chain is equal to the molar fraction of said monomer:

$$[R_1-COOH] + [R_1-COO^-] = \frac{\alpha_1}{\alpha_2} ([R_2-COOH] + [R_2-COO^-]) \quad (\text{A.20})$$

As the ratio of undissociated to dissociated chains is large, the relationship can be approximated as:

$$[R_1-COOH] = \frac{\alpha_1}{\alpha_2} [R_2-COOH] \quad (\text{A.21})$$

Given the following expression for the overall K_a value:

$$\begin{aligned} K_a &= \frac{[H^+][R-COO^-]}{[R-COOH]} = \frac{[H^+][R_1-COO^-] + [H^+][R_2-COO^-]}{[R_1-COOH] + [R_2-COOH]} \\ &= \frac{K_{a1}[R_1-COOH] + K_{a2}[R_2-COO^-]}{[R_1-COOH] + [R_2-COOH]} \\ &= \frac{K_{a1}[R_1-COOH] + K_{a2}\frac{\alpha_2}{\alpha_1}[R_1-COOH]}{[R_1-COOH] + \frac{\alpha_2}{\alpha_1}[R_1-COOH]} \\ &= \frac{K_{a1} + K_{a2}\frac{\alpha_2}{\alpha_1}}{1 + \frac{\alpha_2}{\alpha_1}} = \frac{K_{a1}\alpha_1 + K_{a2}\alpha_2}{\alpha_1 + \alpha_2} = \alpha_1 K_{a1} + \alpha_2 K_{a2} \end{aligned} \quad (\text{A.22})$$

Resulting in a dissociation constant value for a copolymer or polymer blend of $pK_a = -\log_{10}(\alpha_1 K_{a1} + \alpha_2 K_{a2})$ with α_1 and α_2 , the molar/ ratios of the two components of the copolymer or blend and K_{a1} and K_{a2} , their corresponding values of dissociation constant.

A.4 Initial Degree of Pseudo-polymerisation

N_{dp0} (Da Da^{-1}), the initial degree of pseudo-polymerisation, can be expressed as a function of M_{unit} , the molar mass associated to one ester bond (Da) and M_{n0} , the number-average molecular weight at the time origin (Da); or C_{e0} , the concentration of ester bonds in long chains at the time origin in the polymer phase (mol m^{-3}) and $C_{\text{chain}0}$, the initial concentration of long polymer chains in the polymer phase (mol m^{-3}) using $C_{e0} = \frac{\rho_{\text{pol}}}{M_{\text{unit}}}$ and $C_{\text{chain}0} = \frac{\rho_{\text{pol}}}{M_{n0}}$ with ρ_{pol} , the polymer density (kg m^{-3}):

$$\frac{C_{\text{chain0}}}{C_{\text{e0}}} = \frac{\text{Initial number of chains}}{\text{Initial number of ester bonds}} = \frac{\frac{\rho_{\text{pol}}}{M_{\text{n0}}}}{\frac{\rho_{\text{pol}}}{M_{\text{unit}}}} = \frac{M_{\text{unit}}}{M_{\text{n0}}} = N_{\text{dp0}}^{-1} \quad (\text{A.23})$$

A.5 Sauter Mean Diameter

Assuming spherical particles, the diameter which characterises the distribution in terms of volume and surface area, d_{eq} , would satisfy:

$$\begin{aligned} \frac{\text{Overall volume of particles}}{\text{Overall surface of particles}} &= \frac{\sum_{i=1}^N V_i}{\sum_{i=1}^N A_i} = \frac{\sum_{i=1}^N \frac{\pi}{6} d_i^3}{\sum_{i=1}^N \pi d_i^2} \\ &= \frac{N \frac{\pi}{6} d_{\text{eq}}^3}{N \pi d_{\text{eq}}^2} = \frac{d_{\text{eq}}}{6} \quad (\text{m}) \quad (\text{A.24}) \end{aligned}$$

being V_i (m^3), A_i (m^2) and d_i (m), the volume, surface area and diameter of the i th particle respectively.

The Sauter Mean diameter, (SMD or $D[2, 3]$ or d_{32}), is defined as:

$$\begin{aligned} SMD = D[2, 3] = d_{32} &= \frac{d_v^3}{d_s^2} = \frac{N d_v^3}{N d_s^2} = \frac{\sum_{i=1}^N d_i^3}{\sum_{i=1}^N d_i^2} \\ &= 6 \frac{\sum_{i=1}^N \frac{\pi}{6} d_i^3}{\sum_{i=1}^N \pi d_i^2} = 6 \frac{\sum_{i=1}^N V_i}{\sum_{i=1}^N A_i} \quad (\text{m}) \quad (\text{A.25}) \end{aligned}$$

with d_v (m), the volume equivalent particle diameter and d_s (m), the surface equivalent particle diameter.

Thus, $d_{\text{eq}} = SMD = D[2, 3] = d_{32}$.

A.6 Geometric Mean and Geometric Standard Deviation

The geometric mean of a set of numbers $\{x_1, x_2, \dots, x_N\}$ can be defined as:

$$\mu_g = \left(\prod_{i=1}^N x_i \right)^{\frac{1}{N}} = \sqrt[N]{x_1 x_2 \dots x_N} \quad (\text{A.26})$$

And the geometric standard deviation, more appropriately called geometric SD factor [3], can be calculated as:

$$\sigma_g = \exp \left(\sqrt{\frac{\sum_{i=1}^N \left(\ln \frac{x_i}{\mu_g} \right)^2}{N-1}} \right) \quad (\text{A.27})$$

With $\{x_1, x_2, \dots, x_N\}$ characterised by $(\mu_g * \sigma_g)$

A.7 Characterisation of Poly(ethylene:hexamethylene/sebacate)

Figure A.1 includes the different repeating units of the poly(ethylene:hexamethylene/sebacate) block [4]. The copolymer had an estimated 50 mol% sebacic acid, a 25 mol% ethylene glycol and a 25 mol% hexamethylene glycol [5].

A random copolymerisation yields, on average, 1.75 ester bonds available per repeating unit, resulting in a M_{unit} value of:

$$\begin{aligned} M_{\text{unit}} &= \frac{\text{Molar mass of the average repeating unit}}{\text{Number of ester bonds per repeating unit}} \\ &= \frac{0.5M_{\text{sebacic acid}} + 0.25M_{\text{ethylene}} + 0.25M_{\text{hexamethylene}}}{1.75} \\ &= 146.481 \text{ Da} \end{aligned} \quad (\text{A.28})$$

with $M_{\text{sebacic acid}}$, M_{ethylene} and $M_{\text{hexamethylene}}$, the molar masses of the different monomer repeating units.

Table A.1 includes the values of the polymer-specific parameters for poly(ethylene:hexamethylene/sebacate) block copolymer.

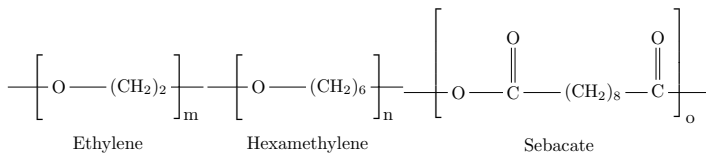


Fig. A.1 Structure of the repeating units of poly(ethylene:hexamethylene/sebacate) block copolymer, taken from [4]

Table A.1 Polymer-specific parameters for poly(ethylene:hexamethylene/sebacate) block copolymer. M_{unit} is the average molar mass associated to one ester bond, $\text{p}K_{\text{a}}$ is the negative base 10 logarithm of the sebacate segment acid dissociation constant at 25 °C and ρ_{pol} is the polymer density reported as midpoint of the typical density interval

Property	Value	Reference
M_{unit}	146.481 Da	See above
$\text{p}K_{\text{a}}$ at 25 °C	4.59	[6]
ρ_{pol}	1085 kg m ⁻³	[7]

Appendix B

This appendix contains information of all the analysed case studies in tabular format. Presented divided by ceramic filler and subdivided by polymeric matrix, each case study occupies a row containing the values of the four composite degradation constants (k_1 , k'_2 , A_d and θ), in addition to several relevant characteristics. These characteristics are split into three different groups: polymer characteristics, such as polymer type and manufacturer; ceramic characteristics, such as ceramic type and manufacturer; and general sample characteristics, such as structure and morphology, fabrication method and degradation protocol. The TCP case studies can be found in Appendix B.1, followed by HA cases in Appendix B.2 and lastly, CC cases in Appendix B.3.

B.1 Tricalcium Phosphate Case Studies Modelling Results

TCP case studies with a poly(L-lactide) matrix are included in Appendix B.1.1, (Table B.1), followed by cases with a poly(D,L-lactide) homo- or copolymer matrix in Appendix B.1.2, (Table B.2), cases with poly(lactide-co-glycolide) matrix in Appendix B.1.3, (Table B.3), and lastly, cases with miscellaneous polymers matrices in Appendix B.1.4, (Table B.4).

B.1.1 Poly(L-lactide) Matrix

Table B.1 Results output by the TCP composites degradation model for cases with a poly(L-lactide) matrix. k_1 is the non-catalytic polymer degradation rate, k_2' is the autocatalytic polymer degradation rate, A_d is the ceramic dissolution rate and θ is the power ceramic dissolution law exponent. Abbreviations used to specify the tricalcium phosphate type can be found in Table 4.2. A dashed line between rows indicates that data displayed in those consecutive rows belong to the same researcher or research group. A solid line between rows indicates no known author relationship for the data

Polymer characteristics			Ceramic characteristics			Sample characteristics			Degradation protocol	Case code and reference	
Type	Manufacturer	k_1 (s^{-1})	k_2' ($m^3 \text{ mol}^{-1} s^{-1}$)	Type	Manufacturer	A_d ($\text{mol m}^{-2} s^{-1}$)	θ (1)	Structure and Morphology			Fabrication method
Poly(L-lactide)	Shimadzu Co., Ltd.; Kyoto, Japan (Lacy#5000)	1.4×10^{-11} (3.7×10^{-12})	2.1×10^{-11} (4.30×10^{-12})	β -TCP	Taihei Chemical Industrial Co., Ltd., Osaka, Japan	5.0×10^{-12}	2.0	Dense film: $10 \times 100 \text{ mm}^2 \times 4 \text{ mm}$	Polymer and ceramic dry-mixed + injection-moulded into final shape	In vitro: PBS (pH = 7.4) at 37°C, w/o shaking, w/o replacement	Kobayashi β -TCP [8]
Poly(L-lactide)	Purac Biomaterials B.V.; Gorinchem, The Netherlands	4.0×10^{-10} (1.1×10^{-11})	3.8×10^{-10} (1.0×10^{-11})	β -TCP	In-house production: granulated + sintered at 1000°C in air	1.0×10^{-12}	2.0	Dense cylinder: $\Phi = 6 \text{ mm}$, $L = 10 \text{ mm}$	Polymer and ceramic mixed in air at 60°C for 24 h + injection-moulded into final shape + γ -irradiation 25 kGy	In vivo: femoral implantation in rabbits following Kathagen protocol	Annoble β -TCP [9]
Poly(L-lactide)	Purac Biomaterials B.V.; Gorinchem, The Netherlands	5.0×10^{-12}	1.3×10^{-11}	β -TCP	Degradable Solution AG, Switzerland	7.0×10^{-14}	2.0	Dense bar: $60 \times 9 \text{ mm}^2 \times 1.5 \text{ mm}$	Polymer and ceramic mixed in twin-screw extruder at 200°C + milled to powder in liquid N_2 + compression-moulded into final shape + sterilised w ethylene oxide	In vitro: Simulated body fluid (SBF) (pH = 7.4) at 37°C, w/o shaking, w/o replacement	Adams β -TCP [10]

(continued)

Table B.1 (continued)

Polymer characteristics			Ceramic characteristics			Sample characteristics			Fabrication method		Degradation protocol	Case code and reference
Type	Manufacturer	k_1 (s^{-1})	k'_2 ($m^3 mol^{-1} s^{-1}$)	Type	Manufacturer	A_d ($mol m^{-2} s^{-1}$)	θ (1)	Structure and Morphology	Fabrication method	Degradation protocol	Case code and reference	
Poly(L-lactide)	Chengdu Organic Chemicals Co. Ltd, China	3.2×10^{-11}	3.8×10^{-11}	β -TCP	In-house production: wet chemistry route using $CaCO_3$ and H_3PO_4 + calcining at $950^\circ C$ for 2 h	3.0×10^{-14}	2.0	Porous (60%) cylinder: $\Phi = 5$ mm, $L = 10$ mm	Polymer dissolved in chloroform + ceramic and porogen addition + composite precipitation in acetone and ethanol + compression moulding at 7 MPa and $25^\circ C$ + porogen leaching	In vitro: SBF (pH = 7.4) at $37^\circ C$, w shaking, w replacement of 30 mL every 2 days, 100 mL/sample	Kang 07 S β -TCP [11]	
Poly(L-lactide)	Chengdu Organic Chemicals Co. Ltd, China	2.2×10^{-11}	2.2×10^{-11}	β -TCP	In-house production: same as Kang 07 S	3.0×10^{-14}	2.0	Porous (60%) cylinder: $\Phi = 5$ mm, $L = 10$ mm	Same as Kang 07 S	In vitro: SBF (pH = 7.4) at $37^\circ C$, w constant buffer replacement at a flow rate of 0.02 mL/(mL.min)	Kang 07 F β -TCP [11]	
Poly(L-lactide)	Chengdu Organic Chemicals Co. Ltd, China	2.8×10^{-11}	3.4×10^{-11}	β -TCP	In-house production: same as Kang 07 S	3.0×10^{-14}	2.0	Porous (60%) cylinder: $\Phi = 5$ mm, $L = 10$ mm	Same as Kang 07 S	In vitro: same as Kang 07 F	Kang 09 F β -TCP [12]	
Poly(L-lactide)	Chengdu Organic Chemicals Co. Ltd, China	4.2×10^{-11}	5.8×10^{-11}	β -TCP	In-house production: same as Kang 09 F	3.0×10^{-14}	2.0	Porous (60%) cylinder: $\Phi = 5$ mm, $L = 10$ mm	Same as Kang 07 S	In vitro: SBF (pH = 7.4) at $37^\circ C$, w constant buffer replacement at a flow rate of 0.02 mL/(mL.min) and dynamic loading at 0.6 Hz and 0.1 MPa	Kang 09 FL- β -TCP [12]	

B.1.2 Poly(D,L-lactide) Homo- and Copolymers Matrixes

Table B.2 Results output by the TCP composites degradation model for cases with a poly(D,L-lactide) or a poly(D,L-lactide) copolymer matrix. k_1 is the non-catalytic polymer degradation rate, k'_2 is the autocatalytic polymer degradation rate, A_d is the ceramic dissolution rate and θ is the power ceramic dissolution law exponent. Abbreviations used to specify the tricalcium phosphate type can be found in Table 4.2. A dashed line between rows indicates that data displayed in those consecutive rows belong to the same researcher or research group. A solid line between rows indicates no known author relationship for the data

Polymer characteristics			Ceramic characteristics			Sample characteristics			Degradation protocol	Case code and reference	
Type	Manufacturer	k_1 (s^{-1})	k'_2 ($m^3 mol^{-1} s^{-1}$)	Type	Manufacturer	A_d ($mol m^{-2} s^{-1}$)	θ (1)	Structure and Morphology			Fabrication method
Poly(D,L-lactide)(4:96)	Purac Biochem B.V.; Gorinchem, The Netherlands	5.0×10^{-11}	5.0×10^{-11}	β -TCP	Biomatlante SA; Vigneux de Bretagne, France	5×10^{-12}	2.0	Dense cylinder: $\phi = 6$ mm; L = 8mm	Polymer + ceramic fed to twin-screw extruder + extruded into cylindrical billet at 50 rpm and (225...235) °C for 2.5 min + self-reinforcing + mached into final shape + γ -radiation (25 kGy)	In vitro: PBS (pH = 7.4) at 37 °C, w formightly replacement	Daculsi Vit β -TCP [13]
Poly(D,L-lactide)(4:96)	Purac Biochem B.V.; Gorinchem, The Netherlands	7.0×10^{-11}	7.0×10^{-11}	β -TCP	Biomatlante SA; Vigneux de Bretagne, France	1.0×10^{-11}	2.0	Dense cylinder: $\phi = 6$ mm; L = 8mm	Same as Daculsi Vit β -TCP	In vivo: condylar implantation in adult New Zealand white rabbits	Daculsi Viv β -TCP [13]
Poly(D,L-lactide)(4:96)	Purac Biochem B.V.; Gorinchem, The Netherlands (Purasorb PLD9655)	4.8×10^{-11}	4.8×10^{-11}	β -TCP	CAM Implants B.V., Leiden, The Nether- lands	3.0×10^{-11}	2.0	Dense cylinder: Pure Polymer: $\phi =$ (2.6...2.7)mm; Composite: $\phi =$ (2.9...3.0)mm; L = 30 mm	Polymer granules mixed w ceramic particles + melt-extrusion into cylindrical billet + solid state die-drawing at 120 °C to final diameter + γ -radiation (25 kGy)	In vitro: PBS (pH = 7.4) at 37 °C, w/o shaking, w formightly replacement	Niemela LD β -TCP [14]

(continued)

Table B.2 (continued)

Polymer characteristics			Ceramic characteristics			Sample characteristics			Degradation protocol	Case code and reference	
Type	Manufacturer	k_1 (s^{-1})	k'_2 ($m^3 mol^{-1} s^{-1}$)	Type	Manufacturer	A_d ($mol m^{-2} s^{-1}$)	θ (1)	Structure and Morphology			Fabrication method
Poly(DL-lactide)(50:50)	In-house production: ring-opening polymerisation of D,L-lactide	7.6×10^{-10}	7.2×10^{-10}	α -TCP	In-house production: wet chemistry route	7.0×10^{-12}	2.0	Dense film: $4 \times 8 mm^2 \times 1 mm$	Polymer dissolved in chloroform + ceramic dispersed in ethanol + addition of ceramic dispersion + press moulding at $85^\circ C$ and (10...15)MPa for 10 min	In vitro: PBS (pH = 7.4) at $37^\circ C$, w shaking at 100 rpm, ceramic dispersion + $100 mL g^{-1}$	Zheng α -TCP [15]
Poly(DL-lactide)(50:50)	Merck; Darmstadt, Germany	2.6×10^{-10}	4.2×10^{-10}	β -TCP	In-house production: as-received TCP (Merck; Darmstadt, Germany) granulated and calcined at $1200^\circ C$ for 2 h + crushing, milling and sieving	7.0×10^{-10}	2.0	Dense cylinder: $\Phi = 5 mm$, $L = 40 mm$	Polymer melted + mixed w ceramic + sintered at $90^\circ C$ and (50...80)MPa for 30 min	In vitro: saline at $37^\circ C$	Lin β -TCP [16]
Poly(DL-lactide)(50:50)	Boehringer Ingelheim, Germany (Resomer R270)	1.0×10^{-10}	9.0×10^{-11}	β -TCP (Assumed)	In-house production: as-received TCP (Fluka AG, Switzerland) milled in ethanol	7.0×10^{-13}	2.0	Dense rod: $20 \times 3 mm^2 \times 2 mm$	Polymer and ceramic mixing + injection moulding + γ -irradiation 25kGy	In vivo: dorsal muscles implantation in rats	Heidemann β -TCP [17]

(continued)

Table B.2 (continued)

Polymer characteristics				Ceramic characteristics			Sample characteristics			Degradation protocol	Case code and reference
Type	Manufacturer	k_1 (s^{-1})	k_2' ($m^3 mol^{-1} s^{-1}$)	Type	Manufacturer	A_d ($mol m^{-2} s^{-1}$)	θ (1)	Structure and Morphology	Fabrication method		
Poly(L-co-D,L-lactide)(70:30)	Boehringer Ingelheim; Ingelheim am Rhein, Germany (Resomer LR70x family)	9.8×10^{-11}	9.6×10^{-11}	β -TCP	CAM Implants B.V., Leiden, The Netherlands	3.0×10^{-11}	2.0	Dense cylinder: Pure Polymer: $\Phi = (2.6...2.7)$ mm; Composite: $\Phi = (2.9...3.0)$ mm; L = 30 mm	Polymer granules mixed w ceramic particles + melt-extrusion into cylindrical billet + solid state die-drawing at 120 °C to final diameter + γ -radiation (25 kGy)	In vitro: PBS (pH = 7.4) at 37 °C, w/o shaking, w fortnightly replacement [18]	Niemela LDL TCP [18]
Poly(L-co-D,L-lactide)(70:30)	Boehringer Ingelheim; Ingelheim am Rhein, Germany (Resomer LR70x family)	5.4×10^{-11}	5.4×10^{-11}	α -TCP	Not reported	3.0×10^{-12}	2.0	Porous cylinder: $\Phi = 10$ mm, L = 15 mm	Ceramic and polymer granules mixed + sintered at 145 °C to final shape + γ -radiation (25 kGy)	In vitro: Sorensen buffer (pH = 7.4) at 37 °C, 50 mL/sample, pH kept constant at 7.4 [19]	Ignatius P α -TCP [19]
Poly(L-co-D,L-lactide)(70:30)	Biovision; Imnerau, Germany (Polypin 2.0) and Boehringer Ingelheim; Ingelheim am Rhein, Germany (Resomer LR70x family)	4.5×10^{-11} (3.0×10^{-11})	3.2×10^{-11} (2.0×10^{-11})	β -TCP	Not reported	3.0×10^{-11}	2.0	Dense cylinder: $\Phi = 2$ mm, L = 35 mm	Ceramic granules added to heated polymer + injection-moulding at 210 °C + γ -radiation (28 kGy)	In vitro: Sorensen buffer (pH = 7.4) at 37 °C, 200 mL g^{-1} , pH kept constant at 7.4 [19]	Ignatius D β -TCP [19]

(continued)

Table B.2 (continued)

Polymer characteristics			Ceramic characteristics				Sample characteristics				
Type	Manufacturer	k_1 (s^{-1})	k'_2 ($m^3 mol^{-1} s^{-1}$)	Type	Manufacturer	A_d ($mol m^{-2} s^{-1}$)	θ (1)	Structure and Morphology	Fabrication method	Degradation protocol	Case code and reference
Poly(L-co-DL-lactide)(70:30)	Purac Biochem B.V.; Gorinchem, The Netherlands (Purasorb PLDL7028)	1.9×10^{-11}	1.8×10^{-11}	β -TCP	Plasma Biotol Ltd.; Tideswell, United Kingdom	5.0×10^{-11}	2.0	Porous disk: $\Phi = 15$ mm, $t = 3$ mm	Polymer dissolved in dioxane at 3 wt % + ceramic addition + stirring of the mixture + freeze drying	In vitro: PBS (pH = 7.4) at 37 °C, 10 mL/sample	Haaparanta 3w β -TCP [20]
Poly(L-co-DL-lactide)(70:30)	Purac Biochem B.V.; Gorinchem, The Netherlands (Purasorb PLDL7028)	2.7×10^{-11}	2.5×10^{-11}	β -TCP	Plasma Biotol Ltd.; Tideswell, United Kingdom	5.0×10^{-11}	2.0	Porous disk: $\Phi = 15$ mm, $t = 3$ mm	Polymer dissolved in dioxane at 2 wt % + ceramic addition + stirring of the mixture + freeze drying	In vitro: PBS (pH = 7.4) at 37 °C, 10 mL/sample	Haaparanta 2w β -TCP [20]

B.1.3 Poly-lactide-co-glycolide Matrix

Table B.3 Results output by the TCP composites degradation model for cases with a poly(lactide-co-glycolide) matrix. k_1 is the non-catalytic polymer degradation rate, k_2 is the autocatalytic polymer degradation rate, A_d is the ceramic dissolution rate and θ is the power ceramic dissolution law exponent. Abbreviations used to specify the tricalcium phosphate type can be found in Table 4.2. A dashed line between rows indicates that data displayed in those consecutive rows belong to the same researcher or research group. A solid line between rows indicates no known author relationship for the data

Polymer characteristics			Ceramic characteristics			Sample characteristics			Degradation protocol	Case code and reference	
Type	Manufacturer	k_1 (s^{-1})	k_2 ($m^3 mol^{-1} s^{-1}$)	Type	Manufacturer	A_d ($mol m^{-2} s^{-1}$)	θ (1)	Structure and Morphology			Fabrication method
Poly(L-lactide-co-glycolide) (70:30)	In-house production: ring-opening polymerisation of L-lactide and glycolide w stannous octoate	4.2×10^{-10}	2.8×10^{-10}	β -TCP (Assumed)	Forth Reagent Factory; Shanghai, China	3.0×10^{-11}	2.0	Porous (90 %) slab: $6 \times 6 mm^2$, $L = 13 mm$	Polymer dissolved in dioxane + ceramic addition + mixing + low-temperature deposition of the slurry using a computer-driven nozzle + freeze-drying	In vitro: distilled water w NaH_2 at $37^\circ C$, w/o shaking, w/o replacement, $50 mL g^{-1}$	Yang F. β -TCP [21]
Poly(L-lactide-co-glycolide) (70:30)	Changchun Institute of Applied Chemistry; Chinese Academy of Sciences, China	3.2×10^{-10}	3.2×10^{-10}	β -TCP	Naitional Engineering Research Centre for Biomaterials; Sichuan University, China	3.0×10^{-13}	2.0	Porous cylinder: $\Phi = 6 mm$, $L = 12 mm$	Polymer dissolved in chloroform + ceramic and porogen addition + freezing + solvent extraction w ethanol at $-18^\circ C$ + porogen leached w water	In vitro: PBS (pH = 7.4) at $37^\circ C$, w shaking at 50 rpm, weekly replacement, $4 mL/sample$	Yang Y. S β -TCP [21]

(continued)

Table B.3 (continued)

Polymer characteristics			Ceramic characteristics			Sample characteristics			Fabrication method		Degradation protocol	Case code and reference
Type	Manufacturer	k_1 (s^{-1})	k'_2 ($m^3 mol^{-1} s^{-1}$)	Type	Manufacturer	A_d ($mol m^{-2} s^{-1}$)	θ (1)	Structure and Morphology				
Poly(L-lactide-co-glycolide) (70:30)	Changchun Institute of Applied Chemistry; Chinese Academy of Sciences, China	3.5×10^{-10}	3.5×10^{-10}	β -TCP	National Engineering Research Centre for Biomaterials; Sichuan University, China	3.0×10^{-13}	2.0	Porous cylinder: $\Phi = 6$ mm, L = 12 mm	Polymer dissolved in chloroform + ceramic and porogen addition + freezing + solvent extraction w ethanol at $-18^\circ C$ + porogen leached w water	In vitro: PBS (pH = 7.4) at $37^\circ C$, w 8 h d^{-1} of cyclic loading at 1 Hz and (0.5...0.6) mm displacement and w 16 h d^{-1} of shaking at 50 rpm, weekly buffer replacement, 4 mL/sample	Yang Y. D β -TCP [22]	
Poly(L-lactide-co-glycolide) (70:7:29:3)	In-house production: polymerisation using preset microwave power of L-lactide and glycolide w stannous octoate	2.0×10^{-9} (1.5×10^{-9})	2.2×10^{-9} (1.5×10^{-9})	β -TCP	In-house production: wet chemistry route using $Ca(NO_3)_2 \cdot 4H_2O$ and $(NH_4)_2HPO_4$ + aging, filtering and drying + calcining at $800^\circ C$ for 2 h	7.0×10^{-12}	2.0	Semi-dense cylinder: $\Phi = 13$ mm, L = 12 mm	Ceramic added to initiator and monomers solution + polymerisation using microwave power	In vitro: SBF (pH = 7.4) at $37^\circ C$, w/o shaking, w/o replacement, 15 mL/sample	Jin β -TCP [23]	
Poly(DL-lactide-co-glycolide) (44:6:55:4)	Mitsui Chemicals; Tokyo, Japan: polymerisation of DL-lactide and glycolide using stannous octoate	3.0×10^{-9}	3.0×10^{-9}	β -TCP (Assumed)	Wako Pure Chemical Industries; Tokyo, Japan	5.0×10^{-11}	2.0	Dense film: $15 \times 15 \text{ mm}^2 \times 0.5 \text{ mm}$	Polymer dissolved in dioxane + ceramic addition + freeze-drying of the solvent east solution + compression moulding at $60^\circ C$ and 4.9 MPa	In vitro: PBS (pH = 7.4) at $37^\circ C$, w/o shaking, weekly replacement, 40 mL/sample	Ara β -TCP [24]	

(continued)

Table B.3 (continued)

Polymer characteristics			Ceramic characteristics			Sample characteristics			Fabrication method		Degradation protocol	Case code and reference
Type	Manufacturer	k_1 (s^{-1})	k'_2 ($m^3 mol^{-1} s^{-1}$)	Type	Manufacturer	A_d ($mol m^{-2} s^{-1}$)	θ (1)	Structure and Morphology				
Poly(D,L-lactide-co-glycolide) (50:50)	Lakeshore Biomaterials; Birmingham, Alabama, USA (5050 DLG 5E)	2.4×10^{-9}	2.4×10^{-9}	α -TCP	In-house production: wet chemistry route using $Ca(OH)_2$ and H_3PO_4 + aging for 1 d and drying + grinding + calcining at $1400^\circ C$ for 4 h + quenching + grinding and sieving through a $100 \mu m$ mesh + ball-milling for 1 h in acetone	1.0×10^{-12}	2.0	Dense disk: $\phi = 3.5$ mm, $t = 1$ mm	Polymer dissolved in acetone + ceramic addition + solvent casting + injection-moulding at $135^\circ C$ and 0.5 MPa into cylinders + cut into final shape	In vitro: PBS (0.01 M, pH = 7.4) at $37^\circ C$, w/o shaking, w/o replacement, $166 mL g^{-1}$	Bennett nC α -TCP [25]	
Poly(D,L-lactide-co-glycolide) (50:50)	Lakeshore Biomaterials; Birmingham, Alabama, USA (5050 DLG 5E)	1.4×10^{-9}	1.4×10^{-9}	α -TCP	In-house production: wet chemistry route. Same as Bennett nC α -TCP	5.0×10^{-13}	2.0	Dense disk: $\phi = 3.5$ mm, $t = 1$ mm	Same as Bennett nC α -TCP	In vitro: PBS (0.01 M, pH = 7.4) w 50 % v/v of D_2O at $37^\circ C$, w/o shaking, w/o replacement, $166 mL g^{-1}$	Bennett nC D ₂ 50 α -TCP [25]	
Poly(D,L-lactide-co-glycolide) (50:50)	Lakeshore Biomaterials; Birmingham, Alabama, USA (5050 DLG 5E)	5.9×10^{-10}	5.9×10^{-10}	α -TCP	In-house production: wet chemistry route. Same as Bennett nC α -TCP	3.0×10^{-13}	2.0	Dense disk: $\phi = 3.5$ mm, $t = 1$ mm	Same as Bennett nC α -TCP	In vitro: PBS (0.01 M, pH = 7.4) w 100 % v/v of D_2O at $37^\circ C$, w/o shaking, w/o replacement, $166 mL g^{-1}$	Bennett nC D ₂ 100 α -TCP [25]	

(continued)

Table B.3 (continued)

Polymer characteristics			Ceramic characteristics				Sample characteristics			Fabrication method		Degradation protocol	Case code and reference
Type	Manufacturer	k_1 (s ⁻¹)	k'_2 (m ³ mol ⁻¹ s ⁻¹)	Type	Manufacturer	A_d (mol m ⁻² s ⁻¹)	θ (1)	Structure and Morphology					
Poly(DL-lactide-co-glycolide) (50:50)	Lakeshore Biomaterials; Birmingham, Alabama, USA (5050 DLG 5E)	2.4×10^{-9}	2.4×10^{-9}	α -TCP	In-house production: wet chemistry route using Ca(OH) ₂ and H ₃ PO ₄ + aging for 1 d and drying + grinding into powder + calcining at 1400 °C for 4 h + grinding and sieving through a 38 μ m mesh	3.0×10^{-12}	2.0	Dense disk: $\Phi = 3.5$ mm, $t = 1$ mm	Polymer dissolved in acetone + ceramic addition + solvent casting + injection-moulding at 135 °C and 0.5 MPa into cylinders + cut into final shape	In vitro: same as Bennett mC α -TCP	Bennett iC α -TCP [25]		
Poly(DL-lactide-co-glycolide) (50:50)	Lakeshore Biomaterials; Birmingham, Alabama, USA (5050 DLG 5E)	2.4×10^{-9}	2.4×10^{-9}	α -TCP	In-house production: wet chemistry route using Ca(OH) ₂ and H ₃ PO ₄ + aging for 1 d and drying + grinding into powder + calcining at 1400 °C for 4 h + quenching + grinding and sieving through a 100 μ m mesh	1.0×10^{-10}	2.0	Dense disk: $\Phi = 3.5$ mm, $t = 1$ mm	Polymer dissolved in acetone + ceramic addition + solvent casting + injection-moulding at 135 °C and 0.5 MPa into cylinders + cut into final shape	In vitro: PBS (0.01 M, pH = 7.4) at 37 °C, w/o shaking, w/o replacement, 166 mL.g ⁻¹	Bennett mC α -TCP [25]		
Poly(DL-lactide-co-glycolide) (50:50)	Lakeshore Biomaterials; Birmingham, Alabama, USA (5050 DLG 5E)	1.4×10^{-9}	1.4×10^{-9}	α -TCP	In-house production: same as Bennett mC	5.0×10^{-11}	2.0	Dense disk: $\Phi = 3.5$ mm, $t = 1$ mm	Same as Bennett mC α -TCP	In vitro: PBS (0.01 M, pH = 7.4) at 37 °C w 50 % v/v of D ₂ O, w/o shaking, w/o replacement, 166 mL.g ⁻¹	Bennett mC D ₂ 50 α -TCP [25]		

(continued)

Table B.3 (continued)

Polymer characteristics				Ceramic characteristics				Sample characteristics				Case code and reference
Type	Manufacturer	k_1 (s^{-1})	k'_2 ($m^2 mol^{-1} s^{-1}$)	Type	Manufacturer	A_d ($mol.m^{-2} s^{-1}$)	θ (1)	Structure and Morphology	Fabrication method	Degradation protocol		
Poly(D,L-lactide-co-glycolide) (50:50)	Lakeshore Biomaterials; Birmingham, Alabama, USA (5050 DLG 5E)	5.9×10^{-10}	5.9×10^{-10}	α -TCP	In-house production; same as Bennett mC	1.0×10^{-11}	2.0	Dense disk; $\Phi = 3.5$ mm, $t = 1$ mm	Same as Bennett mC α -TCP	In vitro: PBS (0.01 M, pH = 7.4) at 37 °C w 100 % v/v of D ₂ O, w/o shaking, w/o replacement, 166 mL.g ⁻¹	Bennett mC D2 100 α -TCP [25]	
Poly(D,L-lactide-co-glycolide) (50:50)	SurModics Pharmaceuticals, USA	1.0×10^{-9}	1.8×10^{-9}	α -TCP	In-house production; wet chemistry route using Ca(OH) ₂ and H ₃ PO ₄ + aging for 1 d and drying + calcining at 1400 °C for 4 h + quenching + grinding and sieving to 75 μ m	3.0×10^{-13}	2.0	Dense disk; $\Phi = 6$ mm, $t = 2$ mm	Ceramic dispersed in polymer using a co-rotating twin screw extruder + cut into pellets + injection-moulded to cylinders at 130 °C and 0.5 MPa + cut into final shape	In vitro: PBS (0.01 M, pH = 7.4) at 37 °C, w/o shaking, w/o replacement	Ege α -TCP [26, 27]	
Poly(D,L-lactide-co-glycolide) (50:50)	Lakeshore Biomaterials; Birmingham, Alabama, USA	2.8×10^{-9}	2.2×10^{-9}	α -TCP	In-house production; wet chemistry route using Ca(OH) ₂ and H ₃ PO ₄ + aging for 1 d and drying + calcining at 1300 °C for 4 h + quenching + grinding and sieving to 75 μ m + attritor-milled for 4 h in acetone	9.0×10^{-13}	2.0	Dense film: 8×8 mm ² \times 1 mm	Polymer addition to milled ceramic + solvent casting + hot-pressed at 150 °C into sheets + cut into final shape	In vitro: PBS (0.01 M, pH = 7.4) at 37 °C, w/o shaking, w/o replacement, 166 mL.g ⁻¹	Yang Z. 5050 nC α -TCP [28, 29]	

(continued)

Table B.3 (continued)

Polymer characteristics			Ceramic characteristics			Sample characteristics			Fabrication method		Degradation protocol	Case code and reference
Type	Manufacturer	k_1 (s^{-1})	k_2^* ($m^3 \text{ mol}^{-1} s^{-1}$)	Type	Manufacturer	A_d ($\text{mol m}^{-2} s^{-1}$)	θ (1)	Structure and Morphology				
Poly(D,L-lactide-co-glycolide) (50:50)	Lakeshore Biomaterials; Birmingham, Alabama, USA	2.8×10^{-9}	2.2×10^{-9}	α -TCP	In-house production; wet chemistry route using $\text{Ca}(\text{OH})_2$ and H_3PO_4 + aging for 1 d and drying + calcining at 1300°C for 4 h + quenching + grinding and sieving to $75 \mu\text{m}$	3.0×10^{-11}	2.0	Dense film: $8 \times 8 \text{ mm}^2 \times 1 \text{ mm}$	Ceramic and polymer mixed + solvent casting + hot-pressed at 150°C into sheets + cut into final shape	In vitro: PBS (0.01 M, pH = 7.4) at 37°C , w/o shaking, w/o replacement, 166 mL g^{-1}	Yang Z. 5(6) mC α -TCP [28, 29]	
Poly(D,L-lactide-co-glycolide) (50:50)	Alkermes Medisorb; Cincinnati, USA	4.5×10^{-9}	2.9×10^{-9}	α -TCP	In-house production; wet chemistry route + drying for 2 d + grinding into powder + calcining at 1200°C for 4 h + ball-milling for 11 h + sieving through a $75 \mu\text{m}$ mesh	3.0×10^{-11}	2.0	Dense disk: $\Phi = 15 \text{ mm}$, $t = 2.3 \text{ mm}$	Polymer and ceramic kneading at 200°C + hot-pressed at 200°C into final shape	In vitro: PBS (0.03 M, pH = 7.4) at 37°C , w/o replacement, periodic agitation, w/o replacement, 30 mL/sample	Mellon α -TCP [30]	
Poly(D,L-lactide-co-glycolide) (50:50)	Lakeshore Biomaterials; Alabama, USA	2.1×10^{-9}	2.1×10^{-9}	α -TCP	In-house production; wet chemistry route using $\text{Ca}(\text{OH})_2$ and H_3PO_4 + aging for 1 d and drying + ball-milling for 4 h + sieving using a $75 \mu\text{m}$ mesh + calcining at 1400°C for 4 h + attritor-milled in acetone for 4 h	5.0×10^{-12}	2.0	Dense disk: $\Phi = 8 \text{ mm}$, $t = 900 \mu\text{m}$	Polymer addition to milled ceramic + drying + hot-pressed at 90°C into sheets + cut w a circular punch into final shape	In vitro: distilled-deionised water at 37°C , w/o shaking, w/o replacement, 40 mL/sample	Barrett α -TCP [31, 32]	

(continued)

Table B.3 (continued)

Polymer characteristics				Ceramic characteristics			Sample characteristics				Sample characteristics		Degradation protocol	Case code and reference
Type	Manufacturer	k_1 (s^{-1})	k'_2 ($m^3 mol^{-1} s^{-1}$)	Type	Manufacturer	A_d ($mol m^{-2} s^{-1}$)	θ (1)	Structure and Morphology	Fabrication method	Degradation protocol	Case code and reference			
Poly(DL-lactide-co-glycolide) (50:50)	SurModics Pharmaceuticals, USA	3.0×10^{-9}	2.4×10^{-9}	α -TCP	In-house production: wet chemistry route using $Ca(OH)_2$ and H_3PO_4 + aging for 1 d and drying + grinding and sieving using a 75 μm mesh + calcining at 1200 °C for 4 h + grinding and sieving to 75 μm	5.0×10^{-10}	2.0	Dense disk: $\Phi = 10$ mm, $t = 1.5$ mm	Polymer and ceramic kneading at 200 °C + hot-pressed at 200 °C into final shape	In vitro: PBS (0.01 M, pH = 7.4) at 37 °C, w periodic agitation, w/o replacement, 166 mL g^{-1}	Ehrenfried α -TCP [33]			
Poly(DL-lactide-co-glycolide) (50:50)	SurModics Pharmaceuticals, USA	1.0×10^{-9}	1.8×10^{-9}	β -TCP	In-house production: wet chemistry route using $Ca(OH)_2$ and H_3PO_4 + aging for 1 d and drying + calcining at 1100 °C for 4 h + grinding and sieving to 75 μm	7.0×10^{-13}	2.0	Dense disk: $\Phi = 6$ mm, $t = 2$ mm	Ceramic dispersed in polymer using a co-rotating twin screw extruder + cut into pellets + injection-moulded to cylinders at 130 °C and 0.5 MPa + cut into final shape	In vitro: PBS (0.01 M, pH = 7.4) at 37 °C, w/o shaking, w/o replacement	Ege β -TCP [26]			
Poly(DL-lactide-co-glycolide) (50:50)	Alkermes Medisorb; Cincinnati, USA	4.5×10^{-9}	2.9×10^{-9}	β -TCP	In-house production: wet chemistry route + drying for 2 d + grinding into powder + calcining at 1000 °C for 4 h + ball-milling for 1 h + sieving through a 75 μm mesh	9.0×10^{-12}	2.0	Dense disk: $\Phi = 15$ mm, $t = 2.5$ mm	Polymer and ceramic kneading at 200 °C + hot-pressed at 200 °C into final shape	In vitro: PBS (0.03 M, pH = 7.4) at 37 °C, w periodic agitation, w/o replacement, 30 mL/sample	Mellon β -TCP [30]			

(continued)

Table B.3 (continued)

Polymer characteristics				Ceramic characteristics			Sample characteristics			Degradation protocol	Case code and reference
Type	Manufacturer	k_1 (s^{-1})	K_2 ($m^3 mol^{-1} s^{-1}$)	Type	Manufacturer	A_d ($mol m^{-2} s^{-1}$)	θ (1)	Structure and Morphology	Fabrication method		
Poly(DL-lactide-co-glycolide) (75:25)	Lakeshore Biomaterials; Birmingham, Alabama, USA	1.6×10^{-9}	1.1×10^{-9}	α -TCP	In-house production: wet chemistry route using $Ca(OH)_2$ and H_3PO_4 + aging for 1 d and drying + calcining at $1300^\circ C$ for 4 h + quenching + grinding and sieving to $75 \mu m$ + attritor-milled for 4 h in acetone	7.0×10^{-13}	2.0	Dense film: $8 \times 8 \text{ mm}^2 \times 1 \text{ mm}$	Polymer addition to milled ceramic + solvent casting + hot-pressed at $150^\circ C$ into sheets + cut into final shape	In vitro: PBS (0.01 M, pH = 7.4) at $37^\circ C$, w/o shaking, w/o replacement, $166 \text{ mL} \cdot g^{-1}$	Yang Z. 7525 nC α -TCP [29]
Poly(DL-lactide-co-glycolide) (75:25)	Lakeshore Biomaterials; Birmingham, Alabama, USA	1.6×10^{-9}	1.1×10^{-9}	α -TCP	In-house production: wet chemistry route using $Ca(OH)_2$ and H_3PO_4 + aging for 1 d and drying + calcining at $1300^\circ C$ for 4 h + quenching + grinding and sieving to $75 \mu m$	3.0×10^{-11}	2.0	Dense film: $8 \times 8 \text{ mm}^2 \times 1 \text{ mm}$	Ceramic and polymer mixed + solvent casting + hot-pressed at $150^\circ C$ into sheets + cut into final shape	In vitro: PBS (0.01 M, pH = 7.4) at $37^\circ C$, w/o shaking, w/o replacement, $166 \text{ mL} \cdot g^{-1}$	Yang Z. 7525 mC α -TCP [29]

B.1.4 Miscellaneous Polymers Matrixes

Table B.4 Results output by the TCP composites degradation model for cases with a miscellaneous polymer matrix. k_1 is the non-catalytic polymer degradation rate, k_2 is the autocatalytic polymer degradation rate, A_d is the ceramic dissolution rate and θ is the power ceramic dissolution law exponent. Abbreviations used to specify the tricalcium phosphate type can be found in Table 4.2. A dashed line between rows indicates that data displayed in those consecutive rows belong to the same researcher or research group. A solid line between rows indicates no known author relationship for the data

Polymer characteristics			Ceramic characteristics			Sample characteristics			Fabrication method		Degradation protocol	Case code and reference
Type	Manufacturer	k_1 (s^{-1})	k_2 ($m^3 mol^{-1} s^{-1}$)	Type	Manufacturer	A_d ($mol m^{-2} s^{-1}$)	θ (1)	Structure and Morphology				
Poly(L-lactide-co- ϵ -caprolactone)(70:30)	Purac Biomaterials, The Netherlands (Purasorb PLC7015)	7.4×10^{-10}	3.4×10^{-10}	β -TCP	Plasma Biotol Ltd., United Kingdom	3.0×10^{-12}	2.0	Dense cylinder: $\Phi = 2.5$ mm, $L = 2.5$ mm	Polymer and ceramic fed to a co-rotating twin screw-extruder and processed at (25...130)°C into rod shape billets + cut to final shape + γ -irradiation (25 kGy)	In vitro: Sörensén buffer (pH = 7.4) at 37 °C, w shaking, w replacement every 2 days, 66 mL.g ⁻¹	Ahola 13 β -TCP [34]	
Poly(L-lactide-co- ϵ -caprolactone)(70:30)	Purac Biomaterials, The Netherlands (Purasorb PLC7015)	5.8×10^{-10} (4.0×10^{-10})	3.8×10^{-10} (2.6×10^{-10})	β -TCP	Plasma Biotol Ltd., United Kingdom	3.0×10^{-12}	2.0	Dense cylinder: $\Phi = 2.5$ mm, $L = 2.5$ mm	Same as Ahola 13	In vitro: Sörensén buffer (pH = 7.4) at 37 °C, w shaking, w fortnightly replacement, 66 mL.g ⁻¹	Ahola 12 β -TCP [35]	
Poly(L-lactide-co- ϵ -caprolactone)(92:8)	In-house production: bulk copolymerisation of L-lactide and ϵ -caprolactone using stannous 2-ethylhexanoate	1.8×10^{-10}	1.4×10^{-10}	β -TCP	In-house production: wet chemistry route using $Ca(OH)_2$ and H_3PO_4 + aging, drying at 120°C, pelleted in 125 μ m + calcining at 800°C	3.0×10^{-12}	2.0	Dense film: 5×20 mm ² $\times 200$ μ m	Polymer and ceramic kneaded in a milled at 180°C and 30 rpm for 10 min + cooled in air for 15 min + hot-pressed at 180°C	In vitro: physiological saline (pH = 7.4) at 37 °C, w/o shaking, w/o replacement	Kikuchi PLCL β -TCP [36]	

(continued)

Table B.4 (continued)

Polymer characteristics			Ceramic characteristics				Sample characteristics			Degradation protocol	Case code and reference
Type	Manufacturer	k_1 (s^{-1})	k_2' ($m^3 mol^{-1} s^{-1}$)	Type	Manufacturer	A_d ($mol m^{-2} s^{-1}$)	θ (1)	Structure and Morphology	Fabrication method		
Poly(L-lactide)/poly(ethylene hexamethylene/sebacate) block copolymer (75.3:24.7)	In-house production: ring-opening polymerisation of L-lactide in the presence of poly(ethylene hexamethylene/sebacate) (60:40 in weight) using stannous octoate	2.3×10^{-10}	2.8×10^{-10}	β -TCP	Taihei Chemical; Tokyo, Japan	5.0×10^{-12}	2.0	Dense film: $20 \times 30 mm^2 \times 100 \mu m$	Blending polymer and ceramic in a plastomill mixer at 180 °C and 50 rpm for 10 min + compression moulding at 180 °C and 19.6 MPa	In vitro: PBS (pH = 7.4) at 37 °C, w/o shaking, weekly replacement, 50 mL/sample	Imai L100 β -TCP [37]
Poly(L-lactide)/poly(ethylene hexamethylene/sebacate) block copolymer (75.3:24.7)	In-house production: same as Imai L100	4.8×10^{-10}	4.9×10^{-10}	β -TCP	Taihei Chemical; Tokyo, Japan	1.0×10^{-11}	2.0	Dense film: $20 \times 30 mm^2 \times 250 \mu m$	Same as Imai L100	In vitro: same as Imai L100	Imai L250 β -TCP [37]
Poly(L-lactide)/poly(ethylene hexamethylene/sebacate) block copolymer (75.3:24.7)	In-house production: ring-opening polymerisation of L-lactide in the presence of poly(ethylene hexamethylene/sebacate) (60:40 in weight) using stannous octoate	1.0×10^{-10}	1.2×10^{-10}	β -TCP	In-house production: wet chemistry route + calcining at 800 °C	5.0×10^{-14}	2.0	Dense film: $5 \times 20 mm^2 \times 3 mm$	Blending polymer and ceramic in mill mixer at 180 °C and 20 rpm for 10 min + cooled in air to room temperature + compression moulding at 180 °C and 30 MPa + cutting to final shape	In vitro: physiological saline (pH = 7.4) at 37 °C, w/o shaking, 100 mL/sample	Kikuchi T800 β -TCP [38]
Poly(L-lactide)/poly(ethylene hexamethylene/sebacate) block copolymer (75.3:24.7)	In-house production: same as Kikuchi T800	5.0×10^{-11}	1.4×10^{-10}	β -TCP	In-house production: wet chemistry route + calcining at 1100 °C	5.0×10^{-13}	2.0	Dense film: $5 \times 20 mm^2 \times 3 mm$	Same as Kikuchi T800	In vitro: same as Kikuchi T800	Kikuchi T1100 β -TCP [38]

(continued)

Table B.4 (continued)

Polymer characteristics			Ceramic characteristics			Sample characteristics			Degradation protocol	Case code and reference	
Type	Manufacturer	k_1 (s^{-1})	k'_2 ($m^3 mol^{-1} s^{-1}$)	Type	Manufacturer	A_d ($mol m^{-2} s^{-1}$)	θ (1)	Structure and Morphology			Fabrication method
Poly(L-lactide)/poly(ethylene hexamethylene/sebacate) block copolymer (98.3:1.7)	In-house production: ring-opening polymerisation of L-lactide in the presence of poly(ethylene hexamethylene/sebacate) (97.3 in weight) using stannous octoate.	6.0×10^{-11}	2.2×10^{-11}	β -TCP	Taihei Chemical; Tokyo, Japan	3.0×10^{-12}	2.0	Dense film: $20 \times 30 mm^2 \times 100 \mu m$	Blending polymer and ceramic in a plastomill mixer at $180^\circ C$ and 50 rpm for 10 min + compression moulding at $180^\circ C$ and 19.6 MPa	In vitro: PBS (pH = 7.4) at $37^\circ C$, w/o shaking, weekly replacement, 50 mL/sample	Imai H100 β -TCP [4]
Poly(L-lactide)/poly(ethylene hexamethylene/sebacate) block copolymer (98.3:1.7)	In-house production: same as Imai H100	6.8×10^{-11}	7.0×10^{-11}	β -TCP	Taihei Chemical; Tokyo, Japan	5.0×10^{-12}	2.0	Dense film: $20 \times 30 mm^2 \times 250 \mu m$	Same as Imai H100	In vitro: same as Imai H100	Imai H250 β -TCP [4]
Poly(L-lactide-co-glycolide-co- ϵ -caprolactone)(75:11:14)	In-house production: bulk copolymerisation of L-lactide, glycolide and ϵ -caprolactone using stannous 2-ethylhexanoate	2.2×10^{-10}	2.8×10^{-10}	β -TCP	In-house production: wet chemistry route using $Ca(OH)_2$ and H_3PO_4 + aging, drying at $120^\circ C$, pelletised in $12.5 \mu m$ + calcining at $800^\circ C$	5.0×10^{-12}	2.0	Dense film: $5 \times 20 mm^2 \times 200 \mu m$	Polymer and ceramic kneaded in a milled at $180^\circ C$ and 30 rpm for 10 min + cooled in air for 15 min + hot-pressed at $180^\circ C$	In vitro: physiological saline (pH = 7.4) at $37^\circ C$, w/o shaking, w/o replacement	Kikuchi PLGC β -TCP [36]

B.2 Hydroxyapatite Case Studies Modelling Results

HA case studies with a poly(L-lactide) matrix are included in Appendix [B.2.1](#), (Table [B.5](#)), followed by cases with a poly(D,L-lactide) matrix in Appendix [B.2.2](#), (Table [B.6](#)), cases with poly(lactide-co-glycolide) matrix in Appendix [B.2.3](#), (Table [B.7](#)) and lastly, cases with miscellaneous polymers matrixes in Appendix [B.2.4](#), (Table [B.8](#)).

B.2.1 Poly(L-lactide) Matrix

Table B.5 Results output by the HA composites degradation model for cases with a poly(L-lactide) matrix. k_1 is the non-catalytic polymer degradation rate, k_2 is the autocatalytic polymer degradation rate, A_d is the ceramic dissolution rate and θ is the power ceramic dissolution law exponent. Abbreviations used to specify the hydroxyapatite type can be found in Table 5.2. A dashed line between rows indicates that data displayed in those consecutive rows belong to the same researcher or research group. A solid line between rows indicates no known author relationship for the data

Polymer characteristics										Ceramic characteristics									
Type	Manufacturer	k_1 (s^{-1})	k_2 ($m^3 mol^{-1} s^{-1}$)	Type	Manufacturer	A_d ($mol m^{-2} s^{-1}$)	θ (1)	Structure and Morphology	Fabrication method	Degradation protocol	Case code and reference								
Poly(L-lactide)	In-house production: ring-opening polymerisation of L-lactide at 140 °C	4.6×10^{-10}	2.0×10^{-10}	hHA	In-house production: wet chemistry route using $Ca(NO_3)_2$ and NH_4PO_3 + sintered + milled	5.0×10^{-14}	2.0	Dense cylinder: $\Phi = 5$ mm, L = 10 mm	Polymer and ceramic mixed at 160 °C + compression moulded	In vitro: PBS (pH = 7.4) at 37 °C, w shaking at 100 rpm	Huang hHA [39]								
Poly(L-lactide)	In-house production: ring-opening polymerisation of L-lactide using stannous octoate	8.0×10^{-11} (1.2 × 10 ⁻¹¹)	8.0×10^{-11} (1.2 × 10 ⁻¹¹)	IHA	In-house production: wet chemistry route w $Ca(OH)_2$ and H_3PO_4	2.0×10^{-14}	2.0	Dense bar: 30 × 5 mm ² × 2 mm	Polymer and ceramic blended at 190 °C for 5 min in the rheometer + hot press moulded at 190 °C and 15 MPa + annealed at 115 °C for 1 h	In vivo: intramuscular implantation in New Zealand white rabbits	Wang IHA [40]								
Poly(L-lactide)	In-house production: same as Wang IHA	8.0×10^{-11} (1.2 × 10 ⁻¹¹)	8.0×10^{-11} (1.2 × 10 ⁻¹¹)	gP-IHA	In-house production: same as Wang nHA + PLLA polymerised on the nHA	5.0×10^{-15}	2.0	Dense bar: 30 × 5 mm ² × 2 mm	Same as Wang nHA	In vivo: same as Wang nHA	Wang gP-IHA [40]								
Poly(L-lactide)	Jamplast; Ellisville, Missouri, USA	3.5×10^{-10} (1.2 × 10 ⁻¹⁰)	2.3×10^{-10} (8.0 × 10 ⁻¹¹)	ICCDHA	In-house production: wet chemistry route using Tris-base SBF	1.0×10^{-15}	2.0	Porous fibrous mat (t: NR)	Polymer dissolved in chloroform + ceramic added + ultrasonicated for 1 h + electrospinning	In vitro: standard conditions (PBS at 37 °C, 25 mL/sample)	Zhou ICCDHA [41]								

(continued)

Table B.5 (continued)

Polymer characteristics			Ceramic characteristics				Sample characteristics				Degradation protocol	Case code and reference
Type	Manufacturer	k_1 (s ⁻¹)	k'_2 (m ³ mol ⁻¹ s ⁻¹)	Type	Manufacturer	A_d (mol m ⁻² s ⁻¹)	θ (1)	Structure and Morphology	Fabrication method			
Poly(L-lactide)	Purac Biochem B.V.; Gorinchem, The Netherlands; polymerised after mixing HA w L-lactide	3.0×10^{-10}	2.4×10^{-10}	hHA	Merck; Darmstadt, Germany; as-received HA + sintered in N ₂ + crushed and milled	1.0×10^{-12}	2.0	Dense film: $15 \times 10 \text{ mm}^2 \times 3 \text{ mm}$	Polymerised in an ampule w ceramic + machined to desired dimensions	In vitro: PBS at 37°C, weekly replacement, 40 mL/sample	Verheyen 92 Vit hHA [42]	
Poly(L-lactide)	Purac Biochem B.V.; Gorinchem, The Netherlands; see Verheyen 92 Vit	1.3×10^{-10}	2.6×10^{-10}	hHA	Merck; Darmstadt, Germany; see Verheyen 92 Vit	5.0×10^{-12}	2.0	Dense film: $15 \times 10 \text{ mm}^2 \times 3 \text{ mm}$	Same as Verheyen 92 Vit	In vitro: PBS at 37°C, weekly replacement, w shaking, 10 mL/sample	Verheyen 93 hHA [43]	
Poly(L-lactide)	Purac Biochem B.V.; Gorinchem, The Netherlands; see Verheyen 92 Vit	3.0×10^{-10}	2.0×10^{-10}	hHA	Merck; Darmstadt, Germany; see Verheyen 92 Vit	3.0×10^{-12}	2.0	Dense film: $15 \times 10 \text{ mm}^2 \times 3 \text{ mm}$	Same as Verheyen 92 Vit	In vitro: subcutaneous implantation in mature female goats	Verheyen 92 Viv hHA [42]	
Poly(L-lactide)	Biomater, Germany (L9000)	1.6×10^{-10}	1.0×10^{-10}	bnHA	Aldrich Chemistry, USA	1.0×10^{-14}	2.0	Porous (90%) disk: $\phi = 15 \text{ mm}$, t: NR	Polymer dissolved in dioxane + blended w ceramic + freeze drying	In vitro: standard conditions (PBS at 37°C, w/6 shaking, 10 mL/sample)	Diaz PP/C bnHA [44]	
Poly(L-lactide)	Biomater, Germany (L9000)	2.0×10^{-10} (1.6×10^{-10})	1.2×10^{-10} (1.0×10^{-10})	bnHA	Aldrich Chemistry, USA	1.0×10^{-14}	2.0	Porous (90%) disk: $\phi = 15 \text{ mm}$, t: NR	Same as Diaz PP/C	In vitro: same as Diaz PP/C	Diaz C bnHA [44]	

(continued)

Table B.5 (continued)

Polymer characteristics			Ceramic characteristics				Sample characteristics			Degradation protocol	Case code and reference
Type	Manufacturer	k_1 (s^{-1})	k'_2 ($m^3 mol^{-1} s^{-1}$)	Type	Manufacturer	A_d ($mol m^{-2} s^{-1}$)	θ (1)	Structure and Morphology	Fabrication method		
Poly(L-lactide)	In-house production: ring-opening polymerisation of L-lactide using tin octoate and lauryl alcohol	1.2×10^{-10} (2.6×10^{-11})	6.0×10^{-11} (1.3×10^{-11})	pCHA	In-house production: wet chemistry route with $CaHPO_4$ and $CaCO_3$ in aqueous solution at $90^\circ C$ + filtered and dried + milled and sieved	3.0×10^{-13}	2.0	Dense cylinder: $\Phi = 3.2$ mm, $L = 30$ mm	Ceramic and polymer mixed + extruded + compression moulded at $103^\circ C$ + lathing	In vivo: standard conditions (PBS ($pH = 7.4$) at $37^\circ C$, w/o shaking)	Shikunami 99 pCHA [45]
Poly(L-lactide)	In-house production: see Shikunami 99	1.2×10^{-10} (7.5×10^{-11})	3.0×10^{-11} (1.7×10^{-11})	pCHA	In-house production: see Shikunami 99	3.0×10^{-13}	2.0	Dense cylinder: $\Phi = 3.2$ mm, $L = 30$ mm	Same as Shikunami 99	In vivo: medullary cavity implantation in Japanese white rabbits	Furukawa Med pCHA [46]
Poly(L-lactide)	In-house production: same as Shikunami 99	1.2×10^{-10}	3.0×10^{-11}	pCHA	In-house production: same as Shikunami 99	3.0×10^{-13}	2.0	Dense cylinder: $\Phi = 3.2$ mm, $L = 50$ mm	Same as Shikunami 99	In vivo: subcutis implantation in Japanese white rabbits	Furukawa Sub pCHA [46]
Poly(L-lactide)	In-house production: same as Shikunami 99	1.2×10^{-10}	3.0×10^{-11}	pCHA	In-house production: same as Shikunami 99	3.0×10^{-13}	2.0	Dense cylinder: $\Phi = 3.2$ mm, $L = 30$ mm	Same as Shikunami 99	In vivo: intercondylar implantation in male Japanese white rabbits	Ishii pCHA [47]
Poly(L-lactide)	In-house production: same as Shikunami 99	8.0×10^{-11}	4.0×10^{-11}	pCHA	In-house production: same as Shikunami 99	5.0×10^{-13}	2.0	Dense screw: $\Phi = 1.6$ mm, $L = 8$ mm	Same as Shikunami 99	In vivo: standard conditions	Shikunami 01 pCHA [48]

(continued)

Table B.5 (continued)

Polymer characteristics				Ceramic characteristics				Sample characteristics				Degradation protocol	Case code and reference
Type	Manufacturer	k_1 (s ⁻¹)	k_2 (m ³ mol ⁻¹ s ⁻¹)	Type	Manufacturer	A_d (mol m ⁻² s ⁻¹)	θ (1)	Structure and Morphology	Fabrication method				
Poly(L-lactide)	In-house production; see Shikunami 99	1.2×10^{-10} (7.5×10^{-11})	3.0×10^{-11} (1.7×10^{-11})	hHA	In-house production: pCHA production (see Shikunami 99) + calcined at 900 °C	5.0×10^{-13}	2.0	Dense cylinder: $\Phi = 3.2$ mm, $L = 30$ mm	Same as Shikunami 99	Same as Furukawa Med CHA	Furukawa Med hHA [46]		
Poly(L-lactide)	In-house production; same as Shikunami 99	8.0×10^{-11}	5.0×10^{-11}	hHA	In-house production: same as Furukawa Med HA	5.0×10^{-13}	2.0	Dense cylinder: $\Phi = 3.2$ mm, $L = 50$ mm	Same as Shikunami 99	In vivo: same as Furukawa Sub CHA	Furukawa Sub hHA [46]		
Poly(L-lactide)	In-house production; same as Shikunami 99	8.0×10^{-11}	5.0×10^{-11}	hHA	In-house production: same as Furukawa Med HA	1.1×10^{-12}	2.0	Dense cylinder: $\Phi = 3.2$ mm, $L = 30$ mm	Same as Shikunami 99	In vivo: same as Ishii CHA	Ishii hHA [47]		
Poly(L-lactide)	In-house production; same as Shikunami 99	1.2×10^{-10}	2.0×10^{-11}	hHA	In-house production: same as Furukawa Med HA	1.0×10^{-13}	2.0	Dense miniplate: $t = 1$ mm	Same as Shikunami 99	In vitro: standard conditions	Shikunami 01 hHA [48]		
Poly(L-lactide)	Shang Dong Institute (Dept. of Med. Pol.), China	6.0×10^{-12}	3.0×10^{-12}	IHA	In-house production: wet chemistry route using $\text{Ca}(\text{NO}_3)_2 \cdot 4\text{H}_2\text{O}$ and $(\text{NH}_4)_2\text{HPO}_4$ w/ ultrasonic stirring	3.0×10^{-15}	2.0	Dense film: $20 \times 20 \text{ mm}^2$ $\times 0.05$ mm	Polymer-ceramic mixture solvent casting	In vitro: PBS (pH = 7.4) at 37 °C, w/o shaking, 15 mL/sample	Deng SC IHA [49]		
Poly(L-lactide)	Shang Dong Institute (Dept. of Med. Pol.), China	1.0×10^{-11}	1.1×10^{-11}	IHA	In-house production: same as Deng SC	9.0×10^{-15}	2.0	Porous fibrous mat: $20 \times 20 \text{ mm}^2$ $\times 0.05$ mm	Polymer-ceramic mixture electrospraying	In vitro: same as Deng SC	Deng ES IHA [49]		
Poly(L-lactide)	Shang Dong Institute (Dept. of Med. Pol.), China	8.0×10^{-12}	1.9×10^{-11}	IHA	In-house production: same as Deng SC	9.0×10^{-15}	2.0	Porous fibrous mat: $20 \times 20 \text{ mm}^2$ $\times 0.05$ mm	Polymer-ceramic mixture electrospraying	In vitro: same as Deng SC	Sui IHA [50]		

(continued)

Table B.5 (continued)

Polymer characteristics			Ceramic characteristics			Sample characteristics			Degradation protocol	Case code and reference	
Type	Manufacturer	k_1 (s^{-1})	k'_2 ($m^3 mol^{-1} s^{-1}$)	Type	Manufacturer	A_d ($mol m^{-2} s^{-1}$)	θ (1)	Structure and Morphology			Fabrication method
Poly(L-lactide)	In-house production: ring-opening polymerisation of L-lactide	5.2×10^{-10}	4.8×10^{-10}	gP-IHA	In-house production: wet chemistry route + grafting with PLLA	2.0×10^{-14}	2.0	Porous fibrous mat: $20 \times 20 mm^2 \times 0.2 mm$	Polymer dissolved in chloroform + ceramic added + electrospinning	In vitro: PBS (pH = 7.4) at $37^\circ C$, w shaking at 90 rpm, 40 mL/sample	Xu PP/C gP-IHA [51]
Poly(L-lactide)	In-house production: same as Xu PP/C	1.1×10^{-9} (5.2×10^{-10})	8.0×10^{-10} (4.8×10^{-10})	gP-IHA	In-house production: same as Xu PP/C	4.0×10^{-15}	2.0	Porous fibrous mat: $20 \times 20 mm^2 \times 0.2 mm$	Same as Xu PP/C	In vitro: same as Xu PP/C	Xu C gP-IHA [51]

B.2.2 Poly(D,L-lactide) Matrix

Table B.6 Results output by the HA composites degradation model for cases with a poly(D,L-lactide) matrix. k_1 is the non-catalytic polymer degradation rate, k_2' is the autocatalytic polymer degradation rate, A_d is the ceramic dissolution rate and θ is the power ceramic dissolution law exponent. Abbreviations used to specify the hydroxyapatite type can be found in Table 5.2. A dashed line between rows indicates that data displayed in those consecutive rows belong to the same researcher or research group. A solid line between rows indicates no known author relationship for the data

Polymer characteristics			Ceramic characteristics			Sample characteristics			Degradation protocol	Case code and reference	
Type	Manufacturer	k_1 (s^{-1})	k_2' ($m^3 mol^{-1} s^{-1}$)	Type	Manufacturer	A_d ($mol m^{-2} s^{-1}$)	θ (1)	Structure and Morphology			Fabrication method
Poly(D,L-lactide)(4:96)	Nature Works LLC (#4042D)	1.7×10^{-10} (3.5×10^{-11})	1.7×10^{-10} (3.4×10^{-11})	ICHA	In-house production: wet chemistry route using bovine bone derived precursors	5.0×10^{-15}	2.0	Dense slab: $10 \times 10 mm^2 \times 4 mm$	Ceramic and polymer mixed at 170°C for 10 min + hot pressed at 180°C and 14 MPa for 5 min	In vitro: SBF (pH = 7.4) at 37°C, weekly replacement, 11.6 mL/sample	Rakmae 13 ICHA [52]
Poly(D,L-lactide)(4:96)	Nature Works LLC (#4042D)	1.7×10^{-10} (3.5×10^{-11})	1.7×10^{-10} (3.4×10^{-11})	pCHA	In-house production: above-mentioned ICHA heat treated at 800°C for 3 h	1.1×10^{-13}	2.0	Dense slab: $10 \times 10 mm^2 \times 4 mm$	Same as Rakmae 13 ICHA	In vitro: same as Rakmae 13 ICHA	Rakmae 13 pCHA [52]
Poly(D,L-lactide)(4:96)	Nature Works LLC (#4042D)	1.7×10^{-10} (3.5×10^{-11})	1.7×10^{-10} (3.4×10^{-11})	hHA	In-house production: above-mentioned ICHA heat treated at 1100°C for 3 h	7.0×10^{-14}	2.0	Dense slab: $10 \times 10 mm^2 \times 4 mm$	Same as Rakmae 13 ICHA	In vitro: same as Rakmae 13 ICHA	Rakmae 13 hHA [52]

(continued)

Table B.6 (continued)

Polymer characteristics			Ceramic characteristics			Sample characteristics			Fabrication method		Degradation protocol	Case code and reference
Type	Manufacturer	k_1 (s^{-1})	k'_2 ($m^3 mol^{-1} s^{-1}$)	Type	Manufacturer	A_d ($mol m^{-2} s^{-1}$)	θ (1)	Structure and Morphology				
Poly(DL-lactide)(4:96)	Nature Works LLC (#4042D)	7.0×10^{-11}	1.6×10^{-10}	hCHA	In-house production: ground and ball-milled bovine bone heat treated at 1100 °C for 3 h	7.0×10^{-14}	2.0	Dense slab: $60 \times 10 mm^2 \times 4 mm$	Ceramic and polymer mixed at 170 °C for 10 min + heated to 180 °C for 15 min + hot pressed at 180 °C and 14 MPa for 5 min	In vitro: PBS (0.1 M, pH = 7.4) at 37 °C, weekly replacement, 11.6 mL/sample	Rakmae 12 hCHA [53]	
Poly(DL-lactide)(4:96)	Nature Works LLC (#4042D)	6.0×10^{-11}	1.7×10^{-10}	sA-hCHA	In-house production: above-mentioned hCHA modified with APES (3-Aminopropyl-triethoxysilane)	5.0×10^{-12}	2.0	Dense slab: $60 \times 10 mm^2 \times 4 mm$	Same as Rakmae 12 C	In vitro: same as Rakmae 12 hCHA	Rakmae 12 sA-hCHA [53]	
Poly(DL-lactide)(4:96)	Nature Works LLC (#4042D)	6.0×10^{-11}	1.7×10^{-10}	sM-hCHA	In-house production: above-mentioned hCHA modified with MPTS (3-methacryloxypropyltrimethoxy-silane)	5.0×10^{-12}	2.0	Dense slab: $60 \times 10 mm^2 \times 4 mm$	Same as Rakmae 12 hCHA	In vitro: same as Rakmae 12 hCHA	Rakmae 12 sM-hCHA [53]	
Poly(DL-lactide)(15:85)	Purac Biochem BV, The Netherlands	2.0×10^{-10}	2.5×10^{-9}	hHA	Not reported: sintered, milled and sieved	3.0×10^{-11}	2.0	Dense film: $t = (0.3...0.5) mm$	Polymer dissolved in chloroform, ceramic added and mixture stirred + solvent evaporation	In vitro: PBS (pH = 7.2) at 37 °C, w/o replacement, w shaking, 11 mL/sample	Vander Meer hHA [54]	

(continued)

Table B.6 (continued)

Polymer characteristics			Ceramic characteristics			Sample characteristics			Fabrication method		Degradation protocol	Case code and reference
Type	Manufacturer	k_1 (s^{-1})	k_2' ($mol^3 mol^{-1} s^{-1}$)	Type	Manufacturer	A_d ($mol m^{-2} s^{-1}$)	θ (1)	Structure and Morphology				
Poly(DL-lactide)(50:50)	In-house production: ring-opening polymerisation of racemic DL-lactide	2.1×10^{-10}	5.0×10^{-11}	pCHA	In-house production: wet chemistry route	1.0×10^{-13}	2.0	Porous cylinder $\phi = 6$ mm, $L = 15$ mm	Composite fiber precipitation	In vivo: male Japanese white rabbits	Hasegawa pCHA [55]	
Poly(DL-lactide)(50:50)	In-house production: ring-opening polymerisation of DL-lactide + modification with gelatin	4.3×10^{-10} (2.6×10^{-10})	8.0×10^{-11} (5.0×10^{-11})	Mixture of pHA and ns-IHA	In-house production: grown in-situ with SBF	5.0×10^{-14}	2.0	Porous fibrous disk: $\phi = 11$ mm, $t = (250...300) \mu m$	Electrospun fibrous mats w in-situ grown ceramic	In vitro: PBS (pH = 7.4) w sodium azide at 37 °C, 20 mL/sample	Zou 12 Vit pns-IHA [56]	
Poly(DL-lactide)(50:50)	In-house production: same as Zou 12 Vit pns-IHA	1.4×10^{-10} (9.0×10^{-11})	1.5×10^{-10} (1.0×10^{-10})	Mixture of pHA and ns-IHA	In-house production: same as Zou 12 Vit pns-IHA	5.0×10^{-15}	2.0	Porous fibrous disk: $\phi = 11$ mm, $t = (250...300) \mu m$	Re-electrospun after dissolving above-mentioned mat	In vitro: same as Zou 12 Vit pns-IHA	Zou 12 Vit R pns-IHA [56]	
Poly(DL-lactide)(50:50)	In-house production: same as Zou 12 Vit pns-IHA	4.4×10^{-10} (3.7×10^{-10})	4.7×10^{-10} (1.8×10^{-10})	Mixture of pHA and ns-IHA	In-house production: same as Zou 12 Vit pns-IHA	5.0×10^{-15}	2.0	Porous fibrous cylinder: $\phi = 5$ mm, $L = 15$ mm	Electrospun fibrous mats w in-situ grown ceramic	In vivo: subcutaneous implantation in male dogs	Zou 12 Viv pns-IHA [56]	
Poly(DL-lactide)(50:50)	In-house production: same as Zou 12 Vit pns-IHA	2.5×10^{-10} (2.0×10^{-10})	5.2×10^{-10} (2.0×10^{-10})	Mixture of pHA and ns-IHA	In-house production: same as Zou 12 Vit pns-IHA	1.1×10^{-14}	2.0	Porous fibrous cylinder: $\phi = 5$ mm, $L = 15$ mm	Re-electrospun after dissolving above-mentioned mat	In vivo: same as Zou 2 Viv pns-IHA	Zou 12 Viv R pns-IHA [56]	
Poly(DL-lactide)(50:50) modified with carboxyl groups	In-house production: ring-opening polymerisation of DL-lactide + modification with carboxyl groups	4.9×10^{-10}	2.3×10^{-10}	Mixture of pHA and ns-IHA	In-house production: grown in-situ using concentrated SBF	3.0×10^{-13}	2.0	Porous fibrous mat: $t = 0.5$ mm	Electrospun mats with in-situ grown ceramic	In vitro: PBS w sodium azide at 37 °C, w shaking at 100 rpm	Zou 11 C pns-IHA [57]	

(continued)

Table B.6 (continued)

Polymer characteristics			Ceramic characteristics				Sample characteristics			Case code and reference	
Type	Manufacturer	k_1 (s^{-1})	k'_2 ($m^3 mol^{-1} s^{-1}$)	Type	Manufacturer	A_d ($mol m^{-2} s^{-1}$)	θ (1)	Structure and Morphology	Fabrication method		Degradation protocol
Poly(DL-lactide)/(50:50) modified with carboxyl groups	In-house production: same as Zou 11 C pns-IHA	2.3×10^{-10}	2.9×10^{-10}	Mixture of pHA and ns-IHA	In-house production: same as Zou 11 C pns-IHA	3.0×10^{-13}	2.0	Porous fibrous mat: $t = 0.5$ mm	Re-electrospun after dissolving above-mentioned mat	In vitro: same as Zou 11 C	Zou 11 CR pns-IHA [57]
Poly(DL-lactide)/(50:50) modified with hydroxyl and carboxyl groups	In-house production: ring-opening polymerisation of DL-lactide + modification with hydroxyl and carboxyl groups	2.8×10^{-10} (2.2×10^{-10})	2.5×10^{-10} (2.1×10^{-10})	Mixture of pHA and ns-IHA	In-house production: same as Zou 11 C pns-IHA	3.0×10^{-14}	2.0	Porous fibrous mat: $t = 0.5$ mm	Electrospun mats with in-situ grown ceramic	In vitro: same as Zou 11 C pns-IHA	Zou 11 HC [57]
Poly(DL-lactide)/(50:50) modified with amino, carboxyl and hydroxyl groups	In-house production: ring-opening polymerisation of DL-lactide + modification with amino, hydroxyl and carboxyl groups	3.5×10^{-10} (2.0×10^{-10})	2.9×10^{-10} (1.8×10^{-10})	Mixture of pHA and ns-IHA	In-house production: same as Zou 11 C pns-IHA	1.1×10^{-14}	2.0	Porous fibrous mat: $t = 0.5$ mm	Electrospun mats with in-situ grown ceramic	In vitro: same as Zou 11 C pns-IHA	Zou 11 AHC pns-IHA [57]
Poly(DL-lactide)(NR)	Jinan Daigang Bio-Technology Co. Ltd., China	6.0×10^{-11}	6.2×10^{-11}	bnHA	Berkeley Advanced Biomaterials Inc, USA	7.0×10^{-15}	2.0	Porous (80 %) cylinder: $\Phi = 10$ mm, $L = 5$ mm	Polymer dissolved in chloroform + ceramic and NaCl addition and dispersion + compression at 10 MPa and 25 °C	In vitro: SBF (pH = 7.4) at 37 °C, weekly replacement, w/o shaking	Chen PPC bnHA [58]
Poly(DL-lactide)(NR)	Jinan Daigang Bio-Technology Co. Ltd., China	8.0×10^{-11} (6.0×10^{-11})	9.0×10^{-11} (6.2×10^{-11})	bnHA	Berkeley Advanced Biomaterials Inc, USA	3.0×10^{-15}	2.0	Porous (80 %) cylinder: $\Phi = 10$ mm, $L = 5$ mm	Same as Chen PPC	In vitro: same as Chen PPC	Chen C bnHA [58]

B.2.3 Poly-lactide-co-glycolide Matrix

Table B.7 Results output by the HA composites degradation model for cases with a poly(lactide-co-glycolide) matrix. k_1 is the non-catalytic polymer degradation rate, k_2^* is the autocatalytic polymer degradation rate, A_d is the ceramic dissolution rate and θ is the power ceramic dissolution law exponent. Abbreviations used to specify the hydroxyapatite type can be found in Table 5.2. A dashed line between rows indicates that data displayed in those consecutive rows belong to the same researcher or research group. A solid line between rows indicates no known author relationship for the data

Polymer characteristics				Ceramic characteristics				Sample characteristics				Degradation protocol	Case code and reference
Type	Manufacturer	k_1 (s^{-1})	k_2^* ($m^3 mol^{-1} s^{-1}$)	Type	Manufacturer	A_d ($mol m^{-2} s^{-1}$)	θ (1)	Structure and Morphology	Fabrication method				
Poly(D,L-lactide-co-glycolide) (44:6:55:4)	Mitsui Chemicals Co.; Tokyo, Japan	2.5×10^{-9}	3.8×10^{-9}	hHA	Mitsui Calcium Co.; Akashi, Japan	2.0×10^{-12}	2.0	Dense disk: $\Phi = 5$ mm, $t = 0.5$ mm	Polymer dissolved in dioxane + ceramic addition + freeze-drying of the solvent cast solution + compression moulding at $60^\circ C$ and 4.9 MPa	In vitro: PBS (0.01 M, pH = 7.4) at $37^\circ C$, weekly replacement, w/o shaking, 30 mL/sample	Tsumoda hHA [59]		
Poly(D,L-lactide-co-glycolide) (44:6:55:4)	Shang Dong Institute (Dept. of Med. Dev.), China	3.2×10^{-10} (2.8×10^{-10})	5.2×10^{-10} (3.2×10^{-10})	hHA	In-house production: wet chemistry route + calcining at $1100^\circ C$ for 6 h	1.1×10^{-11}	2.0	Porous ($\approx 90\%$) disk: $\Phi = 6$ mm, $t = 2$ mm	Polymer-ceramic mixture solvent casting + particulate leaching using NaCl	In vitro: standard conditions (PBS (pH = 7.4) at $37^\circ C$ w/o shaking) 20 mL/sample	Li hHA [60]		
Poly(D,L-lactide-co-glycolide) (47:53)	Purate Biomaterials BV, The Netherlands (Purasorb PDLG 5004)	1.7×10^{-9}	1.5×10^{-9}	bnHA	Aldrich Chemistry, USA	3.0×10^{-13}	2.0	Porous ($\approx 90\%$) film: $0.5 cm^2 \times t$ (t: NR)	Polymer dissolved in dioxane + blended w ceramic + freeze drying	In vitro: standard conditions (PBS at $37^\circ C$ w/o shaking) 20 mL/sample	Diaz PLGA bnHA [61]		
Poly(D,L-lactide-co-glycolide) (50:50)	Birmingham Polymers Inc.; Birmingham, Alabama, USA	4.0×10^{-10}	6.0×10^{-10}	hHA	Sigma Chemicals; St. Louis, Missouri, USA	7.0×10^{-12}	2.0	Dense cylinder: $\Phi = 5$ mm, $L = 5$ mm	Polymer dissolved in acetone + precipitated in ethanol + mixed w ceramic + compression moulding into final shape	In vitro: distilled water at $37^\circ C$ w/o shaking, w/o replacement, 10 mL/sample	Agrawal hHA [62]		

(continued)

Table B.7 (continued)

Polymer characteristics			Ceramic characteristics			Sample characteristics			Degradation protocol	Case code and reference	
Type	Manufacturer	k_1 (s^{-1})	k'_2 ($m^3 mol^{-1} s^{-1}$)	Type	Manufacturer	Ad ($mol m^{-2} s^{-1}$)	θ (1)	Structure and Morphology			Fabrication method
Poly(DL-lactide-co-glycolide) (50:50)	SunModics Pharmaceuticals, USA	1.1×10^{-9}	2.0×10^{-9}	pCHA	In-house production: wet chemistry route using $Ca(OH)_2$ and H_3PO_4 + ground, milled and sieved using a 180 μm mesh	1.0×10^{-12}	2.0	Dense disk: $\Phi = 2$ mm, $t = 2$ mm	Polymer and ceramic mixed in acetone + dried at 40 °C overnight + injection-moulded into cylinders at 140 °C and 0.5 MPa + cut into final shape	In vitro: PBS (0.01 M, pH = 7.4) at 37 °C, w/o shaking, w/o replacement, 166mL g ⁻¹	Naik pCHA [63]
Poly(DL-lactide-co-glycolide) (50:50)	SunModics Pharmaceuticals, USA	1.1×10^{-9}	2.0×10^{-9}	hCHA	In-house production: same as Naik pCHA + calcined at 800 °C for 4 h in air	5.0×10^{-12}	2.0	Dense disk: $\Phi = 2$ mm, $t = 2$ mm	Same as Naik pCHA	In vitro: same as Naik pCHA	Naik ca hCHA [63]
Poly(DL-lactide-co-glycolide) (50:50)	SunModics Pharmaceuticals, USA	1.1×10^{-9}	2.0×10^{-9}	hCHA	In-house production: same as Naik pCHA + calcined at 800 °C for 4 h in a wet argon atmosphere	3.0×10^{-12}	2.0	Dense disk: $\Phi = 2$ mm, $t = 2$ mm	Same as Naik pCHA	In vitro: same as Naik pCHA	Naik cw hCHA [63]

(continued)

Table B.7 (continued)

Polymer characteristics			Ceramic characteristics			Sample characteristics			Degradation protocol	Case code and reference	
Type	Manufacturer	k_1 (s^{-1})	k'_2 ($m^3 mol^{-1} s^{-1}$)	Type	Manufacturer	A_d ($mol m^{-2} s^{-1}$)	θ (1)	Structure and Morphology			Fabrication method
Poly(D,L-lactide-co-glycolide) (50:50)	SurModics Pharmaceuticals, USA	1.1×10^{-9}	2.0×10^{-9}	sMP-pCHA	In-house production: same as Naik pCHA + treatment w physisorbed 1 wt % mercaptopropyl trimethoxy silane (MPTMS) for 1 h	3.0×10^{-11}	2.0	Dense disk: $\Phi = 2$ mm, $t = 2$ mm	Same as Naik pCHA	In vitro: same as Naik pCHA	Naik f-sMP-pCHA [64]
Poly(D,L-lactide-co-glycolide) (50:50)	SurModics Pharmaceuticals, USA	1.1×10^{-9}	2.0×10^{-9}	sMP-pCHA	In-house production: same as Naik pCHA + treatment w chemisorbed 1 wt % mercaptopropyl trimethoxy silane (MPTMS) for 1 h	3.0×10^{-11}	2.0	Dense disk: $\Phi = 2$ mm, $t = 2$ mm	Same as Naik pCHA	In vitro: same as Naik pCHA	Naik e-sMP-pCHA [64]
Poly(D,L-lactide-co-glycolide) (50:50)	SurModics Pharmaceuticals, USA	1.1×10^{-9}	2.0×10^{-9}	sMP-hCHA	In-house production: same as Naik ca-hCHA + treatment w physisorbed 1 wt % mercaptopropyl trimethoxy silane (MPTMS) for 1 h	1.0×10^{-11}	2.0	Dense disk: $\Phi = 2$ mm, $t = 2$ mm	Same as Naik pCHA	In vitro: same as Naik pCHA	Naik fca-sMP-hCHA [64]
Poly(D,L-lactide-co-glycolide) (50:50)	SurModics Pharmaceuticals, USA	1.1×10^{-9}	2.0×10^{-9}	sMP-hCHA	In-house production: same as Naik ca-hCHA + treatment w chemisorbed 1 wt % mercaptopropyl trimethoxy silane (MPTMS) for 1 h	1.0×10^{-11}	2.0	Dense disk: $\Phi = 2$ mm, $t = 2$ mm	Same as Naik pCHA	In vitro: same as Naik pCHA	Naik eca-sMP-hCHA [64]

(continued)

Table B.7 (continued)

Polymer characteristics			Ceramic characteristics				Sample characteristics				Degradation protocol	Case code and reference
Type	Manufacturer	k_1 (s^{-1})	k'_2 ($m^3 mol^{-1} s^{-1}$)	Type	Manufacturer	A_d ($mol m^{-2} s^{-1}$)	θ (1)	Structure and Morphology	Fabrication method			
Poly(D,L-lactide-co-glycolide) (50:50)	SunModics Pharmaceuticals, USA	1.0×10^{-9}	1.8×10^{-9}	hCHA	In-house production: wet chemistry route using $Ca(OH)_2$ and H_3PO_4 + aged for 1 d and dried + calcined at $800^\circ C$ for 2 h + ground and sieved using a $75 \mu m$ mesh	3.0×10^{-14}	2.0	Dense disk: $\Phi = 6$ mm, $t = 2$ mm	Ceramic dispersed in polymer using a co-rotating twin screw extruder + cut into pellets + injection-moulded into cylinders at $130^\circ C$ and 0.5 MPa + cut into final shape	In vitro: PBS (0.01 M, pH = 7.4) at $37^\circ C$, w/o shaking, w/o replacement	Ege hCHA [27]	
Poly(D,L-lactide-co-glycolide) (50:50)	SunModics Pharmaceuticals, USA	1.0×10^{-9}	1.8×10^{-9}	hHA	In-house production: wet chemistry route using $Ca(OH)_2$ and H_3PO_4 + aged for 1 d and dried + calcined at $1200^\circ C$ for 2 h + ground and sieved using a $75 \mu m$ mesh	1.0×10^{-13}	2.0	Dense disk: $\Phi = 6$ mm, $t = 2$ mm	Same as Ege hCHA	In vitro: same as Ege hCHA	Ege hHA [26][27]	
Poly(D,L-lactide-co-glycolide) (75:25)	Boehringer Ingelheim, Germany (Resomer RG750S)	3.4×10^{-10}	2.4×10^{-10}	hHA	Sigma Aldrich, St. Louis, Missouri, USA	1.0×10^{-13}	2.0	Porous fibrous mat: $20 \times 5 mm^2 \times t$ (t: NR)	Polymer dissolved in HPFP + blended w ceramic + electrospinning	In vitro: standard conditions (PBS (0.01 M) at $37^\circ C$ w/o shaking) 5 mL/sample	Lee hHA [65]	
Poly(lactide-co-glycolide) (95:05)	In-house production	1.5×10^{-10} (2.8×10^{-11})	1.0×10^{-10} (1.9×10^{-11})	lHA	In-house production	1.0×10^{-14}	2.0	Dense bar: $60 \times 6 mm^2 \times 4$ mm	Solution mixing + pressed + annealed at $110^\circ C$ for 30 min	In vitro: SBF at $37^\circ C$ (buffered to pH = 7.4), w shaking, 20 mL/sample	Liuyun lHA [66]	
Poly(lactide-co-glycolide) (NR)	Not reported	6.6×10^{-10}	4.8×10^{-10}	hHA	In-house production: hydrothermal-electrochemical deposition using an electrolyte containing the precursors	3.0×10^{-13}	2.0	Dense film: $5 \times 5 mm^2 \times t$ (0.05...0.08) mm	Polymer solvent casting on a Ti substrate w. grown HA + delaminating	In vitro: SBF at $37^\circ C$	Ban hHA [67]	

B.2.4 Miscellaneous Polymers Matrixes

Table B.8 Results output by the HA composites degradation model for cases with a miscellaneous polymer matrix. k_1 is the non-catalytic polymer degradation rate, k'_2 is the autocatalytic polymer degradation rate, A_d is the ceramic dissolution rate and θ is the power ceramic dissolution law exponent. Abbreviations used to specify the hydroxyapatite type can be found in Table 5.2. A dashed line between rows indicates that data displayed in those consecutive rows belong to the same researcher or research group. A solid line between rows indicates no known author relationship for the data

Ceramic characteristics											
Sample characteristics											
Type	Manufacturer	k_1 (s^{-1})	k'_2 ($m^3 \text{ mol}^{-1} s^{-1}$)	Type	Manufacturer	A_d ($\text{mol m}^{-2} s^{-1}$)	θ (1)	Structure and Morphology	Fabrication method	Degradation protocol	Case code and reference
Poly(ϵ -caprolactone)	Daiichi Polymer Ltd.; Tokyo, Japan	6.0×10^{-12} (1.0×10^{-12})	2.3×10^{-11} (4.0×10^{-12})	bnHA	Nanjing Emperor Nano Material Co. Ltd., China	3.0×10^{-16}	2.0	Porous ($\approx 70\%$) film: $10 \times 10 \text{ mm}^2 \times 3 \text{ mm}$	Polymer, ceramic and porogen mixed at 100°C and 90 rpm for 15 min + compression moulding at 80°C + porogen leaching w distilled water	In vitro: PBS (pH = 7.4) at 37°C , monthly replacement, w shaking	Wang PCL bnHA[68]
Poly(L-co-DL-lactide)(70:30) and poly(L-lactide) for pure polymer sample	Boehringer Ingelheim, Germany (Resomer LR708)	4.4×10^{-11} (6.0×10^{-11})	4.4×10^{-11} (5.8×10^{-11})	hHA	CamCeram Coating Powder, The Leiden, The Netherlands	3.0×10^{-13}	2.0	Dense screw: $\Phi = (2.62...4.5) \text{ mm}$, $L = 30 \text{ mm}$	Polymer and ceramic mixed in a rotary blender + injection moulded into screw shape	In vitro: PBS (0.01 M, pH = 7.4) at 37°C , w shaking at 60 rpm, 50 $\text{mL} \cdot \text{g}^{-1}$	Hile hHA [69]
Poly(L-lactide-co- ϵ -caprolactone) (70:30)	Purac Biomaterials BV., The Netherlands (Purasorb PLC 7015)	1.5×10^{-10}	2.8×10^{-10}	bnHA	Aldrich Chemistry, USA	1.0×10^{-14}	2.0	Porous ($\approx 90\%$) film (dimensions: NR)	Polymer dissolved in dioxane + blended w ceramic + freeze drying	In vitro: standard conditions (PBS at 37°C w/o shaking) 10 mL/sample	Diaz PLCL bnHA [70]

(continued)

Table B.8 (continued)

Polymer characteristics			Ceramic characteristics			Sample characteristics			Fabrication method		Degradation protocol	Case code and reference
Type	Manufacturer	k_1 (s^{-1})	k'_2 ($m^3 mol^{-1} s^{-1}$)	Type	Manufacturer	A_d ($mol m^{-2} s^{-1}$)	θ (1)	Structure and Morphology	Fabrication method	Degradation protocol	Case code and reference	
Poly(D,L-lactide-co-ε-caprolactone) (60:40)	In-house production: ring-opening polymerisation at 145 °C in N ₂ atmosphere for 3 h	1.5×10^{-9}	1.2×10^{-9}	hHA	De Puy, Bioland, France (BRKO)	7.0×10^{-12}	2.0	Dense dogbone: DIN Standard 53448 F2. Smallest dimension < 5 mm	Polymer dissolved in methylene chloride + mixed w ceramic + compression moulded at (70, 90 °C and (2500...4500) kg	In vitro: Ringer solution (pH = (5.5...7.0) at 37 °C, w shaking	Ural NS hHA [71]	
Poly(D,L-lactide-co-ε-caprolactone) (60:40)	In-house production: ring-opening polymerisation at 145 °C under vacuum for 19 h	1.5×10^{-9}	1.2×10^{-9}	hHA	De Puy, Bioland, France (BRKO)	5.0×10^{-12}	2.0	Dense dogbone: DIN Standard 53448 F2. Smallest dimension < 5 mm	Same as Ural NS hHA	In vitro: same as Ural NS hHA	Ural VL hHA [71]	
Poly(L-lactide) and poly(ε-caprolactone) blend (28.4:71.6)	Cargill-Dow and Polysciences	8.0×10^{-12}	2.4×10^{-11}	bnHA	Sigma Aldrich	3.0×10^{-15}	2.0	Porous (≈ 90 % disk: $\Phi = 6$ mm, $t = 3.5$ mm)	Polymer dissolved in dioxane + ceramic dispersed in solvent + mixed + freeze-drying + porogen leaching	In vitro: Phosphate buffer (pH = 7.4) at 37 °C, replacement every 2 weeks, w shaking, 200 mL g^{-1}	Rodenas CL bnHA [72]	
Poly(L-lactide) and poly(ε-caprolactone) blend (86.4:13.6)	Cargill-Dow and Polysciences	2.6×10^{-11}	1.2×10^{-11}	bnHA	Sigma Aldrich	3.0×10^{-15}	2.0	Porous (≈ 90 % disk: $\Phi = 6$ mm, $t = 3.5$ mm)	Same as Rodenas CL bnHA	In vitro: same as Rodenas CL bnHA	Rodenas LA bnHA [72]	
Poly(D,L-lactide-co-glycolide)(50:50) and poly(ε-caprolactone) blend (84:16)	Purac Biomaterials BV; Gorinchem, The Netherlands (Purasorb PDLG 5010) and Sigma Aldrich; St. Louis, Missouri, USA	7.8×10^{-10}	9.4×10^{-10}	bnHA	Donated by Prof. Marc Bohner (Budenheim, Tri-Carfos P(C53-80)	1.0×10^{-14}	2.0	Porous mat (dimensions: NR)	Ceramic suspended in solvent + polymer added + electrospinning of the solution	In vitro: PBS (pH = 7.4) at 37 °C, w/o replacement, w shaking at 60 rpm 2 mL/sample	Ji bnHA [73]	

(continued)

Table B.8 (continued)

Polymer characteristics			Ceramic characteristics			Sample characteristics			Degradation protocol	Case code and reference	
Type	Manufacturer	k_1 (s^{-1})	k_2^* ($m^3 mol^{-1} s^{-1}$)	Type	Manufacturer	A_d ($mol m^{-2} s^{-1}$)	θ (1)	Structure and Morphology			Fabrication method
Poly(DL-lactide-co-glycolide)(65:35) and poly(ϵ -caprolactone) blend (71.8:28.2)	Aldrich and Aldrich	2.2×10^{-10}	1.8×10^{-10}	hHA	Aldrich	9.0×10^{-12}	2.0	Porous (80 %) film: $5 \times 5 mm^2 \times 1 mm$	Polymers dissolved in tetrahydrofuran + NaCl and ceramic addition + solvent evaporation + pressed at (41.4, 68.9) MPa	In vitro: sterile water at 37 °C, no replacement	Dunn CLs hHA [74]
Poly(DL-lactide-co-glycolide)(65:35) and poly(ϵ -caprolactone) blend (71.8:28.2)	Aldrich and Aldrich	2.2×10^{-10}	1.8×10^{-10}	hHA	Aldrich	3.0×10^{-11}	2.0	Porous (80 %) film: $5 \times 5 mm^2 \times 1 mm$	Same as Dunn CLs hHA	In vitro: sterile water at 37 °C, replacement every 2/3 days	Dunn CLd hHA [74]
Poly(DL-lactide-co-glycolide)(65:35) and poly(ϵ -caprolactone) blend (71.8:28.2)	Aldrich and Aldrich	3.0×10^{-10}	1.6×10^{-10}	hHA	Aldrich	3.0×10^{-12}	2.0	Porous (80 %) film: $5 \times 5 mm^2 \times 1 mm$	Same as Dunn CLs hHA	In vitro: sterile water at 37 °C, no replacement	Dunn LGs hHA [74]
Poly(DL-lactide-co-glycolide)(65:35) and poly(ϵ -caprolactone) blend (93.9:6.1)	Aldrich and Aldrich	3.0×10^{-10}	2.6×10^{-10}	hHA	Aldrich	5.0×10^{-12}	2.0	Porous (80 %) film: $5 \times 5 mm^2 \times 1 mm$	Same as Dunn CLs hHA	In vitro: sterile water at 37 °C, replacement every 2/3 days	Dunn LGd hHA [74]

B.3 Calcium Carbonate Case Studies Modelling Results

Table B.9 Results output by the CC composites degradation model for all analysed case studies. k_1 is the non-catalytic polymer degradation rate, k'_2 is the autocatalytic polymer degradation rate, A_d is the ceramic dissolution rate and θ is the power ceramic dissolution law exponent. Abbreviations used to specify the calcium carbonate type can be found in Table 7.2. A dashed line between rows indicates that data displayed in those consecutive rows belong to the same researcher or research group. A solid line between rows indicates no known author relationship for the data

Polymer characteristics				Ceramic characteristics				Sample characteristics				Degradation protocol	Case code and reference
Type	Manufacturer	k_1 (s^{-1})	k'_2 ($m^3 mol^{-1} s^{-1}$)	Type	Manufacturer	A_d ($mol m^{-2} s^{-1}$)	θ (1)	Structure and Morphology	Fabrication method				
Poly(L-lactide)	Purate Biochem B.V.; Gorinchem, The Netherlands (Purasorb)	1.1×10^{-10}	9.0×10^{-11}	μ -CC	In-house production: mixture of $Ca(OH)_2$ + aminopropyl-triethoxysilane (APTES) + methanol in a CO_2 atmosphere + dried at $110^\circ C$	3.0×10^{-13}	2.0	Porous mat	Polymer and ceramic kneaded at $200^\circ C$ for 10 min + mixture dissolved in chloroform + electrospinning of the polymer and ceramic mix	In vitro: 5 mmol/L NaOH solution, w/o shaking, w/o replacement	Wakita K μ -CC [75]		
Poly(L-lactide)	Purate Biochem B.V.; Gorinchem, The Netherlands (Purasorb)	1.4×10^{-11}	1.4×10^{-11}	μ -CC	In-house production: mixture of $Ca(OH)_2$ + aminopropyl-triethoxysilane (APTES) + methanol in a CO_2 atmosphere + dried at $110^\circ C$	3.0×10^{-13}	2.0	Porous mat	Ceramic dispersed in chloroform + polymer dissolved in the mixture + electrospinning of the polymer and ceramic mix	In vitro: same as Wakita K μ -CC	Wakita M μ -CC [75]		

(continued)

Table B.9 (continued)

Polymer characteristics			Ceramic characteristics				Sample characteristics			Fabrication method		Degradation protocol	Case code and reference
Type	Manufacturer	k_1 (s^{-1})	k_2' ($m^3 mol^{-1} s^{-1}$)	Type	Manufacturer	A_d ($mol m^{-2} s^{-1}$)	θ (1)	Structure and Morphology					
Poly(L-lactide)	Shandong Medical Device Company, China	5.0×10^{-11}	1.4×10^{-11}	λ -CC	In-house production: freshwater cultured luster pearls (<i>H. camingii</i>) ground into powder using a grinder	5.0×10^{-12}	2.0	Porous cylinder: $\Phi = 9$ mm, $L = 18$ mm	Polymer dissolved in 1,4-dioxane + ceramic addition + stirring and sonication + freezing at $-20^\circ C$ for 24 h + lyophilisation at $-60^\circ C$ for 48 h + drying in a vacuum oven for 72 h	In vitro: PBS (pH = 7.4) at $37^\circ C$, w weekly shaking at 60 rpm, 15 mL/sample	Liu λ -CC [76]		
Poly(L-lactide)	Shandong Medical Device Company, China	5.0×10^{-11}	1.4×10^{-11}	μ -CC	In-house production: freshwater cultured lustrous pearls (<i>H. camingii</i>) ground into powder using a grinder	3.0×10^{-12}	2.0	Porous cylinder: $\Phi = 9$ mm, $L = 18$ mm	Same as Liu λ -CC	In vitro: same as Liu λ -CC	Liu μ -CC [76]		
Poly(D,L-lactide) (50:50)	In-house production: bulk ring-opening polymerisation of D,L-lactide at $145^\circ C$ using zinc powder	1.4×10^{-10}	8.2×10^{-10}	λ -CC	Noteb; France (Biocoral)	3.0×10^{-10}	2.0	Dense film: $12 \times 12 mm^2$ (PDLLA) $\times 2.1 mm$ (PDLLA- λ -CC) $\times 1.4 mm$	Polymer dissolved in acetone + blended w ceramic + vacuum drying at $40^\circ C$ for 2 d + compression at 10 MPa and $130^\circ C$ + cut at $80^\circ C$ into final shape	In vitro: PBS (0.13M, pH = 7.4) w 0.02% NaNO ₃ at $37^\circ C$, w/6 shaking, w/6 replacement, 30 mL/sample	Liu λ -CC [77]		

(continued)

Table B.9 (continued)

Polymer characteristics		Ceramic characteristics				Sample characteristics				Fabrication method		Degradation protocol	Case code and reference
Type	Manufacturer	k_1 (s^{-1})	k_2' ($m^3 mol^{-1} s^{-1}$)	Type	Manufacturer	A_d ($mol m^{-2} s^{-1}$)	θ (1)	Structure and Morphology					
Poly(DL-lactide-co-glycolide) (44:6:55:4)	Mitsui Chemicals; Tokyo, Japan: polymerisation of D,L-lactide and glycolide using stannous octoate	3.0×10^{-9}	3.0×10^{-9}	β -CC (Assumed)	Wako Pure Chemical Industries; Tokyo, Japan	3.0×10^{-10}	2.0	Dense film: $15 \times 15 mm^2 \times 0.5 mm$	Polymer dissolved in dioxane + ceramic addition + freeze-drying of the solvent cast solution + compression moulding at $60^\circ C$ and 4.9 MPa	In vitro: PBS (pH = 7.4) at $37^\circ C$, w/o shaking, weekly replacement, 40 mL/sample	Ara β -CC [24]		
Poly(DL-lactide-co-glycolide) (44:6:55:4)	Mitsui Chemicals Co.; Tokio, Japan	2.5×10^{-9}	3.8×10^{-9}	β -CC	Maruo Calcium Co.; Akashi, Japan	3.0×10^{-11}	2.0	Dense disk: $\Phi = 5 mm$, $t = 0.5 mm$	Polymer dissolved in dioxane + ceramic addition + freeze-drying of the solvent cast solution + compression moulding at $60^\circ C$ and 4.9 MPa	In vitro: PBS (0.01 M, pH = 7.4) at $37^\circ C$, weekly replacement, w/o shaking, 30 mL/sample	Tsumoda β -CC [59]		
Poly(DL-lactide-co-glycolide) (44:6:55:4)	Mitsui Chemicals Co.; Tokio, Japan	2.5×10^{-9}	3.8×10^{-9}	λ -CC	Maruo Calcium Co.; Akashi, Japan	3.0×10^{-11}	2.0	Dense disk: $\Phi = 5 mm$, $t = 0.5 mm$	Same as Tsumoda β -CC	In vitro: same as Tsumoda β -CC	Tsumoda λ -CC [59]		
Poly(DL-lactide-co-glycolide) (50:50)	Birmingham Polymers Inc.; Birmingham, Alabama, USA	4.0×10^{-10}	6.0×10^{-10}	β -CC (Assumed)	Sigma Chemicals; St. Louis, Missouri, USA	3.0×10^{-11}	2.0	Dense cylinder: $\Phi = 5 mm$, $L = 5 mm$	Polymer dissolved in acetone + precipitated in ethanol + mixed w ceramic + compression moulding into final shape	In vitro: distilled water at $37^\circ C$ w/o shaking, w/o replacement, 10 mL/sample	Agrawal β -CC [62]		

(continued)

Table B.9 (continued)

Polymer characteristics			Ceramic characteristics			Sample characteristics			Degradation protocol	Case code and reference	
Type	Manufacturer	k_1 (s^{-1})	k_2' ($m^3 mol^{-1} s^{-1}$)	Type	Manufacturer	A_d ($mol m^{-2} s^{-1}$)	θ (1)	Structure and Morphology			Fabrication method
Poly(DL-lactide-co-glycolide) (50:50)	Lakeshore Biomaterials; Birmingham, Alabama, USA (5050DLG SE)	3.7×10^{-9}	3.2×10^{-9}	β -CC	Shiraishi Calcium Kaisha Ltd; Japan	3.0×10^{-12}	2.0	Dense disk: $\Phi = 3.5$ mm, $t = 2$ mm	Polymer dissolved in acetone + ceramic sonicated in acetone + mixing + solvent casting + cut into pellets + injection-moulding at 135 °C and 0.5 MPa into cylinders + cut into final shape	In vitro: PBS (0.01 M, pH = 7.4) at 37 °C, w/o shaking, w/o replacement, 166 mL g^{-1}	Ch6 β -CC [Chapter 6]
Poly(DL-lactide-co-glycolide) (85:15)	Purac Biochem B.V.; Gorinchem, The Netherlands	1.5×10^{-10}	8.0×10^{-10}	β -CC (Assumed)	Sigma; St. Louis, Missouri, USA (SigmaUltra Grade)	3.0×10^{-10}	2.0	Dense screw: $\Phi = 9$ mm, $L = 30$ mm	Polymer dried under vacuum for 3 h at 40 °C + blended w ceramic in a co-rotating twin extruder + injection moulded into screw shape	In vitro: PBS (0.05 M, pH = 7.4) at 37 °C, w/o shaking, w/o replacement, 250 mL/sample	Cotton β -CC [78]

References

1. Ito, A., Maekawa, K., Tsutsumi, S., Ikazaki, F., & Tateishi, T. (1997). Solubility product of OH-carbonated hydroxyapatite. *Journal of Biomedical Materials Research Part A*, 36(4), 522–528.
2. Atkins, P. (2010). *Shriver and Atkins' inorganic chemistry*. USA: Oxford University Press.
3. Kirkwood, T. B. (1979). Geometric means and measures of dispersion. *Biometrics*, 35, 908–909.
4. Imai, Y., Nagai, M., & Watanabe, M. (1999b). Degradation of composite materials composed of tricalcium phosphate and a new type of block polyester containing a poly(L-lactic acid) segment. *Journal of Biomaterials Science, Polymer Edition*, 10(4), 421–432.
5. Ebato, H., Oya, S., Kakizawa, Y., Furuta, H., and Arai, K. (1995). Process for producing lactic acid-based copolyester and packaging material. US Patent 5,403,897.
6. Speight, J. G., et al. (2005). *Lange's handbook of chemistry* (Vol. 1). New York: McGraw-Hill.
7. Scientific Polymer, Inc. (2013). *Polymer density data*. Retrieved September 1, 2016 from <http://scientificpolymer.com/density-of-polymers-by-density/>.
8. Kobayashi, S., & Yamaji, S. (2014). Analytical prediction of hydrolysis behavior of tricalcium phosphate/poly-L-lactic acid composites in simulated body environment. *Advanced Composite Materials*, 23(3), 211–223.
9. Aunoble, S., Clément, D., Frayssinet, P., Harmand, M. F., & Le Huec, J. C. (2006). Biological performance of a new β -TCP/PLLA composite material for applications in spine surgery: in vitro and in vivo studies. *Journal of Biomedical Materials Research Part A*, 78(2), 416–422.
10. Adamus, A., Jozwiakowska, J., Wach, R., Suarez-Sandoval, D., Ruffieux, K., & Rosiak, J. (2012). In vitro degradation of β -tricalcium phosphate reinforced poly (L-lactic acid). In *Materials Science Forum* (Vol. 714, pp. 283–290). Trans Tech Publication
11. Kang, Y., Xu, X., Yin, G., Chen, A., Liao, L., Yao, Y., et al. (2007). A comparative study of the in vitro degradation of poly(L-lactic acid)/ β -tricalcium phosphate scaffold in static and dynamic simulated body fluid. *European Polymer Journal*, 43(5), 1768–1778.
12. Kang, Y., Yao, Y., Yin, G., Huang, Z., Liao, X., Xu, X., et al. (2009). A study on the in vitro degradation properties of poly(L-lactic acid)/ β -tricalcium phosphate (PLLA/ β -TCP) scaffold under dynamic loading. *Medical Engineering & Physics*, 31(5), 589–594.
13. Daculsi, G., Goyenvalle, E., Cognet, R., Aguado, E., & Suokas, E. O. (2011). Osteoconductive properties of poly(96L/4D-lactide)/beta-tricalcium phosphate in long term animal model. *Biomaterials*, 32(12), 3166–3177.
14. Niemelä, T. (2005). Effect of β -tricalcium phosphate addition on the in vitro degradation of self-reinforced poly-L,D-lactide. *Polymer Degradation and Stability*, 89(3), 492–500.

15. Zheng, X., Zhou, S., Yu, X., Li, X., Feng, B., Qu, S., et al. (2008). Effect of in vitro degradation of poly(D, L-lactide)/ β -tricalcium composite on its shape-memory properties. *Journal of Biomedical Materials Research Part B: Applied Biomaterials*, 86(1), 170–180.
16. Lin, F.-H., Chen, T.-M., Lin, C.-P., & Lee, C.-J. (1999). The merit of sintered PDLLA/TCP composites in management of bone fracture internal fixation. *Artificial Organs*, 23(2), 186–194.
17. Heidemann, W., Jeschkeit, S., Ruffieux, K., Fischer, J. H., Wagner, M., Krüger, G., et al. (2001). Degradation of poly(D, L)lactide implants with or without addition of calcium phosphates in vivo. *Biomaterials*, 22(17), 2371–2381.
18. Niemelä, T., Kellomäki, M., & Törmälä, P. (2004). In vitro degradation of osteoconductive poly-L/DL-lactide/ β -TCP composites. In *Key Engineering Materials* (Vol. 254, pp. 509–512). Trans Tech Publication
19. Ignatius, A. A., Augat, P., & Claes, L. E. (2001 a). Degradation behavior of composite pins made of tricalcium phosphate and poly(L, DL-lactide). *Journal of Biomaterials Science, Polymer Edition*, 12(2), 185–194.
20. Haaparanta, A.-M., Haimi, S., Ellä, V., Hopper, N., Miettinen, S., Suuronen, R., et al. (2010). Porous polylactide/ β -tricalcium phosphate composite scaffolds for tissue engineering applications. *Journal of Tissue Engineering and Regenerative Medicine*, 4(5), 366–373.
21. Yang, F., Cui, W., Xiong, Z., Liu, L., Bei, J., & Wang, S. (2006). Poly(L, L-lactide-co-glycolide)/tricalcium phosphate composite scaffold and its various changes during degradation in vitro. *Polymer Degradation and Stability*, 91(12), 3065–3073.
22. Yang, Y., Zhao, Y., Tang, G., Li, H., Yuan, X., & Fan, Y. (2008). In vitro degradation of porous poly(L-lactide-co-glycolide)/ β -tricalcium phosphate (PLGA/ β -TCP) scaffolds under dynamic and static conditions. *Polymer Degradation and Stability*, 93(10), 1838–1845.
23. Jin, H.-H., Min, S.-H., Song, Y.-K., Park, H.-C., & Yoon, S.-Y. (2010). Degradation behavior of poly(lactide-co-glycolide)/ β -TCP composites prepared using microwave energy. *Polymer Degradation and Stability*, 95(9), 1856–1861.
24. Ara, M., Watanabe, M., & Imai, Y. (2002). Effect of blending calcium compounds on hydrolytic degradation of poly(DL-lactic acid-co-glycolic acid). *Biomaterials*, 23(12), 2479–2483.
25. Bennett, S. M. (2012). *Degradation mechanisms of PLGA/ α -TCP composites for orthopaedic applications*. Ph.D. thesis, Department of Materials Science and Metallurgy, University of Cambridge.
26. Ege, D., Best, S., & Cameron, R. (2014). The degradation behaviour of nanoscale HA/PLGA and α -TCP/PLGA composites. *Bioinspired, Biomimetic and Nanobiomaterials*, 3, BBN2.
27. Ege, D. (2012). *Mechanical and degradation properties of calcium phosphate/biodegradable polymer composites*. Ph.D. thesis, Department of Materials Science and Metallurgy, University of Cambridge.
28. Yang, Z., Best, S. M., & Cameron, R. E. (2009). The influence of α -tricalcium phosphate nanoparticles and microparticles on the degradation of poly(D, L-lactide-co-glycolide). *Advanced Materials*, 21(38–39), 3900–3904.
29. Yang, Z. (2009). *Development and characterisation of bioactive, bioresorbable α -tricalcium phosphate/poly(D,L-lactide-co-glycolide) nanocomposites for bone substitution and fixation*. Ph.D. thesis, Department of Materials Science and Metallurgy, University of Cambridge.
30. Mellon, V. (2003). *Degradation study of poly(lactide-co-glycolic) acid and alpha or beta tricalcium phosphate composites - non porous*. Unpublished research at University of Cambridge.
31. Barrett, C. E., & Cameron, R. E. (2014). X-ray microtomographic analysis of α -tricalcium phosphate-poly(lactic-co-glycolic) acid nanocomposite degradation. *Polymer*, 55(16), 4041–4049.
32. Barrett, C. E. (2013). *The degradation behaviour of tricalcium phosphate - poly(lactide-co-glycolide) nanocomposites*. Ph.D. thesis, Department of Materials Science and Metallurgy, University of Cambridge.
33. Ehrenfried, L. M., Patel, M. H., & Cameron, R. E. (2008). The effect of tri-calcium phosphate (TCP) addition on the degradation of polylactide-co-glycolide (PLGA). *Journal of Materials Science: Materials in Medicine*, 19(1), 459–466.

34. Ahola, N., Männistö, N., Veiranto, M., Karp, M., Rich, J., Efimov, A., et al. (2013). An in vitro study of composites of poly(L-lactide-co- ϵ -caprolactone), β -tricalcium phosphate and ciprofloxacin intended for local treatment of osteomyelitis. *Biomatter*, 3(2), e23162.
35. Ahola, N., Veiranto, M., Rich, J., Efimov, A., Hannula, M., Seppälä, J., et al. (2012). Hydrolytic degradation of composites of poly(L-lactide-co- ϵ -caprolactone)70/30 and β -tricalcium phosphate. *Journal of Biomaterials Applications*, 28(4), 529–543.
36. Kikuchi, M., Koyama, Y., Yamada, T., Imamura, Y., Okada, T., Shirahama, N., et al. (2004). Development of guided bone regeneration membrane composed of β -tricalcium phosphate and poly(L-lactide-co-glycolide-co- ϵ -caprolactone) composites. *Biomaterials*, 25(28), 5979–5986.
37. Imai, Y., Fukuzawa, A., & Watanabe, M. (1999a). Effect of blending tricalcium phosphate on hydrolytic degradation of a block polyester containing poly(L-lactic acid) segment. *Journal of Biomaterials Science, Polymer Edition*, 10(7), 773–786.
38. Kikuchi, M., Koyama, Y., Takakuda, K., Miyairi, H., Shirahama, N., & Tanaka, J. (2002). In vitro change in mechanical strength of β -tricalcium phosphate/copolymerized poly-L-lactide composites and their application for guided bone regeneration. *Journal of Biomedical Materials Research*, 62(2), 265–272.
39. Huang, J., Xiong, J., Liu, J., Zhu, W., & Wang, D. (2013). Investigation of the in vitro degradation of a novel polylactide/nanohydroxyapatite composite for artificial bone. *Journal of Nanomaterials*, 2013, 3.
40. Wang, Z., Wang, Y., Ito, Y., Zhang, P., & Chen, X. (2016). A comparative study on the in vivo degradation of poly(L-lactide) based composite implants for bone fracture fixation. *Scientific Reports*, 6.
41. Zhou, H., Touny, A. H., & Bhaduri, S. B. (2011). Fabrication of novel PLA/CDHA bionanocomposite fibers for tissue engineering applications via electrospinning. *Journal of Materials Science: Materials in Medicine*, 22(5), 1183–1193.
42. Verheyen, C., De Wijn, J., Van Blitterswijk, C., & De Groot, K. (1992). Evaluation of hydroxylapatite/poly(L-lactide) composites: Mechanical behavior. *Journal of Biomedical Materials Research*, 26(10), 1277–1296.
43. Verheyen, C., Klein, C., de Bleeck-Hogervorst, J., Wolke, J., Van Blitterswijk, C., & De Groot, K. (1993). Evaluation of hydroxylapatite/poly(L-lactide) composites: Physico-chemical properties. *Journal of Materials Science: Materials in Medicine*, 4(1), 58–65.
44. Díaz, E., Sandomis, I., Puerto, I., & Ibáñez, I. (2014). In vitro degradation of PLLA/nHA composite scaffolds. *Polymer Engineering & Science*, 54(11), 2571–2578.
45. Shikinami, Y., & Okuno, M. (1999). Bioresorbable devices made of forged composites of hydroxyapatite (HA) particles and poly-L-lactide (PLLA): Part I Basic characteristics. *Biomaterials*, 20(9), 859–877.
46. Furukawa, T., Matsusue, Y., Yasunaga, T., Shikinami, Y., Okuno, M., & Nakamura, T. (2000). Biodegradation behavior of ultra-high-strength hydroxyapatite/poly(L-lactide) composite rods for internal fixation of bone fractures. *Biomaterials*, 21(9), 889–898.
47. Ishii, S., Tamura, J., Furukawa, T., Nakamura, T., Matsusue, Y., Shikinami, Y., et al. (2003). Long-term study of high-strength hydroxyapatite/poly(L-lactide) composite rods for the internal fixation of bone fractures: A 2–4-year follow-up study in rabbits. *Journal of Biomedical Materials Research Part B: Applied Biomaterials*, 66(2), 539–547.
48. Shikinami, Y., & Okuno, M. (2001). Bioresorbable devices made of forged composites of hydroxyapatite (HA) particles and poly L-lactide (PLLA). Part II: Practical properties of miniscrews and miniplates. *Biomaterials*, 22(23), 3197–3211.
49. Deng, X., Sui, G., Zhao, M., Chen, G., & Yang, X. (2007). Poly(L-lactic acid)/hydroxyapatite hybrid nanofibrous scaffolds prepared by electrospinning. *Journal of Biomaterials Science, Polymer Edition*, 18(1), 117–130.
50. Sui, G., Yang, X., Mei, F., Hu, X., Chen, G., Deng, X., et al. (2007). Poly-L-lactic acid/hydroxyapatite hybrid membrane for bone tissue regeneration. *Journal of Biomedical Materials Research Part A*, 82(2), 445–454.

51. Xu, X., Chen, X., Liu, A., Hong, Z., & Jing, X. (2007). Electrospun poly(L-lactide)-grafted hydroxyapatite/poly(L-lactide) nanocomposite fibers. *European Polymer Journal*, 43(8), 3187–3196.
52. Rakmae, S., Lorprayoon, C., Ekgasit, S., & Suppakarn, N. (2013). Influence of heat-treated bovine bone-derived hydroxyapatite on physical properties and in vitro degradation behavior of poly(lactic acid) composites. *Polymer-Plastics Technology and Engineering*, 52(10), 1043–1053.
53. Rakmae, S., Ruksakulpiwat, Y., Sutapun, W., & Suppakarn, N. (2012). Effect of silane coupling agent treated bovine bone based carbonated hydroxyapatite on in vitro degradation behavior and bioactivity of PLA composites. *Materials Science and Engineering: C*, 32(6), 1428–1436.
54. Van der Meer, S., De Wijn, J., & Wolke, J. (1996). The influence of basic filler materials on the degradation of amorphous D- and L-lactide copolymer. *Journal of Materials Science: Materials in Medicine*, 7(6), 359–361.
55. Hasegawa, S., Tamura, J., Neo, M., Goto, K., Shikinami, Y., Saito, M., et al. (2005). In vivo evaluation of a porous hydroxyapatite/poly-DL-lactide composite for use as a bone substitute. *Journal of Biomedical Materials Research Part A*, 75(3), 567–579.
56. Zou, B., Chen, X., Zhi, W., Liu, Y., Cui, W., Hu, S., et al. (2012). Promoted healing of femoral defects with in situ grown fibrous composites of hydroxyapatite and poly(DL-lactide). *Journal of Biomedical Materials Research Part A*, 100(6), 1407–1418.
57. Zou, B., Li, X., Zhuang, H., Cui, W., Zou, J., & Chen, J. (2011). Degradation behaviors of electrospun fibrous composites of hydroxyapatite and chemically modified poly(DL-lactide). *Polymer Degradation and Stability*, 96(1), 114–122.
58. Chen, L., Tang, C. Y., Tsui, C. P., et al. (2013). Mechanical properties and in vitro evaluation of bioactivity and degradation of dexamethasone-releasing poly-D-L-lactide/nano-hydroxyapatite composite scaffolds. *Journal of the Mechanical Behavior of Biomedical Materials*, 22, 41–50.
59. Tsunoda, M. (2003). Degradation of poly(DL-lactic acid-co-glycolic acid) containing calcium carbonate and hydroxyapatite fillers-effect of size and shape of the fillers. *Dental Materials Journal*, 22(3), 371–382.
60. Li, H., & Chang, J. (2005). pH-compensation effect of bioactive inorganic fillers on the degradation of PLGA. *Composites Science and Technology*, 65(14), 2226–2232.
61. Díaz, E., Puerto, I., & Sandomis, I. (2015). The effects of bioactive nanoparticles on the degradation of DLGA. *International Journal of Polymeric Materials and Polymeric Biomaterials*, 64(1), 38–46.
62. Agrawal, C. M., & Athanasiou, K. A. (1997). Technique to control pH in vicinity of biodegrading PLA-PGA implants. *Journal of Biomedical Materials Research*, 38(2), 105–114.
63. Naik, A., Shepherd, D. V., Shepherd, J. H., Best, S. M., & Cameron, R. E. (2017). The effect of the type of HA on the degradation of PLGA/HA composites. *Materials Science and Engineering: C*, 70, 824–831.
64. Naik, A., Best, S. M., & Cameron, R. E. (2015). The influence of silanisation on the mechanical and degradation behaviour of PLGA/HA composites. *Materials Science and Engineering: C*, 48, 642–650.
65. Lee, J. B., Kim, S. E., Heo, D. N., Kwon, I. K., & Choi, B.-J. (2010). In vitro characterization of nanofibrous PLGA/gelatin/hydroxyapatite composite for bone tissue engineering. *Macromolecular Research*, 18(12), 1195–1202.
66. Liuyun, J., Chengdong, X., Lixin, J., & Lijuan, X. (2013). Degradation behavior of hydroxyapatite/poly(lactic-co-glycolic) acid nanocomposite in simulated body fluid. *Materials Research Bulletin*, 48(10), 4186–4190.
67. Ban, S., Watanabe, T., Itoh, T., Nakamura, H., Tsuruta, S., & Kawai, T. (2004). Development of biodegradable composite membrane containing oriented needle-like apatites. *Journal of Oral Tissue Engineering*, 2(1), 1–13.
68. Wang, Y., Liu, L., & Guo, S. (2010b). Characterization of biodegradable and cytocompatible nano-hydroxyapatite/polycaprolactone porous scaffolds in degradation in vitro. *Polymer Degradation and Stability*, 95(2), 207–213.

69. Hile, D. D., Doherty, S. A., & Trantolo, D. J. (2004). Prediction of resorption rates for composite polylactide/hydroxylapatite internal fixation devices based on initial degradation profiles. *Journal of Biomedical Materials Research Part B: Applied Biomaterials*, 71(1), 201–205.
70. Díaz, E., & Puerto, I. (2015). In vitro degradation of PLCL/nHA biodegradable scaffolds. *Polymer-Plastics Technology and Engineering*, 54(6), 556–564.
71. Ural, E., Kesenci, K., Fambri, L., Migliaresi, C., & Piskin, E. (2000). Poly(D, L-lactide/ ϵ -caprolactone)/hydroxyapatite composites. *Biomaterials*, 21(21), 2147–2154.
72. Rodenas-Rochina, J., Vidaurre, A., Cortázar, I. C., & Lebourg, M. (2015). Effects of hydroxyapatite filler on long-term hydrolytic degradation of PLLA/PCL porous scaffolds. *Polymer Degradation and Stability*, 119, 121–131.
73. Ji, W., Yang, F., Seyednejad, H., Chen, Z., Hennink, W. E., Anderson, J. M., et al. (2012). Biocompatibility and degradation characteristics of PLGA-based electrospun nanofibrous scaffolds with nanoapatite incorporation. *Biomaterials*, 33(28), 6604–6614.
74. Dunn, A. S., Campbell, P. G., & Marra, K. G. (2001). The influence of polymer blend composition on the degradation of polymer/hydroxyapatite biomaterials. *Journal of Materials Science: Materials in Medicine*, 12(8), 673–677.
75. Wakita, T., Nakamura, J., Ota, Y., Obata, A., Kasuga, T., & Ban, S. (2011). Effect of preparation route on the degradation behavior and ion releasability of siloxane-poly(lactic acid)-vaterite hybrid nonwoven fabrics for guided bone regeneration. *Dental Materials Journal*, 30(2), 232–238.
76. Liu, Y., Huang, Q., Kienzle, A., Müller, W., & Feng, Q. (2014). In vitro degradation of porous PLLA/pearl powder composite scaffolds. *Materials Science and Engineering: C*, 38, 227–234.
77. Li, S., & Vert, M. (1996). Hydrolytic degradation of coral/poly(DL-lactic acid) bioresorbable material. *Journal of Biomaterials Science, Polymer Edition*, 7(9), 817–827.
78. Cotton, N. J., Egan, M. J., & Brunelle, J. E. (2008). Composites of poly(DL-lactide-co-glycolide) and calcium carbonate: In vitro evaluation for use in orthopedic applications. *Journal of Biomedical Materials Research Part A*, 85(1), 195–205.