process. Apatite-like nanoparticles were observed on the MBG–PCL surface after soaking for 4 h, and soaking for 24 h resulted in a MBG–PCL fully covered with apatite. Likewise, energy dispersive x-ray (EDX) analysis gave the same result. The biocompatibility of the MBG–PCL was evaluated by MTT [3-(4,5-Dimethylthiazol-z-YI)-2,5-Diphenyltetrazolium Bromide] assay. The cell viability on the MBG–PCL surface was found to be significantly larger than that on the PCL surface. These studies demonstrate excellent bioactivity and biocompatibility of this MBG– PCL scaffold, the researchers said.

The method developed in this work was used to fabricate a scaffold for tissue engineering. However, by utilizing alternative materials, the researchers showed that this method can be used in "applications involving biomedical devices, drug delivery systems, filters, catalysis, and optics."

CHANG ZHONG

Atomic Force Microscopy Used to Detect Cancer Cells

Immunohistochemical and cytomorphological analyses are currently used to detect cancer cells, but morphological overlap between tumor and normal cell types often poses problems for these techniques. In the past few years, however, an increase in cell elasticity has been recognized as a marker for disease and associated with cell adhesion and cytoskeletal organization. S.E. Cross and J.K. Gimzewski at the California NanoSystems Institute and the University of California, Los Angeles, and Y.-S. Jin and J. Rao at the University of California, Los Angeles used atomic force microscopy (AFM) to distinguish cancerous cells from normal cells.

As reported in a letter published in the December 2, 2007 issue of Nature Nanotechnology (p. 780; DOI:10.1038/nnano. 2007.388), the researchers used standard protocols to collect from patients with suspected metastatic adenocarcinoma samples of malignant and benign mesothelial cells in pleural effusions (adenocarcinoma is a cancer that originates in glandular tissue; mesothelial cells form part of the membranes covering body cavities, and an effusion is an abnormally large collection of fluid in the space surrounding the lungs). Ex vivo growth during a 12-h incubation period differentiated normal cells, which have a large, flat morphology, from benign cells, which exhibit anchorage-resistant morphology such as rounding. For each cell, elasticity was quantified as Young's modulus E from force-displacement curves recorded from AFM performed at a rate of 1 Hz at 37°C. Using samples from seven patients, average E values of 0.53 \pm 0.10 kPa and 1.97 ± 0.70 kPa were obtained for 40 malignant and 48 benign mesothelial cells, respectively. Similar average *E* values were found for samples from a single patient, showing that the cell stiffness of metastatic cancer cells is about 73 ± 11% less than benign mesothelial cells. Malignant cells displayed a narrow distribution of *E* values while the benign cells displayed a broad peak.

For one particular clinical sample consisting of cells that were difficult to classify as either benign or malignant, ex vivo culturing resulted in two populations of cells with different morphologies. However, nanomechanical analysis based on AFM measurements determined that both populations were malignant cells, which was confirmed with immunohistochemical staining. The researchers said that "the correlation of cytomechanical measurements with immunohistochemical analysis suggests that nanomechanical measurements of cancer cells has potential for the detection of cancer and may aid in personalized selection of medication and drug screening, especially in body cavity effusions where accurate diagnosis based solely on morphology has to date been challenging." STEVEN TROHALAKI

Embedded Capacitive Circuit Detects, Characterizes, and Manipulates Droplets in Microfluidic Systems

Droplets in microfluidic systems are presently detected using optical devices, such as single element photodiodes for counting or high-speed charge-coupled device image sensors with imageprocessing software for imaging. A group of researchers from The Hong Kong University of Science and Technology reported in the October-December 2007 issue of Biomicrofluidics (DOI: 10.1063/1.2795392) on a novel method for real-time detecting, characterizing, and controlling of such droplets through the use of capacitive sensors; the dielectric constant contrast between the fluid of interest and the carrying fluid is the only prerequisite for the method functionality.

The research group, including X. Niu, M. Zhang, S. Peng, W. Wen, and P. Sheng, designed a microfluidic chip that embeds from the fabrication process (using soft lithography) a series of parallel electrodes placed across the microfluidic channel, some of which are connected to a capacitance resonance detection system and others to a square wave signal generator for *in situ* manipulation of the droplets.

The researchers were able to calibrate the device for measurements involving length (and subsequently volume) of nanoand picoliter range aqueous droplets in silicone oil by analyzing the resonance circuit voltage changes as the droplets were passing through the electrodes. Very good accuracy was obtained for droplet lengths bigger than half of the electrode width, and a detection frequency of more than 1 kHz (1100 droplets per second) has been attained, limited in this case only by the sustainable pressure of the chip and not by the detection system. The droplets velocity was also easily calculated by measuring the time required to reach from one set of electrodes to the other.

The system was subsequently calibrated and employed to detect droplets of different composition (ethylene glycol and, respectively, water) and to sort them in two different branches of a T-shaped microchannel using a feedback system that employs a set of electrodes with a positive high potential to charge the droplets and another set that can preferentially change polarity to direct the droplets through either of the branches. The delay time of the feedback system was adjusted for the droplet's velocity so that a perfect match was obtained between detection and manipulation.

The researchers demonstrated that their method of real-time labeling, sorting, and manipulation of droplets can provide a very simple and cost-effective alternative for portable devices involved in biomicrofluidic processing, digital microfluidics, microchemical reactions, and environmental monitoring.

EUGEN PANAITESCU

Quasi-Forbidden Bragg Peaks Demonstrated as Inherent Property in Homogeneous Ordered Soft Materials

Bragg peaks exist in highly ordered soft materials that cannot be indexed assuming homogeneous crystal structures and that have been attributed to changes in the crystal structure induced by the ordering process. S. Förster and co-workers from the University of Hamburg, Germany, together with S.V. Roth from DESY, Germany and P. Lindner from the Institute Laue-Langevin, France, have demonstrated that these unexpected Bragg peaks are an inherent property of homogeneous ordered soft materials and arise from the spontaneous ordering allowed by the weak interaction potential that tolerates imperfections of the constituent structures that limit the coherence of the crystalline lattice. Their results have been reported in the November 2007 issue of Nature Materials (p. 888; DOI: 10.1038/ nmat1995). "This explained the presence of quasi-forbidden Bragg peaks in lyotropic liquid-crystalline phases, mesoporous materials, colloidal dispersions, block copolymers, electrorheological fluids and photonic crystals," the researchers said.

One would expect in such materials Debye-Scherrer rings corresponding to (hkl) lattice planes forbidden in a particular direction to gradually disappear with ordering, whereas those corresponding to allowed (hkl) planes to gradually develop Bragg peaks. However, calculations of the diffraction patterns performed by the researchers showed the existence of a broad intermediate state, where all Debye-Scherrer rings, even those corresponding to forbidden (hkl) planes, developed Bragg peaks. The researchers explained the origin of these quasi-forbidden Bragg peaks by using the Ewald sphere construction with peaks with a width broad enough to partly fulfil the Bragg condition, even if they are located at a position that would not diffract. For soft materials with large unit cells, the presence of these Bragg peaks is more a rule than an exception because of the high ratio of peak broadening with respect to peak position and to the usual multidomain structure of such materials, the researchers said.

The researchers confirmed experimentally those calculations by determining the domain structure by rotating-crystal synchrotron small-angle x-ray scattering and scanning electron microscopy (SEM) of a silica replica of a shear-aligned lyotropic liquid-crystalline fcc phase prepared by swelling it with a pre-hydrolysed solution of tetramethoxysilane, which hydrolysed and condensed into amorphous silica, and calcining it to obtain the mesoporous silica crystal, replica of the initial structure. Before shear orientation, the sample was isotropic and exhibited Debye-Scherrer rings. After a shear rate of $\gamma = 100 \text{ s}^{-1}$, the sample developed a diffraction pattern with more than 50 Bragg peaks characteristic for shear-oriented fcc structures. SEM images revealed noticeable deviations from a perfect single-crystalline structure with well-ordered domains 50-200 nm in size and mismatches at their boundaries, misorientations to adjacent domains, and local bending of the lattice planes that limited the coherence of the lattice.

"These results demonstrate that soft materials do not represent a particularly low state of order, instead, the order was not perfect enough to suppress quasiforbidden Bragg peaks in diffraction experiments," the researchers said.

Based on these results, the researchers showed that with this structural information, they could construct a simple picture of how the line density of a structure was linked to shear orientation and epitaxial relations.

JOAN J. CARVAJAL

Don't Miss MegaMONDAY at the 2008 MRS Spring Meeting For more information on MegaMONDAY see page 100



Janis Research Company 2 Jewel Drive Wilmington, MA 01887 USA TEL +1 978 657-8750 FAX +1 978 658-0349 sales@janis.com Visit our website at www.janis.com.



www.mmr.com

MMR