

# Muromonab CD3

## A Reappraisal of its Pharmacology and Use as Prophylaxis of Solid Organ Transplant Rejection

Michelle I. Wilde and Karen L. Goa

Adis International Limited, Auckland, New Zealand

### Various sections of the manuscript reviewed by:

*D. Abramowicz*, Department of Nephrology, Hôpital Erasme, Brussels, Belgium; *P.N. Bennett*, University of Bath, School of Postgraduate Medicine, Bath, England; *H.A. Bock*, Division of Nephrology, Kantonsspital, Basel, Switzerland; *L. Chatenoud*, Inserm U 25, Hôpital Necker, Paris, France; *J.M. Grinyó*, Institut Català de la Salut, Ciutat Sanitària i Universitària de Bellvitge, Barcelona, Spain; *K. Fukao*, Department of Surgery, University of Tsukuba, Tsukuba City, Japan; *G. Ippoliti*, Clinica Medica II, IRCCS S Matteo, Pavia, Italy; *L.J. Swinnen*, Division of Hematology/Oncology, Stritch School of Medicine, Loyola University of Chicago, Illinois, USA; *E.M. Vasquez*, Department of Pharmacy Practice, University of Illinois at Chicago, Chicago, USA.

### Contents

Summary	866
1. Pharmacodynamic Properties	870
1.1 Mechanism of Action	870
1.2 T Cell-Activating Properties	871
1.3 Immunogenicity	871
1.4 Effects on Rejection Histopathology	872
1.5 Cardiovascular, Pulmonary and Haematological Parameters	872
2. Pharmacokinetic Properties	873
2.1 Pharmacokinetic Parameters	873
2.2 Correlation Between Plasma Concentrations and Efficacy	873
3. Therapeutic Use	874
3.1 Optimal Dosage	875
3.2 Renal Transplant Recipients	875
3.2.1 Comparisons with Triple Therapy	875
3.2.2 Comparisons with Other Antilymphocyte-based Regimens	877
3.3 Hepatic Transplant Recipients	879
3.4 Cardiac Transplant Recipients	879
3.5 Special Patient Groups	881
3.5.1 High Risk Patients	881
3.5.2 Young or Elderly	882
3.6 Reuse of Muromonab CD3	882
4. Pharmacoeconomic Considerations	883
5. Tolerability	883
5.1 First-Dose Effects	883
5.1.1 Late Reactions	884
5.2 Infections	884
5.3 Neoplasia	885
5.4 Other Events	885
6. Drug Interactions	886
7. Dosage and Administration	886
8. Place of Muromonab CD3 in the Prevention of Transplant Rejection	887

## Summary

### Synopsis

*The murine monoclonal antibody muromonab CD3 (OKT3) is directed against the CD3 antigen on peripheral human T cells and effectively blocks all T cell function.*

*Prophylaxis with muromonab CD3 (5mg intravenously once daily for 10 to 14 days) as induction therapy together with corticosteroids, azathioprine and delayed cyclosporin (sequential therapy) optimises early graft function by delaying the potentially nephrotoxic and hepatotoxic effects of cyclosporin until graft function is established.*

*Although clinical data are limited (by inconsistencies in trial design and trial size), prophylactic muromonab CD3-based sequential therapy is significantly more effective than standard triple therapy in the prophylaxis of allograft rejection in renal and hepatic, but not cardiac, transplant recipients. Benefits are particularly notable in patients with delayed graft function. No significant between-treatment differences in patient survival have been observed.*

*The overall efficacy of muromonab CD3- and polyclonal-based prophylactic regimens appears to be similar, although results vary between investigators and confirmation is needed. An anti-interleukin-2 monoclonal antibody-based prophylactic regimen improved graft and patient survival compared with muromonab CD3-based prophylaxis in hepatic transplant recipients.*

*Antimuromonab CD3 antibodies may develop; however, muromonab CD3 may be successfully reused in patients with low titres.*

*Preliminary pharmacoeconomic data suggest that mean drug costs are greater with quadruple immunosuppressive regimens containing muromonab CD3, antithymocyte globulin (ATG) or antilymphocyte globulin (ALG) than with triple therapy. Drug costs with prophylactic muromonab CD3-based regimens were similar or greater than those with polyclonal-based protocols.*

*The first doses of muromonab CD3 are associated with the 'cytokine-release syndrome'. More severe first-dose events include aseptic meningitis, intragraft thromboses, seizures and potentially fatal pulmonary oedema. The incidence and/or severity of cytomegalovirus infection with prophylactic muromonab CD3-based immunosuppression is similar to or greater than that with triple therapy and ATG- or ALG-based regimens. However, the risk of infection and also the observed increase in lymphoproliferative disorders appears to be related to the degree of immunosuppression rather than to the drug itself.*

*Thus, sequential muromonab CD3-based therapy is more effective than standard triple therapy (in renal and hepatic transplant recipients) and appears to be similar to that of polyclonal-based regimens in the prophylaxis of transplant rejection. Although the routine use of prophylactic muromonab CD3 in low-risk patients with primary graft function does not appear to be justified, prophylactic muromonab CD3-based therapy has a role in patients at high risk of rejection.*

### Pharmacodynamic Properties

Muromonab CD3 (OKT3), a murine monoclonal antibody directed against the CD3 antigen (linked to the T cell antigen receptor; TCR) on mature peripheral human T cells, effectively blocks all T cell function. The mechanism of action includes antigenic modulation of the CD3/TCR complex with subsequent opsonisation and removal of circulating T cells; other mechanisms are proposed.

Cytokine release associated with an acute phase reaction occurs after the first doses of muromonab CD3. This is manifest as first-dose adverse events (see 'Tolerability' summary).

A biphasic reversible haemodynamic response and biphasic activation of coagulation and fibrinolysis, both of which coincide with cytokine release and/or complement activation, occur after initiation of muromonab CD3. Evidence suggests that intraoperative administration of the first muromonab CD3 dose is associated with fewer cardiovascular and pulmonary disturbances than administration in the immediate postoperative period.

High antimuromonab CD3 antibody titres ( $\geq 1:1000$ ) were detected in 5.8% of >12 000 serum samples from patients who received muromonab CD3 for the treatment or prevention of transplant rejection. IgG but not IgM antibodies are able to reduce the activity of muromonab CD3. Administration of concomitant immunosuppressants such as corticosteroids and azathioprine reduces the likelihood of antimuromonab CD3 antibody formation.

---

### Pharmacokinetic Properties

Muromonab CD3 is a pure standardised product for which pharmacokinetic data are limited.

Plasma muromonab CD3 concentrations vary according to the muromonab CD3 'antibody status', transplanted organ and age. Muromonab CD3 plasma concentrations were 996  $\mu\text{g/L}$  after 1 hour and 104  $\mu\text{g/L}$  at 24 hours in renal transplant recipients receiving 5mg once daily for 10 to 14 days. Mean trough steady-state serum concentrations range from 500 to 1000  $\mu\text{g/L}$  after 2 to 4 days; approximately 1000  $\mu\text{g/L}$  is required to block cytotoxic T cell function *in vitro*. Steady-state serum muromonab CD3 concentrations are achieved earlier with prophylactic administration than administration for the treatment of rejection. There is evidence of drug accumulation after repeated doses. Muromonab CD3 plasma elimination half-lives of approximately 18 hours (following administration for treatment of rejection) and 36 hours (prophylactic administration) have been reported.

---

### Therapeutic Use

There are 2 main reasons for using muromonab CD3 as induction therapy in sequential immunosuppressive regimens (including azathioprine, methylprednisolone/prednisone and delayed cyclosporin): to optimise early graft function by delaying the administration of potentially nephrotoxic and hepatotoxic cyclosporin until graft function is established; and to reduce and delay the occurrence of rejection episodes.

Data from clinical trials of muromonab CD3-based prophylaxis are limited by trial size and inconsistencies in design. However, compared with triple therapy, prophylactic muromonab CD3-based sequential therapy was significantly more effective as assessed by severity and/or incidence of rejection episodes and time to first rejection episode in renal transplant recipients and by incidence and/or severity of early acute rejection episodes and time to first rejection in hepatic transplant recipients. Benefits appeared to be maintained for up to 3 years in renal transplant recipients and were particularly marked in patients with delayed graft function. In those with delayed renal graft function, overall graft survival was significantly greater with muromonab CD3-based induction therapy than with triple therapy. Three-year graft survival in renal transplant recipients was significantly greater with sequential muromonab CD3-based therapy (delayed cyclosporin) than with cyclosporin-based therapy not containing muromonab CD3; in high-risk patients, sequential therapy was also associated with significantly greater 3-year graft survival rates than simultaneous muromonab CD3 and cyclosporin administration. Muromonab CD3-based prophylaxis appeared to be similar to triple therapy in terms of rejection incidence, time to first rejection episode

and graft survival in cardiac transplant recipients. There were no significant between-treatment differences in patient survival for any type of transplant.

In renal transplant recipients, significantly more of those receiving muromonab CD3-based prophylaxis required postoperative dialysis than those receiving triple therapy; in hepatic transplant recipients, muromonab CD3 improved or maintained renal function in the early postoperative period.

Data on the relative effects of muromonab CD3-based and polyclonal-based prophylactic regimens are conflicting. In renal transplant recipients, some investigators show muromonab CD3- and antilymphocyte globulin (ALG)- or antithymocyte globulin (ATG)-based prophylaxis to be similar in terms of rejection incidence and/or time to first rejection, while others show ALG or ATG to be superior. The incidence of rejection episodes with muromonab CD3-based induction therapy in dual renal-pancreas transplant recipients is either similar to or lower than that with ALG- or ATG-based induction. Similarly, in cardiac transplant recipients, some studies show a longer time to first rejection episode and/or a lower incidence of rejection with muromonab CD3-based prophylaxis; others show no differences or a lower incidence of rejection or a longer time to first rejection episode with ATG-based therapy. Rejection episodes were fewer with ATG than with muromonab CD3 in the 1 available study in hepatic transplant recipients. Generally, no between-treatment differences were apparent in patient and graft survival or the incidence of delayed graft function. However, an anti-interleukin-2 monoclonal antibody-based prophylactic regimen improved graft and patient survival compared with muromonab CD3-based prophylaxis in hepatic transplant recipients.

Several noncomparative trials have shown early rejection in children and adolescents undergoing renal, hepatic or cardiac transplantation to be effectively reduced by prophylactic muromonab CD3 administered as part of a sequential immunosuppressive protocol. Although rejection incidence during the first 14 days was lower with muromonab CD3-based prophylaxis than with standard triple therapy in hepatic transplant recipients, no overall reduction was observed. Further comparative trials are needed to determine the relative benefits of prophylactic muromonab CD3 in this patient group.

Although muromonab CD3 reuse following prophylactic use is not recommended in patients with anti-idiotypic antibody titres  $\geq 1:1000$ , successful retreatment may occur in patients with lower titres.

### Pharmacoeconomic Considerations

In theory, potential cost savings with prophylactic muromonab CD3 because of reduced rejection incidence compared with triple therapy may be offset by the increased incidence of infections and adverse events, but this has not been addressed in formal pharmacoeconomic assessments.

Preliminary data from cost-minimisation studies suggest that mean first-year drug costs are greater with quadruple immunosuppressive regimens containing muromonab CD3, ATG or ALG than with triple therapy, and hospital costs with prophylactic muromonab CD3-based regimens were similar to or greater than those with polyclonal-based protocols.

### Tolerability

The first doses of muromonab CD3 are associated with flu-like 'cytokine-release syndrome' symptoms (e.g. fever, chills, gastrointestinal disturbance), which occur within 45 to 60 minutes and last for 2 to 48 hours. More severe but rare first-dose effects include aseptic meningitis, intragraft thromboses, seizures and potentially fatal pulmonary oedema. The incidence and severity of initial adverse

events with muromonab CD3-based prophylaxis are similar to or greater than those associated with polyclonal antilymphocyte-based regimens.

Cytomegalovirus (CMV), herpes simplex virus and bacterial infections are the primary cause of morbidity and mortality in muromonab CD3 recipients. However, the risk of infection appears to be related to the degree of immunosuppression rather than to the drug itself. The risk of CMV infection is increased by high doses (total doses >75mg) and repeated exposure to muromonab CD3. The incidence and/or severity of CMV infection with prophylactic muromonab CD3-based immunosuppression is similar to or greater than that with triple therapy and ATG- or ALG-based regimens. The overall incidence of bacterial and fungal infections with these treatment options is largely similar.

Muromonab CD3-based immunosuppression has been associated with an increased risk of neoplasia, mainly lymphoproliferative disorders. However, this is most probably a direct result of the degree of immunosuppression; Epstein-Barr virus infection has also been implicated. The risk of lymphoproliferative disease may be increased by the use of high muromonab CD3 doses (total dose >75mg), long durations of administration, multiple courses and early retreatment. While some investigators report the incidence of neoplasia to be similar with muromonab CD3 and polyclonal preparations, others report a trend towards a greater incidence with muromonab CD3.

Other clinically significant pulmonary, cardiovascular or neurological events are uncommon.

Adverse events with muromonab CD3 may be prevented or minimised by the intraoperative administration of the first dose of muromonab CD3, pretreatment with a corticosteroid, administration of an antipyretic and antihistamine, prophylactic use of an antimicrobial(s) and correction of increased temperature and fluid overload before prophylaxis initiation.

---

## Dosage and Administration

Induction therapy with muromonab CD3 together with azathioprine, methylprednisolone/prednisone and delayed cyclosporin therapy is the most accepted muromonab CD3-based regimen for allograft rejection prophylaxis.

Although the optimal dosage of muromonab CD3 has not been established, the currently recommended adult dosage is 5mg administered intravenously once daily for 10 to 14 days, irrespective of the organ transplanted. A dosage of 2.5 mg/day has usually been used in children although higher doses may be needed. The dosage should be adjusted according to the presence of clinical signs of rejection in addition to antimuromonab CD3 antibodies, plasma muromonab CD3 concentrations and CD3+ cell levels.

Patients should be closely monitored during administration of the first few doses. Intraoperative administration of the first dose and the use of preventative measures improve the tolerability of muromonab CD3.

Evidence suggests that procoagulant activity is increased with the concomitant administration of muromonab CD3 and high-dose corticosteroids, and indomethacin may increase the risk of encephalopathy and volatile anaesthetic agents or drugs that decrease cardiac contractility increase the risk of developing cardiovascular problems when administered with muromonab CD3.

Muromonab CD3 (murine monoclonal antibody towards the Cluster of Differentiation 3 antigen; OKT3), an IgG<sub>2a</sub> immunoglobulin, is a purified murine monoclonal antibody directed specifically against the CD3 antigen. This monoclonal antibody is one of a series of monoclonal antibodies (OKT series) directed towards human T cell surface antigens and is the only commercially available anti-CD3 monoclonal antibody.

While muromonab CD3-based immunosuppression is widely accepted as a treatment for acute allograft rejection episodes and as 'rescue' therapy for steroid-resistant allograft rejection,<sup>[1]</sup> the prophylactic use of muromonab CD3 is less well established. Issues central to the prophylactic use of this agent are whether muromonab CD3 is more effective than standard 'triple therapy' (prednisone, azathioprine plus cyclosporin) and whether muromonab CD3 should be used to prevent rejection or be reserved for the treatment of rejection episodes.

The rationale for using muromonab CD3-based therapy (section 3) in the prevention of rejection is to optimise early graft function by delaying administration of potentially toxic cyclosporin until organ function is established and to improve graft

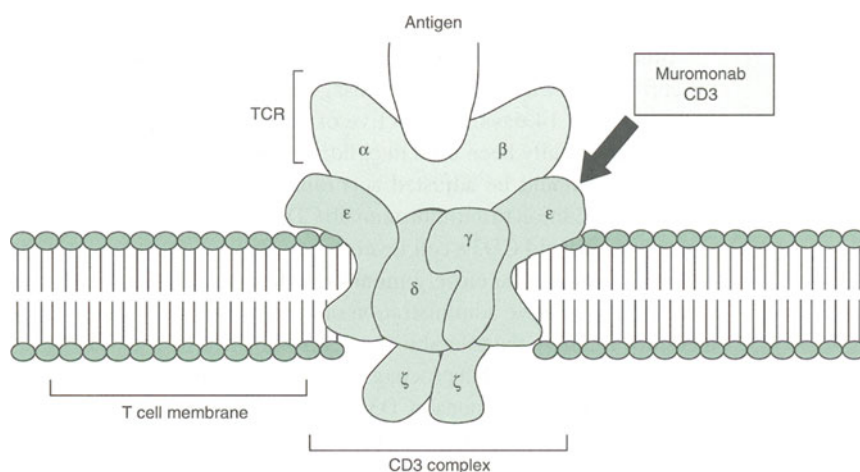
and patient survival by reducing the incidence of early rejection episodes.

The therapeutic potential of muromonab CD3 in both the treatment and prevention of rejection episodes has been previously reviewed.<sup>[1]</sup> This review provides a reappraisal of muromonab CD3 in the prevention of renal, hepatic and cardiac transplant rejection as well as in dual (renal/pancreas) solid organ transplant rejection. The efficacy of prophylactic muromonab CD3-based immunosuppression in patients undergoing cardiac/lung transplantation or nonsolid organ transplantation (e.g. bone marrow transplantation) is not addressed in this review.

## 1. Pharmacodynamic Properties

### 1.1 Mechanism of Action

Muromonab CD3 is directed specifically against the CD3 antigen which is found on mature peripheral human T cells (and medullary thymocytes) and is linked to the T cell antigen receptor (TCR) [fig. 1]<sup>[1-5]</sup> In contrast, polyclonal antilymphocyte antibody preparations are directed against more than one T cell epitope.<sup>[3]</sup> In effect, muromonab CD3 blocks all T cell function and is, there-



**Fig. 1.** Diagrammatic representation of the CD3 complex, T cell antigen receptor (TCR) and site of muromonab CD3 binding (epsilon chain of the CD3 complex).

**Table I.** Known and proposed mechanisms of action of muromonab CD3

Known mechanisms	Proposed mechanisms
<ul style="list-style-type: none"> <li>• Antigenic modulation of the CD3/T cell receptor complex on peripheral T cells resulting in failure of antigen recognition<sup>[1-3,5]</sup></li> <li>• Mediates the opsonisation of circulating T cells and subsequent removal by the reticuloendothelial system<sup>[1-3,5]</sup></li> </ul>	<ul style="list-style-type: none"> <li>• Immunomodulation of graft-infiltrating lymphocytes<sup>[6,7]</sup></li> <li>• Elimination of activated CD3+ cells by induction of apoptosis (programmed cell death)<sup>[3,8]</sup></li> <li>• Modulation of CD3 complex density by shedding CD3 antigens or the whole CD3 complex<sup>[9]</sup></li> <li>• Increasing lymphocyte adhesion molecule expression on peripheral blood lymphocytes resulting in increased adhesion of lymphocytes to vascular endothelium<sup>[10,11]</sup></li> <li>• Induction of cell-mediated cytotoxicity<sup>[12]</sup></li> </ul>

fore, classified as a pan-T cell suppressive monoclonal antibody.

The immunological mechanisms by which muromonab CD3 produces its effects are not completely understood. Known and proposed mechanisms of action are summarised in table I.

Although CD3+ cells are cleared from the circulation within minutes of the first dose of muromonab CD3 (CD2+, CD4+ and CD8+ are also depleted),<sup>[1,5]</sup> CD3+ cells may reappear during continued once-daily administration (usually as a result of antimuromonab CD3 antibody formation; section 1.3). Furthermore, rejection has been observed despite low circulating CD3+ cell levels.<sup>[13]</sup> These findings support the notion that mechanisms other than CD3+ cell depletion and CD3/T cell antigen receptor modulation are involved in the immunological activity of muromonab CD3.

Modulated cells rapidly re-express the CD3+ marker after withdrawal of muromonab CD3 with CD3+ cell levels returning to baseline within 1 week.<sup>[1,5]</sup>

No studies have directly compared the degree of immunosuppression with muromonab CD3 and other agents.

## 1.2 T Cell-Activating Properties

Cytokines including tumour necrosis factor (TNF) alpha, interleukins (IL) 2, 3, 6 and 10 and interferon gamma are released after administration of muromonab CD3. This is associated with an acute phase reaction involving C-reactive protein, neopterin, endothelin-1, complement, transferrin, alpha-1 proteinase inhibitor and neutrophilic gran-

ulocytes.<sup>[1,14-20]</sup> Although this response usually occurs after the first and possibly second and third dose(s), a similar response (particularly IL-6 release<sup>[21]</sup>) may also occur later in the course of treatment if CD3+ cell levels substantially increase (sections 1.3 and 2); this may account for some of the late adverse events observed with muromonab CD3 (section 5.1.1). The T cell-activating properties of muromonab CD3 manifest clinically as first-dose adverse events ('cytokine-release syndrome') [section 5.1].

Monocyte-dependent Fc receptor-mediated cell activation appears to be the main mechanism underlying these events.<sup>[15,16,22-24]</sup>

The release of IL-6 and IL-10 may be involved in the pathogenesis of Epstein-Barr virus (EBV)-associated lymphoproliferative disorders in transplant recipients receiving muromonab CD3 (section 5.3).<sup>[25,26]</sup> Anti-CD3 monoclonal antibodies have been shown to trigger T cell mitogenesis *in vitro*.<sup>[18]</sup>

Several agents including corticosteroids, pentoxifylline and IL-10 can reduce muromonab CD3-induced cytokine release.<sup>[27-32]</sup> However, although corticosteroids can reduce first-dose cytokine-related adverse events (see table V), pretreatment with pentoxifylline does not appear to reduce the cytokine-release syndrome associated with prophylactic muromonab CD3.<sup>[33,34]</sup>

## 1.3 Immunogenicity

Despite improvements in muromonab CD3 dosage regimens, antibodies to this agent may develop. High antibody titres ( $\geq 1:1000$ ) were de-

tected in 5.8% of 12 133 serum samples from patients who received muromonab CD3 for the treatment or prevention of transplant rejection.<sup>[35]</sup> These antibodies may result in decreased muromonab CD3 plasma concentrations (section 2.2) and increased circulating CD3+ cell levels (section 1.1) and may preclude reuse of the agent in some patients (section 3.6).<sup>[36]</sup>

The 2 types of anti-muromonab CD3 antibodies that may be induced are anti-idiotypic and anti-isotypic antibodies.<sup>[1,18,36]</sup> Anti-idiotypic antibodies compete with muromonab CD3 for binding to the CD3 complex and can neutralise the activity of muromonab CD3; IgG but not IgM anti-muromonab CD3 antibodies are able to reduce the activity of this agent. Although anti-isotypic antibodies bind to the constant portion of the muromonab CD3 antibody molecule, they do not block the effects of the drug. Anti-idiotypic antibodies to muromonab CD3 do not cross-react with murine antibodies of similar or different isotypes and cross-react with only 10% of other anti-CD3 monoclonal antibodies.<sup>[37]</sup>

Although variation in antibody test results between centres is significant, the incidence of anti-muromonab CD3 antibodies and the percentage of high antibody titres ( $\geq 1:1000$ ) appears to be greatest in liver or kidney transplant recipients and least in cardiac transplant recipients.<sup>[35,38-40]</sup> The risk of high anti-muromonab CD3 antibody titres also appears to be greatest in patients aged <30 years, in those who have undergone previous transplantation or muromonab CD3 courses, and in those receiving muromonab CD3 for rescue treatment (versus prophylaxis or first-line treatment of rejection).<sup>[35]</sup>

Administration of concomitant immunosuppressants reduces the likelihood of anti-muromonab CD3 antibody formation.<sup>[18,19,36,41-43]</sup>

#### 1.4 Effects on Rejection Histopathology

Although the microscopic appearance of lymphocytic infiltrates in hepatic transplant recipients experiencing acute allograft rejection was similar with prophylactic muromonab CD3 and conven-

tional prophylaxis with triple therapy,<sup>[44]</sup> infiltrate cellularity was reduced with prophylactic muromonab CD3 in cardiac transplant recipients.<sup>[45]</sup> The latter may partly explain the discrepant results reported by different research groups (section 3).

Prophylactic antithymocyte globulin (ATG) has been associated with less cellular infiltration of allograft biopsies, lymphocyte growth and donor-specific cytolytic activity than prophylactic muromonab CD3 in cardiac transplant recipients.<sup>[46]</sup> However, these findings do not appear to have translated into a consistently demonstrable clinical advantage (section 3). Increased adhesion of peripheral blood mononuclear cells to human arterial endothelial cells with muromonab CD3 but not ATG may partly explain why vascular rejection may occur with muromonab CD3 but not ATG.<sup>[47]</sup>

#### 1.5 Cardiovascular, Pulmonary and Haematological Parameters

Following muromonab CD3 administration, a biphasic reversible haemodynamic response has been observed (involving increases in ventricular ejection fractions and cardiac index and decreases in systemic vascular resistance index within the first 2 hours, then reductions in ejection fractions, cardiac index and right atrial and pulmonary capillary wedge pressures after 5 to 6 hours).<sup>[48]</sup> These changes coincide with increases in TNF levels (section 1.2) and are suggestive of a capillary leak syndrome.

Evidence suggests that the intraoperative administration of muromonab CD3 is associated with fewer cardiovascular and pulmonary disturbances (changes in heart rate, blood pressure, mean pulmonary artery pressure, central venous pressure, pulmonary capillary wedge pressure, pulmonary and systemic vascular resistance, cardiac index and blood gases) than administration in the immediate postoperative period.<sup>[49-51]</sup>

Biphasic activation of coagulation and fibrinolysis also occurs after the first dose of muromonab CD3 and other antilymphocyte antibodies. The initial phase appears to be associated with complement activation and the latter with cytokine release



(section 1.2).<sup>[52-55]</sup> This dual activation may explain the general lack of thromboembolic events with recommended muromonab CD3 dosages (although intragraft thromboses may develop; section 5.1). Pentoxifylline inhibits the *in vitro* procoagulant activity of muromonab CD3.<sup>[56]</sup>

## 2. Pharmacokinetic Properties

Muromonab CD3 is a pure standardised product (in contrast to polyclonal antilymphocyte antibody preparations) and is administered via the intravenous route.<sup>[3]</sup>

No studies have specifically compared the pharmacokinetics of muromonab CD3 when administered intraoperatively or in the immediate post-operative period.

### 2.1 Pharmacokinetic Parameters

As plasma muromonab CD3 concentrations are dependent on the presence and degree of sensitisation (section 1.3) and the number of available CD3 molecules, significant interindividual variation exists.<sup>[57,58]</sup> Following a single dose of muromonab CD3 2.5mg (lower than the recommended dosage; section 7) administered 12 to 24 hours before renal transplantation in 7 patients, a maximum serum concentration ( $C_{max}$ ) of 185  $\mu\text{g/L}$  was reached at 1 hour; serum concentrations decreased rapidly thereafter and were undetectable by 12 hours.<sup>[59]</sup> In 66 renal transplant recipients receiving intravenous muromonab CD3 5mg once daily for 10 to 14 days for the prevention of transplant rejection, mean serum muromonab CD3 concentrations were 996  $\mu\text{g/L}$  after 1 hour and decreased to 104  $\mu\text{g/L}$  at 24 hours.<sup>[60]</sup> Plasma muromonab CD3 concentrations increase gradually over the treatment period and decrease sharply after discontinuation.<sup>[61-63]</sup> Mean trough steady-state serum concentrations range from 500 to 1000  $\mu\text{g/L}$  after 2 to 4 days; approximately 1000  $\mu\text{g/L}$  is required to block cytotoxic T cell function *in vitro*.<sup>[60,64]</sup>

Mean serum muromonab CD3 concentrations were significantly higher on days 1 to 6 in patients receiving the drug as prophylaxis than in those being treated for rejection (greatest difference was on

day 1; 678 vs 333  $\mu\text{g/L}$ ) but significantly lower on days 7 to 14 (greatest difference on day 11; 555 vs 784  $\mu\text{g/L}$ ;  $p < 0.05$ ); steady-state serum muromonab CD3 concentrations were achieved earlier (approximately 1 vs 5 days) with prophylactic administration.<sup>[40]</sup> Most patients in each group received a conventional muromonab CD3 dosage of 5 mg/day, while the remainder received increased dosages (dosage not stated). These findings may be explained by muromonab CD3 binding to CD3+ cells associated with rejection. Mean serum muromonab CD3 concentrations were also higher in women than in men, in hepatic transplant recipients compared with renal or cardiac transplant recipients during the first 7 to 10 days of administration and in renal compared with cardiac transplant recipients during the latter part of treatment.<sup>[40]</sup> They were also higher in those aged <10 years compared with older patients, in those who were antimuromonab CD3 antibody-negative before treatment compared with those who had low titres, and in patients who remained antimuromonab CD3 antibody-negative compared with those who seroconverted.<sup>[40]</sup>

The apparent volume of distribution of muromonab CD3 is approximately 6.5L.<sup>[64]</sup> The route of elimination of muromonab CD3 is via binding to lymphocytes and subsequent removal by the reticuloendothelial system.<sup>[40]</sup> There is an initial rapid clearance of the drug (serum concentrations decreased by approximately 60% over the first 4 hours) and unmeasurably low concentrations are reached by 12 hours.<sup>[59,64]</sup> Muromonab CD3 plasma elimination half-lives ( $t_{1/2}$ ) of approximately 18 hours (following administration for the treatment of rejection)<sup>[64]</sup> and 36 hours (with prophylactic administration)<sup>[65]</sup> have been reported; elimination is more rapid in the presence of antimuromonab CD3 antibodies.

### 2.2 Correlation Between Plasma Concentrations and Efficacy

Low plasma muromonab CD3 concentrations have been associated with failure of muromonab CD3 prophylaxis.<sup>[66-68]</sup> The CD3+ cell level alone

is not a reliable indicator of plasma muromonab CD3 concentrations or early sensitisation;<sup>[69-72]</sup> therefore, achieving optimal dosage adjustments and efficacy requires continued surveillance for clinical signs of rejection during muromonab CD3 prophylaxis in addition to monitoring serum muromonab CD3 concentrations, CD3+ cell levels and antimuromonab CD3 antibody titres, as appropriate.<sup>[58,67,68,70,73-78]</sup>

### 3. Therapeutic Use

The end-points of immunosuppressive trials, particularly those involving prophylactic regimens, should include rejection incidence and severity, time to first rejection episode and effects on organ function as well as graft and patient survival. Randomised, double-blind trials are necessary to determine significant between-treatment differences. In this section, the efficacy of prophylactic muromonab CD3-based regimens in terms of these end-points has been investigated in renal, hepatic and cardiac transplant recipients.

Two types of 'induction' therapy for the prevention of rejection are used in transplantation: administration of an antilymphocyte preparation, azathioprine and corticosteroids from the time of transplantation with cyclosporin withheld until renal function is established (sequential therapy); and administration of an antilymphocyte preparation, azathioprine, corticosteroids and low-dose cyclosporin from the time of transplantation. Most trials in this review involved muromonab CD3 administered as part of a sequential regimen followed by maintenance therapy. Muromonab CD3 was usually administered during the immediate post-operative period although some patients received the drug intraoperatively.

Prophylactic muromonab CD3-based immunosuppression has been compared with standard triple therapy (prednisone, azathioprine plus cyclosporin) and with quadruple regimens containing other antilymphocyte preparations (polyclonal preparations) or anti-IL-2 receptor antibodies. The most commonly used polyclonal preparations were antilymphocyte globulin (ALG) and ATG.

Most assessed trials were randomised, but the majority were not double-blind (the first-dose reaction to muromonab CD3 and the different modes of administration of antilymphocyte antibodies precluded double-blinding) and involved small numbers of patients. Furthermore, dosage regimens of concomitant immunosuppressants were varied and complex and administration of muromonab CD3 was preceded by a variety of prophylactic agents (for the prevention of first-dose effects and infections), making between-study comparisons difficult. Most recipients were undergoing cadaveric transplantation for the first time and some underwent dual transplantation. Rejection episodes were documented histologically or with appropriate laboratory tests of organ function.

Factors which may influence the outcome of rejection prophylaxis and should be controlled for in clinical trials include:

- primary disease and severity including baseline organ function;
- age and sex of donor and recipient<sup>[79]</sup>;
- pre-existing antimuromonab CD3 antibody titres (section 1.3);
- delayed graft function<sup>[80,81]</sup>;
- ABO blood group compatibility;
- previous transplantation or blood transfusions;
- donor/recipient cytomegalovirus (CMV) status;
- histocompatibility antigen matching and preformed reactive anti-HLA antibodies, although the potential beneficial effects of HLA matching, especially in those receiving antilymphocyte agents, are controversial<sup>[82,83]</sup>;
- duration of organ cold ischaemia time;
- surgical technique;
- postoperative patient management including treatment of rejection.

Most muromonab CD3 trials selected patients according to primary disease and severity, controlled for ABO blood group compatibility, duration of organ cold ischaemia time, age and sex of donor/recipient, histocompatibility antigen matching and previous transplantation, and used standardised surgical techniques and postoperative patient management. Fewer controlled for donor/recipient

CMV status, delayed graft function or pre-existing antimuromonab CD3 antibody titres. However, clinical judgement should be exercised, as optimal matches are not always possible with the shortage of suitable organs and the usual urgency of transplantation.

### 3.1 Optimal Dosage

The currently recommended dosage of muromonab CD3 for the prevention of rejection is discussed in section 7.

Lower initial dosages (<5 mg/day) may be effective and are better tolerated and less costly than higher dosages of muromonab CD3.<sup>[84,85]</sup> Rejection incidences with low dosage muromonab CD3-based induction therapy in renal transplant recipients were similar to or lower than those with higher dosages.<sup>[84,85]</sup> In cardiac transplant recipients who received muromonab CD3 2.5 or 5mg for 7 days, no significant between-treatment differences were observed; however, rejection incidence tended to be lower and the incidence of adverse events and the number of infectious complications tended to be greater in the 5mg group.<sup>[86]</sup> Whether a low dosage regimen results in a reduction in the incidence of post-transplant lymphoproliferative disease remains to be determined.

High muromonab CD3 dosages (10 mg/day) have been associated with intragraft thromboses (section 5.1). Single high doses (30, 40 or 50mg) provided no efficacy advantages over 5 mg/day and were associated with a high incidence of adverse events.<sup>[87]</sup>

A 14-day course of prophylactic muromonab CD3 in cardiac transplant recipients was associated with a lower incidence of rejection episodes and was more likely to be associated with withdrawal of maintenance corticosteroids than a 10-day course.<sup>[88]</sup> No advantages with shorter-duration (4 to 6 days) muromonab CD3-based quadruple therapy have been reported.<sup>[89]</sup>

According to an abstract report, administration of prophylactic muromonab CD3 via a 2-hour continuous infusion reduced complement activation (section 1.2) and the incidence of adverse events

compared with bolus administration in renal transplant recipients.<sup>[90]</sup> However, administration of muromonab CD3 via continuous infusion is not recommended by the manufacturer.<sup>[91]</sup>

### 3.2 Renal Transplant Recipients

Studies comparing prophylactic muromonab CD3-based sequential immunosuppression with triple therapy or regimens containing other anti-lymphocyte preparations, published since the previous review,<sup>[1]</sup> are summarised in table II.

#### 3.2.1 Comparisons with Triple Therapy

Prophylactic muromonab CD3-based quadruple therapy (with delayed cyclosporin) was significantly more effective than standard triple therapy in terms of severity and/or incidence of rejection episodes (rejection incidence approximately 44 to 68% vs 66 to 78%)<sup>[13,93,94]</sup> and time to first rejection episode.<sup>[13,94]</sup> These benefits appeared to be maintained in the long term (for up to 3 years) and were noted irrespective of antiglobulin crossmatch (AGXM) status.<sup>[93]</sup> In patients with delayed graft function, overall graft survival was significantly increased by muromonab CD3-based therapy compared with triple therapy (the duration of graft nonfunction was decreased with muromonab CD3-based therapy).<sup>[92]</sup> Three-year graft survival rates in renal transplant recipients of first transplants and in those receiving retransplants were significantly greater with sequential muromonab CD3-based induction therapy (with delayed cyclosporin) than with cyclosporin-based prophylactic regimens not containing muromonab CD3 (75 vs 71% and 68 vs 62%, respectively;  $p < 0.001$ ); corresponding survival rates were not greater with simultaneous muromonab CD3 and cyclosporin administration.<sup>[100]</sup> There were no significant between-treatment differences in patient survival (table II).

Compared with triple therapy, the muromonab CD3-containing regimen was associated with greater graft survival rates in patients with 2 mismatches at the HLA DR loci (86 vs 61% at 2 years; total  $n = 51$ ;  $p = 0.04$ ) and in those whose graft had

**Table II.** Efficacy of muromonab CD3 (MCD3)-based immunosuppression (delayed cyclosporin) as prophylaxis of renal allograft rejection versus standard triple therapy (Tr) [immediate cyclosporin] or other antilymphocyte-based regimens

Reference	Study design	No. of evaluable patients (characteristics)	Treatment	Time to initial rejection (days)	Actuarial graft survival (%)	Rejection incidence (%)	Actuarial patient survival (%)	Overall efficacy	Comments
<b>Muromonab CD3-based immunosuppression versus triple therapy</b>									
Abramowicz et al. <sup>[13]</sup>	r	56	MCD3 <sup>ab</sup>	23 (during first 3mo)*	83 (overall; 3y) <sup>c</sup> (92 <sup>nd</sup> )	4 <sup>e</sup> ; 32 (3y) <sup>f</sup>	94.5 (3y)	MCD3 ≥ Tr	Fewer rejections with MCD3 were corticosteroid-resistant (15 vs 30%*)
Benvenisty et al. <sup>[92]</sup>	nb	52	Tr <sup>g</sup>	11 (during first 3mo)	75 (overall; 3y) <sup>c</sup> (79 <sup>th</sup> )	6 <sup>e</sup> ; 22 (3y) <sup>f</sup>	93 (3y)		
		34 (delayed graft function)	MCD3 <sup>a</sup>		80 (1y)*; 74 (2y)*	44*	89	MCD3 ≥ Tr	MCD3 decreased the duration of graft nonfunction (9.4 vs 14.9 days)
		40 (delayed graft function)	Tr <sup>g</sup>		55 (1y); 47 (2y)	82	89		
Dafoe et al. <sup>[93]</sup>	nb	38 (positive AGXM)	MCD3 <sup>a</sup>	35	88*	44 <sup>h</sup>	100	MCD3 ≥ Tr	AGXM-positive patients: incidence of delayed graft function greater with MCD3 (75 vs 40%*)
		10 (positive AGXM)	Tr <sup>g</sup>	13	50	70	100		
		32 (negative AGXM)	MCD3 <sup>a</sup>	22	72	44	97		
		32 (negative AGXM)	Tr <sup>g</sup>	13	92*	73*	100		
Norman et al. <sup>[94]</sup>	r, mc	105	MCD3 <sup>a</sup>	45*	84 (2y); 73 (5y)	51 <sup>ah</sup>	95 (2y); 90 (5y)	MCD3 ≥ Tr	
		102	Tr <sup>g</sup>	8	75 (2y); 64 (5y)	66 <sup>h</sup>	94 (2y); 88 (5y)		
<b>Muromonab CD3-based immunosuppression versus other antilymphocyte-based regimens</b>									
Bock et al. <sup>[66]</sup>	r, nb	51	MCD3 <sup>a</sup>		78 (1y)	45*	92 (1y)	ATG > MCD3	
		53	ATG <sup>i</sup>		91 (1y)*	26	96 (1y)		
Broyer et al. <sup>[95]</sup>	r	77	MCD3 <sup>ab</sup>		79 (1y); 71 (2y); 68 (3y)	11	96	MCD3 = ALG	
		71	ALG <sup>bj</sup>		80 (1y); 77 (2y); 73 (3y)	11	99		
Cole et al. <sup>[96]</sup>	r	83	MCD3 <sup>a</sup>	ATG > MCD3 (1y)*	81 (1y)	31 <sup>k</sup>	95 (1y)	ATG ≥ MCD3	More steroid-resistant rejections with MCD3 (25 vs 12*)
		83	ATG <sup>i</sup>		78 (1y)	57 <sup>ka</sup>	89.5 (1y)		
Frey et al. <sup>[97]</sup>	r	67	MCD3 <sup>ab</sup>	MCD3 = ALG	87 (1y); 83 (2y) <sup>l</sup>	36	96 (1y); 96 (2y) <sup>l</sup>	MCD3 = ALG	
		71	ALG <sup>i</sup>		84 (1y); 80 (2y) <sup>l</sup>	45	93 (1y); 91 (2y) <sup>l</sup>		
Hanto et al. <sup>[98]</sup>	r	59	MCD3 <sup>ab</sup>	35	84 (1y); 79 (2y); 79 (3y)	33 (37)	98 (1, 2 and 3y)	MCD3 = ALG	
		58	ALG <sup>i</sup>	29	81 (1y); 78 (2y); 78 (3y)	27 (31)	96 (1, 2 and 3y)		

Steinmuller et al. <sup>[99]</sup>	r	25	MCD3 <sup>a</sup>	67.5	84 (<6mo)	64	99 (<6mo)	MCD3 ≡ ALG
a		26	ALG <sup>l</sup>	95	85 (<6mo)	35	99 (<6mo)	
b								
c								
d								
e								
f								
g								
h								
i								
j								
k								

MCD3-based regimen: MCD3 5-10 mg/day or 0.05 or 0.1 mg/kg/day IV (7-21 PODs) + Az 1-10 mg/kg/day IV (POD 0-10) + MPr 1-8 mg/kg (250mg-2g) IV (preoperatively) + Pr 0.15-1.5 mg/kg/day, 30-60 mg/m<sup>2</sup> or 30 mg/day + Cs 4-14 mg/kg/day or 400 or 600 mg/day or 150 mg/ml/day PO (starting on POD 4-11).

First antilymphocyte antibody dose administered intraoperatively.

Total graft survival.

Immunological graft survival rate (only losses from rejection were considered).

Number of rejections per patient-month.

Percentage of patients free from rejection.

Triple therapy: Cs 6-14 mg/kg/day PO from POD 1 + Az 1-10 mg/kg/day IV + MPr 1-1.5 mg/kg (375mg-2g) + Pr 0.17-1.5 mg/kg/day (30 mg/day).

The incidence of rejection after the first 2wk was equivalent.

ATG-based regimen: ATG 0.15 mg/kg/day or 4 mg/kg/day (7-14 days) + MPr 1 mg/kg or 1g then 0.25 mg/kg 6-hourly (48h) then Pr 0.25-0.5 mg/kg/day + Az 1 mg/kg IV (preoperatively) then 1 or 2 mg/kg/day + Cs 4 mg/kg/day or 400-600 mg/day (from POD 4 to 7).

ALG-based regimen: ALG 1-20 mg/kg/day IV (7-21 days) + MPr 7 mg/kg then Pr 30 mg/day or 0.3 to 1 mg/kg/day + Az 1-5 mg/kg/day + Cs 5-8 mg/kg/day (started when renal function normal).

Patients with no rejection episodes. The rate of first rejection with MCD3 was 1.8 times higher than with ALG; overall 50 rejection episodes occurred with ALG and 83 with MCD3.

Graft and patients survival rates included data from 35 patients who received kidney-pancreas transplant.

Abbreviations and symbols: AGXM = antiglobulin crossmatch; ALG = antilymphocyte globulin (Minnesota; horse antibody); ATG = antithymocyte globulin; Az = azathioprine; Cs = cyclosporin; IV = intravenous; mc = multicentre; MPr = methylprednisolone; nb = nonblind; PO = orally; POD = postoperative day; Pr = prednisone; r = randomised; ≡ indicates equivalence; > indicates significantly (p < 0.05) more effective than comparator; ≥ indicates significantly (p < 0.05) greater efficacy than comparator in terms of at least 1 efficacy parameter. \* p < 0.05 compared with comparator.

a cold ischaemia time of >24 hours (84 vs 62% at 2 years; n = 98; p = 0.01).<sup>[94]</sup>

Despite delayed cyclosporin administration, significantly more patients receiving muromonab CD3-based therapy required postoperative dialysis than triple therapy recipients;<sup>[13,93]</sup> this was possibly a result of muromonab CD3-induced lymphokine release (section 1.2). Mean serum creatinine levels were similar in the 2 groups.

No studies directly compared the efficacy of intraoperative administration of the first dose of muromonab CD3 with administration in the immediate postoperative period.

### 3.2.2 Comparisons with Other Antilymphocyte-based Regimens

Muromonab CD3 and ALG or ATG were similar in terms of rejection incidence and/or time to first rejection (for up to 3 years' follow-up) in some studies;<sup>[95,97-99]</sup> others show ALG or ATG to be superior (1 year's follow-up) [table II].<sup>[65,96]</sup>

Although most studies showed no significant between-treatment differences in graft and patient survival, the 1-year graft survival rate was significantly greater with ATG than with muromonab CD3 in 1 study.<sup>[65]</sup> The incidence of delayed graft function was similar.<sup>[65,97]</sup>

One-year graft survival in highly sensitised (panel reactive antibodies > 50%) renal transplant recipients was similar with prophylactic muromonab CD3- and ALG-based regimens.<sup>[101]</sup>

In combined renal-pancreas transplant recipients (total of 220 patients) who received muromonab CD3- or ALG/ATG-based quadruple induction therapy, 12- or 15-month actuarial graft survival rates were similar (80 to 91 vs 88 to 96% for renal grafts and 60 to 88.5 vs 73 to 96% for pancreas grafts) as were patient survival rates (80 to 96 vs 89 to 100%).<sup>[102-104]</sup> The incidence of rejection was either similar (40 to 50 for both treatment groups;<sup>[102,104]</sup> mean number of rejection episodes per patient 1.4 vs 0.9<sup>[103]</sup>) or less (mean number of rejection episodes per patient 1.5 vs 2.7; p value not given) with muromonab CD3.<sup>[105]</sup> No studies comparing muromonab CD3-based induc-

**Table III.** Efficacy of muromonab CD3 (MCD3)-based immunosuppression (delayed cyclosporin) as prophylaxis of hepatic allograft rejection versus standard triple therapy (Tr) [immediate cyclosporin], other antilymphocyte-based regimens or anti-interleukin-2 monoclonal antibody-based immunosuppression

Reference	Study design	No. of evaluable patients	Treatment	Time to initial rejection (days)	Actuarial graft survival (%)	Rejection incidence (%)	Actuarial patient survival (%)	Overall efficacy	Comments
<b>Muromonab CD3-based immunosuppression versus triple therapy</b>									
Cosimi et al. <sup>[106]</sup>	r	38	MCD3 <sup>a</sup>		13* (1wk), 39* (2wk), 68 (1y)	84	MCD3 ≥ Tr		Duration of the initial rejection-free period was longer with MCD3
Farges et al. <sup>[107]</sup>	r, mc	41 44	Tr <sup>b</sup> MCD3 <sup>a</sup>	38*	61 (4y)	46(1wk), 71 (2wk), 78 (1y) 34* (2wk); 67 (1y)	73 82 (1y); 69 (4y)	MCD3 ≥ Tr	
McDiarmid et al. <sup>[108,109]</sup>	r	50 46	Tr <sup>b</sup> MCD3 <sup>a,c</sup>	9 12 <sup>b</sup>	54 (4y) 63 (>3mo)	61 (2wk); 75 (1y) 28 (POD 0-14) <sup>d</sup> 46 (<1mo) <sup>e</sup> 91 (>3mo) <sup>e</sup>	78 (1y); 62 (4y) 67 (>3mo)	MCD3 ≥ Tr	Mean duration of follow-up was 648 days for MCD3 and 682 days for Tr
Mühlbacher et al. <sup>[110]</sup>		39	Tr <sup>b</sup>	9.5 <sup>c</sup>	73 (>3mo)	67 (POD 0-14) <sup>d</sup> 31 (rejection free; <1mo) 99 (rejection free; >3mo) 56*	84 (>3mo)	MCD3 > Tr	
Pons et al. <sup>[111]</sup>		58 25 19	Tr <sup>b</sup> MCD3 <sup>a</sup> Tr <sup>b</sup>			79.5 25* 69	45 (1y) 3.5 (2mo) <sup>f</sup> 26 (2mo)		ATG > Cs + Pr and MCD3 (acute rejection) severe rejection was considered
<b>Muromonab CD3-based immunosuppression versus other antilymphocyte-based regimens</b>									
Steininger et al. <sup>[112]</sup>	r	63	Cs + Pr <sup>b</sup>		67 (during first 3wk)				
<b>Muromonab CD3-based immunosuppression versus anti-interleukin-2 receptor monoclonal antibody-based regimen</b>									
Reding et al. <sup>[113]</sup>	r	37 (adults and children)	MCD3 <sup>a</sup>		86 (1y)	81 (first 3mo)	86 (1y)		
		35 28	AIL <sup>h</sup> Tr <sup>b</sup>		97 (1y)* 75 (1y)	91 (first 3mo) 96 (first 3mo)	100 (1y)* 79 (1y)		
<p>a MCD3-based regimen: MCD3 2.5-20 mg/day IV (10-14 days) + Az 1-5 mg/kg/day or 100 mg/day IV + MPr 0.3-5 mg/kg IV then Pr 0.3-2.5 mg/kg/day or 30-200 mg/day + Cs 3-15 mg/kg/day IV (starting on POD 7-11)</p> <p>b Triple/double therapy: Cs 1-4 mg/kg/day IV ± Cs 8-14 mg/kg/day PO ± Az 1-5 mg/kg/day or 100 mg/day IV + MPr 0.3-5 mg/kg (200mg) then Pr 0.3-2.5 mg/kg/day (20-200mg).</p> <p>c First antilymphocyte antibody dose administered intraoperatively.</p> <p>d Based on the results of an initial study involving 25 muromonab CD3 and 27 triple therapy recipients.<sup>[114]</sup></p> <p>e Patients who were rejection free.</p> <p>f Mortality rate.</p> <p>g ATG-based regimen: ATG 2.5 mg/kg/day IV (10 days) + Pr (dosage not given) + Cs (dosage not given) PO (from POD 8).</p> <p>h AIL-based regimen: AIL 10 or 20 mg/day IV (intraoperatively and for 9 PODs) + Cs 3-5 mg/kg/day IV + MPr 1 g/1.73m<sup>2</sup> (children) or 1 g/70kg (adults) IV (starting intraoperatively) then Pr 1 mg/kg/day (children) or 25 mg/day (adults) then 0.5 mg/kg/day (children) or 15 mg/kg (adults) + Az 1.5 mg/kg/day (children) or 1 mg/kg/day (adults).</p> <p>Abbreviations and symbols: AIL = anti-interleukin-2 receptor monoclonal antibody; ATG = antithymocyte globulin; Az = azathioprine; Cs = cyclosporin; IV = intravenous; mc = multicentre; MPr = methylprednisolone; PO = oral; POD = postoperative day; Pr = prednisone; r = randomised; tr = retrospective study; &gt; significantly (p &lt; 0.05) greater efficacy than comparator; ≥ significantly (p &lt; 0.05) greater efficacy in terms of at least one efficacy parameter; * p &lt; 0.05 vs comparator.</p>									

tion therapy with triple therapy in this patient group are available.

### 3.3 Hepatic Transplant Recipients

In addition to the toxic effects of early cyclosporin use, hepatic transplant recipients are at an increased risk of renal impairment in the early postoperative period because of intraoperative haemodynamic instability, hepatorenal syndrome and pre-existing renal or hepatic dysfunction. Studies comparing prophylactic muromonab CD3-based sequential immunosuppression with triple therapy or regimens containing other antilymphocyte preparations or an anti-IL-2 antibody are summarised in table III.

Muromonab CD3-based sequential therapy reduced the incidence and/or severity of early (but not long term) acute rejection,<sup>[106-111]</sup> delayed the time to first rejection<sup>[107]</sup> and improved/maintained renal function or reduced renal dysfunction in the early postoperative period<sup>[106-108,111]</sup> compared with triple therapy. Plasma bilirubin levels were either similar or lower with muromonab CD3-based therapy.<sup>[110,111]</sup> Most studies reported no between-treatment differences in graft or patient survival.

In the one available study, ATG-based prophylaxis achieved a lower incidence of rejection episodes than muromonab CD3-based therapy (table III).<sup>[112]</sup> An anti-IL-2-based prophylactic regimen improved graft and patient survival compared with muromonab CD3-based prophylaxis;<sup>[113]</sup> these data need to be confirmed.

There have been reports of accelerated rejection leading to graft loss<sup>[115]</sup> in hepatic transplant recipients receiving muromonab CD3-based immunoprophylaxis with no evidence of antimuromonab CD3 antibodies in the majority of patients. Prior sensitisation to HLA or other cell surface antigens has been proposed as an explanation.<sup>[115]</sup>

Some investigators have shown that transplantation with ABO-incompatible hepatic grafts can be successful with the administration of prophylactic muromonab CD3 as part of triple or quadruple immunosuppressive regimens and lowering of pre-

formed antibody titres by plasmapheresis or exchange transfusion.<sup>[116,117]</sup>

A prophylactic muromonab CD3-based regimen including alprostadil (prostaglandin E<sub>1</sub>) [10 to 60 µg/h] has shown promise in hepatic transplant recipients.<sup>[118]</sup> No studies to date have compared this regimen with muromonab CD3-based regimens not containing alprostadil.

### 3.4 Cardiac Transplant Recipients

Prophylaxis of rejection is particularly important for cardiac transplant recipients because, unlike renal transplant failure, cardiac graft failure is usually fatal. Studies comparing prophylactic muromonab CD3-based sequential immunosuppression with triple therapy or regimens containing other antilymphocyte preparations are summarised in table IV.

Although direct comparisons are few, on available evidence muromonab CD3-based prophylaxis appears to be similar to cyclosporin plus prednisone<sup>[119]</sup> or triple therapy<sup>[121]</sup> as assessed by rejection incidence, time to first rejection episode and graft and patient survival. Time to rejection was significantly longer with muromonab CD3-based induction therapy than triple therapy in 1 study; however, there was no significant between-treatment difference in rejection incidence.<sup>[120]</sup>

Data on muromonab CD3-based therapy compared with other antilymphocyte-based regimens are conflicting; some investigators report a longer time to first rejection and/or a lower incidence of rejection with muromonab CD3;<sup>[122,129]</sup> others observe no differences<sup>[125,127,128,130]</sup> or a lower incidence of rejection<sup>[123,126]</sup> and/or a longer time to first rejection episode<sup>[123,124]</sup> with ATG-based therapy. No between-treatment differences in graft or patient survival have been reported.

Histological evidence of rejection in heart transplant recipients was observed in more BT563 (an anti-IL-2 receptor monoclonal antibody; n = 22) than muromonab CD3 recipients (n = 11) after 1 week of prophylactic therapy (50 vs 9%); however, the incidence of adverse events related to cytokine release was lower with BT563.<sup>[131]</sup>

**Table IV.** Efficacy of muromonab CD3 (MCD3)-based immunosuppression (delayed cyclosporin) as prophylaxis of cardiac allograft rejection versus standard triple therapy (Tr) (immediate cyclosporin) or other antilymphocyte-based regimens

Reference	Study design	No. of evaluable patients	Treatment	Time to initial rejection (days)	Actuarial graft survival (%)	Rejection incidence (%)	Actuarial patient survival (%)	Overall efficacy
<b>Muromonab CD3-based immunosuppression versus triple therapy</b>								
Balk et al. <sup>[119]</sup>	r	33	MCD3 <sup>a</sup>		91	1.33 (1y) <sup>b</sup> 80 (1mo) <sup>c</sup> 31 (3mo) <sup>c</sup> 28 (6mo) <sup>c</sup>	91 (2y)	MCD3 = Cs + Pr
Barr et al. <sup>[120]</sup>	rt	33	Cs + Pr <sup>d</sup>		94	1.36 (1y) <sup>b</sup> 66 (1mo) <sup>c</sup> 33 (3mo) <sup>c</sup> 27 (6mo) <sup>c</sup>	94	
		26	MCD3 <sup>a</sup>	42*		0.003 <sup>e</sup>	88 (6mo) 81 (18mo)	MCD3 = Tr
		26	Tr <sup>f</sup>	21		0.003 <sup>e</sup>	92 (6mo) 87 (18mo)	
Stapleton et al. <sup>[121]</sup>	rt, nb <sup>g</sup>	8	MCD3 <sup>a</sup>	66		66 (6mo) <sup>c</sup>	100 (1y)	MCD3 = Tr
		33	Tr <sup>f</sup>	57		75 (6mo)	91 (1y)	
<b>Muromonab CD3-based immunosuppression versus other antilymphocyte-based regimens</b>								
Costanzo-Nordin et al. <sup>[122]</sup>	r	12	MCD3 <sup>a</sup>	32*		3.4		MCD3 ≥ ATG
Griffith et al. <sup>[123]</sup>	r	11	ATG <sup>h</sup>	15		5		MCD3 ≤ ATG
		43	MCD3 <sup>a</sup>	33*		0.58/pt* (1mo); 0.43 (2mo); 0.2 (>2mo)	98 (1y)	
		39	ATG <sup>h</sup>	67		0.08/pt (1mo); 0.24 (2mo); 0.2 (>2mo)	95 (1y)	
Ippoliti et al. <sup>[124]</sup>	r	15	MCD3 <sup>a</sup>	24*			80 (6mo)	ATG ≥ MCD3
		15	ATG <sup>h</sup>	28			93 (6mo)	
Kirklin et al. <sup>[125]</sup>	nb	15	MCD3 <sup>a</sup>	21		1.3/pt (2mo)	100 (2y)	MCD3 = ATG
		32	ATG <sup>h</sup>	16		1.5/pt (2mo)	100 (2y)	
Ladowski et al. <sup>[126]</sup>	r	30	MCD3 <sup>a</sup>			0.8/pt (3mo); 1.03/pt (1y)	83 (1y)	ATG ≥ ALG and MCD3
		34	ATG <sup>h</sup>			0.24/pt (3mo)*; 0.26/pt (1y)*	82 (1y)	
		15	ALG <sup>i</sup>			1.14/pt (3mo); 1.27/pt (1y)	80 (1y)	
MacDonald et al. <sup>[127]</sup>	r	20	MCD3 <sup>a</sup>	33		90	83 (1y)	MCD3 = ATG
		21	ATG <sup>h</sup>	27		100	81 (1y)	
Menkis et al. <sup>[128]</sup>	r	20	MCD3 <sup>a</sup>	5.6wk		2.1/pt (6mo)	92 (2y)	MCD3 = ALG
		19	ALG <sup>i</sup>	5.3wk		1.4/pt (6mo)	84 (2y)	



Renlund et al. <sup>[129]</sup>	r <sup>i</sup>	26	MCD3 <sup>a</sup>	76*	1.5/pt (6mo)* 0.2 (1y)*	96 (1y)	MCD3 ≥ ATG
		25	ATG <sup>h</sup>	36	2.2/pt (6mo) 0.8 (1y)	96 (1y)	
Wollenek et al. <sup>[130]</sup>	r		MCD3 <sup>a</sup> ATG <sup>h</sup>		0.5 (1mo); 0.97 (3mo) <sup>k</sup> 0.32 (1mo); 0.53 (3mo) <sup>k</sup>	82 83	MCD3 = ATG
<p>a MCD3-based regimen: MCD3 5-10 mg/day (7-14 days) IV + Az 2-4 mg/kg/day or 50 mg/day IV or PO + MPi 0.2-5 mg/kg IV (375-500mg) then Pr 0.1-1.5 mg/kg/day or 10-60 mg/day + Cs 4-10 mg/kg/day IV (starting on POD 4-11).</p> <p>b Mean number of acute rejections per patient.</p> <p>c Freedom from rejection.</p> <p>d Cs 1-3 mg/kg/day IV (initiated 2-4h preoperatively) then PO + Pr 20mg IV then 10-60 mg/day.</p> <p>e Rejection episodes per patient per month.</p> <p>f Triple therapy: Cs 4-10 mg/kg/day IV + Cs 8-14 mg/kg/day PO + Az 2-4 mg/kg/day IV or PO + MPi 375-500mg then Pr 1 mg/kg/day (10-30mg).</p> <p>g The decision to use muromonab CD3 was based on immediate postoperative renal insufficiency.</p> <p>h ATG-based regimen: ATG 1.5-10 mg/kg/day IV or IM (4-10 days) + MPi 375-1000mg then Pr 0.2-1.5 mg/kg PO + Cs 2-10 mg/kg PO + Az 2-4 mg/kg PO or IV.</p> <p>i ALG-based regimen: ALG 15-20 mg/kg/day IV (7-20 days) + MPi 0.375-1g (0.1-0.2 mg/kg/day) then Pr 30 mg/day or 0.1-0.5 mg/kg/day PO + Az 1-4 mg/kg/day PO or IV + Cs 5-7 mg/kg/day (from POD 4).</p> <p>j According to availability of ATG.</p> <p>k Cumulative incidence of rejection episodes per patient.</p> <p>Abbreviations and symbols: ALG = antilymphocyte globulin; ATG = antithymocyte globulin; Az = azathioprine; Cs = cyclosporin; IM = intramuscular; IV = intravenous; MPi = methylprednisolone; nb = nonblind; PO = oral; POD = postoperative days; Pr = prednisone; pt = patient; r = randomised; rt = retrospective; ≥ significantly (p &lt; 0.05) greater efficacy in terms of at least one parameter; ≤ significantly (p &lt; 0.05) less efficacy in terms of at least one parameter; ≡ indicates equivalence; * p &lt; 0.05 vs comparator.</p>							

According to a meta-analysis of 28 trials of prophylactic muromonab CD3-based therapy in renal, liver and cardiac transplant recipients, muromonab CD3 had no significant effect on patient survival.<sup>[132]</sup> However, the incidence of acute rejection with muromonab CD3 was lower than with triple therapy (11 studies), horse ATG (4 studies) and horse ALG (5 studies) but higher than with rabbit ATG (4 studies). The results of this meta-analysis are limited by the small number and methodological flaws of the available studies.

### 3.5 Special Patient Groups

#### 3.5.1 High Risk Patients

Patients at high immunological risk of rejection and consequent reduced graft survival include those with delayed graft function, increased panel reactive antibody titres, previous but not current anti-donor lymphocytotoxic antibodies or a positive current crossmatch for donor B lymphocytes, those who rejected an earlier graft within the first 6 to 12 months, patients receiving multiple transplants or blood transfusions or who have had multiple pregnancies, patients receiving grafts with prolonged cold ischaemia times (>24 hours) and paediatric patients (section 3.5.2).<sup>[133-135]</sup>

The efficacy of prophylactic muromonab CD3-based therapy appears to be particularly marked in patients with delayed graft function (section 3.2.1). Patient survival was 100% (after a mean follow-up of 12 or 13 months)<sup>[134,136]</sup> and graft survival (at 1 year) was 70<sup>[134]</sup> or 78%<sup>[136]</sup> with muromonab CD3-based prophylaxis in high risk cadaver renal transplant recipients (previous transplant failure and/or panel reactive antibody titres >50%). These graft survival rates are superior to those reported for a historical group of similar patients who did not receive muromonab CD3 (50%) and are comparable to those of primary graft recipients (table II). Rejection incidence within 1 year (0.87 vs 1.35 mean episodes per patient) and 5 years (1.07 vs 1.49) was significantly lower and graft survival was significantly higher at 2 years (84 vs 64%) and 5 years (71 vs 56%) with prophylactic muromonab CD3-based immunosuppression (n = 78) than with triple

therapy ( $n = 81$ ) in patients with renal graft cold ischaemia times  $>24$  hours; no significant between-treatment differences were observed in patients with cold ischaemia times  $\leq 24$  hours.<sup>[135]</sup>

Three-year graft survival rates in high risk renal transplant recipients (whose highest preformed panel reactive lymphocytotoxic antibody level was  $>50\%$ ) were significantly greater with sequential muromonab CD3-based prophylaxis (delayed cyclosporin) than with simultaneous muromonab CD3 and cyclosporin administration and cyclosporin-based prophylaxis not containing muromonab CD3 (72 vs 53 vs 58%;  $p < 0.01$ ).<sup>[100]</sup> However, an immunosuppressive regimen of simultaneous muromonab CD3 and cyclosporin (initiated from the time of transplantation regardless of graft function) plus azathioprine and prednisone was associated with a single rejection incidence of 50%, acute tubular necrosis in 58% of patients, and no graft losses or deaths in 12 immunological high risk renal transplant recipients followed for 3 to 28 months.<sup>[133]</sup>

### 3.5.2 Young or Elderly

The immune response and the propensity to develop acute rejection appear to be greater in children.<sup>[137]</sup> Furthermore, the incidence of high titre antimuromonab CD3 antibodies (section 1.3) is significantly greater in children (particularly those aged  $<10$  years) than in adults ( $>30$  years),<sup>[35]</sup> and increases in muromonab CD3 dosages are required more often in paediatric than adult patients (section 7).<sup>[40,137]</sup>

Several noncomparative trials involving a total of 118 patients show muromonab CD3 induction therapy prophylaxis, administered as part of a quadruple immunosuppressive protocol, to reduce early rejection in children and adolescents undergoing renal, hepatic or cardiac transplantation.<sup>[138-140]</sup> However, a rejection incidence of 75% during the first 2 months and a graft survival incidence of 50% at 1 and 5 years have been reported in 20 paediatric renal transplant recipients receiving muromonab CD3 induction therapy.<sup>[141]</sup> Furthermore, the rejection incidence with muromonab CD3-based prophylaxis was significantly lower

than with standard triple therapy (25 vs 75%) during the first 14 days in 20 liver transplant recipients; however, there were no significant between-treatment differences in overall rejection incidence or graft or patient survival (mean duration of follow-up was 890 days for the muromonab CD3 group and 721 days for the triple therapy group).<sup>[142]</sup>

Quadruple therapy containing muromonab CD3, prednisone, azathioprine and cyclosporin was used successfully in 17 cadaveric renal transplant recipients aged  $\geq 50$  years; no grafts were lost.<sup>[143]</sup>

Further studies directly comparing the effects of muromonab CD3-based therapy with those of either no prophylaxis or prophylaxis with other agents are needed to determine whether these regimens are beneficial in these patient groups.

### 3.6 Reuse of Muromonab CD3

The presence of antimuromonab CD3 antibodies (section 1.3) has the potential to decrease the efficacy of muromonab CD3 (section 2.2) and to hinder its subsequent use.<sup>[75]</sup>

All patients should be tested for the presence and type (section 1.3) of antimuromonab CD3 antibodies 3 to 4 weeks after initiation of the first dose, particularly if repeated or prolonged muromonab CD3 treatment is required. Muromonab CD3 reuse following prophylactic use is not recommended in patients with antimuromonab CD3 antibody titres  $\geq 1:1000$ , but retreatment may be successful in those with lower titres.<sup>[39,63,144,145]</sup>

In addition to the type and titre of antimuromonab CD3 antibodies,<sup>[36]</sup> factors predictive of success with muromonab CD3 retreatment include longer elapsed time after initial treatment and effective T cell depletion/CD3 modulation (section 1.1).<sup>[146]</sup>

Anti-idiotypic antibodies to muromonab CD3 were detected in 70% of paediatric renal transplant recipients ( $n = 40$ ) who received prophylactic muromonab CD3; antibody titres were not given.<sup>[147]</sup>

#### 4. Pharmacoeconomic Considerations

The overall cost of immunosuppression in transplant recipients is substantial and is highly influenced by the use of biological agents.

No formal cost-effectiveness studies have been conducted to determine whether, and to what extent, potential cost savings from the lower incidence of rejection episodes with prophylactic muromonab CD3 than with triple therapy (section 3) are offset by the increased incidence of infections and other adverse events (section 5); adverse events complicate postoperative care and prolong hospitalisation of patients.<sup>[148,149]</sup> The relative pharmacoeconomic merits of prophylactic muromonab CD3 versus administration for the treatment of rejection have not been determined. Nor have the economic consequences of reduced acute rejection incidence on long term graft survival or subsequent requirement for dialysis, or the economic impact of muromonab CD3 reuse and adverse event preventative strategies (see table V) been determined.

Not unexpectedly, in an Australian cost minimisation study, mean first-year drug costs per renal transplant recipient were greater with a quadruple immunosuppressive regimen containing muromonab CD3, ATG or ALG than with triple therapy alone (\$A10 194 vs \$A7679; 1991 prices).<sup>[150]</sup>

According to another cost minimisation analysis, the total cost of 1 week's muromonab CD3 (as induction therapy) administered to a 70kg patient undergoing renal transplantation would be \$US1500 (1992) more than for ALG.<sup>[97]</sup> However, this increased cost is likely to be balanced by savings resulting from the shorter time spent in hospital by patients receiving muromonab CD3 than those treated with ALG. Similarly, the cost of a 14-day prophylactic course of muromonab CD3 in cardiac transplant recipients was higher than the cost of a 10-day course of ALG or a 4-day course of ATG (\$US6650 vs \$US1914 vs \$US2350); despite different administration durations, the overall efficacy of ALG and muromonab CD3 was similar and slightly inferior to that of ATG (table IV).<sup>[126]</sup>

In an abstract report, the mean costs (definition and year of prices not given) of induction therapy based on muromonab CD3 (n = 102) or ALG (n = 511) were similar (\$US63 416 vs \$US62 176) but significantly greater than those with conventional immunotherapy (\$US47 002; n = 970) in renal transplant recipients.<sup>[151]</sup>

Prospective studies assessing the comparative pharmacoeconomics of these agents in the long term are needed. No formal studies have directly assessed the effects of muromonab CD3 on quality of life in transplant recipients.

#### 5. Tolerability

The general complications of immunosuppressants and the problems associated with the narrow therapeutic range of these agents (infection or malignancy because of over immunosuppression) plus the use of multiple agents complicate the determination of the cause of adverse events in transplant patients receiving muromonab CD3.

Although anaphylaxis has been reported (in 1 patient) following a second course of prophylactic muromonab CD3 (high antimuromonab CD3 titres were present before treatment),<sup>[152]</sup> the tolerability profile of muromonab CD3 during reuse (section 3.6) has not formally been determined. Long term tolerability data are also lacking.

Strategies for preventing drug-related toxicity are of critical importance in the management of transplant recipients. Adverse events with muromonab CD3 may be prevented or minimised by several strategies including the intraoperative administration of the first dose of muromonab CD3, pretreatment with a corticosteroid, administration of an antipyretic and antihistamine, prophylactic use of an antimicrobial(s) and correction of increased temperature and fluid overload before initiation of prophylaxis (table V).

##### 5.1 First-Dose Effects

The first doses of muromonab CD3 are associated with a number of adverse events ('cytokine-release syndrome'; section 1.2) which, although usually mild, may be severe and life-threatening.

**Table V.** Approaches to prevent or decrease adverse events associated with muromonab CD3

Adverse event	Approach
First-dose effects	Intravenous methylprednisolone sodium succinate 8 mg/kg 1-4h before the first dose <sup>[150,154-158]</sup>
	Antipyretics, e.g. paracetamol (acetaminophen) <sup>[150]</sup>
	Antihistamines, e.g. diphenhydramine <sup>[150]</sup>
	Reduction of body temperature to <37.8°C before muromonab CD3 administration
	Intraoperative administration of first muromonab CD3 dose <sup>[19,43,159,160]</sup>
Pulmonary oedema	Agents that neutralise cytokine effects, e.g. anti-tumour necrosis factor <sup>[43,157]</sup>
	Correction of pre-existing fluid overload <sup>[43,158,161]</sup>
	Restriction of bodyweight gain to ≤3% in the 7 days before treatment
Infections	Strict management of fluid, electrolyte and mineral balance especially in patients with diabetes and those with delayed graft function <sup>[43,150,161]</sup>
	Cotrimoxazole (trimethoprim-sulfamethoxazole) [ <i>Pneumocystis carinii</i> pneumonia] <sup>[19,162,163]</sup>
	Ganciclovir, aciclovir and cytomegalovirus hyperimmune globulin (cytomegalovirus infection and other viral infections) <sup>[19,162,164,165]</sup>

ing.<sup>[1,19,154,156,165]</sup> The most frequent effect is a 'flu-like' complex consisting of fever and chills (incidence of >50%); other frequently reported effects include dyspnoea, tremor, chest pain/tightness, wheezing, diarrhoea, nausea and vomiting (approximately 10 to 20%). Most adverse events occur within 45 to 60 minutes of administration and last for 2 to 48 hours. More severe first-dose effects are aseptic meningitis (generally self-limiting without sequelae), seizures, intragraft thromboses and potentially fatal pulmonary oedema. Intragraft thromboses have also occurred later (within 2 weeks) during prophylaxis; 9 of 93 renal transplant recipients who received prophylactic muromonab CD3 (10 mg/day) developed a thrombosis with associated graft failure.<sup>[166]</sup> Significant increases in plasma prothrombin fragment 1 and 2 concentrations occurred 4 hours after the first dose. The first-dose response in children appears to be similar to that in adults.<sup>[19,167]</sup>

Risk factors for life- or allograft-threatening adverse events include diabetes mellitus, impaired renal allograft function, uraemia, fluid overload, high-dose muromonab CD3 (dosage not documented), a history of CNS disease, hyponatraemia and hypocalcaemia.<sup>[165,168,169]</sup>

The incidence of initial adverse events with muromonab CD3 (fever, headache, hypotension and/or signs and symptoms of fluid overload) is reported to be similar to or greater than that with

polyclonal agents, and events tend to be more severe with muromonab CD3.<sup>[65,96,97,99,170,171]</sup> The most common adverse events with polyclonal preparations are fever, flushing and chills.

Some investigators report first-dose effects to be more frequent or severe when muromonab CD3 is administered as prophylaxis than when it is used as treatment<sup>[156]</sup> whereas others report the reverse.<sup>[172]</sup>

### 5.1.1 Late Reactions

Cytokine-related adverse events may also occur after more than 4 or 5 doses of muromonab CD3.<sup>[172]</sup> Reactions involving angioedema, hypotension or serum sickness-like syndrome (compatible with IgE-mediated anaphylaxis or serum sickness mediated by antibody formation) 9 to 13 days after the initial dose of prophylactic muromonab CD3 have also occasionally been reported.<sup>[173]</sup> Continued treatment with muromonab CD3 after such a reaction is not recommended.

## 5.2 Infections

In common with other immunosuppressants, muromonab CD3 is associated with an increased risk of infections (the most common pathogens being CMV, herpes simplex virus and bacteria); these infections are the primary cause of morbidity and mortality in muromonab CD3 recipients.<sup>[19]</sup> The risk of infection appears to be related to the overall

degree of immunosuppression rather than to intrinsic factors of the drug.

The risk of CMV infection which is greatest in CMV-positive donor/CMV-negative recipients and in patients seropositive before treatment, is increased by high doses of muromonab CD3 (total doses >75mg) and repeated exposure to the agent.<sup>[1,174,175]</sup> The risk of invasive CMV disease was greater (16 vs 1 patient) when muromonab CD3 was administered during the first 14 post-transplant days than when administered subsequently.<sup>[176]</sup>

The incidence of CMV infection with prophylactic muromonab CD3-based immunosuppression (approximately 30 to 50%) is greater than or similar to that of triple therapy, with the severity of CMV infection tending to be greater in muromonab CD3 recipients.<sup>[107,108,111,119,121,177,178]</sup> The overall incidence of bacterial and fungal infections with these treatment options appears to be similar.

This is also the case for comparisons with ALG- or ATG-based regimens although some investigators have reported more severe infections and/or a greater incidence with muromonab CD3, especially in patients at high risk.<sup>[65,95-99,122-124,126-128,170,171,179]</sup>

The incidence of nosocomial infections (staphylococci, *Candida* sp. and *Enterobacter* sp.) but not community-acquired infections was greater (52 vs 35%) with muromonab CD3-based than ATG-based prophylaxis in 100 cardiac transplant recipients.<sup>[180]</sup>

### 5.3 Neoplasia

Muromonab CD3-based immunosuppression (administered as prophylaxis or treatment of rejection) has been associated with an increased risk of malignancies, mainly lymphoproliferative disorders, compared with conventional triple therapy.<sup>[19,181-188]</sup> The incidence of post-transplantation lymphoproliferative disorder was 9-fold higher with muromonab CD3 plus standard triple therapy (n = 79) than with triple therapy alone (n = 75) [11.4 vs 1.3%] in retrospectively-assessed cardiac transplant recipients who received therapy mainly for the prevention of rejection. The risk of

this disorder was dose-dependent; the incidence was significantly greater in patients who received a cumulative dose >75mg than in patients who received ≤75mg (35.7 vs 6.2%).<sup>[181]</sup> An increased risk of post-transplantation lymphoproliferative disorder in cardiac transplant recipients has also been reported by other research groups;<sup>[182,186,188]</sup> however, some investigators have reported a low incidence with prophylactic muromonab CD3.<sup>[189]</sup>

The factors responsible for the increased risk of lymphoproliferation with muromonab CD3 are unknown but are likely to be related to the overall degree of immunosuppression (including that conferred by concomitant immunosuppressants) rather than to mechanisms specific to muromonab CD3.<sup>[26,184,190,191]</sup> EBV infection and muromonab CD3-induced IL-6 and IL-10 release have been implicated in the development of this disorder.<sup>[26,184,190,192,193]</sup>

In addition to high muromonab CD3 dosages (total cumulative dose >75mg), the risk of lymphoproliferative disease may be increased by long durations of muromonab CD3 treatment,<sup>[187]</sup> multiple courses of the drug in close succession<sup>[194]</sup> and early retreatment.<sup>[195]</sup> No studies have specifically compared the risk of lymphoproliferative disorder with prophylactic muromonab CD3 versus administration for the treatment of rejection. Preliminary evidence suggests that screening for changes in peripheral CD19+ B lymphocytes may represent an effective strategy for identifying transplant recipients at risk of post-transplant lymphoproliferative disease.<sup>[196]</sup>

Some investigators report the incidence of neoplasia to be similar with muromonab CD3 and ALG;<sup>[170,171]</sup> others note a trend towards a greater incidence with muromonab CD3 prophylaxis than with ATG or ALG.<sup>[170,171,188,197]</sup>

### 5.4 Other Events

No clinically significant pulmonary or cardiovascular events have been noted in renal transplant recipients who received intraoperative muromonab CD3.<sup>[158,159]</sup> However, rare adverse events involving these and other systems during post-

operative administration include cardiopulmonary arrest<sup>[198]</sup> and hyperpyrexia-related ventricular tachycardia,<sup>[199]</sup> each in 1 patient, 1 report of reversible aseptic meningoencephalopathy following muromonab CD3 induction therapy<sup>[200]</sup> and 2 reports of aseptic encephalitis, with associated blindness due to bilateral optic nerve palsy in one patient after muromonab CD3 administration for delayed renal graft function.<sup>[201]</sup>

Seven per cent of renal transplant recipients (n = 247) who received muromonab CD3 induction therapy developed 'cytokine encephalopathy' (hallucinations, seizure, confusion, obtundation and coma) during the first 4 postoperative days; all but 1 patient (who died from a cardiac arrest) recovered fully with or without withdrawal of muromonab CD3.<sup>[202]</sup> The presence of delayed renal allograft function and/or insulin-dependent diabetes mellitus was significantly associated with the occurrence of this disorder. Early acute reversible nephrotoxicity associated with cytokine release occurs in some muromonab CD3 recipients,<sup>[157,203-205]</sup> as does *de novo* or recurrent haemolytic uraemic syndrome.<sup>[206,207]</sup>

Biphasic granulocytopenia has been observed after the first dose of muromonab CD3,<sup>[208]</sup> and reversible pancytopenia occurred in 2 renal transplant recipients receiving muromonab CD3 as prophylaxis or treatment of rejection.<sup>[209]</sup>

Prophylactic muromonab CD3 did not appear to adversely affect growth in children receiving a cardiac transplant.<sup>[139]</sup>

## 6. Drug Interactions

Although there is potential for a host of drug interactions between muromonab CD3 and concomitant immunosuppressants, anaesthetics and agents used to prevent or decrease adverse events, data are limited. There are no data on the potential for adverse interactions between muromonab CD3 and agents used to reduce the adverse events associated with this drug (table V).

Median trough blood concentrations of cyclosporin (administered preoperatively and in the immediate postoperative period) were significantly

higher on day 5 in renal transplant recipients who received muromonab CD3 induction therapy than in patients who received ALG.<sup>[210]</sup> Although corticosteroid pretreatment reduces cytokine release and first-dose adverse events with muromonab CD3 (table V), evidence suggests that procoagulant activity (section 1.5) is increased.<sup>[53]</sup>

Coadministration of indomethacin may increase the risk of encephalopathy in muromonab CD3 recipients.<sup>[211]</sup>

Although anaesthetic agents themselves may at least partly account for the reduced incidence or severity of first-dose effects observed with the intraoperative administration of muromonab CD3 (adverse events may be prevented or compensated for during anaesthesia),<sup>[212]</sup> concomitant treatment with muromonab CD3 and volatile anaesthetic agents or drugs that decrease cardiac contractility increases the risk of developing cardiovascular problems.<sup>[19]</sup>

## 7. Dosage and Administration

Induction therapy with muromonab CD3 plus azathioprine, methylprednisolone/prednisone and delayed cyclosporin therapy (section 3) is the most accepted muromonab CD3-based regimen for allograft rejection prophylaxis.

The optimal dosage of muromonab CD3 has not been established (section 3.1). Nevertheless, the currently recommended adult dosage in renal, hepatic and cardiac transplant recipients is 5mg administered intravenously as a once-daily bolus for 10 to 14 days. Although there are no specific dosage recommendations in children, 2.5mg once daily has usually been used in clinical trials (section 3.5.2). Higher doses may, however, be needed; paediatric patients appear to require increased dosages more frequently than adults.<sup>[40]</sup> Surveillance for clinical signs of rejection in addition to anti-muromonab CD3 antibodies, plasma muromonab CD3 concentrations and CD3+ cell levels, as appropriate, is required for optimal muromonab CD3 dosage adjustments (section 2.2). Most patients receive muromonab CD3 from the first postoperative

**Table VI.** Factors to consider when assessing the risk-benefit profile of muromonab CD3 for the prevention of solid organ transplant rejection

Benefits	Limitations
<ul style="list-style-type: none"> <li>• Efficacy (section 3)</li> <li>• Effect on graft dysfunction (section 3)</li> <li>• Minimisation of cyclosporin toxicity</li> <li>• Less batch variability, greater specificity, less cross-reactivity with blood components other than T cells and relative ease of administration (peripheral versus central line administration) compared with polyclonal antibodies (sections 1.1 and 2.1)</li> </ul>	<ul style="list-style-type: none"> <li>• Tolerability: first-dose cytokine-related adverse events (section 5.1), infections (section 5.2), neoplasia (section 5.3)</li> <li>• Immunogenicity (section 1.3) and potential to prevent reuse (section 3.6)</li> <li>• Cost: drug and hospital costs (section 4)</li> </ul>

day; however, intraoperative administration appears to improve the tolerability of the drug.

The first few doses should be administered in a facility equipped for cardiopulmonary resuscitation; vital signs should be closely monitored. Subsequent doses may be administered on an outpatient basis with monitoring as appropriate.

The preventative measures outlined in table V should be observed with muromonab CD3 use, and only the lowest effective dosages of concomitant immunosuppressive agents should be administered (to minimise the potential for malignancies and infections; section 5).

Patients receiving muromonab CD3 should be monitored for signs of lymphoproliferative disorder, and antilymphocyte agents should be used with caution in those at risk of EBV infection and in those with pre-existing tumours.

## 8. Place of Muromonab CD3 in the Prevention of Transplant Rejection

Despite substantial improvements in the surgical and medical management of transplant recipients, acute rejection remains a significant problem.

Although the use of muromonab CD3-based immunosuppression as first-line and 'rescue' therapy of rejection is well established,<sup>[1]</sup> its use as prophylaxis of rejection, particularly as routine therapy, has been controversial. The 2 major immunosuppressive protocols currently in use for the prophylaxis of rejection episodes are standard triple therapy (corticosteroids, azathioprine and cyclosporin) and monoclonal (muromonab CD3) or polyclonal (ALG or ATG) antilymphocyte antibody-based sequential therapy. Cyclosporin-associated nephrotoxicity and hepatotoxicity in the

early postoperative period prompted, in part, the development of the latter more specific regimens. Antilymphocyte-based sequential therapy consists of an induction phase with an antilymphocyte preparation plus azathioprine, corticosteroids and cyclosporin (delayed until graft function is established) and maintenance therapy.

The definitive answer to whether muromonab CD3 should be used routinely as prophylaxis or be reserved for the treatment of rejection episodes has yet to be determined. The risks and benefits that need to be considered when making such a decision are detailed in table VI.

The ideal immunosuppressant agent/regimen would:

- induce specific tolerance to donor antigens and obviate the need for long term immunosuppression;
- be associated with minimal adverse events and achieve a balance between the risks of over-immunosuppression (e.g. infections and malignancies) and underimmunosuppression (e.g. graft loss);
- optimise early graft function;
- reduce rejection incidence and improve graft and patient survival;
- have no sensitising or activating properties;
- minimise hospitalisation.

Muromonab CD3 induction therapy as part of sequential therapy is significantly more effective than standard triple therapy in the prophylaxis of allograft rejection in renal and hepatic, but not cardiac, transplant recipients. The overall efficacy of muromonab CD3- and polyclonal-based prophylactic regimens appears to be similar, although results vary between investigators. Muromonab CD3

also improves overall graft survival in renal transplant recipients with delayed graft function or other high-risk factors, optimises early graft function and may minimise hospitalisation. However, as with other currently available immunosuppressants, rejection may still occur, patient survival does not appear to be improved, adverse events (particularly infections) and immunogenicity limit its use, and the other above-mentioned criteria for the 'ideal' immunosuppressive agent are not met.

Research is now focused on developing more specific immunosuppressive agents. Monoclonal antibodies, other than muromonab CD3, that have either shown promise or are under investigation for the prophylaxis of transplant rejection include antibodies directed against CD4, CD6, CD7, CD8, CD11a, CD18, CD25, CD45 or intracellular adhesion molecules, anti-IL-2 receptor monoclonal antibodies, humanised (and entirely human) pan-lymphocyte monoclonal antibodies, monovalent anti-CD3 antibodies, muromonab CD3 F(ab')<sub>2</sub> fragments, growth factor antagonists and antisense oligonucleotides. Encouragingly, donor-specific tolerance has been induced in animal models by anti-CD4, anti-CD25 and anti-CD54 monoclonal antibodies.<sup>[4,37,131,146,213,214]</sup>

Other potential approaches to improving prophylactic immunosuppression include the development of anti-CD3 monoclonal antibodies of the IgG2b type (to reduce adverse events), humanised muromonab CD3 preparations that are less immunogenic and synthetic peptides that deviate alloreactive T cells; the modification of class I major histocompatibility complex antigens by site-directed mutagenesis; the use of several monoclonal antibodies, each targeting a different TCR epitope; the systemic administration of specific agents or the local administration of nonspecific agents; and the use of different monoclonal antibodies for prophylaxis and treatment.

In the meantime, however, muromonab CD3 is the only clinically available monoclonal antibody for the prevention of solid organ transplant rejection. The efficacy of sequential immunosuppression with this agent in reducing early acute rejection

episodes is greater than that of standard triple therapy in renal and hepatic, but not cardiac, transplant recipients. The overall efficacy of muromonab CD3-based induction regimens appears to be similar to that of polyclonal-based regimens, although this requires confirmation. While the routine use of muromonab CD3 for the prevention of rejection in patients with primary graft function does not appear to be justified, prophylactic sequential muromonab CD3-based therapy has a role in high-risk patients, particularly those with primary allograft dysfunction or renal failure.

## References

1. Todd PA, Brogden RN. Muromonab CD3: a review of its pharmacology and therapeutic potential. *Drugs* 1989; 37: 871-99
2. Chatenoud L, Bach J-F. Selective immunosuppression with anti-T cell monoclonal antibodies. *Clin Nephrol* 1992; 38 Suppl. 1: 53-60
3. Roitt IM. OKT3: immunology, production, purification, and pharmacokinetics. *Clin Transpl* 1993 Aug; 7 (Pt 2): 367-73
4. Cosimi AB. Current and future application of monoclonal antibodies in clinical immunosuppressive protocols. *Clin Transpl* 1995 Jun; 9 (Pt 2): 219-26
5. Norman DJ. Mechanisms of action and overview of OKT3. *Ther Drug Monit* 1995; 17: 615-20
6. Gonçalves LF, Rauber ML, Manfro RC, et al. Fine needle aspiration biopsy in renal transplant patients on prophylactic OKT3 treatment. *Transplant Proc* 1992 Dec; 24: 3085-6
7. Ouwehand AJ, Baan CC, Groeneveld K, et al. Altered specificity of alloreactive cardiac graft-infiltrating cells by prophylactic treatment with OKT3 or horse antilymphocyte globulin. *Transplantation* 1993; 55: 154-8
8. Janssen O, Wesselborg S, Kabelitz D. Immunosuppression by OKT3 - induction of programmed cell death (apoptosis) as a possible mechanism of action. *Transplantation* 1992 Jan; 53: 233-4
9. Magnussen K, Klug B, Moller B. CD3 antigen modulation in T-lymphocytes during OKT3 treatment. *Transplant Proc* 1994 Jun; 26: 1731
10. Buysmann S, van Diepen FJ, van Kooyk Y, et al. The influence of OKT3 on expression of lymphocyte adhesion molecules in vitro. *Transplant Proc* 1994 Dec; 26: 3249-50
11. Buysmann S, Bemelman FJ, Schellekens PT, et al. Activation and increased expression of adhesion molecules on peripheral blood lymphocytes is a mechanism for the immediate lymphocytopenia after administration of OKT3. *Blood* 1996; 87 (1): 404-11
12. Wong JT, Eylath A, Ghobrial I, et al. The mechanism of anti-CD3 monoclonal antibodies. Mediation of cytolysis by inter-T cell bridging. *Transplantation* 1990 Oct; 50: 683-9
13. Abramowicz D, Goldman M, De Pauw L, et al. The long-term effects of prophylactic OKT3 monoclonal antibody in cadaver kidney transplantation - a single-center, prospective, randomized study. *Transplantation* 1992 Sep; 54: 433-7
14. Abramowicz D, Schandene L, Goldman M, et al. Release of tumor necrosis factor, interleukin-2, and gamma-interferon in serum after injection of OKT3 monoclonal antibody in kidney transplant recipients. *Transplantation* 1989; 47: 606-8



15. Ellenhorn JDI, Woodle ES, Ghobreal I, et al. Activation of human T cells in vivo following treatment of transplant recipients with OKT3. *Transplantation* 1990 Oct; 50: 608-12
16. Gaston RS, Deierhoi MH, Patterson T, et al. OKT3 first-dose reaction: association with T-cell subsets and cytokine release. *Kidney Int* 1991 Jan; 39: 141-8
17. Raasveldt MHM, Bemelman FJ, Schellekens PThA, et al. Complement activation during OKT3 treatment: a possible explanation for respiratory side effects. *Kidney Int* 1993 May; 43: 1140-9
18. Chatenoud L. Use of CD3 antibodies in transplantation and autoimmune diseases. *Transplant Proc* 1994 Dec; 26: 3191-3
19. Kreis H. Adverse events associated with OKT3 immunosuppression in the prevention or treatment of allograft rejection. *Clin Transpl* 1993 Aug; 7 (Pt 2): 431-46
20. Goumy L, Ferran C, Merite S, et al. In vivo anti-CD3-driven cell activation. Cellular source of induced tumour necrosis factor, interleukin-1 $\beta$ , and interleukin 6. *Transplantation* 1996; 61: 83-7
21. Bloemena E, ten Berge IJM, Surachno J, et al. Kinetics of interleukin 6 during OKT3 treatment in renal allograft recipients. *Transplantation* 1990 Aug; 50: 330-1
22. Hoffman T, Tripathi AK, Lee YL, et al. Stimulation of human monocytes by anti-CD3 monoclonal antibody: induction of inflammatory mediator release via immobilization of Fc receptor by adsorbed immunoglobulin and T-lymphocytes. *Inflammation* 1992 Dec; 16: 571-85
23. Woodle ES, Thistlethwaite JR, Jolliffe LK, et al. T-cell activation and lymphokine production induced by antihuman CD3 monoclonal antibodies. *Transplant Proc* 1991; 23: 81-2
24. Vossen ACTM, Tibbe GJM, Kroos MJ, et al. Fc receptor binding of anti-CD3 monoclonal antibodies is not essential for immunosuppression, but triggers cytokine-related side effects. *Eur J Immunol* 1995; 25: 1492-6
25. Goldman M, Gérard C, Abramowicz D, et al. Induction of interleukin-6 and interleukin-10 by the OKT3 monoclonal antibody: possible relevance to posttransplant lymphoproliferative disorders. *Clin Transpl* 1992 Jun; 6: 265-8
26. Swinnen LJ, Fisher RI. OKT3 monoclonal antibodies induce interleukin-6 and interleukin-10: a possible cause of lymphoproliferative disorders associated with transplantation. *Curr Opin Nephrol Hypertension* 1993 Jul; 2: 670-8
27. Ferran C, Dy M, Merite S, et al. Reduction of morbidity and cytokine release in anti-CD3 MoAb-treated mice by corticosteroids. *Transplantation* 1990 Oct; 50: 642-8
28. Chatenoud L, Ferran C, Legendre C, et al. In vivo cell activation following OKT3 administration. Systemic cytokine release and modulation by corticosteroids. *Transplantation* 1990 Apr; 49: 697-702
29. Chatenoud L, Legendre C, Ferran C, et al. Corticosteroid inhibition of the OKT3-induced cytokine-related syndrome - dosage and kinetics prerequisites. *Transplantation* 1991 Feb; 51: 334-8
30. Schandené L, Gérard C, Crusiaux A, et al. Interleukin-10 inhibits OKT3-induced cytokine release: in vitro comparison with pentoxifylline. *Transplant Proc* 1993 Apr; 25 (2 Suppl. 1): 55-6
31. Leimenstoll G, Zabel P, Schroeder P, et al. Suppression of OKT3-induced tumor necrosis factor alpha formation by pentoxifylline in renal transplant recipients. *Transplant Proc* 1993 Feb; 25 (Pt 1): 561-3
32. Donckier V, Flament V, Gérard C, et al. Modulation of the release of cytokines and reduction of the shock syndrome induced by anti-CD3 monoclonal antibody in mice by interleukin-10. *Transplantation* 1994 May 27; 57: 1436-9
33. DeVault Jr GA, Kohan DE, Nelson EW, et al. The effects of oral pentoxifylline on the cytokine release syndrome during inductive OKT3. *Transplantation* 1994 Feb 27; 57: 532-40
34. Alegre M-L, Gastaldello K, Abramowicz D, et al. Evidence that pentoxifylline reduces anti-CD3 monoclonal antibody-induced cytokine release syndrome. *Transplantation* 1991 Oct; 52: 674-9
35. Carey G, Lisi PJ, Schroeder TJ. The incidence of antibody formation to OKT3 consequent to its use in organ transplantation. *Transplantation* 1995; 60: 151-8
36. Chatenoud L. Humoral immune response against OKT3. *Transplant Proc* 1993 Apr; 25 Suppl. 1: 68-73
37. Norman DJ. Antilymphocyte antibodies in the treatment of allograft rejection: targets, mechanisms of action, monitoring, and efficacy. *Semin Nephrol* 1992; 12: 315-24
38. Kimball JA, Norman DJ, Shield CF, et al. OKT3 antibody response study: comparative testing of human antimouse antibody. *Transplant Proc* 1993 Apr; 25 Suppl. 1: 74-6
39. O'Connell JB, Bristow MR, Hammond EH, et al. Antimurine antibody to OKT3 in cardiac transplantation: implications for prophylaxis and retreatment of rejection. *Transplant Proc* 1991 Feb; 23: 1157-9
40. Schroeder TJ, Michael AT, First MR, et al. Variations in serum OKT3 concentration based upon age, sex, transplanted organ, treatment regimen, and anti-OKT3 antibody status. *Ther Drug Monit* 1994 Aug; 16: 361-7
41. Taylor DO, Bristow MR, O'Connell JB, et al. A prospective, randomized comparison of cyclophosphamide and azathioprine for early rejection prophylaxis after cardiac transplantation: decreased sensitization to OKT3. *Transplantation* 1994 Sep 27; 58: 645-9
42. Schroeder TJ, First MR, Mansour ME, et al. Antimurine antibody formation following OKT3 therapy. *Transplantation* 1990 Jan; 49: 48-51
43. Norman DJ, Chatenoud L, Cohen D, et al. Consensus statement regarding OKT3-induced cytokine-release syndrome and human antimouse antibodies. *Transplant Proc* 1993 Apr; 25 Suppl. 1: 89-92
44. Rohrer RJ, Jenkins RL, Khettry U, et al. Immunohistology of liver allografts in recipients managed with prophylactic OKT3. *Transplant Proc* 1989; 21: 2249-50
45. Kemnitz J, Cremer J, Schaefer HJ, et al. Some aspects of changed histopathologic appearance of acute rejection in cardiac allografts after prophylactic application of OKT3. *J Heart Lung Transplant* 1991 May-Jun; 10: 366-72
46. Kaufman C, Zeevi A, Zerby T, et al. In vitro studies of endomyocardial biopsies from heart transplant recipients on RATG and OKT3 immunoprophylaxis protocols. *Transplantation* 1989; 48: 621-5
47. Fyfe AI, Harper CM. Anti-thymocyte globulin and OKT3 have opposite effects on adhesion of cardiac transplant recipient mononuclear cells to arterial endothelium [abstract]. *Circulation* 1993; 44: 1-41
48. Breisblatt WM, Schulman DS, Stein K, et al. Hemodynamic response to OKT3 in orthotopic heart transplant recipients: evidence for reversible myocardial dysfunction. *J Heart Lung Transplant* 1991 May-Jun; 10: 359-65
49. Robinson ST, Barry JM, Norman DJ. The hemodynamic effects of intraoperative injection of muromonab CD3. *Transplantation* 1993 Aug; 56: 356-8
50. Stein KL, Ladowski J, Kormos R, et al. The cardiopulmonary response to OKT3 in orthotopic cardiac transplant recipients. *Chest* 1989 Apr; 95: 817-21
51. Beilman GJ, Shield III CF, Hughes JD, et al. The effects of intraoperative administration of OKT3 during renal transplantation. *Transplantation* 1993 Mar; 55: 490-3

52. Raasveld MHM, Hack CE, ten Berge IJM. Activation of coagulation and fibrinolysis following OKT3 administration to renal transplant recipients: association with distinct mediators. *Thromb Haemost* 1992 Sep 7; 68: 264-7
53. Abramowicz D, Pradier O, De Pauw L, et al. High-dose glucocorticosteroids increase the procoagulant effects of OKT3. *Kidney Int* 1994 Dec; 46: 1596-602
54. Pradier O, Marchant A, Abramowicz D, et al. Procoagulant effect of the OKT3 monoclonal antibody: involvement of tumor necrosis factor. *Kidney Int* 1992 Nov; 42: 1124-9
55. Goldman MH, McGrath G, Freeman M, et al. D-Dimer XDP correlates with fibrinolytic shutdown in renal transplant patients treated with anti-T-cell antibodies. *Transplant Proc* 1995 Feb; 27: 1094-6
56. Pradier O, Abramowicz D, Capel P, et al. Procoagulant properties of OKT3 at the monocyte level: inhibition by pentoxifylline. *Transplant Proc* 1993 Apr; 25 Suppl. 1: 39-40
57. Hesse CJ, Heyse P, Stolk BJM, et al. Differences in antibody formation to OKT3 between kidney and heart transplantation recipients. *Transplant Proc* 1989; 21: 979-80
58. Henell KR, Norman DJ. Monitoring OKT3 treatment: pharmacodynamic and pharmacokinetic measures. *Transplant Proc* 1993 Apr; 25 Suppl. 1: 83-5
59. Madden RL, Schroeder TJ, Alexander JW, et al. Single dose OKT3: adverse effects, pharmacokinetics, and anti-OKT3 antibody response. *Transplant Sci* 1994 Sep; 4: 111-4
60. Goldstein G, Fucello AJ, Norman DJ, et al. OKT3 monoclonal antibody plasma levels during therapy and the subsequent development of host antibodies to OKT3. *Transplantation* 1986; 42: 507-11
61. Schroeder TJ, First MR, Hurtubise PE, et al. Immunologic monitoring with Orthoclone OKT3 therapy. *J Heart Transplant* 1989 Sep-Oct; 8: 371-80
62. Norman DJ. The clinical role of OKT3. *Cardiol Clin* 1990 Feb; 8: 97-105
63. Hesse CJ, Heyse P, Stolk BJM, et al. Immune monitoring of heart transplant patients receiving either one or two cycles of OKT3 prophylaxis - induced anti-idiotypic and anti-isotypic anti-OKT3 antibodies do not prohibit depletion of peripheral T-cells due to second OKT3 treatment. *Clin Transpl* 1991 Dec; 5 (Part 1): 446-55
64. Goldstein G, Norman DJ, Henell KR, et al. Pharmacokinetic study of Orthoclone OKT3 serum levels during treatment of acute renal allograft rejection. *Transplantation* 1988; 46: 587-9
65. Bock HA, Gallati H, Zürcher RM, et al. A randomized prospective trial of prophylactic immunosuppression with ATG-Fresenius versus OKT3 after renal transplantation. *Transplantation* 1995 Mar 27; 59: 830-40
66. McDiarmid SV, Millis M, Terashita G, et al. Low serum OKT3 levels correlate with failure to prevent rejection in orthotopic liver transplant patients. *Transplant Proc* 1990 Aug; 22: 1774-6
67. Schroeder TJ, Ryckman FC, Hurtubise PE, et al. Immunological monitoring during and following OKT3 therapy in children. *Clin Transpl* 1991 Apr; 5: 191-6
68. Abramowicz D, Goldman M, Mat O, et al. OKT3 serum levels as a guide for prophylactic therapy: a pilot study in kidney transplant recipients. *Transpl Int* 1994 Jul; 7: 258-63
69. McCarthy C, Light JA, Aquino A, et al. Correlation of CD3<sup>+</sup> lymphocyte depletion with rejection and infection in renal transplants. *Transplant Proc* 1993 Aug; 25: 2477-8
70. Gebel H, Lebeck LL, Jensik SC, et al. Discordant expression of CD3 and T cell receptor antigens on lymphocytes from patients treated with OKT3. *Transplant Proc* 1989; 21: 1745-6
71. Shaefer MS, Stratta RJ, Pirruccello SJ, et al. Peripheral CD3 lymphocyte monitoring of liver transplant recipients being treated with OKT3 for rejection or induction immunosuppression [abstract]. *Pharmacotherapy* 1990; 10 (3): 248
72. Broughan TA, Valenzuela R, Escorcía E, et al. Mouse antibody-coated lymphocytes during OKT3 therapy in liver transplantation. *Clin Transpl* 1994 Oct; 8: 488-91
73. Ohman M, Kotb M, Leathers LK, et al. Multiparameter monitoring of efficacy of OKT3-induced immune suppression in renal allograft recipients [abstract]. *Hum Immunol* 1993; 37 Suppl. 1: 95
74. Gebel HM, Lebeck LK, Jensik SC, et al. T cells from patients successfully treated with OKT3 do not react with the T-cell receptor antibody. *Hum Immunol* 1989 Oct; 26: 123-30
75. Hammond EA, Yowell RL, Greenwood J, et al. Prevention of adverse clinical outcome by monitoring of cardiac transplant patients for murine monoclonal CD3 antibody (OKT3) sensitization. *Transplantation* 1993 May; 55: 1061-3
76. Moore CK, O'Connell JB, Renlund DG, et al. Cardiac allograft cellular rejection during OKT3 prophylaxis in the absence of sensitization. *Transplant Proc* 1991 Feb; 23: 1055-8
77. Ryckman FC, Schroeder TJ, Pedersen SH, et al. Use of monoclonal antibody immunosuppressive therapy in pediatric renal and liver transplantation. *Clin Transplant* 1991; 5: 186-90
78. Toyoda M, Galfayan K, Wachs K, et al. Immunologic monitoring of OKT3 induction therapy in cardiac allograft recipients. *Clin Transplant* 1995; 9: 472-80
79. Wechsler ME, Giardina E-GV, Sciacca RR, et al. Increased early mortality in women undergoing cardiac transplantation. *Circulation* 1995 Feb 15; 91: 1029-35
80. Troppmann C, Gillingham KJ, Benedetti E, et al. Delayed graft function, acute rejection, and outcome after cadaver renal transplantation: a multivariate analysis. *Transplantation* 1995 Apr 15; 59: 962-8
81. Howard RJ, Pfaff WW, Brunson ME, et al. Delayed graft function is associated with an increased incidence of occult rejection and results in poorer graft survival. *Transplant Proc* 1993 Feb; 25 (1 Pt 2): 884
82. Costanzo-Nordin MR, Fisher SG, O'Sullivan EJ, et al. HLA-DR incompatibility predicts heart transplant rejection independent of immunosuppressive prophylaxis. *J Heart Lung Transplant* 1993 Sep-Oct; 12: 779-89
83. Kerman RH, Sullivan K, Tejani A. Impact of HLA matching, type of crossmatch and immunosuppressive therapy on primary pediatric cadaver renal allograft survival [abstract]. *Hum Immunol* 1994; 40 Suppl. 1: 17
84. Norman DJ, Kimball JA, Bennett WM, et al. A prospective, double-blind, randomized study of high-versus low-dose OKT3 induction immunosuppression in cadaveric renal transplantation. *Transpl Int* 1994 Aug; 7: 356-61
85. Norman DJ, Barry JM, Bennett WM, et al. OKT3 for induction immunosuppression in renal transplantation: a comparative study of high versus low doses. *Transplant Proc* 1991 Feb; 23: 1052-4
86. Alonso-Pulpón L, Serrano-Fiz S, Rubio JA, et al. Efficacy of low-dose OKT3 as cytolytic induction therapy in heart transplantation. *J Heart Lung Transplant* 1995 Jan/Feb; 14 (1 Pt 1): 136-42
87. Welter HF, Illner W-D, Schleibner S, et al. Pilot study on induction treatment with high-dose OKT3: preliminary observations in kidney transplantation. *Transplant Proc* 1990 Oct; 22: 2272
88. Hegewald MG, O'Connell JB, Renlund DG, et al. OKT3 monoclonal antibody given for ten versus fourteen days as immunosuppressive prophylaxis in heart transplantation. *J Heart Transplant* 1989 Jul-Aug; 8: 303-10

89. Kobashigawa JA, Stevenson LW, Brownfield E, et al. Does short-course induction with OKT3 improve outcome after heart transplantation? A randomized trial. *J Heart Lung Transplant* 1993 Mar-Apr; 12: 205-8
90. Buysmann S, Hack CE, van Diepen FNJ, et al. Administration of OKT3 via continuous infusion attenuates first-dose side-effects. *Kidney Int* 1995; 48: 1368-9
91. Ortho Biotech Inc. Orthoclone OKT3 prescribing information. New Jersey, USA, 1995
92. Benvenisty AI, Cohen D, Stegall MD, et al. Improved results using OKT3 as induction immunosuppression in renal allograft recipients with delayed graft function. *Transplantation* 1990 Feb; 49: 321-7
93. Dafoe DC, Bromberg JS, Grossman RA, et al. Renal transplantation despite a positive antiglobulin crossmatch with and without prophylactic OKT3. *Transplantation* 1991 Apr; 51: 762-8
94. Norman DJ, Kahana L, Stuart Jr FP, et al. A randomized clinical trial of induction therapy with OKT3 in kidney transplantation. *Transplantation* 1993 Jan; 55: 44-50
95. Broyer M, Gagnadoux M-F, Guest G, et al. Prophylactic OKT3 monoclonal antibody versus antilymphocyte globulins: a prospective, randomized study in 148 first cadaver kidney grafts. *Transplant Proc* 1993 Feb; 25 (Pt 1): 570-1
96. Cole EH, Cattran DC, Farewell VT, et al. A comparison of rabbit antithymocyte serum and OKT3 as prophylaxis against renal allograft rejection. *Transplantation* 1994 Jan; 57: 60-7
97. Frey DJ, Matas AJ, Gillingham KJ, et al. Sequential therapy – a prospective randomized trial of malg versus OKT3 for prophylactic immunosuppression in cadaver renal allograft recipients. *Transplantation* 1992 Jul; 54: 50-6
98. Hanto DW, Jendrisak MD, So SKS, et al. Induction immunosuppression with antilymphocyte globulin or OKT3 in cadaver kidney transplantation. Results of a single institution prospective randomized trial. *Transplantation* 1994 Feb 15; 57: 377-84
99. Steinmuller DR, Hayes JM, Novick AC, et al. Comparison of OKT3 with ALG for prophylaxis for patients with acute renal failure after cadaveric renal transplantation. *Transplantation* 1991 Jul; 52: 67-71
100. Opelz G. Efficacy of rejection prophylaxis with OKT3 in renal transplantation. *Transplantation* 1995; 60: 1220-4
101. Vela C, Cristol JP, Chong G, et al. Antilymphocyte globulins versus OKT3 as prophylactic treatment in highly sensitized renal transplant recipients [abstract]. *Nephrol Dial Transplant* 1993; 8 (9): 1052
102. Knechtle SJ, Pirsch JD, Groshek M, et al. OKT3 vs ALG induction therapy in combined pancreas-kidney transplantation. *Transplant Proc* 1991 Feb; 23: 1581-2
103. Lefrançois N, Raffaele P, Martinenghi S, et al. Prophylactic polyclonal versus monoclonal antibodies in kidney and pancreas transplantation. *Transplant Proc* 1990; 22: 632-3
104. Illner W-D, Theodorakis J, Abendroth D, et al. Quadruple-drug induction therapy in combined renal pancreatic transplantation – OKT3 versus ATG. *Transplant Proc* 1990 Aug; 22: 1586-7
105. Melzer JS, D'Alessandro AM, Kalayoglu M, et al. The use of OKT3 in combined pancreas-kidney allotransplantation. *Transplant Proc* 1990 Apr; 22: 634-5
106. Cosimi AB, Jenkins RL, Rohrer RJ, et al. A randomized clinical trial of prophylactic OKT3 monoclonal antibody in liver allograft recipients. *Arch Surg* 1990 Jun; 125: 781-5
107. Farges O, Ericzon B-G, Bresson-Hadni S, et al. A randomized trial of OKT3-based versus cyclosporine-based immunoprophylaxis after liver transplantation: long-term results of a European and Australian multicenter study. *Transplantation* 1994 Oct 27; 58: 891-8
108. McDiarmid SV, Millis MJ, Terasaki PI. OKT3 prophylaxis in liver transplantation. *Dig Dis Sci* 1991 Oct; 36: 1418-26
109. McDiarmid SV, Busuttill RW, Levy P, et al. The long-term outcome of OKT3 compared with cyclosporine prophylaxis after liver transplantation. *Transplantation* 1991 Jul; 52: 91-7
110. Mühlbacher F, Steininger R, Längle F, et al. OKT3 immunoprophylaxis in human liver transplantation. *Transplant Proc* 1989; 21: 2253-4
111. Pons JA, Bueno F, Parrilla P, et al. Cyclosporine vs OKT3 prophylaxis after orthotopic liver transplantation. *Transplant Proc* 1993 Apr; 25: 1949
112. Steininger R, Mühlbacher F, Hamilton G, et al. Comparison of CyA, OKT3, and ATG immunoprophylaxis in human liver transplantation. *Transplant Proc* 1991 Aug; 23: 2269-71
113. Reding R, Vraux H, de Goyet J de V, et al. Monoclonal antibodies in prophylactic immunosuppression after liver transplantation. A randomized controlled trial comparing OKT3 and anti-IL-2 receptor monoclonal antibody LO-Tact-1. *Transplantation* 1993 Mar; 55: 534-41
114. Millis JM, McDiarmid S, Hiatt JR, et al. Randomized prospective trial of okt3 for early prophylaxis of rejection after liver transplantation. *Transplantation* 1989; 47: 82-8
115. Sasaki AW, Lee RG, Porayko MK, et al. Accelerated liver allograft rejection during prophylactic immunosuppression with OKT3. *Transplantation* 1993 Jan; 55: 216-9
116. Mor E, Skerrett D, Manzarbeitia C, et al. Successful use of an enhanced immunosuppressive protocol with plasmapheresis for ABO-incompatible mismatched grafts in liver transplant recipients. *Transplantation* 1995 Apr 15; 59: 986-90
117. Tokunaga Y, Tanaka K, Fujita S, et al. Living related liver transplantation across ABO blood groups with FK506 and OKT3. *Transpl Int* 1993 Nov; 6: 313-8
118. Fisher RA, Posner M, Shiffman ML, et al. Induction with OKT3 and prostaglandin E<sub>1</sub> in liver transplantation. *Transplant Sci* 1994; 4 Suppl. 1: S1-8
119. Balk AHMM, Simoons ML, Jutte NHPM, et al. Sequential OKT3 and cyclosporine after heart transplantation. A randomized study with single and cyclic OKT3. *Clin Transpl* 1991 Aug; 5: 301-5
120. Barr ML, Sanchez JA, Seche LA, et al. Anti-CD3 monoclonal antibody induction therapy. Immunological equivalency with triple-drug therapy in heart transplantation. *Circulation* 1990 Nov; 82 Suppl. IV: IV-291-4
121. Stapleton DD, Ventura HO, Grundtner SE, et al. Induction immunosuppression with the monoclonal antibody OKT3 after cardiac transplantation. *Am J Med Sci* 1993 Jul; 306: 16-9
122. Costanzo-Nordin MR, O'Sullivan EJ, Johnson MR, et al. Prospective randomized trial of OKT3 versus horse antithymocyte globulin based immunosuppressive prophylaxis in heart transplantation. *J Heart Transplant* 1990 May-Jun; 9: 306-15
123. Griffith BP, Kormos RL, Armitage JM, et al. Comparative trial of immunoprophylaxis with RATG versus OKT3. *J Heart Transplant* 1990 May-Jun; 9: 301-5
124. Ippoliti G, Negri M, Abelli P, et al. Preoperative prophylactic OKT3 vs RATG. A randomized clinical study in heart transplant patients. *Transplant Proc* 1991 Aug; 23: 2272-4
125. Kirklin JK, Bourge RC, White-Williams C, et al. Prophylactic therapy for rejection after cardiac transplantation – a comparison of rabbit antithymocyte globulin and OKT3. *J Thorac Cardiovasc Surg* 1990 Apr; 99: 716-24
126. Ladowski JS, Dillon T, Schatzlein MH, et al. Prophylaxis of heart transplant rejection with either antithymocyte globulin-,

- Minnesota antilymphocyte globulin-, or an OKT3-based protocol. *J Cardiovasc Surg* 1993 Apr; 34: 135-40
127. Macdonald PS, Mundy J, Keogh AM, et al. A prospective randomized study of prophylactic OKT3 versus equine antithymocyte globulin after heart transplantation – increased morbidity with OKT3. *Transplantation* 1993 Jan; 55: 110-6
  128. Menkis AH, Powell A-M, Novick RJ, et al. A prospective randomized controlled trial of initial immunosuppression with ALG versus OKT3 in recipients of cardiac allografts. *J Heart Lung Transplant* 1992 May-Jun; 11: 569-76
  129. Renlund DG, O'Connell JB, Gilbert EM, et al. A prospective comparison of murine monoclonal CD-3 OKT3 antibody-based and equine antithymocyte globulin-based rejection prophylaxis in cardiac transplantation decreased rejection and less corticosteroid use with OKT3. *Transplantation* 1989; 47: 599-605
  130. Wollenek G, Laufer G, Laczkovics A, et al. Comparison of a monoclonal anti-T cell antibody vs ATG as prophylaxis after heart transplantation. *Transplant Proc* 1989; 21: 2499-501
  131. van Gelder T, Mulder AH, Balk AHMM, et al. Intra-graft monitoring of rejection after prophylactic treatment with monoclonal anti-interleukin-2 receptor antibody (BT563) in heart transplant recipients. *J Heart Lung Transplant* 1995; 14: 346-50
  132. Carrier M, Jenicek M, Pelletier LC. Value of monoclonal antibody OKT3 in solid organ transplantation: a meta-analysis. *Transplant Proc* 1992 Dec; 24: 2586-91
  133. Induhara R, Khauli RB, Menon M. Simultaneous quadruple immunosuppression with cyclosporine induction therapy in high risk renal transplant recipients. *J Urol* 1994 Aug; 152 (Pt 1): 307-11
  134. Schroeder TJ, First MR, Mansour ME, et al. Prophylactic use of OKT3 in immunologic high-risk cadaver renal transplant recipients. *Am J Kidney Dis* 1989 Nov; 14 Suppl. 2: 14-8
  135. Abramowicz D, Norman DJ, Vereerstraeten P, et al. OKT3 prophylaxis in renal grafts with prolonged cold ischemia times: association with improvement in long-term survival. *Kidney Int*. In press
  136. Cardella CJ, Blake P, Cattran D, et al. Prophylactic OKT3 in renal retransplantation. *Transplant Proc* 1989 Apr; 21: 3373-4
  137. Schroeder TJ, First MR, Gaber AO. Monitoring and management of immunosuppression in paediatric transplant patients. *Clin Immunother* 1995; 4 (6): 425-44
  138. Shaddy RE, Bullock EA, Morwessel NJ, et al. Murine monoclonal CD3 antibody (OKT3)-based early rejection prophylaxis in pediatric heart-transplant. *J Heart Lung Transplant* 1993 May-Jun; 12: 434-9
  139. Brown JW, Turrentine MW, Kesler KA, et al. Triple-drug immunosuppression for heart transplantation in infants and children. *J Heart Lung Transplant* 1993 Nov-Dec; 12: S265-74
  140. Conley SB, al-Uzri A, So S, et al. Prevention of rejection and graft loss with an aggressive quadruple immunosuppressive therapy regimen in children and adolescents. *Transplantation* 1994 Feb 27; 57: 540-4
  141. Bartosh SM, Aronson AJ, Swanson-Pewitt EE, et al. OKT3 induction in pediatric renal transplantation. *Pediatr Nephrol* 1993 Feb; 7: 45-9
  142. McDiarmid SV, Millis MJ, Terasaki P, et al. Induction of immunosuppression in pediatric orthotopic liver transplantation. *Clin Transpl* 1991; 5: 174-80
  143. Peters TG, Charlton RK, Jones KW, et al. Kidney transplantation in the older patient. *J Fla Med Assoc* 1994 Aug; 81: 535-8
  144. Schroeder TJ, Rossi SJ, First MR. OKT3 monoclonal antibody therapy. International Consensus Conference on Immunosuppressive Drugs. 1995 5 May
  145. Shield III CF. Consequences of anti-OKT3 antibody development: OKT3 reuse and long-term graft survival. *Transplant Proc* 1993 Apr; 25 Suppl. 1: 81-2
  146. Hayes JM. The immunobiology and clinical use of current immunosuppressive therapy for renal transplantation. *J Urol* 1993 Mar; 149: 437-48
  147. Niaudet P, Jean G, Broyer M, et al. Anti-OKT3 response following prophylactic treatment in paediatric kidney transplant recipients. *Pediatr Nephrol* 1993 Jun; 7: 263-7
  148. Mozes MF, Venkat KK, Kupin W, et al. Is the routine use of induction immunosuppression with ALG or OKT3 justified in cadaveric renal transplantation? *Transplant Proc* 1993 Feb; 25 (Pt 1): 575-6
  149. Rossi SJ, Schroeder TJ, Hariharan S. Prevention and management of the adverse effects associated with immunosuppressive therapy. *Drug Saf* 1993 Aug; 9: 104-31
  150. Barclay PG, Allen RDM, Stewart JH, et al. Costs of immunosuppressive therapies used in renal transplantation. *Transplant Proc* 1992 Feb; 24: 165-6
  151. Mendez R, Aswad S, Khetan U, et al. Renal transplantation induction therapy – OKT3 vs MALG – outcome, cost [abstract]. *Nephrol Dial Transplant* 1995; 10 (6): 1069
  152. Abramowicz D, Crusiaux A, Goldman M. Anaphylactic shock after retreatment with OKT3 monoclonal antibody. *N Engl J Med* 1992 Sep 3; 340: 736
  153. Peces R, Urra JM, Escalada P, et al. High-dose methylprednisolone inhibits the OKT3-induced cytokine-related syndrome. *Nephron* 1993 Jan; 63: 118
  154. Costanzo-Nordin MR. Cardiopulmonary effects of OKT3: determinants of hypotension, pulmonary edema, and cardiac dysfunction. *Transplant Proc* 1993 Apr; 25 Suppl. 1: 21-4
  155. Bemelman FJ, Buysmann S, Surachno J, et al. Pretreatment with divided doses of steroids strongly decreases side effects of OKT3. *Kidney Int* 1994 Dec; 46: 1674-9
  156. Jeyarajah DR, Thistlethwaite Jr JR. General aspects of cytokine-release syndrome: timing and incidence of symptoms. *Transplant Proc* 1993 Apr; 25 (2 Suppl. 1): 16-20
  157. Abramowicz D, De Pauw L, Le Moine A, et al. Prevention of OKT3 nephrotoxicity after kidney transplantation. *Kidney Int* 1996; 49 Suppl. 53: S39-43
  158. Shield III CF, Beilman G. Safety of OKT3 use in the operating room. *Transplant Proc* 1993 Apr; 25 (2 Suppl. 1): 43-4
  159. Kupin W, Venkat KK, Ikemiyashiro D, et al. Morbidity of intraoperative OKT3 administration in primary cadaveric renal transplant recipients. *Transplant Proc* 1993 Feb; 25 (Pt 1): 572-4
  160. Robinson ST. Administration of OKT3 in the operating room. *Transplant Proc* 1993 Apr; 25 (2 Suppl. 1): 41-2
  161. Bolman III RM, Saffitz J. Early postoperative care of the cardiac transplantation patient: Routine considerations and immunosuppressive therapy. *Prog Cardiovascular Dis* 1990; 33: 137-48
  162. Hayes MJ, Torzillo PJ, Sheil AGR, et al. *Pneumocystis carinii* pneumonia after liver transplantation in adults. *Clin Transplant* 1994 Dec; 8: 499-503
  163. Hibberd PL, Tolkoff-Rubin NE, Conti D, et al. Preemptive ganciclovir therapy to prevent cytomegalovirus disease in cytomegalovirus antibody-positive renal transplant recipients. A randomized controlled trial. *Ann Intern Med* 1995 Jul 1; 123: 18-26
  164. Rubin RH, Tolkoff-Rubin N. Minireview. Antimicrobial strategies in the care of organ transplant recipients. *Antimicrob Ag Chemother* 1993; 37: 619-24
  165. Radhakrishnan J, Cohen D. Cytokine-release syndrome: General risk-factor modification – preparation of high-risk patients for use of OKT3. *Transplant Proc* 1993; 25 (2) Suppl. 1: 60-2

166. Abramowicz D, Pradier O, Marchant A, et al. Induction of thromboses within renal grafts by high-dose prophylactic OKT3. *Lancet* 1992 Mar 28; 339: 777-8
167. Woodle ES, Thistlethwaite Jr J, Emond JC, et al. OKT3 therapy for hepatic allograft rejection. Differential response in adults and children. *Transplantation* 1991 Jun; 51: 1207-12
168. Kehinde EO, Scriven SD, Feehally J, et al. Adverse effects of OKT3 therapy: increased risk with impaired renal function. *Transplant Proc* 1994 Aug; 26: 1945-7
169. Kehinde EO, Veitch PS, Scriven SD, et al. Complications of using OKT3 for induction of immunosuppression in recipients of kidneys from nonheart beating donors. *Transplant Proc* 1994 Dec; 26: 3123-5
170. Russ GR. Complications of immunosuppressive therapy in transplantation. 2: Specific immunosuppressive agents. *Med J Aust* 1992; 157: 264-7
171. Min DI, Monaco AP. Complications associated with immunosuppressive therapy and their management. *Pharmacotherapy* 1991; 11 (5): 119S-25S
172. Vasquez EM, Fabrega AJ, Pollak R. OKT3-induced cytokine-release syndrome: occurrence beyond the second dose and association with rejection severity. *Transplant Proc* 1995 Feb; 27: 873-4
173. Turner MC, Holman Jr JM. Late reactions during initial OKT-3 treatment. *Clin Transpl* 1993 Feb; 7 (Pt 1): 1-3
174. Kirklin JK, Naftel DC, Levine TB, et al. Cytomegalovirus after heart transplantation. Risk factors for infection and death: a multiinstitutional study. *J Heart Lung Transplant* 1994 May-Jun; 13: 394-404
175. Hibberd PL, Tolkoff-Rubin NE, Cosimi AB, et al. Symptomatic cytomegalovirus disease in the cytomegalovirus antibody seropositive renal transplant recipient treated with OKT3. *Transplantation* 1992 Jan; 53: 68-72
176. Hooks MA, Perlino CA, Henderson JM, et al. Prevalence of invasive cytomegalovirus disease with administration of muromonab CD-3 in patients undergoing orthotopic liver transplantation. *Ann Pharmacother* 1992 Apr; 19 Suppl. 33: 617-20
177. Tejada F, Gomez E, Aguado S, et al. OKT3 treatment induces higher prevalence and greater severity of cytomegalovirus disease in renal transplant [abstract]. *Kidney Int* 1994 Aug; 46: 582
178. Johnson MR, Mullen GM, O'Sullivan EJ, et al. Risk/benefit ratio of perioperative OKT3 in cardiac transplantation. *Am J Cardiol* 1994 Aug 1; 74: 261-6
179. Bailey TC, Powderly WG, Storch GA, et al. Symptomatic cytomegalovirus infection in renal transplant recipients given either Minnesota antilymphoblast globulin (MALG) or OKT3 for rejection prophylaxis. *Am J Kidney Dis* 1993 Feb; 21: 196-201
180. Waser M, Maggiorini M, Lüthy A, et al. Infectious complications in 100 consecutive heart transplant recipients. *Eur J Clin Microbiol Infect Dis* 1994 Jan; 13: 12-8
181. Swinnen LJ, Costanzo-Nordin MR, Fisher SG, et al. Increased incidence of lymphoproliferative disorder after immunosuppression with the monoclonal antibody OKT3 in cardiac-transplant recipients. *N Engl J Med* 1990 Dec 20; 323: 1723-8
182. Burtin P, Boman F, Pinelli G, et al. Cancers following thoracic organ transplantation: a single center study. *Transplant Proc* 1995 Apr; 27: 1765
183. Stempfle HU, Mudra H, Angermann CE, et al. Rapid growth of cutaneous neuroendocrine (Merkel cell) carcinoma during treatment of refractory cardiac allograft rejection with OKT3 monoclonal antibody. *J Heart Lung Transplant* 1993 May-Jun; 12: 501-3
184. Penn I. Cancers complicating organ transplantation. *N Engl J Med* 1990 Dec 20; 323: 1767-9
185. Reyes CV, Jensen J, Chinoy M. Pulmonary lymphoma in cardiac transplant patients treated with OKT3 for rejection: diagnosis by fine-needle aspiration. *Diagn Cytopathol* 1995 Feb; 12: 32-6
186. Rinde-Hoffman D, Cintron GB, Ferguson JE, et al. Lymphoproliferative disorder early after cardiac transplantation [published erratum appears in *Am J Cardiol* 1992 Mar 15;69(8):844]. *Am J Cardiol* 1991 Dec 15; 68: 1724-5
187. Morgan G, Superina RA. Lymphoproliferative disease after pediatric liver transplantation. *J Pediatr Surg* 1994 Sep; 29: 1192-6
188. Opelz G, Henderson R. Incidence of non-Hodgkin lymphoma in kidney and heart transplant recipients. *Lancet* 1993 Dec 18-25; 342: 1514-6
189. Emery RW, Lake KD. Post-transplantation lymphoproliferative disorder and OKT3 [letter]. *N Engl J Med* 1991 May 16; 324: 1437
190. Hardy MA, Benvenisty AI, Cohen D, et al. Incidence of malignancies in renal and cardiac transplant recipients in relation to treatment with biologic agents. *Clin Transplantation* 1992; 6 Special issue: 269-71
191. Haverty TP, Sanders M, Carey G. A 2-year follow-up of post-transplantation malignancy in renal allograft recipients receiving muromonab-CD3 for immunosuppressive induction therapy. *Drug Invest* 1992; 4 (5): 403-8
192. Jones C, Bleau B, Buskard N, et al. Simultaneous development of diffuse immunoblastic lymphoma in recipients of renal transplants from a single cadaver donor: transmission of Epstein-Barr virus and triggering by OKT3. *Am J Kidney Dis* 1994 Jan; 23: 130-4
193. Cockfield SM, Preiksaitis JK, Jewell LD, et al. Post-transplant lymphoproliferative disorder in renal allograft patients. Clinical experience and risk factor analysis in a single center. *Transplantation* 1993 Jul; 56: 88-96
194. Canfield CW, Hudnall SD, Colonna JO, et al. Fulminant Epstein-Barr virus-associated post-transplant lymphoproliferative disorders following OKT3-therapy. *Clin Transpl* 1992 Feb; 6: 1-9
195. Ratkovec RM, O'Connell JB, Bristow MR, et al. Post-transplant lymphoproliferative disease in cardiac transplant patients receiving OKT3 therapy. *Clin Transpl* 1992 Jun; 6: 260-4
196. Morrissey PE, Lorber KM, Marcarelli M, et al. Posttransplant Epstein-Barr virus infection is associated with elevated levels of CD19<sup>+</sup> B lymphocytes. *Transplantation* 1995 Feb 27; 59: 637-40
197. Bernstein D, Baum D, Berry G, et al. Neoplastic disorders after pediatric heart transplantation. *Circulation* 1993 Nov; 88 (Pt 2): 230-7
198. Lee CW, Logan JL, Zukoski CF. Cardiovascular collapse following orthoclone OKT3 administration: a case report. *Am J Kidney Dis* 1991 Jan; 17: 73-5
199. Hall KA, Dale EJ, Hunter GC, et al. Hyperpyrexia-related ventricular tachycardia during OKT3 induction therapy. *Transplantation* 1992 Dec; 54: 1112-3
200. Coleman AE, Norman DJ. OKT3 encephalopathy. *Ann Neurol* 1990 Dec; 28: 837-8
201. Marks WH, Perkal M, Bia M. Aseptic encephalitis and blindness complicating OKT3 therapy. *Clin Transpl* 1991 Dec; 5 (Pt 1): 435-8
202. Shihab FS, Barry JM, Norman DJ. Encephalopathy following the use of OKT3 in renal allograft transplantation. *Transplant Proc* 1993 Apr; 25 Suppl. 1: 31-4

203. Alegre M-L, Depierreux M, Florquin S, et al. Nephrotoxicity of anti-CD3 monoclonal antibodies [in French]. *Nephrologie* 1991; 12: 42-5
204. Batiuk TD, Bennett WM, Norman DJ. Cytokine nephropathy during antilymphocyte therapy. *Transplant Proc* 1993 Apr; 25 Suppl. 1: 27-30
205. First MR, Schroeder TJ, Hariharan S. OKT3-induced cytokine-release syndrome: renal effects (cytokine nephropathy). *Transplant Proc* 1993 Apr; 25 Suppl. 1: 25-6
206. Dussol B, Brunet P, Vacher-Coponat H, et al. Haemolytic uraemic syndrome in a renal transplant recipient during OKT3 therapy. *Nephrol Dial Transplant* 1994; 9 (8): 1191-3
207. Doutrelepont J-M, Abramowicz D, Florquin S, et al. Early recurrence of hemolytic uremic syndrome in a renal transplant recipient during prophylactic OKT3 therapy. *Transplantation* 1992; 53: 1378-9
208. Bemelman FJ, Buysmann S, Yong SL, et al. Biphasic granulocytopenia after administration of the first dose of OKT3 [abstract]. *Kidney Int* 1995 Apr; 47: 1218
209. Burke III GW, Vercellotti GM, Simmons RL, et al. Reversible pancytopenia following OKT3. Use in the context of multi-drug immunosuppression for kidney allografting. *Transplantation* 1989 Sep; 48: 403-7
210. Vasquez EM, Pollak R. Effect of OKT3 therapy on cyclosporine blood levels [abstract]. *Pharmacotherapy* 1995 Jan-Feb; 15: 118-9
211. Chan GL, Weinstein SS, Wright CE, et al. Encephalopathy associated with OKT3 administration. Possible interaction with indomethacin. *Transplantation* 1991 Jul; 52: 148-50
212. Doutrelepont JM, Abramowicz D, Borre B, et al. Prophylactic OKT3: practical considerations for the prevention of first-dose reactions. *Transplant Proc* 1993 Apr; 25 Suppl. 1: 45-6
213. Dantal J, Soulillou J-P. Use of monoclonal antibodies in human transplantation. *Curr Opin Immunol* 1991 Oct; 3: 740-7
214. Krensky AM, Clayberger C. Transplantation immunology. *Pediatr Clin North Am* 1994 Aug; 41: 819-39

---

Correspondence: *Michelle I. Wilde*, Adis International Limited, 41 Centorian Drive, Private Bag 65901, Mairangi Bay, Auckland 10, New Zealand.

---

## Erratum

**Vol. 51, No. 2, page 314:** In section 5.2, the second sentence in the third paragraph should read: 'Dialyser clearances of urea . . . (mean dosage 4335IU 3 times weekly) . . .'.

[*Dunn CJ, Markham A. Epoetin beta: a review of its pharmacological properties and clinical use in the management of anaemia associated with chronic renal failure. Drugs* 1996 Feb; 51 (2): 299-318]