

## Genetic Mapping of Gibberellic Acid-sensitive Genes for Semi-dwarfism in Durum Wheat

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Semi-dwarf varieties in wheat associated with gibberellic acid ( $GA_3$ )-insensitive height reducing genes have led to significant increases in yield but often fall below this potential because of poor seedling emergence after deep sowing. Alternative semi-dwarf genes may have the potential to reduce plant height without compromising early plant growth. In durum wheat, bulk segregant analysis was used to screen microsatellite markers linked with the  $GA_3$ -sensitive genes *Rht14* in cv. Castelporziano, *Rht16* in cv. Edmore M1 and *Rht18* in cv. Icaro. Molecular marker *Xbarc3-6A* for *Rht14*, *Rht16* and *Rht18* showed significant polymorphic differences among DNA bulks for height classes. The genes *Rht14*, *Rht16* and *Rht18* were linked with *Xbarc3* (11.7–28.0 cM) on the short arm of chromosome 6A and they appear to be allelic. Semi-dwarf genes on chromosome 6AS may potentially be used in breeding for improved establishment.

**Keywords:** *Triticum durum*,  $GA_3$ -sensitive, plant height, reduced-height

### Introduction

The introduction of stem-shortening genes into commercial bread wheat (Perry and D'Antuono 1989) and durum wheat (Waddington et al. 1987) has led to substantial yield increases per unit area. The dwarfing genes *Rht-B1b* (*Rht1*) and *Rht-D1b* (*Rht2*) from *Triticum aestivum* cv. Norin 10 resulted in an increased harvest index and reduced risk of lodging under high nitrogen fertilization (Evans 1993). Although these genes can lead to yield increases in favorable environments, they have been associated with yield reduction in unfavorable environments (Laing and Fisher 1977; Richards 1992). For successful crop establishment, it is necessary to achieve a high proportion of emerged plants relative to the number of seeds sown. This has become an important issue in wheat because crop establishment is poor when seeds of varieties containing the gibberellic acid ( $GA_3$ )-insensitive dwarfing genes *Rht-B1b* and *Rht-D1b* are sown deep to gain early access to deep soil moisture and when conservation farming systems and stubble retention practices have

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been adopted (Schillinger et al. 1998). These GA<sub>3</sub>-insensitive varieties have short coleoptiles and will not establish well if sown too deep. Addisu et al. (2009) reported that the GA<sub>3</sub>-sensitive *Rht12* and *Rht8* were not associated with reduced coleoptile length. It has been suggested that lines containing dwarfing genes that are responsive to applied GA<sub>3</sub> and have longer coleoptiles will emerge more successfully when sown deep or when used in conservation farming systems (Schillinger et al. 1998; Rebetzke et al. 2005, 2007).

A number of accessions containing alternative dwarfing genes (i.e. unrelated to *Rht1* and *Rht2*) are known in wheat. They reduce plant height to varying degrees, and are sensitive to GA<sub>3</sub>. Semi-dwarf wheats containing these genes produce long coleoptiles, which assist rapid germination and plant establishment. Therefore, there is potential to utilize these semi-dwarf genes in wheat breeding programs (Rebetzke et al. 1999). Ellis et al. (2004) assessed the effect of different height-reducing genes on the early growth of wheat, and Ellis et al. (2005) mapped GA<sub>3</sub>-sensitive dwarfing genes in bread wheat (*T. aestivum*) on chromosomes 3BS (*Rht5*), 5AL (*Rht12*), 7BS (*Rht13*), 2BL (*Rht4*), 2DS (*Rht8*) and 5AL (*Rht9*). Semi-dwarf durum wheat accessions, var. Castelporziano, Edmore M1 and Icaro, were identified as GA<sub>3</sub>-sensitive (Ellis et al. 2004). These accessions were expected to have the potential to produce long coleoptiles and to be utilized in breeding tetraploid and hexaploid wheats. However, no map locations were reported for the genes conferring semi-dwarf stature in these accessions.

Molecular markers are widely used for mapping genes of agricultural importance. Microsatellite markers have been widely used in such applications in wheat. Numerous polymorphic microsatellite markers have been integrated into genetic framework maps (Röder et al. 1998; Song et al. 2005; Torada et al. 2006). The aim of the present study was to map GA<sub>3</sub>-sensitive genes for semi-dwarfism in three durum wheat accessions.

### Materials and Methods

The three reduced height sources studied here were GA<sub>3</sub>-sensitive. *T. durum* mutant line Edmore M1 (PI 499362) was derived from cv. Edmore by mutagenesis with methylnitrourea (C.F. Konzak, Washington State University, USA). *T. durum* cv. Castelporziano (PI 347731) was derived from mutagenesis of cv. Capelli, whereas cv. Icaro (PI 503555) was obtained by fast-neutron treatment of cv. Anhinga. The semi-dwarf genes in these materials were designated as *Rht14* for Castelporziano (Gale et al. 1985), *Rht16* for Edmore M1 (Konzak et al. 1984), and *Rht18* for Icaro (Konzak 1988). The mean culm lengths of mutant lines Castelporziano, Edmore M1 and Icaro were 73.8 ± 4.2 cm, 76.8 ± 4.5 cm, and 63.5 ± 3.4 cm, respectively (measured after ripening). The culm length of *T. durum* cv. LD222, which does not carry any known reduced height alleles, was 150 ± 8.1 cm.

All materials for study were grown in the experimental field at the College of Agriculture, Ibaraki University, Japan. Culm lengths of individual genotypes were measured after ripening. F<sub>2</sub> hybrid populations of Castelporziano × LD222 and Icaro × LD222 for genetic analysis were sown in October 2006 and those of Edmore M1 × LD222 in October 2007.

### *Bulk segregant analysis*

Bulk segregant analysis (BSA) (Michelmore et al. 1991) was used to identify markers linked to the GA<sub>3</sub>-sensitive reduced-height genes in specific chromosomal regions. Pooled DNA samples of the semi-dwarf and tall progeny from the F<sub>2</sub> mapping populations were screened for polymorphisms using 87 microsatellite markers known to be located on A or B genome chromosomes (*Triticum aestivum* is a hexaploid containing genomes A, B and D, whereas *T. durum* is tetraploid and contains only genomes A and B). DNA samples from the parents, six individual semi-dwarf F<sub>2</sub> plants, and six individual tall F<sub>2</sub> plants with heights similar to the respective parents were also used to check the amplification characteristics of the linked markers. Markers close to a semi-dwarfing gene should be polymorphic between both parents and bulks. If unlinked, a similar banding pattern will be seen in each of the two bulks.

### *Allelic relationships of the Rht genes*

Castelporziano, Edmore M1 and Icaro were crossed with each other to test for allelism among the *Rht14*, *Rht16* and *Rht18* genes using the F<sub>2</sub> populations.

### *Microsatellite mapping of the GA<sub>3</sub>-sensitive Rht genes*

Genomic DNA was extracted from seedlings of 94 individuals per F<sub>2</sub> hybrid population according to the method of Dellaporta et al. (1983). The microsatellite markers *Xgwm*, *Xbarc* and *Xhbg* located on chromosomes 6A were chosen based on the results of BSA and the potentially allelic relationship detected between the semi-dwarf genes under study. These markers were obtained from Röder et al. (1998), Song et al. (2005), and Torada et al. (2006), respectively. PCR analysis was performed as described by Kosuge et al. (2008). Linkage values in centiMorgans (cM) were calculated using Map Manager QTX (<http://mapmgr.roswellpark.org/>). Minimum LOD scores of at least 3.0 were used to develop the linkage map. The software calculated genetic distances in cM using the Kosambi (1944) mapping function.

## **Results**

### *Allelism test*

No tall plants were observed in F<sub>2</sub> populations derived from the three crosses between Castelporziano, Edmore M1 and Icaro (n = 152–158 for each F<sub>2</sub> population), thereby indicating that *Rht14*, *Rht16* and *Rht18* are allelic.

### *Chromosomal location of GA<sub>3</sub>-sensitive genes*

Figure 1 shows the segregation of culm length in F<sub>2</sub> populations of Castelporziano × LD222, Edmore M1 × LD222, and Icaro × LD222. The culm lengths of three F<sub>2</sub> hybrid populations were classified 94 semi-dwarf: 25 tall for the F<sub>2</sub> of Castelporziano × LD222, 94 semi-dwarf: 40 tall for the F<sub>2</sub> of Edmore M1 × LD222 and 108 semi-dwarf : 26 tall for

the F<sub>2</sub> of Icaro × LD222. The observed segregation ratios in all three F<sub>2</sub> populations fit a model of 3 semi-dwarf: 1 tall ( $\chi^2$  values of 1.011, 1.681 and 2.239, respectively; df = 1). The semi-dwarf mutant lines were, on average, 50% shorter than the LD222.

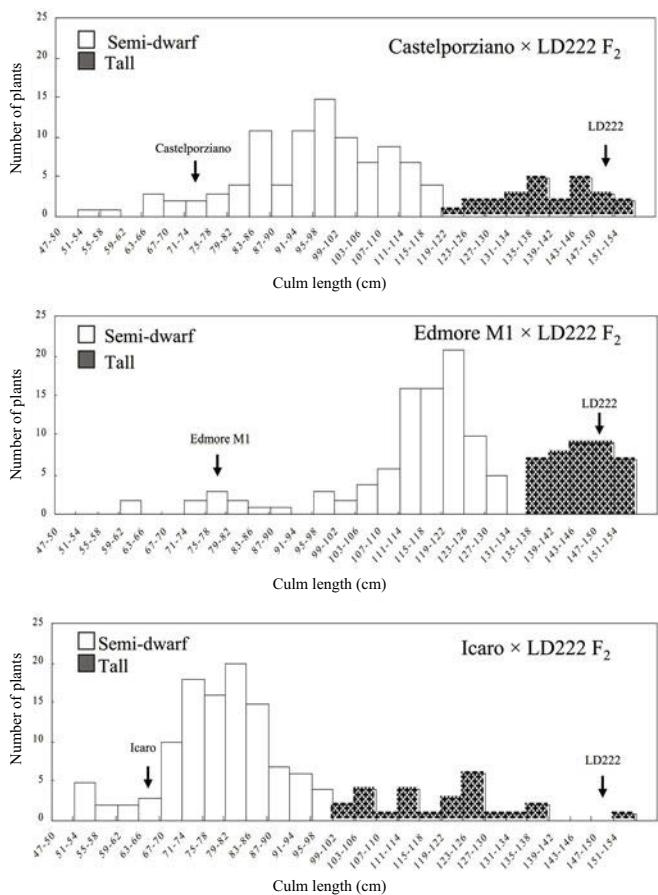


Figure 1. Segregation of culm length in F<sub>2</sub> populations of Castelporziano × LD222, Edmore M1 × LD222 and Icaro × LD222. Plant height was categorized as semi-dwarf (white bars) or tall (grey bars). Arrows indicate mean culm length of the parents

The distributions in culm length shown in Figure 1 indicate that the semi-dwarf phenotypes were incompletely dominant. Selection of more extreme phenotypes for BSA resulted in plants homozygous for the contrasting alleles being chosen.

Amplification using microsatellite marker *Xbarc3*, located on chromosome 6A, showed polymorphism between the parents and bulks as well as individuals in BSA (Fig. 2). Based on the results of the BSA, 22 microsatellite markers on chromosome 6A were

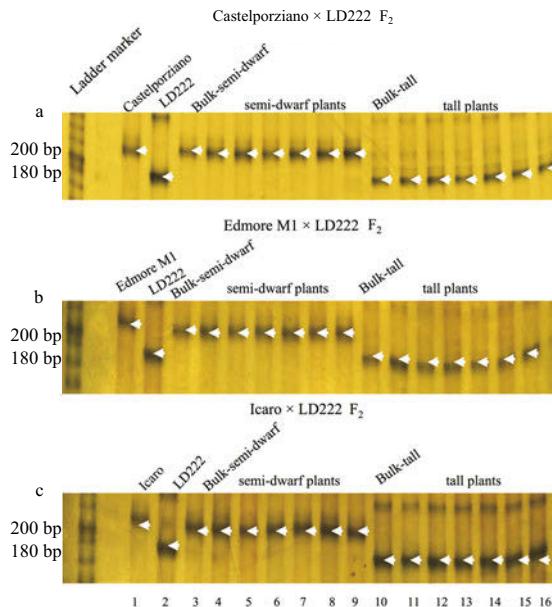


Figure 2. Amplification profiles of microsatellite marker *Xbarc3-64* by bulk segregant analysis in  $F_2$  populations of (a) Castelporziano × LD222, (b) Edmore M1 × LD222 and (c) Icaro × LD222.

*Left to right:* Semi-dwarf parent (lane 1), LD222 (lane 2), bulk of semi-dwarf plants (lane 3), individual semi-dwarf  $F_2$  plants (lanes 4 to 9), bulk of tall plants (lane 10), and individual tall  $F_2$  plants (lanes 11 to 16). Arrows indicate bands in common between a given parent, individual homozygous progeny with the same phenotype as that parent and the corresponding bulk

used to map the *Rht14*, *Rht16* and *Rht18* genes. Markers *Xbarc3*, *Xbarc171* and *Xgwm256* were polymorphic in all three combinations.

*Rht14*, *Rht16* and *Rht18* each mapped distal to marker *Xbarc3* on the short arm of chromosome 6A (Fig. 3). These mapping data provide further evidence for an allelic relationship between *Rht14*, *Rht16* and *Rht18*.

## Discussion

The GA<sub>3</sub>-insensitive semi-dwarf genes *Rht-B1b* and *Rht-D1b* produce short coleoptiles, which may reduce seedling vigour and crop yield stability (Richards 1992; Rebetzke et al. 1999). It was proposed that GA<sub>3</sub>-sensitive semi-dwarf genes could resolve these problems in durum wheat (Giorgi et al. 1984). Rebetzke et al. (2004) reported that coleoptile lengths of progeny derived from GA<sub>3</sub>-sensitive *Rht8*, *Rht9* and *Rht12* semi-dwarf genes were an average 47% longer coleoptiles than comparative materials with *Rht-B1b* and *Rht-D1b*, thus offering the potential of longer coleoptiles as well as reduced height for deep sowing. The present study focused on mapping three GA<sub>3</sub>-sensitive semi-dwarfing genes that came from different mutation events. *Rht14*, *Rht16* and *Rht18* were located to similar positions on chromosome 6AS and tests of allelism based on relatively small populations of

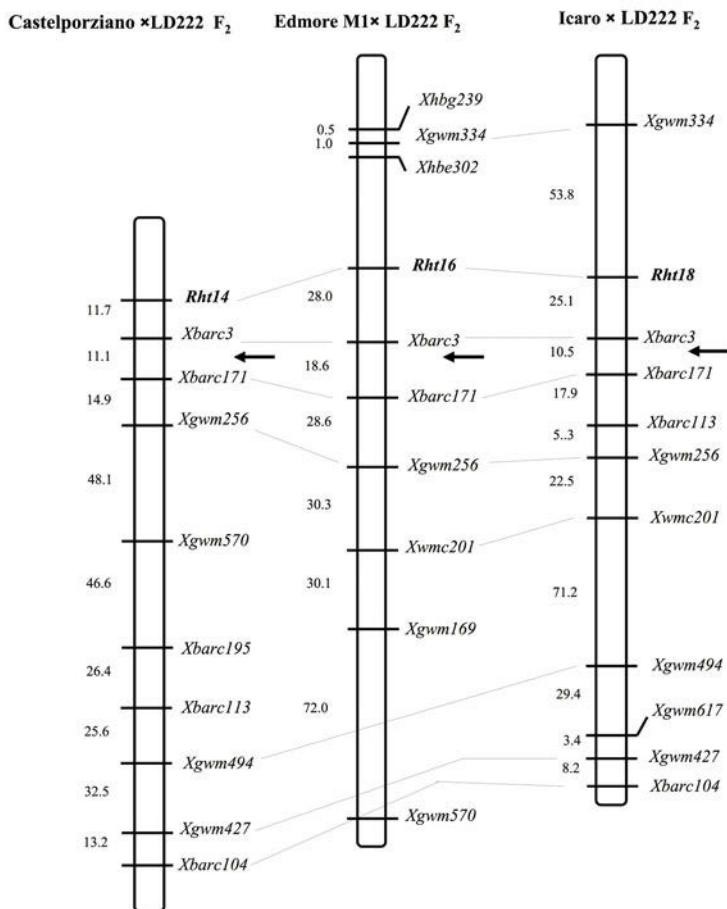


Figure 3. Linkage maps for *Rht14*, *Rht16* and *Rht18* on chromosome 6A. Arrows indicate the supposed position of the centromere. Distances between markers are shown in cM

$F_2$  plants indicated the possibility of allelism. Spielmeyer et al. (2007) located a QTL for coleoptile length on chromosome 6AS, and they indicated that the QTL on this region promoted coleoptile length during early plant growth. Further investigation is needed to assess the potential of these semi-dwarf genes in dryland wheat production.

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### References

- Addisu, M., Snape, J.W., Simmonds, J.R., Gooding, M.J. 2009. Reduced height (*Rht*) and photoperiod insensitivity (*Ppd*) allele associations with establishment and early growth of wheat in contrasting production system. *Euphytica* **166**:249–267.
- Dellaporta, S.L., Wood, J., Hicks, J.B. 1983. A plant DNA mini preparation: Version II. *Plant Mol. Biol. Rep.* **1**:19–21.
- Ellis, M.H., Rebetzke, G.J., Azanza, F., Richards, R.A., Spielmeyer, W. 2005. Molecular mapping of gibberellin-responsive dwarfing genes in bread wheat. *Theor. Appl. Genet.* **111**:423–430.
- Ellis, M.H., Rebetzke, G.J., Chandler, P., Bonnett, D.G., Spielmeyer, W., Richards, R.A. 2004. The effect of different height reducing genes on the early growth of wheat. *Funct. Plant Biol.* **31**:583–589.
- Evans, L.T. 1993. *Crop Evolution, Adaptation, and Yield*. Cambridge University Press, Cambridge, UK.
- Gale, M.D., Youssefian, S. 1985. Dwarfing genes in wheat. In: Russell, G.E. (ed.), *Progress in Plant Breeding*. Butterworths, London, UK, pp. 1–35.
- Giorgi, B., Barbera, F., Bitti, O., Cavicchioni, G. 1984. Yield performance of  $F_3$  progenies from a durum wheat involving two different semidwarfing genes: *Rhl1* and *Sd* mutation. In: *Semi-dwarf Cereal Mutants and Their Use in Cross-breeding II*. Research Coordination Meeting 1982. IAEA, Vienna, Austria, pp. 91–95.
- Konzak, C.F. 1988. Genetic analysis, genetic improvement and evaluation of induced semi-dwarf mutants in wheat. In: *Semi-dwarf Cereal Mutants and Their Use in Cross-breeding III*. Research Coordination Meeting 1985. IAEA, Vienna, Austria, pp. 77–94.
- Konzak, C.F., Wilson, M.R., Franks, P.A. 1984. Progress in the evaluation, use in breeding, and genetic analysis of semidwarf mutants of wheat. In: *Semi-dwarf Cereal Mutants and Their Use in Cross-breeding II*. Research Coordination Meeting 1982. IAEA, Vienna, Austria, pp. 39–50.
- Kosambi, D.D. 1944. The estimation of map distances from recombination values. *Ann. Eugen.* **12**:172–175.
- Kosuge, K., Watanabe, N., Kuboyama, T., Melnik, V.M., Yanchenko, V.I., Rosova, M.A., Goncharov, N.P. 2008. Cytological and microsatellite mapping of mutant genes for spherical grain and compact spikes in durum wheat. *Euphytica* **159**:289–296.
- Laing, D.R., Fischer, R.A. 1977. Adaptation of semidwarf wheat cultivars to rainfed conditions. *Euphytica* **26**:129–139.
- Michelmore, R.W., Para, I., Kesselli, R.V. 1991. Identification of markers linked to disease-resistance genes by bulked segregant analysis – a rapid method to detect markers in specific genomic regions by using segregating populations. *Proc. Nat. Acad. Sci. USA* **88**:9828–9832.
- Perry, M.W., D'Antuono, M.F. 1989. Yield improvement and associated characteristics of some Australian spring wheat cultivars introduced between 1860 and 1982. *Aust. J. Agric. Res.* **40**:457–472.
- Rebetzke, G.J., Richards, R.A., Fischer, V.M., Mickelson, B.J. 1999. Breeding long coleoptile, reduced height wheats. *Euphytica* **106**:158–168.
- Rebetzke, G.J., Richards, R.A., Sirault, X.R.R., Morrison, A.D. 2004. Genetic analysis of coleoptile length and diameter of wheat. *Aust. J. Agric. Res.* **55**:733–743.
- Rebetzke, G.J., Bruce, S.E., Kirkegaard, J.A. 2005. Longer coleoptiles improve emergence through crop residues to increase seedling number and biomass in wheat (*Triticum aestivum* L.). *Plant Soil* **272**:87–100.
- Rebetzke, G.J., Richards, R.A., Fettell, N.A., Long, M., Condon, A.G., Forrester, R.I., Botwright, T.L. 2007. Genotypic increases in coleoptile length improves stand establishment, vigour and grain yield of deep-sown wheat. *Field Crops Res.* **100**:10–23.
- Richards, R.A. 1992. The effect of dwarfing genes in spring wheat in dry environments. II. Growth, water use and water use efficiency. *Aust. J. Agric. Res.* **43**:529–539.
- Röder, M.S., Korzun, V., Gill, B.S., Ganap, M.W. 1998. The physical mapping of microsatellite markers in wheat. *Genome* **41**:278–283.
- Schillinger, W.F., Donaldson, E., Allan, R.E., Jones, S.S. 1998. Winter wheat seedling emergence from deep sowing depths. *Agron. J.* **90**:582–586.

- Song, Q.J., Shi, J.R., Singh, S., Fikus, E.W., Costa, J.M., Lewis, J., Gill, B.S., Ward, R., Cregan, P.B. 2005. Development and mapping of microsatellite (SSR) markers in wheat. *Theor. Appl. Genet.* **110**:550–560.
- Spielmeyer, W., Hyles, J., Joaquim, P., Azanza, F., Bonnett, D., Ellis, M.E., Moore, C., Richards, R.A. 2007. A QTL on chromosome 6A in bread wheat (*Triticum aestivum*) is associated with longer coleoptiles, greater seedling vigour and final plant height. *Theor. Appl. Genet.* **115**:59–66.
- Torada, A., Koike, M., Mochida, K., Ogihara, Y. 2006. SSR-based linkage map with new markers using an intraspecific population of common wheat. *Theor. Appl. Genet.* **112**:1042–1051.
- Waddington, S.R., Osmanzai, M., Yoshida, M., Ransom, J.K. 1987. The yield of durum wheats released in Mexico between 1960 and 1984. *J. Agric. Sci. (Camb.)* **108**:469–477.