

Research Article

Emissions of Volatile Organic Compounds from Dairy Cattle Manure in a Cattle Shed in Japan

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ABSTRACT The livestock industry is a major source of atmospheric volatile organic compounds (VOCs), but details on these emissions are not well documented in Japan. In particular, it remains unclear how the rearing method affects the emissions of VOCs from livestock, which originate primarily from feces and urine. Here we aimed to estimate the amounts of VOCs emitted from the feces and urine of tethered Holstein dairy cattle in a cattle shed in Japan. Dimethyl sulfide and acetone accounted for about 60% of the total VOCs emitted from feces, followed by formaldehyde and acetaldehyde. Also, dimethyl sulfide and acetone were the dominant VOCs emitted from urine, accounting for 90% of the total VOCs. The VOCs from manure were considered to be emitted between the excretion and removal of the manure during the cleaning of the shed. As a result of analyzing images from three cameras installed in the shed, the average time between excretion and cleaning during the daytime (8:00 am–5:00 pm) was 80 min for feces and urine, whereas at night (5:00 pm–7:00 am), the average time between excretion and cleaning was 480 min. Based on the above findings, the emissions of VOCs in the interval between excretion and cleaning of the shed were estimated. As a result, the emissions of VOCs from feces and urine per head of cattle in the shed were estimated to be 1.75 and 1.52 g day⁻¹, respectively. Furthermore, contribution of VOCs emitted from manure to odor activity value (OAV) and hydroxyl radical reactivity (OHR) were also estimated. Volatile fatty acids and sulfur compounds emitted from feces estimated to have high contribution to OAV, whereas aldehydes contributed mainly for OHR from manure.

KEY WORDS Volatile organic compounds, Dairy cattle, Manure, Chemical composition, Emission rate, Emissions

1. INTRODUCTION

Volatile organic compounds (VOCs) are emitted from human activities, nature, and other sources, and these compounds contaminate the atmosphere. The annual emissions of VOCs in Japan in 2018 were estimated to be 641 Gg (Ministry of the Environment, Japan, 2020), and various emission sources such as paint, fuel, transportation, and the chemical and manufacturing industries have been identified as the dominant sources. In general, atmospheric VOCs are odorous (Ito, 2005). In addition, some VOCs in the atmosphere form photochemical oxidants (Ox) and

secondary organic aerosol (SOA) via reaction with ozone and free radicals (Giganek *et al.*, 2008). The achievement rate of environmental standards for Ox in Japan is less than 1% (Ministry of the Environment, Japan, 2020). In addition, the values for atmospheric organic carbon calculated by numerical models tend to be lower than the measured values (Fushimi *et al.*, 2011). One of the reasons for this may be the existence of emission sources that are not taken into account in the models. One such source is the livestock industry. In fact, the annual average concentrations of non-methane hydrocarbons (NMHC) in the dairy area in Hokkaido and the area where poultry and pig farming are active in the northern Kanto region were almost the same as those in urban areas (National Institute for Environmental Studies). This suggests that the contribution of NMHC from livestock and agriculture to atmospheric concentrations was significant.

In the United States, it has been estimated that about 10% of atmospheric VOCs are derived from livestock farming (CDPR, 2006). Several reports on VOC emissions from the livestock industry have been published. Ni *et al.* (2012) reported that approximately 300 VOCs were detected and quantified in animal facilities. Alanis *et al.* (2010) reported that the feces and urine of swine were major source of volatile fatty acids (VFAs) in a swine shed. There are a number of reports concerning the VOCs emitted from swine manure. Parker *et al.* (2012) evaluated the volatilization of VOCs from swine manure in a chamber. They found that the emission rates (ERs) of the VOCs rapidly declined after land application and were below or near the detection limit within 4 to 8 h. Laor *et al.* (2007) reported that the amount and composition of VOCs emitted from the manure of dairy calves varied significantly with changes in feed content and rumen development as the calves grew.

VOCs emitted from livestock industry are one of the main sources of odor in the atmosphere around the world (e.g., Hwang *et al.*, 2018; Orzi *et al.*, 2010). Also, there are many complaints of odor caused by livestock industry in Japan (Ministry of Agriculture, Forestry and Fisheries, 2020). In addition, there have been several reports on the contribution of VOCs from livestock industry to oxidant production in the troposphere. Howard *et al.* (2010) estimated the ozone formation potential (OFP) of VOCs emitted from livestock industry in the San Joaquin Valley (SJV) of California. They reported that, livestock industry was accounted for 40% of OFP of

small gasoline vehicles. Hu *et al.* (2012) estimated that the cumulative mass of 8-hour average ozone derived from VOCs from mobile sources and livestock feed at SJV for the period 2000–2020. They showed that the VOC emissions from mobile sources will produce less O₃ in the future, making the VOC emissions from livestock feed more important.

There are only a few reports on emissions of VOCs from the livestock industry in Japan (Tanaka *et al.*, 2020, 2019; Osaka *et al.*, 2018). These studies estimated the VOC emissions from livestock sheds based on the VOC concentrations and amount of ventilation in the shed. The sources of VOC emissions from the livestock include feces, urine, and feed, as mentioned before. To ascertain the actual emissions of VOCs from manure, it is necessary to take into account the differences in the way manure is handled, which depends on the rearing method. Dairy cattle, one of the main livestock in Japan, can be classed into two types in terms of rearing method, namely, tethered rearing and grazing, with the former accounting for 90% of the total in Japan (Takahashi, 2018). For tethered dairy cattle, the periods of VOC emissions from feces and urine can be grouped into three main categories: (1) after excretion and before removal from the shed, (2) after transportation and during the composting process, and (3) after the compost is applied to fields. By contrast, in the case of grazing, the emissions of VOCs from feces and urine are considered to be basically confined to the pastureland. Therefore, in order to clarify the actual situation regarding VOC emissions from the manure of tethered dairy cattle, it is necessary to estimate emission amounts of VOCs at each stage. However, there is no report clarifying the details of the emissions of VOCs from the manure of tethered dairy cattle. In this context, the aim of present work is to estimate the amount of VOC emissions from the feces and urine of tethered dairy cattle in a cattle shed. In addition, odor and oxidant production will be discussed as environmental effects of VOCs emitted to the atmosphere from livestock industry, and the components that contribute to these effects will be clarified.

2. EXPERIMENTAL CONDITIONS

2.1 Target Compounds

The target compounds were eight VFAs (acetic acid, propanoic acid, isobutyric acid, butyric acid, isovaleric

acid, valeric acid, hexanoic acid, heptanoic acid), three phenols (phenol, *p*-cresol, 4-ethyl phenol), two indoles (indole, skatole), two sulfur compounds (dimethyl sulfide, dimethyl disulfide), eleven aldehydes (formaldehyde, acetaldehyde, acrolein, propionaldehyde, crotonaldehyde, methacrolein, *n*-butylaldehyde, benzaldehyde, valeraldehyde, *m*-tolualdehyde, hexaldehyde), five alcohols (methanol, ethanol, 1-butanol, 1-propanol, 2-butanol), and three ketones (acetone, 2-butanone, 2-pentanone). These compounds are typically detected in livestock sheds (e.g., Alanis *et al.*, 2010; Parker *et al.*, 2012; Osaka *et al.*, 2018).

2.2 Sampling

Samples of feces and urine were collected from a cattle shed where Holstein dairy cattle were raised in a shed, at Asahi Agricultural High School (latitude 35°43'00"N, longitude 140°39'36"E) located in the northeast of Chiba Prefecture, Japan. The volume of the shed was 522.7 m³ (11.0 m width × 14.4 m depth × 3.3 m height). Holstein is major breed of dairy cattle raised in Japan. The amounts of feed per day and per head were as follows: alfalfa: 2 kg; bermudagrass: 4 kg; Italian ryegrass and Phleum pratense: a little; *Miraku-yu70* (Marubeni Nissin Feed Co., Ltd): 2 kg/time × 4 times; corn: 2 kg; corn silage: 2 kg. The manure was removed every 2 h during the day. Feces and urine were collected from the cattle shed on 13 March 2020, when the standard diet was fed and the temperature in the shed (17°C) was close to the annual average temperature (16°C) at Yokoshiba-hikari national weather station located 16 km WSW from the sampling point. Feces and urine were collected once from each of two lactating dairy cattle (average weight: about 650 kg). After measuring the weight of the collected manure, portions of the manure were placed in a polyethylene (PE) bag and a polypropylene (PP) bottle and sealed and then stored in a freezer at -20°C. Since the color and viscosity of the collected feces were almost uniform in appearance, it was assumed that the ingredients were also almost uniform, and a portion of the sample was fractionated without mixing. Freezing the manure samples would not likely alter the odorous compound flux (Hales *et al.*, 2015; Miller *et al.*, 2006). During the sampling campaign, three cameras were installed in the cattle shed. Photos were taken every 10 s to monitor the defecation behavior of the dairy cattle and worker activity.

2.3 Evaluation of VOC Emissions from Manure

The VOC emissions from the manure were evaluated with reference to Parker *et al.* (2012). A schematic outlining the approach for evaluating of VOC emissions from manure is shown in Fig. 1. A known amount of feces and urine were placed on a tray in a stainless steel (SS) chamber (300 mm × 68 mm × 120 mm). Since dairy cattle feces contains a lot of water, it naturally becomes flat when it falls on the floor. Therefore, in this experiment, feces was flattened on a tray (5 cm × 10 cm × 1 cm) in the chamber to simulate the condition of feces falling on the floor. Chamber experiment was conducted at 22°C, which is 6°C higher than the annual average temperature (16°C) at the Yokoshiba-hikari national weather station near the sampling point in 2020. On the other hand, the temperature and humidity inside the shed tends to be higher than outside the shed, so the temperature condition in this experiment is considered to roughly simulate the annual average temperature inside the shed. Sorbent tubes filled with silica gel and activated carbon were installed upstream of the chamber, and the room air was passed through the tubes to remove moisture and organic compounds in the air. It was confirmed that the target VOCs were not detected in the air downstream the tube by GC/MS measurement. Clean air was flowed into the chamber at 2.5 L min⁻¹, and the VOCs emitted from the feces and urine were collected downstream of the chamber. The linear velocity of clean air in the chamber was 0.11 cm s⁻¹, which is considered close to the shed environment where there is usually little wind. Since the volume of the chamber was about 2.5 L, ventilation rate at 2.5 L min⁻¹ corresponds to 60 times h⁻¹ of ventilation times. Note that the average ventilation rate in the shed calculated using temperature and humidity data inside and outside the shed on the day of manure collection, referring to Tanaka *et al.* (2019), was 104 times h⁻¹, so the ventilation rate in this experiment roughly simulates that in the shed. The VFAs, alcohols, indoles, sulfur compounds, phenols, and 2-pentanone were collected using a SS tube filled with Tenax TA sorbent (3.5 inch × 1/4 inch OD, 60/80, COMSCO). Prior to sample collection, all the sampling tubes were conditioned by flowing a stream of pure nitrogen gas at a flow rate of 50 mL min⁻¹ for 60 min at 300°C. Sampled air was pulled through the sorbent tube at a flow rate of 0.1 L min⁻¹ for 10 min with a sampling pump (MP-Σ30NII, Shibata Scientific Technology). After sampling, the sorbent tubes were sealed at

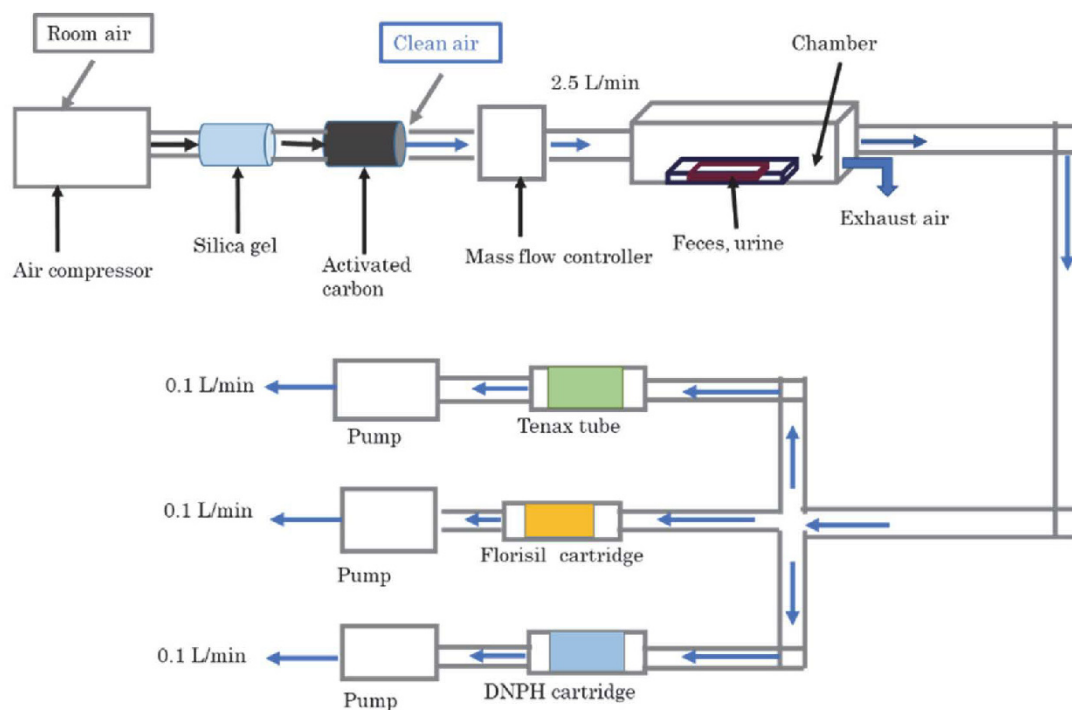


Fig. 1. Schematic of chamber experiment to evaluate VOC emissions from manure.

both ends with SS caps and put into an aluminum bag with a zipper and kept in a freezer at -20°C . Aldehydes and ketones except 2-pentanone were collected with two cartridges containing 2,4-dinitrophenylhydrazine (DNPH) as a derivatizing agent (InertSep mini AERO DNPH-LG, GL Sciences). An ozone scrubber cartridge (InertSep mini AERO Ozone Scrubber, GL Sciences) was connected upstream of the two DNPH cartridges. The sampled air was flowed through the cartridges at a flow rate of 0.1 L min^{-1} . After sampling, the cartridges were sealed at both ends with caps and put into an aluminum bag with a zipper and kept in a freezer at -20°C . Methanol and ethanol were collected with a cartridge filled with silica gel and magnesium oxide (Presep-C Florisil, Fujifilm Wako Pure Chemicals). The sampled air was flowed through the cartridge at a flow rate of 0.1 L min^{-1} . After sampling, the cartridge was sealed at both ends with caps and put into an aluminum bag with a zipper and kept in a freezer at -20°C . After the feces and urine was placed in the chamber, the samples were collected four times over the period between 3–483 min. Each sampling procedure was carried out for 10 min for compounds collected with the Tenax tube and the Florisil cartridge and for 50 min for compounds collected with the DNPH cartridges.

2.4 Analytical Procedures

The samples were pretreated and analyzed as described by Osaka *et al.* (2018). The standards for the target compounds and the reagents for pretreatment were special grade, dioxane grade, or HPLC grade reagents (Fujifilm Wako Pure Chemicals, Kanto Chemical or Sigma-Aldrich). The VOCs collected with Tenax tubes were analyzed by gas chromatography-mass spectrometry (GC/MS) equipped with a thermal desorption injector (TD-GC/MS; GCMS-QP2020 and TD-20, Shimadzu Corporation). Analytical condition of TD-GC/MS was shown in Table 1. The aldehydes and ketones collected by the DNPH cartridge were processed before analysis. A strong cation exchange resin (InertSep mini AERO SC, GL Sciences) was conditioned with 5 mL of acetonitrile, 5 mL of purified (ion-exchange) water, 20 mL of 0.1 M hydrochloric acid solution, 20 mL of purified (ion-exchange) water, and 5 mL of acetonitrile. After conditioning, the strong cation exchange cartridge was connected downstream of the DNPH cartridge and the DNPH derivatives were eluted with 5 mL acetonitrile at a flow rate of 1 mL min^{-1} . The eluate was concentrated, and the volume was adjusted to 1 mL with acetonitrile. The samples were analyzed by high-performance liquid chromatography (HPLC). Analytical condition of HPLC

Table 1. Analytical condition of TD-GC/MS.

TD		GC/MS	
Heating temperature on conditioning tube	230°C	Column	CP-Wax58 (50 m × 0.25 mm × 0.20 μm, GL science)
Purge flow rate	50 mL/min (3min)	GC oven temperature program	35°C (hold 5 min) → (ramp 10°C/min) → 200°C (hold 10 min)
Cooling temperature on trapped tube	-20°C	Injection mode	Spitless
Heating temperature on trapped tube	230°C	Career gas	Helium (1.4 mL/min)
Line temperature	230°C	Temperature of interface	200°C
		Temperature of ion source	210°C
		Measurement method	Select ion monitoring (SIM) and scan mode

Table 2. Analytical condition of HPLC.

Column	Deltabond Resolution AK 200 mm × 4.6 mm × 5 μm (Thermo Fisher Scientific)	
Column oven temperature	40°C	
Eluents	A: acetonitrile B: Acetonitrile solution containing water (10% by distilled water)	
Gradient program	Time (min)	Eluent A/B-(%)
	0.0-35.0	35.0 / 65.0 → 65.0 / 35.0
	35.0-35.2	35.0 / 65.0 → 80.0 / 20.0
	35.2-40.0	80.0 / 20.0
	40.0-40.2	80.0 / 20.0 → 35.0 / 65.0
40.2-45.0	35.0 / 65.0	
UV wavelength for detection	365.8 nm	

was shown in Table 2. Methanol and ethanol collected by the Florisil cartridge were processed before analysis. 3 mL of purified water were added to the Florisil cartridge to extract the methanol and ethanol. The eluate was analyzed by GC/MS (GCMS-QP2020, Shimadzu Corporation). Analytical condition of methanol and ethanol by GC/MS was shown in Table 3.

2.5 Estimation of VOC Emissions from Feces and Urine of Dairy Cattle

The ERs of VOCs from manure were estimated using the following procedure. From the results of the chamber experiment (see section 2.3), the amount of component i of the VOCs emitted from the manure at sampling time t_n to t_{n+1} ($= T_n$), M_{i,T_n} (μg), can be expressed as follows:

$$M_{i,T_n} = C_{i,T_n} \times V_{T_n} / V_{x,T_n} \quad (1)$$

Here, C_{i,T_n} (μg) is the amount of component i collected in the sampling tube during T_n , V_{T_n} (L min⁻¹) is ventilation rate in the chamber during T_n , V_{x,T_n} (L min⁻¹) is the flow rate for sample collection during T_n . The ER of component i during T_n (ER_{i,T_n} in μg min⁻¹ g-feces or urine⁻¹) from the manure is expressed as

$$ER_{i,T_n} = M_{i,T_n} / n / T_n \quad (2)$$

Where n (g) is amount of feces or urine used in the chamber experiment. Based on the calculation described above, the emission of component i from the feces or urine in the shed was estimated as follows:

$$E_i = \int_0^x ER_{i,T_n}(t) dt \quad (3)$$

Here, E_i (μg g⁻¹) is total emission of component i from 1 g of feces or urine, x is the average time between excretion ($= 0$) and removal by cleaning of the shed, $ER_{i,T_n}(t)$ is a function for the approximation curve describing the relationship between the ER of component i and time, and can be obtained from the above calculation. Based on the analysis of the camera images, the values for x were estimated as 80 min during the day, and 400 min at night (see section 3.3).

The emission of component i from the manure per head of cattle in the daytime in the shed ($E_{i,daytime}$ in μg head⁻¹) is expressed as

$$E_{i,daytime} = E_i \times N \times T_{daytime} \quad (4)$$

Where N (g times⁻¹) is average amount of feces or urine per unit time, $T_{daytime}$ (times head⁻¹) is the number of excretions per head in the daytime. Similarly, the emission of component i from the manure per head of cattle in the nighttime in the shed ($E_{i,night}$ in μg head⁻¹) can be

Table 3. Analytical condition of methanol and ethanol by GC/MS.

Column	CP-Wax58 (50 m × 0.25 mm × 0.20 μm, GL science)
Column oven temperature	40°C (4 min) → (5°C/min) → 56°C → (100°C/min) → 200°C (10 min)
Injection port	200°C
Ion source	210°C
Measurement method	SIM mode

expressed as

$$E_{i,\text{night}} = E_i \times N \times T_{\text{night}} \quad (5)$$

Where T_{night} (times head⁻¹) is number of excretions per head in the nighttime. Finally, the emission of component i from the manure per day per head ($E_{i,\text{day}}$) of cattle can be expressed as follows:

$$E_{i,\text{day}} = E_{i,\text{daytime}} + E_{i,\text{night}} \quad (6)$$

The daily emissions of total VOCs from the manure per head in the shed (E) is the sum of the emissions of each VOC. The VOC emissions during the time period when sampling was not conducted were supplemented by an approximate equation between time and VOC emissions obtained from the experiment. Note that, the total amount of the emission characteristics of VOCs from excrements depends on a variety of factors, including feed, breed, and life stages of the cattle (Laor *et al.*, 2007).

2.6 Evaluation of the Contribution of VOCs Emitted from Dairy Cattle Manure to Odor

Odor activity value (OAV) was used as an evaluation index for odor. OAV of compound i (OAV_i) can be expressed as follows.

$$OAV_i = C_i / OTV_i \quad (7)$$

Here, C_i (μg/m³) is concentration of compound i , OTV_i (μg/m³) is odor threshold of compound i . In order to evaluate the relative contribution of VOCs emitted from manure to OAV in this study, C_i was determined by dividing the total emission of i from manure by 1 m³. Hence, it is important to note that the absolute value of OAV calculated in this study is not meaningful. OTV of each VOC used the value reported by Nagata *et al.* (2003).

C_i (μg/m³) can be expressed as follows.

$$C_i = E_{i,\text{day}} \times n_{\text{dairy cattle}} / V \quad (8)$$

Here, $E_{i,\text{day}}$ is emission of i from manure per day per head as shown in (6), $n_{\text{dairy cattle}}$ is head of dairy cattle in the shed, and V (m³) is 1 m³.

2.7 Evaluation of the Contribution of VOCs Emitted from Dairy Cattle Manure to Hydroxyl Radical Reactivity

Hydroxyl radical (OH) reactivity (OHR) was used as an evaluation index for oxidant formation ability of VOCs. OHR of compound i (OHR_i) can be expressed as follows (Atkinson *et al.*, 2006).

$$OHR_i = C_i \times K_i \quad (9)$$

Here, C_i (mol/m³) is concentration of compound i , K_i (m³ mol⁻¹ s⁻¹) is reaction rate constant of the compound i with OH. In order to evaluate the relative contribution of VOCs emitted from manure to OHR in this study, C_i was determined by dividing the total emission of i from manure by 1 m³. Hence, note that the absolute value of OHR calculated in this study is not meaningful.

3. RESULTS AND DISCUSSION

3.1 Composition of the Emissions of VOCs from Feces and Urine

Composition of VOCs average distribution of each sample from feces and urine is shown in Fig. 2 ($n = 2$). Sulfur compounds and ketones accounted for about 60% of the total VOCs emitted from feces, followed by aldehydes. Sulfur compounds and ketones were the dominant VOCs emitted from urine and accounted for 90% of the total VOCs. Hales *et al.* (2015) reported that VFAs such as acetic acid and propanoic acid were the major compounds emitted from the manure of Jersey cattle. Shaw *et al.* (2007) reported that methanol, ethanol, and acetone were the predominant VOCs emitted from the manure of Holstein cattle. These differences in composition are thought to be influenced by the feed, breed, and life stage of the cattle. Hales *et al.* (2015) compared the fluxes of the VOCs emitted from the manure of cattle fed conventional livestock diets: steam-flaked corn (SFC), and dry-rolled corn (DRC). Hales *et al.* found that the total flux (μg m⁻² min⁻¹) for 13 VOCs, such as phenols and VFAs emitted from the manure was

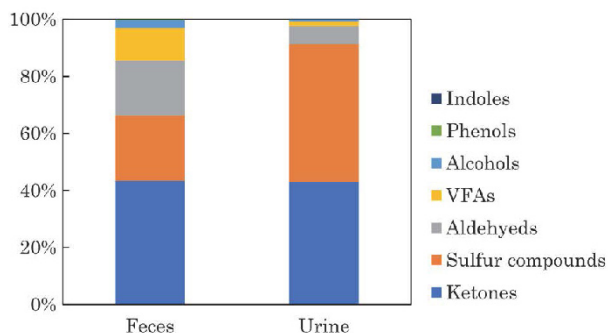


Fig. 2. The average composition of VOCs mitted from feces and urine. Four samples were collected intermittently between 3–483 min in the experiment. The data shown in the figure is the average of four samples.

2.1 times higher for cattle fed DRC compared with SFC. Shaw *et al.* (2007) also compared the fluxes of manure from milking cows at different growth stages and reported that the flux of methanol was about 3 times higher than that of far-off cows. In contrast, manure was collected from lactating dairy cattle in this study. Therefore, the difference in the composition of VOCs emitted from the manure in the present study compared with previous reports may be due to the differences in feed, type of livestock, and growth stage of the cows.

3.2 ERs of VOCs from Manure

The temporal variation in the ERs of VOCs from feces is shown in Fig. 3. 2-pentanone is not shown in the figure because it had a minimal effect on the ERs. The ERs of aldehydes and ketones decreased over time (VOCs except aldehydes and ketones: $y = 0.21e^{-0.0008x}$, $R^2 = 0.21$; aldehydes and ketones: $y = 2.9 \times 10^{-2}e^{-0.002x}$, $R^2 = 0.73$; x : time from excretion (min), y : ER ($\mu\text{g min}^{-1} \text{g}^{-1}$)). The temporal variation in the ERs of VOCs from urine is shown in Fig. 4. It can be seen that the ERs of the VOCs tended to decrease rapidly with time, with the exception of some compounds such as VFAs. Lee *et al.* (2003) mentioned that VOC emissions from varnish were fitted using a first order decay model, instead of a double-exponential model previously used (Guo and Murray, 2000a; Guo *et al.*, 2000b). They reported that comparatively reliable estimates of emission parameters when VOC emissions from indoor materials were mainly controlled by evaporation. Parker *et al.* (2012) also reported that an exponential decrease in VOCs emissions from manure. Based on the above, we assumed that the emissions of VOCs from manure also decrease expo-

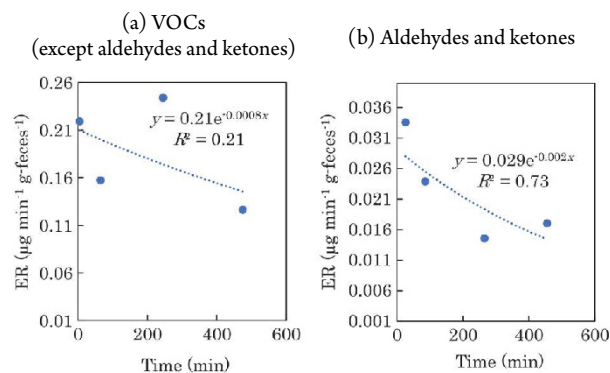


Fig. 3. Temporal variation in ERs of VOCs from feces.

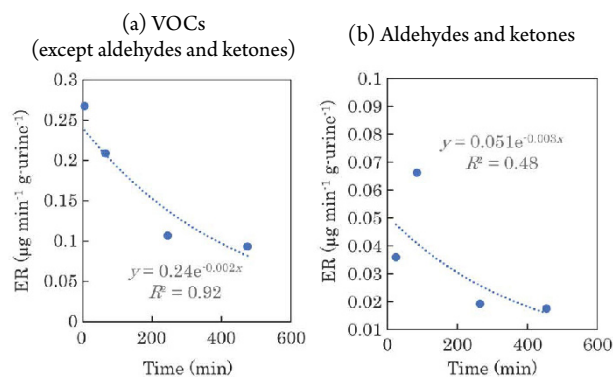


Fig. 4. Temporal variation in ERs of VOCs from urine.

entially with time, and approximated it as an exponential function. The regression equations are as follows; VOCs except aldehydes and ketones: $y = 0.24e^{-0.002x}$, $R^2 = 0.92$; aldehydes and ketones: $y = 0.051e^{-0.003x}$, $R^2 = 0.48$. Indoles and 2-pentanone were not detected from urine. A tendency of the ERs from feces to decrease over time was observed by Hales *et al.* (2015), and the results of this study are consistent with those previous results. The temporal variation in the ERs of each chemical compound emitted from feces and urine is shown in Fig. 5 and Fig. 6, respectively. Fig. 5 shows that the ERs for alcohols, ketones, and phenols tended to decrease over time. The ERs of sulfur compounds and aldehydes, on the other hand, were almost constant over time. Furthermore, the ERs of VFAs and indoles tended to increase with time. As shown in Fig. 6, the ERs of alcohols, ketones, phenols, and sulfur compounds from urine tended to decrease with time. In contrast, for VFAs and aldehydes, there was no clear correlation between the elapsed time and the ERs. The trends of temporal varia-

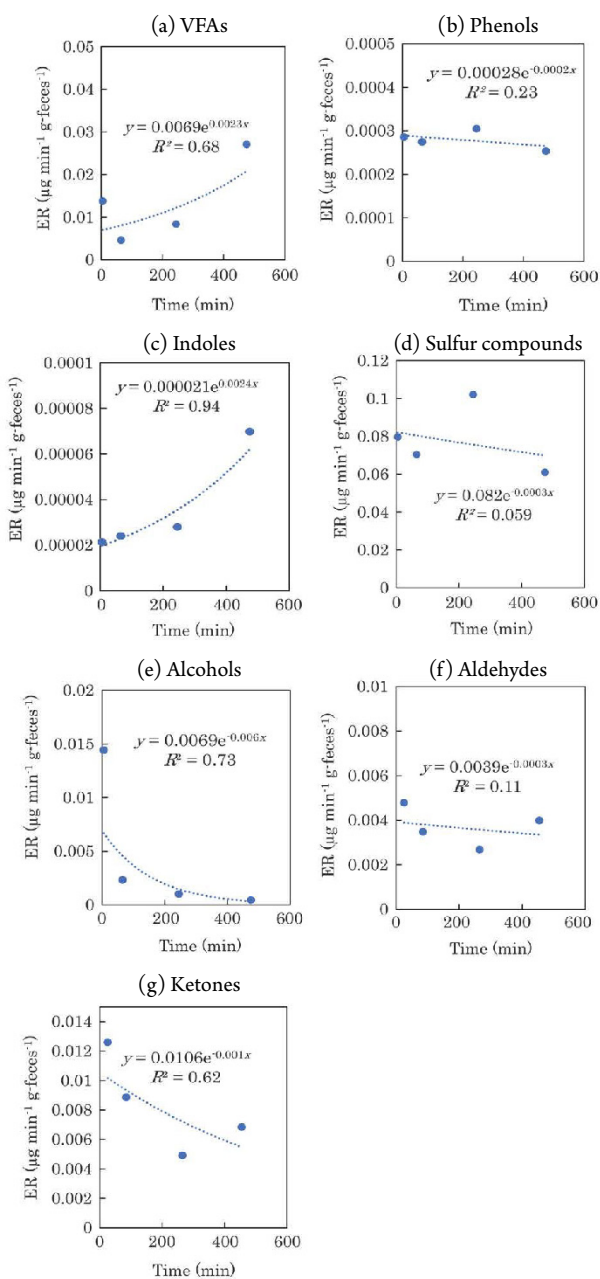


Fig. 5. Temporal variation in ERs of each chemical class emitted from feces. (a) VFAs, (b) Phenols, (c) Indoles, (d) Sulfur compounds, (e) Alcohols, (f) Aldehydes, (g) Ketones.

tion in the ERs differed according to the class of chemical compound. This may reflect the differences in the saturation vapor pressure of each VOC. For example, VFAs, which showed a tendency of increasing ERs with time, had low saturation vapor pressure (saturation vapor pressure of the target VFAs at 20°C: 0.23–15.2 hPa), whereas ketones, those ERs decreased with time, had

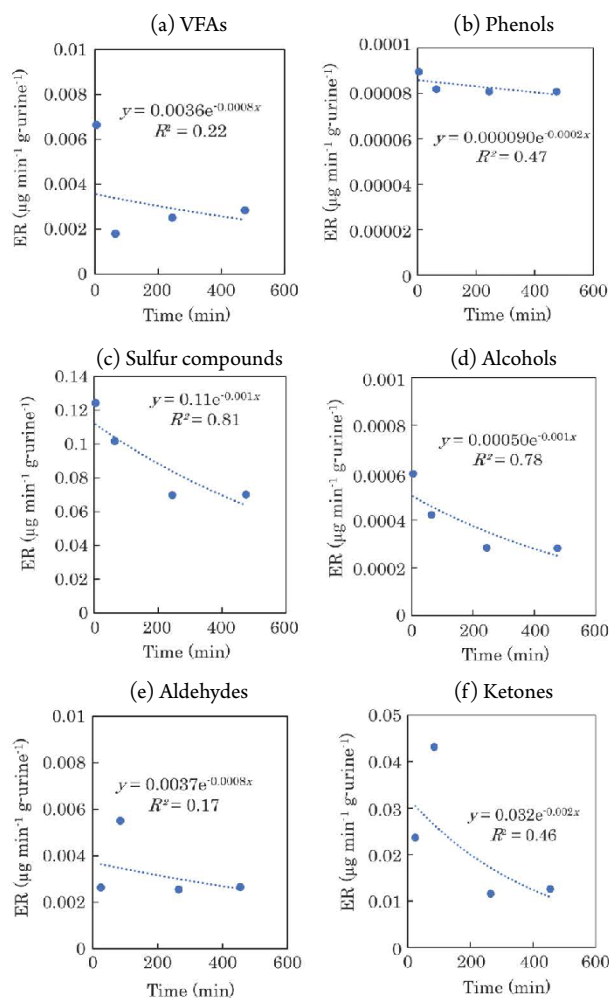


Fig. 6. Temporal variation in ERs of each chemical class emitted from urine. Indoles were not detected. (a) VFAs, (b) Phenols, (c) Sulfur compounds, (d) Alcohols, (e) Aldehydes, (f) Ketones.

high saturation vapor pressure (saturation vapor pressure of target ketones at 20°C: 103–239.5 hPa). In addition, urine is by nature liquid, while feces consist of solids and liquid. These differences in phase may affect the volatility characteristics of the VOCs. In fact, the ERs of VOCs from swine manure, such as phenols, indoles, and sulfur compounds decayed over time, whereas that of the eight VFAs exhibited high variability (Parker *et al.*, 2012). Thus, although the ERs may not decay monotonically with time for some components, the overall trend was that the ERs of VOCs tended to decline as time elapsed. In addition, during the composting process of dairy cattle manure, anaerobic microorganisms such as *Methanotrix* and *Acetobacter* in the manure decompose the easily degradable organic matter (Yokoyama, 2009). Altho-

Table 4. Emissions of VOCs from 1 g of feces and percentage of each VOC.

	Day ¹⁾		Night ²⁾	
	Emissions (g g ⁻¹)	Percentage (%)	Emissions (g g ⁻¹)	Percentage (%)
VFAs	9.04×10^{-7}	5.53	8.21×10^{-6}	11.8
Phenols	3.94×10^{-6}	0.24	2.28×10^{-7}	0.328
Indoles	2.12×10^{-9}	1.30×10^{-2}	1.55×10^{-8}	2.22×10^{-2}
Sulfur compounds	6.04×10^{-6}	37.0	1.41×10^{-5}	20.3
Aldehydes	2.25×10^{-6}	13.7	1.34×10^{-5}	19.2
Alcohols	6.23×10^{-7}	3.81	1.65×10^{-6}	2.37
Ketones	6.48×10^{-6}	39.7	4.83×10^{-5}	45.9
Total	1.63×10^{-5}	100	6.96×10^{-5}	100

¹⁾8:00 am–5:00 pm.

²⁾5:00 pm–7:00 am.

ugh composting is generally carried out mainly at 50–60°C, useful fermenting microorganisms such as acetic acid bacteria generally prefer at 15–25°C (Matsushita, 2010). This temperature range is comparable to the temperature (20–25°C) at which the chamber experiment in this study was conducted. As shown in Fig. 5(a), the increased ERs of VFAs over time from the feces may be due to the production of VFAs by the microorganisms in the feces.

3.3 Estimation of VOC Emissions from Manure in a Dairy Cattle Shed

Based on the results described above, the VOC emissions from feces and urine in the cattle shed were estimated. The dairy cattle shed targeted in this study had a tethered system and excrement was regularly removed from the shed. Thus, the VOCs emitted from the manure were considered to be emitted between excretion and cleaning of the shed. As a result of analyzing the images of the three cameras which were installed in the shed, the average time between excretion and cleaning during the daytime (8:00 am–5:00 pm) was 80 min for feces and urine. At night (5:00 pm–7:00 am), when the staff were not available and cleaning was not performed, the average time between excretion and cleaning was 480 min for both feces and urine. In other words, the VOCs derived from excretion in the shed during the daytime and nighttime can be considered to be emitted on average for time durations of 80 and 480 min, respectively. Therefore, by integrating the approximation equation specified integrating as shown in (3) with respect to the average time between excretion and cleaning, the amount of VOC

emissions originating from excretions in the shed during the day and at night can be estimated. First, the data for the emissions of the VOCs from 1 g of feces and urine for one dairy cow are shown in Tables 4, 5. The VOC emissions were calculated as 1.63×10^{-5} g (g-feces)⁻¹ and 2.90×10^{-5} g (g-urine)⁻¹ during the daytime. At nighttime, the emissions of the VOCs from the feces and urine were 6.96×10^{-5} g g⁻¹ and 1.30×10^{-4} g g⁻¹, respectively. For both feces and urine, sulfur compounds and ketones accounted for 60–90% of the total VOCs, followed by aldehydes. Table 4 indicates that the composition of VOCs emitted from the feces is different during the daytime and at night. For example, the percentage of VFAs and indoles that tended to have higher ERs over time was relatively high. This is because the ERs of VFAs and indoles from the feces increased with time, while the ERs of aldehydes and ketones tended to decrease. On the other hand, Table 5 shows that the composition of VOCs emitted from urine was almost the same during the daytime and night. This is because the ERs of most compounds from urine decreased with time as shown in Fig. 6. From Fig. 6, we can see that the ERs of sulfur compounds did not decay much with time compared to the other compounds, whereas the ERs of ketones decayed with time after excretion. In other words, when the excrement was left for a relatively long time, such as overnight, the composition of each component changed over time. In contrast, the ERs of VOCs from urine decayed uniformly with time for many components. Therefore, the composition of the components emitted from urine was almost the same for both daytime and nighttime.

Next, the data for the total weight of feces and urine

Table 5. Emissions of VOC from 1 g of urine and percentage of each VOC.

	Day ¹⁾		Night ²⁾	
	Emissions (g g ⁻¹)	Percentage (%)	Emissions (g g ⁻¹)	Percentage (%)
VFAs	4.60 × 10 ⁻⁷	1.59	2.10 × 10 ⁻⁶	1.60
Phenols	9.53 × 10 ⁻⁹	3.29 × 10 ⁻²	5.10 × 10 ⁻⁸	3.91 × 10 ⁻²
Indoles	N.D. ³⁾	N.D. ³⁾	N.D. ³⁾	N.D. ³⁾
Sulfur compounds	1.39 × 10 ⁻⁵	47.8	6.26 × 10 ⁻⁵	48.0
Aldehydes	1.56 × 10 ⁻⁶	5.39	8.45 × 10 ⁻⁶	6.48
Alcohols	2.24 × 10 ⁻⁷	0.775	8.61 × 10 ⁻⁷	0.660
Ketones	1.29 × 10 ⁻⁵	44.4	5.63 × 10 ⁻⁵	43.2
Total	2.90 × 10 ⁻⁵	100	1.30 × 10 ⁻⁴	100

¹⁾8:00 am–5:00 pm.

²⁾5:00 pm–7:00 am.

³⁾Not detected.

Table 6. Total weight of feces and urine per unit time and the average number of feces and urine per head of cattle per day.

	Weight			Number					
	Total weight (g)	n ¹⁾	SD ²⁾	Day	n ¹⁾	SD ²⁾	Night	n ¹⁾	SD ²⁾
Feces	3.0 × 10 ³	4	0.14	3.0	6	0.069	6.1	6	0.88
Urine	2.8 × 10 ³	4	0.45	3.4	6	0.68	3.4	6	0.21

¹⁾Number of samples.

²⁾Standard deviation.

Table 7. Emissions of VOC per head of dairy cattle (g day⁻¹) from the cattle shed for feces and urine.

	Feces	Urine
VFAs	0.158	0.0244
Phenols	0.0396	5.76 × 10 ⁻⁴
Indoles	3.03 × 10 ⁻⁴	N.D. ¹⁾
Sulfur compounds	0.312	0.728
Aldehydes	0.265	0.0953
Alcohols	0.0358	0.0103
Ketones	0.942	0.659
Total	1.75	1.52

¹⁾Not detected.

per unit time and the average amount of feces and urine per head of cattle per day based on the analysis of the camera images are presented in Table 6. These data indicated that the weights of both feces and urine were almost constant regardless of the time, the individual cattle, or the season in which they were collected. The total weight of the feces and urine was taken as the average weight collected in November 2019 and March 2020. Number of times of feces and urine was taken as the

average time during the sampling campaigns in August 2019, November 2019, and March 2020. Based on these results, it was, therefore, assumed that the amount of excretion was constant regardless of the time, the individual, and the season.

Emissions of the VOCs from the feces and urine in the shed were estimated by applying the values in the table to equations (4)–(6). From Table 7, the emissions of the VOCs per head from the shed derived from the feces and urine were 1.75 and 1.52 g day⁻¹, respectively. Except for sulfur compounds, emissions from feces were higher than from urine. The emissions of sulfur compounds from urine were 2.3 times higher than from feces. In this study, the total weight of feces and urine per head per day was estimated to be 27 kg and 19 kg, respectively, and total weight of manure was estimated to be 46 kg. In an earlier study, Yamada *et al.* (2009) reported that the average total weight of manure per head was 64.1 ± 7.1 kg day⁻¹, where the cattle were reared in a free stall and the average body weight was 700 kg. This weight of manure was about 1.1 times higher than the weight in the present study. The amounts of feed per head per day in the present study and that of Yamada *et al.* were 18 kg

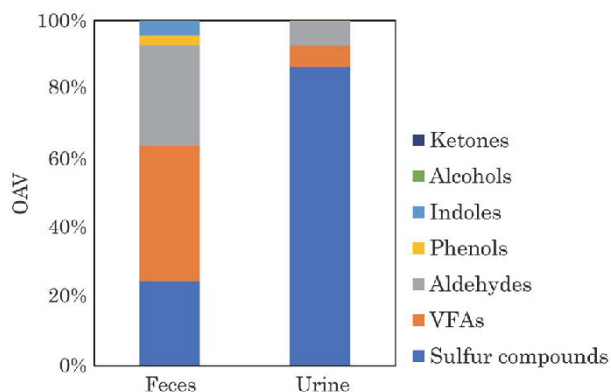


Fig. 7. Contribution of each chemical group emitted from dairy cattle manure to OAV.

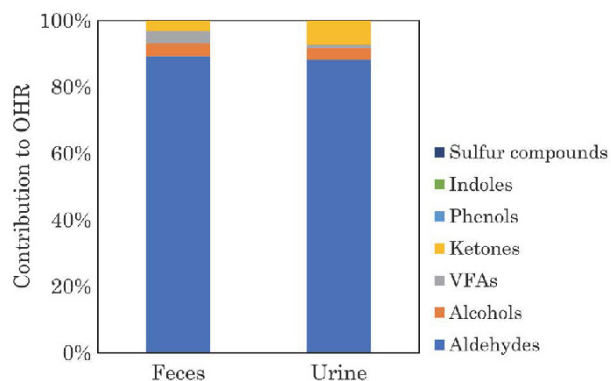


Fig. 8. Contribution of each chemical group emitted from dairy cattle manure to OHR.

and 22.5 kg, respectively; that is, the previous amount was 1.25 times higher than in the present study. The amount of excretion by the dairy cattle estimated for this study was less than that reported by Yamada *et al.*, and this difference may be due in part to the difference in the amount of feed.

This study is the first attempt to estimate the VOC emissions from the manure of tethered dairy cattle in Japan. It is thought that the emissions of VOCs from excretions in the shed can be evaluated more accurately by estimating the time it takes for manure to be removed from the shed.

3.4 Contribution of Each VOC from Manure of Dairy Cattle to OAV and OHR

The contribution of VOCs derived from manure to the OAV is shown in Fig. 7. VFAs, aldehydes, and sulfur

compounds accounted for about 90% of OAV from feces, while sulfur compounds accounted for 86% of OAV from urine. Ketones, which were highly emitted from feces and urine, made little contribution to OAV because their OTV is very high compared to other substances (e.g., the OTV of acetone is about 14,000 times higher than that of dimethyl sulfide). Yuan *et al.* (2017) reported that sulfur compounds accounted for about 80% of OAV in the dairy cattle shed. Hobbs *et al.* (1999) and Nie *et al.* (2020) reported that H₂S was main compounds of OAV in pig slurry. Therefore, sulfur compounds may contribute to OAV mainly regardless live-stock species.

The contribution of VOCs derived from manure to the OHR is shown in Fig. 8. Aldehydes were dominantly contributed to OHR, followed by alcohols. Yuan *et al.* (2017) reported that alcohols were main compounds of OHR in dairy cattle shed whereas alcohols were secondary main compounds of OHR in this study. The high contribution of alcohols to OHR in the shed reported by Yuan *et al.* may be due to the high ethanol emissions from silage and total mixed ration (TMR) (Bonifacio *et al.*, 2017).

4. CONCLUSIONS

In this study, the concentrations of 34 VOCs emitted from the feces and urine of dairy cattle were determined to clarify the chemical composition of the VOCs and to estimate the VOC emissions from manure in a dairy cattle shed. Dimethyl sulfide and acetone accounted for about 60% of the total VOCs from feces, followed by formaldehyde and acetaldehyde. Dimethyl sulfide and acetone were also dominant VOCs emitted from the manure, which accounted for 90% of the total VOCs. The ERs of VOCs from the manure showed a tendency to decay exponentially with time after excretion. Furthermore, the average time between excretion and cleaning during the daytime (8:00 am–5:00 pm) and during the nighttime (5:00 pm–7:00 am) was estimated to be 80 min and 480 min, respectively. Finally, the amounts of VOC emissions derived from the feces and urine in the cattle shed were estimated to be 1.75 and 1.52 g day⁻¹, respectively. The contribution of VOCs emitted from manure to OAV and OHR were also evaluated. VFAs, sulfur compounds, aldehydes accounted for about 90% of OAV from feces whereas sulfur compounds were

86% of OAV from urine. In addition, sulfur compounds were main compounds of OAV from manure regardless of livestock species. In contrast, aldehydes had the highest contribution to OHR from manure.

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