# Human dosimetry of free ${ }^{211}$ At and meta[ ${ }^{211} \mathrm{At}$ ]astatobenzylguanidine ( ${ }^{211} \mathrm{At}-\mathrm{MABG}$ ) estimated using preclinical biodistribution from normal mice 

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#### Abstract

Background: ${ }^{211}$ At is one of the ideal nuclides for targeted radionuclide therapies (TRTs). Meta-[ $\left.{ }^{211} \mathrm{At}\right]$ astatobenzylguanidine ( $\left.{ }^{211} \mathrm{At}-\mathrm{MABG}\right)$ has been proposed for the treatment of pheochromocytoma. To effectively use these radiopharmaceuticals, dosimetry must be performed. It is important to determine the absorbed doses of free ${ }^{211} \mathrm{At}$ and ${ }^{211} \mathrm{At}-\mathrm{MABG}$ to determine the organs that may be at risk when using TRTs. The aim of this study was to estimate human dosimetry from preclinical biodistribution of free ${ }^{211}$ At and ${ }^{211}$ At-MABG in various organs in normal mice. Methods: Male C57BL/ 6 N mice were administered 0.13 MBq of free ${ }^{211}$ At or 0.20 MBq of ${ }^{211} \mathrm{At}$-MABG by tail-vein injection. The mice were sacrificed at 5 min , and at $1,3,6$, and 24 h after the injection ( $n=5$ for each group). The percentage of injected activity per mass in organs and blood (\% $\mathrm{I} \mathrm{A} / \mathrm{g}$ ) was determined. The human absorbed doses of free ${ }^{211}$ At and ${ }^{211}$ At-MABG were calculated using the Organ Level INternal Dose Assessment/EXponential Modeling (OLINDA/EXM) version 2.0 and IDAC-Dose 2.1. Results: High uptake of free ${ }^{211}$ At was observed in the lungs, spleen, salivary glands, stomach, and thyroid. The absorbed doses of free ${ }^{211}$ At in the thyroid and several tissues were higher than those of ${ }^{211} \mathrm{At}$-MABG. The absorbed doses of ${ }^{211} \mathrm{At}-\mathrm{MABG}$ in the adrenal glands, heart wall, and liver were higher than those of free ${ }^{211} \mathrm{At}$. Conclusions: The absorbed doses of ${ }^{211}$ At-MABG in organs expressing the norepinephrine transporter were higher than those of free ${ }^{211}$ At. In addition, the biodistribution of free ${ }^{211}$ At was different from that of ${ }^{211} \mathrm{At}-\mathrm{MABG}$. The absorbed dose of free ${ }^{211}$ At may help predict the organs potentially at risk during TRTs using ${ }^{211}$ AtMABG due to deastatination.


Keywords: Alpha-emitter, Meta-[ ${ }^{211}$ At]astatobenzylguanidine, Dosimetry, Radionuclide therapy

## Background

A new generation of targeted radionuclide therapies (TRTs) involves the use of alphaparticles. One of the TRTs that uses alpha particles is called targeted alpha therapy (TAT). TAT is exclusively cytotoxic and not affected by many of the limitations associated with conventional chemotherapy and radionuclide therapy using electrons. The alpha particles have high-energy deposition [linear energy transfer (LET)] and a limited range in tissue, resulting in strong therapeutic effects with minimal adverse effects on normal organs [1,2]. Studies on the therapeutic application of alpha-emitters have been carried out using various nuclides [1, 3-9].

Pheochromocytoma originates from the adrenal medulla and sympathetic ganglia, and approximately $10-15 \%$ of patients with pheochromocytoma have systemic metastasis, progressing into malignant pheochromocytoma [10, 11]. Meta- $\left[{ }^{131}\right.$ I]iodobenzylguanidine ( ${ }^{131}$ I-MIBG) is a radiopharmaceutical for the systemic treatment of patients with metastatic pheochromocytoma and paraganglioma. ${ }^{131}$ I-MIBG, an analog of guanethidine, concentrates in adrenergic tissue by the same mechanism as that of norepinephrine through the norepinephrine transporter (NET) [12]. Treatment with ${ }^{131}$ I-MIBG has shown limited efficacy even when administered at a high radioactivity such as more than 7.4 GBq [10]. Given the limited treatment approaches currently available for patients with metastatic pheochromocytoma, new effective approaches are being sought out. The alpha-emitting radiopharmaceutical meta- $\left[{ }^{211} \mathrm{At}\right]$ astatobenzylguanidine $\left({ }^{211} \mathrm{At}-\mathrm{MABG}\right)$ is an alternative to ${ }^{131}$ I-MIBG for the treatment of malignant pheochromocytoma, because the uptake mechanisms of these radiopharmaceuticals are similar. Astatine is the heaviest element of the halogen group, which also contains iodine; therefore, both astatine and iodine share the same chemical properties. Theoretically, ${ }^{211}$ At-MABG should be more effective and have fewer side effects than ${ }^{131}$ I-MIBG, because alpha-particles have a high LET and a very short range in tissues compared with electrons [1].

Since the decay pathway is $100 \%$ alpha-particle emission ( 5.87 and 7.45 MeV in $42 \%$ and $58 \%$ of the decays, respectively) during the decay of ${ }^{211} \mathrm{At}$ at a half-life of $7.2 \mathrm{~h},{ }^{211} \mathrm{At}$ is one of the nuclides available for TAT [13, 14]. However, the biochemical properties of ${ }^{211} \mathrm{At}$ and ${ }^{211}$ At-labeled compounds have not been clarified owing to the absence of stable astatine isotopes. In addition, the ${ }^{211}$ At-labeled compounds for TAT can be problematic owing to their rapid deacidification in vivo [15, 16]. Free ${ }^{211}$ At generated by deastatination may accumulate in specific tissues through redistribution. Therefore, it is important to determine the human dosimetry of free ${ }^{211}$ At and ${ }^{211}$ At-labeled compounds in normal tissues to predict the organ potentially at risk when using radiopharmaceuticals for TAT.
Several studies based on planar images of ${ }^{211}$ At obtained using a gamma camera have been reported $[17,18]$. However, there are still issues with the spatial resolution of the images and the quantification of the images because of the lack of attenuation correction. Therefore, the absorbed doses in humans were assessed by measuring and extrapolating the in vivo distribution in mice.

Several types of absorbed dose calculation software have been developed for nuclear medicine. The Organ Level INternal Dose Assessment/EXponential Modeling (OLINDA/EXM) versions 1.0, 1.1 (Vanderbilt University, Nashville, TN, USA) and 2.0 (Hermes Medical Solutions, Stockholm, Sweden) were developed as dosimetry software, as reported by Stabin et al. [19, 20]. In addition, IDAC-Dose 2.1 was also created as an internal dosimetry computer program by Anderssen et al. [21].

OLINDA/EXM version 2.0 has realistic human computational phantoms that were based on the International Commission on Radiological Protection (ICRP) Publication 89, i.e., nonuniform rational B-splines voxel-based models. It also includes phantoms for the mouse, rat, and dog. IDAC-Dose 2.1 has a voxel phantom installed describing two adults that were reported in ICRP Publication 110, and the specific absorbed fraction (SAF) values are presented in ICRP Publication 133. IDAC-Dose 2.1 and OLINDA/EXM version 2.0 use radiation spectra obtained with the Medical Internal Radiation Dose (MIRD) program [22]. However, the two programs have differences in calculable absorbed doses in organs and tissues. The range of alpha-particles is short, and the SAF in organs is strongly affected by the absorbed dose calculation.

The aim of this study was to calculate and compare the absorbed doses of free ${ }^{211} \mathrm{At}$ and ${ }^{211} \mathrm{At}$-MABG in various organs in normal mice using two software programs, OLINDA/EXM and IDAC-Dose 2.1.

## Materials and methods

## Production of ${ }^{211} \mathrm{At}$ and radiosynthesis of ${ }^{211} \mathrm{At}$-MABG

${ }^{211}$ At was produced through the ${ }^{209} \mathrm{Bi}(\alpha, 2 \mathrm{n})^{211}$ At reaction using a Sumitomo multipurpose cyclotron (MP-30, Sumitomo Heavy Industries, Ltd. Japan) in the Advanced Clinical Research Center at Fukushima Medical University, Japan. A 30 MeV alpha-particle beam was degraded to $26.5 \pm 0.9 \mathrm{MeV}$ by inserting $70 \mu \mathrm{~m}$ of aluminum foil to prevent the production of ${ }^{210}$ At. The degraded beam was used to bombard the bismuth ( $99.999 \%$, Goodfellow Cambridge Ltd., Huntingdon, England) layer on an aluminum backing for 110 min with $10.9 \mathrm{e} \mu \mathrm{A} .{ }^{211}$ At was isolated from the irradiated target using the dry distillation procedure reported by Lindegren et al. [23] with slight modifications. Briefly, the target was inserted in a quartz tube placed in a tube furnace preheated to $700^{\circ} \mathrm{C}$. Under helium gas flow in a $700^{\circ} \mathrm{C}$ oven, vaporized ${ }^{211} \mathrm{At}$ was transported from the quartz tube to an externally connected PTFE tube that was immersed in dry ice/ethanol bath. The cooled ${ }^{211}$ At was trapped in the PTFE tube, eluted, and recovered using 0.5 mL of chloroform. The radioactivity of ${ }^{211}$ At was measured using a dose calibrator (CRC-25R, Capintec Inc., Ramsey, NJ, USA), which was previously calibrated by measuring a highly radioactive ${ }^{211}$ At source using a Ge detector (GEM30-70, ORTEC, Oak Ridge, TN, USA) and a dose calibrator. For the quantification of ${ }^{211}$ At radioactivity on a Ge detector, we selected 687.0 keV (gamma-ray intensity: $\operatorname{Ir}=0.261 \%)$ of gamma-ray from ${ }^{211} \mathrm{At}$, and $569.65 \mathrm{keV}\left(0.311 \%\right.$, against the decay of $\left.{ }^{211} \mathrm{At}\right)$ and $897.8 \mathrm{keV}\left(0.321 \%\right.$, against the decay of $\left.{ }^{211} \mathrm{At}\right)$ from the daughter nuclide ${ }^{211} \mathrm{Po}$. Gamma-ray spectrometry was also performed using a Ge detector to assign the radionuclides produced in the target and the recovery solution of ${ }^{211} \mathrm{At}$. The radioactivity of ${ }^{211} \mathrm{At}$ was 263.3 MBq at the end of the bombardment, and the radiochemical purity of ${ }^{211} \mathrm{At}$ was more than $99.9 \%$ at the end of recovery; there was no contamination of ${ }^{210} \mathrm{At}$. Chloroform, the recovery solvent of ${ }^{211} \mathrm{At}$, was added to $50 \mu \mathrm{~L}$ of 0.1 M NaOH aqueous solution and then removed with nitrogen gas. The remaining ${ }^{211}$ At was redissolved in 3 mL of saline and administered to mice. In this study, ${ }^{211}$ At refers to "free astatine," which likely consists not only of ${ }^{211} \mathrm{At}^{-}$, but also, to some extent, other oxidation states [24].
${ }^{211} \mathrm{At}$-MABG was prepared in accordance with a slightly modified previously published method [25, 26]. Namely, ${ }^{211}$ At in chloroform and meta-trimethylsilylbenzylguanidine hemisulfate with $N$-chlorosuccinimide were dissolved in trifluoroacetic acid and
heated at $70^{\circ} \mathrm{C}$ for 10 min . Crude ${ }^{211} \mathrm{At}-\mathrm{MABG}$ was purified with a Sep-Pak tC18 Plus Light Cartridge (Waters, Milford, MA). After washing with water, ${ }^{211}$ At-MABG was eluted with $5 \%$ ethanol aqueous solution, providing a radiochemical yield of $36.3 \%$ (decay corrected). The eluate was diluted with saline, and sodium ascorbate was added at a final concentration of $2.5 \%$. The radiochemical purity of ${ }^{211}$ At-MABG was determined using reverse-phase radio-high-performance liquid chromatography (radio-HPLC), and the value was $>98 \%$.

## Biodistribution study

The experimental procedures and care of animals were carried out with the approval of the Fukushima Medical University Institute of Animal Care and Use Committee. Normal male mice (C57BL/6 N, 9 weeks old) were administered 0.13 MBq of free ${ }^{211} \mathrm{At}$ or 0.20 MBq of ${ }^{211} \mathrm{At}-\mathrm{MABG}$ by tail-vein injection. The mice were sacrificed at 5 min , and at $1,3,6$, and 24 h after each tracer injection ( $n=5$ in each group). The radioactivities in organs (muscle, heart, lung, spleen, pancreas, white adipocyte, testis, stomach, small intestine inclusive of contents, large intestine inclusive of contents, kidneys, adrenal glands, liver, brown adipocyte, salivary gland, thyroid gland, bone, and brain) and blood were measured using a $\gamma$-counter (Wizard2 ${ }^{\circ}$, Perkin Elmer, MA, USA). The activities in the abovementioned organs and blood were determined as the percentage of injected activity per mass ( $\% \mathrm{IA} / \mathrm{g}$ ), whereas that in the thyroid gland was determined as \%IA because the gland could not be weighed accurately.

## Radiation absorbed dose calculations

The data of biodistribution in the mice were used to estimate not only the mouse radiation absorbed doses but also the human radiation absorbed doses for both free ${ }^{211} \mathrm{At}$ and ${ }^{211} \mathrm{At}-\mathrm{MABG}$. The mean radioactivity in mouse organs at 5 min , and at $1,3,6$, and 24 h ( $n=5$ in each group) was used to calculate the time-integrated activity coefficient ( $\mathrm{Bq}-\mathrm{h} / \mathrm{Bq}$ ) for each organ. The calculated \%IA/organ in mouse and human organs was fitted with an exponential function and integrated to obtain the number of disintegrations (time-integrated activity coefficient) for source organs using the OLINDA/EXM version 1.1 software. One to three exponential terms can be selected for the modeling process [19]. An indirect blood-based method using patient-based red marrow-to-blood ratio (RMBLR) and bone marrow mass was used to determine bone marrow self-dose. The red marrow cumulated activity $\left(A_{R M}\right)$ is generally determined using the following equation (1) [27, 28]:

$$
\begin{equation*}
A_{R M}=\left[A_{b l o o d}\right] \times R M B L R \times m_{R M-\text { phantom }}, \tag{1}
\end{equation*}
$$

where [ $\mathrm{A}_{\text {blood }}$ ] is the blood cumulated activity concentration obtained from serial whole-blood sampling and analysis of the resulting blood activity concentration-time curve, and $m_{R M \text {-phantom }}$ is the red marrow mass ( kg ) of the male human phantom. The RMBLR is a correction factor representing the marrow-to-blood activity concentration ratio. The RMBLR reported by previous studies was 0.36 [27, 28].

Using the percent $\mathrm{kg} / \mathrm{g}$ method [29] with mass extrapolation of 73.0 kg (ICRP 89 adult male phantom), we extrapolated data to human dosimetry. In this method, the human \%ID/organ is calculated using the following equation (2) [29, 30]:

$$
\begin{equation*}
(\% I D)_{\text {human }}=\left[\left(\frac{\% I D}{m_{\text {organ }}}\right)_{\text {animal }} \times\left(m_{T B}\right)_{\text {animal }}\right] \times\left(\frac{m_{\text {organ }}}{m_{T B}}\right)_{\text {human }} \tag{2}
\end{equation*}
$$

where $m_{\text {organ }}$ is the organ mass and $m_{\mathrm{TB}}$ is the total body mass. The human body mass and organ masses were taken from OLINDA/EXM version 2.0 for adult male phantoms. The mouse body mass and organ masses were measured. For the thyroid, a mass of 14 mg for a 25 g mouse model installed in OLINDA/EXM version 2.0 was used for calculations because the gland could not be weighed accurately by the $\mathrm{kg} / \mathrm{g}$ method. The 25 g mouse model is the closest to the average body mass $(23.3 \pm 1.2 \mathrm{~g})$ of the mice used in this experiment.
OLINDA/EXM version 2.0 and IDAC-Dose 2.1 were used for human absorbed dose calculations. For animal absorbed dose assessment, only OLINDA/EXM version 2.0 was used. The absorbed dose contribution from the daughter nuclides can also be included in the absorbed dose calculations.

## Results

## Biodistribution study

The biodistributions of free ${ }^{211} \mathrm{At}$ and ${ }^{211} \mathrm{At}$-MABG are shown in Figs. 1 and 2. The activity concentration of free ${ }^{211}$ At was higher than that of ${ }^{211}$ At-MABG in several organs, such as the stomach, lungs, spleen, and salivary gland. The stomach had a high concentration of free ${ }^{211} \mathrm{At}$ for up to 24 h , with the highest activity concentration observed after 1 h .
On the other hand, the accumulation of ${ }^{211}$ At-MABG was higher than that of free ${ }^{211}$ At in the heart and adrenal glands. The biodistributions of the two compounds at 6 $h$ post intravenous injection were specifically different in the heart ( $3.34 \pm 0.33$ vs.


Fig. 1 Biodistribution data of free ${ }^{211}$ At in mice at 5 min , and at $1,3,6$, and 24 h . Results are shown as $\% I A / g($ mean $\pm$ SD)


Fig. 2 Biodistribution data of ${ }^{211} \mathrm{At}-\mathrm{MABG}$ in mice at 5 min , and at $1,3,6$, and 24 h . Results are shown as \%IA/g (mean $\pm$ SD)
$9.92 \pm 1.33 \% \mathrm{IA} / \mathrm{g})$ and adrenal gland ( $2.52 \pm 1.06$ vs. $14.24 \pm 2.59 \% \mathrm{IA} / \mathrm{g}$ ). ${ }^{211} \mathrm{At}-\mathrm{MABG}$ showed faster clearance in each organ as well as in blood and plasma.

The thyroid gland showed the highest accumulation at 6 h after the injection of free ${ }^{211}$ At with $1.777 \%$ IA and the highest accumulation at 24 h for ${ }^{211}$ At-MABG with an uptake of $0.506 \% \mathrm{IA}$ (Fig. 3). The free ${ }^{211} \mathrm{At}$ concentration in the thyroid gland was markedly increased compared with ${ }^{211} \mathrm{At}$-MABG concentration (four times higher).

## Dosimetry

The time-integrated activity coefficients calculated using OLINDA/EXM version 1.1 for the organs are listed in Table 1 for the 25 g mouse model in OLIDA/EXM version 2.0, in Table 2 for the adult male model in OLINDA/EXM version 2.0, and in Table 3 for the adult male model in IDAC-Dose 2.1. The time-integrated activity coefficients in Tables 2 and 3 were calculated by extrapolation from the distribution data from mice. The mean absorbed doses per unit injection activity for the 25 g mouse phantom estimated using free ${ }^{211} \mathrm{At}$ and ${ }^{211} \mathrm{At}$-MABG biodistribution data are given in Table 4. The dosimetric calculations for free ${ }^{211} \mathrm{At}$ and ${ }^{211} \mathrm{At}$-MABG showed that the thyroid received the highest absorbed dose per injection activity, with free ${ }^{211} \mathrm{At}=15.1 \mathrm{~Gy} / \mathrm{MBq}$ and ${ }^{211} \mathrm{At}-\mathrm{MABG}=4.08 \mathrm{~Gy} / \mathrm{MBq}$ followed by the stomach wall in the mouse model (Table 4). Relatively higher absorbed doses in extrathyroidal tissues and organs were found in the heart, lung, and stomach wall for free ${ }^{211} \mathrm{At}$ than for ${ }^{211} \mathrm{At}$-MABG.

For the adult male human model, the mean absorbed doses per unit injection activity calculated using OLINDA/EXM version 2.0 and IDAC-Dose 2.1 were equivalent in major high-uptake organs such as the adrenal gland (634 vs. $517 \mu \mathrm{~Gy} / \mathrm{MBq}$ ), heart ( 526 vs. $443 \mu \mathrm{~Gy} / \mathrm{MBq}$ ), and salivary gland ( 458 vs. $438 \mu \mathrm{~Gy} /$ MBq) (Tables 5 and 6). On the other hand, the absorbed doses in the left and right colon ( 21.4 vs. $134 \mu \mathrm{~Gy} / \mathrm{MBq}$ ), small intestine wall ( 21.7 vs. $195 \mu \mathrm{~Gy} / \mathrm{MBq}$ ), and stomach wall ( 21.3 vs. $211 \mathrm{~Gy} / \mathrm{MBq}$ ) calculated using OLINDA/EXM version 2.0 were markedly lower than those calculated using IDAC-Dose 2.1.


Fig. 3 Thyroid gland biodistribution data of free ${ }^{211} \mathrm{At}$ and ${ }^{211} \mathrm{At}-\mathrm{MABG}$. Results are shown as \%IA (mean $\pm$ SD)

Table 1 Time-integrated activity coefficient (Bq-h/Bq) for 25 g mouse model calculated using OLINDA/EXM version 1.1 for free ${ }^{211} \mathrm{At}$ and ${ }^{211} \mathrm{At}-\mathrm{MABG}$ in the main target organs. Data are based on biodistribution in mouse organs

| Source organ | ${ }^{211} \mathrm{At}-$ Free | ${ }^{211} \mathrm{At}-\mathrm{MABG}$ |
| :--- | :--- | :--- |
| Brain | $1.17 \mathrm{E}-02$ | $3.90 \mathrm{E}-03$ |
| LLI* contents | $5.93 \mathrm{E}-02$ | $7.99 \mathrm{E}-02$ |
| Small intestine | $2.92 \mathrm{E}-01$ | $3.42 \mathrm{E}-01$ |
| Stomach contents | $2.02 \mathrm{E}-01$ | $5.89 \mathrm{E}-02$ |
| Heart contents | $2.03 \mathrm{E}-02$ | $5.50 \mathrm{E}-03$ |
| Kidney | $1.08 \mathrm{E}-01$ | $9.67 \mathrm{E}-02$ |
| Liver | $1.83 \mathrm{E}-01$ | $3.81 \mathrm{E}-01$ |
| Lung | $1.15 \mathrm{E}-01$ | $5.99 \mathrm{E}-02$ |
| Pancreas | $3.05 \mathrm{E}-02$ | $2.90 \mathrm{E}-02$ |
| Cortical bone | $3.24 \mathrm{E}-01$ | $2.02 \mathrm{E}-01$ |
| Spleen | $5.02 \mathrm{E}-02$ | $3.12 \mathrm{E}-02$ |
| Testis | $4.12 \mathrm{E}-02$ | $1.18 \mathrm{E}-02$ |
| Thyroid | $1.49 \mathrm{E}-01$ | $4.02 \mathrm{E}-02$ |
| Total body | $1.56 \mathrm{E}+00$ | $1.08 \mathrm{E}+00$ |

*LLI low large intestine

Table 2 Time-integrated activity coefficient ( $\mathrm{Bq}-\mathrm{h} / \mathrm{Bq}$ ) for input to OLINDA/EXM version 2.0 for adult male model calculated using OLINDA/EXM version 1.1 for free ${ }^{211} \mathrm{At}$ and ${ }^{211} \mathrm{At}$-MABG in the main target organs. It was calculated by extrapolation from the distribution data of mice

| Source organ | ${ }^{211} \mathrm{At}-\mathrm{Free}$ | ${ }^{211} \mathrm{At}-\mathrm{MABG}$ |
| :--- | :--- | :--- |
| Adrenal gland | $1.17 \mathrm{E}-03$ | $6.13 \mathrm{E}-03$ |
| Brain | $1.67 \mathrm{E}-02$ | $5.85 \mathrm{E}-03$ |
| Left colon | $1.20 \mathrm{E}-02$ | $1.77 \mathrm{E}-02$ |
| Small intestine | $8.72 \mathrm{E}-02$ | $1.12 \mathrm{E}-01$ |
| Stomach contents | $8.66 \mathrm{E}-02$ | $2.75 \mathrm{E}-02$ |
| Right colon | $1.20 \mathrm{E}-02$ | $1.77 \mathrm{E}-02$ |
| Heart contents | $3.23 \mathrm{E}-02$ | $8.88 \mathrm{E}-03$ |
| Heart wall | $3.62 \mathrm{E}-02$ | $1.17 \mathrm{E}-01$ |
| Kidney | $3.09 \mathrm{E}-02$ | $3.14 \mathrm{E}-02$ |
| Liver | $1.03 \mathrm{E}-01$ | $2.13 \mathrm{E}-01$ |
| Lung | $3.86 \mathrm{E}-01$ | $2.11 \mathrm{E}-01$ |
| Pancreas | $1.09 \mathrm{E}-02$ | $1.04 \mathrm{E}-02$ |
| Salivary gland | $4.64 \mathrm{E}-02$ | $2.69 \mathrm{E}-02$ |
| Red marrow | $2.56 \mathrm{E}-02$ | $6.97 \mathrm{E}-03$ |
| Cortical bone | $2.03 \mathrm{E}-01$ | $1.31 \mathrm{E}-01$ |
| Trabecular bone | $5.08 \mathrm{E}-02$ | $3.27 \mathrm{E}-02$ |
| Spleen | $3.69 \mathrm{E}-02$ | $2.73 \mathrm{E}-02$ |
| Testis | $6.04 \mathrm{E}-03$ | $1.70 \mathrm{E}-03$ |
| Thyroid | $6.69 \mathrm{E}-02$ | $1.83 \mathrm{E}-02$ |
| Total body | $1.49 \mathrm{E}+00$ | $1.03 \mathrm{E}+00$ |

## Discussion

In this study, we investigated the biodistribution of free ${ }^{211} \mathrm{At}$ in normal mice and compared it with that of ${ }^{211}$ At-MABG. We estimated the human internal radiation absorbed doses using biodistribution data from normal mice. High uptake of free ${ }^{211} \mathrm{At}$ was observed in the lung, spleen, salivary gland, stomach, and thyroid, whereas ${ }^{211}$ AtMABG was observed in the heart and adrenal gland. In normal tissues, relatively high concentrations of ${ }^{211} \mathrm{At}$ were found in the heart and adrenal gland. The low radioactivity and low retention of ${ }^{211} \mathrm{At}$-MABG in the organs where free ${ }^{211} \mathrm{At}$ accumulates (stomach, spleen, salivary gland, etc.) suggest that ${ }^{211} \mathrm{At}$-MABG was relatively stable and did not undergo ${ }^{211}$ At deastatination in the body. Absorbed dose calculations showed that the mean absorbed dose of ${ }^{211}$ At was highest in the thyroid gland.

Note that the biodistribution of free ${ }^{211}$ At showed high free ${ }^{211}$ At activity concentrations in the lung and spleen. This supports the findings of a previous study where the uptake of free ${ }^{211}$ At was high in the thyroid gland, lung, spleen, and stomach in nude mice [31]. Previous studies also demonstrated higher activity concentrations of free ${ }^{211}$ At than of radioiodine in extrathyroidal organs and tissues, suggesting that the uptake/transport of free ${ }^{211} \mathrm{At}$ is dependent on mechanisms other than the sodium iodide symporter (NIS) [3, 24, 31, 32].

Concerning the physical properties of ${ }^{211}$ At [33], the energy released from electrons and photons per decay was negligible compared with that from alpha-particles. However,

Table 3 Time-integrated activity coefficient ( $\mathrm{Bq}-\mathrm{h} / \mathrm{Bq}$ ) for input to IDAC-Dose 2.1 for adult male model calculated using OLINDA/EXM version 1.1 for free ${ }^{211} \mathrm{At}$ and ${ }^{211} \mathrm{At}$-MABG in the main target organs. It was calculated by extrapolation from the distribution data of mice

| Source organ | ${ }^{211} \mathrm{At}-$ Free | ${ }^{211} \mathrm{At}-\mathrm{MABG}$ |
| :--- | :--- | :--- |
| Adipose | $1.49 \mathrm{E}+00$ | $1.03 \mathrm{E}+00$ |
| Adrenal gland | $1.17 \mathrm{E}-03$ | $6.13 \mathrm{E}-03$ |
| Brain | $1.67 \mathrm{E}-02$ | $5.85 \mathrm{E}-03$ |
| Blood | $3.57 \mathrm{E}-01$ | $9.92 \mathrm{E}-02$ |
| Cortical bone mineral, volume | $2.03 \mathrm{E}-01$ | $1.31 \mathrm{E}-01$ |
| Heart wall | $3.62 \mathrm{E}-02$ | $1.17 \mathrm{E}-01$ |
| Kidney | $3.09 \mathrm{E}-02$ | $3.14 \mathrm{E}-02$ |
| Left colon wall | $1.20 \mathrm{E}-02$ | $1.77 \mathrm{E}-02$ |
| Liver | $1.03 \mathrm{E}-01$ | $2.13 \mathrm{E}-01$ |
| Lung | $3.86 \mathrm{E}-01$ | $2.11 \mathrm{E}-01$ |
| Pancreas | $1.09 \mathrm{E}-02$ | $1.04 \mathrm{E}-02$ |
| Red marrow | $2.56 \mathrm{E}-02$ | $6.97 \mathrm{E}-03$ |
| Right colon wall | $1.20 \mathrm{E}-02$ | $1.77 \mathrm{E}-02$ |
| Salivary gland | $4.64 \mathrm{E}-02$ | $2.69 \mathrm{E}-02$ |
| Small intestine wall | $8.72 \mathrm{E}-02$ | $1.12 \mathrm{E}-01$ |
| Spleen | $3.69 \mathrm{E}-02$ | $2.73 \mathrm{E}-02$ |
| Stomach wall | $8.66 \mathrm{E}-02$ | $2.75 \mathrm{E}-02$ |
| Testis | $6.04 \mathrm{E}-03$ | $1.70 \mathrm{E}-03$ |
| Thyroid | $6.69 \mathrm{E}-02$ | $1.83 \mathrm{E}-02$ |
| Trabecular bone mineral, volume | $5.08 \mathrm{E}-02$ | $3.27 \mathrm{E}-02$ |

Table 4 Radiation dosimetry estimates per unit administered activity [Gy/MBq] for free ${ }^{211} \mathrm{At}$ and ${ }^{211}$ At-MABG for 25 g mouse phantom in OLINDA/EXM ver. 2.0, based on mouse biodistribution data

| Target organ | ${ }^{211}$ At-Free | ${ }^{211} \mathrm{At}-\mathrm{MABG}$ |
| :---: | :---: | :---: |
| Brain | 0.037 | 0.012 |
| Large intestine | 0.240 | 0.263 |
| Small intestine | 0.336 | 0.348 |
| Stomach wall | 5.370 | 1.600 |
| Heart | 0.218 | 0.099 |
| Kidney | 0.518 | 0.463 |
| Liver | 0.153 | 0.317 |
| Lung | 1.910 | 0.994 |
| Pancreas | 0.145 | 0.138 |
| Skeleton | 0.094 | 0.065 |
| Spleen | 0.652 | 0.405 |
| Testis | 0.372 | 0.107 |
| Thyroid | 15.100 | 4.080 |
| Urinary bladder | 0.094 | 0.065 |
| Total body | 0.189 | 0.145 |

Table 5 Radiation dosimetry estimates per unit administered activity [ $\mu \mathrm{Gy} / \mathrm{MBq}$ ] from OLINDA/ EXM ver. 2.0 for free ${ }^{211}$ At and ${ }^{211}$ At-MABG for human adult male phantom. It was calculated by extrapolation from the distribution data of mice

| Target organ | $\mathbf{2 1 1}^{\text {At-Free }}$ | $\mathbf{2 1 1}^{\mathbf{A t}-\mathrm{MABG}}$ |
| :--- | :--- | :--- |
| Adrenal gland | 122.00 | 634.00 |
| Brain | 16.80 | 5.94 |
| Esophagus | 30.40 | 21.00 |
| Eyes | 29.70 | 20.60 |
| Gallbladder wall | 30.00 | 21.10 |
| Left colon | 30.40 | 21.40 |
| Small intestine | 30.60 | 21.70 |
| Stomach wall | 31.40 | 21.30 |
| Right colon | 30.10 | 21.10 |
| Rectum | 29.90 | 20.70 |
| Heart | 205.00 | 526.00 |
| Kidney | 145.00 | 147.00 |
| Liver | 83.40 | 172.00 |
| Lung | 466.00 | 255.00 |
| Pancreas | 113.00 | 108.00 |
| Prostate | 29.80 | 20.60 |
| Salivary gland | 790.00 | 458.00 |
| Red marrow | 103.00 | 59.00 |
| Skeleton | 1140.00 | 741.00 |
| Spleen | 357.00 | 264.00 |
| Testis | 250.00 | 70.40 |
| Thymus | 30.10 | 20.90 |
| Thyroid | 4840.00 | 1320.00 |
| Urinary bladder | 29.80 | 20.60 |
| Total body | 50.20 | 37.30 |
|  |  | 2 |

preclinical results should be translated to humans with caution, as the photon contribution may be greater in clinical situations.

Results from the absorbed dose calculations for the animal model showed that the thyroid received the maximum mean absorbed dose per unit injected activity of both tracers. This was expected owing to the biodistribution pattern. The mean absorbed dose in the thyroid was higher for free ${ }^{211} \mathrm{At}$ than for ${ }^{211} \mathrm{At}-\mathrm{MABG}$, which is explained by the much higher uptake of free ${ }^{211}$ At than of ${ }^{211}$ At-MABG. In the group of mice injected with ${ }^{211}$ At-MABG, ${ }^{211}$ At accumulation in the thyroid gland increased over time. This is possibly due to the deastatination of ${ }^{211} \mathrm{At}$-MABG. The adrenal gland, heart, thyroid gland, and salivary gland seem to be potential absorbed dose-limiting organs for ${ }^{211}$ At-MABG. In clinical settings, the tissue accumulation of ${ }^{211}$ At may be blocked, which would potentially reduce the mean absorbed doses in extrathyroidal tissues [3].
The uptake of ${ }^{211}$ At-MABG was higher in the heart and adrenal gland, which have higher densities of NET than in other organs and glands. The biodistribution data of ${ }^{211} \mathrm{At}$-MABG were consistent with those reported in previous studies [4, 5].

The absorbed dose in human have been calculated for two different tracers with radiopharmaceuticals using OLINDA/EXM version 2.0 and IDCA-Dose 2.1. Absorbed

Table 6 Radiation dosimetry estimates per unit administered activity [ $\mu \mathrm{Gy} / \mathrm{MBq}$ ] from IDAC-dose 2.1 for free ${ }^{211}$ At and ${ }^{211}$ At-MABG for human adult male phantom. It was calculated by extrapolation from the distribution data of mice

| Target organs | ${ }^{\mathbf{2 1 1}} \mathbf{A t}$-Free | $\mathbf{2 1 1}^{\mathbf{A t}-\mathrm{MABG}}$ |
| :--- | :--- | :--- |
| Adrenal gland | 116.00 | 517.00 |
| Brain | 20.20 | 6.80 |
| Esophagus | 21.90 | 6.50 |
| Eye lenses | 0.14 | 0.08 |
| Gallbladder wall | 4.79 | 1.94 |
| Left colon wall | 110.00 | 135.00 |
| Small intestine wall | 170.00 | 195.00 |
| Stomach wall | 667.00 | 211.00 |
| Right colon wall | 110.00 | 135.00 |
| Recto-sigmoid colon wall | 23.40 | 6.68 |
| Heart wall | 150.00 | 443.00 |
| Kidney | 131.00 | 115.00 |
| Liver | 85.70 | 137.00 |
| Lung | 31.10 | 9.24 |
| Pancreas | 110.00 | 92.40 |
| Prostate | 4.47 | 1.38 |
| Salivary gland | 758.00 | 438.00 |
| Red (active) bone marrow | 32.80 | 9.68 |
| Endosteum (bone surface) | 22.80 | 9.91 |
| Spleen | 266.00 | 182.00 |
| Testis | 241.00 | 67.80 |
| Thymus | 4.84 | 1.59 |
| Thyroid | 4160.00 | 1140.00 |
| Urinary bladder wall | 2.42 | 0.85 |
|  |  |  |

doses were calculated using the animal biodistribution data for free ${ }^{211} \mathrm{At}$ and ${ }^{211} \mathrm{At}$ MABG, and calculations with IDAC-Dose 2.1 were validated using OLINDA/EXM version 2.0 with identical results in major organs. Calculations of the two programs were based on the same computational framework, so that identical radiation exposures give the same absorbed doses independently of the situation for which they are estimated. On the other hand, the absorbed doses in the left and right colon, small intestine wall, and stomach wall calculated using OLINDA/EXM version 2.0 were quite different and markably lower than those calculated using IDAC-Dose 2.1. Although the absorbed doses in these organs were relatively low, attention should be paid to the discrepancy. IDAC-Dose 2.1 can be used to calculate the absorbed doses in 47 different organs and tissues. Therefore, it can be divided into smaller parts and calculated.
The relative biological effectiveness (RBE) of alpha-particle radiation has already been discussed in reports from the US Department of Energy [34] and the MIRD Committee [35]. However, uncertainties have also been reported for the RBE of the alpha-particle radiation [9]. Therefore, in this study, the weight factor of the alpha-particle radiation is 1 and the RBE is not considered.

It is important to consider whether a similar dosimetry can be obtained even in human studies. There are also many reports in which the results were extrapolated to
human dosimetry by the $\mathrm{kg} / \mathrm{g}$ method with reference to the results of animal experiments [36-38]. The $\mathrm{kg} / \mathrm{g}$ method is discussed in a previous report [29]. Lee et al. estimated the human-equivalent internal radiation absorbed doses of ${ }^{124}$ I-MIBG using PET/CT data in a murine xenograft model [38]. In their experiment, they showed that preclinical ${ }^{124}$ I-MIBG data can predict reasonably precise radiation dose estimates relevant to clinical situations. The results suggest the relevance of our study in mice to human dosimetry. Further studies are needed to clarify whether our results are general.

## Conclusions

${ }^{211} \mathrm{At}$-MABG is a promising radiopharmaceutical for the treatment of malignant pheochromocytoma. The distribution of ${ }^{211}$ At-MABG showed different uptakes in several organs compared with free ${ }^{211} \mathrm{At}$. It is suggested that ${ }^{211} \mathrm{At}$-MABG was relatively stable and did not undergo ${ }^{211}$ At deastatination in the body. The higher mean absorbed doses of ${ }^{211}$ At-MABG in the heart and adrenal glands, which have higher NET densities than in other organs and glands, was reasonable to characterize the radiopharmaceutical. Note that free ${ }^{211}$ At has higher mean absorbed doses in the thyroid, salivary gland, stomach, lung, and spleen than ${ }^{211}$ At-MABG. This finding may contribute to the understanding of the instability of ${ }^{211} \mathrm{At}$-labeled compounds in the body and at-risk organs. Some tissues is analyzed using IDAC-Dose 2.1, and OLINDA/EXM version 2.0 show differences in alpha-particle dosimetry. The characteristics of each program should be understood and taken into consideration when they are used.

## Supplementary information

Supplementary information accompanies this paper at https://doi.org/10.1186/s40658-020-00326-7.

Additional file 1: Table S1. Biodistribution of free ${ }^{211}$ At and ${ }^{211}$ At-MABG in normal mouse.

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## Authors' contributions

Conception and design of the study: NU. Analysis and interpretation of data: NU. Collection and assembly of data: NU, SZ, CT, SS, MA. Drafting of the article: NU, SZ, NO, MA. Critical revision of the article for important intellectual content: KW. Final approval of the article: NO, SZ, HK, KT, HI

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## Availability of data and materials

The datasets supporting the conclusions of this article are included within the article.

## Ethics approval and consent to participate

Not applicable

## Consent for publication

Not applicable

## Competing interests

The authors declare that they have no competing interests.

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## References

1. Ohshima Y, Sudo H, Watanabe S, Nagatsu K, Tsuji AB, Sakashita T, et al. Antitumor effects of radionuclide treatment using alpha-emitting meta- ${ }^{211}$ At-astato-benzylguanidine in a PC12 pheochromocytoma model. Eur J Nucl Med Mol Imaging. 2018;45:999-1010. https://doi.org/10.1007/s00259-017-3919-6.
2. McDevitt MR, Sgouros G, Finn RD, Humm JL, Jurcic JG, Larson SM, et al. Radioimmunotherapy with alpha-emitting nuclides. Eur J Nucl Med. 1998;25:1341-51. https://doi.org/10.1007/s002590050306.
3. Watabe T, Kaneda-Nakashima K, Liu Y, Shirakami Y, Ooe K, Toyoshima A, et al. Enhancement of ${ }^{211}$ At uptake via the sodium iodide symporter by the addition of ascorbic acid in targeted alpha-therapy of thyroid cancer. J Nucl Med. 2019;60:1301-7. https://doi.org/10.2967/jnumed.118.222638.
4. Vaidyanathan G, Zhao XG, Larsen RH, Zalutsky MR. 3-[ ${ }^{211}$ At]astato-4-fluorobenzylguanidine: a potential therapeutic agent with prolonged retention by neuroblastoma cells. Br J Cancer. 1997;76:226-33. https://doi.org/10.1038/bjc.1997.366.
5. Vaidyanathan G, Strickland DK, Zalutsky MR. Meta-[ ${ }^{211}$ At]astatobenzylguanidine: further evaluation of a potential therapeutic agent. Int J Cancer. 1994;57:908-13. https://doi.org/10.1002/ijc. 2910570622.
6. Kratochwil C, Schmidt K, Afshar-Oromieh A, Bruchertseifer F, Rathke H, Morgenstern A, et al. Targeted alpha therapy of mCRPC: dosimetry estimate of ${ }^{213}$ Bi-PSMA-617. Eur J Nucl Med Mol Imaging. 2018;45:31-7. https://doi.org/10.1007/ s00259-017-3817-y.
7. Parker C, Nilsson S, Heinrich D, Helle SI, O'Sullivan JM, Fossa SD, et al. Alpha emitter radium-223 and survival in metastatic prostate cancer. N Engl J Med. 2013;369:213-23. https://doi.org/10.1056/NEJMoa1213755.
8. Dizdarevic S, McCready R, Vinjamuri S. Radium-223 dichloride in prostate cancer: proof of principle for the use of targeted alpha treatment in clinical practice. Eur J Nucl Med Mol Imaging. 2020;47:192-217. https://doi.org/10.1007/ s00259-019-04475-5.
9. Kratochwil C, Bruchertseifer F, Rathke H, Bronzel M, Apostolidis C, Weichert W, et al. Targeted alpha-therapy of metastatic castration-resistant prostate cancer with ${ }^{225}$ Ac-PSMA-617: dosimetry estimate and empiric dose finding. J Nucl Med. 2017;58:1624-31. https://doi.org/10.2967/jnumed.117.191395.
10. Lenders JWM, Eisenhofer G, Mannelli M, Pacak K. Phaeochromocytoma. Lancet. 2005;366:665-75. https://doi.org/10.1016/ s0140-6736(05)67139-5.
11. Ayala-Ramirez M, Feng L, Johnson MM, Ejaz S, Habra MA, Rich T, et al. Clinical risk factors for malignancy and overall survival in patients with pheochromocytomas and sympathetic paragangliomas: primary tumor size and primary tumor location as prognostic indicators. J Clin Endocrinol Metab. 2011;96:717-25. https://doi.org/10.1210/jc.2010-1946.
12. Sisson JC, Frager MS, Valk TW, Gross MD, Swanson DP, Wieland DM, et al. Scintigraphic localization of pheochromocytoma. N Engl J Med. 1981;305:12-7. https://doi.org/10.1056/NEJM198107023050103.
13. Guerard F, Gestin JF, Brechbiel MW. Production of $\left.{ }^{[211} \mathrm{At}\right]$-astatinated radiopharmaceuticals and applications in targeted alpha-particle therapy. Cancer Biother Radiopharm. 2013;28:1-20. https://doi.org/10.1089/cbr.2012.1292.
14. Zalutsky MR, Reardon DA, Pozzi OR, Vaidyanathan G, Bigner DD. Targeted alpha-particle radiotherapy with ${ }^{211}$ At-labeled monoclonal antibodies. Nucl Med Biol. 2007;34:779-85. https://doi.org/10.1016/j.nucmedbio.2007.03.007.
15. Ayed T, Pilme J, Teze D, Bassal F, Barbet J, Cherel M, et al. ${ }^{211}$ At-labeled agents for alpha-immunotherapy: on the in vivo stability of astatine-agent bonds. Eur J Med Chem. 2016;116:156-64. https://doi.org/10.1016/j.ejmech.2016.03.082.
16. Teze D, Sergentu DC, Kalichuk V, Barbet J, Deniaud D, Galland N, et al. Targeted radionuclide therapy with astatine-211: oxidative dehalogenation of astatobenzoate conjugates. Sci Rep. 2017;7:2579. https://doi.org/10.1038/ s41598-017-02614-2.
17. Zalutsky MR, Reardon DA, Akabani G, Coleman RE, Friedman AH, Friedman HS, et al. Clinical experience with alphaparticle emitting ${ }^{211}$ At: treatment of recurrent brain tumor patients with ${ }^{211}$ At-labeled chimeric antitenascin monoclonal antibody 81C6. J Nucl Med. 2008;49:30-38. doi:10.2967/jnumed.107.046938.
18. Andersson H, Cederkrantz E, Back T, Divgi C, Elgqvist J, Himmelman J, et al. Intraperitoneal alpha-particle radioimmunotherapy of ovarian cancer patients: pharmacokinetics and dosimetry of ${ }^{211} \mathrm{At}-\mathrm{MX} 35 \mathrm{~F}\left(\mathrm{ab}^{\prime}\right) 2-$-a phase I study. J Nucl Med. 2009;50:1153-60. https://doi.org/10.2967/jnumed.109.062604.
19. Stabin MG, Sparks RB, Crowe E. OLINDA/EXM: the second-generation personal computer software for internal dose assessment in nuclear medicine. J Nucl Med. 2005;46:1023-7.
20. Stabin MG, Siegel JA. RADAR dose estimate report: a compendium of radiopharmaceutical dose estimates based on OLINDA/EXM version 2.0. J Nucl Med. 2018;59:154-60. https://doi.org/10.2967/jnumed.117.196261.
21. Andersson M, Johansson L, Eckerman K, Mattsson S. IDAC-Dose 2.1, an internal dosimetry program for diagnostic nuclear medicine based on the ICRP adult reference voxel phantoms. EJNMMI Res. 2017;7:88. https://doi.org/10.1186/ s13550-017-0339-3.
22. Ku A, Facca VJ, Cai Z, Reilly RM. Auger electrons for cancer therapy - a review. EJNMMI Radiopharm Chem. 2019;4:27. https://doi.org/10.1186/s41181-019-0075-2.
23. Lindegren S, Bäck T, Jensen HJ. Dry-distillation of astatine-211 from irradiated bismuth targets: a time-saving procedure with high recovery yields. Appl Radiat Isot. 2001;55:157-60. https://doi.org/10.1016/s0969-8043(01)00044-6.
24. Spetz J, Rudqvist N, Forssell-Aronsson E. Biodistribution and dosimetry of free ${ }^{211} \mathrm{At},{ }^{125} \mathrm{I}$ - and ${ }^{131}$ - in rats. Cancer Biother Radiopharm. 2013;28:657-64. https://doi.org/10.1089/cbr.2013.1483.
25. Vaidyanathan G, Zalutsky MR. 1-(m-[ ${ }^{[111}$ At]astatobenzyl)guanidine: synthesis via astato demetalation and preliminary in vitro and in vivo evaluation. Bioconjug Chem. 1992;3:499-503. https://doi.org/10.1021/bc00018a006.
26. Sudo H, Tsuji AB, Sugyo A, Nagatsu K, Minegishi K, Ishioka NS, et al. Preclinical evaluation of the acute radiotoxicity of the alpha-emitting molecular-targeted therapeutic agent ${ }^{211} \mathrm{At}-\mathrm{MABG}$ for the treatment of malignant pheochromocytoma in normal mice. Transl Oncol. 2019;12:879-88. https://doi.org/10.1016/j.tranon.2019.04.008.
27. Sgouros G. Bone marrow dosimetry for radioimmunotherapy: theoretical considerations. J Nucl Med. 1993;34:689-94.
28. Hindorf C, Glatting G, Chiesa C, Linden O, Flux G. Committee ED. EANM dosimetry committee guidelines for bone marrow and whole-body dosimetry. Eur J Nucl Med Mol Imaging. 2010;37:1238-50. https://doi.org/10.1007/ s00259-010-1422-4..
29. Kirschner ASIR, Beierwaltes WH. Radiation dosimetry of ${ }^{131}$ I-19-lodocholesterol: the pitfalls of using tissue concentration data-reply. J Nucl Med. 1975;16:248-9.
30. Macey DJ, Williams LE, Breitz HB, Liu A, Johnson TK, Zanzonico PB. Report No. 071 - a primer for radioimmunotherapy and radionuclide therapy (2001). American Association of Physicists in. Medicine. 2001.
31. Lundh C, Lindencrona U, Schmitt A, Nilsson M, Forssell-Aronsson E. Biodistribution of free ${ }^{211}$ At and ${ }^{125}$ I- in nude mice bearing tumors derived from anaplastic thyroid carcinoma cell lines. Cancer Biother Radiopharm. 2006;21:591-600 https://doi.org/10.1089/cbr.2006.21.591.
32. Josefsson M, Grunditz T, Ohlsson T, Ekblad E. Sodium/iodide-symporter: distribution in different mammals and role in entero-thyroid circulation of iodide. Acta Physiol Scand. 2002;175:129-37. https://doi.org/10.1046/j.1365201X.2002.00968.X.
33. Nuclear structure and decay data on-line library. National Nuclear Data Center; 2017.
34. Feinendegen LE, McClure JJ. Alpha-emitters for medical therapy: workshop of the United States department of energy: Denver, Colorado, May 30-31, 1996. Radiat Res. 1997;148. https://doi.org/10.2307/3579579.
35. Sgouros G, Roeske JC, McDevitt MR, Palm S, Allen BJ, Fisher DR, et al. MIRD pamphlet No. 22 (abridged): radiobiology and dosimetry of alpha-particle emitters for targeted radionuclide therapy. J Nucl Med. 2010;51:311-28. https://doi.org/ 10.2967/jnumed.108.058651.
36. Oxboel J, Brandt-Larsen M, Schjoeth-Eskesen C, Myschetzky R, El-Ali HH, Madsen J, et al. Comparison of two new angiogenesis PET tracers ${ }^{68} \mathrm{Ga}-$ NODAGA-E[c(RGDyK)]2 and ${ }^{64} \mathrm{Cu}-$ NODAGA-E[c(RGDyK)]2; in vivo imaging studies in human xenograft tumors. Nucl Med Biol. 2014;41:259-67. https://doi.org/10.1016/j.nucmedbio.2013.12.003.
37. Constantinescu CC, Sevrioukov E, Garcia A, Pan ML, Mukherjee J. Evaluation of $\left[{ }^{18} \mathrm{~F}\right]$ Mefway biodistribution and dosimetry based on whole-body PET imaging of mice. Mol Imaging Biol. 2013;15:222-9. https://doi.org/10.1007/s11307-012-0582-y.
38. Lee CL, Wahnishe H, Sayre GA, Cho HM, Kim HJ, Hernandez-Pampaloni M, et al. Radiation dose estimation using preclinical imaging with ${ }^{124}$--metaiodobenzylguanidine (MIBG) PET. Med Phys. 2010;37:4861-7. https://doi.org/10.1118/1. 3480965.

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