

REVIEW

Open Access



# Human Identical Sequences, hyaluronan, and hymecromone — the new mechanism and management of COVID-19

Shuai Yang<sup>1,2†</sup>, Ying Tong<sup>1,2†</sup>, Lu Chen<sup>1,2†</sup> and Wenqiang Yu<sup>1,2\*</sup>

## Abstract

COVID-19 caused by SARS-CoV-2 has created formidable damage to public health and market economy. Currently, SARS-CoV-2 variants has exacerbated the transmission from person-to-person. Even after a great deal of investigation on COVID-19, SARS-CoV-2 is still rampaging globally, emphasizing the urgent need to reformulate effective prevention and treatment strategies. Here, we review the latest research progress of COVID-19 and provide distinct perspectives on the mechanism and management of COVID-19. Specially, we highlight the significance of Human Identical Sequences (HIS), hyaluronan, and hymecromone (“Three-H”) for the understanding and intervention of COVID-19. Firstly, HIS activate inflammation-related genes to influence COVID-19 progress through NamiRNA-Enhancer network. Accumulation of hyaluronan induced by HIS-mediated *HAS2* upregulation is a substantial basis for clinical manifestations of COVID-19, especially in lymphocytopenia and pulmonary ground-glass opacity. Secondly, detection of plasma hyaluronan can be effective for evaluating the progression and severity of COVID-19. Thirdly, spike glycoprotein of SARS-CoV-2 may bind to hyaluronan and further serve as an allergen to stimulate allergic reaction, causing sudden adverse effects after vaccination or the aggravation of COVID-19. Finally, antisense oligonucleotides of HIS or inhibitors of hyaluronan synthesis (hymecromone) or antiallergic agents could be promising therapeutic agents for COVID-19. Collectively, Three-H could hold the key to understand the pathogenic mechanism and create effective therapeutic strategies for COVID-19.

**Keywords:** Human Identical Sequences, Hyaluronan, Hymecromone, COVID-19, Ground-glass opacity

## Introduction

The ongoing pandemic of coronavirus disease 2019 (COVID-19) caused by SARS-CoV-2 infection has resulted in more than 6.2 million deaths globally according to the WHO Coronavirus Dashboard by April 27, 2022. Along with the increase in global infections and deaths, the economic burden and health threats caused

by COVID-19 have been extremely acute [1–4]. In the world, there are different kinds of vaccines available for COVID-19 in various populations [5–9], including inactivated vaccines, live-attenuated vaccines, subunit vaccines, virus-like particles vaccines, viral vector-based vaccines, mRNA vaccines, and DNA vaccines. Especially, DNA vaccines are under development because of its long-term stability [10–12]. Recently, one DNA vaccine named INO-4800 against SARS-CoV-2 could stimulate durable immune responses in the phase 1 trial [13]. Unfortunately, the situation of COVID-19 pandemic is exacerbated by the emergence of SARS-CoV-2 variants. Since April 2021, the B.1.617.2 (Delta) variant, which has higher morbidity and transmissibility [14, 15]. Another B.1.1.529 (Omicron) variant was identified as the fifth

<sup>†</sup>Shuai Yang, Ying Tong and Lu Chen contributed equally to this work.

\*Correspondence: wenqiangyu@fudan.edu.cn

<sup>1</sup>Laboratory of RNA Epigenetics, Institutes of Biomedical Sciences & Shanghai Public Health Clinical Center & Department of General Surgery, Huashan Hospital, Cancer Metastasis Institute, Shanghai Medical College, Fudan University, Shanghai 200032, People's Republic of China  
Full list of author information is available at the end of the article

variants of concern (VOC) by WHO on November 26, 2021 [16]. Currently, Omicron variant of SARS-CoV-2 has caused rapid epidemic expansion in many countries [17–20]. Specially, the mutational sites of receptor binding domain (RBD) regions in Omicron variant leads to its widespread escape from the responses of neutralizing antibodies [21–24]. Although vaccines theoretically prevent the transmission and infection of SARS-CoV-2 and are considered by the many to be the ultimate weapon against COVID-19 [14, 25–28], an increasing number of confirmed COVID-19 patients are alarmingly also vaccinated [15, 29–31]. Besides, some adverse reactions are reported in individuals vaccinated against COVID-19 [32], including myocarditis [33–36], thrombosis [37–41], adenopathy [42–44], abnormal cutaneous manifestations [45–47], and so on. Therefore, current circumstances indicate that there is still a long way to go to overcome COVID-19.

As the causative agent for COVID-19, SARS-CoV-2 is a new type of  $\beta$ -coronavirus ( $\beta$ -CoV) with a genome of about 30kb and encodes at least 29 proteins [48–50]. The recent outbreaks of two viral pneumonia induced by  $\beta$ -CoVs infection are SARS-CoV in 2002 [51] and MERS-CoV in 2012 [52], respectively. Angiotensin-converting enzyme 2 (ACE2) is considered to be the common receptor for the cell entry of SARS-CoV and SARS-CoV-2 by binding to their surface spike (S) glycoprotein [53–55] while dipeptidyl peptidase 4 (DPP4) is the receptor for MERS-CoV-2 entry into cells [56–58]. In this case, DPP4 is considered as a potential receptor to binding the Spike protein of SARS-CoV-2 [59, 60]. Of note, a molecular docking study showed that the RBD of spike in SARS-CoV-2 had weakened interactions with DPP4 compared to MERS-CoV [61], indicating DPP4 was not a dominant receptor for SARS-CoV-2. Recently, numerous excellent reviews and comments related to COVID-19 have been published [62–69], including the methods for medical and laboratorial diagnosis [70–75], the transmission and epidemiology [76–79], the potential pathological mechanisms and clinical manifestations [80–86], the potential therapeutic strategies and management [87–93]. Excitingly, Dacheng Wei et al. developed a rapid and ultra-sensitive method for the detection of SARS-CoV-2 [94], which just need 4 min and detected an ultralow concentration (one to ten copies in 100  $\mu$ L biofluids). Notably, some experts thought that omicron variant of SARS-CoV-2 may overturn the COVID-19 pandemic based on its genetic mutation and clinical peculiarities [95–97]. Instead, the other thought that we need to face a grim reality for COVID-19 caused by omicron variant due to its rapid spread and evasion from the immune response [98–103]. Besides, most researches on the potential mechanisms underlying COVID-19 focus on the way for

SARS-CoV-2 entry into host cells [104–107], the non-specific and specific immune responses to SARS-CoV-2 infection [107–112], and the pathological consequences caused by SARS-CoV-2 infection [113–117]. However, there are still some important and fundamental scientific issues to be resolved. For instance, what are the vital pathogenic factors derived from SARS-CoV-2 for causing COVID-19? What is the material foundation of clinical manifestations of COVID-19?

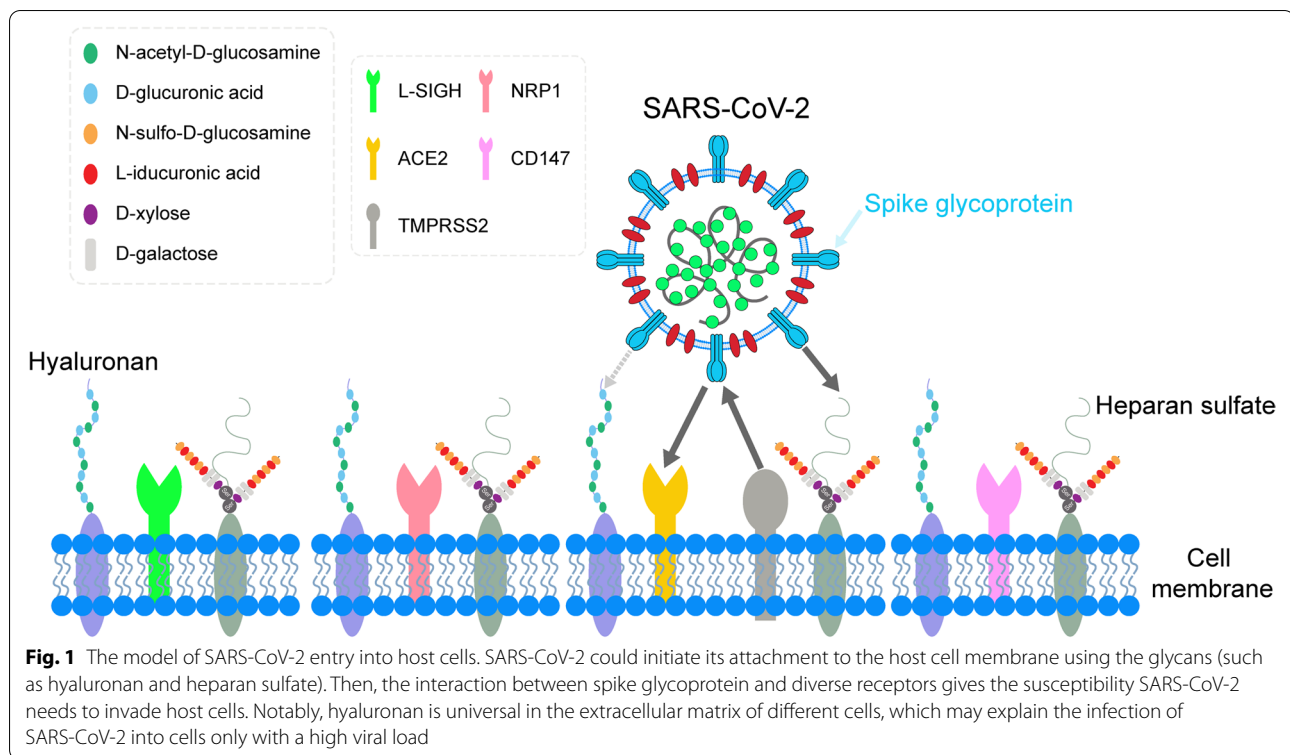
It is well-known that  $\beta$ -CoVs replicate in double-membrane vesicles (DMVs) of cell cytoplasm [118–121]. Paulina Pawlica et al. found that SARS-CoV-2 could generate microRNA-like small RNA to silence host transcripts in cytoplasm and thus contribute to its pathogenesis [122]. However, recent researches indicated that SARS-CoV-2 RNA could be located in the host mitochondria and nucleolus [123, 124]. Moreover, numerous unknown transcripts of SARS-CoV-2 in infected Vero cells have been identified [125, 126]. Significantly, SARS-CoV-2 infection could cause human death even though it is not fatal to its potential hosts, including bats and pangolins [127, 128]. Frankly speaking, little is known regarding the function of SARS-CoV-2 RNA located in nucleus and the key factors involved in determining its pathogenicity in hosts. The recent research revealed that Human Identical Sequences of SARS-CoV-2 (HIS-SARS2) can promote COVID-19 progression by inducing hyaluronan accumulation through activating *HAS2* expression [129], which offers a novel insight into understanding the pathogenic mechanism of SARS-CoV-2.

To date, complete recovery from COVID-19 is still not optimistic despite the tremendous efforts that have been made. Based on previous literature and our understanding of COVID-19, this review discusses pertinent topics of public concern and provides an intensive exposition on SARS-CoV-2, especially with regards to the pathogenic mechanism and potential therapeutic strategies.

## The pathogenesis and intervention therapy of COVID-19 before Three-H strategy

### The infection and transmission of SARS-CoV-2

The basic reproduction number  $R_0$  (also called the basic reproductive ratio), refers to the average number of secondary infected individuals directly linked to the primary infected individual [130, 131]. Generally,  $R_0$  is applied to evaluate the spread ability of communicable diseases, which indicates the intensity of infection and transmission for the infectious source (such as viruses and bacteria). Compared with the  $R_0$  (2 to 5) of severe acute respiratory syndrome coronavirus (SARS) outbreak in 2003, the  $R_0$  of COVID-19 caused by SARS-CoV-2 reached 1.5 to 6.49 [132, 133]. Remarkably, the mean  $R_0$  (5.08) of Delta variant is much higher than the  $R_0$  (2.79)



of its ancestral strain [134], indicating the high communicability of COVID-19. Currently, the Omicron variant has rapidly become a dominating variant of SARS-CoV-2 instead of Delta variant [135]. Which factors then determine the infection and transmission of SARS-CoV-2?

The cell entry of SARS-CoV-2 is reported to be dependent on the ACE2 receptor with the help of TMPRSS2 [136–138]. Especially, the affinity between the S protein of SARS-CoV-2 and ACE2 receptor is 10–20 times that of SARS-CoV-1 [139–141], which gives the susceptibility SARS-CoV-2 needs to invade cells. Surprisingly, the expression of ACE2 is upregulated in the lungs of patients with severe COVID-19 [142–144]. Consistent with this finding, HIS of SARS-CoV-2 can stimulate ACE2 expression [129], implying that SARS-CoV-2 can promote cell entry through upregulating receptor expression by itself. As one of the glycans, heparan sulfate can interact with the RBD of the SARS-CoV-2 S glycoprotein to facilitate the binding of S protein to ACE2 [145]. Consistent with this result, the infection of SARS-CoV-2 pseudovirus is significantly decreased in 293 T-hACE2 cells after treatment with heparin [146]. Noteworthy, SARS-CoV-1 and MERS-CoV can also utilize glycans mediated by their initial attachment to the host cell membranes [147, 148], indicating the importance of glycans in virus infection. Interestingly, endothelial cells can be infected by SARS-CoV-2 only with a high viral load [149]. Moreover, the

diversity of receptors facilitates their facile invasion into cells (Fig. 1), such as ACE2 [136], NRP1 [150], CD147 [151], and L-SIGN [152]. As one of the RNA viruses, the emergence of variants also enhances their high infectivity. For example, the widespread transmission of G614 variants in Europe and America is 3–9 times higher than that of the original D614 strain [153].

#### The re-positive appearance of SARS-CoV-2

The re-positive cases of COVID-19 are usually defined as confirmed cases with SARS-CoV-2 infection after discharge. Different cohort studies have shown that the re-positive ratio of SARS-CoV-2 ranges from 2.4% to 69.2% [154–158]. Notably, most re-positive cases are asymptomatic or mild after discharge [154, 155, 159]. Moreover, the re-positive cases tend to be younger populations, given its median average age at 28 years old [156]. Additionally, the viral load of SARS-CoV-2 was very low in the re-positive cases, as is their transmission risk [158, 159]. From these characteristics of re-positive cases, we speculated that a small amount of SARS-CoV-2 is still residual in the epithelial cells of the nasopharynx even if the clinical symptoms of COVID-19 have receded after treatment. In other words, the patients of re-positive cases had always carried SARS-CoV-2 rather than being truly “re-positive.” This gap in knowledge can be caused by

the limited sensitivity of current detection methods for SARS-CoV-2.

Notably, the re-positive appearance of SARS-CoV-2 may hint at the “friendship” between SARS-CoV-2 and the epithelial cells of the nasopharynx. It is well-known that influenza viruses can disrupt the mucosal barrier resulting in coinfection with common strains of bacteria [160–162], indicating that influenza viruses may cause damage to epithelial cells in the respiratory tract. However, the rate of COVID-19 cases with coinfection of bacteria is considerably low [163–166], further supporting the “friendship” between SARS-CoV-2 and the epithelial cells of the nasopharynx. Accordingly, one of the most common symptoms of COVID-19 is dry cough without sputum [167–169], which has been used to accurately diagnose COVID-19 via artificial intelligence [48].

### The cytokine storm in COVID-19

The hyperactive immune responses in COVID-19 patients promote the release of a large number of pro-inflammatory cytokines and further stimulate excessive inflammatory reaction, also known as the cytokine storm [170], which induces acute respiratory distress syndrome (ARDS). It is reported that high concentrations of cytokines (such as IL-1 $\beta$ , TNF $\alpha$ , IL-6, and IL-8) and chemokines (such as CCL2, CCL-5, IFN $\gamma$ -induced protein 10 (IP-10), and CCL3) are detected in plasma and BALF and are associated with the occurrence and poor clinical outcomes of ARDS, such as mortality rate [171–176]. Apoptosis and other type of cell deaths could be another factors to cause ARDS, which has been reviewed in detail elsewhere [177]. For example, apoptosis mediated by the activation of Fas/Fas ligand pathway contributes to ARDS [178–180]. Currently, many systematic reviews have concentrated on cytokine storm in COVID-19 and discussed its potential mechanisms including signaling pathways [181–187], which mainly emphasized the important roles of immune cells in cytokine storm. However, it isn't still clearly elucidated about the pathogenesis of cytokine storm. In the following section, we will discuss the distinct perspectives on the underlying mechanisms for cytokine storm in COVID-19 based on current clinical observations and experimental findings.

### Spike protein of SARS-CoV-2 may stimulate allergic reaction during COVID-19

The aberrant cutaneous manifestations in COVID-19 has been found in a multitude of retrospective studies [188–190], such as the erythematous rash, urticaria and maculopapular eruptions, which are also the typical symptoms of allergic reactions. It is well-known that abnormal levels of IgE and histamine are universal indicators to assess allergic reactions [191–193]. In a retrospective study on

COVID-19 [194], the level of IgE is significantly increased in non-survivors (71.30 IU/mL), compared to survivors (42.25 IU/mL). Similarly, 119 of 303 (39%) COVID-19 patients with elevated serum IgE [195]. Particularly, the dynamic change of IgE is closely similar to IgM against SARS-CoV-2 [196]. Some excellent reviews have recently highlighted the importance of histamine and its receptors in COVID-19 [197, 198]. Particularly, SARS-CoV-2 could activate mast cells to secrete histamine [199]. At present, there are four known receptors of histamine, designated as H1/H2/H3/H4 receptors (H1R/H2R/H3R/H4R). Although H1 and H2 receptors are relevant to allergic inflammation and gastric acid secretion, respectively [200], both antagonists of H1R and H2R are therapeutic agents for acute allergic reactions in a clinical set [201]. Excitingly, the antagonists of H1/H2 receptor have been proven to improve outcomes of COVID-19 [198, 202]. Above all, allergic responses could appear in some COVID-19 patients, which were potentially elicited by SARS-CoV-2 infection. In this case, what are the potential allergens in COVID-19 after SARS-CoV-2 infection?

Here, the S protein of SARS-CoV-2 may have a potential role as an allergen by analyzing the phenotype and potential molecular mechanisms after vaccination. Since the beginning of COVID-19 outbreak, the spike protein of SARS-CoV-2 has been considered as a foremost target for COVID-19 vaccine development. For example, one of the nCoV-19 vaccines, ChAdOx1, is a full-length virus vector of S-protein [203]. The occurrence of thrombocytopenia and thrombosis in individuals after ChAdOx1 vaccination has attracted public attention [204, 205]. In particular, survivors are discharged from the hospital on day 12 after prednisolone treatment [205], a typical drug used for allergic diseases. Besides, anti-spike binding was detectable in all these individuals while the levels of antibodies against the S protein are varied [205], suggesting the S protein was produced alongside the emergence of the syndrome. Another nCoV-19 vaccine, NVX-CoV2373 is a recombinant nanoparticle vaccine containing the trimeric full-length spike glycoproteins of SARS-CoV-2 (rSARS-CoV-2) and Matrix-M1 adjuvant [206]. During the phase 1–2 trial of NVX-CoV2373, only vaccination of 25  $\mu$ g rSARS-CoV-2 can induce some mild symptoms after the second dose of vaccination including erythema, redness, induration, or swelling [206], which provides direct evidence for the S protein as an allergen. In line with these findings, the safety assessment on the COVID-19 mRNA vaccine showed that the suspected adverse reactions primarily occurred after the second dose of COVID-19 vaccination [207]. Surprisingly, the spike protein of SARS-CoV-2 could directly induce the release of proinflammatory cytokines (such as TNF $\alpha$ ) and apoptosis in THP-1-like macrophages [208], indicating that S



protein may serve as a pathogenic substance. Moreover, the S protein accelerated the expression of pro-thrombotic molecules in pulmonary endothelial cells [209], further hinting the pathogenicity of S protein. Notably, a recent work found that high-sulfated hyaluronan could inactivate SARS-CoV-2 including Alpha and Beta variants by stable binding [210], emphasizing the interaction between hyaluronan and SARS-CoV-2. The structure of monomers is similar between hyaluronan and heparin. Given that heparan sulfate can steadily bind to the RBD of S proteins [145], there could be a similar interaction between hyaluronan and the S protein. Alarmingly, some individuals with hyaluronan dermal fillers had hypersensitivity reactions after SARS-CoV-2 infection or COVID-19 vaccination [211–217], indicating the combination of hyaluronan and S protein may trigger the allergic reactions. Besides, COVID-19 vaccination led a distinct hepatitis mediated by CD8 T cell-dominant immune [218], which may attribute to the long-term expression of S protein from the mRNA vaccination. Interestingly, the binding of hyaluronan to CD44 could regulate the CD8 T cell response [219], further suggesting that the complex of hyaluronan and S protein may induce the hepatitis in individuals after COVID-19 vaccination. Collectively, all these signs suggest that the S protein of SARS-CoV-2 could serve as allergen and cause the allergic response in certain patients, further aggravating their COVID-19 symptoms or inducing adverse reactions toward vaccination.

#### Current therapeutic strategies for COVID-19

At present, symptomatic supportive treatment is still the primary therapeutic strategy for COVID-19 clinically [62]. Many potential therapeutic strategies have been proposed to combat COVID-19 [220–229], such as antiviral treatment, immunological therapy, Chinese medicinal therapy, and anti-inflammatory therapy. Interestingly, electric stimulation may be a subsidiary approach to improve COVID-19 outcomes by increasing the penetration of antiviral drugs [230]. Recently, these therapeutics for COVID-19 have been well-summarized in an updated review [231]. Given the emergency circumstances of COVID-19, some drugs have been urgently approved for COVID-19 treatment (Table 1), which still need further clinical trials to confirm their effectiveness. The development of vaccines against SARS-CoV-2 is considered a key approach to fighting the COVID-19 outbreak. As of April 23, 2022, 153 COVID-19 vaccine candidates are in human clinical trials while 197 COVID-19 vaccine candidates are in preclinical development [232]. Numerous clinical trials of potential therapeutic drugs on COVID-19 are ongoing in the world. Yet unfortunately, there are still few drugs available for COVID-19

treatment in clinical practice other than dexamethasone, which promotes us to further investigate the pathogenic mechanism of COVID-19 and develop new therapeutic strategies.

#### HIS as a nuclear acid factor from SARS-CoV-2 for the progression of COVID-19

It is well known that protein encoded from the virus genome is the main driving factor in virus infection and disease development. Since the virus also contain nuclear acid, it would be interesting to investigate whether nuclear acid, especially in its non-coding regions, can also act as a pathogenic factor in virus infection, especially for SARS-CoV-2.

#### HIS-SARS2 could be a crucial pathogenic factor of SARS-CoV-2

Human Identical Sequences of SARS-CoV-2 (HIS-SARS2) are greater than 20bp and are entirely identical sequences between SARS-CoV-2 and human genomes [129]. They function as miRNA-like RNA. Intriguingly, it has been reported that multiple viruses could produce miRNA-like non-coding RNAs during their infection [265–269]. For instance, the small viral RNAs (svRNAs) encoded by SARS-CoV could repress host mRNA expression by targeting 3'-UTR of specific transcripts [270]. Specially, SARS-CoV-2 can generate viral miRNAs in relation to the cellular metabolism and biosynthesis in host cells [271]. Recently, a noncoding RNA produced from the ORF7a in SARS-CoV-2 genome was demonstrated to decrease the host transcripts (such as BATF2) via target slicing [122]. Inversely, HIS-SARS2 could activate gene expression by targeting enhancer in human [129], which significantly overlap with the aberrant expressed genes found in bronchoalveolar lavage fluid (BALF) of COVID-19 patients [272]. Particularly, HIS-SARS2 could upregulate inflammation-associated genes in transformed human embryonic kidney cell HEK293T, human fetal lung fibroblast cell MRC5, and human umbilical vein endothelial cell (HUVEC), which is consistent with the finding that the infection of SARS-CoV-2 could lead to multiple organ damage (such as lung, kidney, and liver) by stimulating an inflammation response [273–276]. Moreover, HIS-SARS2 destroyed the function of mitochondria by increasing *CYB5A* and *TIMM21*, which may cause the mitochondria dysfunction related to COVID-19 pathogenesis [277–280]. Moreover, the major enzyme for hyaluronan synthesis, *HAS2*, was activated by HIS-SARS2, which promoted hyaluronan accumulation in severe COVID-19 patients [129, 281, 282]. Notably, SARS-CoV-2 RNA was detected in the plasma of COVID-19 patients using droplet-based digital PCR [283], raising a possibility that HIS-SARS2

**Table 1** The urgent approved drugs for COVID-19 treatment

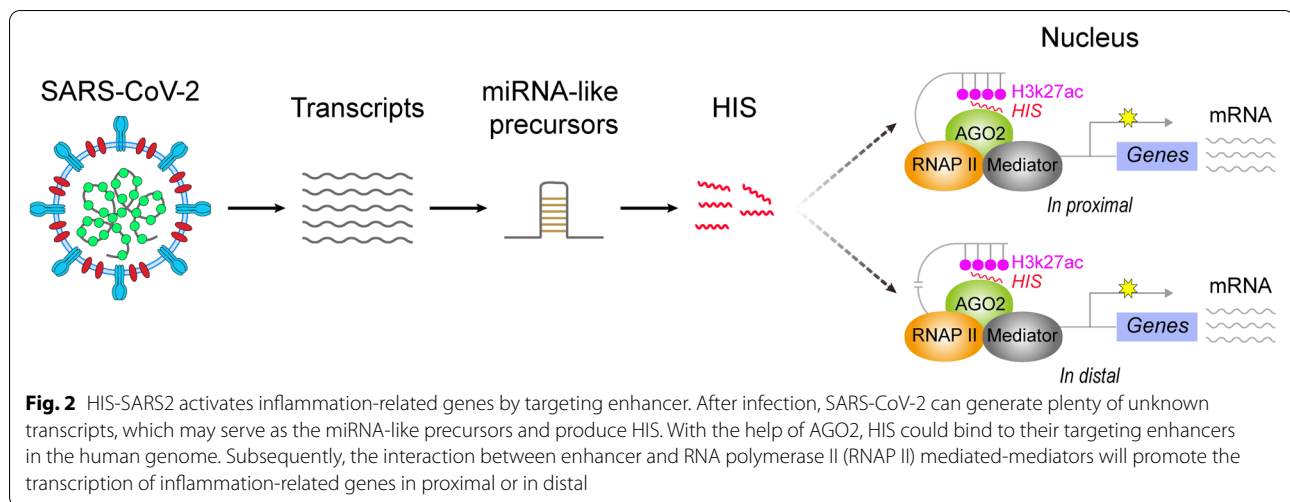
Schemes	Classes	Dosage forms	Clinical trial numbers	References
Remdesivir	Antiviral drug	I.V.	NCT04292899 (Phase 3) NCT04292730 (Phase 3) NCT04401579 (Phase 3) NCT04280705 (Phase 3)	[233–236]
Baricitinid plus Remdesevir	Antiviral drug	I.V./Oral	NCT04970719 (Phase 3) NCT04401579 (Phase 3) NCT04640168 (Phase 3)	[237]
Paxlovid	Antiviral drug	Oral	NCT04960202 (Phase 3) NCT05011513 (Phase 2/3) NCT05047601 (Phase 3)	[237–239]
Molnupiravir	Antiviral drug	Oral	NCT04575584 (Phase 2/3) NCT04575597 (Phase 2/3) NCT04939428 (Phase 3) NCT04405570 (Phase 2) NCT05195060 (Phase 3)	[240–242]
BR11-196/BR11-198	Monoclonal antibodies	I.V.	NCT04787211 (Phase 2) NCT04518410 (Phase 2/3) NCT04501978 (Phase 3)	[237, 243]
Bebtelovimab	Monoclonal antibodies	I.V.	NCT04634409 (Phase 2)	[237, 238]
Bamlanivimad plus Etesevimab	Monoclonal antibodies	I.V.	NCT05205759 (Phase 3) NCT04790786 (Phase 3) NCT04634409 (Phase 2) NCT04427501 (Phase 2)	[237, 238, 244–246]
Casirivimab plus imdevimab	Monoclonal antibodies	I.V.	NCT05205759 (Phase 3) NCT05074433 (Phase 3) NCT04425629 (Phase 3) NCT04790786 (Phase 3) NCT04452318 (Phase 3) NCT04518410 (Phase 2/3)	[244, 247–251]
Sotrovimab	Monoclonal antibodies	I.V.	NCT04913675 (Phase 3) NCT04779879 (Phase 2) NCT04790786 (Phase 3) NCT04381936 (Phase 2/3)	[237, 244]
Convalescent plasma	Plasma	I.V.	NCT04747158 (Phase 2/3) NCT04649879 (Phase 2/3) NCT04433910 (Phase 2) NCT04355767 (Phase 3) NCT04547660 (Phase 3) NCT04345523 (Phase 2) NCT04359810 (Phase 2) NCT04381858 (Phase 3) NCT04747158 (Phase 2/3) NCT04425915 (Phase 3) NCT04362176 (Phase 3) NCT04332835 (Phase 2/3)	[252–261]
Evusheld	Monoclonal antibodies	I.M.	NCT04625725 (phase 3) NCT04625972 (phase 3)	[262]
VV116	Antiviral drug	Oral	NCT05242042 (phase 2/3) NCT05341609 (phase 3)	[263, 264]

could be transported into the distal cells and via hematologic system. Therefore, HIS-SARS2 exert an important role in the pathogenicity of SARS-CoV-2 during infection (Fig. 2).

Strikingly, SARS-CoV-2 infection does not affect the health of bats and pangolins [127, 128]. Some identical sequences were also found between SARS-CoV-2 and its potential hosts' genomes, which were termed as "Host

Identical Sequences (HIS)", while no identical sequences were identified between the genome of SARS-CoV-2 and chicken. This indicates that HIS from SARS-CoV-2 may be helpful when tracing to its mediated hosts.

Most importantly, we put forward the hypothesis of nucleic acid pathogenicity based on the discovery of HIS, which is that the identical sequences between the genome of viruses and hosts (such as human) probably



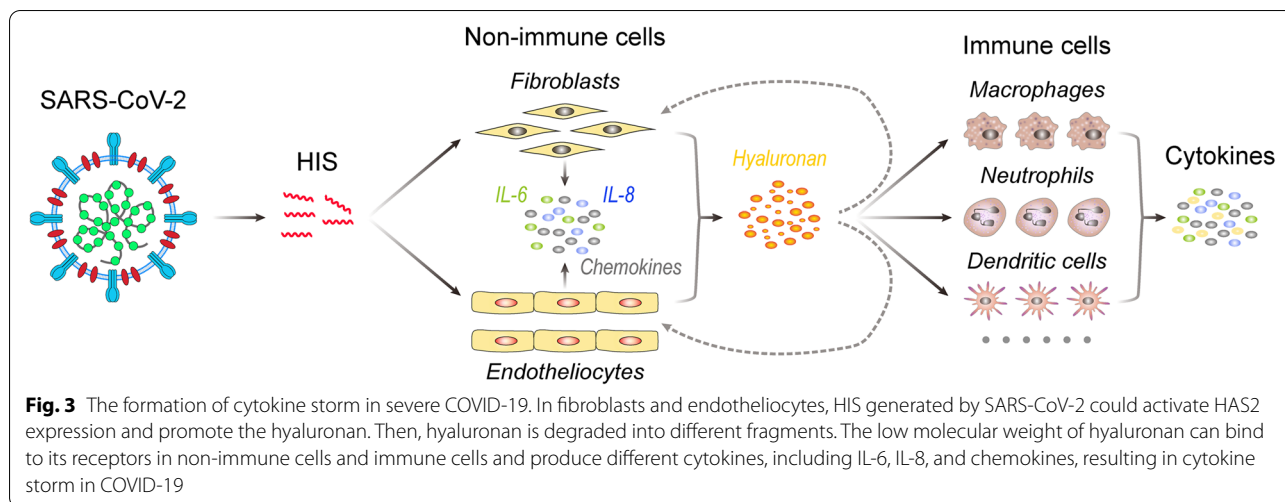
hold the key for viruses to infect hosts and cause diseases. In fact, there are also identical sequences between the genome of human and other pathogenic viruses, including HIV, Ebolavirus, and Zika virus [129], which provide additional support for our hypothesis. The HIS in different viruses deserve further efforts toward clarifying their potential functions, such as acting as important therapeutic targets for these associated diseases.

#### Cytokine storm may be triggered by SARS-CoV-2 rather than the passive response of host

As we all know, the immune responses divide into innate and adaptive responses, which are indisputably activated by SARS-CoV-2 infection in COVID-19. Particularly, the adaptive immune response is believed to be the most potent approach to clearing SARS-CoV-2 [110, 284–286], which is contradictory to a recent report that no IgG antibody against SARS-CoV-2 is detected in 18% of COVID-19 patients even though the average testing time between IgG positive and negative groups is close [287]. Moreover, COVID-19 patients with the second infection have more severe clinical presentation than their first infection [288–290], indicating that adaptive immune response may not exert a dominant role in combating SARS-CoV-2. In fact, many individuals with SARS-CoV-2 infection are capable of clearing the virus under asymptomatic situations [291–293], which implies that innate immunity may hold the key to defeating SARS-CoV-2 as the first defense line against environmental pathogenic substances. In addition, different single-cell omics analyses of COVID-19 patients suggest that the cellular components of innate immune (such as macrophages and monocytes) could determine COVID-19 severity [294, 295]. Thus, innate immunity may play a dominant role in responding to SARS-CoV-2 infection rather than

adaptive immunity during the COVID-19 process, which could be harnessed to mitigate COVID-19.

Currently, it is a common consensus that various cytokines in COVID-19 are generated by diverse immune cells involved in innate immunity (such as neutrophils and macrophages) and adaptive immunity (such as adaptive B and T lymphocytes) [296]. There are many significant increases of cytokines in severe COVID-19 patients [296–298], including TNF- $\alpha$ , IL-6, IL-8, and IL-10. Honestly, we have unwittingly ignored that non-immune cells (such as endothelial cells and fibroblasts) can also produce cytokines [299, 300], which may contribute to the cytokine storm in COVID-19. For example, pulmonary endothelial cells could produce IL-6 by sensing SARS-CoV-2 infection in the adjacent epithelium [149]. Similarly, HIS-SARS2 could activate inflammation-associated genes in human umbilical vein endothelial cells [129]. In line with these findings, circulating endothelial cells were significantly more relevant to IL-6 in severe COVID-19 patients [301]. Notably, fibroblasts could also produce pro-inflammatory cytokines and participate in the persistence of inflammation [302], which may underly the multi-organ fibrosis of COVID-19 patients [303]. Especially, HIS-SARS2 could upregulate HAS2 in human fetal lung fibroblast cells and promote the synthesis of hyaluronan [129], a crucial mediator for inflammation, which may be connected to the pulmonary fibrosis of COVID-19 [304]. Notably, the binding of hyaluronan to CD44 can induce the production of IL-6 and IL-8 in human dermal fibroblasts [305]. In addition, the interaction between hyaluronan and its receptors facilitates cytokine production (such as IL-6 and IL-8) in immune cells [306–308], including macrophages, neutrophils, and dendritic cells. Together, these findings hint that



cytokines released by non-immune cells may trigger cytokine storm in severe COVID-19 (Fig. 3). Given that fibroblasts are widely distributed in various tissues and organs, we believe that fibroblasts may be a major source of cytokine in COVID-19.

**Hyaluronan functions as an essential inducer for the development and severity of COVID-19**  
**Hyaluronan could be a main contributor underlying the clinical manifestations of COVID-19**

COVID-19 has various clinical symptoms. The most common symptoms of COVID-19 are fever, dry cough, and shortness of breath [167]. Based on chest CT, ground-glass opacity (GGO) or consolidations exist in the lungs of COVID-19 patients [309]. Lymphopenia and elevated C-reaction protein (CRP) are two of the most common laboratory abnormalities in hospitalized COVID-19 patients [310]. Meanwhile, severe COVID-19 patients develop ARDS [311]. Notably, COVID-19 can impair the function of multiple organs (such as heart, brain, lung, liver, and kidney) and the coagulation system [62]. Additionally, COVID-19 patients show some neurological complications and symptoms (such as headache, encephalitis, and intracerebral hemorrhage) [312]. However, the foundation underlying the clinical manifestations of COVID-19 is still ambiguous.

Recently, a few studies on metabolic profiles have revealed that the metabolism of carbohydrates, fats, and proteins are dysregulated in COVID-19 patients [313–317], which may provide us some important indication. Specially, gene alterations involved in the metabolism of hyaluronan, glycosaminoglycan, and mucopolysaccharides are excessive in SARS-CoV-2 infected bronchoalveolar cells [317]. In line with this finding, there is a significant increase of hyaluronan in

patients with severe and critical COVID-19 [129, 281], indicating that hyaluronan is related to the COVID-19 clinical process. According to the recent results combined with previous literature, hyaluronan may be crucial to the material foundation of COVID-19 clinical symptoms (Fig. 4).

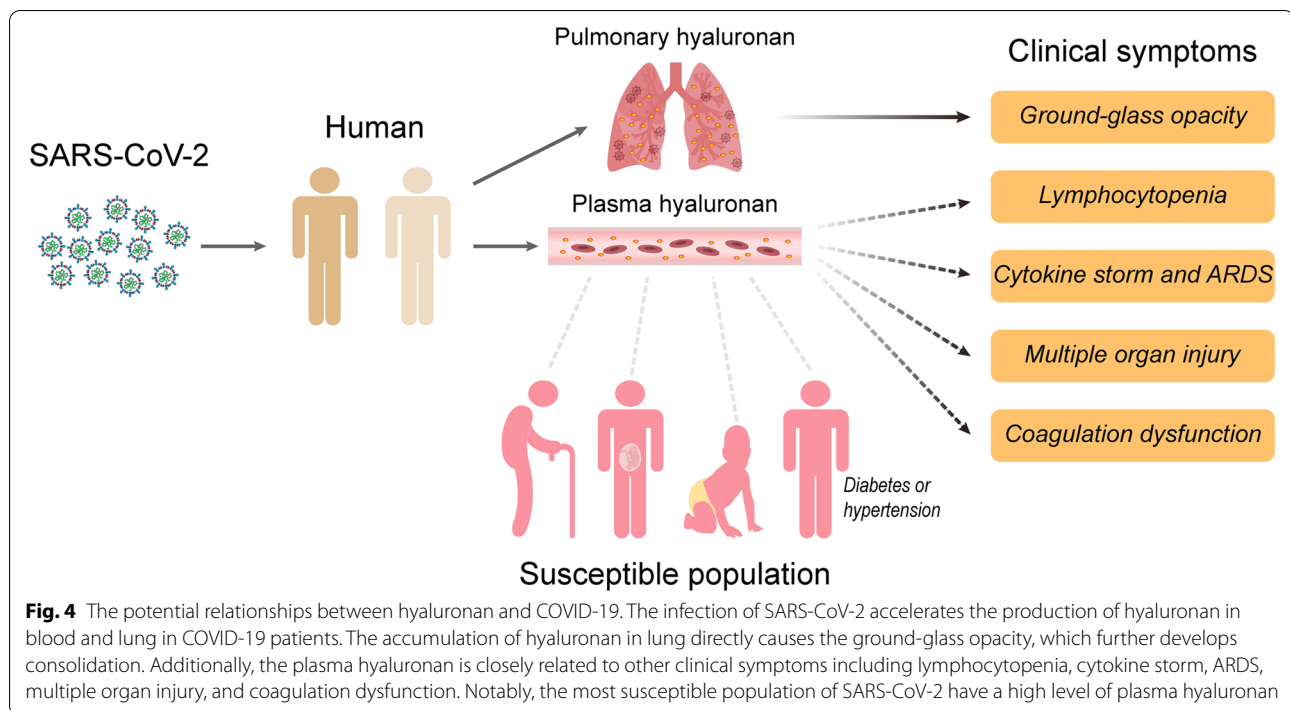
***Hyaluronan may be an important substance for ground-glass opacity in lung***

GGO is a typical presentation of COVID-19 patients based on chest CT, which can further develop into consolidations. Autopsy from three patients with COVID-19 showed that hyaluronan is accumulated in the lung alveoli [318]. Similarly, hyaluronan is also abundant in the respiratory secretions of COVID-19 patients [319]. Strikingly, hyaluronan positively relates to the volume of extravascular water in normal animal lungs [320], which may be due to its ability to absorb a large volume of water [321]. This water absorption characteristic of hyaluronan may be a cause for the jelly-like substance formation present in the lungs of severe COVID-19 patients [322]. Moreover, intratracheal instillation of hyaluronan directly causes pulmonary ground-glass opacity (GGO) or consolidations in mice [323]. Therefore, these evidences suggest that the GGO or consolidations in COVID-19 patients’ lungs contribute to the increase in hyaluronan.

***Hyaluronan may cause the lymphocytopenia in patients with COVID-19***

Lymphocytopenia is a syndrome defined as the loss of lymphocyte in peripheral blood. The reduction of lymphocytes is revealed to be lower in COVID-19 patients with higher levels of hyaluronan [129]. The increased hyaluronan is positively related to the elevated CRP, but





negatively related to decreased lymphocytes in COVID-19 patients, which may be due to the reduction in total T lymphocytes [324]. Intriguingly, the interaction between hyaluronan and its ligand CD44 can cause T cells death when they are activated [325]. In fact, the infection of SARS-CoV-2 can rapidly activate CD4+ T lymphocytes [326]. Thus, the reduction of T lymphocytes mediated by hyaluronan may underlie the foundation of lymphocytopenia in patients with COVID-19.

#### **Hyaluronan may promote acute respiratory distress syndrome in COVID-19**

ARDS is a clinical syndrome characterized by hypoxemia and nonhydrostatic pulmonary edema [327], one of the leading causes of COVID-19 death. Generally, ARDS is thought to be the impairment of pulmonary vascular permeability, which increases lung mass as caused by acute diffuse inflammatory lung injury. Plenty of studies have revealed that hyaluronan can serve as an essential mediator for inflammation and vascular homeostasis [328, 329], which may underlie the inflammatory and vascular permeability impairment in COVID-19 patients with ARDS. These insights are supported by the emergence of hyaluronan in the lung alveoli of patients with ARDS [318]. Especially, HIS-SARS2 can upregulate the expression of a key hyaluronan synthase *HAS2* in MRC5 and HUVEC [129], suggesting that the SARS-CoV-2 infection could directly induce hyaluronan accumulation in lungs.

Consistent with these results, pulmonary microvascular endothelial cells could produce hyaluronan to directly destroy the endothelial barrier when they are exposed to the COVID-19 environment [330]. The binding of hyaluronan may mediate the disruption of endothelial cell barrier to its receptor HABP2 [331]. Hence, beyond triggering the pulmonary inflammatory, dysfunction of the pulmonary endothelial barrier induced by hyaluronan could account for ARDS.

#### **Hyaluronan may trigger multiple organ injury and coagulation system dysfunction**

SARS-CoV-2 has been demonstrated to infect diverse cell types in different tissues [150, 332–334], including lungs, kidneys, brain, and heart. Interestingly, hyaluronan is widely distributed in all parts of the body and exercises a myriad of biological functions in different cell types [335]. Particularly, HIS-SARS2 can induce hyaluronan production via stimulating *HAS2* expression in diverse cells associated with lung, blood vessel, and kidney [17], implying the inflammation triggered by hyaluronan may be the cause of multiple organ injury. In addition, 71% of 183 COVID-19 patients who passed away had diffuse intravascular coagulation [336]. Notably, HABP2, a receptor of hyaluronan is involved in blood coagulation [337], which may also play a key role in the dysfunction of the coagulation system in response to the increased hyaluronan level in COVID-19 patients.

### **Hyaluronan is a pivotal connection between COVID-19 and its risk factors**

The case fatality rate (CFR) of COVID-19 worldwide in 219 countries is approximately 2.1% based on the WHO Coronavirus Dashboard by 22 August 2021. There are many risk factors (such as advanced age, diabetes, hypertension, and cancer) for COVID-19 mortality [338]. Specially, pregnant women and newborns are susceptible to SARS-CoV-2 and can develop severe COVID-19 [339, 340]. Additionally, there are some long-term sequelae of COVID-19 [341, 342], such as severe fatigue, loss of sense of smell or taste, and skin rash. Of note, female sex and older age are related to the risk of persistent symptoms in long COVID [343]. However, the intrinsic relationship between COVID-19 and its risk factors are still unclear.

Remarkably, hyaluronan could help explain the connection between COVID-19 and some of its risk factors. It is reported that serum hyaluronan level of newborns (0–7 day) and elderly people (>60 years) are higher than the other ages in 585 healthy individuals [344]. The higher level of hyaluronan could be associated with the risk of long-term sequelae of COVID-19 in older people. Similarly, the serum hyaluronan level in diabetic patients ( $83.6 \pm 5.6$  ng/mL) was significantly higher than in normal subjects ( $41.7 \pm 12$  ng/mL) [345]. In addition, the elevation of plasma hyaluronan was found in patients with pulmonary hypertension and cancer [346, 347]. Moreover, there is a gradual increase of serum hyaluronan level in women during pregnancy [348], which may partly explain the higher probability of COVID-19 sequelae in female. And lastly, individuals who have received vaccination but died from COVID-19 did so from major cerebral hemorrhage [204]. All these individuals have a high level of platelet factor 4 (PF4). Notably, PF4 can stimulate the release of histamine from basophils and mast cells [349, 350], which could further accelerate the hyaluronan synthesis [351]. Surprisingly, hyaluronan is capable of regulating vascular integrity [352], which may caused the fatal intracranial hemorrhage in vaccinated individuals. Recently, it is reported that the hepatitis of unknown aetiology occurred in five young children infected with SARS-CoV-2 in Scotland [353], which may be attributable to the accumulation of hyaluronan in liver [354]. Thus, we can reasonably infer that a high level of hyaluronan in these individuals may provoke a more violent response after SARS-CoV-2 infection, further exacerbating COVID-19 symptoms (Fig. 4).

### **Hyaluronan could serve as an important indicator for COVID-19**

Serum hyaluronan is usually a non-invasive test to diagnose liver cirrhosis [355], which means that it is exceptionally convenient to detect hyaluronan in clinical.

Combined with the potential roles of hyaluronan in COVID-19, hyaluronan may become a useful clinical indicator for COVID-19. For one, hyaluronan can predict the progression of COVID-19, which is helpful for physicians to determine which patients would require special attention. Secondly, hyaluronan can act as a biomarker to appraise the prognosis of COVID-19. And lastly, hyaluronan can initially screen individuals vaccinated against COVID-19, which may reduce adverse reactions in certain individuals.

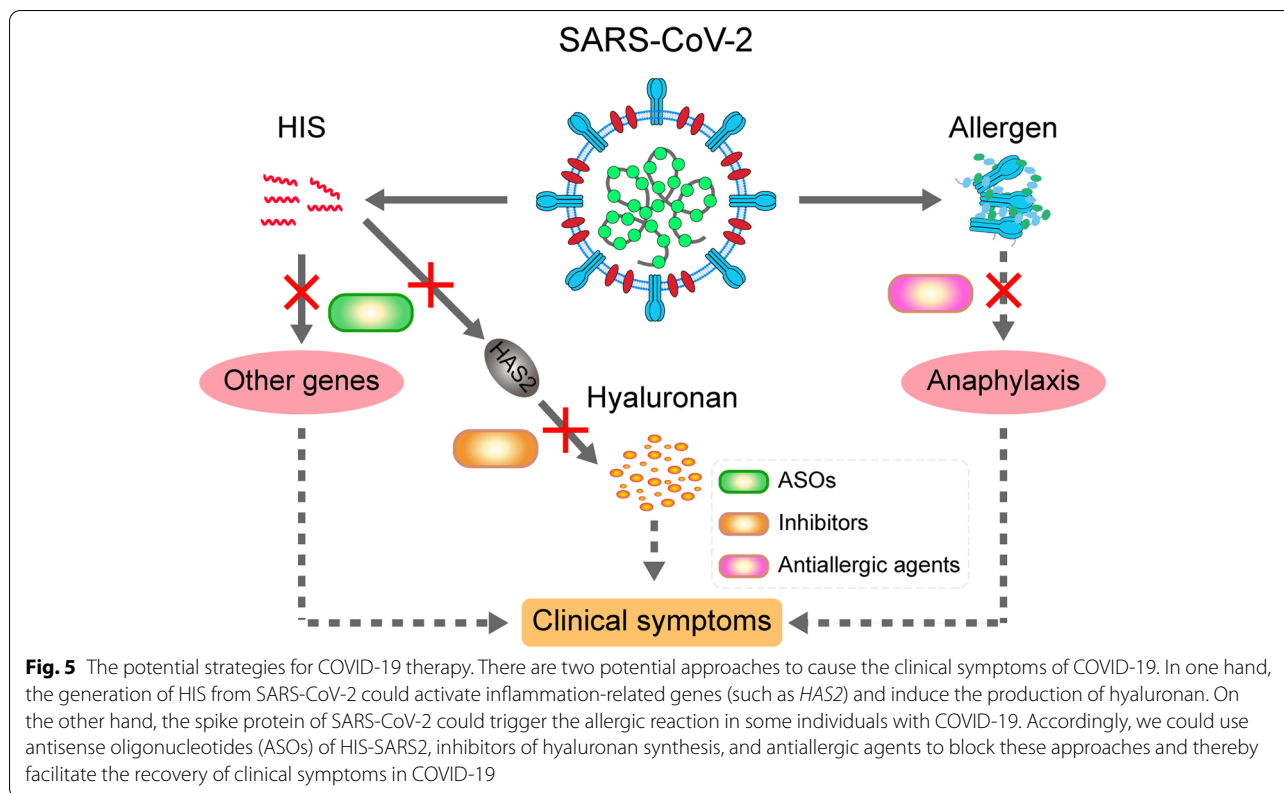
### **Hymecromone as a prospective therapeutic agent for overcome with COVID-19**

Here, we proposed three promising therapeutic agents for COVID-19 treatment on the basis of our recent studies and novel insights into COVID-19, including antisense oligonucleotides (ASOs) of HIS-SARS2, inhibitors of hyaluronan synthesis, and antiallergic agents (Fig. 5).

#### **ASOs of HIS-SARS2**

ASOs are defined as synthesized oligonucleotides measuring 12–30 nucleotides, which are designed to bind to RNA based on base pairing rules [356]. Along with the improvement in technology, ASOs have been used as a therapeutic drug since the late 1980s. To date, several ASO drugs have been approved by the Food and Drug Administration (FDA) to treat different diseases [357]. For example, a 30-mer morpholino ASO, eteplirsen obtained the provisional approval by FDA to treat Duchenne Muscular Dystrophy [358]. Notably, antisense morpholino oligomers targeting the transcription-regulatory sequence (TRS) regions of SARS-CoV could inhibit the production of SARS-CoV [359]. Similarly, the antisense peptide nucleic acid targeting the highly conserved PRF signal of SARS-CoV could significantly suppress its replication [360]. Recently, the importance of ASOs in combating COVID-19 have been recognized due to their high target specificity and rapid development [361]. Especially, the appearance of nanotechnology could facilitate the delivery of ASOs to their target sites [362]. However, it is crucial to identify the potential targets of SARS-CoV-2.

The recent research has clarified the key roles of HIS-SARS2 in response to SARS-CoV-2 infection. In particular HIS-SARS2 upregulated genes associated with inflammation whereas their antagonists abolished their activation [129], suggesting the blocking of HIS-SARS2 could be conducive to alleviating the inflammatory response in COVID-19. Therefore, HIS-SARS2 are candidate targets when designing ASOs, which hold great potential in treating COVID-19.



**Inhibitors of hyaluronan synthesis**

The concentration of hyaluronan is significantly higher in patients with severe COVID-19 [129, 281]. As mentioned above, elevated hyaluronan in COVID-19 could cause most clinical manifestations (such as GGO, lymphopenia, and ARDS) and establish a subtle connection with the risk factors, indicating that hyaluronan could be a key therapeutic target for COVID-19. In fact, this insight is also supported by certain medications for COVID-19 that have already been proven effective in clinical trials. Dexamethasone and Metformin are reported to significantly reduce the CFR of severe COVID-19 patients [363–365], which may be partly attributed to their effects in rapid decrease of hyaluronan [366, 367]. Consequently, inhibitors of hyaluronan synthesis are promising therapeutic agents for COVID-19.

4-methylumbelliferone (4-MU) is a coumarin derivative that can suppress the hyaluronan synthesis by down-regulating the mRNA levels of hyaluronan synthases and depleting their substrate UDP-glucuronic acid [368]. Fortunately, there is an approved prescription drug of 4-MU, called hymecromone, which is used for biliary spasm treatment [369]. In a small sample clinical trial, hyme-cromone has been verified to accelerate the recovery of COVID-19 patients via the promotion of lymphocyte recovery and pulmonary lesion absorption [323]. In other

words, hyme-cromone could be an efficient clinical prescription to block COVID-19 progression.

**Antiallergic agents**

The elevation of serum IgE in COVID-19 patients indicates SARS-CoV-2 infection can stimulate an allergic reaction in some individuals. We proposed above that the S protein of SARS-CoV-2 may serve as an allergen to stimulate an allergic reaction. As such, antiallergic agents could be potential candidates for COVID-19. In general, common clinical antiallergics are antihistamines (such as Diphenhydramine, Promethazine, and Chlorpheniramine) and corticosteroids (such as Dexamethasone) [370, 371]. In addition, the monoclonal antibodies of anti-IgE are alternative agents for anaphylaxis [372]. These antiallergic agents may be new weapons against COVID-19.

**Concluding remarks**

In order to end the COVID-19 pandemic, scientists around the world have conducted a great deal of research on SARS-CoV-2 since its initial outbreak. However, some important issues on the COVID-19 and SARS-CoV-2 are still waiting to be solved. Here, we summarized five concerned problems and discussed the possible answers based on the Three-H strategy.

Why does the pathogenicity of SARS-CoV-2 have species specificity? The interaction between HIS-SARS2 and enhancer activates expression of genes associated with inflammation and further promotes the COVID-19 progression. The sequences of HIS-SARS2 showed the higher conservation in primates (such as Rhesus and Green monkey) [129], which could explain the similar pulmonary damage to COVID-19 in rhesus macaques infected with SARS-CoV-2 [373]. Thus, the interaction between HIS-SARS2 and enhancer determines the pathogenicity of SARS-CoV-2 in specific species. Accordingly, primates could be the best choices to establish animal models of COVID-19 for mechanism research and drug development.

What is the potential mechanism of the clinical manifestations (such as cytokine storm, GGO, thrombosis, and anosmia) in COVID-19? On one hand, HIS-SARS2 activates the expression of HAS2 and induces the accumulation of hyaluronan in the SARS-CoV-2 infected host cells through targeting enhancer. As the key inflammatory mediator, hyaluronan stimulates the non-immune cells (such as fibroblasts) to release cytokines. Meanwhile, the increased hyaluronan in lung leads to the GGO in COVID-19 patients. The binding of hyaluronan to its receptor HBP2 mediates the abnormal thrombosis, which may cause the anosmia in COVID-19 [374]. On the other hand, HIS-SARS2 may promote the production of hyaluronan in distal host cells without SARS-CoV-2 infection via the transportation in exosomes. In this situation, HIS-SARS2 may activate the release of cytokines in fibroblasts through the NamiRNA-Enhancer network.

Why are the specific subpopulations (such as pregnant women, and elderly people, and male sex) susceptible to SARS-CoV-2? As one of the glycans, heparin in the surfaces of the cell facilitate the cell entry of SARS-CoV-2. Likewise, hyaluronan in the cell surfaces may stick SARS-CoV-2 and promote its infection. The pregnant women, and elderly people have higher hyaluronan level, which might enhance the invasion of SARS-CoV-2. Surprisingly, testosterone could elevate the expression of hyaluronan synthase 1 in fibroblasts [375], suggesting that the androgen in the male sex may further increase the hyaluronan production and cause the susceptibility to SARS-CoV-2.

How do we estimate the progression of COVID-19 and decrease the risk of severe COVID-19 in specific subpopulations (such as elderly people and diabetes patients)? Given that hyaluronan is closely associated with the progression of COVID-19 [281, 323], the detection of plasma hyaluronan level may be an important approach to judge the COVID-19 progression. Moreover, hyaluronan level is relatively higher in elderly people and diabetes patients. After SARS-CoV-2 infection, the complexes of hyaluronan and S protein may together aggravate the COVID-19.

Which are the novel strategies to develop the effective drugs for COVID-19 and prevent the sequelae of COVID-19? Focus on HIS and hyaluronan is one of the novel strategies for the drug development of COVID-19. Specifically, we could design the ASO drugs targeting HIS and develop small molecular inhibitors against hyaluronan, which may block the interaction between SARS-CoV-2 and human. Similar, prevention of the hyaluronan elevation by inhibiting androgen pathway is also a strategy for the treatment of COVID-19. When it comes to the intervention therapy of COVID-19, the combination of diverse drugs (such as antiviral drugs and hymecromone) may reach a better therapeutical effect [376]. In addition, reduction of hyaluronan into the normal range might be a potential strategy to prevent the sequelae of COVID-19 though the oral administration of hyaluronan inhibitor drugs (such as hymecromone).

Taken together, Three-H is crucial for us to reconsider the underlying relationship between COVID-19 and SARS-CoV-2. The oligonucleotide drugs, inhibitors of hyaluronan synthesis, or antiallergic agents deserve our time and effort towards further validating their effectiveness in treating COVID-19 in the future.

#### Abbreviations

ACE2: Angiotensin-converting enzyme 2; ARDS: Acute respiratory distress syndrome; ASOs: Antisense oligonucleotides; BALF: Bronchoalveolar lavage fluid; CFR: Case fatality rate; COVID-19: Coronavirus disease 2019; CRP: C-reactive protein; DPP4: Dipeptidyl peptidase 4; DMVs: Double-membrane vesicles; FDA: Food and Drug Administration; GGO: Ground-glass opacity; HIS: Human Identical Sequences; HIS-SARS2: Human Identical Sequences of SARS-CoV-2; HUVEC: Human umbilical vein endothelial cell; IP-10: IFN $\gamma$ -induced protein 10; 4-MU: 4-methylumbelliferone; NamiRNA: Nuclear activating miRNAs; PF4: Platelet factor 4; svRNAs: Small viral RNAs; RBD: Receptor binding domain; R0: R nought; TRS: Transcription-regulatory sequence; Three-H: HIS, hyaluronan, and hymecromone; VOC: Variants of concern.

#### Acknowledgements

We sincerely thank Wei Li, Kaicheng Zhou, Daoping Ru, Peng Xu, Baolong Zhang, Zhicong Yang, Mengxing Liu, and Wenxuan Li from Fudan University for their valuable comments during manuscript preparation. And we appreciate Yue Yu for her kind support in language editing.

#### Code availability

Not applicable.

#### Authors' contributions

W.Y. and S.Y. discussed and proposed the outline of this reviews. S.Y., Y.T., and L.C. finished the manuscript. All the authors participated in manuscript revision and approved the final submission of this manuscript.

#### Funding

This work is supported by the National Key R&D Program of China (2018YFC1005004), Major Special Projects of Basic Research of Shanghai Science and Technology Commission (18JC1411101), and the National Natural Science Foundation of China (31872814).

#### Availability of data and materials

Not applicable.



## Declarations

### Ethics approval and consent to participate

Not applicable.

### Consent for publication

Not applicable.

### Competing interests

All authors have declared that no competing interest exists.

### Author details

<sup>1</sup>Laboratory of RNA Epigenetics, Institutes of Biomedical Sciences & Shanghai Public Health Clinical Center & Department of General Surgery, Huashan Hospital, Cancer Metastasis Institute, Shanghai Medical College, Fudan University, Shanghai 200032, People's Republic of China. <sup>2</sup>Shanghai Key Laboratory of Medical Epigenetics, Shanghai 200032, People's Republic of China.

Received: 9 April 2022 Accepted: 4 May 2022

Published online: 20 May 2022

## References

- Miller IF, Becker AD, Grenfell BT, Metcalf CJE. Disease and healthcare burden of COVID-19 in the United States. *Nat Med.* 2020;26(8):1212–7. <https://doi.org/10.1038/s41591-020-0952-y>.
- Antonio-Villa NE, Bello-Chavolla OY, Vargas-Vazquez A, Fermin-Martinez CA, Marquez-Salinas A, Pisanty-Alatorre J, et al. Assessing the burden of coronavirus disease 2019 (COVID-19) among healthcare workers in Mexico City: a data-driven call to action. *Clin Infect Dis.* 2021;73(1):e191–8. <https://doi.org/10.1093/cid/ciaa1487>.
- Haleem A, Javaid M, Vaishya R. Effects of COVID-19 pandemic in daily life. *Curr Med Res Pract.* 2020;10(2):78–9. <https://doi.org/10.1016/j.cmrp.2020.03.011>.
- Prime H, Wade M, Browne DT. Risk and resilience in family well-being during the COVID-19 pandemic. *Am Psychol.* 2020;75(5):631–43. <https://doi.org/10.1037/amp0000660>.
- Calina D, Docea AO, Petrakis D, Egorov AM, Ishmukhametov AA, Gabibov AG, et al. Towards effective COVID-19 vaccines: updates, perspectives and challenges (review). *Int J Mol Med.* 2020;46(1):3–16. <https://doi.org/10.3892/ijmm.2020.4596>.
- Karpinski TM, Ozarowski M, Seremak-Mrozikiewicz A, Wolski H, Wlodkowic D. The 2020 race towards SARS-CoV-2 specific vaccines. *Theranostics.* 2021;11(4):1690–702. <https://doi.org/10.7150/thno.53691>.
- Zhang J, Zeng H, Gu J, Li H, Zheng L, Zou Q. Progress and prospects on vaccine development against SARS-CoV-2. *Vaccines (Basel).* 2020;8(2):153. <https://doi.org/10.3390/vaccines8020153>.
- Feikin DR, Higdon MM, Abu-Raddad LJ, Andrews N, Araos R, Goldberg Y, et al. Duration of effectiveness of vaccines against SARS-CoV-2 infection and COVID-19 disease: results of a systematic review and meta-regression. *Lancet.* 2022;399(10328):924–44. [https://doi.org/10.1016/S0140-6736\(22\)00152-0](https://doi.org/10.1016/S0140-6736(22)00152-0).
- Tamming LA, Duque D, Tran A, Zhang W, Pfeifle A, Laryea E, et al. DNA based vaccine expressing SARS-CoV-2 spike-CD40L fusion protein confers protection against challenge in a Syrian hamster model. *Front Immunol.* 2022;12:785349. <https://doi.org/10.3389/fimmu.2021.785349>.
- Silveira MM, Moreira G, Mendonca M. DNA vaccines against COVID-19: perspectives and challenges. *Life Sci.* 2021;267:118919. <https://doi.org/10.1016/j.lfs.2020.118919>.
- Jeyanathan M, Afkhami S, Smalf F, Miller MS, Lichty BD, Xing Z. Immunological considerations for COVID-19 vaccine strategies. *Nat Rev Immunol.* 2020;20(10):615–32. <https://doi.org/10.1038/s41577-020-00434-6>.
- Tejaro JR, Farber DL. COVID-19 vaccines: modes of immune activation and future challenges. *Nat Rev Immunol.* 2021;21(4):195–7. <https://doi.org/10.1038/s41577-021-00526-x>.
- Kraynyak KA, Blackwood E, Agnes J, Tebas P, Giffear M, Amante D, et al. SARS-CoV-2 DNA vaccine INO-4800 induces durable immune responses capable of being boosted in a phase 1 open-label trial. *J Infect Dis.* 2022;jiac016. <https://doi.org/10.1093/infdis/jiac016>.
- Thanh Le T, Andreadakis Z, Kumar A, Gomez Roman R, Tollefsen S, Saville M, et al. The COVID-19 vaccine development landscape. *Nat Rev Drug Discov.* 2020;19(5):305–6. <https://doi.org/10.1038/d41573-020-00073-5>.
- Williams SV, Vusirikala A, Ladhani SN, Fernandez Ruiz De Olano E, Iyanger N, Aiano F, et al. An outbreak caused by the SARS-CoV-2 Delta (B.1.617.2) variant in a care home after partial vaccination with a single dose of the COVID-19 vaccine Vaxzevria, London, England, April 2021. *Euro Surveill.* 2021;26(27):2100626. <https://doi.org/10.2807/1560-7917.ES.2021.26.27.2100626>.
- He X, Hong W, Pan X, Lu G, Wei X. SARS-CoV-2 omicron variant: characteristics and prevention. *MedComm.* 2021;2(4):838–45. <https://doi.org/10.1002/mco2.110>.
- Viana R, Moyo S, Amoako DG, Tegally H, Scheepers C, Althaus CL, et al. Rapid epidemic expansion of the SARS-CoV-2 omicron variant in southern Africa. *Nature.* 2022;603(7902):679–86. <https://doi.org/10.1038/s41586-022-04411-y>.
- Araf Y, Akter F, Tang YD, Fatemi R, Parvez MSA, Zheng C, et al. Omicron variant of SARS-CoV-2: genomics, transmissibility, and responses to current COVID-19 vaccines. *J Med Virol.* 2022;94(5):1825–32. <https://doi.org/10.1002/jmv.27588>.
- Karim SSA, Karim QA. Omicron SARS-CoV-2 variant: a new chapter in the COVID-19 pandemic. *Lancet.* 2021;398(10317):2126–8. [https://doi.org/10.1016/S0140-6736\(21\)02758-6](https://doi.org/10.1016/S0140-6736(21)02758-6).
- Wong SC, Au AK, Chen H, Yuen LL, Li X, Lung DC, et al. Transmission of omicron (B.1.1.529) - SARS-CoV-2 variant of concern in a designated quarantine hotel for travelers: a challenge of elimination strategy of COVID-19. *Lancet Reg Health West Pac.* 2022;18:100360. <https://doi.org/10.1016/j.lanwpc.2021.100360>.
- Dejnirattisai W, Huo J, Zhou D, Zahradnik J, Supasa P, Liu C, et al. SARS-CoV-2 omicron-B.1.1.529 leads to widespread escape from neutralizing antibody responses. *Cell.* 2022;185(3):467–84.e15. <https://doi.org/10.1016/j.cell.2021.12.046>.
- Flemming A. Omicron, the great escape artist. *Nat Rev Immunol.* 2022;22(2):75. <https://doi.org/10.1038/s41577-022-00676-6>.
- Cao Y, Wang J, Jian F, Xiao T, Song W, Yisimayi A, et al. Omicron escapes the majority of existing SARS-CoV-2 neutralizing antibodies. *Nature.* 2022;602(7898):657–63. <https://doi.org/10.1038/s41586-021-04385-3>.
- Planas D, Saunders N, Maes P, Guivel-Benhassine F, Planchais C, Buchrieser J, et al. Considerable escape of SARS-CoV-2 omicron to antibody neutralization. *Nature.* 2022;602(7898):671–5. <https://doi.org/10.1038/s41586-021-04389-z>.
- Ndwanjwe D, Wiysonge CS. COVID-19 vaccines. *Curr Opin Immunol.* 2021;71:111–6. <https://doi.org/10.1016/j.coi.2021.07.003>.
- Tregoning JS, Brown ES, Cheeseman HM, Flight KE, Higham SL, Lemm NM, et al. Vaccines for COVID-19. *Clin Exp Immunol.* 2020;202(2):162–92. <https://doi.org/10.1111/cei.13517>.
- Buonsenso D, von Both U. Ensuring global access to COVID-19 vaccines: deployment strategies for refugees and migrants must not be forgotten. *Infection.* 2022;50(1):273–5. <https://doi.org/10.1007/s15010-021-01631-8>.
- Yamey G, Schaferhoff M, Hatchett R, Pate M, Zhao F, McDade KK. Ensuring global access to COVID-19 vaccines. *Lancet.* 2020;395(10234):1405–6. [https://doi.org/10.1016/S0140-6736\(20\)30763-7](https://doi.org/10.1016/S0140-6736(20)30763-7).
- Pomara C, Sessa F, Ciaccio M, Dieli F, Esposito M, Giammanco GM, et al. COVID-19 vaccine and death: causality algorithm according to the WHO eligibility diagnosis. *Diagnostics (Basel).* 2021;11(6):955. <https://doi.org/10.3390/diagnostics11060955>.
- Team CC-VBCL. COVID-19 vaccine breakthrough infections reported to CDC—United States, January 1–April 30, 2021. *MMWR Morb Mortal Wkly Rep.* 2021;70(21):792–3. <https://doi.org/10.15585/mmwr.mm7021e3>.
- Madhi SA, Kwatra G, Myers JE, Jassat W, Dhar N, Mukendi CK, et al. Population immunity and Covid-19 severity with omicron variant in South Africa. *N Engl J Med.* 2022;386:1314–26. <https://doi.org/10.1056/NEJMoa2119658>.
- Rutkowski K, Mirakian R, Till S, Rutkowski R, Wagner A. Adverse reactions to COVID-19 vaccines: a practical approach. *Clin Exp Allergy.* 2021;51(6):770–7. <https://doi.org/10.1111/cea.13880>.

33. Verma AK, Lavine KJ, Lin CY. Myocarditis after Covid-19 mRNA vaccination. *N Engl J Med*. 2021;385(14):1332–4. <https://doi.org/10.1056/NEJM.2109975>.
34. Choi S, Lee S, Seo JW, Kim MJ, Jeon YH, Park JH, et al. Myocarditis-induced sudden death after BNT162b2 mRNA COVID-19 vaccination in Korea: case report focusing on histopathological findings. *J Korean Med Sci*. 2021;36(40):e286. <https://doi.org/10.3346/jkms.2021.36.e286>.
35. Luk A, Clarke B, Dahdah N, Ducharme A, Krahn A, McCrindle B, et al. Myocarditis and pericarditis after Covid-19 mRNA vaccination: practical considerations for care providers. *Can J Cardiol*. 2021;37(10):1629–34. <https://doi.org/10.1016/j.cjca.2021.08.001>.
36. Oster ME, Shay DK, Su JR, Gee J, Creech CB, Broder KR, et al. Myocarditis cases reported after mRNA-based COVID-19 vaccination in the US from December 2020 to August 2021. *JAMA*. 2022;327(4):331–40. <https://doi.org/10.1001/jama.2021.24110>.
37. Ramesh R, Saffar N, Czako B, Agarwal A, Batta K. Cutaneous thrombosis associated with skin necrosis following Oxford-AstraZeneca COVID-19 vaccination. *Clin Exp Dermatol*. 2021;46(8):1610–2. <https://doi.org/10.1111/ced.14819>.
38. Hunter PR. Thrombosis after covid-19 vaccination. *BMJ*. 2021;373:n958. <https://doi.org/10.1136/bmj.n958>.
39. Garcia-Azorin D, Do TP, Gantenbein AR, Hansen JM, Souza MNP, Obermann M, et al. Delayed headache after COVID-19 vaccination: a red flag for vaccine induced cerebral venous thrombosis. *J Headache Pain*. 2021;22(1):108. <https://doi.org/10.1186/s10194-021-01324-5>.
40. Haakonsen HB, Nystedt A. Deep vein thrombosis more than two weeks after vaccination against COVID-19. *Tidsskr Nor Laegeforen*. 2021;141. <https://doi.org/10.4045/tidsskr.21.0274>.
41. Mungmunpantipantip R, Wiwanitit V. Thrombosis after adenovirus-vectored COVID-19 vaccination: a concern on underlying illness. *Clin Appl Thromb Hemost*. 2021;27:10760296211060446. <https://doi.org/10.1177/10760296211060446>.
42. Washington T, Bryan R, Clemow C. Adenopathy following COVID-19 vaccination. *Radiology*. 2021;299(3):E280–1. <https://doi.org/10.1148/radiol.2021210236>.
43. Keshavarz P, Yazdanpanah F, Rafiee F, Mizandari M. Lymphadenopathy following COVID-19 vaccination: imaging findings review. *Acad Radiol*. 2021;28(8):1058–71. <https://doi.org/10.1016/j.acra.2021.04.007>.
44. Garreffa E, Hamad A, O'Sullivan CC, Hazim AZ, York J, Puri S, et al. Regional lymphadenopathy following COVID-19 vaccination: literature review and considerations for patient management in breast cancer care. *Eur J Cancer*. 2021;159:38–51. <https://doi.org/10.1016/j.ejca.2021.09.033>.
45. Gambichler T, Boms S, Susok L, Dickel H, Finis C, Abu Rached N, et al. Cutaneous findings following COVID-19 vaccination: review of world literature and own experience. *J Eur Acad Dermatol Venereol*. 2022;36(2):172–80. <https://doi.org/10.1111/jdv.17744>.
46. Bellinato F, Maurelli M, Gisondi P, Girolomoni G. Cutaneous adverse reactions associated with SARS-CoV-2 vaccines. *J Clin Med*. 2021;10(22):5344. <https://doi.org/10.3390/jcm10225344>.
47. Hussain K, Kawsar A, Weir J, Au L, Turajlic S, Larkin J, et al. Severe cutaneous adverse reaction following COVID-19 vaccination and immunotherapy: a second hit? *Clin Exp Dermatol*. 2022;47(1):149–51. <https://doi.org/10.1111/ced.14852>.
48. Yao H, Song Y, Chen Y, Wu N, Xu J, Sun C, et al. Molecular architecture of the SARS-CoV-2 virus. *Cell*. 2020;183(3):730–8.e13. <https://doi.org/10.1016/j.cell.2020.09.018>.
49. Andersen KG, Rambaut A, Lipkin WI, Holmes EC, Garry RF. The proximal origin of SARS-CoV-2. *Nat Med*. 2020;26(4):450–2. <https://doi.org/10.1038/s41591-020-0820-9>.
50. Fernandes JD, Hinrichs AS, Clawson H, Gonzalez JN, Lee BT, Nasar LR, et al. The UCSC SARS-CoV-2 genome browser. *Nat Genet*. 2020;52(10):991–8. <https://doi.org/10.1038/s41588-020-0700-8>.
51. Graham RL, Donaldson EF, Baric RS. A decade after SARS: strategies for controlling emerging coronaviruses. *Nat Rev Microbiol*. 2013;11(12):836–48. <https://doi.org/10.1038/nrmicro3143>.
52. Zumla A, Hui DS, Perlman S. Middle East respiratory syndrome. *Lancet*. 2015;386(9997):995–1007. [https://doi.org/10.1016/S0140-6736\(15\)60454-8](https://doi.org/10.1016/S0140-6736(15)60454-8).
53. Li H, Liu SM, Yu XH, Tang SL, Tang CK. Coronavirus disease 2019 (COVID-19): current status and future perspectives. *Int J Antimicrob Agents*. 2020;55(5):105951. <https://doi.org/10.1016/j.ijantimicag.2020.105951>.
54. Song W, Gui M, Wang X, Xiang Y. Cryo-EM structure of the SARS coronavirus spike glycoprotein in complex with its host cell receptor ACE2. *PLoS Pathog*. 2018;14(8):e1007236. <https://doi.org/10.1371/journal.ppat.1007236>.
55. Turner AJ, Hiscox JA, Hooper NM. ACE2: from vasoepitidase to SARS virus receptor. *Trends Pharmacol Sci*. 2004;25(6):291–4. <https://doi.org/10.1016/j.tips.2004.04.001>.
56. Yin Y, Wunderink RG. MERS, SARS and other coronaviruses as causes of pneumonia. *Respirology*. 2018;23(2):130–7. <https://doi.org/10.1111/resp.13196>.
57. Wang N, Shi X, Jiang L, Zhang S, Wang D, Tong P, et al. Structure of MERS-CoV spike receptor-binding domain complexed with human receptor DPP4. *Cell Res*. 2013;23(8):986–93. <https://doi.org/10.1038/cr.2013.92>.
58. Letko M, Miazgowiec K, McMinn R, Seifert SN, Sola I, Juanes L, et al. Adaptive evolution of MERS-CoV to species variation in DPP4. *Cell Rep*. 2018;24(7):1730–7. <https://doi.org/10.1016/j.celrep.2018.07.045>.
59. Li Y, Zhang Z, Yang L, Lian X, Xie Y, Li S, et al. The MERS-CoV receptor DPP4 as a candidate binding target of the SARS-CoV-2 spike. *iScience*. 2020;23(6):101160. <https://doi.org/10.1016/j.isci.2020.101160>.
60. Xi CR, Di Fazio A, Nadvi NA, Patel K, Xiang MSW, Zhang HE, et al. A novel purification procedure for active recombinant human DPP4 and the inability of DPP4 to bind SARS-CoV-2. *Molecules*. 2020;25(22):5392. <https://doi.org/10.3390/molecules25225392>.
61. Cameron K, Rozano L, Falasca M, Mancera RL. Does the SARS-CoV-2 spike protein receptor binding domain interact effectively with the DPP4 (CD26) receptor? A molecular docking study. *Int J Mol Sci*. 2021;22(13):7001. <https://doi.org/10.3390/ijms22137001>.
62. Wiersinga WJ, Rhodes A, Cheng AC, Peacock SJ, Prescott HC. Pathophysiology, transmission, diagnosis, and treatment of coronavirus disease 2019 (COVID-19): a review. *JAMA*. 2020;324(8):782–93. <https://doi.org/10.1001/jama.2020.12839>.
63. Anka AU, Tahir MI, Abubakar SD, Alsabbagh M, Zian Z, Hamedifar H, et al. Coronavirus disease 2019 (COVID-19): An overview of the immunopathology, serological diagnosis and management. *Scand J Immunol*. 2021;93(4):e12998. <https://doi.org/10.1111/sji.12998>.
64. Madewell ZJ, Yang Y, Longini IM Jr, Halloran ME, Dean NE. Household secondary attack rates of SARS-CoV-2 by variant and vaccination status: an updated systematic review and meta-analysis. *JAMA Netw Open*. 2022;5(4):e229317. <https://doi.org/10.1001/jamanetworkopen.2022.9317>.
65. Alam T, Qamar SJIUP. Coronavirus disease (COVID-19): reviews, applications, and current status. *Jurnal Informatika Universitas Pamulang*. 2020;5(3). <https://doi.org/10.32493/informatika.v5i3.6563>.
66. Hatmi ZN. A systematic review of systematic reviews on the COVID-19 pandemic. *SN Compr Clin Med*. 2021;3(2):419–36. <https://doi.org/10.1007/s42399-021-00749-y>.
67. To KK, Sridhar S, Chiu KH, Hung DL, Li X, Hung IF, et al. Lessons learned 1 year after SARS-CoV-2 emergence leading to COVID-19 pandemic. *Emerg Microbes Infect*. 2021;10(1):507–35. <https://doi.org/10.1080/22221751.2021.1898291>.
68. Mogensen TH. Human genetics of SARS-CoV-2 infection and critical COVID-19. *Clin Microbiol Infect*. 2022. <https://doi.org/10.1016/j.cmi.2022.02.022>.
69. Cascella M, Rajnik M, Aleem A, Dulebohn SC, Di Napoli R. Features, evaluation, and treatment of coronavirus (COVID-19). *StatPearls Publishing*; 2022. Available from: <https://www.ncbi.nlm.nih.gov/books/NBK54776/?report=classic>.
70. Taleghani N, Taghipour F. Diagnosis of COVID-19 for controlling the pandemic: a review of the state-of-the-art. *Biosens Bioelectron*. 2021;174:112830. <https://doi.org/10.1016/j.bios.2020.112830>.
71. Yuce M, Filiztekin E, Ozkaya KG. COVID-19 diagnosis - a review of current methods. *Biosens Bioelectron*. 2021;172:112752. <https://doi.org/10.1016/j.bios.2020.112752>.
72. Singh B, Datta B, Ashish A, Dutta G. A comprehensive review on current COVID-19 detection methods: from lab care to point of care diagnosis. *Sens Int*. 2021;2:100119. <https://doi.org/10.1016/j.sintl.2021.100119>.

73. Lai CKC, Lam W. Laboratory testing for the diagnosis of COVID-19. *Biochem Biophys Res Commun*. 2021;538:226–30. <https://doi.org/10.1016/j.bbrc.2020.10.069>.
74. Wan DY, Luo XY, Dong W, Zhang ZW. Current practice and potential strategy in diagnosing COVID-19. *Eur Rev Med Pharmacol Sci*. 2020;24(8):4548–53. [https://doi.org/10.26355/eurrev\\_202004\\_21039](https://doi.org/10.26355/eurrev_202004_21039).
75. Carpenter CR, Mudd PA, West CP, Wilber E, Wilber ST. Diagnosing COVID-19 in the emergency department: a scoping review of clinical examinations, laboratory tests, imaging accuracy, and biases. *Acad Emerg Med*. 2020;27(8):653–70. <https://doi.org/10.1111/acem.14048>.
76. Yesudhas D, Srivastava A, Gromiha MM. COVID-19 outbreak: history, mechanism, transmission, structural studies and therapeutics. *Infection*. 2021;49(2):199–213. <https://doi.org/10.1007/s15010-020-01516-2>.
77. Khan M, Adil SF, Alkhatlan HZ, Tahir MN, Saif S, Khan M, et al. COVID-19: a global challenge with old history, epidemiology and progress so far. *Molecules*. 2020;26(1):39. <https://doi.org/10.3390/molecules26010039>.
78. Du Z, Liu C, Wang C, Xu L, Xu M, Wang L, et al. Reproduction numbers of SARS-CoV-2 variants: a systematic review and meta-analysis. *Clin Infect Dis*. 2022;ciac137. <https://doi.org/10.1093/cid/ciac137>.
79. Reich P, Elward A. Infection prevention during the coronavirus disease 2019 pandemic. *Infect Dis Clin N Am*. 2022;36(1):15–37. <https://doi.org/10.1016/j.jidc.2021.12.002>.
80. Gusev E, Sarapultsev A, Solomatina L, Chereshev V. SARS-CoV-2-specific immune response and the pathogenesis of COVID-19. *Int J Mol Sci*. 2022;23(3):1716. <https://doi.org/10.3390/ijms23031716>.
81. Bechmann N, Barthel A, Schedl A, Herzig S, Varga Z, Gebhard C, et al. Sexual dimorphism in COVID-19: potential clinical and public health implications. *Lancet Diabetes Endocrinol*. 2022;10(3):221–30. [https://doi.org/10.1016/S2213-8587\(21\)00346-6](https://doi.org/10.1016/S2213-8587(21)00346-6).
82. Mehandru S, Merad M. Pathological sequelae of long-haul COVID. *Nat Immunol*. 2022;23(2):194–202. <https://doi.org/10.1038/s41590-021-01104-y>.
83. Shirvaliloo M. The unfavorable clinical outcome of COVID-19 in smokers is mediated by H3K4me3, H3K9me3 and H3K27me3 histone marks. *Epi-genomics*. 2022;14(3):153–62. <https://doi.org/10.2217/epi-2021-0476>.
84. Shen Q, Li J, Zhang Z, Guo S, Wang Q, An X, et al. COVID-19: systemic pathology and its implications for therapy. *Int J Biol Sci*. 2022;18(1):386–408. <https://doi.org/10.7150/ijbs.65911>.
85. Khezri MR, Varzandeh R, Ghasemnejad-Berenji M. The probable role and therapeutic potential of the PI3K/AKT signaling pathway in SARS-CoV-2 induced coagulopathy. *Cell Mol Biol Lett*. 2022;27(1):6. <https://doi.org/10.1186/s11658-022-00308-w>.
86. Jiang Y, Rubin L, Peng T, Liu L, Xing X, Lazarovici P, et al. Cytokine storm in COVID-19: from viral infection to immune responses, diagnosis and therapy. *Int J Biol Sci*. 2022;18(2):459–72. <https://doi.org/10.7150/ijbs.59272>.
87. Ambrosino P, Calcaterra IL, Mosella M, Formisano R, D'Anna SE, Bachetti T, et al. Endothelial dysfunction in COVID-19: a unifying mechanism and a potential therapeutic target. *Biomedicines*. 2022;10(4):812. <https://doi.org/10.3390/biomedicines10040812>.
88. Mindt BC, DiGiandomenico A. Microbiome modulation as a novel strategy to treat and prevent respiratory infections. *Antibiotics (Basel)*. 2022;11(4):474. <https://doi.org/10.3390/antibiotics11040474>.
89. Chilamakuri R, Agarwal S. COVID-19: characteristics and therapeutics. *Cells*. 2021;10(2):206. <https://doi.org/10.3390/cells10020206>.
90. Gavriatopoulou M, Ntanasis-Stathopoulos I, Korompoki E, Fotiou D, Migkou M, Tzanninis IG, et al. Emerging treatment strategies for COVID-19 infection. *Clin Exp Med*. 2021;21(2):167–79. <https://doi.org/10.1007/s10238-020-00671-y>.
91. Pascarella G, Strumia A, Piliago C, Bruno F, Del Buono R, Costa F, et al. COVID-19 diagnosis and management: a comprehensive review. *J Intern Med*. 2020;288(2):192–206. <https://doi.org/10.1111/joim.13091>.
92. Stawski R, Nowak D, Perdas E. Cell-free DNA: potential application in COVID-19 diagnostics and management. *Viruses*. 2022;14(2):321. <https://doi.org/10.3390/v14020321>.
93. Chekol Abebe E, Tiruneh GMM, Behaile TMA, Asmamaw Dejenie T, Mengie Ayele T, Tadele Admasu F, et al. Mutational pattern, impacts and potential preventive strategies of omicron SARS-CoV-2 variant infection. *Infect Drug Resist*. 2022;15:1871–87. <https://doi.org/10.2147/IDR.S360103>.
94. Wang L, Wang X, Wu Y, Guo M, Gu C, Dai C, et al. Rapid and ultrasensitive electromechanical detection of ions, biomolecules and SARS-CoV-2 RNA in unamplified samples. *Nat Biomed Eng*. 2022;6(3):276–85. <https://doi.org/10.1038/s41551-021-00833-7>.
95. Tiecco G, Storti S, Degli Antoni M, Focà E, Castelli F, Quiros-Roldan E. Omicron genetic and clinical peculiarities that may overturn SARS-CoV-2 pandemic: a literature review. *Int J Mol Sci*. 2022;23(4):1987. <https://doi.org/10.3390/ijms23041987>.
96. Meo SA, Meo AS, Al-Jassir FF, Klonoff DC. Omicron SARS-CoV-2 new variant: global prevalence and biological and clinical characteristics. *Eur Rev Med Pharmacol Sci*. 2021;25(24):8012–8. [https://doi.org/10.26355/eurrev\\_202112\\_27652](https://doi.org/10.26355/eurrev_202112_27652).
97. Zhao H, Lu L, Peng Z, Chen L-L, Meng X, Zhang C, et al. SARS-CoV-2 omicron variant shows less efficient replication and fusion activity when compared with Delta variant in TMPRSS2-expressed cells. *Emerg Microbes Infect*. 2022;11(1):277–83. <https://doi.org/10.1080/22221751.2021.2023329>.
98. Arora S, Grover V, Saluja P, Algarni YA, Saquib SA, Asif SM, et al. Literature review of omicron: a grim reality amidst COVID-19. *Microorganisms*. 2022;10(2):451. <https://doi.org/10.3390/microorganisms10020451>.
99. Callaway E. Heavily mutated omicron variant puts scientists on alert. *Nature*. 2021;600(7887):21. <https://doi.org/10.1038/d41586-021-03552-w>.
100. Callaway E. Omicron likely to weaken COVID vaccine protection. *Nature*. 2021;600(7889):367–8. <https://doi.org/10.1038/d41586-021-03672-3>.
101. Chen LL, Chua GT, Lu L, Chan BP, Wong JS, Chow CC, et al. Omicron variant susceptibility to neutralizing antibodies induced in children by natural SARS-CoV-2 infection or COVID-19 vaccine. *Emerg Microbes Infect*. 2022;11(1):543–7. <https://doi.org/10.1080/22221751.2022.2035195>.
102. Mostafavi E, Dubey AK, Teodori L, Ramakrishna S, Kaushik A. SARS-CoV-2 omicron variant: a next phase of the COVID-19 pandemic and a call to arms for system sciences and precision medicine. *MedComm*. 2022;3(1):e119. <https://doi.org/10.1002/mco2.119>.
103. Zhang L, Li Q, Liang Z, Li T, Liu S, Cui Q, et al. The significant immune escape of pseudotyped SARS-CoV-2 variant omicron. *Emerg Microbes Infect*. 2022;11(1):1–5. <https://doi.org/10.1080/22221751.2021.2017757>.
104. Behl T, Kaur I, Aleya L, Sehgal A, Singh S, Sharma N, et al. CD147-spike protein interaction in COVID-19: get the ball rolling with a novel receptor and therapeutic target. *Sci Total Environ*. 2022;808:152072. <https://doi.org/10.1016/j.scitotenv.2021.152072>.
105. Gadanec LK, McSweeney KR, Qaradakhli T, Ali B, Zulli A, Apostolopoulos V. Can SARS-CoV-2 virus use multiple receptors to enter host cells? *Int J Mol Sci*. 2021;22(3):992. <https://doi.org/10.3390/ijms22030992>.
106. Latini A, Agolini E, Novelli A, Borgiani P, Giannini R, Gravina P, et al. COVID-19 and genetic variants of protein involved in the SARS-CoV-2 entry into the host cells. *Genes*. 2020;11(9):1010. <https://doi.org/10.3390/genes11091010>.
107. Jackson CB, Farzan M, Chen B, Choe H. Mechanisms of SARS-CoV-2 entry into cells. *Nat Rev Mol Cell Biol*. 2022;23(1):3–20. <https://doi.org/10.1038/s41580-021-00418-x>.
108. Kellam P, Barclay W. The dynamics of humoral immune responses following SARS-CoV-2 infection and the potential for reinfection. *J Gen Virol*. 2020;101(8):791–7. <https://doi.org/10.1099/jgv.0.001439>.
109. Pierce CA, Preston-Hurlburt P, Dai Y, Aschner CB, Cheshenko N, Galen B, et al. Immune responses to SARS-CoV-2 infection in hospitalized pediatric and adult patients. *Sci Transl Med*. 2020;12(564):eabd5487. <https://doi.org/10.1126/scitranslmed.abd5487>.
110. Zhang F, Gan R, Zhen Z, Hu X, Li X, Zhou F, et al. Adaptive immune responses to SARS-CoV-2 infection in severe versus mild individuals. *Signal Transduct Target Ther*. 2020;5(1):156. <https://doi.org/10.1038/s41392-020-00263-y>.
111. Wheatley AK, Juno JA, Wang JJ, Selva KJ, Reynaldi A, Tan H-X, et al. Evolution of immune responses to SARS-CoV-2 in mild-moderate COVID-19. *Nat Commun*. 2021;12(1):1162. <https://doi.org/10.1038/s41467-021-21444-5>.
112. Kudlay D, Kofadi I, Khaïtov M. Peculiarities of the T cell immune response in COVID-19. *Vaccines (Basel)*. 2022;10(2):242. <https://doi.org/10.3390/vaccines10020242>.

113. Song E, Zhang C, Israelow B, Lu-Culligan A, Prado AV, Skriabine S, et al. Neuroinvasion of SARS-CoV-2 in human and mouse brain. *J Exp Med*. 2021;218(3):e20202135. <https://doi.org/10.1084/jem.20202135>.
114. Wang J, Jiang M, Chen X, Montaner LJ. Cytokine storm and leukocyte changes in mild versus severe SARS-CoV-2 infection: review of 3939 COVID-19 patients in China and emerging pathogenesis and therapy concepts. *J Leukoc Biol*. 2020;108(1):17–41. <https://doi.org/10.1002/JLB.3COV0520-272R>.
115. Higgins V, Sohaei D, Diamandis EP, Prassas I. COVID-19: from an acute to chronic disease? Potential long-term health consequences. *Crit Rev Clin Lab Sci*. 2021;58(5):297–310. <https://doi.org/10.1080/10408363.2020.1860895>.
116. Wu CT, Lidsky PV, Xiao YH, Lee IT, Cheng R, Nakayama T, et al. SARS-CoV-2 infects human pancreatic beta cells and elicits beta cell impairment. *Cell Metab*. 2021;33(8):1565–76.e5. <https://doi.org/10.1016/j.cmet.2021.05.013>.
117. Gassen NC, Papiés J, Bajaj T, Emanuel J, Dethloff F, Chua RL, et al. SARS-CoV-2-mediated dysregulation of metabolism and autophagy uncovers host-targeting antivirals. *Nat Commun*. 2021;12(1):3818. <https://doi.org/10.1038/s41467-021-24007-w>.
118. Snijder EJ, Limpens R, de Wilde AH, de Jong AWM, Zevenhoven-Dobbe JC, Maier HJ, et al. A unifying structural and functional model of the coronavirus replication organelle: tracking down RNA synthesis. *PLoS Biol*. 2020;18(6):e3000715. <https://doi.org/10.1371/journal.pbio.3000715>.
119. Alsaadi EAJ, Jones IM. Membrane binding proteins of coronaviruses. *Future Virol*. 2019;14(4):275–86. <https://doi.org/10.2217/fvl-2018-0144>.
120. Mohan J, Wollert T. Membrane remodeling by SARS-CoV-2 - double-enveloped viral replication. *Fac Rev*. 2021;10:17. <https://doi.org/10.12703/r/10-17>.
121. Wolff G, Melia CE, Snijder EJ, Bárcena M. Double-membrane vesicles as platforms for viral replication. *Trends Microbiol*. 2020;28(12):1022–33. <https://doi.org/10.1016/j.tim.2020.05.009>.
122. Pawlica P, Yario TA, White S, Wang J, Moss WN, Hui P, et al. SARS-CoV-2 expresses a microRNA-like small RNA able to selectively repress host genes. *Proc Natl Acad Sci U S A*. 2021;118(52):e2116668118. <https://doi.org/10.1073/pnas.2116668118>.
123. Wu KE, Fazal FM, Parker KR, Zou J, Chang HY. RNA-GPS predicts SARS-CoV-2 RNA residency to host mitochondria and nucleolus. *Cell Syst*. 2020;11(1):102–8.e3. <https://doi.org/10.1016/j.cels.2020.06.008>.
124. Burke JM, St Clair LA, Perera R, Parker R. SARS-CoV-2 infection triggers widespread host mRNA decay leading to an mRNA export block. *RNA*. 2021;27(11):1318–29. <https://doi.org/10.1261/rna.078923.121>.
125. Kim D, Lee JY, Yang JS, Kim JW, Kim VN, Chang H. The architecture of SARS-CoV-2 transcriptome. *Cell*. 2020;181(4):914–21.e10. <https://doi.org/10.1016/j.cell.2020.04.011>.
126. Brant AC, Tian W, Majerciak V, Yang W, Zheng ZM. SARS-CoV-2: from its discovery to genome structure, transcription, and replication. *Cell Biosci*. 2021;11(1):136. <https://doi.org/10.1186/s13578-021-00643-z>.
127. Xiao K, Zhai J, Feng Y, Zhou N, Zhang X, Zou JJ, et al. Isolation of SARS-CoV-2-related coronavirus from Malayan pangolins. *Nature*. 2020;583(7815):286–9. <https://doi.org/10.1038/s41586-020-2313-x>.
128. Zhou P, Yang X-L, Wang X-G, Hu B, Zhang L, Zhang W, et al. A pneumonia outbreak associated with a new coronavirus of probable bat origin. *Nature*. 2020;579(7798):270–3. <https://doi.org/10.1038/s41586-020-2012-7>.
129. Li W, Yang S, Xu P, Zhang D, Tong Y, Chen L, et al. SARS-CoV-2 RNA elements share human sequence identity and upregulate hyaluronan via NamiRNA-enhancer network. *EBioMedicine*. 2022;76:103861. <https://doi.org/10.1016/j.ebiom.2022.103861>.
130. Dietz K. The estimation of the basic reproduction number for infectious diseases. *Stat Methods Med Res*. 1993;2(1):23–41. <https://doi.org/10.1177/096228029300200103>.
131. Delamater PL, Street EJ, Leslie TF, Yang YT, Jacobsen KH. Complexity of the basic reproduction number (R0). *Emerg Infect Dis*. 2019;25(1):1–4. <https://doi.org/10.3201/eid2501.171901>.
132. Liu Y, Gayle AA, Wilder-Smith A, Rocklöv J. The reproductive number of COVID-19 is higher compared to SARS coronavirus. *J Travel Med*. 2020;27(2):taaa021. <https://doi.org/10.1093/jtm/taaa021>.
133. Sanche S, Lin YT, Xu C, Romero-Severson E, Hengartner N, Ke R. High contagiousness and rapid spread of severe acute respiratory syndrome coronavirus 2. *Emerg Infect Dis*. 2020;26(7):1470–7. <https://doi.org/10.3201/eid2607.200282>.
134. Liu Y, Rocklöv J. The reproductive number of the Delta variant of SARS-CoV-2 is far higher compared to the ancestral SARS-CoV-2 virus. *J Travel Med*. 2021:taab124. <https://doi.org/10.1093/jtm/taab124>.
135. Jiahui C, Guo-Wei W. Omicron BA.2 (B.1.1.529.2): high potential to becoming the next dominating variant. *J Phys Chem Lett*. 2022;13:3840–9. <https://doi.org/10.1021/acs.jpcclett.2c00469>.
136. Hoffmann M, Kleine-Weber H, Schroeder S, Kruger N, Herrler T, Erichsen S, et al. SARS-CoV-2 cell entry depends on ACE2 and TMPRSS2 and is blocked by a clinically proven protease inhibitor. *Cell*. 2020;181(2):271–80.e8. <https://doi.org/10.1016/j.cell.2020.02.052>.
137. Ragia G, Manolopoulos VG. Inhibition of SARS-CoV-2 entry through the ACE2/TMPRSS2 pathway: a promising approach for uncovering early COVID-19 drug therapies. *Eur J Clin Pharmacol*. 2020;76(12):1623–30. <https://doi.org/10.1007/s00228-020-02963-4>.
138. Zhou L, Xu Z, Castiglione GM, Soiberman US, Eberhart CG, Duh EJ. ACE2 and TMPRSS2 are expressed on the human ocular surface, suggesting susceptibility to SARS-CoV-2 infection. *Ocul Surf*. 2020;18(4):537–44. <https://doi.org/10.1016/j.jtos.2020.06.007>.
139. Wrapp D, Wang N, Corbett KS, Goldsmith JA, Hsieh CL, Abiona O, et al. Cryo-EM structure of the 2019-nCoV spike in the prefusion conformation. *Science*. 2020;367(6483):1260–3. <https://doi.org/10.1126/science.abb2507>.
140. Ali A, Vijayan R. Dynamics of the ACE2-SARS-CoV-2/SARS-CoV spike protein interface reveal unique mechanisms. *Sci Rep*. 2020;10(1):14214. <https://doi.org/10.1038/s41598-020-71188-3>.
141. Medina-Enriquez MM, Lopez-Leon S, Carlos-Escalante JA, Aponte-Torres Z, Cuapio A, Wegman-Ostrosky T. ACE2: the molecular doorway to SARS-CoV-2. *Cell Biosci*. 2020;10(1):148. <https://doi.org/10.1186/s13578-020-00519-8>.
142. Pinto BGG, Oliveira AER, Singh Y, Jimenez L, Goncalves ANA, Ogava RLT, et al. ACE2 expression is increased in the lungs of patients with comorbidities associated with severe COVID-19. *J Infect Dis*. 2020;222(4):556–63. <https://doi.org/10.1093/infdis/jiaa332>.
143. Reindl-Schwaighofer R, Hodlmoser S, Eskandary F, Poglitsch M, Bonderman D, Strassl R, et al. ACE2 elevation in severe COVID-19. *Am J Respir Crit Care Med*. 2021;203(9):1191–6. <https://doi.org/10.1164/rccm.202101-0142LE>.
144. Xiao L, Sakagami H, Miwa N. ACE2: the key molecule for understanding the pathophysiology of severe and critical conditions of COVID-19: demon or angel? *Viruses*. 2020;12(5):491. <https://doi.org/10.3390/v12050491>.
145. Clausen TM, Sandoval DR, Spliid CB, Pihl J, Perrett HR, Painter CD, et al. SARS-CoV-2 infection depends on cellular heparan sulfate and ACE2. *Cell*. 2020;183(4):1043–57.e15. <https://doi.org/10.1016/j.cell.2020.09.033>.
146. Tu B, Wang H, An X, Qu J, Li Q, Gao Y, et al. Inhaled heparin polysaccharide nanodecoy against SARS-CoV-2 and variants. *Acta Pharm Sin B*. 2022. <https://doi.org/10.1016/j.apsb.2022.01.019>.
147. Lang J, Yang N, Deng J, Liu K, Yang P, Zhang G, et al. Inhibition of SARS pseudovirus cell entry by lactoferrin binding to heparan sulfate proteoglycans. *PLoS One*. 2011;6(8):e23710. <https://doi.org/10.1371/journal.pone.0023710>.
148. Li W, Hulswit RJG, Widjaja I, Raj VS, McBride R, Peng W, et al. Identification of sialic acid-binding function for the Middle East respiratory syndrome coronavirus spike glycoprotein. *Proc Natl Acad Sci U S A*. 2017;114(4):E8508–17. <https://doi.org/10.1073/pnas.1712592114>.
149. Schimmel L, Chew KY, Stocks C, Yordanov T, Essebier P, Kulasinghe A, et al. Endothelial cells elicit a pro-inflammatory response to SARS-CoV-2 without productive viral infection. *bioRxiv*. 2021. <https://doi.org/10.1101/2021.02.14.431177>.
150. Cantuti-Castelvetri L, Ojha R, Pedro LD, Djannatian M, Franz J, Kuivanen S, et al. Neuropilin-1 facilitates SARS-CoV-2 cell entry and infectivity. *Science*. 2020;370(6518):856–60. <https://doi.org/10.1126/science.abd2985>.
151. Zhou YQ, Wang K, Wang XY, Cui HY, Zhao Y, Zhu P, et al. SARS-CoV-2 pseudovirus enters the host cells through spike protein-CD147 in an Arf6-dependent manner. *Emerg Microbes Infect*. 2022;11(1):1135–44. <https://doi.org/10.1080/22221751.2022.2059403>.



152. Kondo Y, Larabee JL, Gao L, Shi H, Shao B, Hoover CM, et al. L-SIGN is a receptor on liver sinusoidal endothelial cells for SARS-CoV-2 virus. *JCI Insight*. 2021;6(14):e148999. <https://doi.org/10.1172/jci.insight.148999>.
153. Korber B, Fischer WM, Gnanakaran S, Yoon H, Theiler J, Abfalterer W, et al. Tracking changes in SARS-CoV-2 spike: evidence that D614G increases infectivity of the COVID-19 virus. *Cell*. 2020;182(4):812–27.e19. <https://doi.org/10.1016/j.cell.2020.06.043>.
154. Du HW, Chen JN, Pan XB, Chen XL, Yixian Z, Fang SF, et al. Prevalence and outcomes of re-positive nucleic acid tests in discharged COVID-19 patients. *Eur J Clin Microbiol Infect Dis*. 2021;40(2):413–7. <https://doi.org/10.1007/s10096-020-04024-1>.
155. Habibzadeh P, Sajadi MM, Emami A, Karimi MH, Yadollahie M, Kucheki M, et al. Rate of re-positive RT-PCR test among patients recovered from COVID-19. *Biochem Med (Zagreb)*. 2020;30(3):030401. <https://doi.org/10.11613/BM.2020.030401>.
156. Lu J, Peng J, Xiong Q, Liu Z, Lin H, Tan X, et al. Clinical, immunological and virological characterization of COVID-19 patients that test re-positive for SARS-CoV-2 by RT-PCR. *EBioMedicine*. 2020;59:102960. <https://doi.org/10.1016/j.ebiom.2020.102960>.
157. Kang Y-J. South Korea's COVID-19 infection status: from the perspective of re-positive test results after viral clearance evidenced by negative test results. *Diaster Med Public Health Prep*. 2020;14(6):762–4. <https://doi.org/10.1017/dmp.2020.168>.
158. Liang L, Guo Q, Zhang H, Lin S, Zheng H, Li B, et al. Low infectious risk of re-positive COVID-19 patients: a single-center study. *Int J Infect Dis*. 2021;111:5–9. <https://doi.org/10.1016/j.ijid.2021.08.019>.
159. Tang X, Musa SS, Zhao S, He D. Reinfection or reactivation of severe acute respiratory syndrome coronavirus 2: a systematic review. *Front Public Health*. 2021;9:663045. <https://doi.org/10.3389/fpubh.2021.663045>.
160. Cauley LS, Vella AT. Why is coinfection with influenza virus and bacteria so difficult to control? *Discov Med*. 2015;19(102):33–40.
161. McCullers JA. The co-pathogenesis of influenza viruses with bacteria in the lung. *Nat Rev Microbiol*. 2014;12(4):252–62. <https://doi.org/10.1038/nrmicro3231>.
162. Klein EY, Monteforte B, Gupta A, Jiang W, May L, Hsieh YH, et al. The frequency of influenza and bacterial coinfection: a systematic review and meta-analysis. *Influenza Other Respir Viruses*. 2016;10(5):394–403. <https://doi.org/10.1111/irv.12398>.
163. Adler H, Ball R, Fisher M, Mortimer K, Vardhan MS. Low rate of bacterial co-infection in patients with COVID-19. *Lancet Microbe*. 2020;1(2):e62. [https://doi.org/10.1016/S2666-5247\(20\)30036-7](https://doi.org/10.1016/S2666-5247(20)30036-7).
164. Gerver SM, Guy R, Wilson K, Thelwall S, Nsonwu O, Rooney G, et al. National surveillance of bacterial and fungal coinfection and secondary infection in COVID-19 patients in England: lessons from the first wave. *Clin Microbiol Infect*. 2021;27(11):1658–65. <https://doi.org/10.1016/j.cmi.2021.05.040>.
165. Lardaro T, Wang AZ, Bucca A, Croft A, Globler N, Holt DB, et al. Characteristics of COVID-19 patients with bacterial coinfection admitted to the hospital from the emergency department in a large regional healthcare system. *J Med Virol*. 2021;93(5):2883–9. <https://doi.org/10.1002/jmv.26795>.
166. Hughes S, Troise O, Donaldson H, Mughal N, Moore LSP. Bacterial and fungal coinfection among hospitalized patients with COVID-19: a retrospective cohort study in a UK secondary-care setting. *Clin Microbiol Infect*. 2020;26(10):1395–9. <https://doi.org/10.1016/j.cmi.2020.06.025>.
167. Mao R, Qiu Y, He JS, Tan JY, Li XH, Liang J, et al. Manifestations and prognosis of gastrointestinal and liver involvement in patients with COVID-19: a systematic review and meta-analysis. *Lancet Gastroenterol Hepatol*. 2020;5(7):667–78. [https://doi.org/10.1016/S2468-1253\(20\)30126-6](https://doi.org/10.1016/S2468-1253(20)30126-6).
168. Clemency BM, Varughese R, Scheafer DK, Ludwig B, Welch JV, McCormack RF, et al. Symptom criteria for COVID-19 testing of health care workers. *Acad Emerg Med*. 2020;27(6):469–74. <https://doi.org/10.1111/acem.14009>.
169. Calica Utku A, Budak G, Karabay O, Guclu E, Okan HD, Vatan A. Main symptoms in patients presenting in the COVID-19 period. *Scott Med J*. 2020;65(4):127–32. <https://doi.org/10.1177/0036933020949253>.
170. Ragab D, Salah Eldin H, Taeimah M, Khattab R, Salem R. The COVID-19 cytokine storm; what we know so far. *Front Immunol*. 2020;11:1446. <https://doi.org/10.3389/fimmu.2020.01446>.
171. McClintock D, Zhuo H, Wickersham N, Matthay MA, Ware LB. Biomarkers of inflammation, coagulation and fibrinolysis predict mortality in acute lung injury. *Crit Care*. 2008;12(2):R41. <https://doi.org/10.1186/cc6846>.
172. Parsons PE, Matthay MA, Ware LB, Eisner MD, National Heart LBIARD-SCTN. Elevated plasma levels of soluble TNF receptors are associated with morbidity and mortality in patients with acute lung injury. *Am J Physiol Lung Cell Mol Physiol*. 2005;288(3):L426–31. <https://doi.org/10.1152/ajplung.00302.2004>.
173. Pugin J, Ricou B, Steinberg KP, Suter PM, Martin TR. Proinflammatory activity in bronchoalveolar lavage fluids from patients with ARDS, a prominent role for interleukin-1. *Am J Respir Crit Care Med*. 1996;153(6 Pt 1):1850–6. <https://doi.org/10.1164/ajrccm.153.6.8665045>.
174. Cameron MJ, Bermejo-Martin JF, Danesh A, Muller MP, Kelvin DJ. Human immunopathogenesis of severe acute respiratory syndrome (SARS). *Virus Res*. 2008;133(1):13–9. <https://doi.org/10.1016/j.virusres.2007.02.014>.
175. Jiang Y, Xu J, Zhou C, Wu Z, Zhong S, Liu J, et al. Characterization of cytokine/chemokine profiles of severe acute respiratory syndrome. *Am J Respir Crit Care Med*. 2005;171(8):850–7. <https://doi.org/10.1164/rccm.200407-857OC>.
176. Reghunathan R, Jayapal M, Hsu LY, Chng HH, Tai D, Leung BP, et al. Expression profile of immune response genes in patients with severe acute respiratory syndrome. *BMC Immunol*. 2005;6:2. <https://doi.org/10.1186/1471-2172-6-2>.
177. Tang PS, Mura M, Seth R, Liu M. Acute lung injury and cell death: how many ways can cells die? *Am J Physiol Lung Cell Mol Physiol*. 2008;294(4):L632–41. <https://doi.org/10.1152/ajplung.00262.2007>.
178. Albertine KH, Soulier MF, Wang Z, Ishizaka A, Hashimoto S, Zimmerman GA, et al. Fas and fas ligand are up-regulated in pulmonary edema fluid and lung tissue of patients with acute lung injury and the acute respiratory distress syndrome. *Am J Pathol*. 2002;161(5):1783–96. [https://doi.org/10.1016/S0002-9440\(10\)64455-0](https://doi.org/10.1016/S0002-9440(10)64455-0).
179. Hashimoto S, Kobayashi A, Kooguchi K, Kitamura Y, Onodera H, Nakajima H. Upregulation of two death pathways of perforin/granzyme and FasL/Fas in septic acute respiratory distress syndrome. *Am J Respir Crit Care Med*. 2000;161(1):237–43. <https://doi.org/10.1164/ajrccm.161.1.9810007>.
180. Herrero R, Tanino M, Smith LS, Kajikawa O, Wong VA, Mongovin S, et al. The Fas/FasL pathway impairs the alveolar fluid clearance in mouse lungs. *Am J Physiol Lung Cell Mol Physiol*. 2013;305(5):L377–88. <https://doi.org/10.1152/ajplung.00271.2012>.
181. Hu B, Huang S, Yin L. The cytokine storm and COVID-19. *J Med Virol*. 2021;93(1):250–6. <https://doi.org/10.1002/jmv.26232>.
182. Kim JS, Lee JY, Yang JW, Lee KH, Effenberger M, Szpirt W, et al. Immunopathogenesis and treatment of cytokine storm in COVID-19. *Theranostics*. 2021;11(1):316–29. <https://doi.org/10.7150/thno.49713>.
183. Ye Q, Wang B, Mao J. The pathogenesis and treatment of the 'cytokine storm' in COVID-19. *J Inf Secur*. 2020;80(6):607–13. <https://doi.org/10.1016/j.jinf.2020.03.037>.
184. Soy M, Keser G, Atagunduz P, Tabak F, Atagunduz I, Kayhan S. Cytokine storm in COVID-19: pathogenesis and overview of anti-inflammatory agents used in treatment. *Clin Rheumatol*. 2020;39(7):2085–94. <https://doi.org/10.1007/s10067-020-05190-5>.
185. Hsu RJ, Yu WC, Peng GR, Ye CH, Hu S, Chong PCT, et al. The role of cytokines and chemokines in severe acute respiratory syndrome coronavirus 2 infections. *Front Immunol*. 2022;13:832394. <https://doi.org/10.3389/fimmu.2022.832394>.
186. Luo XH, Zhu Y, Mao J, Du RC. T cell immunobiology and cytokine storm of COVID-19. *Scand J Immunol*. 2021;93(3):e12989. <https://doi.org/10.1111/sji.12989>.
187. Nile SH, Nile A, Qiu J, Li L, Jia X, Kai G. COVID-19: pathogenesis, cytokine storm and therapeutic potential of interferons. *Cytokine Growth Factor Rev*. 2020;53:66–70. <https://doi.org/10.1016/j.cytogfr.2020.05.002>.
188. Recalcati S. Cutaneous manifestations in COVID-19: a first perspective. *J Eur Acad Dermatol Venereol*. 2020;34(5):e212–3. <https://doi.org/10.1111/jdv.16387>.
189. Galvan Casas C, Catala A, Carretero Hernandez G, Rodriguez-Jimenez P, Fernandez-Nieto D, Rodriguez-Villa Lario A, et al. Classification of the cutaneous manifestations of COVID-19: a rapid prospective nationwide consensus study in Spain with 375 cases. *Br J Dermatol*. 2020;183(1):71–7. <https://doi.org/10.1111/bjd.19163>.

190. De Giorgi V, Recalcati S, Jia Z, Chong W, Ding R, Deng Y, et al. Cutaneous manifestations related to coronavirus disease 2019 (COVID-19): a prospective study from China and Italy. *J Am Acad Dermatol*. 2020;83(2):674–5. <https://doi.org/10.1016/j.jaad.2020.05.073>.
191. Gould HJ, Sutton BJ, Beavil AJ, Beavil RL, McCloskey N, Coker HA, et al. The biology of IGE and the basis of allergic disease. *Annu Rev Immunol*. 2003;21:579–628. <https://doi.org/10.1146/annurev.immunol.21.120601.141103>.
192. Lieberman P. The basics of histamine biology. *Ann Allergy Asthma Immunol*. 2011;106(2 Suppl):S2–5. <https://doi.org/10.1016/j.anai.2010.08.005>.
193. Du H, Dong X, Zhang JJ, Cao YY, Akdis M, Huang PQ, et al. Clinical characteristics of 182 pediatric COVID-19 patients with different severities and allergic status. *Allergy*. 2021;76(2):510–32. <https://doi.org/10.1111/all.14452>.
194. Zhao Y, Nie HX, Hu K, Wu XJ, Zhang YT, Wang MM, et al. Abnormal immunity of non-survivors with COVID-19: predictors for mortality. *Infect Dis Poverty*. 2020;9(1):108. <https://doi.org/10.1186/s40249-020-00723-1>.
195. Liu X, Lv J, Gan L, Zhang Y, Sun F, Meng B, et al. Comparative analysis of clinical characteristics, imaging and laboratory findings of different age groups with COVID-19. *Indian J Med Microbiol*. 2020;38(1):87–93. [https://doi.org/10.4103/ijmm.IJMM\\_20\\_133](https://doi.org/10.4103/ijmm.IJMM_20_133).
196. Song Y, Zhong H, Li L, Yin M, Yin Y, Guo X, et al. Dynamic monitoring of immune function indexes in COVID-19 patients. *Aging (Albany NY)*. 2020;12(24):24596–603. <https://doi.org/10.18632/aging.202362>.
197. Ennis M, Tiligada K. Histamine receptors and COVID-19. *Inflamm Res*. 2021;70(1):67–75. <https://doi.org/10.1007/s00011-020-01422-1>.
198. Qu C, Fuhler GM, Pan Y. Could histamine H1 receptor antagonists be used for treating COVID-19? *Int J Mol Sci*. 2021;22(11):5672. <https://doi.org/10.3390/ijms22115672>.
199. Conti P, Caraffa A, Tete G, Gallenga CE, Ross R, Kritas SK, et al. Mast cells activated by SARS-CoV-2 release histamine which increases IL-1 levels causing cytokine storm and inflammatory reaction in COVID-19. *J Biol Regul Homeost Agents*. 2020;34(5):1629–32. <https://doi.org/10.23812/20-2EDIT>.
200. Tiligada E, Ennis M. Histamine pharmacology: from sir Henry dale to the 21st century. *Br J Pharmacol*. 2020;177(3):469–89. <https://doi.org/10.1111/bph.14524>.
201. Wechsler JB, Schroeder HA, Byrne AJ, Chien KB, Bryce PJ. Anaphylactic responses to histamine in mice utilize both histamine receptors 1 and 2. *Allergy*. 2013;68(10):1338–40. <https://doi.org/10.1111/all.12227>.
202. Hogan Ii RB, Hogan Iii RB, Cannon T, Rappai M, Studdard J, Paul D, et al. Dual-histamine receptor blockade with cetirizine - famotidine reduces pulmonary symptoms in COVID-19 patients. *Pulm Pharmacol Ther*. 2020;63:101942. <https://doi.org/10.1016/j.pupt.2020.101942>.
203. Abdalla M, El-Arabey AA, Jiang X. Progress in research on the S protein as the target of COVID-19 vaccines. *Expert Rev Vaccines*. 2021;1-4. <https://doi.org/10.1080/14760584.2021.1918003>.
204. Greinacher A, Thiele T, Warkentin TE, Weisser K, Kyrle PA, Eichinger S. Thrombotic thrombocytopenia after ChAdOx1 nCov-19 vaccination. *N Engl J Med*. 2021;384(22):2092–101. <https://doi.org/10.1056/NEJMoA2104840>.
205. Schultz NH, Sorvoll IH, Michelsen AE, Munthe LA, Lund-Johansen F, Ahlen MT, et al. Thrombosis and thrombocytopenia after ChAdOx1 nCov-19 vaccination. *N Engl J Med*. 2021;384(22):2124–30. <https://doi.org/10.1056/NEJMoa2104882>.
206. Keech C, Albert G, Cho I, Robertson A, Reed P, Neal S, et al. Phase 1-2 trial of a SARS-CoV-2 recombinant spike protein nanoparticle vaccine. *N Engl J Med*. 2020;383(24):2320–32. <https://doi.org/10.1056/NEJMoA2026920>.
207. Hernández AF, Calina D, Poulas K, Docea AO, Tsatsakis AM. Safety of COVID-19 vaccines administered in the EU: should we be concerned? *Toxicol Rep*. 2021;8:871–9. <https://doi.org/10.1016/j.toxrep.2021.04.003>.
208. Barhoumi T, Alghanem B, Shaibah H, Mansour FA, Alamri HS, Akiel MA, et al. SARS-CoV-2 coronavirus spike protein-induced apoptosis, inflammatory, and oxidative stress responses in THP-1-like-macrophages: potential role of angiotensin-converting enzyme inhibitor (perindopril). *Front Immunol*. 2021;12:728896. <https://doi.org/10.3389/fimmu.2021.728896>.
209. Perico L, Morigi M, Galbusera M, Pezzotta A, Gastoldi S, Imberti B, et al. SARS-CoV-2 spike protein 1 activates microvascular endothelial cells and complement system leading to platelet aggregation. *Front Immunol*. 2022;13:827146. <https://doi.org/10.3389/fimmu.2022.827146>.
210. Moller S, Theiss J, Deinert TIL, Golat K, Heinze J, Niemyer D, et al. High-sulfated glycosaminoglycans prevent Coronavirus replication. *Viruses*. 2022;14(2):413. <https://doi.org/10.3390/v14020413>.
211. Rowland-Warmann MJ. Hypersensitivity reaction to hyaluronic acid dermal filler following novel coronavirus infection - a case report. *J Cosmet Dermatol*. 2021;20(5):1557–62. <https://doi.org/10.1111/jocd.14074>.
212. Ortigosa LCM, Lenzone FC, Suarez MV, Duarte AA, Prestes-Carneiro LE. Hypersensitivity reaction to hyaluronic acid dermal filler after COVID-19 vaccination: a series of cases in Sao Paulo, Brazil. *Int J Infect Dis*. 2022;116:268–70. <https://doi.org/10.1016/j.ijid.2022.01.024>.
213. Savva D, Battineni G, Amenta F, Nittari G. Hypersensitivity reaction to hyaluronic acid dermal filler after the Pfizer vaccination against SARS-CoV-2. *Int J Infect Dis*. 2021;113:233–5. <https://doi.org/10.1016/j.ijid.2021.09.066>.
214. Kato K, Inoue E, Tanaka S, Kawamoto H. Increase in the incidence of acute inflammatory reactions to injectable fillers during COVID-19 era. *J Cosmet Dermatol*. 2022. <https://doi.org/10.1111/jocd.14886>.
215. Calvisi L. Hyaluronic acid delayed inflammatory reaction after third dose of SARS-CoV-2 vaccine. *J Cosmet Dermatol*. 2022. <https://doi.org/10.1111/jocd.14970>.
216. Hamed Azzam S, Hamed M, Mukari A. COVID-19 vaccine in patients with dermal hyaluronic acid fillers in the tear trough: a retrospective study. *J Cosmet Dermatol*. 2022. <https://doi.org/10.1111/jocd.14875>.
217. Beamish IV, Bogoch II, Carr D. Delayed inflammatory reaction to dermal fillers after COVID-19 vaccination: a case report. *CJEM*. 2022. <https://doi.org/10.1007/s43678-022-00289-x>.
218. Boettler T, Csernalabics B, Salié H, Luxenburger H, Wischer L, Alizei ES, et al. SARS-CoV-2 vaccination can elicit a CD8 T-cell dominant hepatitis. *J Hepatol*. 2022;S0168-8278(22):00234–3. <https://doi.org/10.1016/j.jhep.2022.03.040>.
219. Lesley J, Howes N, Perschl A, Hyman R. Hyaluronan binding function of CD44 is transiently activated on T cells during an in vivo immune response. *J Exp Med*. 1994;180(1):383–7. <https://doi.org/10.1084/jem.180.1.383>.
220. Zhu S, Guo X, Geary K, Zhang D. Emerging therapeutic strategies for COVID-19 patients. *Discoveries (Craiova)*. 2020;8(1):e105. <https://doi.org/10.15190/d.2020.2>.
221. Salián VS, Wright JA, Vedell PT, Nair S, Li C, Kandimalla M, et al. COVID-19 transmission, current treatment, and future therapeutic strategies. *Mol Pharm*. 2021;18(3):754–71. <https://doi.org/10.1021/acs.molpharmaceut.0c00608>.
222. Narozna M, Rubis B. Anti-SARS-CoV-2 strategies and the potential role of miRNA in the assessment of COVID-19 morbidity, recurrence, and therapy. *Int J Mol Sci*. 2021;22(16):8663. <https://doi.org/10.3390/ijms22168663>.
223. Parray HA, Shukla S, Perween R, Khatri R, Shrivastava T, Singh V, et al. Inhalation monoclonal antibody therapy: a new way to treat and manage respiratory infections. *Appl Microbiol Biotechnol*. 2021;105(16–17):6315–32. <https://doi.org/10.1007/s00253-021-11488-4>.
224. Xu J, Zhang Y. Traditional Chinese medicine treatment of COVID-19. *Complement Ther Clin Pract*. 2020;39:101165. <https://doi.org/10.1016/j.ctcp.2020.101165>.
225. Allawadhi P, Khurana A, Allawadhi S, Joshi K, Packirisamy G, Bharani KK. Nanoceria as a possible agent for the management of COVID-19. *Nano Today*. 2020;35:100982. <https://doi.org/10.1016/j.nano.2020.100982>.
226. Khurana I, Allawadhi P, Khurana A, Srivastava AK, Navik U, Banothu AK, et al. Can bilirubin nanomedicine become a hope for the management of COVID-19? *Med Hypotheses*. 2021;149:110534. <https://doi.org/10.1016/j.mehy.2021.110534>.
227. Allawadhi P, Singh V, Khurana I, Rawat PS, Renushe AP, Khurana A, et al. Decorin as a possible strategy for the amelioration of COVID-19. *Med Hypotheses*. 2021;152:110612. <https://doi.org/10.1016/j.mehy.2021.110612>.
228. Allawadhi P, Singh V, Khurana A, Khurana I, Allawadhi S, Kumar P, et al. Silver nanoparticle based multifunctional approach for combating

- COVID-19. *Sens Int.* 2021;2:100101. <https://doi.org/10.1016/j.sintl.2021.100101>.
229. Kalkal A, Allawadhi P, Pradhan R, Khurana A, Bharani KK, Packirisamy G. Allium sativum derived carbon dots as a potential theranostic agent to combat the COVID-19 crisis. *Sens Int.* 2021;2:100102. <https://doi.org/10.1016/j.sintl.2021.100102>.
230. Allawadhi P, Khurana A, Allwadi S, Navik US, Joshi K, Banothu AK, et al. Potential of electric stimulation for the management of COVID-19. *Med Hypotheses.* 2020;144:110259. <https://doi.org/10.1016/j.mehy.2020.110259>.
231. Basu D, Chavda VP, Mehta AA. Therapeutics for COVID-19 and post COVID-19 complications: an update. *Curr Res Pharmacol Drug Discov.* 2022;3:100086. <https://doi.org/10.1016/j.crphar.2022.100086>.
232. World Health Organization. Draft landscape and tracker of COVID-19 candidate vaccines. Available from: <https://www.who.int/publications/m/item/draft-landscape-of-covid-19-candidate-vaccines>
233. Goldman JD, Lye DCB, Hui DS, Marks KM, Bruno R, Montejano R, et al. Remdesivir for 5 or 10 days in patients with severe Covid-19. *N Engl J Med.* 2020;383(19):1827–37. <https://doi.org/10.1056/NEJMoa2015301>.
234. Spinner CD, Gottlieb RL, Criner GJ, Arribas Lopez JR, Cattelan AM, Soriano Viladomiu A, et al. Effect of Remdesivir vs standard care on clinical status at 11 days in patients with moderate COVID-19: a randomized clinical trial. *JAMA.* 2020;324(11):1048–57. <https://doi.org/10.1001/jama.2020.16349>.
235. Kalil AC, Patterson TF, Mehta AK, Tomashek KM, Wolfe CR, Ghazaryan V, et al. Baricitinib plus remdesivir for hospitalized adults with Covid-19. *N Engl J Med.* 2021;384(9):795–807. <https://doi.org/10.1056/NEJMoa2031994>.
236. Beigel JH, Tomashek KM, Dodd LE, Mehta AK, Zingman BS, Kalil AC, et al. Remdesivir for the treatment of Covid-19 - final report. *N Engl J Med.* 2020;383(19):1813–26. <https://doi.org/10.1056/NEJMoa2007764>.
237. Kreuzberger N, Hirsch C, Chai KL, Tomlinson E, Khosravi Z, Popp M, et al. SARS-CoV-2-neutralising monoclonal antibodies for treatment of COVID-19. *Cochrane Database Syst Rev.* 2021;9:CD013825. <https://doi.org/10.1002/14651858.CD013825.pub2>.
238. Nathan R, Shawa I, De La Torre I, Pustizzi JM, Hastrup N, Patel DR, et al. A narrative review of the clinical practicalities of bamlanivimab and etesevimab antibody therapies for SARS-CoV-2. *Infect Dis Ther.* 2021;10(4):1933–47. <https://doi.org/10.1007/s40121-021-00515-6>.
239. Hammond J, Leister-Tebbe H, Gardner A, Abreu P, Bao W, Wisemandle W, et al. Oral nirmatrelvir for high-risk, nonhospitalized adults with Covid-19. *N Engl J Med.* 2022;386(15):1397–408. <https://doi.org/10.1056/NEJMoa2118542>.
240. Lee CC, Hsieh CC, Ko WC. Molnupiravir-a novel oral anti-SARS-CoV-2 agent. *Antibiotics (Basel).* 2021;10(11):1294. <https://doi.org/10.3390/antibiotics10111294>.
241. Jayk Bernal A, Gomes da Silva MM, Musungaie DB, Kovalchuk E, Gonzalez A, Delos Reyes V, et al. Molnupiravir for oral treatment of Covid-19 in nonhospitalized patients. *N Engl J Med.* 2022;386(6):509–20. <https://doi.org/10.1056/NEJMoa2116044>.
242. Mollan KR, Eron JJ, Krajewski TJ, Painter W, Duke ER, Morse CG, et al. Infectious severe acute respiratory syndrome coronavirus 2 (SARS-CoV-2) virus in symptomatic coronavirus disease 2019 (COVID-19) outpatients: host, disease, and viral correlates. *Clin Infect Dis* 2021:ciab968. <https://doi.org/10.1093/cid/ciab968>.
243. Group AC-TflwC-S. Efficacy and safety of two neutralising monoclonal antibody therapies, sotrovimab and BRII-196 plus BRII-198, for adults hospitalised with COVID-19 (TICO): a randomised controlled trial. *Lancet Infect Dis.* 2021;22(5):622–35. [https://doi.org/10.1016/S1473-3099\(21\)00751-9](https://doi.org/10.1016/S1473-3099(21)00751-9).
244. Huang DT, McCreary EK, Bariola JR, Wadas RJ, Kip KE, Marroquin OC, et al. The UPMC OPTIMISE-C19 (OPTimizing treatment and impact of monoclonal antibodies through evaluation for COVID-19) trial: a structured summary of a study protocol for an open-label, pragmatic, comparative effectiveness platform trial with response-adaptive randomization. *Trials.* 2021;22(1):363. <https://doi.org/10.1186/s13063-021-05316-3>.
245. Gottlieb RL, Nirula A, Chen P, Boscica J, Heller B, Morris J, et al. Effect of Bamlanivimab as monotherapy or in combination with Etesevimab on viral load in patients with mild to moderate COVID-19: a randomized clinical trial. *JAMA.* 2021;325(7):632–44. <https://doi.org/10.1001/jama.2021.0202>.
246. Chen P, Nirula A, Heller B, Gottlieb RL, Boscica J, Morris J, et al. SARS-CoV-2 neutralizing antibody LY-CoV555 in outpatients with Covid-19. *N Engl J Med.* 2021;384(3):229–37. <https://doi.org/10.1056/NEJMoa2029849>.
247. Weinreich DM, Sivapalasingam S, Norton T, Ali S, Gao H, Bhore R, et al. REGEN-COV antibody combination and outcomes in outpatients with Covid-19. *N Engl J Med.* 2021;385(23):e81. <https://doi.org/10.1056/NEJMoa2108163>.
248. Weinreich DM, Sivapalasingam S, Norton T, Ali S, Gao H, Bhore R, et al. REGEN-COV, a neutralizing antibody cocktail, in outpatients with Covid-19. *N Engl J Med.* 2021;384(3):238–51. <https://doi.org/10.1056/NEJMoa2035002>.
249. O'Brien MP, Forleo-Neto E, Sarkar N, Isa F, Hou P, Chan KC, et al. Effect of subcutaneous Casirivimab and Imdevimab antibody combination vs placebo on development of symptomatic COVID-19 in early asymptomatic SARS-CoV-2 infection: a randomized clinical trial. *JAMA.* 2022;327(5):432–41. <https://doi.org/10.1001/jama.2021.24939>.
250. Irvin SC, Ganguly S, Weiss R, Elango C, Zhong X, Mao Y, et al. REGEN-COV(R) antibody cocktail bioanalytical strategy: comparison of LC-MRM-MS and immunoassay methods for drug quantification. *Bioanalysis.* 2021;13(24):1827–36. <https://doi.org/10.4155/bio-2021-0190>.
251. Chew KW, Moser C, Daar ES, Wohl DA, Li JZ, Coombs R, et al. Bamlanivimab reduces nasopharyngeal SARS-CoV-2 RNA levels but not symptom duration in non-hospitalized adults with COVID-19. *medRxiv.* 2021. <https://doi.org/10.1101/2021.12.17.21268009>.
252. Joyner MJ, Bruno KA, Klassen SA, Kunze KL, Johnson PW, Lesser ER, et al. Safety update: COVID-19 convalescent plasma in 20,000 hospitalized patients. *Mayo Clin Proc.* 2020;95(9):1888–97. <https://doi.org/10.1016/j.mayocp.2020.06.028>.
253. Yoon HA, Bartash R, Gendlina I, Rivera J, Nakouzi A, Bortz RH 3rd, et al. Treatment of severe COVID-19 with convalescent plasma in the Bronx, NYC. *medRxiv.* 2020. <https://doi.org/10.1101/2020.12.02.20242909>.
254. Dillner J, Ursing J. Convalescent plasma for treatment of COVID-19: study protocol for an open randomised controlled trial in Sweden. *BMJ Open.* 2021;11(12):e048337. <https://doi.org/10.1136/bmjopen-2020-048337>.
255. Korper S, Weiss M, Zickler D, Wiesmann T, Zacharowski K, Corman VM, et al. Results of the CAPSID randomized trial for high-dose convalescent plasma in patients with severe COVID-19. *J Clin Invest.* 2021;131(20):e152264. <https://doi.org/10.1172/JCI152264>.
256. Korley FK, Durkalski-Mauldin V, Yeatts SD, Schulman K, Davenport RD, Dumont LJ, et al. Early convalescent plasma for high-risk outpatients with Covid-19. *N Engl J Med.* 2021;385(21):1951–60. <https://doi.org/10.1056/NEJMoa2103784>.
257. Salazar E, Perez KK, Ashraf M, Chen J, Castillo B, Christensen PA, et al. Treatment of coronavirus disease 2019 (COVID-19) patients with convalescent plasma. *Am J Pathol.* 2020;190(8):1680–90. <https://doi.org/10.1016/j.ajpath.2020.05.014>.
258. Avendano-Sola C, Ramos-Martinez A, Munez-Rubio E, Ruiz-Antoran B, Malo de Molina R, Torres F, et al. A multicenter randomized open-label clinical trial for convalescent plasma in patients hospitalized with COVID-19 pneumonia. *J Clin Invest.* 2021;131(20):e152740. <https://doi.org/10.1172/JCI152740>.
259. O'Donnell MR, Grinsztejn B, Cummings MJ, Justman JE, Lamb MR, Eckhardt CM, et al. A randomized double-blind controlled trial of convalescent plasma in adults with severe COVID-19. *J Clin Invest.* 2021;131(13):e150646. <https://doi.org/10.1172/JCI150646>.
260. Self WH, Stewart TG, Wheeler AP, El Atrouni W, Bistran-Hall AJ, Casey JD, et al. Passive immunity trial for our nation (PassITON): study protocol for a randomized placebo-control clinical trial evaluating COVID-19 convalescent plasma in hospitalized adults. *Trials.* 2021;22(1):221. <https://doi.org/10.1186/s13063-021-05171-2>.
261. Axfors C, Janiaud P, Schmitt AM, Van't Hooft J, Smith ER, Haber NA, et al. Association between convalescent plasma treatment and mortality in COVID-19: a collaborative systematic review and meta-analysis of randomized clinical trials. *BMC Infect Dis.* 2021;21(1):1170. <https://doi.org/10.1186/s12879-021-06829-7>.

262. Levin MJ, Ustianowski A, De Wit S, Launay O, Avila M, Templeton A, et al. Intramuscular AZD7442 (Tixagevimab-Cilgavimab) for prevention of Covid-19. *N Engl J Med*. 2022. <https://doi.org/10.1056/NEJMoa2116620>.
263. Xie Y, Yin W, Zhang Y, Shang W, Wang Z, Luan X, et al. Design and development of an oral remdesivir derivative WV116 against SARS-CoV-2. *Cell Res*. 2021;31(11):1212–4. <https://doi.org/10.1038/s41422-021-00570-1>.
264. Qian HJ, Wang Y, Zhang MQ, Xie YC, Wu QQ, Liang LY, et al. Safety, tolerability, and pharmacokinetics of WV116, an oral nucleoside analog against SARS-CoV-2, in Chinese healthy subjects. *Acta Pharmacol Sin*. 2022. <https://doi.org/10.1038/s41401-022-00895-6>.
265. Mishra R, Kumar A, Ingle H, Kumar H. The interplay between viral-derived miRNAs and host immunity during infection. *Front Immunol*. 2019;10:3079. <https://doi.org/10.3389/fimmu.2019.03079>.
266. Tagawa T, Serquina A, Kook I, Ziegelbauer J. Viral non-coding RNAs: stealth strategies in the tug-of-war between humans and herpesviruses. *Semin Cell Dev Biol*. 2021;111:135–47. <https://doi.org/10.1016/j.semcdb.2020.06.015>.
267. Madhry D, Pandey KK, Kaur J, Rawat Y, Sapra L, YSR, et al. Role of non-coding RNAs in dengue virus-host interaction. *Front Biosci (Schol Ed)*. 2021;13(1):44–55. <https://doi.org/10.52586/S552>.
268. Zhang X, Ma X, Jing S, Zhang H, Zhang Y. Non-coding RNAs and retroviruses. *Retrovirology*. 2018;15(1):20. <https://doi.org/10.1186/s12977-018-0403-8>.
269. Lung RW, Tong JH, To KF. Emerging roles of small Epstein-Barr virus derived non-coding RNAs in epithelial malignancy. *Int J Mol Sci*. 2013;14(9):17378–409. <https://doi.org/10.3390/ijms140917378>.
270. Morales L, Oliveros JC, Fernandez-Delgado R, tenOever BR, Enjuanes L, Sola I. SARS-CoV-encoded small RNAs contribute to infection-associated lung pathology. *Cell Host Microbe*. 2017;21(3):344–55. <https://doi.org/10.1016/j.chom.2017.01.015>.
271. Meng F, Siu GK, Mok BW, Sun J, Fung KSC, Lam JY, et al. Viral micro-RNAs encoded by nucleocapsid gene of SARS-CoV-2 are detected during infection, and targeting metabolic pathways in host cells. *Cells*. 2021;10(7):1762. <https://doi.org/10.3390/cells10071762>.
272. Xiong Y, Liu Y, Cao L, Wang D, Guo M, Jiang A, et al. Transcriptomic characteristics of bronchoalveolar lavage fluid and peripheral blood mononuclear cells in COVID-19 patients. *Emerg Microbes Infect*. 2020;9(1):761–70. <https://doi.org/10.1080/22221751.2020.1747363>.
273. Li S, Wang J, Yan Y, Zhang Z, Gong W, Nie S. Clinical characterization and possible pathological mechanism of acute myocardial injury in COVID-19. *Front Cardiovasc Med*. 2022;9:862571. <https://doi.org/10.3389/fcvm.2022.862571>.
274. Gupta A, Madhavan MV, Sehgal K, Nair N, Mahajan S, Sehrawat TS, et al. Extrapulmonary manifestations of COVID-19. *Nat Med*. 2020;26(7):1017–32. <https://doi.org/10.1038/s41591-020-0968-3>.
275. Chen YM, Zheng Y, Yu Y, Wang Y, Huang Q, Qian F, et al. Blood molecular markers associated with COVID-19 immunopathology and multi-organ damage. *EMBO J*. 2020;39(24):e105896. <https://doi.org/10.15252/embj.2020105896>.
276. Ottolina D, Zazzeron L, Trevisi L, Agarossi A, Colombo R, Fossali T, et al. Acute kidney injury (AKI) in patients with Covid-19 infection is associated with ventilatory management with elevated positive end-expiratory pressure (PEEP). *J Nephrol*. 2022;35(1):99–111. <https://doi.org/10.1007/s40620-021-01100-3>.
277. Saleh J, Peyssonnaud C, Singh KK, Edeas M. Mitochondria and microbiota dysfunction in COVID-19 pathogenesis. *Mitochondrion*. 2020;54:1–7. <https://doi.org/10.1016/j.mito.2020.06.008>.
278. Moreno Fernandez-Ayala DJ, Navas P, Lopez-Lluch G. Age-related mitochondrial dysfunction as a key factor in COVID-19 disease. *Exp Gerontol*. 2020;142:111147. <https://doi.org/10.1016/j.exger.2020.111147>.
279. Shenoy S. Coronavirus (Covid-19) sepsis: revisiting mitochondrial dysfunction in pathogenesis, aging, inflammation, and mortality. *Inflamm Res*. 2020;69(11):1077–85. <https://doi.org/10.1007/s00011-020-01389-z>.
280. Gibellini L, De Biasi S, Paolini A, Borella R, Boraldi F, Mattioli M, et al. Altered bioenergetics and mitochondrial dysfunction of monocytes in patients with COVID-19 pneumonia. *EMBO Mol Med*. 2020;12(12):e13001. <https://doi.org/10.15252/emmm.202013001>.
281. Ding M, Zhang Q, Li Q, Wu T, Huang YZ. Correlation analysis of the severity and clinical prognosis of 32 cases of patients with COVID-19. *Respir Med*. 2020;167:105981. <https://doi.org/10.1016/j.rmed.2020.105981>.
282. Ontong P, Prachayasittikul V. Unraveled roles of hyaluronan in severe COVID-19. *EXCLI J*. 2021;20:117–25. <https://doi.org/10.17179/excli2020-3215>.
283. Monchi M, Bruneau T, Jochmans S, Veyer D, Pitsch A, Ellrodt O, et al. Association of high SARS-CoV-2 RNAemia with diabetes and mortality in critically ill COVID-19 patients. *iScience*. 2022;25(5):104075. <https://doi.org/10.1016/j.isci.2022.104075>.
284. O'Connell P, Aldhamen YA. Systemic innate and adaptive immune responses to SARS-CoV-2 as it relates to other coronaviruses. *Hum Vaccin Immunother*. 2020;16(12):2980–91. <https://doi.org/10.1080/21645515.2020.1802974>.
285. Grifoni A, Sidney J, Vita R, Peters B, Crotty S, Weiskopf D, et al. SARS-CoV-2 human T cell epitopes: adaptive immune response against COVID-19. *Cell Host Microbe*. 2021;29(7):1076–92. <https://doi.org/10.1016/j.chom.2021.05.010>.
286. Sette A, Crotty S. Adaptive immunity to SARS-CoV-2 and COVID-19. *Cell*. 2021;184(4):861–80. <https://doi.org/10.1016/j.cell.2021.01.007>.
287. Liu J, Guo J, Xu Q, Cai G, Chen D, Shen Y. Detection of IgG antibody during the follow-up in patients with COVID-19 infection. *Crit Care*. 2020;24(1):448. <https://doi.org/10.1186/s13054-020-03138-4>.
288. Brouqui P, Colson P, Melenotte C, Houhamdi L, Bedotto M, Devaux C, et al. COVID-19 re-infection. *Eur J Clin Investig*. 2021;51(5):e13537. <https://doi.org/10.1111/eci.13537>.
289. He S, Zhou K, Hu M, Liu C, Xie L, Sun S, et al. Clinical characteristics of "re-positive" discharged COVID-19 pneumonia patients in Wuhan, China. *Sci Rep*. 2020;10(1):17365. <https://doi.org/10.1038/s41598-020-74284-6>.
290. Elzein F, Ibrahim A, Alshahrani F, Mahrous M, Murshid E, Aldhehyan T, et al. Reinfection, recurrence, or delayed presentation of COVID-19? Case series and review of the literature. *J Infect Public Health*. 2021;14(4):474–7. <https://doi.org/10.1016/j.jiph.2021.01.002>.
291. Oran DP, Topol EJ. Prevalence of asymptomatic SARS-CoV-2 infection: a narrative review. *Ann Intern Med*. 2020;173(5):362–7. <https://doi.org/10.7326/M20-3012>.
292. Barboza JJ, Chambergo-Michilot D, Velasquez-Sotomayor M, Silva-Rengifo C, Diaz-Arocutipa C, Caballero-Alvarado J, et al. Assessment and management of asymptomatic COVID-19 infection: a systematic review. *Travel Med Infect Dis*. 2021;41:102058. <https://doi.org/10.1016/j.tmaid.2021.102058>.
293. Yu C, Zhou M, Liu Y, Guo T, Ou C, Yang L, et al. Characteristics of asymptomatic COVID-19 infection and progression: a multicenter, retrospective study. *Virulence*. 2020;11(1):1006–14. <https://doi.org/10.1080/21505594.2020.1802194>.
294. Chua RL, Lukassen S, Trump S, Hennig BP, Wendisch D, Pott F, et al. COVID-19 severity correlates with airway epithelium-immune cell interactions identified by single-cell analysis. *Nat Biotechnol*. 2020;38(8):970–9. <https://doi.org/10.1038/s41587-020-0602-4>.
295. Schulte-Schrepping J, Reusch N, Paclik D, Bassler K, Schlickeiser S, Zhang B, et al. Severe COVID-19 is marked by a dysregulated myeloid cell compartment. *Cell*. 2020;182(6):1419–40.e23. <https://doi.org/10.1016/j.cell.2020.08.001>.
296. Moore JB, June CH. Cytokine release syndrome in severe COVID-19. *Science*. 2020;368(6490):473–4. <https://doi.org/10.1126/science.abb8925>.
297. Huang C, Wang Y, Li X, Ren L, Zhao J, Hu Y, et al. Clinical features of patients infected with 2019 novel coronavirus in Wuhan, China. *Lancet*. 2020;395(10223):497–506. [https://doi.org/10.1016/s0140-6736\(20\)30183-5](https://doi.org/10.1016/s0140-6736(20)30183-5).
298. Zhang X, Tan Y, Ling Y, Lu G, Liu F, Yi Z, et al. Viral and host factors related to the clinical outcome of COVID-19. *Nature*. 2020;583(7816):437–40. <https://doi.org/10.1038/s41586-020-2355-0>.
299. Hamada A, Torre C, Drancourt M, Ghigo E. Trained immunity carried by non-immune cells. *Front Microbiol*. 2018;9:3225. <https://doi.org/10.3389/fmicb.2018.03225>.
300. Teijaro JR, Walsh KB, Cahalan S, Fremgen DM, Roberts E, Scott F, et al. Endothelial cells are central orchestrators of cytokine amplification during influenza virus infection. *Cell*. 2011;146(6):980–91. <https://doi.org/10.1016/j.cell.2011.08.015>.
301. Guerville C, Burtsey S, Sabatier F, Cauchois R, Lano G, Abdili E, et al. Circulating endothelial cells as a marker of endothelial injury in severe COVID-19. *J Infect Dis*. 2020;222(11):1789–93. <https://doi.org/10.1093/infdis/jiaa528>.



302. Buckley CD. Why does chronic inflammation persist: an unexpected role for fibroblasts. *Immunol Lett.* 2011;138(1):12–4. <https://doi.org/10.1016/j.imlet.2011.02.010>.
303. Nie X, Qian L, Sun R, Huang B, Dong X, Xiao Q, et al. Multi-organ proteomic landscape of COVID-19 autopsies. *Cell.* 2021;184(3):775–91.e14. <https://doi.org/10.1016/j.cell.2021.01.004>.
304. John AE, Joseph C, Jenkins G, Tatler AL. COVID-19 and pulmonary fibrosis: a potential role for lung epithelial cells and fibroblasts. *Immunol Rev.* 2021;302(1):228–40. <https://doi.org/10.1111/immr.12977>.
305. Vistejnova L, Safrankova B, Nesporova K, Slavkovsky R, Hermannova M, Hosek P, et al. Low molecular weight hyaluronan mediated CD44 dependent induction of IL-6 and chemokines in human dermal fibroblasts potentiates innate immune response. *Cytokine.* 2014;70(2):97–103. <https://doi.org/10.1016/j.cyto.2014.07.006>.
306. Baeva LF, Lyle DB, Rios M, Langone JJ, Lightfoote MM. Different molecular weight hyaluronic acid effects on human macrophage interleukin 1beta production. *J Biomed Mater Res A.* 2014;102(2):305–14. <https://doi.org/10.1002/jbm.a.34704>.
307. Lu CH, Lin CH, Li KJ, Shen CY, Wu CH, Kuo YM, et al. Intermediate molecular mass hyaluronan and CD44 receptor interactions enhance neutrophil phagocytosis and IL-8 production via p38- and ERK1/2-MAPK signalling pathways. *Inflammation.* 2017;40(5):1782–93. <https://doi.org/10.1007/s10753-017-0622-5>.
308. Haegel-Kronenberger H, de la Salle H, Bohbot A, Oberling F, Cazenave JP, Hanau D. Adhesive and/or signaling functions of CD44 isoforms in human dendritic cells. *J Immunol.* 1998;161(8):3902–11.
309. Shi W, Peng X, Liu T, Cheng Z, Lu H, Yang S, et al. A deep learning-based quantitative computed tomography model for predicting the severity of COVID-19: a retrospective study of 196 patients. *Ann Transl Med.* 2021;9(3):216. <https://doi.org/10.21037/atm-20-2464>.
310. Levi M, Thachil J, Iba T, Levy JH. Coagulation abnormalities and thrombosis in patients with COVID-19. *Lancet Haematol.* 2020;7(6):e438–40. [https://doi.org/10.1016/S2352-3026\(20\)30145-9](https://doi.org/10.1016/S2352-3026(20)30145-9).
311. Wang D, Hu B, Hu C, Zhu F, Liu X, Zhang J, et al. Clinical characteristics of 138 hospitalized patients with 2019 novel coronavirus-infected pneumonia in Wuhan, China. *JAMA.* 2020;323(11):1061–9. <https://doi.org/10.1001/jama.2020.1585>.
312. Singh V, Allawadhi P, Khurana A, Banothu AK, Bharani KK. Critical neurological features of COVID-19: role of imaging methods and biosensors for effective diagnosis. *Sens Int.* 2021;2:100098. <https://doi.org/10.1016/j.sintl.2021.100098>.
313. Shen B, Yi X, Sun Y, Bi X, Du J, Zhang C, et al. Proteomic and metabolomic characterization of COVID-19 patient sera. *Cell.* 2020;182(1):59–72.e15. <https://doi.org/10.1016/j.cell.2020.05.032>.
314. Song JW, Lam SM, Fan X, Cao WJ, Wang SY, Tian H, et al. Omics-driven systems interrogation of metabolic dysregulation in COVID-19 pathogenesis. *Cell Metab.* 2020;32(2):188–202.e5. <https://doi.org/10.1016/j.cmet.2020.06.016>.
315. Thomas T, Stefanoni D, Reisz JA, Nemkov T, Bertolone L, Francis RO, et al. COVID-19 infection alters kynurenine and fatty acid metabolism, correlating with IL-6 levels and renal status. *JCI Insight.* 2020;5(14):e140327. <https://doi.org/10.1172/jci.insight.140327>.
316. Matsuyama T, Yoshinaga SK, Shibue K, Mak TW. Comorbidity-associated glutamine deficiency is a predisposition to severe COVID-19. *Cell Death Differ.* 2021;28(12):3199–213. <https://doi.org/10.1038/s41418-021-00892-y>.
317. Andonegui-Elguera S, Taniguchi-Ponciano K, Gonzalez-Bonilla CR, Torres J, Mayani H, Herrera LA, et al. Molecular alterations prompted by SARS-CoV-2 infection: induction of hyaluronan, glycosaminoglycan and mucopolysaccharide metabolism. *Arch Med Res.* 2020;51(7):645–53. <https://doi.org/10.1016/j.arcmed.2020.06.011>.
318. Hellman U, Karlsson MG, Engstrom-Laurent A, Cajander S, Dorofte L, Ahlm C, et al. Presence of hyaluronan in lung alveoli in severe Covid-19: an opening for new treatment options? *J Biol Chem.* 2020;295(45):15418–22. <https://doi.org/10.1074/jbc.AC120.015967>.
319. Kaber G, Kratochvil MJ, Burgener EB, Peltan EL, Barlow G, Yang S, et al. Hyaluronan is abundant in COVID-19 respiratory secretions. *medRxiv.* 2020. <https://doi.org/10.1101/2020.09.11.20191692>.
320. Bhattacharya J, Cruz T, Bhattacharya S, Bray BA. Hyaluronan affects extravascular water in lungs of unanesthetized rabbits. *J Appl Physiol* (1985). 1989;66(6):2595–9. <https://doi.org/10.1152/jappl.1989.66.6.2595>.
321. Laurent TC, Laurent UB, Fraser JR. The structure and function of hyaluronan: an overview. *Immunol Cell Biol.* 1996;74(2):A1–7. <https://doi.org/10.1038/icb.1996.32>.
322. Xu Z, Shi L, Wang Y, Zhang J, Huang L, Zhang C, et al. Pathological findings of COVID-19 associated with acute respiratory distress syndrome. *Lancet Respir Med.* 2020;8(4):420–2. [https://doi.org/10.1016/s2213-2600\(20\)30076-x](https://doi.org/10.1016/s2213-2600(20)30076-x).
323. Yang S, Ling Y, Zhao F, Li W, Song Z, Wang L, et al. Hymecromone: a clinical prescription hyaluronan inhibitor for efficiently blocking COVID-19 progression. *Signal Transduct Target Ther.* 2022;7(1):91. <https://doi.org/10.1038/s41392-022-00952-w>.
324. Qin C, Zhou L, Hu Z, Zhang S, Yang S, Tao Y, et al. Dysregulation of immune response in patients with coronavirus 2019 (COVID-19) in Wuhan, China. *Clin Infect Dis.* 2020;71(15):762–8. <https://doi.org/10.1093/cid/ciaa248>.
325. McKallip RJ, Do Y, Fisher MT, Robertson JL, Nagarkatti PS, Nagarkatti M. Role of CD44 in activation-induced cell death: CD44-deficient mice exhibit enhanced T cell response to conventional and superantigens. *Int Immunol.* 2002;14(9):1015–26. <https://doi.org/10.1093/intimm/14/9/1015>.
326. Zhang W, Li L, Liu J, Chen L, Zhou F, Jin T, et al. The characteristics and predictive role of lymphocyte subsets in COVID-19 patients. *Int J Infect Dis.* 2020;99:92–9. <https://doi.org/10.1016/j.ijid.2020.06.079>.
327. Bernard GR, Artigas A, Brigham KL, Carlet J, Falke K, Hudson L, et al. The American-European consensus conference on ARDS. Definitions, mechanisms, relevant outcomes, and clinical trial coordination. *Am J Respir Crit Care Med.* 1994;149(3 Pt 1):818–24. <https://doi.org/10.1164/ajrccm.149.3.7509706>.
328. Hascall VC. The journey of hyaluronan research in the journal of biological chemistry. *J Biol Chem.* 2019;294(5):1690–6. <https://doi.org/10.1074/jbc.TM118.005836>.
329. Grandoch M, Bollyky PL, Fischer JW. Hyaluronan: a master switch between vascular homeostasis and inflammation. *Circ Res.* 2018;122(10):1341–3. <https://doi.org/10.1161/CIRCRESAHA.118.312522>.
330. Queisser KA, Mellema RA, Middleton EA, Portier I, Manne BK, Denorme F, et al. COVID-19 generates hyaluronan fragments that directly induce endothelial barrier dysfunction. *JCI Insight.* 2021:e147472. <https://doi.org/10.1172/jci.insight.147472>.
331. Mambetsariev N, Mirzapoiazova T, Mambetsariev B, Sammani S, Lennon FE, Garcia JG, et al. Hyaluronic acid binding protein 2 is a novel regulator of vascular integrity. *Arterioscler Thromb Vasc Biol.* 2010;30(3):483–90. <https://doi.org/10.1161/ATVBAHA.109.200451>.
332. Farkash EA, Wilson AM, Jentzen JM. Ultrastructural evidence for direct renal infection with SARS-CoV-2. *J Am Soc Nephrol.* 2020;31(8):1683–7. <https://doi.org/10.1681/ASN.2020040432>.
333. Nicin L, Abplanalp WT, Mellentin H, Kattih B, Tombor L, John D, et al. Cell type-specific expression of the putative SARS-CoV-2 receptor ACE2 in human hearts. *Eur Heart J.* 2020;41(19):1804–6. <https://doi.org/10.1093/eurheartj/ehaa311>.
334. Xiao F, Tang M, Zheng X, Liu Y, Li X, Shan H. Evidence for gastrointestinal infection of SARS-CoV-2. *Gastroenterology.* 2020;158(6):1831–3.e3. <https://doi.org/10.1053/j.gastro.2020.02.055>.
335. Necas J, Bartosikova L, Brauner P, Kolar JJV. Hyaluronic acid (hyaluronan): a review. *Vet Med-Czech.* 2008;53(8):397–411. <https://doi.org/10.17221/1930-VETMED>.
336. Tang N, Li D, Wang X, Sun Z. Abnormal coagulation parameters are associated with poor prognosis in patients with novel coronavirus pneumonia. *J Thromb Haemost.* 2020;18(4):844–7. <https://doi.org/10.1111/jth.14768>.
337. Stavenuiter F, Ebberink E, Mertens K, Meijer AB. Role of glycine 221 in catalytic activity of hyaluronan-binding protein 2. *J Biol Chem.* 2017;292(15):6381–8. <https://doi.org/10.1074/jbc.M116.757849>.
338. Lu L, Zhong W, Bian Z, Li Z, Zhang K, Liang B, et al. A comparison of mortality-related risk factors of COVID-19, SARS, and MERS: a systematic review and meta-analysis. *J Inf Secur.* 2020;81(4):e18–25. <https://doi.org/10.1016/j.jinf.2020.07.002>.
339. Chen H, Guo J, Wang C, Luo F, Yu X, Zhang W, et al. Clinical characteristics and intrauterine vertical transmission potential of COVID-19 infection in nine pregnant women: a retrospective review of medical records. *Lancet.* 2020;395(10226):809–15. [https://doi.org/10.1016/S0140-6736\(20\)30360-3](https://doi.org/10.1016/S0140-6736(20)30360-3).

340. Barrero-Castillero A, Beam KS, Bernardini LB, Ramos EGC, Davenport PE, Duncan AR, et al. COVID-19: neonatal-perinatal perspectives. *J Perinatol*. 2021;41(5):940–51. <https://doi.org/10.1038/s41372-020-00874-x>.
341. Whitaker M, Elliott J, Chadeau-Hyam M, Riley S, Darzi A, Cooke G, et al. Persistent COVID-19 symptoms in a community study of 606,434 people in England. *Nat Commun*. 2022;13(1):1957. <https://doi.org/10.1038/s41467-022-29521-z>.
342. Group P-CC. Clinical characteristics with inflammation profiling of long COVID and association with 1-year recovery following hospitalisation in the UK: a prospective observational study. *Lancet Respir Med*. 2022. [https://doi.org/10.1016/S2213-2600\(22\)00127-8](https://doi.org/10.1016/S2213-2600(22)00127-8).
343. Sudre CH, Murray B, Varsavsky T, Graham MS, Penfold RS, Bowyer RC, et al. Attributes and predictors of long COVID. *Nat Med*. 2021;27(4):626–31. <https://doi.org/10.1038/s41591-021-01292-y>.
344. Lindqvist U, Laurent TC. Serum hyaluronan and aminoterminal propeptide of type III procollagen: variation with age. *Scand J Clin Lab Invest*. 1992;52(7):613–21. <https://doi.org/10.3109/00365519209115504>.
345. Mine S, Okada Y, Kawahara C, Tabata T, Tanaka Y. Serum hyaluronan concentration as a marker of angiopathy in patients with diabetes mellitus. *Endocr J*. 2006;53(761–6):0609110032. <https://doi.org/10.1507/endocrj.K05-119>.
346. Kalay N, Elcik D, Canatan H, Kaya MG, Yarliogluoglu M, Oguzhan A, et al. Elevated plasma hyaluronan levels in pulmonary hypertension. *Tohoku J Exp Med*. 2013;230(1):7–11. <https://doi.org/10.1620/tjem.230.7>.
347. Karbownik MS, Nowak JZ. Hyaluronan: towards novel anti-cancer therapeutics. *Pharmacol Rep*. 2013;65(5):1056–74. [https://doi.org/10.1016/S1734-1140\(13\)71465-8](https://doi.org/10.1016/S1734-1140(13)71465-8).
348. Kobayashi H, Sun GW, Tanaka Y, Kondo T, Terao T. Serum hyaluronic acid levels during pregnancy and labor. *Obstet Gynecol*. 1999;93(4):480–4. [https://doi.org/10.1016/S0029-7844\(98\)00526-2](https://doi.org/10.1016/S0029-7844(98)00526-2).
349. Brindley LL, Sweet JM, Goetzl EJ. Stimulation of histamine release from human basophils by human platelet factor 4. *J Clin Invest*. 1983;72(4):1218–23. <https://doi.org/10.1172/JCI111077>.
350. Rosenkranz AR, Wekkeli M, Hippmann G, Benda H, Jarisch R, Gotz M. Cold urticaria as a model of mediator release: platelet factor 4, eosinophil cationic protein and histamine. *Allergy*. 1992;47(4 Pt 2):366–70. <https://doi.org/10.1111/j.1398-9995.1992.tb02073.x>.
351. Soderberg M, Bjermer L, Hallgren R, Lundgren R. Increased hyaluronan (hyaluronic acid) levels in bronchoalveolar lavage fluid after histamine inhalation. *Int Arch Allergy Appl Immunol*. 1989;88(4):373–6. <https://doi.org/10.1159/000234719>.
352. Lennon FE, Singleton PA. Hyaluronan regulation of vascular integrity. *Am J Cardiovasc Dis*. 2011;1(3):200–13.
353. Marsh K, Tayler R, Pollock L, Roy K, Lakha F, Ho A, et al. Investigation into cases of hepatitis of unknown aetiology among young children, Scotland, 1 January 2022 to 12 April 2022. 2022;27(15):2200318. <https://doi.org/10.2807/1560-7917.ES.2022.27.15.2200318>.
354. Neuman MG, Cohen LB, Nanau RM. Hyaluronic acid as a non-invasive biomarker of liver fibrosis. *Clin Biochem*. 2016;49(3):302–15. <https://doi.org/10.1016/j.clinbiochem.2015.07.019>.
355. Plevris JN, Haydon GH, Simpson KJ, Dawkes R, Ludlum CA, Harrison DJ, et al. Serum hyaluronan—a non-invasive test for diagnosing liver cirrhosis. *Eur J Gastroenterol Hepatol*. 2000;12(10):1121–7. <https://doi.org/10.1097/00042737-200012100-00009>.
356. Bennett CF. Therapeutic antisense oligonucleotides are coming of age. *Annu Rev Med*. 2019;70:307–21. <https://doi.org/10.1146/annur-ev-med-041217-010829>.
357. Dhuri K, Bechtold C, Quijano E, Pham H, Gupta A, Vikram A, et al. Antisense oligonucleotides: an emerging area in drug discovery and development. *J Clin Med*. 2020;9(6):2004. <https://doi.org/10.3390/jcm9062004>.
358. Aartsma-Rus A, Krieg AM. FDA approves eteplirsen for duchenne muscular dystrophy: the next chapter in the eteplirsen saga. *Nucleic Acid Ther*. 2017;27(1):1–3. <https://doi.org/10.1089/nat.2016.0657>.
359. Neuman BW, Stein DA, Kroeker AD, Churchill MJ, Kim AM, Kuhn P, et al. Inhibition, escape, and attenuated growth of severe acute respiratory syndrome coronavirus treated with antisense morpholino oligomers. *J Virol*. 2005;79(15):9665–76. <https://doi.org/10.1128/JVI.79.15.9665-9676.2005>.
360. Ahn DG, Lee W, Choi JK, Kim SJ, Plant EP, Almazan F, et al. Interference of ribosomal frameshifting by antisense peptide nucleic acids suppresses SARS coronavirus replication. *Antivir Res*. 2011;91(1):1–10. <https://doi.org/10.1016/j.antiviral.2011.04.009>.
361. Le TK, Paris C, Khan KS, Robson F, Ng WL, Rocchi P. Nucleic acid-based technologies targeting coronaviruses. *Trends Biochem Sci*. 2021;46(5):351–65. <https://doi.org/10.1016/j.tibs.2020.11.010>.
362. Khurana A, Allawadhi P, Khurana I, Allawadhi S, Weiskirchen R, Banothu AK, et al. Role of nanotechnology behind the success of mRNA vaccines for COVID-19. *Nano Today*. 2021;38:101142. <https://doi.org/10.1016/j.nantod.2021.101142>.
363. Group RC, Horby P, Lim WS, Emberson JR, Mafham M, Bell JL, et al. Dexamethasone in hospitalized patients with Covid-19 – preliminary report. *N Engl J Med*. 2020;NEJMoa2021436. <https://doi.org/10.1056/NEJMoA2021436>.
364. Tomazini BM, Maia IS, Cavalcanti AB, Berwanger O, Rosa RG, Veiga VC, et al. Effect of dexamethasone on days alive and ventilator-free in patients with moderate or severe acute respiratory distress syndrome and COVID-19: the CoDEX randomized clinical trial. *JAMA*. 2020;324(13):1307–16. <https://doi.org/10.1001/jama.2020.17021>.
365. Luo P, Qiu L, Liu Y, Liu XL, Zheng JL, Xue HY, et al. Metformin treatment was associated with decreased mortality in COVID-19 patients with diabetes in a retrospective analysis. *Am J Trop Med Hyg*. 2020;103(1):69–72. <https://doi.org/10.4269/ajtmh.20-0375>.
366. Gebhardt C, Averbek M, Diedenhofen N, Willenberg A, Anderegg U, Sleeman JP, et al. Dermal hyaluronan is rapidly reduced by topical treatment with glucocorticoids. *J Invest Dermatol*. 2010;130(1):141–9. <https://doi.org/10.1038/jid.2009.210>.
367. Sainio A, Takabe P, Oikari S, Salomaki-Myftari H, Koulu M, Soderstrom M, et al. Metformin decreases hyaluronan synthesis by vascular smooth muscle cells. *J Investig Med*. 2020;68(2):383–91. <https://doi.org/10.1136/jim-2019-001156>.
368. Kultti A, Pasonen-Seppanen S, Jauhiainen M, Rilla KJ, Karna R, Pyoria E, et al. 4-Methylumbelliferone inhibits hyaluronan synthesis by depletion of cellular UDP-glucuronic acid and downregulation of hyaluronan synthase 2 and 3. *Exp Cell Res*. 2009;315(11):1914–23. <https://doi.org/10.1016/j.yexcr.2009.03.002>.
369. Nagy N, Kuipers HF, Frymoyer AR, Ishak HD, Bollyky JB, Wight TN, et al. 4-methylumbelliferone treatment and hyaluronan inhibition as a therapeutic strategy in inflammation, autoimmunity, and cancer. *Front Immunol*. 2015;6:123. <https://doi.org/10.3389/fimmu.2015.00123>.
370. Simons FE. Advances in H1-antihistamines. *N Engl J Med*. 2004;351(21):2203–17. <https://doi.org/10.1056/NEJMr033121>.
371. Liyanage CK, Galappaththy P, Seneviratne SL. Corticosteroids in management of anaphylaxis; a systematic review of evidence. *Eur Ann Allergy Clin Immunol*. 2017;49(5):196–207. <https://doi.org/10.23822/EurAnnACI.1764-1489.15>.
372. Chang TW. The pharmacological basis of anti-IgE therapy. *Nat Biotechnol*. 2000;18(2):157–62. <https://doi.org/10.1038/72601>.
373. Song TZ, Zheng HY, Han JB, Jin L, Yang X, Liu FL, et al. Delayed severe cytokine storm and immune cell infiltration in SARS-CoV-2-infected aged Chinese rhesus macaques. *Zool Res*. 2020;41(5):503–16. <https://doi.org/10.24272/j.issn.2095-8137.2020.202>.
374. Baudar C, Duprez T, Kassab A, Miller N, Rutgers MP. COVID-19 as triggering co-factor for cortical cerebral venous thrombosis? *J Neuroradiol*. 2021;48(1):65–7. <https://doi.org/10.1016/j.neurad.2020.06.008>.
375. Mukudai S, Matsuda KI, Nishio T, Sugiyama Y, Bando H, Hirota R, et al. Differential responses to steroid hormones in fibroblasts from the vocal fold, trachea, and esophagus. *Endocrinology*. 2015;156(3):1000–9. <https://doi.org/10.1210/en.2014-1605>.
376. Yang S, Chen L, Tong Y, Yu W. Viral miRNA-mediated activation of hyaluronan production as a drug target against COVID-19. *Acta Pharm Sin B*. 2022. <https://doi.org/10.1016/j.apsb.2022.03.022>.

## Publisher's Note

Springer Nature remains neutral with regard to jurisdictional claims in published maps and institutional affiliations.