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# Ameliorating effect of blueberry consumption on energy drink-induced testicular damage in rats: histological and immunohistochemical study

Turki M. Al-Shaikh<sup>1\*</sup>  and Nisreen Abdullah Rajeh<sup>2</sup>

## Abstract

**Background** Energy drinks prevent fatigue and increase physical and cognitive performance; however, they also inflict toxic effects on the body. Blueberry (BB) possesses antioxidant and anti-inflammatory properties. The present study aimed to assess the possible therapeutic effects of BB on testicular damage in adult male Wistar albino rats induced by administering the energy drink Code Red.

**Results** Thirty adult rats were used in the study, divided into five groups; Group 1 (Control), where rats were fed on distilled water and basal rodent diet only. The other four groups received different energy drink Code Red doses for 8 weeks and BB supplementation for another 6 weeks. Administration of low and high doses of Code Red induced a decline in serum levels of testosterone and antioxidant enzymes like superoxide dismutase (SOD), and glutathione (GSH), while malondialdehyde (MDA) was significantly increased relative to controls. A low dose of Code Red led to sporadic and scattered appearance of seminiferous tubules with loss of spermatogenic germ cells and marked degeneration of interstitial cells. A high dose of Code Red exhibited increased degenerative changes in the tubules with highly congested thick vessels in the interstitial tissue. Also, testis from rats consuming either low or high doses of Code Red showed increased caspase-3 immunostaining in seminiferous tubules with early degeneration features. However, the deleterious effect of the administration of Code Red was remarkably ameliorated with the supplementation of BB. A reversal in the mutilative effect of Code Red was observed where with BB supplementation, the histopathology of the testis displayed recovery of most of the seminiferous tubules to normal structure. BB administration in both groups also showed negative or mild immunostaining for caspase-3.

**Conclusions** Oral exposure of rats to Code Red produced noticeable testicular damage, especially in high doses, probably due to increased oxidative stress and inflammation. Blueberry administration exhibited therapeutic effects through its anti-inflammatory and antioxidative properties.

**Keywords** Energy drinks, Blueberry, Histopathology, Wistar albino rats, Testis, Oxidative stress, Caspase-3

\*Correspondence:

Turki M. Al-Shaikh  
tmaalshaikh@uj.edu.sa

Full list of author information is available at the end of the article



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## Background

Energy drinks (EDs) are non-alcoholic drinks composed of high concentrations of caffeine, sugar, amino acids, vitamins, and plant or herbal derivatives, ultimately enhancing the consumer's energy levels (Nadeem et al., 2021). The first beverage of this kind was produced in Austria in 1987, and 10 years later, comparable drinks with high amounts of caffeine and taurine were introduced in the United States of America (Malinauskas et al., 2007). Energy drink consumption is primarily prevalent among young people and adolescents (Heckman et al., 2010). Al-Hazzaa et al. (2011), in a study, showed that 16.3% of male teenagers (14–19 years) in the three major Saudi cities, viz. Riyadh, Jeddah, and Al Khobar consumed energy drinks more than three times a week (Al-Hazzaa et al., 2011). Another study from Saudi Arabia established that 40% of consumers consumed more than three cans per week, with more than half of consumers being youngsters (13–35 years) (Elsoadaa et al., 2016). Local energy drink manufacturers in Saudi Arabia include Boom, Bugzi, and Code Red. However, Code Red was the most preferred energy drink among the participating youngsters (Alrasheedi, 2016).

Although energy drinks proclaim to boost physical and mental performance, they have questionable and transient benefits. Caffeine (1,3,7-trimethylxanthine) is a major ingredient in energy drinks that stimulates cardiovascular and neurological systems (Burke, 2008). In addition to poisoning by caffeine, energy drink consumption can cause seizures, strokes, and gonadotoxic effects (Dias et al., 2015; Subaiea et al., 2019; Worrall et al., 2005). ED consumption can also lead to epithelial hydrops, intertubular bleeding, and inflammatory alterations in the renal tubules (Jahedi et al., 2014). Rats administered with ED exhibited severe necrotic alterations, vacuolization, and nuclear karyolysis, in addition to clogged blood arteries and perivascular infiltration in the pancreatic cells. In another study, ED-treated rats showed extensive glandular atrophy, gastric ulcers, and vascular congestion, accompanied by injury to the physiological barrier separating the gastric lumen and underlying mucosa (Khayyat et al., 2014). A significant rise in the incidence of male infertility has been observed worldwide (Okonofua et al., 2022), and thus necessitates the need to study this issue. Earlier reports demonstrate that ingested caffeine can cross the blood-testis barrier, yet our knowledge about the inimical effect of ED on the reproductive system is limited. Data from *in vitro* experiments demonstrate that the effect of caffeine on human sperm motility, number, and shape varied with dosage. Males who consumed one or two cups of coffee each day had more active and dense sperms compared to those who did not. However, sperm motility and density decreased in males whose

consumption was more than two cups of coffee daily (Dlugosz & Bracken, 1992).

Blueberries (BB) plants belong to the genus *Vaccinium* (*Vaccinium corymbosum*) and Ericaceae family (Nardi et al., 2016). Proanthocyanidins, flavonols, anthocyanins, and phenolic acids like ferulic, p-Coumaric, chlorogenic, and caffeic acid are the antioxidant that are found to be abundant in blueberries (Yuan et al., 2016). Due to the presence of several antioxidant molecules, particularly anthocyanins, blueberries possess several therapeutic health benefits (Zhou et al., 2020). Zepeda et al. reported that the administration of blueberries resulted in lowered lipid peroxidation, reduced apoptosis, and elevated glutathione reductase (GR) and superoxide dismutase (SOD) levels in the testis of the rats under hypobaric hypoxia conditions (Zepeda et al., 2012). The widespread high consumption level of energy drinks by people necessitates a detailed investigation regarding the harmful impact on health and their potential side effects. Here, in this study, we aimed to assess the possible therapeutic and ameliorating effects of blueberry administration on the Code Red energy drink-induced toxic and harmful effects on the testis of Wistar albino rats.

## Methods

### Preparation of blueberry extract

Blueberry powder was purchased from Xi'an Pincredit Bio-tech Co., Ltd, Xian, China. The blueberry extract was prepared using the reported protocol (Zheng et al., 2013), which was used to administer the experimental Wistar rats. Briefly, the blueberry powder was frozen at 5–10 °C for 4 h and then sliced and loaded onto a plate. Then it was subsequently frozen at –30 °C for 4 h and freeze-dried at 65–85 °C for 20 h. The foreign body was sorted from the material, followed by grinding at 20 °C, with a relative humidity of 35%. The resultant dry extract was stored at 4 °C until further use.

### Code Red energy drink

The energy drink, brand name “Code Red” was used in this study, and it was obtained from a local store in Jeddah, Saudi Arabia.

### Experimental animals

Thirty adult male Wistar albino rats aged 90–120 days and weighing 200–250 g were used in this study. The rats were maintained in the animal house facility of the King Fahd Medical Research Center, King Abdulaziz University (KAU), Jeddah, Saudi Arabia. The rats were maintained in the facility following the recommendations laid by the International Laboratory Animal Use and Care throughout the experiment [National Research Council (US), 2011]. King Abdulaziz University Research Ethics

Committee approved the study protocols and ethics. All the experiments were performed at the King Fahd Medical Research Center, KAU, Jeddah, Saudi Arabia. The animals were kept in a standard cage (6 rats per cage) at an ambient temperature of  $21 \pm 1$  °C with a 12 h light–dark cycle. The animals were provided unrestricted access to water *ad libitum* and a chow diet.

### Study design

The rats were housed under standard laboratory conditions. They were fed a normal commercial chow diet and water *ad libitum* for 1 week for acclimatization before the start of the experiment. Any rats that showed abnormal behaviors were excluded from the study. Later, the rats were randomly divided into 5 groups (6 rats per group), and each group was housed in separate cages. Group 1 (Control) rats were only fed with distilled water and a basal diet. Group 2 (Low dose Code Red) received a small dose of Code Red (0.72 ml/100 g/day) for 8 weeks. Group 3 (High dose Code Red) received a high dose of Code Red (1.44 ml/100 g/day) for 8 weeks (Backer & Baeissa, 2014). Group 4 (Low dose Code Red + Blueberry) received a low dose of Code Red for 8 weeks, followed by blueberry (250 mg/kg/day) extracts for 6 weeks. Group 5 (High dose Code Red + Blueberry) received a high dose of Code Red for 8 weeks, followed by blueberry (250 mg/kg/day) extracts for 6 weeks (Larrosa et al., 2010). All treatments were given orally via gastric gavage. The low dose of energy drink (0.72 ml/100 g/day) was determined considering that an adult intakes two cans of energy drink, or 500 ml, each day with the assumption that the average weight of an adult is 70 kg. A high dose (1.44 ml/100 g/day) for the treatment was determined based on the consideration that the maximum quantity of energy drink allowed for intake by an adult is four cans, or 1000 ml each day, with an average weight of 70 kg (Backer & Baeissa, 2014). The total body weights of rats were recorded weekly, while the testis weight was determined at the end of the experiment.

### Sample collection

At the end of experiment, experimental rats were subjected to overnight fasting (12 h), and their blood samples were obtained from the retro-orbital veins into plain dry tubes. The samples were stored at room temperature for 15 min and then centrifuged for 10 min at  $3000 \times g$  to separate serum. The serum was aspirated and transferred into dry tubes using a Pasteur pipette and then frozen at  $-20$  °C until further analysis. Testosterone hormone and oxidative stress markers such as superoxide dismutase (SOD), glutathione (GSH), and malondialdehyde (MDA) were estimated in the central laboratory using

commercially available kits according to the manufacturer's protocol.

### Histology examination

After the withdrawal of blood samples, rats of all groups were euthanized by cervical dislocation under deep ether anesthesia. The abdomen and pelvis were opened, then both testes were excised and washed with saline. The testes were weighed, opened, fixed immediately in 10% formalin, and prepared for microscopic study in the histopathology laboratory in KAUH. For microscopic evaluation, paraffin sections of 5  $\mu$ m thickness were stained with hematoxylin and eosin (H&E).

### Caspase-3 staining

The testes sections mounted on positive slides were immersed in 10 mM sodium citrate buffer. The endogenous peroxidase activity in the tissue samples was blocked by treatment with hydrogen peroxide, followed by the addition of an anti-caspase 3 blocking solution to the testis tissues, respectively.

### Statistical analysis

The results obtained from the experiments were analyzed using IBM SPSS Statistics software for Windows, version 23 (IBM SPSS, IBM Corp., Armonk, N.Y., USA). The data obtained in the study are presented as mean  $\pm$  standard deviation (SD). One-way ANOVA (analysis of variance) was performed for the statistical comparisons, and Tukey's test was used to calculate the significance between groups. The  $P$ -value  $< 0.05$  was considered statistically significant.

## Results

### Effect of ED on testosterone levels and oxidative stress

The data from Table 1 showed that the serum testosterone levels were significantly decreased in experimental group 2 (Low dose Code Red), group 3 (High dose Code Red), and group 4 (Low dose Code Red + Blueberry) compared to the control group ( $P < 0.0001$ ,  $P < 0.0001$  and  $P = 0.013$ , respectively) (Table 1). Also, the levels of serum testosterone dropped significantly further in group 3 (High dose Code Red) compared to group 2 (Low dose Code Red) ( $P = 0.005$ ), depicting that the increased consumption of ED can have a more devastating effect on the testes (Table 1). Overall, the results indicated the adverse effects of continued consumption of low and high doses of ED for 8 weeks on the testis of rats. Thus, regular consumption of either low or high amounts of ED can have detrimental effects on the testes, thereby increasing the risk of male fertility.

Meanwhile, with the administration of blueberry for 6 weeks, serum levels of testosterone were found to be

**Table 1** Comparison of testosterone levels between different experimental groups

Parameters	Groups				
	Group 1 (control)	Group 2 (low dose code red)	Group 3 (high dose code red)	Group 4 (low dose code red + blueberry)	Group 5 (high dose code red + blueberry)
Testosterone (ng/dl)	464.50 ± 47.33	184.83 ± 3.66	134.17 ± 17.51	422.29 ± 33.05	434.83 ± 19.43
Significance	–	<sup>1</sup> P = 0.0001	<sup>1</sup> P = 0.0001; <sup>2</sup> P = 0.005	<sup>1</sup> P = 0.013; <sup>2</sup> P = 0.0001	<sup>1</sup> P = 0.084; <sup>3</sup> P = 0.0001; <sup>4</sup> P = 0.437

Data are presented as mean ± SD derived from six samples obtained from the six rats in each group. The statistical significance (*P* value) between different groups is calculated, and the *P* < 0.05 are considered statistically significant

<sup>1</sup> *P*: significance versus group 1 (Control); <sup>2</sup> *P*: significance versus group 2 (Low dose Code Red); <sup>3</sup> *P*: significance versus group 3 (High dose Code Red); <sup>4</sup> *P*: significance versus Group 4 (Low dose Code Red + Blueberry)

significantly increased in rats from group 4 (Low dose Code Red + Blueberry) compared to group 2 (Low dose Code Red) (*P* < 0.0001) and also in group 5 (High dose Code Red + Blueberry) compared to group 3 (High dose Code Red) (*P* < 0.0001) (Table 1). The BB administration in groups 4 and 5 showed restoration of the normal serum testosterone levels as in group 1 (Table 1). These results suggest the ameliorating and therapeutic effects of blueberry on the testicular functions against the damaging effects of energy drinks.

Further, to assess the impact of energy drinks on the reactive oxygen species (ROS) levels of the experimental rats, we measured the levels of oxidative stress markers (MDA) and antioxidants (SOD and GSH) in the blood serum, as shown in Table 2. Serum levels of MDA were significantly increased in groups 2, 3, 4, and 5 compared with control (*P* < 0.0001, *P* < 0.0001, *P* < 0.0001, and *P* = 0.027, respectively); in group 3 (High dose Code Red) compared with group 2 (Low dose Code Red) (*P* < 0.0001); meanwhile, it was significantly decreased in group 5 (High dose

Code Red + Blueberry) compared with groups 3 and 4 (*P* < 0.0001 for both) (Table 2). Serum levels of SOD were significantly decreased in groups 2 and 3 versus controls (*P* < 0.0001, for both); in group 3 compared with group 2 (*P* < 0.0001); meanwhile, it was significantly increased in group 4 compared with groups 2 (*P* < 0.0001) and in group 5 compared with group 3 (*P* < 0.0001) (Table 2). Serum levels of GSH were significantly decreased in groups 2 and 3 as compared to control (*P* < 0.0001, for both); in group 3 compared with group 2 (*P* < 0.0001); meanwhile, it was significantly increased in group 4 compared with control and group 2 (*P* = 0.023; *P* < 0.0001, respectively) and in group 5 as compared to control and group 3 (*P* = 0.008; *P* < 0.0001, respectively) (Table 2). These results indicated that the administration of low and high doses of energy drinks for 8 weeks increased oxidative stress and decreased anti-oxidants and that improvement was observed after the administration of blueberry for a period of 6 weeks (Table 2).

**Table 2** Comparison of oxidative stress levels between different experimental groups

Parameters	Groups				
	Group 1 (control)	Group 2 (low dose code red)	Group 3 (high dose code red)	Group 4 (low dose code red + blueberry)	Group 5 High dose code red + blueberry)
MDA (nmol/ml)	0.24 ± 0.06	0.75 ± 0.11	1.02 ± 0.06	0.75 ± 0.19	0.41 ± 0.14
Significance	–	<sup>1</sup> P < 0.0001	<sup>1</sup> P < 0.0001; <sup>2</sup> P < 0.001	<sup>1</sup> P = 0.0001; <sup>2</sup> P = 0.915	<sup>1</sup> P = 0.027; <sup>3</sup> P < 0.0001; <sup>4</sup> P < 0.0001
SOD (U/ml)	176.67 ± 8.24	147.67 ± 7.61	105.3 ± 9.85	181.29 ± 9.21	179.00 ± 14.44
Significance	–	<sup>1</sup> P < 0.0001	<sup>1</sup> P < 0.0001; <sup>2</sup> P < 0.0001	<sup>1</sup> P = 0.420; <sup>2</sup> P < 0.0001	<sup>1</sup> P = 0.693; <sup>3</sup> P < 0.0001; <sup>4</sup> P = 0.688
GSH (ng/ml)	16.03 ± 1.61	8.20 ± 1.09	3.43 ± 1.25	18.53 ± 2.47	19.10 ± 2.30
Significance	–	<sup>1</sup> P < 0.0001	<sup>1</sup> P < 0.0001; <sup>2</sup> P < 0.0001	<sup>1</sup> P = 0.023; <sup>2</sup> P < 0.0001	<sup>1</sup> P = 0.008; <sup>3</sup> P < 0.0001; <sup>4</sup> P = 0.585

Data are shown as mean ± SD derived from six blood samples obtained from the six rats in each group. The statistical significance (*P* value) between different groups is calculated, and the *P* < 0.05 are considered statistically significant

<sup>1</sup> *P*: significance versus group 1 (Control); <sup>2</sup> *P*: significance versus group 2 (Low dose Code Red); <sup>3</sup> *P*: significance versus group 3 (High dose Code Red); <sup>4</sup> *P*: significance versus Group 4 (Low dose Code Red + Blueberry)

SOD: superoxide dismutase, GSH: glutathione, MDA: malondialdehyde



## Histological monitoring of the effect of ED on testes

### H&E stain

In the present study, it was observed that consuming energy drinks (Code Red) by albino rats resulted in drastic changes in their testis compared to that of the control animals, whose testis showed normal seminiferous tubules with regular outlines, intact complete germ cell layers thickness, with most showing mature spermatozoa, besides the appearance of interstitial cells with its normal population and histological features and appearance (Fig. 1). Meanwhile, the rats consuming energy drinks (Code Red) showed marked histological alterations that were dose-dependent. Low dosage Code Red treatment led to the sporadic scattered appearance of seminiferous tubules with marked degeneration and loss of spermatogenic germ cells series (dividing stages), sparing only basal spermatogonia (stem precursor cells) (Fig. 1). Interstitial cells looked sparse and degenerated. High dose Code Red group showed increase in the number of tubules exhibiting similar degenerative changes in addition to highly congested thick vessels in the interstitial tissue.

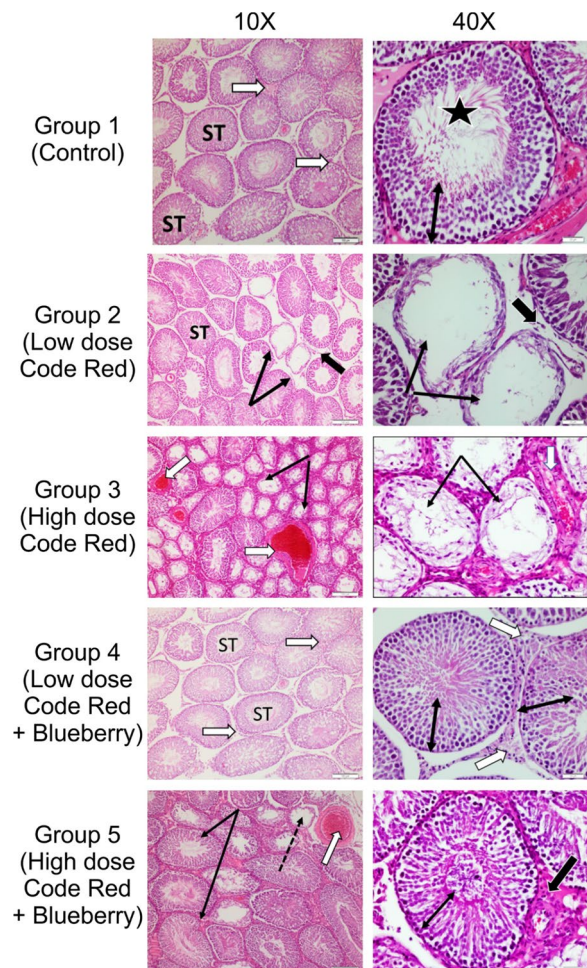
Administration of Code Red, followed by blueberry (*Vaccinium corymbosum L.*) extract for 6 weeks, showed markedly amelioration effects, such as histological changes where most seminiferous tubules showed nearly normal appearance with more impact on low energy drink group. Interstitial tissue (cells and blood vessels) of the treated group restored their normal appearance compared to the non-treated groups (Fig. 1).

### Immunohistochemistry staining for the apoptotic marker (caspase-3)

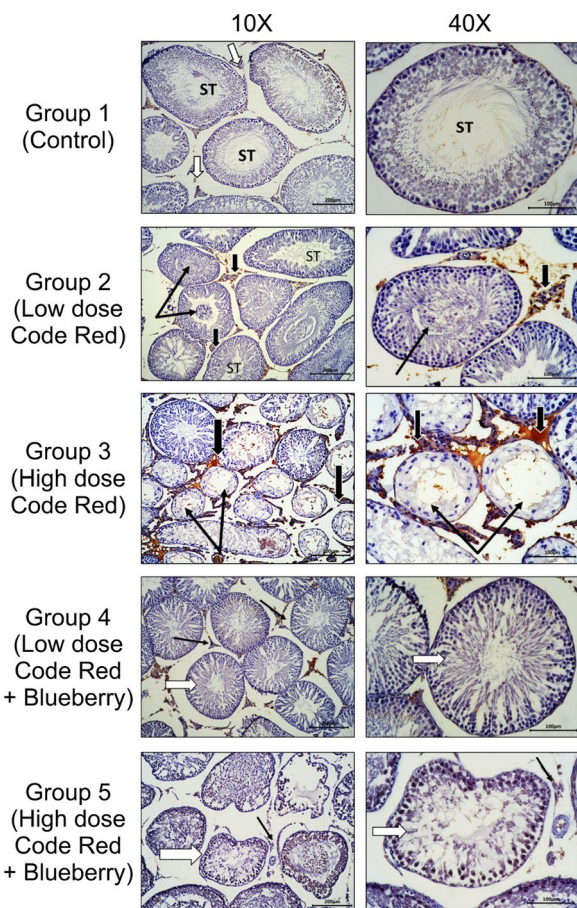
Figure 2 showed negative or sporadic positive immunostaining for caspase-3 (the apoptotic marker) in the control testis (in normally degenerated cells). On the other hand, testis from rats consuming either low or high doses of Code Red energy drink showed increased caspase-3 immunostaining in seminiferous tubules with early degeneration features, while negative impact in the seminiferous tubules exhibited loss (necrosis) of germ cells (Fig. 2). Administration of blueberry extract to both groups showed negative or mild immunostaining for caspase-3, which was similar to the control group (Fig. 2).

## Discussion

The testosterone hormone secreted by testicular interstitial Leydig cells regulates gene expression related to sexual function, which plays a vital role in male reproductive



**Fig. 1** Paraffin sections from adult Wistar albino rat testis stained by H&E and photographed at low and high powers ( $\times 100$  bar = 200  $\mu$ m and  $\times 400$  bar = 50  $\mu$ m, respectively). Group 1 (Control): Normal control with seminiferous tubules (ST) showing regular outlines and fully-thickened intact germ cell layers (double heads arrow) with mature sperm tails extending to the lumen (star). Interstitial cells show normal morphology (white arrows). Group 2 (Low dose Code Red): shows sporadically scattered tubules with marked degeneration and loss of germ cell layers (thin black arrows). Interstitial cells appear sparse and degenerated (thick black arrow). Group 3 (High dose Code Red): shows marked interstitial vascular congestion (white arrows), numerous tubules with degenerated germ cells (black arrows), and thickened fibrous interstitial tissue (white stars). Group 4 (Low dose Code Red + Blueberry): shows marked protection against Code Red-induced trophic changes on spermatogenic cells. Group 5 (High dose Code Red + Blueberry): shows a decrease in the degeneration of seminiferous tubules (black arrows), with most of the tubules having near fully-thickened germ cell layers (black arrows and double heads arrows). A few tubules show degenerated cells (dotted arrows) with nearby congested blood vessels (thick white arrow). Also, the interstitial tissue shows thickened blood vessels and degenerated cells (thick black arrow)



**Fig. 2** Sections from rat testis immunostained for caspase-3 (apoptotic marker) and photographed at  $\times 100$  bar = 200  $\mu\text{m}$  and  $\times 200$  bars = 100  $\mu\text{m}$ . Group 1 (Control): with negative immunostaining for caspase-3 in seminiferous tubules (ST) except for mild reaction in interstitial tissue (white arrows). Group 2 (Low dose Code Red): showing desquamated germ cells within ST lumina (thin black arrows) and increased immunostaining for caspase-3 in interstitial cells (thick black arrow). Group 3 (High dose Code Red): showing marked degenerative changes in most ST (Black arrows) and a marked increase in immunostaining for caspase-3 in the degenerated germ cells (black arrows) as well as the interstitial cells (thick black arrows). Group 4 (Low dose Code Red + Blueberry): showing a marked decrease in caspase-3 immunostaining in the interstitial cells (arrows) as well as germ cells (white arrows). Group 5 (High dose Code Red + Blueberry) showed decreased caspase-3 immunoexpression, although some seminiferous tubules still showed deformity or germ cell desquamation (white arrows)

ability. This study demonstrated that oral administration of the energy drink Code Red in low and high doses to rats for 8 weeks led to a considerable decline in serum testosterone levels versus the control group that was dose-dependent. These results indicated the harmful effects of consumption of low as well as a high dose of energy drinks on rat testis. Meanwhile, the administration of blueberry to Code Red treated groups led to an

increase in serum testosterone levels versus untreated groups, indicating the beneficial effects of blueberry on the testicular functions against the damaging impact of the energy drink. The decrease in testosterone production in rats receiving energy drinks could be explained by energy drink-induced damage of the interstitial cells of Leydig that produce testosterone. Al-Eryani et al. (2018) reported a reduction in serum testosterone levels in rats treated with ED for 7 weeks. Male reproductive function is affected by various factors in vivo and in vitro, and the hypothalamic-pituitary-gonad axis is the key in vivo regulatory pathway (Drobnis & Nangia, 2017; Liu et al., 2016; Nabi et al., 2020). Testosterone cooperates with luteinizing hormone and follicle-stimulating hormone to enhance the development of spermatogenic tubules, spermatogenesis, and maturation, thus affecting male reproductive capacity (Drobnis & Nangia, 2017; Kaplan et al., 2016). Meanwhile, administration of BB leads to an improvement in stromal interstitial Leydig cell structure and function, which in turn results in improvement of testosterone serum levels as is observed in this study.

The present study revealed that the administration of low and high doses of Code Red energy drinks for 8 weeks led to elevated oxidative stress markers. The results indicate that Code Red intake led to a notable reduction in serum activities of SOD and GSH and a significant elevation in MDA levels. Meanwhile, the administration of blueberry to these groups led to a marked reduction in the MDA levels and elevated SOD and GSH levels; however, these levels were still significantly changed versus the control group. In this respect, Al-Eryani (2018) revealed a significantly higher concentration of lipid peroxides in the testis of male rats that were administered energy drinks for 7 weeks (Al-Eryani et al., 2018). Also, Abdollahi et al. (2004) reported that energy drink administration induced a significant reduction in the antioxidant activity of SOD, catalase (CAT), and glutathione peroxidase (GPx) enzymes (Abdollahi et al., 2004). Together with the non-enzymatic antioxidant system, these enzymes guard cells from free radical oxidative damage. The highly reactive superoxide anions are neutralized by SOD by converting to hydrogen peroxide, which GPx and CAT subsequently break down into water (Sharma & Sangha, 2014). The considerable decrease in enzyme levels in the blood, particularly in rats given medium, low, and high doses of ED, may be caused by an ED-induced increase in superoxide radicals, which exceeds enzymes' antioxidant ability to neutralize these radicals. Studies have also shown that higher caffeine exposure causes a pro-oxidant environment in human cells, which promotes protein oxidation, whereas low amounts of caffeine do not affect the antioxidant capacity of the cells (Dias et al., 2015). Caffeine



causes a decrease in the antioxidant defense system (SOD, GPx, and CAT), resulting in elevated free radical activities and subsequently leading to oxidative stress, as evident by increased MDA concentration (Ekaluo et al., 2016). When blood urea nitrogen levels are dramatically increased by caffeine, xanthine oxidase is activated, stimulating the conversion of xanthine to uric acid and the production of superoxide anion and H<sub>2</sub>O<sub>2</sub>. On the other hand, several studies have shown that many ED ingredients, including taurine, ginseng, caffeine, and guarana, have antioxidant qualities (Obochi et al., 2010). Blueberries are abundant in antioxidants that possess anti-inflammatory properties. Anthocyanins, flavanols, and phenolic acid are among the antioxidant polyphenols that are known to be present in BB. Studies have shown that eating a diet rich in anthocyanins lowers the incidence of myocardial infarction in women and suggests that anthocyanins may act as an anti-inflammatory agent (Cassidy et al., 2013; Johnson et al., 2013).

In the present study, it was observed that consuming energy drinks (Code Red) by albino rats resulted in drastic changes in their testis compared to the control testis. The rats consuming energy drinks showed marked histological alterations, which were dose-dependent. Low-dose cod red treatment led to the sporadic scattered appearance of seminiferous tubules with marked degeneration and loss of spermatogenic germ cells series sparing only basal spermatogonia as well as degeneration of interstitial cells. Meanwhile, the high-dose cod red group showed more tubules exhibiting similar degenerative changes in addition to highly congested thick wall vessels in the interstitial tissue. Administration of Code Red and then blueberry extract for 6 weeks markedly ameliorated such histological changes, where most seminiferous tubules showed a nearly normal appearance with more effect on low energy drink group. These findings were in agreement with the findings of a similar study (Elshennawy & Elwafa, 2011). A recent study showed that male adult rats' testicles had serious histopathological alterations after consuming Red Bull energy drinks in excess and for an extended period of time (Ahmed, 2016). According to Al-Eryani (2018), male rats exposed to Red Bull and Power Horse energy drinks continuously for 7 weeks reported several histological alterations, including epithelial cell sloughing, atrophic changes, and a decrease in germ cell count as a result of cytotoxicity (Al-Eryani et al., 2018). Degenerative alterations in the seminiferous tubules and a decline in spermatozoa in the testis showed genotoxicity. Previous research showed that the consumption of energy drinks caused histological alterations in many organs, such as the testis (Dias et al., 2015), pancreas, fundus of the stomach (Ayuob & ElBeshbeishy, 2016), and liver (Khayyat et al., 2012).

These lesions may be caused by an unbalanced oxidant/antioxidant environment in these tissues, which results in increased oxidative stress due to the generation of tumor necrosis factor-alpha (TNF-alpha) and inducible nitric oxide synthase (iNOS). These mechanisms are triggered as a result of high content of caffeine in energy drinks (Ayuob & ElBeshbeishy, 2016). According to Ali (2015), a safe amount of caffeine is 200 to 300 mg; however, the caffeine concentration in popular energy drinks is in the 500 mg range (Ali et al., 2015). Numerous epidemiologic studies have linked caffeine to an increased incidence of infertility (Wilcox et al., 1988). A study in Sprague-Dawley rats found that administering a high dosage of caffeine-200 mg/kg of body weight adversely alters the histoarchitecture of the testis' seminiferous tubules, resulting in a significant loss of spermatogenic cells (Basssey et al., 2011). Another research using the same coffee dosage also noted histological changes such as atrophic cells with necrosis, increased spermatid degeneration, and nearly no spermatozoa (Ekaluo et al., 2014). Adult albino rats who received 30 mg/kg/day of caffeine administered by gavage for 15–38 days in a row experienced a breakdown of the germinal epithelium (Pollard & Smallshaw, 1988). Caffeine, given daily to adult male rabbits (30–60 mg/kg) for 4 weeks in a row, resulted in a reduction in the size of the seminiferous tubules and an inhibition of spermatogenesis (Ezzat & El-Gohary, 1994). In another study, 85–100% of 4–6-week-old male rats administered caffeine for 14–75 weeks showed severe bilateral testicular atrophy with aspermiogenesis or oligospermatogenesis. Reduced spermatogenesis caused by coffee in mature animals resulted in testicular atrophy (Friedman et al., 1979). Additionally, one of the effects caused was that the testicles shrink and lighten, and Leydig cells produce less testosterone (Park et al., 2015).

In our study, the expression of the cytoplasmic immunoreactivity caspase-3 was highly positive in testicular tissues of Code Red-treated groups. Administration of blueberry extract to low and high doses ED groups showed negative or mild immunostaining for caspase-3. This was in harmony with the results of a similar study (Ayuob & ElBeshbeishy, 2016), where it was reported that the administration of power horse, one of the energy drinks, to rats resulted in a strong cytoplasmic caspase-3 reaction in the pancreas and stomach.

## Conclusions

The findings of this study showed that 8 weeks of exposure to low and high doses of Code Red causes damage to the testicles and decreases blood testosterone levels in rats. A significant decrease in the levels of key antioxidant enzymes (SOD and GSH) in blood and elevation of MDA with increased caspase-3 expression in testicular tissue

clearly indicate that the deleterious effects of Code Red occur as a result of enhanced ROS generation, oxidative stress, and inflammation. Blueberries had a marked therapeutic effect against Code Red-induced testicular damage via their anti-inflammatory and antioxidant effects. Therefore, the use of Code Red with BB is recommended. These findings urge moderation and caution in the intake of Code Red and other energy drinks if animal-to-man extrapolation is considered. Therefore, the need for having appropriate public information regarding energy drinks cannot be understated.

#### Abbreviations

ED	Energy drink
BB	Blueberry
SOD	Superoxide dismutase
GSH	Glutathione
MDA	Malondialdehyde
GR	Glutathione reductase
SD	Standard deviation
ROS	Reactive oxygen species
ST	Seminiferous tubules
CAT	Catalase
GPx	Glutathione peroxidase
TNF-alpha	Tumor necrosis factor-alpha
iNOS	Inducible nitric oxide synthase
KAU	King Abdulaziz University
ST	Seminiferous tubules
H&E	Hematoxylin and eosin

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#### Author contributions

TMA conceived the idea, TMA and NAR performed the experiments, and TMA and NAR wrote the manuscript. Both authors read and approved the manuscript.

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Not applicable.

#### Availability of data and materials

This published paper contains all of the data collected throughout the research.

#### Declarations

##### Ethics approval and consent to participate

All the experiments and animal care were carried out under the guidelines of King Abdulaziz University Research Ethics Committee that approved the study protocols and ethics.

##### Consent for publication

Not applicable.

##### Competing interests

All the authors declare that there is no conflict of interest.

##### Author details

<sup>1</sup>Department of Biology, College of Science and Arts at Khulis, University of Jeddah, Jeddah, Saudi Arabia. <sup>2</sup>Department of Anatomy, Faculty of Medicine, King Abdulaziz University, Jeddah, Saudi Arabia.

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