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The association between toll-like receptor 4 (TLR4) genotyping and the risk of epilepsy in children



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Abstract

Background: Epilepsy is one of the most widely recognized neurological disorders; unfortunately, twenty to thirty percent of patients do not get cured from epilepsy, despite many trials of antiepileptic drug (AED) therapy. Immunotherapy may be a viable treatment strategy in a subset of epileptic patients. The association between Tolllike receptor polymorphisms and epilepsy clarifies the role of the immune system in epilepsy and its response to the drug. Thus, this study will focus on the relation between TLR4 rs1927914, rs11536858, rs1927911SNPs, and epilepsy in an Egyptian case-control study to assess their link to antiepileptic drug response.

Results: According to TLR4 rs1927914, there is a significant association between the SNP and the development of epilepsy, as CC genotype is 15.3 times more at risk for developing epilepsy than TT genotype, and CT is 11.1 times more at risk for developing epilepsy than TT. Also, patients with CC genotypes are 6.3 times more at risk for developing primary epilepsy than TT genotype.

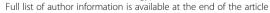
According to rs11536858, there is a significant association between cases and control groups, as AA genotypes are found to be more at risk for developing epilepsy than GG genotypes. Also, there is a statistically significant association between clonazepam resistance and rs11536858, as p value < 0.001* with the highest frequency of TT genotypes at 4.3%.

According to rs1927911, there are no significant results between the cases and the control groups or between drug-responsive and drug resistance.

Conclusion: Possible involvement of the Toll-like receptor clarifies the importance of innate immunity in initiating seizures and making neuronal hyperexcitability. In this work, multiple significant associations between TLR SNPs and epilepsy, epileptic phenotype, and drug-resistant epilepsy have been found. More studies with bigger sample sizes and different techniques with different SNPs are recommended to find the proper immunotherapy for epilepsy instead of the treatment by antiepileptic drugs.

Keywords: Antiepileptic drug-resistant, Egyptian, Innate immunity, PCR, Pediatric, RFLP

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Background

Epilepsy is one of the most widely recognized neurological disorders, and seizures are its main symptom with emotional and cognitive dysfunction, which affects ≈ 50 million people worldwide. Many experimental and clinical research studies showed the active role of the two arms of the immune system (innate and adaptive) in the pathogenesis of epilepsy via inflammation. Also, inflammatory cytokines and immune cells were reported to have a significant role in the initiation and relapse of epileptic seizures and the association of other diseases with epilepsy [1]. In the brain, TLR4 is expressed by astrocytes and microglia; the presence of TLR on glial cells and their limited expression on neurons have been evidenced by various studies. The expression of TLR mRNA on human microglial cells has been demonstrated with high cell surface and intracellular expression of TLR2 and TLR3, respectively [1].

The capacity of ligand binding with TLR4 may be changed due to alteration in the signals caused by single-nucleotide polymorphisms (SNPs) in the extracellular domain of TLR4 which will then alter the level of cytokines either pro- or anti-inflammatory cytokines and hence alter the susceptibility of chronic inflammation. Three SNPs of TLR4 gene, namely, rs11536858 (now merged into rs10759931), rs1927911, and rs1927914, are reported to be associated with inflammatory diseases [2]. The regions of rs1927911 (7764C>T), rs1927914 (2437T>C), and rs10759931 (2688A>G) are Intron 1, 5'UTR, and promoter respectively; their locations are Chromosome 9:117707776, Chromosome 9:117702447, and Chromosome 9:117701869 respectively, and their minor allele frequencies (MAF) are 0.40, 0.49, and 0.35 respectively. All data were collected from the Ensemble Genome Browser (www.ensembl.org). These SNPs rs1927911, rs1927914, and rs10759931 have not been previously studied in Egypt.

An experimental study on murine microglia using real-time PCR has shown an expression of TLRs 1–9 and a condensed expression of TLRs 2, 4, 6, 8, and 9 mRNA in response to IFN-g, as well as a reaction to lipopolysaccharide (LPS). During pathogen recognition, TLR4 receptors interacted with intracellular protein MyD88 after the stimulation of the IL-1b/IL-1b R axis.

HMGB1 is one of the damage-associated molecular patterns (DAMPs) that is passively released from necrotic cells and actively secreted by cells that are exposed to significant stress, which are recognized by TLR-signaling pathways. Also, an increase in the expression of TLR4 and HMGB1 in neurons has been observed in chemical epilepsy models [3].

HMGB1-TLR4 signaling has a significant role in the pathogenesis of epilepsy, which was confirmed by studies that were performed on human hippocampal tissue from intractable temporal lobe epilepsy patients, but with unknown source of HMGB1 secretion, or the cause of initiation of this pathway. Also, seizure frequency and duration had been limited by trials using HMGB1-TLR4 antagonists. Those antagonists have been studied on the bicuculline-induced non-lesional model of seizures, leading to an increase in the onset of the latency period [3].

Suleiman et al. documented many epileptic cases with severe resistance to *antiepileptic* drugs (AED), but at the same time, they responded to immune therapy; these cases confirm the link between immune dysregulation and epilepsy, especially drug-resistant epilepsy [4]. Thus, a strong study should be constructed to highlight the relation between immune dysregulation, epilepsy, and the epileptic drug. Antiepileptic drugs (AEDs) are commonly available, but unfortunately, they have a risk of idiosyncratic effects, such as allergic reactions and organ damage. Also, about one third of epileptic patients have AEDs resistance; thus, an urgent need exists for effective therapies to be developed [5].

Therefore, in our current study, we aimed to determine the relation between TLR4 *rs1927914*, rs11536858, rs1927911, and epilepsy in an Egyptian case-control study and to study their relation to clinical data and response to antiepileptic drug (AED) therapy.

Methods

Patients

The study involved 133 Egyptian children divided into two groups: group I (83 pediatric patients), who were recruited to the inpatient and outpatient clinic of the pediatric neurology unit of a children hospital in 2017–2018 (40 boys and 43 girls), and group II. Group I has

Table 1 Primers for PCR-RFLP of the TLR4, restriction enzymes used, and RFLP products size and genotypes

SNP ID	Prime	Sequence	Restriction enzyme used	RFLP product size and genotypes
rs11536858	Forward	ATAACCTCAGTGGGCTCTGG	Kpnl	241 = AA
	Reverse	ATGTTCTGGCATCTGGGAAG		241, 190, 51 = AG 190, 51 = GG
rs1927911	Forward	TCACTTTGCTCAAGGGTCAA	Styl	203 = TT
	Reverse	AAACCTGCATGCTCTGCAC		203, 178, 25 = TC 178, 25 = CC
rs1927914	Forward	ACAAAATGGTCCCTCACAGC	Sphl	150 = TT
	Reverse	TGGAAAGTAGCAAGTGCAATG		157, 90, 67 = TC 90, 67 = CC

Table 2 rs1927914 genotype genetic polymorphism distribution between cases and control

Genotype rs1927914	Total			χ²	p value	OR (95% CI)
	number <i>n</i> = 133	Control n = 50 (%)	Cases n = 83 (%)			
CC	63	8 (12.7)	55 (87.3)	39.9	< 0.001*	15.3 (6.05–38.6)
CT	12	2 (16.7)	10 (83.3)	11.3	0.007*	11.1 (2.21–55.9)
TT (r)	58	40 (69.0)	18 (31.0)			1
HW equilibrium		< 0.001*	< 0.001*			

Significant association has been found as the homozygous CC genotype is more at risk for developing epilepsy than homozygous TT by 15.3 (ρ value < 0.001* and OR 15.3). The heterozygous CT genotype is more at risk for developing epilepsy than TT by 11.1 (ρ value 0.007* and OR 11.1)

two divisions, according to response to the antiepileptic drug: group Ia with 40 AED responders and a mean age of 6.83 ± 4.57 and group Ib with 43 drug resistance and a mean age of 8.58 ± 4.14 .

Group II included 50 clinically healthy subjects from the same geographical area.

Our patients were diagnosed and classified according to the guidelines of the International League Against Epilepsy.

Exclusion criteria consisted of any significant medical disorders as hepatic disease or renal disease, brain cancer, history of pseudo-seizures, side effects of AEDs, and any progressive degenerative or metabolic brain disorders.

By the International League Against Epilepsy 2010, drug-resistant epilepsy was diagnosed as a nonresponse to AED after 2 or more adequate trials which properly chosen and tolerated either one drug or more.

The local ethical committee approved the study.

Information on epilepsy diagnosis, age of onset, duration, types of drugs, family history of epilepsy or other autoimmune diseases, and response to therapy were abstracted from medical reports.

Genotyping

The DNA was extracted from whole blood that was collected on EDTA tubes and reserved at – 20 °C, until use. The genomic extraction was done by using Thermo Scientific DNA Purification mini kit for whole blood. The genotyping was done for TLR4 rs11536858 (now merged into rs10759931), rs1927911, and rs1927914 by the polymerase chain reaction-restriction fragment length polymorphism (PCR–RFLP) method according to [2].

The sequences of both forward and reverse primers (purchased from Applied Biosystems) and restriction enzymes are described in Table 1. The final reaction

volume was 25 µl which was composed of 4.0 µl H2O, 0.5 µl from each primer (forward and reverse), 15.0 µl DreamTaq Green PCR master mix (Tiagen, China), and 5 µl DNA. The PCR reactions were done by using Thermocycler PTC-100 (Bio-Rad, USA). The first step was initial denaturation for 5 min at 95 °C, then 35 cycles for 40 s at 94 °C, and 58 °C for 45 s and 72 °C for 40 s, and lastly the final extension step for 10 min at 72 °C. Before using restriction enzymes and to check PCR product, 5 µl of the products mixed with 2 µl of loading buffer were migrated on 2% agarose gel. For TLR4, rs11536858 was checked at 241 bp, rs1927911 at 203 bp, and rs1927914 at 157 bp.

The amplified PCR products for rs11536858, rs1927911, and rs1927914 were digested using the following restriction enzymes: KpnI, StyI, and SphI (New England Biolabs, USA) respectively. The total reaction volume was 30 μ l formed from 17 μ l nuclease-free water, 10 μ l of PCR products, 2.0 μ l 10X Fast Digest green buffer, and 1.0 μ l of restriction enzyme, and then the mixture was incubated. The temperature and duration of incubation was according to the manufacturer instructions. The DNA fragments produced from the reaction were revealed using agarose gel 3%. The genotypes for each SNP are mentioned in Table 1.

Statistical analysis

The analysis of the data was done using IBM SPSS software package version 20.0. Quantitative data were described using mean and median range for non-parametric data, after testing normality using the Kolmogorov-Smirnov test. The significance of the obtained results was judged at 5% level.

The used tests were as follows: comparison of variables between studied groups by using the chi-square test and Kruskal-Wallis test for comparing non-parametric

 Table 3 Association between rs1927914 alleles and epileptic cases

Genotype rs1927914	Total	Studied groups		χ²	p value	OR (95% CI)
	number n = 266	Control n = 100	Cases n = 166			
С	138	18 (13.1)	120 (86.9)	73.7	< 0.001*	1
T(r)	128	82 (64.1)	46 (35.9)			11.9 (6.4–21.9)

The total number of cases is 266; as it is the number of alleles, we duplicated the number of cases to be 266 instead of 133; a significant association between cases and control has been found as C allele is more at risk for developing epilepsy than T allele

Table 4 Association between TLR genetic polymorphism rs 1927914 and epilepsy type

Genotype rs1927914	Total cases	Epilepsy type		χ²	р	OR (95% CI)
	n = 83	Primary n = 61 (%)	Secondary n = 22 (%)			
CC	55	39 (70.9)	16 (29.1)	10.5	0.001*	6.3 (1.9–20.7)
CT	10	9 (90.0)	1 (10.0)	1.21	0.27	0.29 (0.03-2.9)
TT (r)	18	13 (72.2)	5 (27.8)			1
HW equilibrium		< 0.001*	< 0.001*			

Significant association was detected as p value 0.001*; cases with CC genotype are more at risk for developing primary epilepsy than TT cases by 6.3 times (OR = 6.3)

continuous variables of more than 2 studied groups. Mann-Whitney U test was used for comparing non-parametric continuous variables between the 2 studied groups. Hardy-Weinberg equilibrium was used to compare the frequency of genotype deviation from the general population. The odd ratio was calculated using cross tabulation through Epi Info. In allele statistics, we did the duplication for the studied groups for calculation of each allele frequency (number of cases = 166 and control = 100), and in allele combination or haplotype combination, we did the combination between the studied alleles (cases = 332 and control = 200).

Results

According to rs1927914 genotypes in Table 2, the homozygous CC genotypes are 15.3 times (p value < 0.001* and OR 15.3) more at risk for developing epilepsy than homozygous TT. The heterozygous CT genotype is 11.1 times (p value 0.007* and OR 11.1) more at risk for developing epilepsy than TT. Also, C alleles are 11.9 times (p value < 0.001* and OR 11.9) more at risk for developing epilepsy than the T allele, as shown in Table 3.

The association between TLR genetic polymorphism rs1927914 and epilepsy types were presented in Table 4; a significant association was detected as p value was 0.001^* , cases with CC genotype are more risky for developing primary epilepsy than TT cases by 6.3 times (OR = 6.3). Hardy-Weinberg equilibrium illustrates a statistically significant deviation of rs1927914 genotype among primary and secondary epilepsy cases. However, the rs1927914 allele demonstrates a non-statistically significant association with epilepsy types (primary and secondary), as mentioned in Table 5.

A significant association was found in homozygoze AA genotypes of rs11536858, being more risky for developing epilepsy than homozygoze GG by 14.5 times with p value < 0.001* and OR 14.5. A allele is 3.8 times more at risk for developing epilepsy than G allele with p value < 0.001* and OR 3.8, as mentioned in Tables 6 and 7, respectively.

Analysis of the association between drug resistance and genotypes was analyzed in Table 8. It had a statistically significant association between clonazepam resistance and rs11536858 as a p value $< 0.001^*$ with the highest frequency with AA genotypes (4.3%).

According to rs1927911, no significant results were found between study cases and control groups, or between drug-responsive and drug resistance, as shown in Tables 9 and 10.

According to the rs11536858 and rs1927914 allele combination, there is a statistically significant difference between the study cases and control groups regarding all combinations. The highest combination frequency among cases was CG (82.2%), and then CA (36.8%), as shown in Table 11.

The highest frequency of genotype combination among case was CC/TT/CT (22.9%) followed by CC/CT/CT (15.7%) while the among control group, TT/CT/CT (32%) was the highest followed by TT/CC/CT (16.0%). Genotype combinations have a statistically significant difference between study cases and control groups which were CC/TT/CT, TT/CC/CT, TT/CT/CC, and TT/CT/CT, as shown in Table 12.

Discussion

Epilepsy is one of the most common neurological disorders. It represents about 27% of all neurological diagnosis. Different studies from Egypt reported the prevalence of epilepsy

Table 5 Association between TLR genetic polymorphism *rs1927914* alleles and epilepsy type

Genotype rs1927914	Total	Studied groups		χ²	р	OR (95% CI)
	number <i>n</i> = 166	Primary n = 122 (%)	Secondary n = 44 (%)		value	
С	120	87 (71.3)	33 (75.0)	0.22	0.64	1.21 (0.55–2.65)
T(r)	46	35 (38.7)	11 (25.0)			1

Table 6 rs11536858 Genotype distribution between cases and control

Genotype rs11536858	Total number n = 133	Studied groups		χ²	p value	OR (95% CI)
		Control n = 50 (%)	Cases n = 83 (%)			
GG (r)	19	13 (68.4)	6 (31.6)			1
GA	60	30 (50.0)	30 (50.0)	1.97	0.16	2.2 (0.73-6.5)
AA	54	7 (13.0)	47 (87.0)	21.7	< 0.001*	14.5 (4.2–50.8)
HW equilibrium		0.88	0.69			

Significant association has been found. Homozygoze AA genotype is more at risk for developing epilepsy than homozygoze GG by 14.5 with p value < 0.001* and OR 14.5)

among Egyptian children. A study reported that the lifetime prevalence of epilepsy in children below 18 years old was 9.7/1000 [6]. Another two studies reported a prevalence of 7–10/1000 among children at school who were less than 15 years of age [7, 8], while another study reported a higher rate of 12.9/1000 among children < 12 years [9].

Multiple findings encourage us to study the TLR4 in epilepsy. Firstly, TLR4 expression in the brain tissue was proven in different studies [10–12]. Secondly, growing evidence supports the hypothesis that inflammation within brain tissue is a cornerstone for the development of seizures [13–17]. Thirdly, the relation between TLR4, as a key player in innate immune response, and epilepsy has been found on both molecular and pharmacological levels [18–20]. Previous researches were lacking proper studies on children, so we decided to do our pediatric study.

In the present study on a cohort of Egyptian pediatric patients, we tried to strengthen the previous findings concerning the relation between TLR4 and epilepsy by investigating this relation on the genetic level. We focused on 3 SNPs in TLR4 (rs1927911, rs1927914, and rs11536858). These SNPs have been studied in different disorders with or without immunological bases like prostate cancer [21], Alzheimer [22], normal-tension glaucoma [23], and kidney transplant rejection [24]. To our knowledge, this is the first study investigating TLR4 gene polymorphisms and their relation to epilepsy development and antiepileptic drug resistance.

Our results showed a significant association between homozygous CC and heterozygous CT genotypes of TLR4 rs1927914, homozygous AA genotype of TLR4 rs11536858, and epilepsy (p < 0.05). These genotypes can be considered as a risk factor for epilepsy

Table 7 rs11536858 allele distribution between cases and control

rs11536858	Studied g	roups	χ²	p value	OR (95%	
allele	Control n = 100	Cases n = 166			CI)	
G (r)	56 (57.1)	42 (42.9)	25.3	< 0.001*	1	
A	44 (26.2)	124 (73.8)			3.8 (2.2–6.4)	

Significant association has been found as A allele is more at risk for developing epilepsy than G allele by 3.8 with p value < 0.001* and OR 3.8)

development. Also, the C allele for rs1927914 can be considered a risky allele (p < 0.001); on the other hand, the G allele for rs11536858 can be considered as a protective allele (p < 0.001). For epilepsy subtypes, either primary or secondary, a significant association was found between CC genotype of rs1927914 and primary epilepsy (p < 0.001). There was no significant association between rs1927911 (genotype and alleles) and epilepsy (p > 0.05).

In literature, different studies supported the role of TLR4 in one of the following: occurrence, severity, and/ or persistence of epilepsy. TLR4 astrocyte mediates the development of the excitatory synapse in young mice after strong immune activation, and this effect could be blocked by TLR4 antagonists [25]. The same findings were recorded by an earlier study after intraperitoneal injections of LPS in 2-week-old mice [26]. High mobility group box 1 (HMGB1), the ligand for TLR4, together with TLR4, was found to have more expression in drugresistant temporal lope epilepsy; moreover, they found an increase in the frequency of seizures in the mice model with increased expression of HMGB1 [3]. The same result was obtained when HMGB1 serum level was measured in epilepsy cases [27]. In the same context, a

Table 8 Association between rs11536858 genotype and epilepsy drugs used

Drug	rs115368	58 genotype	2	χ²	р
	GG n = 6(%)	GA n = 30(%)	AA n = 47(%)		
Na valproate	6 (100)	29 (96.7)	39 (83.0)	4.3	0.11
Carbamazepine	1 (16.7)	4 (13.3)	14 (29.8)	2.9	0.23
Levitracetapine	5 (83.3)	17 (56.7)	24 (51.1)	2.3	0.32
Oxacarbamazepine	0	4 (13.3)	6 (1.8)	0.9	0.64
Topiramate	3 (50.0)	10 (33.3)	12 (25.5)	1.7	0.42
Clonazepam	4 (66.7)	1 (3.3)	2 (4.3)	28.4	< 0.001*
Phenytoin	0	2 (6.7)	1 (2.1)	1.3	0.52
Lacosamide	0	0	1 (2.1)	0.78	0.68
Lamotrigine	0	1 (3.3)	3 (6.4)	0.69	0.71

Statistically significant association between clonazepam resistance and rs11536858 as p value < 0.001* with the highest frequency with AA genotypes (4.3) %

Table 9 rs1927911 genetic polymorphism distribution between cases and control

Genotype	Studied g	roups	χ²	р	OR (95% CI)	
rs1927911	Control n = 50	Cases n = 83		value		
CC	13 (31.0)	29 (69)	1.5	0.22	2.2 (0.6–8.2)	
CT	31 (39.2)	48 (60.8)	0.49	0.37	1.55 (0.46-5.2)	
TT (r)	6 (50.0)	6 (50.0)			1	

No significant results were found between cases and control

higher dose of kinic acid was needed to induce epileptic fits in TLR4 knockout mice [28]. Another study found that increased TLR4 expression was associated with an increase in the frequency of fits [29].

Regarding AEDs resistance, patients with multiple AEDs usually develop drug resistance. We examined the relation between TLR4 SNPs and resistance to different AEDs like Na valproate, carbamazepine, clonazepam, phenytoin, and topiramate. No relation was found between the studied SNPs and drug resistance except for the rs11536858 with clonazepam (p < 0.001). This association may be explained by the association of this genotype with benzodiazepine receptor action in epilepsy. The relation between gene polymorphisms and AEDs resistance was investigated in different studies. In previous studies, for example, they focused on ABCB1 C343T polymorphism with different findings [30–32].

Stimulation of TLR4 either by exogenous agent (LPS) or endogenous ligand (heat shock protein) enhances the production of pro-inflammatory mediators like IL1, TNF, prostaglandins, and complement cascade, which has an active role in seizures development and exacerbation [33-35]. Myeloid differentiation 88 (MYD88) is an essential intracellular mediator for TLR4 signaling; after TLR4 stimulation by LPS in young animals, the formation of excitatory synapses is increased by the activation of TLR4-MyD88-ERK (excitatory signal regulated kinase) signaling pathway [26]. Another theory explains that the role of TLR4 in epilepsy involves HMGB1. Activation of the TLR4-HMGB1 pathway enhances the release of Ca, which increases the excitability of neurons, as well as neuronal damage, and decreases the threshold for fit development, which means more frequency of epileptic fits [36, 37]. In a pilocarpine rat model of epilepsy, early

Table 10 rs1927911 allele polymorphism distribution between cases and control

Allele	Total	Studied o	groups	χ²	Р.	OR	
rs1927911	number n = 266	Control n = 100	trol Cases 100 <i>n</i> = 166		value	(95% CI)	
C (r)	163	57 (34.9)	106 (65.03)	1.24	0.27	1	
Т	103	43 (41.7)	60 (58.3)			1.33 (0.8–2.2)	

No significant results were found between cases and control (the total number of cases is 266; as it is the number of alleles, we duplicated the number of cases to be 266 instead of 133)

Table 11 rs11536858 and *rs1927914* allele combination distribution between cases and control

Haplotype combination	Control N = 400 (%)	Cases N = 664 (%)	Test of significance	Bonferroni correction
CG	74 (18.5)	162 (82.2)	p = 0.02*	p = 0.01*
CA	62 (15.5)	244 (36.8)	p < 0.001*	p = 0.001*
TG	138 (34.5)	88 (13.3)	p < 0.001*	p = 0.001*
TA	126 (31.5)	170 (25.6)	p = 0.03*	p = 0.015*

There is a statistically significant difference between cases and control regarding all combinations with the highest combination frequency among cases being CG (82.2%) and then CA (36.8%)

activation of TLR4 and TLR2, probably through preserving normal hippocampal cytokine profile and neuronal function, attenuates seizure severity [38]. A new therapeutic approach of epilepsy by using TLR7 inhibitors has been suggested after observing the neuroinflammatory cascade using the microRNAs TLR7 activator [39]. Accordingly, Liao et al. suggested that electronic

Table 12 Genotype combination among studied cases and control groups

control groups			
Genotype combination	Control n = 50	Cases n = 83	
CC/CC/CC	1 (2.0)	0 (0.0)	FET, p = 0.38
CC/CC/CT	1 (2.0)	3 (3.6)	FET, $p = 1.0$
CC/CT/CC	1 (2.0)	7 (8.4)	$\chi^2 = 2.26, p = 0.13$
CC/CT/CT	3 (6.0)	13 (15.7)	$\chi^2 = 2.75, p = 0.09$
CC/CT/TT	1 (2.0)	0 (0.0)	FET, $p = 0.38$
CC/TT/CC	1 (2.0)	9 (10.8)	$\chi^2 = 3.5, p = 0.06$
CC/TT/CT	0 (0.0)	19 (22.9)	$\chi^2 = 13.4, p = 0.002*$
CC/TT/TT	1 (2.0)	4 (4.8)	FET, $p = 0.65$
CT/CC/TT	1 (2.0)	0 (0.0)	FET, $p = 0.38$
CT/CT/CC	1 (2.0)	2 (2.4)	FET, $p = 1.0$
CT/CT/TT	0 (0.0)	1 (1.2)	FET, $p = 1.0$
CT/TT/CC	0 (0.0)	4 (4.8)	FET, $p = 0.29$
CT/TT/CT	0 (0.0)	2 (2.4)	FET, $p = 0.53$
CT/TT/TT	0 (0.0)	1 (1.2)	FET, $p = 1.0$
TT/CC/CC	0 (0.0)	3 (3.6)	FET, $p = 0.29$
TT/CC/CT	8 (16)	0 (0.0)	$\chi^2 = 14.12, p = 0.002*$
TT/CC/TT	2 (4)	0 (0.0)	FET, $p = 0.14$
TT/CT/CC	7 (14)	2 (2.4)	$\chi^2 = 6.64, p = 0.009*$
TT/CT/CT	16 (32)	5 (6.0)	$\chi^2 = 15.83, p = 0.001*$
TT/CT/TT	1 (2)	0 (0.0)	FET, $p = 0.38$
TT/TT/CC	2 (4)	2 (2.4)	FET, $p = 0.63$
TT/TT/CT	3 (6)	6 (7.2)	$\chi^2 = 0.07, p = 0.78$
Total	50 (100.0)	83 (100.0)	

The highest frequency of genotype combination among cases was CC/TT/CT (22.9%) followed by CC/CT/CT (15.7%) while among the control group, TT/CT/CT (32%) was the highest followed by TT/CC/CT (16.0%). Genotype combinations have a statistically significant difference between cases and control groups which were CC/TT/CT, TT/CC/CT, TT/CT/CC, and TT/CT/CT

stimulation of the ear could control epileptic seizures by TLR4 signaling pathway regulation, which may be a novel therapeutic approach for epileptic seizures [40].

Conclusion

Possible involvement of the Toll-like receptor clarifies the importance of innate immunity in initiating seizures and making neuronal hyperexcitability. In this work, multiple significant associations between TLR SNPs and epilepsy, epileptic phenotype, and drug-resistant epilepsy have been found. More studies with larger samples sizes and different techniques along with different SNPs are recommended to find potential immunotherapeutic approaches as a sole or adjuvant therapy for epilepsy.

Abbreviations

AEDs: Antiepileptic drugs; DAMPs: Damage-associated molecular patterns; HMGB1: High mobility group box 1 protein; LPS: Lipopolysaccharide; MAF: Minor allele frequencies; PCR: Polymerase chain reaction-restriction fragment length; RFLP: Restriction fragment length polymorphism; SNP: Single-nucleotide polymorphisms; TLR4: Toll-like receptor 4

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Authors' contributions

MA wrote the paper and analyzed and interpreted the patient data regarding the disease. DS performed the examination of the patient and collection of the samples. HM made the statistical analysis of the data. AM participated in the writing, MM was a major contributor in writing the manuscript. All authors read and approved the final manuscript.

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Availability of data and materials

The data that support the findings of this study are available from the corresponding author upon reasonable request.

Ethics approval and consent to participate

This study was approved by the ethics committee of Faculty of Medicine, Mansoura University, with approval number [R.20.04.808]. The patient provided written consent.

Consent for publication

Written parental consent had been taken, and this was approved by the ethics committee.

Competing interests

The authors declare that they have no competing interests.

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