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Serum neutrophil gelatinase-associated lipocalin in obese adolescents

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Abstract

Background: Nonalcoholic fatty liver disease (NAFLD) is an emerging clinical problem among obese adolescents.

Methods: This study was conducted on 40 obese adolescents, previously diagnosed by ultrasonography to determine liver status and confirmed to have NAFLD, and 40 non-obese healthy controls matching in age and sex. Anthropometric measurements were taken and the Homeostatic Model Assessment-Insulin Resistance (HOMA-IR) test was performed. Levels of C-reactive protein (CRP) and neutrophil gelatinase-associated lipocalin (NGAL) were measured, and the biochemical parameters like blood glucose, insulin, lipid profile, and liver enzymes (ALT and AST) were also measured.

Results: Anthropometric measurements were significantly higher in the obese group than in the control group ($P < 0.05$). Obese adolescents had significantly higher values of serum insulin, HOMA-IR, CRP, ALT, triglycerides, total cholesterol ($P < 0.05$), and NGAL ($P < 0.01$) and a lower HDL-C value than the control group. Also, we found a highly significant correlation between the CRP, insulin, and NGAL levels where $P < 0.01$.

Conclusion: NGAL can be used as a biomarker for predicting incidence of NAFLD in obese adolescents.

Trial registration: NRC17006. Registered: 19 January 2017

Keywords: NGAL, NAFLD, Obesity, CRP, Adolescents

Background

Obesity is a major worldwide health problem and is strongly associated with a number of diseases. There is an increasing evidence that obesity is causally linked to a chronic low-grade inflammatory state as shown by the high levels of inflammatory markers (Polyzos et al. 2017; Wu et al. 2016).

Obesity is frequently associated with insulin resistance which is regarded as a critical factor that contributes to the development of nonalcoholic fatty liver disease (NAFLD) (Neuschwander-Tetri 2017). Neutrophil gelatinase-associated lipocalin (NGAL) is secreted by adipocytes and immune cells such as neutrophils and macrophages and was initially described to play a role in innate immunity, perhaps by sequestering iron (Bulluck et al. 2017). Since NGAL is abundantly

expressed in adipose tissue and the liver, we suggest that it might be useful as a noninvasive biomarker of NAFLD and in evaluating the outcomes of various clinical interventions in such obesity-related metabolic disease. The aim of our study is to determine the NGAL role in the pathogenesis of NAFLD and to assess its relationship with CRP and insulin resistance among adolescent obese Egyptians.

Methods

The study group included 40 obese adolescents (BMI > 90th percentile) recruited from an ultrasonography clinic in Al-Zahraa Hospital, Cairo, Egypt. They were 15 males and 25 females, and their ages ranged from 12 to 17 years. Obese participants were previously examined by ultrasonography to detect liver status and confirmed to have NAFLD. The control group comprised 40 non-obese adolescents who were age- and sex-matched with the study group.

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Biochemical analysis

Morning 5-ml venous blood samples were withdrawn after a 12-h overnight fasting into plane tubes and were left to clot; a small portion of blood was separated in sodium fluoride-coated tubes for glucose level determination. The serum was separated by centrifugation for 10 min at 5000 rpm and was stored at -20°C until assayed.

Triglyceride (TG), total cholesterol (TC), and high-density lipoprotein cholesterol (HDL-C) levels were assayed by enzymatic colorimetric methods according to Wahlefeld (1974), Allain et al. (1974), and Warnick et al. (1982) respectively using kits purchased from Spinreact, Girona, Spain. Low-density lipoprotein cholesterol (LDL-C) was calculated using the Friedewald equation [$\text{LDL-C} = \text{total cholesterol} - \text{HDL-C} - (\text{triglyceride}/5)$] when the triglyceride concentrations did not exceed 4.8 mmol/L (Friedewald et al. 1972).

Plasma glucose level was determined by the glucose oxidase method (Barham and Trinder 1972) using a kit purchased from Spinreact, Girona, Spain, and serum insulin was measured by using the quantitative enzyme-linked immunosorbent assay (ELISA) using a commercial kit provided by DIA source, Belgium (Turkington et al. 1982).

The Homeostatic Model Assessment-Insulin Resistance (HOMA-IR) method was used for the calculation of insulin resistance. This method has been validated as a reliable measure of insulin resistance in vivo in humans (Michael 2008).

HOMA-IR

$$= \frac{\text{Fasting serum insulin } (\mu\text{U/ml}) \times \text{fasting plasma glucose (mmol/L)}}{22.5}$$

Serum CRP was measured using the ELISA kit provided by Immunospec, CA, USA (Ridker et al. 1998).

Serum ALT and AST activities were measured by a colorimetric procedure using a kit provided by Linear Chemicals S.L., Barcelona, Spain (Bergmeyer et al. 1985).

Serum NGAL levels were measured using the ELISA kit provided by Bio-Rad, Hercules, CA, USA (Mitsnefes et al. 2007).

Anthropometric parameters

According to the recommendations of the International Biological Program (Hiernaux and Tanner 1969), anthropometric measurements including height, weight,

waist circumferences, and hip circumferences were measured by well-trained anthropologists. Body weight was determined to the nearest 0.01 kg using a Seca scale balance, Hamburg, Germany, with the subject wearing minimal clothing and no shoes. The body height was measured to the nearest 0.1 cm in a standing position using a Holtain portable anthropometer with fixed stadiometers. Circumferences were obtained with a flexible non-stretchable measuring tape. The waist circumference (WC) was measured at the midpoint between the lower rib margin and the iliac crest with the subject standing at the end of normal expiration. The hip circumference (HC) was measured at the level of the greater trochanters with the subject wearing minimum clothing. The mean of the two readings was taken in for calculating the waist-to-hip ratio (WHR).

The body mass index (BMI) was calculated by dividing the weight (kg) by the squared value of height in meters. Obese subjects are defined as having a BMI above the 95th percentile for age and sex based on the established percentile curves for Egyptian children and adolescents (Ghalli et al. 2008). Statistical analysis was performed using independent-sample *T* test for comparison between means and bivariate Pearson test for correlations by SPSS v.16, SPSS Inc. Chicago, USA.

Results

Sex and age of study subjects are presented in Table 1 and their biochemical parameters are presented in Table 2 and Figs. 1, 2, and 3. Statistics show that the obese group has significantly higher values of serum insulin, HOMA-IR, CRP, ALT, triglycerides, total cholesterol ($P < 0.05$), and NGAL ($P < 0.01$) and lower HDL value than the control group. Also, a highly significant ($P < 0.01$) positive correlation between serum NGAL and CRP ($r = 0.88$) (Fig. 4), CRP and insulin ($r = 0.84$) (Fig. 5), and NGAL and insulin ($r = 0.84$) (Fig. 6) levels were found.

Anthropometric data

Table 3 shows comparison of the anthropometric parameters between the control and the obese groups. Statistics show that obese subjects have significantly higher anthropometric measurements than control subjects where $P < 0.05$.

Table 1 Sex and age distribution in control and obese groups

| | Control | Mean age (years) | <i>P</i> | Obese | Mean age (years) | <i>P</i> |
|--------|------------|------------------|----------|------------|------------------|----------|
| Male | 13 (32.5%) | 14.23 | 0.42 | 15 (37.5%) | 14.73 | 0.42 |
| Female | 27 (67.5%) | 14.07 | 0.32 | 25(62.5%) | 14.6 | 0.29 |
| Total | 40 (100%) | | | 40 (100%) | | |

(%) : percent from total subjects, *P* : SE value

Table 2 Comparison of laboratory data between control and obese groups

| Parameter | Control | Obese |
|----------------------------|---------------|-----------------------------|
| Glucose (mg/dl) | 90.53 ± 6.18 | 95.30 ± 8.23 |
| Insulin (μIU/ml) | 5.01 ± 0.20 | 11.37 ± 0.89 ^a |
| HOMA-IR | 0.89 ± 0.021 | 4.973 ± 0.29 ^a |
| AST(U/L) | 24.62 ± 2.12 | 25.19 ± 3.22 |
| ALT(U/L) | 14.06 ± 0.84 | 22.23 ± 1.51 ^a |
| HDL-cholesterol (mg/dl) | 49.00 ± 4.35 | 46.75 ± 5.41 |
| Triglycerides (mg/dl) | 80.35 ± 1.63 | 123.49 ± 3.79 ^a |
| Total cholesterol (mg/dl) | 160.13 ± 8.82 | 174.66 ± 11.93 ^a |
| LDL-cholesterol (mg/dl) | 98.56 ± 5.74 | 103.37 ± 4.65 |
| C-reactive protein (ng/ml) | 1320 ± 29.2 | 3319 ± 84.9 ^a |
| NGAL (ng/ml) | 74.02 ± 4.78 | 324.98 ± 12.3 ^b |

All data are presented as mean ± SE

^aSignificant versus control group at $P < 0.05$

^bSignificant versus control group at $P < 0.01$

Discussion

Obesity is almost always associated with a low-grade chronic inflammation due to proinflammatory cytokine release (Georgio et al. 2013). This inflammation truly represents the critical “link” between obesity and insulin resistance (IR). In turn, IR is the main backbone player in NAFLD pathogenesis. Researches have reported that 76% of obese adolescents have NAFLD (Georgio et al. 2013; Seung-Hoi 2013; Basaranoglu et al. 2015).

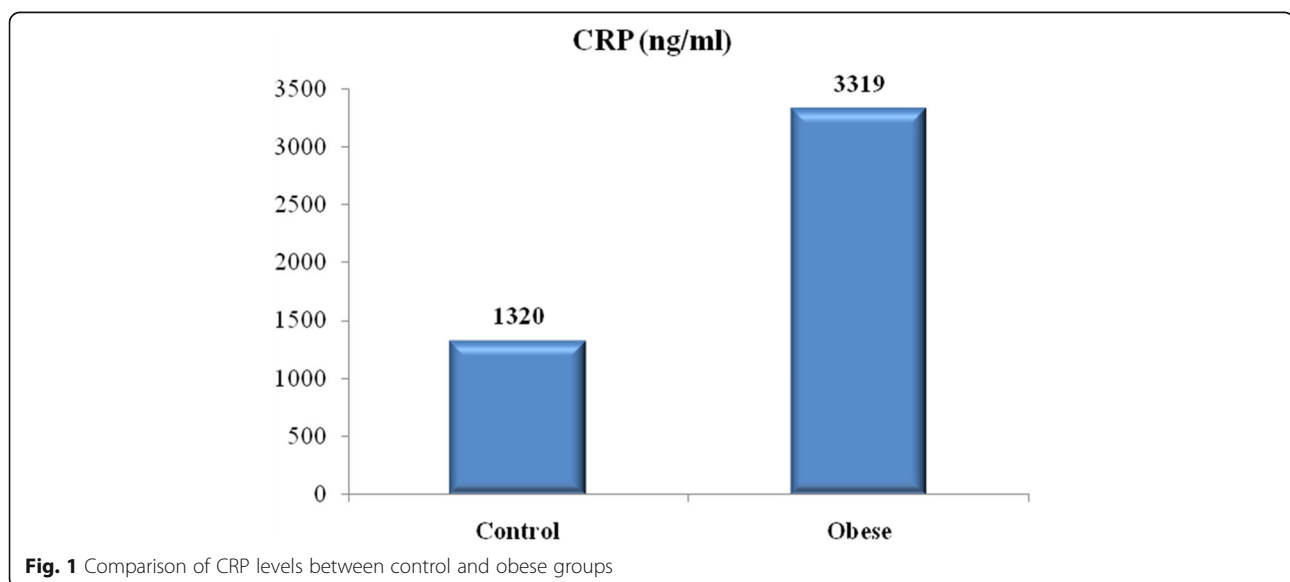
In this study, inflammation was displayed through the significant elevation of the CRP level in the obese group compared to the control group. That striking elevation might not only be due to obesity but also due to NAFLD, where NAFLD could progress from simple steatosis up to steatohepatitis (NASH). Accordingly, there is

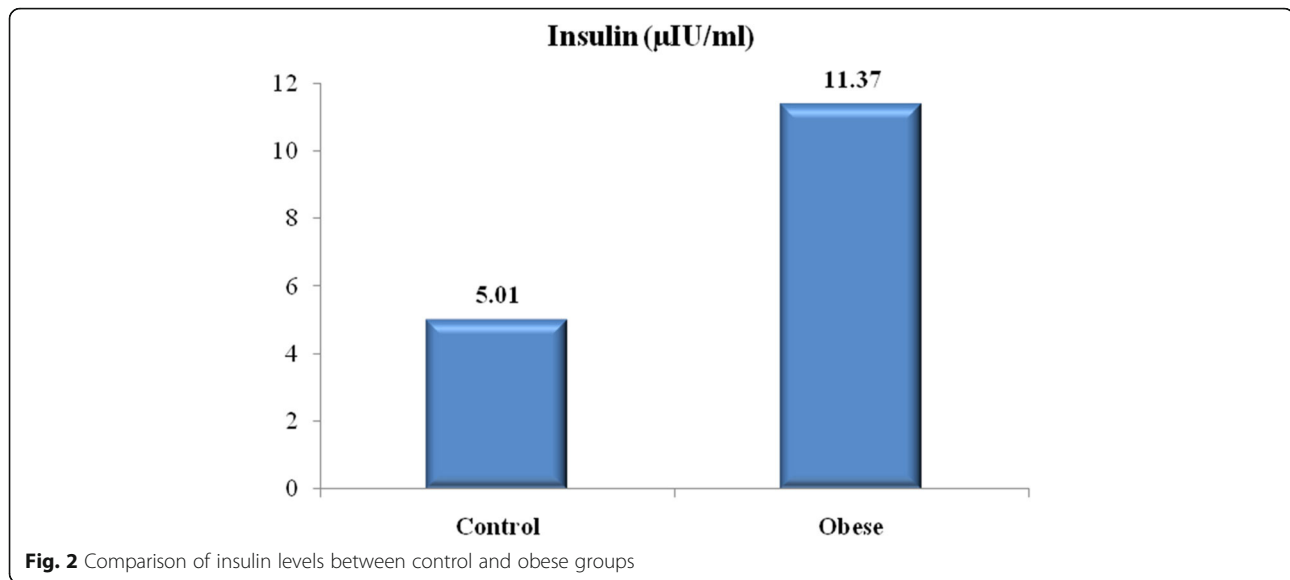
the additional liver cell inflammation besides that of the adipose tissue cells. Similarly, other studies reported a marked rise of the CRP levels in their obese adolescents compared to the non-obese ones (Esser et al. 2014; Lichtenauer et al. 2015).

Concerning the critical issue of IR, the studied obese adolescents exhibited higher blood glucose levels and significantly higher triglycerides and cholesterol levels compared to their control peers. Consequently, their blood insulin levels were also much higher and their “IR states” were so profound as proved by the HOMA-IR test. In accordance with these findings, many researchers emphasized the presence of IR states in their obese subjects (Lichtenauer et al. 2015; Puspitadewi et al. 2013; Chandrasekhar et al. 2014).

At this point, it is worth noting that our work revealed a significant positive correlation between the CRP and the insulin levels in the obese group ($P = 0.001$). Some studies were matching with this finding and found significant positive correlations between the CRP and the increased risk of type 2 diabetes in their obese subjects (Esser et al. 2014; Wang 2013). Our obese subjects were found to be suffering from NAFLD depending on abdominal ultrasonography examination. This was an expected finding due to several reasons that were confirmed by our investigations and is in agreement with El-Koofy et al. (2012), Seung-Hoi (2013), Georgio et al. (2013), and Huang and Yang (2013).

It was reported that NAFLD incidence increases in the obese and the extremely obese patients (up to 65 and 85% respectively) whose BMI is 30–39.9 kg/m² respectively (Basaranoglu et al. 2015). In addition Schwimmer et al. (2014) found out that their NAFLD subjects displayed evidences of inflammation and oxidative stress





(OS). This study showed that O.S. is well known to be a driver towards IR. This finding was reported by other work studies.

NAFLD by definition is the excessive accumulation of triglycerides in the liver without alcohol intake (Seung-Hoi 2013), and in our study, obese subjects did have highly significant levels of serum triglycerides and cholesterol.

Our obese group adolescents suffered from visceral adiposity (also called central or abdominal adiposity). This was elicited through their WC, HC, and WHR. All measurements were significantly much higher than those of their control mates. In accordance with us, many researchers have emphasized that the visceral adiposity is much more crucial than mere subcutaneous adiposity concerning the risk of NAFLD occurrence (Fishbein et al. 2006; Fan and Farrell 2008). This may be due to the finding that the visceral adipose tissue plays a pivotal role in NAFLD pathogenesis because it participates in the production of most of the adipokines that are involved in causing inflammation and IR (Georgio et al. 2013).

Matching with our findings and verifying the former explanations, Suarez-Alvarez et al. (2013) and Nappo et al. (2013) found out a significant positive correlation between the high CRP level (inflammation) and the increased waist-to-hip ratio. Several studies emphasized significant positive correlations between IR and waist circumference and BMI (Chandrasekhar et al. 2014; Gobato et al. 2014).

Moreover, in our work, the AST and the ALT enzymes were higher in the obese NAFLD group more than the control group with special referral to the ALT enzyme because it was significantly higher. The “liver fat infiltration” explains that increase in both liver enzymes but as the ALT was reported by some studies to be more

sensitive, so its rise was a significant one. Actually, the ALT enzyme is not commonly found outside the liver; accordingly, its high levels mostly accompany the increased liver fat fraction. Many researches confirmed that there was a significant positive correlation between ALT level and liver size in children (Rodriguez et al. 2010; Lenzi et al. 2015).

In our study, the NGAL serum levels were significantly higher in the obese group subjects more than their control peers ($P < 0.01$). In addition, there were highly significant positive correlations between NGAL and the insulin level and also between NGAL and the CRP level ($P = 0.001$). All that was very much expected because in obesity there is a low-grade chronic inflammation and NGAL is an acute-phase protein, like CRP, and an independent predictor of inflammation (Martins et al. 2014; Zaki et al. 2015).

Actually, many research studies declared NGAL to be an inflammatory marker closely related not only to obesity but also to its metabolic complications of which “IR” is an alarming issue (Wang et al. 2007; Shieh et al. 2014). Concerning the positive correlation between NGAL and insulin resistance (IR), it is worth noting to mention that lipopolysaccharides induce the expression of NGAL in the liver (Martins et al. 2014; Zaki et al. 2015). In obesity, there are high levels of lipopolysaccharides which not only act as inflammatory stimuli but are also one of the reasons why IR occurs. Accordingly, we can consider that lipopolysaccharides are one single common factor or explanation between NGAL high levels and IR state elevation. Another explanation is that NGAL antagonizes the effects of TNF- α in adipocytes and macrophages where TNF- α is well known to promote IR (Martins et al. 2014; Shieh et al. 2014; Dimitrova et al. 2014).

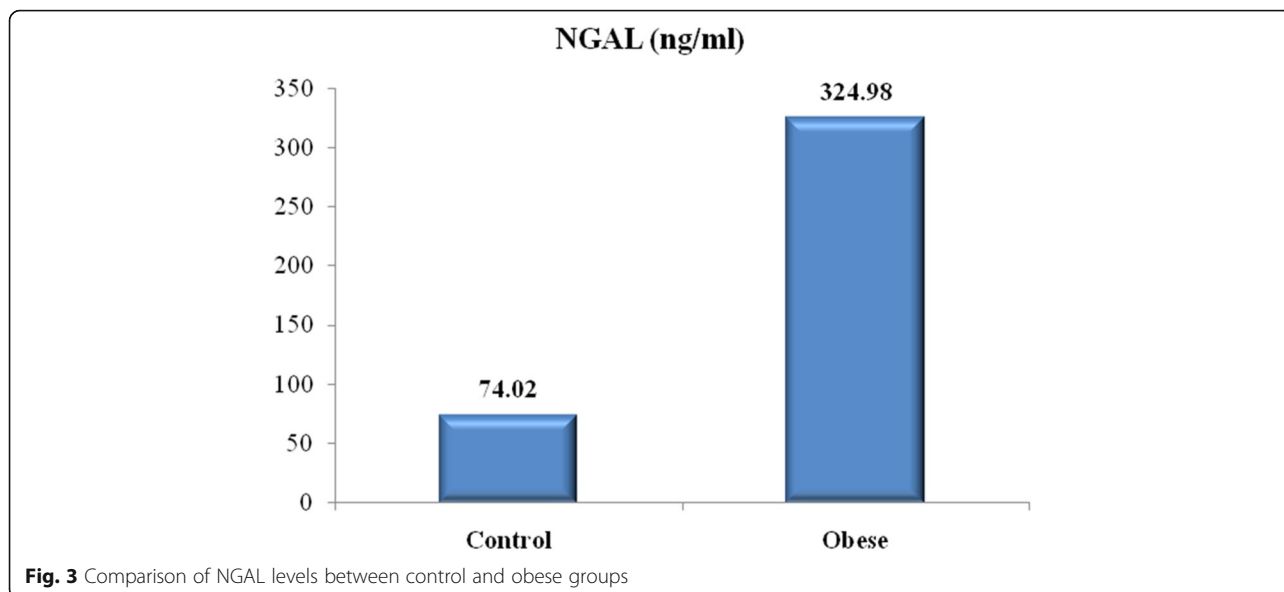


Fig. 3 Comparison of NGAL levels between control and obese groups

In accordance with our results, two studies found out that NGAL expression is increased by agents which promote IR, and this emphasizes its role in obesity IR (Yan et al. 2007; Fried and Greenberg 2012).

Still matching with our findings, Wang et al. (2007) reported that in human beings, NGAL levels correlated positively with IR, hypertriglyceridemia, and BMI. It correlated negatively with HDL-C. However, and contrary

to our results, some researchers did not find any relationship between serum NGAL and the anthropometric or the metabolic parameters ($P > 0.05$). Thus, they do not suggest routine NGAL measurements in obese subjects (Akelma et al. 2012).

Liver neutrophil activation is a “double-edged sword” (Xu et al. 2014). Normally, neutrophils (and other cell subsets) have a particular pivotal role in the liver innate

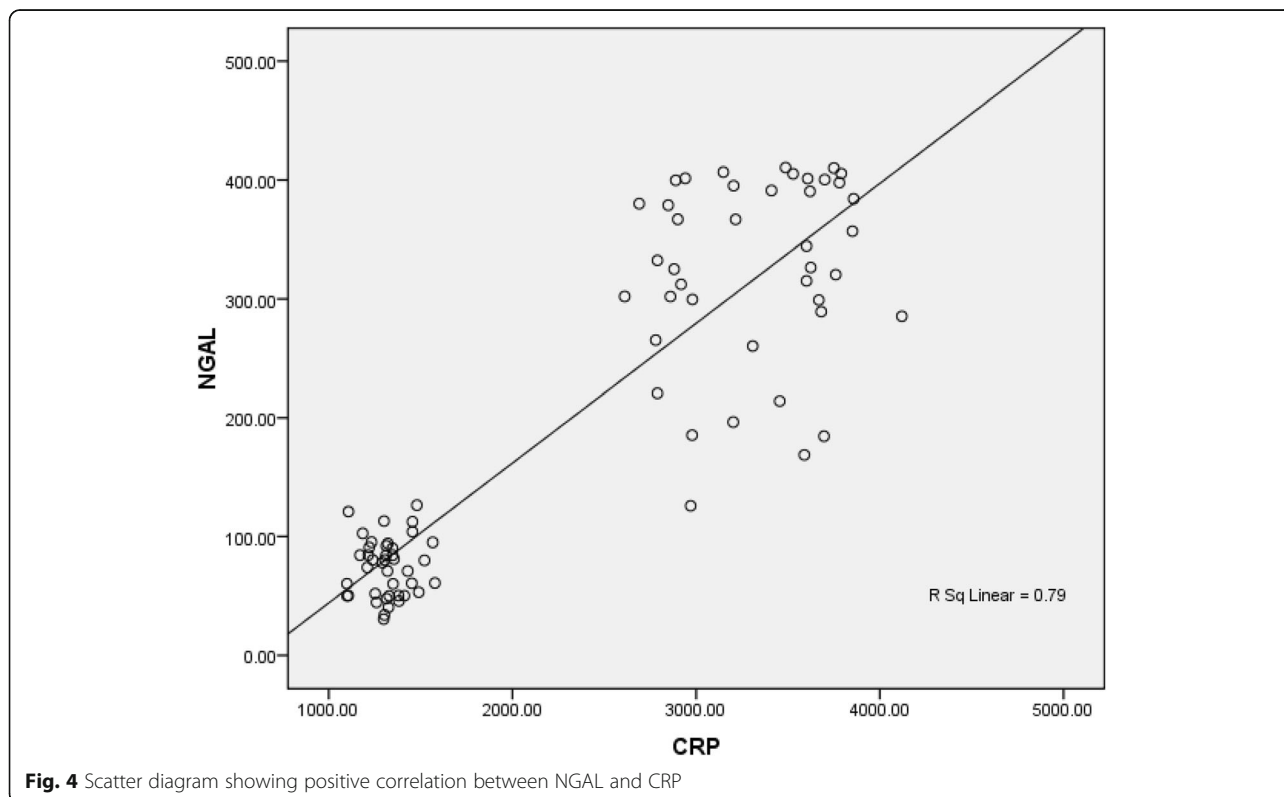


Fig. 4 Scatter diagram showing positive correlation between NGAL and CRP

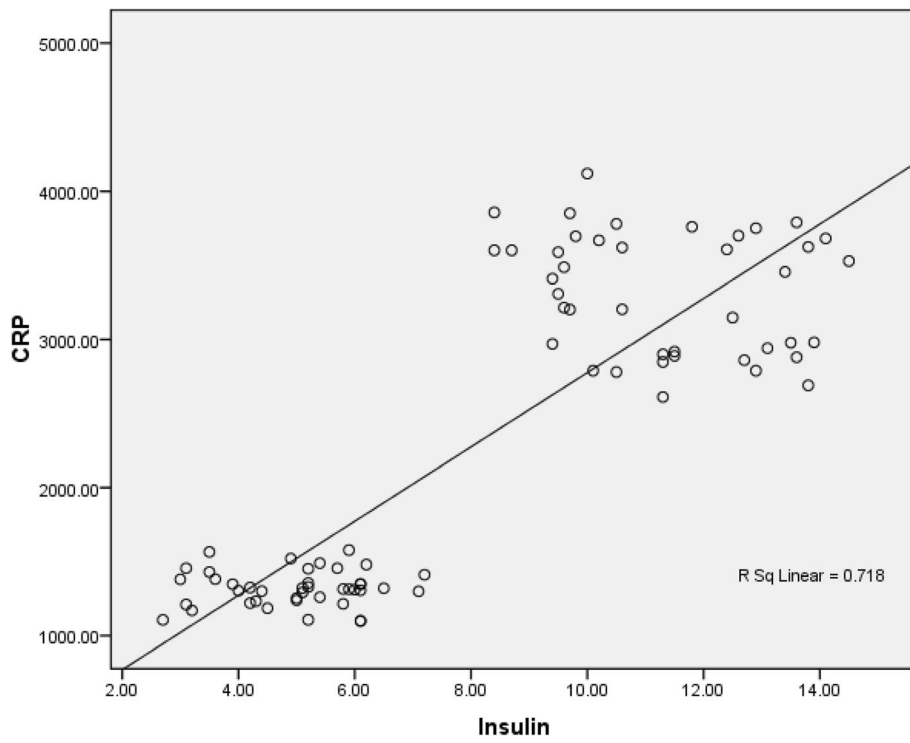


Fig. 5 Scatter diagram showing positive correlation between CRP and insulin

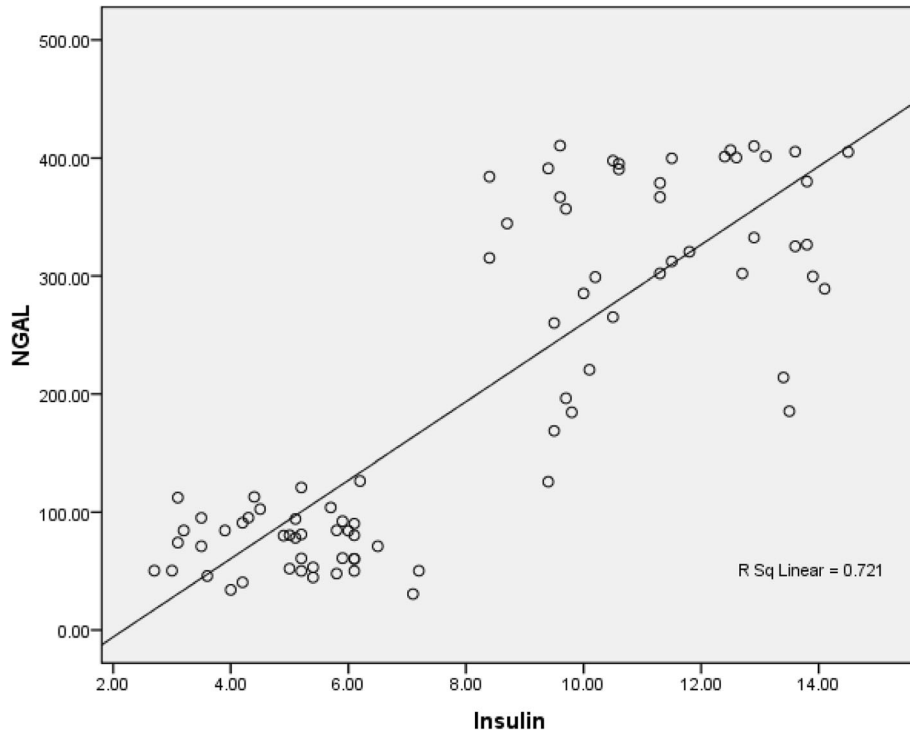


Fig. 6 Scatter diagram showing positive correlation between NGAL and insulin

Table 3 Comparison of the anthropometric parameters between control and obese groups

| Parameter | Control | Obese |
|--------------------------|----------------|----------------------------|
| BMI (kg/m ²) | 19.55 ± 3.763 | 38.09 ± 3.493 ^a |
| Waist circumference (cm) | 71.21 ± 2.56 | 106.65 ± 4.25 ^a |
| Hip circumference (cm) | 89.75 ± 3.78 | 123.21 ± 4.48 ^a |
| Waist-to-hip ratio | 0.7951 ± 0.062 | 0.8670 ± 0.08 ^a |

All data are presented as mean ± SE

^aSignificant versus control group at $p < 0.05$

immune system. However, on the other side when an overwhelming activation of neutrophils occurs, here liver damage occurs. This “aberrant neutrophil activity” has been evidenced from both human patients and animal investigations as a cause of liver diseases.

In our own words, we can only guess that since NGAL is abundantly expressed in the liver neutrophil granules and has been declared to be implicated in innate immunity, inflammation, and apoptosis (Martins et al. 2014; Zaki et al. 2015); therefore, maybe NGAL too acts as a “double-edged sword”. Also, Hafner et al. (2013) suggested that chronic inflammation causes “pre-conditioning” against excessive acute hyper-inflammation.

Conclusions

NGAL can be used as a biomarker for predicting incidence of NAFLD in obese adolescents.

Abbreviations

ALT: Alanine transaminase; AST: Aspartate transaminase; BMI: Body mass index; CRP: C-reactive protein; HDL: High-density lipoprotein; HOMA: Homeostatic Model Assessment; IR: Insulin resistance; LDL: Low-density lipoprotein; NAFLD: Nonalcoholic fatty liver disease; NGAL: Neutrophil gelatinase-associated lipocalin; TNF- α : Tumor necrosis factor- α

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Availability of data and materials

The datasets used and/or analyzed during the current study are available from the corresponding author on reasonable request.

Authors' contributions

WA analyzed and interpreted the patients' anthropological data and contributed in writing the manuscript. HAE analyzed and interpreted the patients' biochemical data and contributed in writing the manuscript. OM was a major contributor in writing the manuscript. HAM analyzed and interpreted the patients' biochemical data. GA contributed in analyzing the patients' biochemical data. EM contributed in the laboratory determination and analysis of the patients' biochemical parameters. NA contributed in the interpretation and selection of the suitable cases for the study. MM contributed in the laboratory determination of patients' biochemical parameters. MA contributed in the laboratory determination of patients' biochemical parameters and in writing and editing the manuscript. All authors read and approved the final manuscript.

Ethics approval and consent to participate

The study protocol was approved by the Ethical Committee Board of the National Research Centre, Cairo, Egypt, with registration number 17006. An informed written consent was obtained from all participants.

Consent for publication

Not applicable

Competing interests

The authors declare that they have no competing interests.

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