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Medicinal pteridophytes: ethnopharmacological, phytochemical, and clinical attributes

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Abstract

Background: Almost from the very beginning of human existence, man has been interacting with plants. Throughout human history, plants have provided humans with basic needs such as sustenance, firewood, livestock feed, and wood. The world has approximately 3 million vascular plants. The treatment of primary health problems is provided primarily by traditional medicines by around 80% of the world's population. Compared to other vascular plants, pteridophytes remain underexplored in ethnobotanical aspects, despite being regarded as a valuable component of healthcare for centuries. As an alternative medicine, pteridophytes are being investigated for their pharmacological activity. Almost 2000 years ago, humans were exploring and using plant species from this lineage because of its beneficial properties since pteridophytes were the first vascular plants.

Main body of the abstract: All popular search engines such as PubMed, Google Scholar, ScienceDirect, and Scopus were searched to retrieve the relevant literature using various search strings relevant to the topic. Pteridophytes belonging to thirty different families have been documented as medicinal plants. For instance, *Selaginella* sp. has been demonstrated to have numerous therapeutic properties, including antioxidative, inflammation-reducing, anticarcinogenic, diabetes-fighting, virucidal, antibacterial, and anti-senile dementia effects. In addition, clinical trials and studies performed on pteridophytes and derived compounds are also discussed in details.

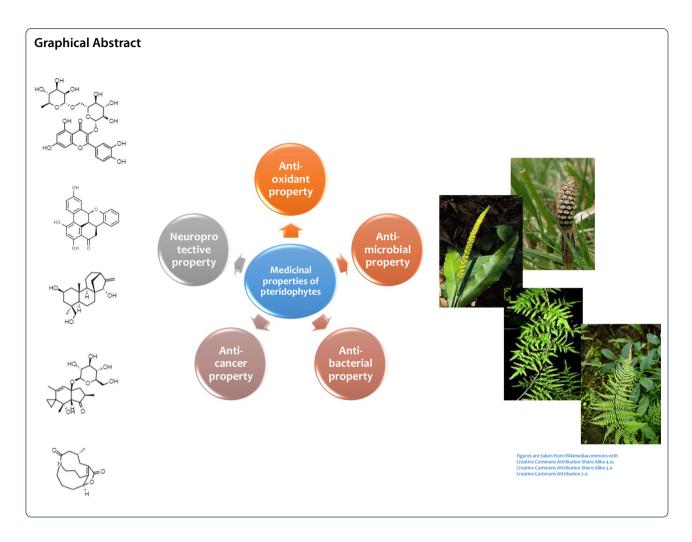
Short conclusion: This review offers a compilation of therapeutically valuable pteridophytes utilized by local ethnic groups, as well as the public.

Keywords: Pteridophytes, Ethnobotany, Pharmacology, Phytochemistry

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1 Background

People from all continents have utilized medicinal plants as a remedy for several ailments since ancient times [47, 48, 114]. While synthetic organic chemistry has experienced significant growth in the twentieth century, plants are responsible for producing nearly one-quarter of all pharmaceuticals approved by the drug administration boards [120]. Phytochemicals that have antioxidant, antibacterial, or inflammation-fighting abilities have become increasingly sought after for their prospective applications in combating a variety of chronic diseases and infections [6, 70, 134]. Alternative sources of antimicrobial agents, such as plants, might offer potent bioactivity against microbes with pathogenicity, while having fewer adverse effects [11, 13]. There are a number of secondary metabolites in plants, including phenols, which are biologically active [8, 16]. Antimicrobial and antioxidant properties are known to be associated with plant secondary metabolites, with some of them receiving general safety approval [135]. Due to the increasing demand for phyto-compounds, there is a great demand for plants with biomedical properties [3, 33].

Many plants contain phytochemicals that are used as components of the diets of humans and animals. Fruits, nuts, and greens, for example, contain phytochemicals [123]. As plants proceed through their normal metabolic processes, they form phytochemicals. The term "secondary metabolites" refers to the many kinds of chemicals which can be found in plants, namely alkaloids, coumarins, flavonoids, glycosides, gums, phenols, polysaccharides, tannins, terpenes, and terpenoids [80, 126]. Plants also contain other chemical compounds in addition to these. They can be used as adjuvants to reduce unwanted side effects of the active ingredients or as means of assisting with their absorption. Plants are capable of synthesizing an almost infinite number of aromatic compounds, of which 12,000 have been found [104]. The researchers estimate that this is only a small fraction of the total. The active compounds of plants are present in different parts, which can influence the functioning of organs in both humans and animals. Phytochemicals can protect plants from predation by microbes, pests, and grazers, while also exhibiting therapeutic properties for a variety of ailments.

There are over 12,000 species in Ferns and their allies, distributed over 250 different genera in the Pteridophyta [31]. According to Sushruta and Charka, some ferns are prescribed in the Samhita texts of the Ayurvedic systems of medicine. In Unani medicine as well, pteridophytes are used [164]. Native doctors in China recommend ferns in the traditional Chinese medical system [82]. Recent studies have investigated ferns and their allies from an ethnobotanical and advanced pharmacological point of view [34, 35, 50, 51, 140, 169]. Inhibition of the cyclo oxygenase pathway can be associated with anti-inflammatory properties of fern secondary metabolites, flavonoids [95]. Moreover, they possess antioxidative, cancer-preventive, and antibacterial activities [2, 67, 97, 99, 189]. There are a variety of bioactive flavonoids in spider brake fern (Pteris multifida), that have thermoregulation, antifever, cleansing, antibiotic, inflammation-relieving, and mutagen-suppressive properties [93]. Various bioactive compounds are present in the plant of P. multifida, namely, 16-hydroxy-kaurane-2-β-D-glucoside [98], apigenin 4-O-α-L-rhamnoside [100, 137], apigenin-7-O-β-D-glucoside [73, 100], hyperin, isoquercitrin, kaempferol, luteolin, luteolin-7-O-glucoside [117], palmitic acid, quercetin and rutin [73, 100, 175]. Furthermore, P. multifida contains bioactive compounds that are cytotoxic to cells, pterosin sesquiterpenes [152]. Terpenoids with bioactive properties, particularly monocyclic sesquiterpene α-caryophyllene, and extracts of Pteris tripartita were found containing these compounds. In addition to acting as anti-carcinogenic, inflammation-reducing, cytotoxic, phytoregulatory, and antibacterial substances, sesquiterpenes are also an important group of secondary metabolites [9]. Humans have known for more than 2000 years that the pteridophytes are medicinal plants. In modern chemotherapy, however, there are very few applications compared to the angiosperms. Drynaria quercifolia has been reported to help with flatulence, heartburn, constipation, inflammatory conditions, typhoid, flu, osteoarthritis, migraines, and nausea. Hemoptysis is treated with a rhizome decoction either fresh or dried. Astringents and antiparasitic drugs are also used as a result of its properties. Digestive problems can also be treated using its leaves.

2 Ethnomedicinal properties of pteridophytes

All types of diseases are treated with traditional medicine in urban, rural, and areas rich in aboriginal people [49, 58]. Pteridophytes were first studied by Caius in India for their medicinal properties [20]. The knowledge

of ethnobotanical and medicinal uses of pteridophytes was exposed by [38, 124, 124, 170]. The spores and rhizomes of Sporophytes have been proven to have medicinal properties. Many diseases can be cured with the plant extract of pteridophytes. Several pteridophytes are reported to cure human disorders by Theophrastus and Dioscorides in their works from 327 to 287 BC. Also mentioned by Shushurta and Charak are specific ferns such as Marsilea minuta and Adiantum capillus-veneris. There is evidence that Selaginella bryopteris and Lycopodium clavatum are frequently prescribed for neurological disorders and to address the effects of heat stroke in the homeopathic system of medicine. Splinted bones can be treated with L. clavatum. Herb Ayurvedic formulations contain Helenthostachys zeylanica (Kamraj) as a sexual stimulant and a tonic. Known to boost immunity, marsiline is isolated from Marsilea minuta and is also known to treat a variety of conditions, including sleeplessness, flatulence, cough and indigestion. A number of gastrointestinal bacterial strains have been shown to be resistant to the antimicrobial properties of Pteris vittata. In addition, pteridophytes would prove themselves to be a vital biological resource for the benefit of humanity due to their enormous importance and vast medicinal scope. Plants, such as ferns, are used as fodder and food in tropical countries. In addition to serving as a supplementary food source, dried fern biomass is used as a strong insulation against extreme temperatures and as an effective absorber of urinal excreta in cattle sheds. Listed below are some important plants and their ethnobotanical significance.

2.1 Acrostichum aureum L. (Pteridaceae)

A paste made from leaves is used to treat headaches. Tender leaves can be eaten raw or cooked as a vegetable. Besom and fish attractors can be made from the fronds. Paste of rhizomes is frequently used for healing wounds and boils [149].

2.2 Actiniopteris dichotoma Kuhn. (Actiniopteridaceae)

A number of conditions are treated with rhizomes, including chronic cough, gallstones, gastrointestinal problems, infection, Hansen's disease, skin conditions, high blood sugar, and hyperthermia. For sore throats, the leaves are chewed, and to remove dandruff, the rhizomes are boiled and then applied [158].

3 A. radiata (J. Koenig ex Sw.) Link (Actiniopteridaceae)

Rhizomes and leaves are employed as disinfectants, to treat asthma, and for disorders related to the female reproductive system. Tuberculosis is treated with dried fronds [149, 156]. Watery stool and high temperatures

are treated using the juice from the stem twice daily [79]. For treatment of leukorrhoea and fertility enhancement, fresh leaves paste or powdered leaves are taken twice a day with honey [20]. Five to six leaves are taken twice a day with sugar as a philter and tonic to stimulate fertility in women [127].

3.1 Adiantum capillus-veneris L. (Adiantaceae)

Aphrodisiac properties are associated with rhizomes and leaves [12]. India's indigenous community uses the juice of fresh plants to treat cough and blood sugar levels. The juice is also administered to children as a nutritional supplement. High temperatures and acute respiratory infections may be treated by leaf extraction. Chicken pox can be controlled by placing rhizomes and leaves next to the bed [156]. In combination with honey, the leaves are used to make ophthalmic to speed up the healing process, applying Aloe vera gel along with paste made from leaves, stems, and rhizomes is prevalent among several communities. To promote hair growth, the leaves, stems, and roots of the plant are mashed together [79]. Astringent and diuretic properties have also been attributed to it [149]. Leaf chewing is used to treat mouth ulcers. Leaf, stem, and rhizome pastes are also used as urine promoters, stimulants, mucoprotective agent, mucoactive agent, and anti-pyretic agent [166].

3.2 A. caudatum L.

Foliage extracts are used to treat wounds, to get relief in cough and fever [144]. Antihelmintic properties are associated with rhizomes [149]. In addition to its uses as an Mucoactive agent, it is used in the treatment of dermatological conditions, blood sugar disorders, coughing, and high temperatures [158, 161]. Blisterd, wounds, and abrasions can be treated with leaf paste.

3.3 A. incisum forsk

Leaf dust is consumed with butter, thus relieving the feeling of burning in the body [144]. Additionally, it is used for cough relief, lowering blood sugar levels, and for treating dermatological disorders [156, 166].

3.4 A. lunulatum Burm.

Acute abdominal cramps, Hansen's disease, and flu are treated by this plant. For a centipede bite, fronds and rhizomes are mixed together into a paste. Blood-related diseases, such as epilepsy and rabies, can also be treated with it. Rhizomes are recommended for strange symptoms and fever caused by elephantiasis. Loose motion, soreness, and discomfort are treated with crushed leaves and juice [161].

3.5 A. philippense L.

Chronic nasal congestion, cough, bronchitis, swelling of the throat, and swollen glands of the face can be treated with the plant. The fresh frond extract is used for treating dysentery, blood diseases, ulcers, and burning sensations. For immediate relief from gastric problems, fresh frond paste about 1 gm can be given twice daily on an unfilled stomach for two weeks.

Curing glandular swelling with rhizome is an effective remedy. A powdered rhizome of about one gram combined with water can be used as a contraceptive by tribal women once every 3 to 5 days during menstruation.

Menstrual irregularities are treated with fresh leaf extracts [158–160]. In addition to treating respiratory diseases, it also helps in treating pyrexia, Hansen's disease and thinning hair [155, 166]. This substance is found in Hansraj, the Indian cough medicine. As a remedy for pyrexia and Filariasis, rhizome is considered useful [83]. Also, rhizome is used as an antidote when treating Snakebite envenoming and dog bites [158].

Cheilanthes bicolor (Roxb.) Griff. ex Fras.-Jenk. (Sinopteridaceae)

In Indian medicine, leaves and rhizomes are used as a stimulant and also as a remedy for preventing fear among children. Young girls use the stems as nasal and ear ornaments [156].

3.7 C. farinosa (Forsk.) Kaulf

The leaves of the plant are brewed into a decoction, which is taken orally for seven days to treat menstrual irregularities. Additionally, the decoction is used to treat colds and pyrexia. To cure wounds and Atopic dermatitis, rhizome paste is applied. Stomach pain is treated with extracts of rhizome [158].

3.8 C. tenuifolia (Burm.f.) Sw.

To liberate pus from abscesses, leaves paste is applied as a cataplasm or used in pyrexia. Indian tribal people drink a bouillon of roots and rhizomes to improve their immunity [53].

3.9 Diplazium esculentum (Retz.) Sw. (Athyriaceae)

The leaves and stems of this plant are either fried or consumed raw. Rhizome and root extracts are used as general stimulants by indigenous people [53]. Roots have medicinal properties. The infusion of the plant is used as a hair cleanser and nourisher. Pests and insects are kept at bay by storing rhizome in granaries. As a green vegetable, young fronds are suitable for a salad or for cooking [156, 161]. Sarcoptic mange is treated with a tincture of leaves.

Various respiratory ailments, cough, and icterus can be treated with the juice. Broth made from of young fronds is effective in treating toothaches and dental caries. As a protection against labor pains, pregnant women eat fronds and leaves [122].

3.10 *Helminthostachys zeylanica* (L.) Hook. (Helminthostachyaceae)

Sciatic neuritis is treated using this plant because it is sedative, pain-relieving, and inflammation-reducing. Fronds have been used as sexual stimulant, euphoriant, tranquilizer [156]. Rhizomes are used to treat pertussis and in sexual dysfunction.

3.11 Hypodematium crenatum (Forssk.) Kuhn (Woodsiaceae)

A powder made from dried fronds blended within cow milk can be taken by women after five days of period for about a week to improve conception [122].

3.12 Nephrolepis cordifolia (L.) Presl, (Nephrolepidaceae)

To prevent bleeding from wounds, frost's paste is applied. Phytochemicals in pinna have bactericidal and fungicidal properties and are used in cough, cuts, and icterus treatment [88]. Approximately 10–15 ml of rhizome extract is administered once during menstruation to sterilize a woman permanently. It is used as a birth control method [50, 51, 159, 160]. The rhizome of this plant has bactericidal activity and is used in, Joint pain, seasonal allergies, nasal congestion, and appetite loss. [157]. Tuber extracts are used for digestive problems. Fresh tubers used to treat ulcers, and in indigestion it is simmered in brine water and consumed to soothe stomach upset [156, 166].

3.13 Tectaria cicutaria (L.) Copal (Tectariaceae)

Antibacterial properties are present in the plant. Chest cold, stings from honeybees, and breathing problems are treated with it [157]. Children with irregular bowel movements and dysentery can benefit from a decoction made from the fresh root. [54]. The roots can be used as a stimulant, blood cleanser, and infectious diarrhea. In cases of menstrual problems, rhizomes containing *Zingiber purpureum* and *Croton roxburgii* are prescribed [158]. A stomach-ache is treated with the fresh boiled stem [105].

3.14 Tectaria coadunata C. Chr.

A condition that causes inflammatory conditions of the colon's inner lining can be treated with an extract from the plant. Children suffering from stomach problems are given root extracts [166]. For those suffering from chest cold, leaves are extracted or leaves combined with honey are administered. Woodcutters use leaves paste to alleviate pain caused by centipede and honey bee stings [150].

3.15 Tectaria wightii (Clarke) Ching.

Disinfectant are made from the roots of this plant [54].

3.16 Thelypteris arida D.Don (Thelypteridaceae)

On lesions and abrasions, leaves and rhizome paste are applied [166].

3.17 Vittaria elongata Sw. (Vittariaceae)

Its leaves are used to treat joint pain in Andaman and Nicobar Islands. [88].

3.18 Woodwardia unigemmata (Makino) Nakai (Blechnaceae)

Diseases like dysentery, abdominal pain may be treated with it [62]

4 Medicinal properties of pteridophytes

4.1 Antioxidant property

There is a growing body of evidence that phenolic compounds in plants have many potential health benefits. Phenolic compounds are by far the most widespread secondary metabolites in plants. When plants are stressed, they produce phenolic compounds. A recent study showed the signaling enzyme phenylalanine ammonia lyase (PAL) is involved in the synthesis of flavonoids, especially anthocyanins and flavones [138] and the presence of phenolics protects DNA from UV-B damage and thereby prevents DNA dimerization and degradation [43]. Thus, plants growing in high mountains must cope with issues such as low temperatures, low partial pressure of oxygen, high UV levels, and arid conditions which result in the accumulation of flavonoids [145]. A high total phenolic content (TPC) can partly be explained by the presence of this conditions in places where the ferns are found. Like, C. latebrosa, C. barometz, D. quercifolia, B. orientale and D. linearis. Their habitats include areas with direct sunlight, as well as mountainous terrain up to an altitude of 1500-1700 m.

Several fern species have been reported to have antioxidant activity, including *A. penangiana, Braomea* insignis, *Cheilanthes anceps* Swartz, *Davallia divaricata*,

Table 1 Pteridophytes and their antioxidant potential

Family	Species	Method	References
Thelypteridaceae	Abacopteris penangiana	TEAC assay	Zhao et al. [194]
Sinopteridaceae	Cheilanthes anceps	DPPH assay	Chowdhary et al. [37]
Selaginellaceae	Selaginella sp.	In vitro lipid peroxidation	Gayathri et al. [63]
Selaginellaceae	Selaginella labordei	In vitro xanthine oxidase inhibition	Tan et al. [163]
Pteridaceae	Pteris ensiformis	TEAC assays	Wei et al. [180]
Pteridaceae	Pteris multifida	Superoxide scavenging activity	Wang et al[177]
Pteridaceae	Pteris tripartita	Metal chelating activity	Baskaran and Jeyachandran [10]
Pteridaceae	Pteris vittata	DPPH assays	Lai and Lim, [90]
Polypodiaceae	Polypodium leucotomus	FRAP assays	Gombau et al. [65]
Marsileaceae	Marsilea quadrifolia	DPPH assay	Ripa et al. [142]
Gleicheniaceae	Dicranopteris linearis	DPPH assay	[112]
Equisetaceae	Equesitum sp.	Total antioxidant capacity	Milovanovic et al. [113]
Dryopteridaceae	Polystichum semifertile	ABTS• + assay	Chen et al. [34, 35]
Dryopteridaceae	Nothoperanema hendersonii	ABTS• + assay	Chen et al. [34, 35]
Blechnaceae	Blechnum orientale	DPPH assay	Lai et al. [89]
Athyriaceae	Diplazium polypodioides	DPPH assay	Kshirsagar and Upadhyay [87]

Davallia mariesii, Davallia solida, Dicranopteris linearis, Drynaria fortunei, Drynaria quercifolia, E. arvense, Equesitum sylvaticum, Humata griffithiana, Marsilea quadrifolia, Nothoperanema hendersonii, P. multifida, P. tripartita, Polyopodium leucotomus, Polystichum semifertile, Pseudodrynaria coronans, Pteris ensiformis Burm., Selaginella sp. [10, 30, 35–37, 61, 63, 65, 112, 113, 142, 153, 177, 180, 191, 194]. Table 1 represents a list of pteridophytes and their antioxidant potential.

4.2 Anti-microbial property

Antibiotics are available for treating a wide range of conditions currently, namely chloramphenicol, tetracycline, erythromycin, penicillin G, ampicillin, cephalosporin, ciprofloxacin, kanamycin, gentamicin, neomycin, amoxycillin, nystatin, amphotericin-B, and ketaconazole. Despite the availability of several antibiotics to treat bacterial and fungal infections, these medications may not always be effective at combating pathogens. A number of ferns were used in ancient Indian medicine to treat a variety of ailments. Some ferns were recommended for medicinal use by Susruta and Charaka.

In high concentrations of extract and their solvents, pteridophyte extracts and their solvents effectively inhibited microbial growth. *Lygodium altum* extracts in both ethanol and acetone displayed good activity against *B. cerus. Salvinia molesta* showed considerable inhibitory activity against *B. cerus* only in its methanolic extract. *Salvinia cuculata* and *Helminthostachys zeylanica*, two plants whose solvent extracts have no

antibacterial activity, don't produce such results. Its antimicrobial properties cover both gram-positive and gramnegative bacteria, with *Dryopteris filixmas* showing the strongest antimicrobial effects. Antimicrobial substances are produced by three species of ferns. Folk medicine commonly uses the ferns to treat dermal infections, tonsillitis, abscesses, blisters, ulcers, and skin wounds [7].

4.3 Antibacterial property

An alcoholic and an aqueous extract of the leaves of A. lunulatum and A. pectinatum, as well as aqueous leaf extracts from D. cochleata and M. minuta and alcoholic extractions of C. dentatus and H. crenatum did not show any inhibitory effects on A. tumefaciens. With the exception of water-soluble and alcoholic extracts of A. pectinatum, aqueous extract of A. incisum and H. crenatum and alcoholic extract of C. dentatus, each of which was effective for combating E.coli. Furthermore, both water-soluble and alcoholic extracts of the leaves of M. minuta inhibited the disease-causing strain of E. coli better than tetracycline. Additionally, water-soluble and alcoholic extracts of A. incisum and D. cochleata, as well as aqueous extracts from A. capillus-veneris and M. minuta and alcoholic extract of leaves of C. albomarginata, C. dentatus and H. crenatum failed to inhibit Salmonella arizonae. But few other extracts showed effectiveness in combating bacteria. In addition, with the exception of the alcoholic and aqueous extracts of A.pectinatum and the aqueous extract of leaves of C. albomarginata, each of these extracts has demonstrated inhibition of *S. typhi*. Also found to be active were water soluble and alcoholic extracts of the leaves of *A. pectinatum*, aqueous extract of leaves of *A. incisum* and *D. cochleata* and alcoholic extract of leaves of *C. dentatus* and *D. cochleata*, neither of these compounds inhibited the growth of *S. aureus*. [128]

4.4 Anti-carcinogenic properties

Plants, marine organisms, and microorganisms are some of the most common natural sources of anticancer agents [41, 119]. Most cytostatic drugs are derived from plants—paclitaxel, vinblastine and vincristine, topotecan and irinotecan, the camptothecin derivatives, and etoposide.

There are reports that Microsorum grossum can cure liver cancer [44, 130]. A. evecta [44] and Pteris polyphylla [93] also possess anticancer properties. In Chinese traditional medicine, Gushuibu is regarded as having anticancer and anti-inflammatory properties. Various ferns' rhizomes are used to make it, including *D. fortunei* (Kze.) J. Sm., P. coronans (Wall. Ex Mett.) Ching, D. divaricata BL., D. mariesii Moore ex-Bak, D. solida (Forst.) Sw., and H. griffithiana (Hk.) C. Chr. [19, 32, 39, 42]. Medicinal uses of M. quadrifolia include treating various cancers, diabetic conditions, and diseases related to inflammation and diuresis [165]. Phlebodium decumanum, also known as the bear paw fern, has been used for centuries as an anticancer and ulcer remedy [136]. Foliage and rootstocks of *H. arifolia* (Burm.) Moore and complete specimens of Adiantum capillus-veneris L. are used by the tribal communities of Tamil Nadu for hypoglycaemia and cancer prevention [146].

4.5 Lycopodiales and Isoetales

Lycopodiales, which contain more than 1,200 known existing species, and the associated orders Sellaginelales and Isoetales [21], are characterized by a distinctive set of secondary metabolic compounds. Lycopodiaceae has been reported to contain more than 200 quinolizidine, pyridine, and alpha-pyridone alkaloids, including lycopodine, lycodine, and fawcettimine [55, 102]. In terms of their anticholinergic and inflammation-reducing properties, some of the alkaloids show potential [102, 174]. There are also anticancer flavones in the Lycopodiaceae and Isoetaceae, including apigenin, chrysoeriol, luteolin and tricin, as well as their O-glycosylated or C-glycosylated forms [22, 66, 107]. Furthermore, lycopodine was found to reduce viability with a concentration of 50 μg/mL that can trigger cell death in HeLa cells [106]. In looking into the mechanisms of action, Bishayee et al. [14] revealed that *L. clavatum* spore-derived lycopodine inhibited cell growth and caused apoptosis in prostate cancer cells. In turn, this occurs because of downregulation of 5-lipoxygenase, the 5-oxo-ETE receptor (OXE receptor1) and the EGF receptor, which results in depolarization of the membrane potential of the mitochondria, but without affecting p53 activity.

4.6 Phenolic compounds

A majority of the anticancer activity of ferns is reported to be attributed to phenolic compounds, particularly flavonoids and their O-glycosides. In vivo as well as in vitro, flavonoids are powerful antioxidants with anticancer properties. Flavonoids from different plants, including ferns, show both of these properties [24]. Ferns have been studied extensively for their patterning, and they are probably present in all of them. The amentoflavone found in Psilotaceae and its monomer apigenin is primarily contained in this family of plants [22, 172]. It has only been discovered that biflavonoids occur in Osmundaceae and Cyatheaceae so far [66]. Flavonols are the main constituents of Ophioglossaceae [66, 179]. Flavonoids have been found to be abundant in Helminthostachys zeylanica (L.) Hook [75]. Marratiales contain flavones in large amounts [173], flavonols are less common. In almost all of the leptosporangiate fern families, different flavonoid types can be detected, including flavonols. Several flavonoids derived from ferns along with their O-glycosides can be found in most plant kingdoms. Flavonoids such as quercetin and kaempferol; the flavanone naringenin; the flavones apigenin, isoorientin, tricetin, vicenin-2, and vitexin; and the 3-deoxyanthocyanins apigenidin and luteolinidin, etc. Among the mechanisms by which they work are a reduction in DNA polymerase activity, suppression of NF-κB, decreased Bcl-2 expression, as well as suppression of MMP-1, p38 MAPK, and c-Jun N-terminal kinase (JNK activation. There is consequently an increase in apoptosis, a decline in cell growth, and migration and invasion of cells [74, 96, 109, 118, 151, 181, 184, 185]. Table 2 presents the diverse phytochemical compositions of pteridophytes.

4.7 Selaginella

There are approximately 700 extant species of *Selaginella* (spikemoss) found in the tropics [131]. Species found in primary tropical rainforests have the widest diversity of spikemoss, as well as xerophytes that can withstand drought and plants indigenous to tundra biomes in the arctic and alpine regions of the world. Many morphological [76] and molecular phylogenetic classifications have been published [85]. In some cases, it has also been shown that extracts of some plants of various taxa, mostly originating in Asia, including South, Southeast, and East Asia, but also from Australia and the Pacific, can inhibit the growth of cancer cells. *S.doederleinii* Hieron's crude extract inhibited cancer cell viability [86] in addition to reversing multidrug resistance [59]. Patients with

Table 2 Diverse phytochemical compositions of Pteridophytes

Family	Species	Biologically active chemicals	References
Equisetaceae	i. Equisetum arvense	Equise to lic acid, kaempferol, peteosins n, querce tinglucoside	Dos Santos Jr. et al. [56], Singh et al. [154]
Gleicheniaceae	i. D. prelata ii. Dicranopteris dichotoma, iii. G. blotiana, iv. G. hirta, v. Gleichenia quadriparitta,	Ecdysteroids	Cao et al. [23]
Helminthostachyaceae	i. Helminthostachys zeylanica	Prenylated flavonoids, thermalic acids, ugonins	Cao et al. [23]
Huperziaceae	i. Huperzia serrata ii. Lycopodium serratum iii. Phlegariurus spp.	Huperzine, lycobeline alkaloids, lycoclavanol, lycopodine, serratine, serratine-diols	Jiang et al.[77] Ma et al. [103] Yang et al. [186] Ying et al.[188]
Lygodiaceae	i. L. japonicum ii. Lygodium venustum	Acacetin, phenyl propanoids, glucosides, rutinoside	Cao et al. [23] Morais-Braga et al. [116]
Ophioglossaceae	i. Botrychium ternatum ii. Ophioglossum. petiolotum, iii. O. thermale, iv.O. pedunculosum,	Glucopyranosides, kaempferol, ophioglonin, quercetin glycosides	Cao et al. [23]
Polypodiaceae	i. Polypodium hastata ii. P. triloba iii. P. leucotomos	Caffeic acid, coumaric acid, gallic acid, glucopyranoside	Cao et al. [23]
Pteridaceae	i. P. aquilinum ii. P. esculentum iii. P. Multifida, iv. P. Semipinnata v. Pityrogramma calomelanos vi. Pteridium spp. vii. Pteris semipinnata	Apigenin, caffeic acid, kaurene, kauroic acid, luteolin, multifedoside ptaquiloside, pterosine, rutin	Bai et al. [5] Reinaldo et al. [141] Wang et al. [178]
Selaginellaceae	i . Selaginella longistrobilina ii. S. bryopteris, iii. S. delicatula, iv. S. involvens, v. S. tamariscina, vi. S. wangpeishanii vii. S. amassae,	Biflavonoids-amentoflavone, hinokiflavone, involvenflavone; phenolics, selaginellins, terpenoids	Chandran and Muralidhara [27, 28] Chandran et al. [26, 29] Girish and Muralidhara [64] Ha et al. [68] Lee et al. [92]

cancer who were administered it produced intrinsic tumour necrosis factor [183]. Cao et al. [25] evaluated the cytotoxic potential of flavonoid extracts of several ferns and their allies against adenocarcinoma cells. There is a Southeast-Asian variant that has proven to be particularly effective, the *Selaginella frondosa* Warb. The mitotic activity of glandular stomach epithelium was reduced in rats exposed to a carcinogen followed by feeding *S.tamariscina* (Beauv.) Spring [94].

4.8 Neuroprotective properties

First, antioxidants and anti-inflammatory effects are evaluated in order to determine the neuromodulatory potency of phytochemicals. Among the most widespread fern families, Pteridaceae plants grow in a range of habitats, including tropical to temperate, on flat to hilly terrain, in arid and irrigated conditions, and along the coast and in interior areas. The pharmacological applications of members of the Pteridaceae have been cited in tribal medicine. Researchers have isolated a

variety of bioactive compounds from the family Pteridaceae, including alkaloids, flavonoids, and derivatives of their glucosides including kaurene, kauroic acid, apigenin, caffeic acid, rutin, luteolin, ptaquiloside, and pterosine [5, 178]. The cognitive symptoms of Alzheimer's disease can be alleviated with acetylcholinesterase inhibitors. Phytoextracts of Huperzia can be used to treat schizophrenia [77], strains, swells, and contusions [77]. A notable fern bioactive is huperzine A, from Huperzia spp. that has been found to inhibit Acetylcholinesterase (AChE) activity [103]. As an acetylcholinesterase inhibitor (AChEI), hyperzine A can be used to treat mild-moderate cognitive decline, and its safety, tolerability, and efficacy have also been evaluated in clinical trials [139]. Patients using this alkaloid reported improvements in learning, and it has been reported that rats using it demonstrated neuroprotective properties by protecting cortical neurons against oxidative stress and amyloid beta-induced apoptosis, which is an inherent feature of Alzheimer's disease.

Accordingly, huperzine A has emerged as a major lead for finding a new anti-Alzheimer's drug [162, 182]. There are currently drugs available in China and Europe for Alzheimer's disease and related diseases based on hyperzine A. Research shows that hyperzine A is a good AChEI that has a higher in-mouth bioavailability than some AChEIs, crosses the blood—brain barrier easily, and also has a longer in vivo half-life. In the

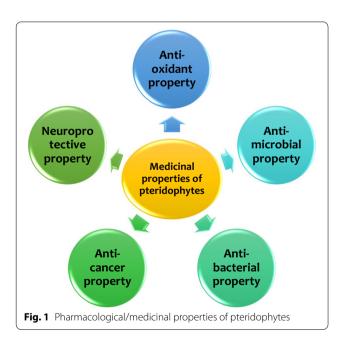
same manner as huperzine, its analogues like 12-deoxyhuperzine O also inhibit NMDA receptors with an impressively low IC50 value (0.92 μ M) [186]. *Huperzia* bioactive has neuroameliorating properties that extend beyond its AChEI activity. Huperzine A and specific triterpenoids in alcohol extractions of *H. serrata* have been demonstrated to possess growth inhibitory effects against human leukemia cells (HL-60) through

Table 3 Pharmacologically significant pteridophyte species and their few representative bioactive chemical compounds

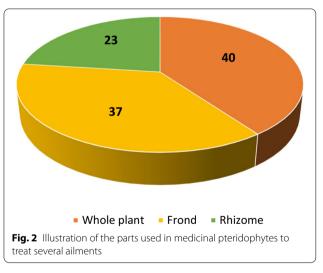
Class	Bioactive compounds	IUPAC name	Family
Alkaloids	Huperzine A (C ₁₅ H ₁₈ N ₂ O)	(1R,9R,13E)-1-Amino-13-ethylidene-11-methyl-6-azatricy-clo[7.3.1.0 ^{2,7}]trideca-2(7),3,10-trien-5-one	Lycopodiaceae
	Huperzine B (C ₁₆ H ₂₀ N ₂ O)	(1R,9R,10R)-16-Methyl-6,14-diazatetracyclo[7.5.3.0 ^{1,10} .0 ^{2,7}] heptadeca-2(7),3,16-trien-5-one	
	Huperzine R (C ₁₅ H ₂₁ NO ₃₎	(1S,6R)-6-Methyl-2-oxa-9-azatricyclo[7.4.3.0 ^{4,13}]hexadec-4(13)-ene-3,8-dione	
	Cernuine (C ₁₆ H ₂₆ N ₂ O)	(3aS,5S,6aR,7aS,12aS)-5-Methyltetradecahydro- 11H-pyrido[1',2':3,4]pyrimido[2,1,6-de]quinolizin-11-one	
	Lycocernuine (C ₁₄ H ₁₄ O ₂₎	3-[2-(4-Hydroxyphenyl)ethyl]phenol	
	Lycopodine (C ₁₆ H ₂₅ NO)	(15R)-15-Methyllycopodan-5-one	
Flavonoids	Apigenin (C ₁₅ H ₁₀ O ₅₎	5,7-Dihydroxy-2-(4-hydroxyphenyl)-4H-chromen-4-one	Marattiaceae
	Luteolin (C ₁₅ H ₁₀ O ₆₎	2-(3,4-Dihydroxyphenyl)-5,7-dihydroxy-4H-chromen-4-one	
	Violanthin $(C_{27}H_{30}O_{14)}$	5,7-Dihydroxy-2-(4-hydroxyphenyl)-6-[(2S,3R,4R,5S,6R)-3,4,5-trihydroxy-6-(hydroxymethyl)tetrahydro-2H-pyran-2-yl]-8-[(2S,3R,4R,5R,6S)-3,4,5-trihydroxy-6-methyltetrahydro-2H-pyran-2-yl]-4H-chromen-4-on e	
	Isoviolanthin $(C_{27}H_{30}O_{14)}$	5,7-Dihydroxy-2-(4-hydroxyphenyl)-8-[(25,3R,4R,55,6R)-3,4,5-trihydroxy-6-(hydroxymethyl)tetrahydro-2H-pyran-2-yl]-6-[(25,3R,4R,5R,6S)-3,4,5-trihydroxy-6-methyltetrahydro-2H-pyran-2-yl]-4H-chromen-4-on e	
	Muxiangrine III (C ₂₃ H ₂₄ O ₆)	2-[3,4-dihydroxy-5-(3-methylbut-2-enyl)phenyl]-5-hydroxy-7-methoxy-6,8-dimethylchromen-4-one	
Phenolics	Rutin (C ₂₇ H ₃₀ O ₁₆₎	2-(3,4-Dihydroxyphenyl)-5,7-dihydroxy-4-oxo-4H-chromen-3-yl 6-O-(6-deoxy-α-L-mannopyranosyl)-β-D-glucopyranoside	Pteridaceae, Adiantaceae Aspleniaceae, Daval-
	Cinnamic acids (C ₉ H ₈ O ₂₎	(E)-3-Phenylprop-2-enoic acid	liaceae
	caffeic acid (C ₉ H ₈ O ₄)	(2E)-3-(3,4-Dihydroxyphenyl)acrylic acid	
	Quinic acid (C ₇ H ₁₂ O)	(1S,3R,4S,5R)-1,3,4,5-Tetrahydroxycyclohexanecarboxylic acid	
	Catechin (C ₁₅ H ₁₄ O ₆₎	(2R,3S)-2-(3,4-Dihydroxyphenyl)-3,5,7-chromanetriol	
	Coumarin (C ₉ H ₆ O ₂₎	2H-Chromen-2-one	
	Anthraquinone (C ₁₄ H ₈ O ₂₎	9,10-Anthracenedione	
	Dihydrochalcone (C ₁₅ H ₁₄ O)	1,3-Diphenyl-1-propanone	
Diterpenoid	Pterokaurane M1(C ₂₀ H ₃₂ O ₃₎	(2β,5β,8α,9β,10α,13α,15α)-Kaur-16-ene-2,15,19-triol	Pteridaceae
Sesquiterpenoids	2,5,7-Trimethyl-indan-1-one (C ₁₂ H ₁₄ O)	2,5,7-Trimethyl-1-indanone	Pteridaceae
	Pterosin Z ($C_{15}H_{20}O_{2}$)	6-(2-Hydroxyethyl)-2,2,5,7-tetramethyl-1-indanone	
	Ptaquiloside (C ₂₀ H ₃₀ O ₈₎	(2'R,3a'R,4'S,7a'R)-4'-Hydroxy-2',4',6'-trimethyl-3'-oxo-2',3',3a',4'-tetrahydrospiro[cyclopropane-1,5'-inden]-7a'(1'H)-yl β-D-glucopyranoside	
Triterpenoids	$β$ -sitosterol ($C_{29}H_{50}O$)	(3β)-Stigmast-5-en-3-ol	Polypodiaceae
	Scaphopetalone (C ₂₁ H ₂₆ O ₆₎	4-[(1R,2S,3S)-2,3-Bis(hydroxymethyl)-6,7-dimethoxy-1,2,3,4-tetrahydro-1-naphthalenyl]-2-methoxyphenol	
	Neohop-13(18)-ene (C ₃₀ H ₅₀₎	(3R,3aR,5aS,5bR,7aS,11aS,11bR)-3-Isopropyl-3a,5a,5b,8,8,11a-hexamethyl-2,3,3a,4,5,5a,5b,6,7,7a,8,9,10,11,11a,11b,12,13-octadecahydro-1H-cyclopenta[a]chrysene	
	Diploptene (C ₃₀ H ₅₀₎	Hop-22(29)-ene	

Table 4 Medicinal properties of some pteridophytes

Name	Part used	Medicinal properties	
Adiantum capillusveneris	Entire plant	Treats constipation, common cold, growth of hair follicles, Irregular menstruation, snake bite	
Adiantum caudatum	Entire plant	High blood glucose, dermal disorders, common cold	
Adiantum incisum	Entire plant	Urinary tract infection, high blood glucose, dermal disorders	
Adiantum philippens	Entire plant	Infectious diarrhea, promotes diuresis, growth of hair follicles, cuts and bruises, induces femal infertility	
Adiantum venustum	Entire plant	Promotes diuresis, insect bites, cuts and bruises, cluster headache	
Marsilea minuta	Entire plant	Dermatitis, sleeplessness, sore muscle, tranquilizer	
Onychium japonicum	Entire plant	Dermatitis, pyrexia	
Diplazium esculentum	Entire plant, fronds	Pyrexia, Infectious diarrhea, stimulant, promotes bowel movement, fatigue	
Pteris cretica	Entire plant, fronds	Cuts and bruises, pyrexia	
Selaginella chrysocaulos	Entire plant, Spores	Pyrexia	
Asplenium dalhousiae	Fronds	Burns and blisters, dermal disorders, pyrexia	
Asplenium tricomanes	Fronds	Treats constipation, Irregular menstruation, hepatic disorders, treats parasitic infections	
Onychium contiguum	Fronds	Urinary tract infections	
Pteris vittata	Fronds	Burns and blisters	
Hypodematium crenatum	Fronds	Female fertility, insect bite	
Athyrium pectinatum	Rhizome	Treats parasitic infections	
Cheilanthes bicolor	Rhizome	Pyrexia, stimulant	
Noodwardia unigemmata	Rhizome, frond	Infectious diarrhea	
Dryopteris cochleata	Rhizome, fronds	Dermatitis, blood cleanser, stimulant, cuts and bruises, infections in GI tract, sore muscle	



induction of proapoptotic mechanisms [71]. Furthermore, huperzine A has been shown to stimulate the production of new hippocampal neurons and alleviate cognitive impairment in rats undergoing acute hypobaric hypoxia [101]. An interesting finding was that in a spinal cord injury model it significantly ameliorated



chronic pain phenotypes in rats [190]. Table 3 presents the pharmacologically significant pteridophyte species and their few representative bioactive chemical compounds. Table 4 presents medicinal properties of some pteridophytes. Figure 1 presents some pharmacological/medicinal properties of pteridophytes. Figure 2 illustrates the parts used in medicinal pteridophytes to treat several ailments. Figure 3 represents some phytochemicals of pteridophytes (from ChemDraw).

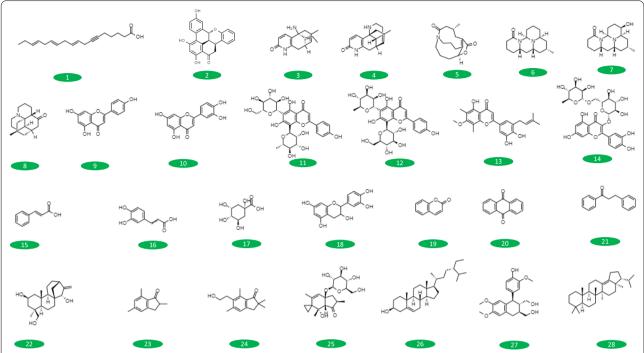


Fig. 3 Some phytochemicals of pteridophytes (from ChemDraw). 1. Dicranin, 2. Ohioensin A, 3. Huperzine A, 4. Huperzine B, 5. Huperzine R, 6. Cernuine, 7. Lycocernuine, 8. Lycopodine, 9. Apigenin, 10. Luteolin, 11. Violanthin, 12. Isoviolanthin, 13. Muxiangrine III, 14. Rutin, 15. Cinnamic acid, 16. Caffeic acid, 17. Quinic acid, 18. Catechin, 19. Coumarin, 20. Anthraquinone, 21. Dihydrochalcone, 22. Pterokaurane M1, 23. 2,5,7-Trimethyl-indan-1-one, 24. Pterosin Z, 25. Ptaquiloside, 26. β-sitosterol, 27. Scaphopetalone, 28. Neohop-13(18)-ene

4.9 Clinical trials

The saponin calagualine was shown to have anti-tumor activity in clinical trials and studies performed in vitro and in vivo in 1967 [18]. Also, humans who have uncontrolled, abnormal growth of tissues or cells benefited from the procedure without causing any complications. When the extract was administered orally to patients at high risk of melanoma, sensitivity to UV radiation was significantly reduced. [1]. The anti-neoplasic activity of antioxidants and immunomodulators is a topic of further study. Rhizome saponins inhibited the insertion of nucleoproteins into cells in an antagonistic manner to that of cytostatics [168]. On hairless mice, researchers observed reductions in UV-induced expression of Cox-2, inflammatory responses, and gene mutation [192]; suppression of excessive epithelial cell growth, elevated p53 activity, and increased antioxidant capacity in the blood [143]; and retardation of skin tumor formation.

Currently, only dermatological benefits of *Phlebodium* spp extract are known. In the treatment of psoriasis, it has been validated in clinical trials since 1974 [125]. When administered to normal volunteers, it reduced lymphocyte proliferation in response to mutation-triggering agents, OKT8+cells, and immunoglobulin levels. It may explain why the extract benefited patients with psoriasis

[78]. A placebo-controlled, randomized study found the extract to be more effective than standard treatment in treating psoriasis [45]. Volunteers also reported protection from UV-induced skin damage [111] and from skin cancer caused by ultraviolet radiation [110].

Apart from this, pteridophyte-derived compounds are also effective against neurodegenerative disorders. Despite their distinct clinical and etiopathogenic differences, neurodegenerative disorders share common traits, including protein deposition abnormalities, dysfunctional cellular transport, impaired mitochondrial function, inflammatory response, excessive Ca2+ inside the cell, uncontrolled ROS production, and overstimulation of receptors [52, 84]. In addition, reactive astroglia and microglia play an important role in the pathogenesis of all essential neurodegenerative disorders [4, 57]. It has been suggested that natural substances can be used in conjunction with conventional medicines to help treat neurodegenerative disorders [52]. Since Alzheimer's disease is a neurodegenerative disease with numerous causes and a complex pathophysiology, the development of drugs that offer a combination of multiple targets is becoming increasingly important for treating Alzheimer's disease. Omega-3 fatty acids, for example, have anti-inflammatory properties and inhibit cell toxicity in Alzheimer's [17]. Plant-derived compounds, including lunasin and tannins, may be useful in treating Alzheimer's disease [46]. The Hup A treatment has been used on more than 1 million people in China. Based on these studies, Hup A improves cognitive functioning and is a feasible medication. 202 Alzheimer's disease patients were enrolled in a double-blind, random, trial involving numerous hospitals in 2002 by Zhang et al. Hup A was administered to 100 patients for 12 weeks at 400 mg per day, and placebos were given to 102 patients. Alzheimer's disease Assessment Scale results showed cognitive improvements in the treatment group, as well as behavior improvements and improvements in daily living skills. [193].

In the same way, treatments for Parkinson's Disorder often involve Levodopa, which, in the brain, is almost exclusively transformed into dopamine with the help of the enzyme dopa decarboxylase, thus producing therapeutic benefits [69, 91]. Neuronal mitochondrial dysfunction has been linked to the progression of Parkinson's Disorder [115, 129]. A major contributing factor to the malfunction is excessive buildup of alpha-synuclein. The mitochondrial dysfunction leads to oxidative stress and neuronal degeneration. [81, 171]. A second peculiar feature of Parkinson's Disorder is the presence of neuroinflammation, which appears to contribute to the progression of the disease. Furthermore, inflammation is also dependent on mitochondrial dysfunction, which activates microglia and produces proinflammatory mediators, namely cytokines, chemokines, complement, and ROS [40]. In addition, most Parkinson's Disorder patients also struggle with non-motor complications, including sleep-wake cycle disorders, cognitive impairments, and mood disorders. All of these symptoms are often worsened by impaired sensory perception and pain. There has been a recent increase in the use of functional foods or nutritional supplements in the management of diseases associated with aging, such as Parkinson's. There is no doubt that diets high in vitamins, dietary minerals, and dietary salts may reduce symptoms of Parkinson's disease and its associated pathologies [40]. Pteridophyte-derived phytochemicals have been shown to improve motor function, reduce oxidative stress, and enhance neurogenesis [15]. Patients with Parkinson's disease were treated with Hup A (0.1 mg) in conjunction with cognitive retraining exercises, as demonstrated by Yao and his colleagues. Yao et al. found that the patients' cognitive function enhanced after six weeks of three daily treatments. [187]. Additionally, Wang et al. administered 0.3 mg Hup A subcutaneously 30 min in advance of general anesthesia to 15 patients. During recovery, acetylcholine levels in the brain were found to be elevated, a factor that helped restore cholinergic nerve conduction [176]. Diabetes patients who took Hup A (0.1 mg) three times daily for eight weeks exhibited improved Mini-mental state examination scores, latency and amplitude. The trial course lengths and sample numbers were small, and positive controls did not exist for most of the trials.

4.10 Negative effect of pteridophytes

Cancer has been linked to only one plant in the vascular system, Pteridium aquilinum (L.) Kuhn. Bracken is the most common fern in the world today, and it is also an extremely aggressive weed. The toxic effects of bracken, including its carcinogenic, mutagenic and teratogenic effects on animals and human beings, have been thoroughly examined several times. Bovine enzootic haematuria is caused by feeding on bracken in cattle and sheep. It causes tumours in the bladder which can be either benign or malignant. Additionally, it leads to the formation of rumen papillomas, squamous cell malignancies of the gastrointestinal tract, and cancer of fibroblasts of teeth [108]. A water extract of bracken has been shown to cause carcinoma of the bladder, breast, and intestine in rodents [60, 72, 147, 148]. In addition, the norsesquiterpene glucoside ptaquiloside has been identified as a mutation-inducing, chromosome disrupting and cancercausing component of bracken [121, 167]. Upon exposure to alkaline conditions, ptaquiloside decomposes into an intermediate dienone known as 'activated ptaquiloside. This dienone is decomposed in weakly acidic conditions, leading to the production of pterosin B, which can additionally be synthesized directly from ptaquiloside at acidic pH values [148]. By utilizing its cyclopropyl ring, the unstable dienone forms covalently bonded DNA adducts, mostly at adenine's N-3 [132]. Initially alkylating adenine with ptaquiloside or its aglycon ptaquilosin in codon 61 of H-ras accompanied by depurination and faulty DNA synthesis results in the triggering of the H-ras proto-oncogene [133].

4.11 Future prospects

It is estimated that there are 13,000 species belonging to nearly 48 plant families in Pteridophytes; however, few species have been studied and 30 families have been studied. As a result, we strongly encourage pteridophyte researchers to explore the therapeutic potential of the plants and their bioactive components. With the help of biotechnology, researchers could increase the effectiveness of biologically active substances by studying in vitro culture, biochemical processes, mechanism of action, and genetic modification. Multidrug therapies should be developed from early tracheophytes and pteridophytes, as well as studying their biological processes, drug kinetics, and toxicity evaluations. A secondary metabolite analysis could help in identifying the genetic relationships among pteridophytes. Extraction of phytochemicals

from B. orientale, C. thalictroides, D. heterophyllum, D. linearis, H. arifolia, L. ensifolia, N. multiflora, P. calomelanos, P. confusa and the rhizomes and leaves of D. quercifolia were found to contain sugars, steroids, flavonoids, tannins, saponins, carboxylic acid, xanthoprotein, and phenols. Research on the antimicrobial properties of B. orientale, D. linearis, H. arifolia and roots of D. quercifolia suggests they contain one or more of these phytoconstituents. Numerous phytochemicals discovered have been documented to have medicinal and industrial benefits. There is huge therapeutic potential in plants that contain antimicrobial compounds because they are free of side effects common to synthesized antimicrobials. After extensive investigation, it appears that plants can be used as antimicrobial bioactive agents with broadspectrum activity. In future work, the objective will be to identify and characterize the underlying mechanisms for bioefficacy and bioactivity.

5 Conclusion

Phytochemical and pharmacological characteristics of pteridophytes by Indian minority groups are poorly understood as are their medicinal uses. Research on pteridophytes has grown over time in recent decades, and more reports have been published on ethnobotany of pteridophytes. The objective of this review is to provide baseline data for scientific research on medicinally important pteridophytes and to categorize them. Pteridophytes haven't been studied extensively in several mountainous regions, particularly the central Western Ghats and the seven sister states. Pteridophyte species are found in large numbers in the North East Indian states, but ethnomedicinal uses are in short supply. The lack of knowledge about pteridophyte species by people residing in these regions may be due to the ecological conditions in these states. Unless these states' underexplored and unexplored territories are explored, researchers must document the ethnomedicinal uses of pteridophytes. It should be given top priority to preserve traditional knowledge records to ensure that resources are conserved and the knowledge of these practices is protected before they disappear from future generations. Pteridophytes are widely used in India, but there are few publications about their therapeutic effects. The scientific literature has not explored much about a neglected group of pteridophytes, so further studies are needed to focus on them. In order to experimentally validate prevailing traditional knowledge as a solution to healthcare problems, the existence of the scientific steps discussed in this review needs to be taken into account. Based on the conclusions of this review, monographs and recommendations about Indian ethnomedicinally important pteridophytes could be formulated based on the data presented here. Pteridophytes are a group of plants with medicinal applications that by studying their phytochemical and pharmacological properties may reveal the presence of active compounds that can be developed into novel therapeutics. Pteridophytes are poorly understood in terms of phytochemistry, pharmacology, and pharmacognosy. To develop new drugs from underexplored plant groups, it is essential to encourage intensive scientific research on unexplored plant species to contribute to the on-going search for alternative and affordable treatments in India. For the purposes of conducting preclinical and clinical trials, a thorough investigation of toxicology is necessary. The study suggests that ethnobotanical investigations are crucial among indigenous communities, as well as the need for detailed studies of available cryptogams, potentially leading to the development of new and novel pharmaceuticals. Therefore, if ethnobotanical research is combined with biochemical analysis or physiological investigations, useful drugs could be discovered.

Abbreviations

AChEl: Acetylcholinesterase inhibitors; PAL: Phenylalanine ammonia lyase; TPC: Total phenolic content.

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Authors' contributions

AB: original draft; AD: Conceptualization, guidance. Both authors read and approved the final manuscript.

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Consent for publication

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References

- Aguilera P, Carrera C, Puig-Butille JA (2012) Benefits of oral Polypodium leucotomos extraction MM high-risk patients. J Eur Acad Detamatol Venereol 27:1095–1100. https://doi.org/10.1111/j.1468-3083.2012. 04659.x
- 2. Amaral S, Mira L, Nogueira JM et al (2009) Plant extracts with antiinflammatory properties—a new approach for characterization of their bioactive compounds and establishment of structure—antioxidant

- activity relationships. Bioorg Med Chem 17(5):1876–1883. https://doi.org/10.1016/j.bmc.2009.01.045
- Anand U, Tudu CK, Nandy S, Sunita K, Tripathi V, Loake GJ, Dey A, Proćków J (2022) Ethnodermatological use of medicinal plants in India: From ayurvedic formulations to clinical perspectives—a review. J Ethnopharmacol 10(284):114744
- Angeloni C, Vauzour D (2019) Natural products and neuroprotection. Int J Mol Sci 20:5570
- Bai R, Zhou Y, Deng S, Dong P-P, Zhang B, Huang S, Wang C, Zhang H-L, Zhao Y-Y, Wang L, Ma X (2013) Two new ent-kaurane diterpenoids from Pteris semipinnata. J Asian Nat Prod Res 15:1107–1111
- Bandopadhyay S, Anand U, Gadekar VS, Jha NK, Gupta PK, Behl T, Kumar M, Shekhawat MS, Dey A (2022) Dioscin: a review on pharmacological properties and therapeutic values. BioFactors 48(1):22–55
- 7. Banerjee RD, Sen SP (1980) Antibiotic activity of pteridophytes. Ecol Bot 34(3):284–298
- Banerjee S, Anand U, Ghosh S, Ray D, Ray P, Nandy S, Deshmukh GD, Tripathi V, Dey A (2021) Bacosides from *Bacopa monnieri* extract: an overview of the effects on neurological disorders. Phytother Res 35(10):5668–5679
- Baruah NC, Sarma JC, Barua NC et al (1994) Germination and growth inhibitory sesquiterpene lactones and a flavornes from *Tithonia* diversifolia. Phytochemitry 36(1):29–36. https://doi.org/10.1016/S0031-9422(00)97006-7
- Baskaran X, Jeyachandran R (2010) Evaluation of antioxidant and phytochemical analysis of *Pteris tripartita* Sw. a critically endangered fern from South India. J Fairy Lake Bot Gard 9(3):28–34
- Batiha GE, Hussein DE, Algammal AM, George TT, Jeandet P, Al-Snafi AE, Tiwari A, Pagnossa JP, Lima CM, Thorat ND, Zahoor M (2021) Application of natural antimicrobials in food preservation: recent views. Food Control 1(126):108066
- 12. Benjamin A, Manickam VS (2007) Medicinal pteridophytes from the Western Ghats. Ind J Tradit Knowl 6(4):611–618
- Berahou AA, Auhmani A, Fdil N et al (2007) Antibacterial activity of Quercus ilex bark's extracts. J Ethnopharmacol 112(3):426–429. https://doi.org/10.1016/j.jep.2007.03.032
- Bishayee K, Chakraborty D, Ghosh S, Boujedaini N, Khuda-Bukhsh AR (2013) Lycopodine triggers apoptosis by modulating 5-lipoxygenase, and depolarizing mitochondrial membrane potential in androgen sensitive and refractory prostate cancer cells without modulating p53 activity: signaling cascade and drug-DNA interaction. Eur J Pharmacol 698:110–121
- Bisht R, Joshi BC, Kalia AN, Prakash A (2017) Antioxidant-rich fraction of urtica dioica mediated rescue of striatal mito-oxidative damage in MPTP-induced behavioral, cellular, and neurochemical alterations in rats. Mol Neurobiol 54:5632–5645
- Biswas D, Mandal S, Chatterjee Saha S, Tudu CK, Nandy S, Batiha GE, Shekhawat MS, Pandey DK, Dey A (2021) Ethnobotany, phytochemistry, pharmacology, and toxicity of *Centella asiatica* (L.) Urban: a comprehensive review. Phytother Res 35(12):6624–6654
- 17. Braak H, Del Tredici K (2015) The preclinical phase of the pathological process underlying sporadic alzheimer's disease. Brain 138:2814–2833
- Caceres A, Gupta M, Ocampo R, Mendoza J, Herrera M, Solis P, Cruz S, Martinez A, José V (2006) Multidisciplinary development of phytotherapeutic products from native Central American plants. Acta Hortic 720:149–155. https://doi.org/10.17660/ActaHortic.2006.720.14
- Cai Y, Luo Q, Sun M et al (2004) Antioxidant activity and phenolic compounds of 112 traditional Chinese medicinal plants associated with anti-cancer. Life Sci 74(17):2157–2184. https://doi.org/10.1016/j.lfs.2003. 09.047
- 20. Caius JF (1935) The medicinal and poisonous ferns of India. J Bombay Nat. I Hist Soc 38:341–361
- 21. Callow RS, Cook LM (1999) Genetic and evolutionary diversity: the sport of nature. S. Thornes: Cheltenham; 8. ISBN 0-7487-4336-4337.
- 22. Cambie RC (1988) A New Zealand phytochemical register. J R Soc N Z 18:137–184
- Cao H, Chai T-T, Wang X, Morais-Braga MFB, Yang J-H, Wong F-C, Wang R, Yao H, Cao J, Cornara L, Burlando B, Wang Y, Xiao J, Coutinho HDM (2017) Phytochemicals from fern species: potential for medicine applications. Phytochem Rev 16:1–62

- 24. Cao J, Xia X, Chen X, Xiao J, Wang Q (2013) Characterization of flavonoids from Dryopteris erythrosora and evaluation of their antioxidant, anticancer and acetylcholinesterase inhibition activities. Food Chem Toxicol 51:242–250
- 25. Cao J, Xia X, Chen X, Xiao J, Wang Q (2012) Flavonoids contents, antioxidant and anticancer activities of 72 species of ferns from China. Biomed Pap Med Fac Univ Palacky Olomouc Czech Repub 156(Supp 1):S33
- Chandran G, Leelabai S, Srinivas Bharath M, Rajini P, Muralidhara M
 (2014) Understanding the role of neuronal thioredoxin and glutathione
 systems in motor disorder pathophysiology: relevance to natural product therapy. Mov Disord 29:S307
- Chandran G, Muralidhara (2013) Neuroprotective effect of aqueous extract of Selaginella delicatula as evidenced by abrogation of rotenone-induced motor deficits, oxidative dysfunctions, and neurotoxicity in mice. Cell Mol Neurobiol 33:929–942
- Chandran G, Muralidhara (2014) Insights on the neuromodulatory propensity of Selaginella (Sanjeevani) and its potential pharmacological applications. CNS Neurol Disord Drug Targets 13:82–95
- Chandran G, Venkareddy L, Muralidhara (2015) Biflavonoids: therapeutic potential of novel flavonoid dimers. In: Govil JN, Pathak M (eds) Recent progress in medicinal plants Volume 40 - Flavonoids and antioxidants. Studium Press LLC, USA. pp 316–330
- Chang HC, Agrawal DC, Kuo CL et al (2007) In vitro culture of *Drynaria fortunei*, a fern species source of Chinese medicine "Gu-Sui-Bu."
 In vitro Cell Dev Biol Plant 43(2):133–139. https://doi.org/10.1007/ s11627-007-9037-6
- Chang HC, Gupta SK, Tasay HS (2011) Studies on folk medicinal fern: an example of "Gu Sui-Bu." In: Fernandez H, Kumar A, Revilla MA (eds) Working with ferns, issues and applications. Springer, New York Dordrecht Heidelberg, London, pp 285–304
- 32. Chang HC, Huang GJ, Agrawal DC et al (2007) Antioxidant activities and polyphenol contents of six folk medicinal ferns used as "Gusuibu." Bot Stud 48:397–406
- 33. Chen IN, Chang CC, Wang CY et al (2008) Antioxidant and antimicrobial activity of Zingiberaceae plants in Taiwan. Plant Foods Hum Nutr 63(1):15–20. https://doi.org/10.1007/s11130-007-0063-7
- 34. Chen JJ, Duh CY, Chen JF (2005) New cytotoxic bioflavonoids from Selaginella delicatula. Plant Med 71(7):659–665. https://doi.org/10. 1055/s-2005-871273
- 35. Chen K, Plumb GW, Bennett RN et al (2005) Antioxidant activities of extracts from five anti-viral medicinal plants. J Ethnopharmacol 96(1–2):201–205. https://doi.org/10.1016/j.jep.2004.09.020
- Chen YH, Chang FR, Lin YJ et al (2007) Identification of phenolic antioxidants from Sword Brake fern (*Pteris ensiformis* Burm.). Food Chem 105:48–56. https://doi.org/10.1016/j.foodchem.2007.03.055
- Chowdhary S, Verma DL, Pande R et al (2010) Antioxidative properties of flavonoids from Cheilanthes anceps Swartz. J Am Sci 6(5):203–207
- Chowdhary NP (1973) The pteridophytic flora of the upper gangetic plain. Navyug Traders, New Delhi
- ChPC (Pharmacopoeia Commission of People's Republic of China),
 (2005) Pharmacopoeia of People's Republic of China (ChP). Chemical Industry Press, Beijing, China, pp 49-50
- 40. Ciulla M, Marinelli L, Cacciatore I, Stefano AD (2019) Role of dietary supplements in the management of Parkinson's disease. Biomolecules 9:271
- 41. Cragg GM, Newman DJ (2005) Plants as a source of anti-cancer agents. J Ethnopharmacol 100:72–79
- 42. Cui CB, Tezuka Y, Kikuchi T et al (1990) Constituents of fern, *Davallia mariesii* Moore. I. Isolation and structures of Davallialactone and a new flavanone glucuronide. Chem Pharm Bull 38:3218–3225. https://doi.org/10.1248/cpb.38.3218
- Strack D (1997) Phenolic metabolism. In: Dev PM, Harborne JB (eds) Plant biochemistry. Academic Press, London, UK, pp 387–416
- 44. Defilpps RA, Maina SL, Pray LA (1988) The Palauan and Yap Medicinal Plant Studies of Masayoshi okabe. The National Museum of Natural History Smithsonian Institution, Washington DC, USA, pp 1941–1943.
- Del Pino GJ, de SambricioGuiu F, Colomo GC (1982) Comparison of Polypodium leucotomos extract with placebo in 37 cases of psoriaris. Med Cut Ibero-Lat Amer 10:203–208

- Deshpande P, Gogia N, Singh A (2019) Exploring the efficacy of natural products in alleviating Alzheimer's disease. Neural Regen Res 14:1321–1329
- Dey A, De JN (2012) Ethnomedicinal plants used by the tribals of Purulia district, West Bengal, India against gastrointestinal disorders. J Ethnopharmacol 143(1):68–80
- Dey A, Gorai P, Mukherjee A, Dhan R, Modak BK (2017) Ethnobiological treatments of neurological conditions in the Chota Nagpur Plateau, India. J Ethnopharmacol 23(198):33–44
- Dey A, Nandy S, Mukherjee A, Modak BK (2021) Sustainable utilization of medicinal plants and conservation strategies practiced by the aboriginals of Purulia district, India: a case study on therapeutics used against some tropical otorhinolaryngologic and ophthalmic disorders. Environ Dev Sustain 23(4):5576–5613
- 50. Dhiman AK (1998) Ethnomedicinal uses of some pteridophyitc species in India. Ind Fern J 15(1–2):61–64
- 51. Dhiman AK (1998) Ethnomedicinal uses of some Pteridophytic species in India. Indian Fern J 15(1–2):61–64
- Di Paolo M, Papi L, Gori F, Turillazzi E (2019) Natural products in neurodegenerative diseases: a great promise but an ethical challenge. Int J Mol Sci 20:5170
- 53. Dixit RD (1989) Ecology and taxonomy of pteridophytes of Madhya Pradesh. Indian Fern J 6:140–159
- 54. Dixit RD, Vohra JN (1984) A dictionary of the pteridophytes of India. Botanical Survey of India, Howrah
- Dong LB, Yang J, He J et al (2012) Lycopalhine A, a novel sterically congested Lycopodium alkaloid with an unprecedented skeleton from Palhinhaea cernua. Chem Commun (Camb) 48:9038–9040
- Dos Santos Jr J, Blanco MM, Do Monte FHM, Russi M, Lanziotti VMNB, Leal LKAM, Cunha GM (2005) Sedative and anticonvulsant effects of hydroalcoholic extract of Equisetum arvense. Fitoterapia 76:508–513
- Durrenberger PF, Fernando FS, Kashefi SN, Bonnert TP, Seilhean D, Nait-Oumesmar B, Schmitt A, Gebicke-Haerter PJ, Falkai P, Grunblatt E et al (2015) Common mechanisms in neurodegeneration and neuroinflammation: a brainnet europe gene expression microarray study. J Neural Transm 122:1055–1068
- Dutta T, Nandy S, Dey A (2022) Urban ethnobotany of Kolkata, India: a case study of sustainability, conservation and pluricultural use of medicinal plants in traditional herbal shops. Environ Dev Sustain 24(1):1207–1240
- Engi H, Hohmann J, Gang G et al (2008) Chemoprevention and inhibition of P-glycoprotein in cancer cells by Chinese medicinal herbs. Phytother Res 22:1671–1676
- 60. Freitas RN, Brasileiro-Filho G, Silva ME, Pena SD (2002) Bracken ferninduced malignant tumors in rats: absence of mutations in p53, H-ras and K-ras and no microsatellite instability. Mutat Res 499:189–196
- Garcia F, Pivel JP, Guerrero A et al (2006) Phenolic components and antioxidant activity of Fernblock, an aqueous extract of the aerial parts of the fern Polypodium leucotomos. Methods Find Exp Clin Pharmacol 28(3):157–160. https://doi.org/10.1358/mf.2006.28.3.985227
- 62. Gaur RD, Bhatt BP (1994) Folk utilization of some Pteridophytes of deoprayag area in Garhwal Himalaya: India. Econ Bot 48(2):146–151
- Gayathri V, Asha V, Subromaniam A (2005) Preliminary studies on the immunomodulatory and antioxidant properties of Selaginella species. Ind J Pharmacol 37(6):381–385. https://doi.org/10.4103/0253-7613. 19075
- Girish C, Muralidhara (2012) Propensity of Selaginella delicatula aqueous extract to offset rotenone-induced oxidative dysfunctions and neurotoxicity in Drosophila melanogaster: implications for Parkinson's disease. Neurotoxicology 33:444–456
- Gombau L, Garcia F, Lahoz A et al (2006) Polypodium leucotomos extract: antioxidant activity and disposition. Toxicol Vitro 20(4):464–471. https://doi.org/10.1016/j.tiv.2005.09.008
- Gottlieb OR, Kaplan MAC, Zocher DHT, Kubitzki K (1990) A chemosystematic overview of pteridophyta and gymnosperms. In: Kubitzki K (ed)
 The families and genera of vascular plaNTs, vol 1. Springer-Verlag, Berlin, pp. 2–10
- 67. Govindappa M, Naga Sravya S, Poojashri MN et al (2011) Antimicrobial, antioxidant and in vitro anti-inflammatory activity of ethanol extract and active phytochemical screening of *Wedelia trilobata* (L.) Hitchc. J Pharmacogn Phytother 3(3):43–51

- Ha L, Thao D, Huong H, Minh C, Dat N (2012) Toxicity and anticancer effects of an extract from Selaginella tamariscina on a mice model. Nat Prod Res 26:1130–1134
- 69. Haddad F, Sawalha M, Khawaja Y, Najjar A, Karaman R (2018) Dopamine and levodopa prodrugs for the treatment of parkinson's disease. Molecules 23:40
- 70. Halliwell B (1996) Antioxidants in human health and disease. Ann Rev Nutr 16(1):33–50. https://doi.org/10.1146/annurev.nu.16.070196.000341
- Ham Y-M, Yoon W-J, Park S-Y, Jung Y-H, Kim D, Jeon Y-J, Wijesinghe JP, Kang S-M, Kim K-N (2012) Investigation of the component of *Lycopo-dium serratum* extract that inhibits proliferation and mediates apoptosis of human HL-60 leukemia cells. Food Chem Toxicol 50:2629–2634
- Hirono I, Ushimaru Y, Kato K, Mori H, Sasaoka I (1978) Carcinogenicity of boiling water extract of bracken, *Ptereidium aquilinum*. Gann 69:383–388
- Hoang L, Tran H (2014) In vitro antioxidant and anti-cancer properties of active compounds from methanolic extract of *Pteris multifida* Poir. Leaves Eur J Med Plants 4(3):292–302. https://doi.org/10.9734/EJMP/ 2014/7053
- 74. Hsu YL, Uen YH, Chen Y, Liang HL, Kuo PL (2009) Tricetin, a dietary flavonoid, inhibits proliferation of human breast adenocarcinoma mcf-7 cells by blocking cell cycle progression and inducing apoptosis. J Agric Food Chem 57:8688–8695
- Huang YL, Yeh PY, Shen CC, Chen CC (2003) Antioxidant flavonoids from the rhizomes of *Helminthostachys zeylanica*. Phytochemistry 64:1277–1283
- Jermy AC (1986) Subgeneric names in Selaginella. Fern Gazette 13:117–118
- 77. Jiang J, Liu Y, Min K, Jing B, Wang L, Zhang Y, Chen Y (2010) Two new Lycopodine alkaloids from *Huperzia serrata*. Helv Chim Acta 93:1187–1191
- 78. Jiménez D, Naranjo R, Doblaré E et al (1987) Anapsos, an antipsoriatic drug in atopic dermatitis. Allergol Immunopathol 15:185–189
- Karthik V, Raju K, Ayyanar M, Gowrishankar K, Sekar T (2011) Ethnomedicinal uses of pteridophytes in kolli hills, Eastern Ghats of Tamil Nadu, India. J Nat Prod Plant Resour 1(2):50–55
- Kaur P, Gupta RC, Dey A, Malik T, Pandey DK (2020) Optimization of salicylic acid and chitosan treatment for bitter secoiridoid and xanthone glycosides production in shoot cultures of Swertia paniculata using response surface methodology and artificial neural network. BMC Plant Biol 20(1):1–3
- 81. Kaushik S, Cuervo AM (2015) Proteostasis and aging. Nat Med 21:1406–1415
- 82. Kimura K, Noro Y (1965) Pharmacognostical studies on Chinese drug "Gu-Sui-Bu". I. Consideration on "Gu-Sui-Bu" in old herbals (pharmacognostical studies on fern drugs XI). Syoy akugaku Zasshi 19:25–31
- 83. Kirtikar KR, Basu BD, An ICS (1935) Indian medicinal plants, Part IV, 2nd edn. Lalit Mohan Basu, Allahabad, India
- Koh EJ, Seo YJ, Choi J, Lee HY, Kang DH, Kim KJ, Lee BY (2017) Spirulina maxima extract prevents neurotoxicity via promoting activation of BDNF/CREB signaling pathways in neuronal cells and mice. Molecules 22:1363
- Korall P, Kenrick P (2002) Phylogenetic relationships in Selaginellaceae based on RBCL sequences. Am J Bot 89:506–517
- 86. Kosuge T, Yokota M, Sugiyama K, Yamamoto T, Ni MY, Yan SC (1985) Studies on antitumor activities and antitumor principles of Chinese herbs. I. Antitumor activities of Chinese herbs. Yakugaku Zasshi 105:791–795 ([Article in Japanese])
- 87. Kshirsagar R, Upadhyay S (2009) Free radical scavenging activity screening of medicinal plants from Tripura, Northeast India. Nat Prod Radiance 8(2):117–122
- Kumar M, Ramesh M, Sequiera S (2003) Medicinal pteridophytes of Kerala, South India. Indian Fern J 21:1–28
- Lai HY, Lim YY, Kim KH (2010) Blechnum orientale Linn—a fern with potential as antioxidant, anti-cancer and antibacterial agent. BMC Complement Altern Med 10(15):1–8
- Lai H, Lim Y (2011) Evaluation of antioxidant activities of the methanolic extracts of selected ferns in Malaysia. Int J Environ Sci Dev 2(6):442
- Lan J, Liu Z, Liao C, Merkler DJ, Han Q, Li J (2019) A study for therapeutic treatment against parkinson's disease via Chou's 5-steps rule. Curr Top Med Chem 19:2318–2333

- Lee C-W, Choi H-J, Kim H-S, Kim D-H, Chang I-S, Moon HT, Lee S-Y, Oh WK, Woo E-R (2008) Biflavonoids isolated from Selaginella tamariscina regulate the expression of matrix metalloproteinase in human skin fibroblasts. Bioorg Med Chem 16:732–738
- Lee H, Lin JY (1988) Antimutagenic activity of extracts from anti-cancer drugs in Chinese medicine. Mutat Res 204(2):229–234. https://doi.org/ 10.1016/0165-1218(88)90093-6
- 94. Lee IS, Nishikawa A, Furukawa F, Kasahara K, Kim SU (1999) Effects of Selaginella tamariscina on in vitro tumor cell growth, p53 expression, G1 arrest and in vivo gastric cell proliferation. Cancer Lett 144:93–99
- 95. Liang YC, Huang YT, Tsau SH et al (1999) Suppression of inducible cyclo oxygenase and inducible nitric oxide synthase by apigenia and related flavonoid in mouse macrophages. Carcinogenesis 20(10):1945–1952. https://doi.org/10.1093/carcin/20.10.1945
- Lim H, Kim HP (2007) Inhibition of mammalian collagenase, matrix metalloproteinase-1, by naturally-occurring flavonoids. Planta Med 73:1267–1274
- 97. Lin Y, Shi R, Wang X et al (2008) Luteolin, a flavonoid with potential for cancer prevention and therapy. Curr Can Drug Targets 8(7):634–646. https://doi.org/10.2174/156800908786241050
- 98. Liu Q, Qin M (2002) Studies on chemical constituents of rhizomes of Pteris multifidi Poir. Chin Tradit Herb Drugs 33(2):114
- Lopez-Lazaro M (2009) Distribution and biological activities of the flavonoid luteolin. Mini-Rev Med Chem 9(1):31–59. https://doi.org/10.2174/ 138955709787001712
- Lu H, Xu J, Zhang LX et al (1999) Bioactive constituents from *Pteris multifida*. Plant Med 65(6):586–587. https://doi.org/10.1055/s-2006-960835
- Ma T, Gong K, Yan Y, Zhang L, Tang P, Zhang X, Gong Y (2013) Huperzine a promotes hippocampal neurogenesis in vitro and in vivo. Brain Res 1506:35–43
- Ma X, Gang DR (2004) The Lycopodium alkaloids. Nat Prod Rep 21:752–772
- Ma X, Tan C, Zhu D, Gang DR, Xiao P (2007) Huperzine a from Huperzia species—an ethnopharmacological review. J Ethnopharmacol 113:15–34
- Mallikharjuna PB, Rajanna LN, Seetharam YN, Sharanabasappa GK (2007)
 Phytochemical studies of Strychnos potatorum L.f.-A medicinal plant. E-J Chem 4(4):510–518
- Manandhar PN (1996) Ethnobotanical observations on ferns and fern allies of Nepal. J Econ Taxon Bot 12:414–422
- Mandal SK, Biswas R, Bhattacharyya SS et al (2010) Lycopodine from Lycopodium clavatum extract inhibits proliferation of HeLa cells through induction of apoptosis via caspase-3 activation. Eur J Pharmacol 626:115–122
- Markham KR, Moore NA (1980) Comparative flavonoid glycoside biochemistry as a chemotaxonomic tool in the subdivision of the classical "genus" Lycopodium. BiochemSyst Ecol 8:17–20
- 108. Masuda EK, Kommers GD, Martins TB, Barros CS, Piazer JV (2011) Morphological factors as indicators of malignancy of squamous cell carcinomas in cattle exposed naturally to bracken fern (*Pteridium aquilinum*). J Comp Pathol 144:48–54
- Mazzio EA, Soliman KF (2009) In vitro screening for the tumoricidal properties of international medicinal herbs. Phytother Res 23:385–398
- Middelkamp-Hup MA, Bos JD, Rius-Diaz F et al (2007) Treatment of vitiligo vulgaris with narrow-band UVB and oral *Polypodium leucotomos* extract: a randomized double-blind placebocontrolled study. J Eur Acad Dermatol Venereol 21:942–950. https://doi.org/10.1111/j.1468-3083.2006.02132.x
- Middlekamp-Hup MA, Pathak MA, Parrado C et al (2004) Orally administered *Polypodium leocutomos* extract decreases psoralen-UVA-induced phototoxicity, pigmentation, and damage of human skin. J Am Acad Dermatol 50:41–49. https://doi.org/10.1016/S0190-9622(03)02732-4
- Milan CM, Avijit D, Abdur R et al (2013) Evaluation of antioxidant, cytotoxic and antimicrobial properties of *Drynaria quercifolia*. Int Res J Pharm 4(7):46–48. https://doi.org/10.7897/2230-8407.04710
- 113. Milovanovic V, Radulovic N, Todorovic Z et al (2007) Antioxidant, antimicrobial and genotoxicity screening of hydro-alcoholic extracts of five Serbian *Equisetum* species. Plant Foods Hum Nutr 62(3):113–119. https://doi.org/10.1007/s11130-007-0050-z
- 114. Modak BK, Gorai P, Dhan R, Mukherjee A, Dey A (2015) Tradition in treating taboo: Folkloric medicinal wisdom of the aboriginals of Purulia

- district, West Bengal, India against sexual, gynaecological and related disorders. J Ethnopharmacol 1(169):370–386
- Moon HE, Paek SH (2015) Mitochondrial dysfunction in Parkinson's disease. Exp Neurobiol 24:103
- Morais-Braga MFB, Souza TM, Santos KKA, Guedes GMM, Andrade JC, Tintino SR, Sobral-Souza CE, Costa JGM, Saraiva AAF, Coutinho HDM (2012) Phenolic compounds and interaction between aminoglycosides and natural products of *Lygodium venustum* SW against multiresistant bacteria. Chemotherapy 58:337–340
- Murakami T, Machashi NT (1985) Chemical and chemotaxonomical studies on filices. J Pharm Soc Japan 105(7):640–648. https://doi.org/10. 1248/yakushi1947.105.7 640
- 118. Nagaprashantha LD, Vatsyayan R, Singhal J et al (2011) Anticancer effects of novel flavonoid vicenin-2 as a single agent and in synergistic combination with docetaxel in prostate cancer. Biochem Pharmacol 82:1100–1109
- Newman DJ, Cragg GM, Holbeck S, Sausville EA (2002) Natural products as leads to cell cycle pathway targets in cancer chemotherapy. Curr Cancer Drug Targets 2:279–308
- Newman DJ, Cragg GM, Snader KM (2000) The influence of natural products upon drug discovery. Nat Prod Rep 17(2):175–191. https://doi. org/10.1039/a809402k
- Niwa H, Ojika M, Wakamatsu K, Yamada K, Hirono I, Matsushita K
 (1983) Ptaquiloside, a novel norsesquiterpene glucoside from bracken
 (Pteridium aquilinum var. latiusculum). Tetrahedron Lett 24:4117–4120
- 122. Nwosu MO (2002) Ethnobotanical studies on some pteridophytes of Southern Nigeria. Econ Bot 56:255–259
- 123. Okwu DE (2005) Phytochemicals, vitamins and mineral contents of two Nigeria medicinal plants. Int J Mol Med Adv Sci 1(4):375–381
- 124. Padala S (1988) Ethnobotanical euphony in some pteridophytes. In: Kaushik P (ed) Indigenous medicinal plants including microbes and fungi. Today and tomorrow's printers and publishers, New Delhi, pp 67–69
- Padilla HC, Laínez H, Pacheco JA (1974) A new agent (hydrophilic fraction of Polypodium leucotomos) for management of psoriasis. Int J Dermatol 13:276–282
- 126. Pandey DK, Kaur P, Kumar V, Banik RM, Malik T, Dey A (2021) Screening the elite chemotypes of *Gloriosa superba* L. in India for the production of anticancer colchicine: simultaneous microwave-assisted extraction and HPTLC studies. BMC Plant Biol 21(1):1–8
- 127. Parihar P, Parihar L (2006) Some pteridophytes of medicinal importance from Rajasthan. Nat Prod Radiance 5(4):293–301
- 128. Parihar P, Parihar L, Bohra A (2010) In vitro antibacterial activity of fronds (leaves) of some important pteridophytes. J Microbiol Antimicrob
- Park JS, Davis RL, Sue CM (2018) Mitochondrial dysfunction in Parkinson's disease: new mechanistic insights and therapeutic perspectives. Curr Neurol Neurosci Rep 18:21
- Petard P, Raau T (1972) The Use of Polynesia Medicinal Plants in Tahitian Medicine. Technical Paper No. 167. South Pacific Commission. Noumea, New Caledonia.
- Pichi-Sermolli REG (1977) Tentamen pteridophytorum genera in taxonomicum ordinem redigendi. Webbia 31:313–512
- 132. Potter DM, Baird MS (2000) Carcinogenic effects of ptaquiloside in bracken fern and related compounds. Br J Cancer 83:914–920
- Prakash AS, Pereira TN, Smith BL, Shaw G, Seawright AA (1996) Mechanism of bracken fern carcinogenesis: evidence for H-ras activation via initial adenine alkylation by ptaquiloside. Nat Toxins 4(5):221–227
- 134. Prakash S, Kumar M, Kumari N, Thakur M, Rathour S, Pundir A, Sharma AK, Bangar SP, Dhumal S, Singh S, Thiyagarajan A (2021) Plant-based antioxidant extracts and compounds in the management of oral cancer. Antioxidants 10(9):1358
- Proestos C, Chorianopoulos N, Nychas GJ et al (2005) RP-HPLC analysis
 of the phenolic compounds of plant extracts. Investigation of their
 antioxidant capacity and antimicrobial activity. J Agric Food Chem
 53(4):1190–1195. https://doi.org/10.1021/jf040083t
- Punzon C, Alcaide A, Fresno M (2003) In vitro anti-inflammatory activity of *Phlebodium decumanum*. Modulation of tumor necrosis factor and soluble TNF receptors. Int Immonopharmacol 3(9):1293–1299. https:// doi.org/10.1016/S1567-5769(03)00117-6

- 137. Qin B, Zhu D, Jiang S et al (2006) Chemical constituents of Pteris multifida and their inhibitory effects on growth of rat prostatic epithelial cells in vitro. Chin J Nat Med 4(6):428–431
- 138. Dixon RA, Paiva NL (1995) Stress-induced phenylpropanoid metabolism. Plant Cell 7:1085–1097
- 139. Rafii MS, Walsh S, Little JT, Behan K, Reynolds B, Ward C, Jin S, Thomas R, Aisen PS, Alzheimer's Disease Cooperative Study (2011) A phase II trial of huperzine a in mild to moderate Alzheimer disease. Neurology 76:1380–1394
- 140. Reddy VL, Ravikanth V, Rao TP et al (2001) A new triterpenoid from the fern Adiantum lunulatum and evaluation of antibacterial activity. Phytochemistry 56(2):173–175. https://doi.org/10.1016/S0031-9422(00) 00334-4
- Reinaldo RC, Santiago ACP, Medeiros PM, Albuquerque UP (2015) Do ferns and lycophytes function as medicinal plants? a study of their low representation in traditional pharmacopoeias. J Ethnopharmacol 175:39–47
- Ripa FA, Nahar L, Haque M et al (2009) Antibacterial, cytotoxic and antioxidant activity of crude extract of Marsilea quadrifolia. Eur J Sci Res 33(1):123–129
- Rodríguez-Yanes E, Juarranz A, Cuevas J et al (2012) Polypodium leucotomos decreases UV-induced epidermal cell proliferation and enhances p53 expression and plasma antioxidant capacity in hairless mice. Exp Dermatol 21:630–642. https://doi.org/10.1111/exd.12454
- Rout SD, Panda T, Mishra N (2009) Ethnomedicinal studies on some pteridophytes of Similipal Biosphere Reserve, Orissa, India. Int J Med Med Sc 1(5):192–197
- Chanishvili S, Badridze G, Janukashvili N (2007) Effect of altitude on the contents of antioxidants in leaves of some herbaceous plants. Russ J Ecol 38:367–373
- Santhosh KS, Samydurai P, Nagarajan N (2014) Indigenous knowledge on some medicinal pteridophytic plant species among the Malasar tribe's in Valparai Hills, Western Ghats of Tamil Nadu. Am J Ethnomed 1(3):164–173
- Santos RC, Brasileiro-Filho G, Hojo ES (1987) Induction of tumors in rats by bracken fern (Pteridium aquilinum) from Ouro Preto (Minas Gerais, Brazil). Braz J Med Biol Res 20:73–77
- 148. Shahin M, Smith BL, Worral S, Moore MR, Seawright AA, Prakash AS (1998) Bracken fern carcinogenesis: multiple intravenous doses of activated ptaquiloside induce DNA adducts, monocytosis, increased TNF alpha levels, and mammary gland carcinoma in rats. Biochem Biophys Res Commun 244:192–197
- Shaikh SD, Masal VP, Shaikh AS (2014) Ethnomedicinal uses of some pteridophytes from Konkan region of Maharashtra, India. Int J Pharm Chem Sci 3:2277–5005
- 150. Sharma BD, Vyas MS (1985) Ethnobotanical studies on ferns and fernallies of Rajasthan. Bull Bot Survey India 27(1–4):90–91
- Shih CH, Siu SO, Ng R et al (2007) Quantitative analysis of anticancer 3-deoxyanthocyanidins in infected sorghum seedlings. J Agric Food Chem 55:254–259
- 152. Shu JC, Liu JQ, Zhong YQ et al (2012) Two new pterosin sesquiterpenes from *Pteris multifida* Poir. Phytochem Lett 5(2):276–279
- Shyur LF, Tsung JH, Chen JH et al (2005) Antioxidant properties of extracts from medicinal plants popularly used in Taiwan. Int J Appl Sci Eng 3(3):195–202
- 154. Singh N, Kaur S, Bedi PMS, Kaur D (2011) Anxiolytic effects of Equisetum arvense Linn. extracts in mice. Indian J Exp Biol 49:352–356
- Singh BP, Upadhyay R (2010) Observations on some ferns of Pachmarhi Biosphere Reserve in traditional veterinary uses. Indian Fern J 27:94–100
- Singh BP, Upadhyay R (2012) Ethno-botanical importance of Pteridophytes used by the tribe of Pachmarhi, Central India. J Med Plants Res 6(1):14–18
- 157. Singh HB (1999) Potential medicinal pteridophytes of India and their chemical constituents. J Econ Taxon Bot 23(1):63–78
- Singh S, Dixit RD, Sahu TR (2005) Ethnobotanical use of Pteridophytes of Amarkantak (MP). Indian J Tradit Knowl 4(4):392–395
- Singh S, Dixit RD, Sahu TR (2003) Some medicinally important Pteridophytes of Central India. Int J For Usufruct Manag 4(2):41–51
- 160. Singh S, Dixit RD, Sahu TR (2007) Ethnomedicinal pteridophytes of Pachmarhi Biosphere Reserve, Madhya Pradesh. Indigenous knowledge: an application. Scientific Publisher, Jodhpur, pp 121–147

- Srivastava SK, Gautam RP, Singh SK, Rajkumar SD (2015) Ethnomedicinal uses of Pteridophytes of Tikri Forest, Gonda, Uttar Pradesh. Int J Pharma Bio Sci 6(3):88–94
- 162. Sun Z-K, Yang H-Q, Chen S-D (2013) Traditional Chinese medicine: a promising candidate for the treatment of Alzheimer's disease. Transl Neurodegener 2:6
- Tan WJ, Xu JC, Li L et al (2009) Bioactive compounds of inhibiting xanthine oxidase from Selaginella labordei. Nat Prod Res 23(4):393–398. https://doi.org/10.1080/14786410802228736
- Uddin MG, Mirza MM, Pasha MK (1998) The medicinal uses of pteridophytes of Bangladesh. Bangladesh J Plant Taxon 5(2):29–41
- Uma R, Pravin B (2013) In vitro cytotoxic activity of Marsilea quadrifolia Linn. of MCF-7 cells of human breast cancer. Int Res J Med Sci 1(1):10–13
- Upreti K, Jalal JS, Tewari LM, Joshi GC, Pangtey YPS, Tewari G (2009) Ethnomedicinal uses of Pteridophytes of Kumaun Himalaya, Uttarakhand. India J Am Sci 5(4):167–170
- van der Hoeven JC, Lagerweij WJ, Posthumus MA, van Veldhuizen A, Holterman HA (1983) Aquilide A, a new mutagenic compound isolated from bracken fern (*Pteridium aquilinum* (L.) Kuhn). Carcinogenesis 4:1587–1590
- Vargas J, García E, Gutiérrez F, Osorio C (1981) Síntesis de ácidos nucleicos y niveles de AMP cíclico en tumores murinos después del tratamiento in vitro con anapsos. Arch Fac Med Madrid 40:39–46
- Vasudeva SM (1999) Economic importance of pteridophytes. Ind Fern J 16(1–2):130–152
- 170. Vyas MS, Sharma BD (1988) Ethnobotanical importance of ferns of Rajasthan. Indigenous Medicinal Plants of Gurukul Kangri. Today and Tomorrow Printers and Publishers, New Delhi, India, pp 61–66
- 171. Wales P, Pinho R, Lázaro DF, Outeiro TF (2013) Limelight on alpha-synuclein: pathological and mechanistic implications in neurodegeneration. J Parkinsons Dis 3:415–459
- Wallace JW, Markham KR (1978) Apigenin and amentoflavone glycosides in the psilotaceae and their phylogenetic significance. Phytochemistry 17:1313–1317
- 173. Wallace JW, Yopp DL, Besson E, Chopin J (1981) Apigenin di-Cglycosylflavones of Angiopteris (Marattiales). Phytochemistry 20:2701–2703
- Wang H, Tang XC (1998) Anticholinesterase effects of huperzine A, E2020, and tacrine in rats. Zhongguo Yao Li Xue Bao 19:27–30
- Wang HB, Wong MH, Lan CY et al (2010) Effect of arsenic on flavonoid contents in Pteris species. Biochem Syst Ecol 38(4):529–537. https://doi. org/10.1016/j.bse.2010.05.009
- 176. Wang R, Yan H, Tang XC (2006) Progress in studies of huperzine A, a natural cholinesterase inhibitor from Chinese herbal medicine. Acta Pharmacol Sin 27(1):1–26
- 177. Wang TC, Lee HI, Yang CC (2009) Evaluation of in vitro antioxidant and antilipid peroxidation activities of Ching- Pien-Tsao (Pteris multifida Poiret). J Taiwan Agric Res 58(1):55–60
- 178. Wang Y-S, Li F-Y, Huang R, Li Y, Feng X-F, Yang J-H (2013) Chemical constituents of *Pteris multifida*. Chem Nat Compd 49:629–631
- 179. Warashina T, Umehara K, Miyase T (2012) Flavonoid glycosides from Botrychium ternatum. Chem Pharm Bull (Tokyo) 60:1561–1573
- 180. Wei HA, Lian TW, Tu YC et al (2007) Inhibition of lowdensity lipoprotein oxidation and oxidative burst in polymorphonuclear neutrophils by caffeic acid and hispidin derivatives isolated from sword brake fern (*Pteris ensiformis* Burm.). J Agric Food Chem 55(26):10579–10584. https://doi. org/10.1021/jf071173b
- Woo HJ, Oh IT, Lee JY et al (2012) Apigeninidin induces apoptosis through activation of Bak and Bax and subsequent mediation of mitochondrial damage in human promyelocytic leukemia HL-60 cells. Process Biochem 47:1861–1871
- Wu T-Y, Chen C-P, Chen C-P, Jinn T-R (2011) Traditional Chinese medicines and Alzheimer's disease. Taiwan J Obstet Gynecol 50:131–135
- 183. Xu Q, Mori H, Sakamoto O, Uesugi Y, Koda A (1989) Immunological mechanisms of antitumor activity of some kinds of crude drugs on tumor necrosis factor production. Int J Immunopharmacol 11:607–613
- Yang JH, Kondratyuk TP, Jermihov KC et al (2011) Bioactive compounds from the fern Lepisorus contortus. J Nat Prod 74:129–136
- Yang SH, Liao PH, Pan YF, Chen SL, Chou SS, Chou MY (2012) The novel p53-dependent metastatic and apoptotic pathway induced by Vitexin

- in human oral cancer OC2 cells, Phytother Res. https://doi.org/10.1002/ptr.4841 ([Epub ahead of print])
- Yang Y-F, Qu S-J, Xiao K, Jiang S-H, Tan J-J, Tan C-H, Zhu D-Y (2010)
 Lycopodium alkaloids from Huperzia serrata. J Asian Nat Prod Res 12:1005–1009
- Yao LQ, Ren H, Ai QL, Chang LH (2006) Efficacy of rehabilitation practice together with Huperzine A on the cognitive impairment in patients with Parkinson's disease. Foreign Med Sci Psychiatry 33(3):204–206
- Ying Y-M, Liu X-S, Tong C-P, Wang J-W, Zhan Z-J, Shan W-G (2014) Lycopodium Alkaloids from Huperzia serrata. Helv Chim Acta 97:1433–1439
- Yoshida T, Konishi M, Horinaka M et al (2008) Kaempferol sensitizes colon cancer cells to TRAIL-induced apoptosis. Biochem Biophys Res Commun 375(1):129–133. https://doi.org/10.1016/j.bbrc.2008.07.131
- Yu D, Thakor DK, Han I, Ropper AE, Haragopal H, Sidman RL, Zafonte R, Schachter SC, Teng YD (2013) Alleviation of chronic pain following rat spinal cord compression injury with multimodal actions of huperzine A. Proc Natl Acad Sci U S A 110:E746–E755
- Zakaria ZA, Mohamed AM, Jamil NSM et al (2011) In vitro cytotoxic and antioxidant properties of the aqueous, chloroform and methanol extracts of Dicranopteris linearis leaves. Afr J Biotechnol 10(2):273–282
- 192. Zattra E, Coleman C, Arad S et al (2009) Polypodium leucotomos extract decreases UV-induced Cox-2 expression and inflammation, enhances DNA repair, and decreases mutagenesis in hairless mice. Am J Pathol 175:1952–1961. https://doi.org/10.2353/ajpath.2009.090351
- 193. Zhang ZX, Wang XD, Chen QT, Shu L, Wang JZ, Shan GL (2002) Clinical efficacy and safety of huperzine A in treatment of mild to moderate Alzheimer disease, a placebo- controlled, double-blind, randomized trial. Natl Med J China 82(14):941–944
- 194. Zhao Z, Jin J, Ruan J et al (2007) Antioxidant flavonoid glycosides from aerial parts of the fern *Abacopteris penangiana*. J Nat Prod 70(10):1683–1686. https://doi.org/10.1021/np0703850

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