REVIEW

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The autonomic nervous system in septic shock and its role as a future therapeutic target: a narrative review

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Abstract

The autonomic nervous system (ANS) regulates the cardiovascular system. A growing body of experimental and clinical evidence confirms significant dysfunction of this regulation during sepsis and septic shock. Clinical guidelines do not currently include any evaluation of ANS function during the resuscitation phase of septic shock despite the fact that the severity and persistence of ANS dysfunction are correlated with worse clinical outcomes. In the critical care setting, the clinical use of ANS-related hemodynamic indices is currently limited to preliminary investigations trying to predict and anticipate imminent clinical deterioration. In this review, we discuss the evidence supporting the concept that, in septic shock, restoration of ANS-mediated control of the cardiovascular system or alleviation of the clinical consequences induced by its dysfunction (e.g., excessive tachycardia, etc.), may be an important therapeutic goal, in combination with traditional resuscitation targets. Recent studies, which have used standard and advanced monitoring methods and mathematical models to investigate the ANS-mediated mechanisms of physiological regulation, have shown the feasibility and importance of monitoring ANS hemodynamic indices at the bedside, based on the acquisition of simple signals, such as heart rate and arterial blood pressure fluctuations. During the early phase of septic shock, experimental and/or clinical studies have shown the efficacy of negative-chronotropic agents (i.e., betablockers or ivabradine) in controlling persistent tachycardia despite adequate resuscitation. Central α-2 agonists have been shown to prevent peripheral adrenergic receptor desensitization by reducing catecholamine exposure. Whether these new therapeutic approaches can safely improve clinical outcomes remains to be confirmed in larger clinical trials. New technological solutions are now available to non-invasively modulate ANS outflow, such as transcutaneous vagal stimulation, with initial pre-clinical studies showing promising results and paving the way for ANS modulation to be considered as a new potential therapeutic target in patients with septic shock.

Keywords: Sepsis, Septic shock, Autonomic nervous system, Autonomic dysfunction, Dysautonomia, Sympathetic overstimulation, Baroreflex, Tachycardia, Vagal stimulation, Desensitization

Introduction

Sepsis is defined as life-threatening organ dysfunction caused by a dysregulated host response to infection [1]. Although guidelines for septic shock resuscitation have

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been extensively revisited over recent years [1-4], they have not included the potential role of autonomic nervous system (ANS) dysregulation.

Several studies have documented altered sympathetic activity with the progression of circulatory shock severity, using direct measures of sympathetic activity during surgical or pharmacological interventions in animal models [5, 6], or indirect estimations of autonomic activity, such as heart rate variability (HRV), blood pressure variability (BPV), or baroreflex sensitivity, in patients/animal



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models [7-12]. ANS response can occur rapidly, over seconds to minutes, to restore homeostasis. Clinical signs of ANS dysregulation may therefore be an early warning sign of an imminent decline in cardiovascular function, occurring before any deterioration in more global markers, such as a decrease in arterial blood pressure, which may occur too late to enable preventive action to be taken. Including variables associated with ANS regulation of the cardiovascular system, along with variables traditionally displayed on bedside monitors, has already been proposed as a useful tool for anticipating potential clinical deterioration in critical care settings [13-16]. One example of this approach is the HeRO (Heart Rate Observation) monitor, which displays a score indicating the risk of an infant deteriorating with sepsis in the next 24 h to prompt earlier interventions [17]. Display of this HeRO score resulted in a 22% relative reduction in mortality in a randomized clinical trial in neonatal intensive care unit (ICU) patients [17]. Similarly, the hypotension prediction index (HPI), based on dynamic changes in the variability and complexity of variables estimated from the arterial pressure waveform, has recently been proposed [18], and clinical trials are ongoing to validate the index and evaluate its impact on outcomes.

During the resuscitation phase in septic shock, little attention has been given so far to the assessment of ANSmediated control of the cardiovascular system. However, the ANS has a key role in maintaining cardiovascular homeostasis in both physiological and pathological conditions, and its dysfunction during sepsis and septic shock has been extensively demonstrated (Table 1).

The objective of this narrative review is therefore to discuss the recent evidence, from studies using standard or advanced monitoring methods and mathematical models, to support the importance of ANS monitoring in patients with sepsis and septic shock and the potential for the ANS to become an important therapeutic target in this context.

The autonomic nervous system

The parasympathetic and sympathetic branches

The ANS is part of the central nervous system (CNS) and provides unconscious control of vital physiological functions, ensuring body homeostasis. The ANS is centrally regulated by the hypothalamus, which acts as an integrator for autonomic functions. The two efferent branches of the ANS system are the parasympathetic (PNS) and sympathetic (SNS) nervous systems, both of which are characterized by two types of neuron: pre-ganglionic and post-ganglionic fibers (Fig. 1).

The PNS is composed of several nerves, e.g., the glossopharyngeal, the vagus, and the pelvic splanchnic nerves. Unlike the SNS, all the pre- and post-ganglionic nerves of the PNS are exclusively cholinergic, activating nicotinic receptors in the synapses between pre- and post-ganglionic neurons (as for the SNS), but muscarinic receptors at the target organ level. The PNS innervates the heart through the vagus nerve until it reaches the parasympathetic cardiac ganglia lying in the fat pad on the heart's surface, from which arise very short post-ganglionic neurons towards the heart itself. Hence stimulation of the PNS and vagus nerves directly influences only the activity of the heart, with negative chronotropic and inotropic effects, but can also modify the blood pressure (BP) as a secondary effect of cardiac output (CO) regulation [19].

The pre-ganglionic SNS neurons originate from the thoracolumbar region of the spinal cord. They are all cholinergic and travel to the paravertebral sympathetic ganglia, activating nicotinic receptors in the synapses between pre- and post-ganglionic neurons. These latter are usually longer neurons, extending through the body until they reach the surface of their target organs. They are typically adrenergic, releasing norepinephrine on the target organs' adrenergic receptors. Moreover, SNS can also promote epinephrine secretion in the bloodstream by the adrenal medulla stimulation, acting as a circulating hormone.

Lifecycle of adrenergic receptors

Adrenergic receptors are characteristic only of the SNS, and can be divided into two types, α -receptors and β -receptors, which can be further divided into subtypes and subclasses [19, 20]. From the cardiovascular standpoint, α 1-receptors are mostly located on the blood vessels and when activated are responsible for vasoconstriction; β 1 (and β 2) receptors are mostly present on the heart (the β 1/ β 2 ratio in the myocardium is around 3:1 to 4:1) where their stimulation mediates positive chronotropic, dromotropic and inotropic effects. At the vessel level, β 2-receptor stimulation causes vasodilation [20]. Of note, in the CNS, presynaptic α 2-receptor activation inhibits the release of norepinephrine and thus produces a global sympatholytic action.

Adrenergic receptors undergo frequent desensitization, a process that leads to reduced receptor responsiveness after prolonged stimulation. This action represents an important physiological "feedback" mechanism to protect the receptors against overstimulation. This desensitization process occurs by means of three mechanisms that begin soon after stimulation: phosphorylation (within seconds), endocytosis (within minutes), and downregulation (within hours) [21, 22]. Cells have elaborated a complex mechanism to dampen or turn off adrenergic stimuli through phosphorylation of their adrenergic receptors, which produces functional uncoupling of the activated

Markers	Main results	Type of studies	References
Catecholamines/ neuroendocrine mediators	Endogenous catecholamine increase after a sympathetic insult (e.g., head-up tit) was larger in sepsis survivors than in non-survivors. Mortality and morbidity were higher in patients with higher and more prolonged catecholamine burden	Clinical studies	Benedict and Rose [44], Ostrowski et al. [28], Boldt et al. [27], Schmittinger et al. [29]
	In the late phase of sepsis, animals showed reduced vaso- pressin release. Animal models of sinoartic denervation and bilateral carotid neurotomy showed an increase in catecholamines and TNF-alpha release	Animal studies	Pancoto et al. [8], Shi et al. [9], Nardocci et al. [57]
Adrenergic receptors	Endotoxemia caused systemic a1-receptor downregula- tion in all organs investigated; tissue concentrations of interleukin-1ß and TNF-a were markedly increased	Animal study	Bucher et al. [47]
	Patients presenting with sepsis or septic shock had extended deficits in β-adrenergic post-receptor signal transduction	Clinical study	Bernardin et al. [48]
HRV, BPV, BRS and chemoreflex sensitivity	BRS, chemoreflex sensitivity, and almost all HRV indexes were attenuated in MOF patients. Patients in whom organ function (SOFA score) increased significantly showed an increase in mean BP value and LF power in BP time series	Clinical studies	Schmidt et al. [7], Pontet et al. [77], Carrara et al. [84]
	Reduced HRV indexes preceded sepsis-induced macro- hemodynamic alterations. HRV reduction was associ- ated with pronounced parasympathetic inhibition and a sympatho-vagal balance shift. BRS was reduced after fecal pertonitis; overall variability, LF power, LF/HF ratio of HR, and LF power of MAP were all reduced. Denervated rats had the lowest BRS and survived the shortest time after the peritonitis. Endotoxic shock induced a very rapid impairment in baroreflex function, independent of the BP level. Septic shock induced an inversion of real pulse pressure amplification; aortic time constant tau, aortic compliance and TPR were decreased; BRS and aortic time constant tau alterations were correlated	Animal studies	Pancoto et al. [8], Jarkovska et al. [78], Shi et al. [9], Shen et al. [56], Nardocci et al. [57] Radaelli et al. [5], Carrara et al. [82]
Autonomic centers	Septic shock is associated with neuronal and glial apoptosis within the autonomic centers, which is strongly associated with endothelial iNOS expression	Post mortem patient study	Sharshar et al. [55]

Table 1 Relevant clinical and pre-clinical investigations supporting autonomic dysfunction in response to sepsis and septic shock (or endotoxic shock)

BRS baroreflex sensitivity, TPR total peripheral resistance, HRV heart rate variability, MAP mean arterial pressure, LF low frequency, HF high frequency



receptors from their cognate G protein. Several enzymes can mediate β -adrenoreceptor phosphorylation: protein kinase A and protein kinase C phosphorylate active and inactive receptors, whereas the G protein-coupled receptor kinases (GRK) phosphorylate only agonist-occupied receptors. Phosphorylation by GRK induces adrenergic receptor binding to cytosolic proteins called β -arrestins, which further enhances the uncoupling of the receptors from their G-proteins, promoting receptor endocytosis and their subsequent degradation in lysosomes [23]. Downregulation of genes encoding G proteincoupled receptors also leads to reduced β -agonism efficacy. Finally, receptors can also undergo resensitization through specific phosphatases and be recycled back to the cell membrane (Fig. 2).

As sepsis is characterized by excessive activation of the SNS and increased levels of circulating endogenous catecholamines, the increased stimulation of adrenergic receptors activates all previously described mechanisms, leading to desensitization. These processes are enhanced



further by the effect of inflammatory mediators and bacterial toxins. Indeed, several studies have shown that endotoxin, tumor necrosis factor (TNF)- α , and several interleukins, all act synergistically to reduce the responsiveness of adrenergic receptors [24].

It is important to stress that the two major pathways involved in the immune response to sepsis are the hypothalamic–pituitary–adrenal axis (HPA) and the SNS. As a consequence, most immune cells express adrenoceptors (particularly $\beta 2$, but also $\alpha 1$ -adrenergic receptors) and most functional processes in the immune system can be influenced (enhanced or attenuated) by adrenergic receptor agonists or antagonists [25] (Table 2). The complex timing of catecholamine release during sepsis has not yet been clarified and achieving the correct balance between providing cardiovascular support and causing undesirable immunomodulation remains a challenge. Indeed, the beneficial effect of norepinephrine, the most frequently used vasopressor in septic shock, is regularly questioned for several reasons: (i) it may contribute to a dysregulated immune response leading to immunoparalysis in sepsis [26]; (ii) a higher burden of endogenous [27, 28] or exogenous [29] catecholamine exposure in the ICU is associated with a worse clinical outcome; (iii) adrenergic stimulation has been extensively shown to inhibit both innate [30] and adaptative [30, 31] immune cell activity in various contexts.

The short-term ANS-mediated cardiovascular regulatory mechanism

The ANS-mediated cardiovascular regulatory system involves several afferent pathways and reflex mechanisms [19]. These mechanisms provide appropriate responses to rapid changes in cardiovascular function to maintain the BP within a physiological range of values, provide adequate blood flow to privileged organs (e.g., heart, kidneys, and brain), and redistribute it to specific regions according to metabolic demand. For these reasons, these mechanisms are known as short-term reflexes (occurring over seconds to minutes) and they include the arterial or

Table 2	Mechanisms affecting the ANS th	nat provide clinically	relevant effects and may	represent potential therapeuti	c targets during
sepsis					

Implicated receptors	Main endogenous and exogenous agonists	Main clinically relevant effects of receptors agonism	Potential therapeutic targets under investigation	
β1	Norepinephrine, epinephrine, dobutamine, milrinone	Sinoatrial node and ectopic pacemaker acceleration Cardiac contractility increase Renin release by juxtaglomerular cells Potential pro-inflammatory action through macrophage activation	 Patient "decatecholaminization" from a global standpoint: Adrenergic burden decrease by reducing both duration and intensity (i.e., dose) exposure to exogenous catecholamines Partial exogenous catecholamine substitu- 	
β ₂	Norepinephrine, epinephrine, dobutamine, milrinone	Arterial and venous dilation Skeletal muscle arteriole relaxation Sinoatrial node and ectopic pacemaker acceleration Cardiac contractility increase Macrophage inhibition (anti-inflammatory and immunosuppressive effects) Neutrophil activity and endothelial adhe- sion inhibition NK cell activity inhibition T-lymphocyte activity and proliferation inhibition (N/A for the Th2 cells) Variable influences on B-lymphocytes and antibody production	 tion by adding/shifting to alternative non-adrenergic agents (e.g., vasopressin or levosimendan) 2) Myocardial oxygen consumption reduction and ventricular filling optimization by HR control: Cardioselective β₁-receptor antagonists (e.g., esmolol or landiolol) Selective bradycardic agent (ivabradine) 3) Attenuation of β-adrenergic induced immune cell inhibition: Cardioselective β₁-receptor antagonists as above (e.g., esmolol or landiolol) Less cardioselective β-blockers? 	
β_3	Norepinephrine, epinephrine, mirabegron	Variable actions on cardiac contractility Arterial and venous vasodilation Neutrophil activity and endothelial adhe- sion inhibition	- Central α -agonist agent promotion (e.g., clonidine and dexmedetomidine) to reduce peripheral α_1 -adrenergic receptor desensitization and restore vascular tone adrenergic control	
α ₁	Norepinephrine, epinephrine	Arterial and venous vasoconstriction Cardiac contractility slight increase Sodium reabsorption in renal tubules Glomerular arteriole (afferent > efferent) vasoconstriction Neutrophil activity and endothelial adhe- sion inhibition T-lymphocyte activity and proliferation inhibition		
a ₂	Norepinephrine, epinephrine	Sympatholytic effect in the CNS by presyn- aptic inhibition Coronary artery and arteriole vasoconstric- tion Sodium reabsorption in renal tubules Pro- and anti-inflammatory action on macrophages		
nACh	Acetylcholine	Macrophage activity reduction through the inflammatory reflex (spleen, liver, gut, heart)	External vagus nerve stimulation to reduce MOF and excessive pro-inflammatory reactions by the inflammatory reflex mechanism.	

 β_1 beta-1 adrenergic receptors, β_2 beta-2 adrenergic receptors, β_3 beta-3 adrenergic receptors, α_1 alpha-1 adrenergic receptors, α_2 alpha-2 adrenergic receptors, *nACh* nicotinic acetylcholine receptors, *CNS* central nervous system

cardiac baroreflex, the cardiopulmonary baroreflex, and the chemoreceptor reflex. Along with these short-term mechanisms, there are also long-term mechanisms of cardiovascular homeostasis (over minutes to hours or days), such as the renin-angiotensin system, to control blood volume and vascular tone.

The arterial baroreflex

The baroreflex is the most important short-term feedback mechanism responsible for BP regulation. It is initiated by the arterial baroreceptors, usually known as high-pressure baroreceptors, which are mainly located in the aortic arch and carotid sinuses. Mechanosensitive ion channels are present on baroreceptor nerve endings, and the influx of sodium and calcium through these channels is responsible for baroreceptor depolarization during increases in BP [32]. Neural signals from arterial baroreceptors are transmitted to the nucleus of the solitary tract (NST) in the medullary area of the brainstem through two pathways: (i) signals generated by the

carotid baroreceptors are transmitted through Hering's nerve and the glossopharyngeal nerve; (ii) signals produced by the aortic baroreceptors are transmitted through the vagus nerve. The generated neural signal is transmitted to the vasomotor center through the NST in the medullary area of the brain stem. At this level, neurons project to the medullary vasomotor center, which mediates the sympathetic and parasympathetic outflows to the heart and the circulation [19]. Thus, an increase in BP induces sympathetic center inhibition, with decreased outflow to the heart and blood vessels, and parasympathetic center stimulation, with increased vagal outflow to the heart. The immediate consequence is a change in BP by vascular tone and CO modulation. The net effects are vasodilation of the veins and the arterioles and a reduction in heart rate (HR) and contractility. A decrease in arterial BP generates the opposite effects.

Importantly, baroreceptor sensitivity is not constant but depends on the BP value; specifically, baroreceptor sensitivity is maximal for BP values near the physiological range, where even a small variation in BP induces a large reflex response. Moreover, the response rate of the baroreceptors is not constant: the more rapid the BP variation, the more rapid the receptor response, regardless of the absolute BP value [33].

The cardiopulmonary baroreflex

The cardiopulmonary baroreceptors, also known as volume receptors or low-pressure baroreceptors, are located in the cardiac atria and ventricles, in large systemic veins and in the pulmonary arteries. They behave similarly to the arterial baroreceptors as they attempt to minimize the changes in BP due to changes in blood volume, mostly contained in the veins. Indeed, changes in atrial pressure are often associated with changes in venous return, which is altered by changes in CO and blood volume. The atria contain receptors activated by the tension developed during atrial contraction and receptors activated by the stretch of the atria during atrial filling; when stimulated, they send impulses up the vagal fibers to the vagal center in the medulla. Consequently, sympathetic activity is decreased to the kidney and increased to the sinoatrial (SA) node. These changes in sympathetic activity result in increased renal blood flow, diuresis, and HR.

Cardiopulmonary receptors stimulation can also lower BP by inhibiting the vasoconstrictor center in the medulla and inhibits angiotensin, aldosterone, and vasopressin (ADH) release; interruption of the reflex pathway has the opposite effects. Changes in diuresis elicited by changes in cardiopulmonary baroreceptor activation are important in the regulation of blood volume. For example, hypovolemia enhances sympathetic vasoconstriction in the kidney and increases secretion of renin, angiotensin, aldosterone, and ADH [19]. However, compared to the changes induced by the arterial baroreflex, cardiopulmonary receptor-induced variations in HR have a much smaller effect [34].

It is important to note that the direct effect of the cardiopulmonary reflex on contractility has not yet been clarified. The Bainbridge reflex is, indeed, still a matter of debate. This reflex, in contrast with the two types of baroreflex discussed earlier, consists of a transient increase in HR in case of a rapid increase of blood volume, leading to transient tachycardia concomitant with high right atrial pressures. The Bainbridge reflex has been demonstrated in animals like dogs and rats, but is not fully understood in humans; nonetheless, some studies suggest a possible role for this reflex in cardiovascular regulation when there are large variations in venous return [35, 36].

The chemoreflex

The chemoreceptors represent another mechanism involved in the regulation of BP and respiratory activity. These receptors are located peripherally (carotid bodies and aortic arch) and centrally (respiratory center of brain medulla) and they contribute to maintaining the arterial pH and both arterial partial pressures of oxygen and carbon dioxide within appropriate physiological ranges. Different from baroreceptors, chemoreceptors respond to chemical stimuli. In particular, hypercapnia detected by central chemoreceptors or hypoxia detected by peripheral chemoreceptors leads to an increase in respiratory rate and tidal volume, mediated by the neural respiratory center. Anatomically, the chemoreceptors transmit afferent signals through the vagus nerve to the NST where the respiratory center is located. Furthermore, the respiratory center also stimulates the vasomotor center; hence the net effect is a concomitant increase in arterial BP by means of vasoconstriction, although this reflex has a limited effect compared to the arterial baroreflex [19, 37]. The chemoreceptor reflex has been widely studied and reviewed [38, 39] and can be summarized as an inhibitory feedback process that interacts closely with the baroreflex [40, 41].

ANS-mediated cardiovascular regulation in septic shock

Sympathetic overstimulation

In septic shock, cardiovascular regulatory mechanisms responsible for counteracting the hypotensive stress induced by the septic immune response are impaired. The normal compensatory response to hypotension includes increased sympathetic outflow to the heart and peripheral vessels to restore BP to normal values [42]. Several studies have demonstrated dysfunction of the sympathetic branch of the ANS in septic shock, i.e., a maladaptive response to the hypotensive and inflammatory stress, which leads to impaired autonomic control of the heart and vessels, contributing to circulatory failure. The exact pathophysiological mechanism underlying this autonomic imbalance is not yet clear, but excessive, uncontrolled or prolonged SNS activation and/or inappropriate downregulation of the PSNS are key factors [42, 43]. Overall, these alterations are generally referred to as autonomic dysfunction or dysautonomia.

Life-threatening illness, such as septic shock, is one of the most potent stimuli of the SNS and extensive sympathetic activation is documented by elevated concentrations of endogenous circulating catecholamines-i.e., plasma epinephrine and norepinephrine-during shock and persisting into the post-resuscitation phase. Persistently high plasma catecholamine concentrations have been demonstrated to be associated with increased morbidity and mortality in critically ill populations [27, 29] and, in particular, in patients with septic shock [28, 44, 45], compared to progressive normalization of concentrations. Protracted and overwhelming adrenergic stress may exceed in time and scope the beneficial short-term compensatory effects. The entity of damage depends on the vulnerability of the different organs to adrenergic overstimulation and to the presence of coexisting chronic diseases. For example, the heart, which has abundant β -adrenergic receptors, seems to be most susceptible to sympathetic overstimulation, with detrimental consequences such as impaired diastolic function, tachyarrhythmia, myocardial ischemia, vasospasm, impaired coronary microcirculation, stunning, and apoptosis [20]. Other organs are also affected by adrenergic stress, with associated consequences such as pulmonary edema (with subsequent right ventricular dysfunction/failure), increased thrombosis formation, gastrointestinal hypoperfusion, immunosuppression, increased cell energy expenditure and microvascular dysfunction [42, 43, 46].

Dysfunctions of the afferent, central and efferent ANS pathways

In addition to its overstimulation, the SNS is widely disturbed by inflammatory mediators during sepsis and septic shock, with dysfunction of afferent, central, and efferent pathways, including massive desensitization of its adrenergic receptors.

Pro-inflammatory cytokine release and overproduction of nitric oxide (NO) have been shown to downregulate adrenergic receptors [47–49], including the cardiac β 1-adrenergic receptors, which could contribute to the reduced cardiovascular responsiveness to adrenergic stimuli that is frequently observed in septic shock [50]. Downregulation of α 1-adrenergic receptor gene expression during sepsis has been demonstrated in vivo and in vitro as a result of pro-inflammatory cytokine release and was associated with circulatory failure [51].

ANS chemoreceptor dysregulation also occurs in sepsis. In vitro and animal experiments in endotoxic shock have reported that inflammatory mediators activate the chemosensitive glomus cells of the carotid body, leading to increased and excessive respiratory activity [52], explaining in part why a high respiratory rate is a hallmark of sepsis onset. In a small clinical study in patients with multiorgan failure (MOF), as a result of sepsis in two thirds, cardiac chemoreflex sensitivity was blunted in proportion to disease severity (i.e. the higher the severity score, the more the variations in arterial oxygen partial pressure were not followed by the expected variations in heart rate, as in the controls subjects) [53].

Continuous communication among all vital organs is guaranteed through neural signals mediated by the ANS, which allows constant adaptation to the different physiological and pathological conditions. In the fundamental work by Godin and Buchman [54], this communication was considered in terms of "coupled oscillators": i.e., the vital organs could be seen as biological oscillators that are coupled to one another. The overwhelming inflammatory reaction associated with sepsis disrupts this communication by initiating uncoupling of these oscillators. Progression into MOF may reflect further progressive uncoupling that becomes irreversible, whereas recovery may be related to restoration of oscillator coupling. Bacterial toxins and inflammatory sepsis mediators can alter neural reflexes and consequently cause uncoupling of these inter-organ connections [55]. However, the exact mechanism responsible for this autonomic dysfunction has not been determined, and it is not clear whether it is due to a reduction in central vasomotor activity, to altered peripheral neuroeffector transmission, or depressed end-organ responsiveness as a result of desensitization.

Several studies documented how baroreflex often plays the major driver role of this autonomic imbalance. Indeed, in animal models of septic shock, reduced baroreflex function has been associated with reduced survival times [9, 56, 57]. This sepsis-related dysfunction of BP regulation may involve the baroreflex arch at several levels, including (i) a reduction in baroreceptor sensitivity; (ii) a shift in the baroreflex set point to lower levels of BP with impaired efferent sympathetic activity; (iii) reduced responsiveness of the target organ.

A recent study on endotoxic shock induced in rats by *Escherichia coli* lipopolysaccharide (LPS) infusion, demonstrated that the impairment of the baroreflex occurred almost immediately after induction of the inflammatory reaction, without changes in BP, and persisted after the LPS infusion had been stopped [5]. The authors hypothesized a direct effect of LPS through the production of NO and reactive oxygen species (ROS) (extrapolating their hypothesis from similar observations in a carotid arteriosclerosis animal model [58]). Circulating cytokines released after LPS infusion may also play an important role since they were shown to induce inflammation of the carotid body with a consequent reduction in arterial distensibility and, therefore, of baroreceptor engagement [59].

The limited knowledge about the role of the SNS on cardiovascular system control during sepsis is partly related to the difficulty in directly assessing sympathetic activity. Indirect measures of autonomic activity, such as HRV, do not enable discrimination between compromised sympathetic outflow at the central level or at the level of peripheral neuroeffector transmission. Nevertheless, sympathetic outflow can be directly measured in animal experiments through microneurographic measurements of muscle, cardiac, or renal sympathetic nerve activity (MSNA, CSNA, and RSNA, respectively). These measures enable quantification of the centrally regulated sympathetic outflow to different peripheral organs and thus for the ANS-mediated mechanisms activated in response to sepsis and septic shock to be unraveled [60, 61]. Ramchandra et al. [61] studied the changes in regional sympathetic activity following *E. coli* infusion in conscious sheep. They reported an increase in CSNA highly correlated with the increase in HR, and a transient decrease in RSNA during the first 3 h after infusion followed by a sustained increase. Vayssettes et al. [62] also reported a strong correlation between increased RSNA and tachycardia in anesthetized rats after LPS infusion; by contrast, arterial baroreceptor denervation had a minimal effect on the RSNA increase induced by LPS. However, in healthy human volunteers, Sayk et al. [60] showed that injection of endotoxin was associated with suppressed MSNA, concomitant with an increased HR and a blunted MSNA baroreflex-mediated response. The increased HR did not change with BP modulation, indicating that HR was uncoupled from baroreflex regulation. This finding suggests that the immune response in sepsis directly suppresses sympathetic outflow to the muscle vascular bed via central nervous mechanisms, leading to blunted baroreflex sensitivity [60].

Together these studies highlight that altered baroreflex control drives the sympatho-excitation elicited by sepsis, supporting previous research that demonstrated that the sympathetic activation observed in septic shock was greater than that expected with the simple unloading of baroreceptors because of the hypotensive stress [63, 64].

Inflammatory reflex

The term "inflammatory reflex"-or sometimes "cholinergic anti-inflammatory response"-highlights the increasing awareness that the PNS reflexly regulates the inflammatory response in real-time by inhibiting tissue macrophage activation [65, 66], just as it controls HR and other vital functions. Briefly, efferent signals in the vagus nerve lead to acetylcholine release in target organs of the reticuloendothelial system, such as the liver, heart, spleen, and gastrointestinal tract. Inside these organs, acetylcholine interacts with its nicotinic receptors on tissue macrophages to inhibit further release of TNF, interleukin (IL)-1, high mobility group B1 (HMGB1) and other pro-inflammatory cytokines. Of note, this cholinergic stimulation of macrophages does not reduce antiinflammatory cytokine release (e.g., IL-10) [65]. Figure 3 illustrates this mechanism for the spleen.

Important work by Fairchild and colleagues [67] has confirmed that the PNS and immune system interaction is actually bilateral, with vagal efferent signaling but also sensory afferent signaling, supporting the idea of a complete reflex loop. Indeed, mice exposed to pathogens by induction of peritonitis, demonstrated activation of both vagal efferent and afferent signaling pathways; these latter pathways activated cholinergic visceromotor neurons including the dorsal motor nuclei of the solitary tract in the brain stem.

The crucial role of PNS-mediated inflammatory modulation in sepsis has been illustrated by several studies with research demonstrating that electrical stimulation of the efferent vagus nerve inhibits the pro-inflammatory cytokine cascade, limiting the potentially damaging systemic inflammatory response and improving outcomes in animal models of endotoxic shock (opposite results being observed after vagotomy) [66, 68, 69]. This beneficial effect of the inflammatory reflex extends beyond infectious disease and pathogen-induced inflammation, as it has been shown, for example, to produce cardioprotection after acute myocardial infarction, significantly reducing myocardial ischemia/reperfusion injuries in pre-clinical investigations [70, 71].

Indices of autonomic dysfunction

Clinical tools that can be used to characterize autonomic dysfunction include HRV, BPV, baroreflex sensitivity (Fig. 4) and, less frequently, chemoreflex sensitivity [72–76]. A reduction in physiologic variability, measured with the standard indices of HRV, BPV and the BRS index, has been demonstrated to be directly correlated with septic shock severity and mortality in several experimental and clinical studies [7, 8, 77–79].

Reduced HRV and compromised vagal activity at the cardiac level have been documented in patients with



septic shock. The association between inflammation and HRV has been investigated in a rodent endotoxemia model. The authors reported that LPS-induced cytokine elevation was closely linked to HRV changes, as the major cytokine peaks were concomitant with maximal HRV depression [80]. In another study, the high levels of circulating catecholamines reported in septic shock were inversely correlated with indices of HRV [78]. Explanations for this finding may be that, similar to the situation in patients with heart failure, high sympathetic drive may lead to saturation of low-frequency oscillatory systems [81], or that excessive concentrations of circulating catecholamines may compromise central autonomic control [10, 60].

In a recent study by our group [82], analysis of cardiovascular variables in an experimental polymicrobial septic shock model in pigs suggested a pattern of autonomic dysfunction typical of septic shock. There was a significant positive correlation between an increase in aortic arterial stiffness and depressed vagal activity, together with decreased total peripheral resistance (TPR), decreased Windkessel time constant, τ , and impaired sympathetic activity at the peripheral sites. Interestingly, this abnormal condition generated by septic shock did not resolve after resuscitation (i.e., volume expansion and norepinephrine perfusion) and correction of hypotension. Although global hemodynamic markers were restored by the resuscitation maneuvers, the indices of cardiovascular function mediated by the ANS were still impaired, suggesting that the baseline homeostatic control had not completely recovered, similar to findings in an earlier study in an experimental model of hemorrhagic shock [83].

In another study by our group [84], the early response to standard resuscitation in septic shock patients was analyzed; patients were stratified according to the change in the sequential organ failure assessment (SOFA) score within 48 h of ICU admission. All the patients reached resuscitation goals, i.e., a mean arterial pressure (MAP) > 65 mmHg, but only the patients who significantly improved their SOFA score (responders) showed an increase in the BP oscillations associated with the sympathetic outflow. In the non-responders, the fluid balance was higher and, despite a higher dose of vasopressors, there was no increase in the BP oscillations associated with the sympathetic outflow. This observation may indicate poor responsiveness to the vasopressor therapy in these patients, despite similar BP values to the SOFA score responders.

From these results, BP variability, HRV, and baroreflex analyses can be considered valuable tools to understand the responsiveness of patients to sympathomimetic drugs and/or fluid administration and to identify patients with a worse prognosis who require more invasive or frequent monitoring. Resuscitation strategies should consider the balance of sympathetic and autonomic tone, considering the potential role of redirecting and maximizing sympathetic activity [43].



Fig. 4 Basic concepts to estimate heart rate and blood pressure variabilities. **a** Identification of R peaks in the ECG signal—in sinus rhythm permits assessment of the RR interval duration for consecutive beats. **b** The tachogram displays RR intervals for each beat; this time series analysis permits a first visual inspection of the variability in RR interval duration, computed as the mean RR interval and its standard deviation. **c** Before the spectral analysis, the time series needs to be resampled at evenly spaced time intervals $(T_c = \text{sampling interval which corresponds to } f_c = 1/T_c$ sampling frequency). **d** Power spectrum density (PSD) computation and spectral analysis: as the parasympathetic nervous system (PNS) acts at higher frequencies than the sympathetic nervous system (SNS), the region associated with high frequency (HF, 0.15–0.4 Hz) estimates the PNS contribution to RR interval modulation, whereas the low frequency (LF, 0.04–0.15 Hz) represents mainly the SNS contribution; LF and HF indexes represent the PSD areas in those bands. **e** Similar to HRV, beat-to-beat series of systolic (SBP) and diastolic (DBP) blood pressure values lead first to the time domain analysis and, after resampling, to PSD computation and spectral analysis (NB: in case of blood pressure, HF variations are not related to the PNS, but mainly to the respiratory activity). The simplest way to assess the baroreflex sensitivity consists of calculating the ratio between LF components of both SBP and RR series, in their respective PSD computation. **f** The spectral analysis permits to disentangle the individual contributions of several modulators (in this example, the signal y(t) is composed of three harmonics and the PSD can clearly identify those harmonics with the low-frequency signal, y1(t), contributing more than the other two)

Therapeutic interventions to modulate the ANS in septic shock

Table 2 summarizes some innovative therapeutic paradigms for modulating ANS in septic shock, although more robust clinical trials are needed to confirm whether they have beneficial effects on clinically-relevant outcomes.

Cardioselective **B1-blockers** and ivabradine

Although the concept of "adrenergic toxicity" in acute illness and in septic shock is well established in the scientific literature, there are no clear clinical recommendations for ANS modulation. For example, tachycardia persisting despite fluid and vasopressor administration is a hallmark of septic shock and an independent risk factor for increased mortality [85, 86]. The consequences of excessive tachycardia are multifactorial and include: (i) less diastolic time available for ventricular filling (i.e., impaired diastolic function) and for left ventricular myocardial coronary perfusion, while myocardial oxygen consumption is actually increased (i.e., potential functional ischemia); (ii) a higher risk of developing tachyarrhythmia; (iii) a certain degree of heart failure by cellular exhaustion when tachycardia is excessively high and prolonged (i.e., tachycardiomyopathy). One of the sources of this persistent tachycardia has been demonstrated to be protracted adrenergic stress at the cardiac level, which exceeds in time and degree the beneficial short-term compensatory effects [87].

For this reason, drugs that attenuate tachycardia are currently under investigation, assuming that the reduction in HR will be hemodynamically compensated for by longer ventricular filling, with higher stroke volume, and hence, a reasonably preserved CO. To date, the most advanced investigations involve ultra-short acting cardioselective \beta1-blockers, such as esmolol or landiolol. Esmolol administration has been tested in animal experiments [88, 89] and in a single center, open-label randomized clinical trial in septic shock patients with persistent tachycardia despite 24 h of full resuscitation [90]. Esmolol was associated with reduced HR and an unexpected faster vasopressor weaning together with an improvement in the 28-day mortality. This result still needs confirmation in a larger, multicenter clinical trial, however, because a recent attempt to replicate these observations was disappointing, mostly from a safety standpoint because of insufficient stroke volume compensation [91]. This risk related to the negative inotropic effect of esmolol was previously illustrated in another septic shock animal experiment, in which esmolol also had a global hypotensive effect [92]. Landiolol has also been studied in septic shock patients. In a multicenter Japanese randomized clinical trial with a similar design, landiolol was associated with HR reduction but with no statistically significant reduction in mortality, although mortality was not the primary endpoint of the study [93].

To overcome the negative inotropic effect of betablockers, another negative chronotropic drug was recently proposed, ivabradine, which specifically inhibits the pacemaker current (funny current, If) of the sinoatrial node cells, resulting in therapeutic HR lowering, without any negative inotropic or hypotensive side effects [94]. In acutely ill patients, ivabradine has been shown to effectively lower HR, but a beneficial effect on more relevant outcomes, such as major adverse cardiovascular events and mortality, has yet to be demonstrated [95]. A large randomized clinical trial aimed at controlling HR with ivabradine in septic shock is currently ongoing (IRISS trial; NCT04031573).

In addition to the already mentioned pathophysiological mechanisms (e.g., an attenuation of impairments in ventricular filling and myocardial perfusion), a possible mechanism explaining some of the beneficial effects of these β 1 selective antagonists is an indirect immunomodulatory effect. These agents were shown to stimulate vagal nerve activity in an endotoxemia animal model [96] and to preserve normal splenic T-lymphocyte counts in an animal sepsis model [97]. Moreover, in an animal endotoxemia model, landiolol infusion suppressed the high-mobility group box 1 (HMGB-1) expression, while improving lung injury and cardiac function [98].

Central a2-agonists

In addition to peripheral adrenergic modulation, it is important to mention the more recent interest in pharmacologic interventions acting on central a2-adrenergic receptors located in the brain and, in particular, on vascular pre-junctional terminals, where they can inhibit the release of norepinephrine in a form of negative feedback. Although conventional understanding of α 2-agonists has considered these drugs as antihypertensive agents (e.g., clonidine), robust evidence now highlights their role in CNS outflow as light sedative agents with a positive impact on peripheral adrenergic receptor regulation (e.g., dexmedetomidine). Intravenous infusion of central α 2-agonists in animal models of septic shock almost completely restored vascular responsiveness to catecholamines and angiotensin to pre-septic conditions [99]. This raises the hypothesis that central α 2-agonists inhibit central sympathetic outflow and consequently reduce the high levels of norepinephrine released at sympathetic nerve terminals. This effect would reduce α 1-receptor desensitization and transform it into a gradual resensitization. In other words, vascular α 1-receptors that were down-regulated in septic shock become progressively

up-regulated on the administration of α 2-agonists [100]. This hypothesis also suggests that the more the α 1-receptors are desensitized by high concentrations of endogenous catecholamines (e.g., in refractory septic shock), the larger the expected improved responsiveness to vasopressors. In recent studies and subgroup analyses, dexmedetomidine appeared to be associated with lower vasopressor requirements to maintain the same MAP target in septic shock [101, 102].

Vagal stimulation

In the last few years, a new field, "bioelectronic medicine", has been rapidly evolving, with the discovery and development of innovative nerve stimulating and sensing technologies to diagnose and regulate biological processes and treat disease. The idea is that by manipulating the neural signals, it may be possible to change the way physicians treat pathological conditions, including autoimmune disorders (e.g., rheumatoid arthritis), inflammatory pathologies (e.g., Crohn's disease), and potentially also sepsis and bleeding [65].

This new technology has been applied to the field of the "inflammatory reflex"-described earlier-and exploited in numerous studies, which have shown the beneficial effect of vagal stimulation in treating animal models of endotoxemia and septic shocks [68, 103, 104]. In a recent experimental swine study of polymicrobial sepsis [105], animals were divided into three groups: sham, sepsis, and sepsis with vagus nerve cervical electrical stimulation (initiated 6 h after peritonitis induction). Animals treated with vagal nerve stimulation required less fluid and norepinephrine administration to achieve the resuscitation targets. Vagus nerve stimulation reduced the number of activated macrophages and partially or completely prevented the development of hyperlactatemia, hyperdynamic hemodynamic status, septic cardiomyopathy, and cardiac mitochondrial dysfunction.

Conclusion and future perspectives

ANS dysfunction has an impact on the responsiveness of septic shock patients to fluids and to adrenergic drug administrations. BPV, HRV, and baroreflex sensitivity can be considered valuable tools to understand and quantify the ANS dysregulation associated with the potential loss of cardiovascular homeostasis. They could therefore be integrated into standard hemodynamic monitoring systems, to strengthen our ability to anticipate deterioration in patient condition, or to better modulate some therapies, such as vasopressors, including their dose and selection (e.g., whether or not to give adrenergic agents). Adrenergic vasoactive drug prescriptions should be tailored and personalized, to obtain a better balance between cardiovascular support and undesirable immunomodulation during septic shock.

A better understanding of ANS-mediated control of the cardiovascular system during septic shock may translate into future therapeutic expansion towards negative chronotropic agents (e.g., beta-blockers or ivabradine) or central α 2 agonists. Availability of novel technologies, such as noninvasive transcutaneous vagus nerve stimulation, in the clinical setting, will help pave the way for other innovative therapies to selectively stimulate the nervous system.

Abbreviations

ANS: Autonomic nervous system; BP: Blood pressure; BPV: Blood pressure variability; CO: Cardiac output; CSNA: Cardiac sympathetic nerve activity; GRKs: G protein-coupled receptor kinase; HR: Heart rate; HRV: Heart rate variability; ICU: Intensive care unit; iNOS: Inducible nitric oxide synthase; LPS: Lipopolysaccharide; MAP: Mean arterial pressure; MOF: Multi-organ failure; MSNA: Muscle sympathetic nerve activity; NO: Nitric oxide; NST: Nucleus of the solitary tract; PNS: Parasympathetic nervous system; PP: Pulse pressure; ROS: Reactive oxygen species; RSNA: Renal sympathetic nerve activity; SAD: Sino aortic denervation; SNS: Sympathetic nervous system; SOFA: Sequential organ failure assessment; SV: Stroke volume; TPR: Total peripheral resistance.

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References

1. Rhodes A, Evans LE, Alhazzani W, Levy MM, Antonelli M, Ferrer R, et al. Surviving sepsis campaign. Crit Care Med. 2017;45:486–552.

- Bone RC, Balk RA, Cerra FB, Dellinger RP, Fein AM, Knaus WA, et al. Definitions for sepsis and organ failure and guidelines for the use of innovative therapies in sepsis. Chest. 1992;101:1644–55.
- Rivers E, Nguyen B, Havstad S, Ressler J, Muzzin A, Knoblich B, et al. Early goal-directed therapy in the treatment of severe sepsis and septic shock. N Engl J Med. 2001;345:1368–77.
- Dellinger RP, Levy MM, Rhodes A, Annane D, Gerlach H, Opal SM, et al. Surviving sepsis campaign: international guidelines for management of severe sepsis and septic shock: 2012. Crit Care Med. 2013;41:580–637.
- Radaelli A, Castiglioni P, Cerrito MG, De Carlini C, Soriano F, Di Rienzo M, et al. Infusion of *Escherichia coli* lipopolysaccharide toxin in rats produces an early and severe impairment of baroreflex function in absence of blood pressure changes. Shock. 2013;39:204–9.
- Penn AH, Schmid-Schönbein GW. Severe intestinal ischemia can trigger cardiovascular collapse and sudden death via a parasympathetic mechanism. Shock. 2011;36:251–62.
- Schmidt H, Müller-Werdan U, Hoffmann T, Francis DP, Piepoli MF, Rauchhaus M, et al. Autonomic dysfunction predicts mortality in patients with multiple organ dysfunction syndrome of different age groups*. Crit Care Med. 2005;33:1994–2002.
- Pancoto JAT, Corrêa PBF, Oliveira-Pelegrin GR, Rocha MJA. Autonomic dysfunction in experimental sepsis induced by cecal ligation and puncture. Auton Neurosci. 2008;138:57–63.
- Shi K-Y, Shen F-M, Liu A-J, Chu Z-X, Cao Y-L, Su D-F. The survival time post-cecal ligation and puncture in sinoaortic denervated rats. J Cardiovasc Pharmacol. 2007;50:162–7.
- Annane D, Trabold F, Sharshar T, Jarrin I, Blanc AS, Raphael JC, et al. Inappropriate sympathetic activation at onset of septic shock: a spectral analysis approach. Am J Respir Crit Care Med. 1999;160:458–65.
- Piepoli M, Garrard CS, Kontoyannis DA, Bernardi L. Autonomic control of the heart and peripheral vessels in human septic shock. Intensive Care Med. 1995;21:112–9.
- Korach M, Sharshar T, Jarrin I, Fouillot J-P, Raphaël J-C, Gajdos P, et al. Cardiac variability in critically ill adults: influence of sepsis. Crit Care Med. 2001;29:1380–5.
- De Pasquale M, Moss TJ, Cerutti S, Calland JF, Lake DE, Moorman JR, et al. Hemorrhage prediction models in surgical intensive care: bedside monitoring data adds information to lab values. IEEE J Biomed Heal Informatics IEEE. 2017;21:1703–10.
- Moorman JR, Delos JB, Flower AA, Cao H, Kovatchev BP, Richman JS, et al. Cardiovascular oscillations at the bedside: early diagnosis of neonatal sepsis using heart rate characteristics monitoring. Physiol Meas. 2011;32:1821–32.
- Pinsky MR, Payen D. Functional hemodynamic monitoring. Crit Care. 2005;9:566–72.
- Vincent JL, Rhodes A, Perel A, Martin GS, Rocca GD, Vallet B, et al. Clinical review: update on hemodynamic monitoring—a consensus of 16. Crit Care. 2011;15:229.
- Moorman JR, Carlo WA, Kattwinkel J, Schelonka RL, Porcelli PJ, Navarrete CT, et al. Mortality reduction by heart rate characteristic monitoring in very low birth weight neonates: a randomized trial. J Pediatr. 2011;159:900-906.e1.
- Hatib F, Jian Z, Buddi S, Lee C, Settels J, Sibert K, et al. Machine-learning algorithm to predict hypotension based on high-fidelity arterial pressure waveform analysis. Anesthesiology. 2018;129:663–74.
- Hall JE. Guyton and hall textbook of medical physiology thirteenth edition. Elsevier: Saunders; 2011.
- Belletti A, Landoni G, Lomivorotov VV, Oriani A, Ajello S. Adrenergic downregulation in critical care: molecular mechanisms and therapeutic evidence. J Cardiothorac Vasc Anesth. 2020;34:1023–41.
- Vasudevan NT, Mohan ML, Goswami SK, Prasad SVN. Regulation of β-adrenergic receptor function. Cell Cycle. 2011;10:3684–91.
- 22. Rockman HA, Koch WJ, Lefkowitz RJ. Seven-transmembrane-spanning receptors and heart function. Nature. 2002;415:206–12.
- Ferguson SSG. Evolving concepts in G protein-coupled receptor endocytosis: the role in receptor desensitization and signaling. Pharmacol Rev. 2001;53:1–24.
- 24. Wu L-L, Yang S-L, Yang R-C, Hsu H-K, Hsu C, Dong L-W, et al. G Protein and adenylate cyclase complex-mediated signal transduction in the rat heart during sepsis. Shock. 2003;19:533–7.

- Elenkov IJ, Wilder RL, Chrousos GP, Vizi ES. The sympathetic nerve—an integrative interface between two supersystems: the brain and the immune system. Pharmacol Rev. 2000;52:595–638.
- 26. Stolk RF, van der Pasch E, Naumann F, Schouwstra J, Bressers S, van Herwaarden AE, et al. Norepinephrine dysregulates the immune response and compromises host defense during sepsis. Am J Respir Crit Care Med. 2020;202:830–42.
- Boldt J, Menges T, Kuhn D, Diridis C, Hempelmann G. Alterations in circulating vasoactive substances in the critically ill—a comparison between survivors and non-survivors. Intensive Care Med. 1995;21:218–25.
- Ostrowski SR, Gaïni S, Pedersen C, Johansson PI. Sympathoadrenal activation and endothelial damage in patients with varying degrees of acute infectious disease: an observational study. J Crit Care. 2015:30:90–6.
- Schmittinger CA, Torgersen C, Luckner G, Schröder DCH, Lorenz I, Dünser MW. Adverse cardiac events during catecholamine vasopressor therapy: a prospective observational study. Intensive Care Med. 2012;38:950–8.
- Kenney MJ, Ganta CK. Autonomic nervous system and immune system interactions. In: Comprehensive physiology. USA: Wiley; 2014. p. 1177–200.
- 31. Scanzano A, Cosentino M. Adrenergic regulation of innate immunity: a review. Front Pharmacol. 2015;6:171.
- Chapleau MW, Li Z, Meyrelles SS, Ma X, Abboud FM. Mechanisms determining sensitivity of baroreceptor afferents in health and disease. Ann N Y Acad Sci. 2001;940:1–19.
- Chapleau MW. Baroreceptor reflexes. In: Primer on the autonomic nervous system. The Netherlands: Elsevier; 2012. p. 161–5.
- Desai TH, Collins JC, Snell M, Mosqueda-Garcia R. Modeling of arterial and cardiopulmonary baroreflex control of heart rate. Am J Physiol Circ Physiol. 1997;272:H2343–52.
- 35. Crystal GJ, Salem MR. The Bainbridge and the "Reverse" Bainbridge reflexes. Anesth Analg. 2012;114:520–32.
- 36. Vatner SF, Zimpfer M. Bainbridge reflex in conscious, unrestrained, and tranquilized baboons. Am J Physiol Circ Physiol. 1981;240:H164–7.
- O'Regan RG, Majcherczyk S. Role of peripheral chemoreceptors and central chemosensitivity in the regulation of respiration and circulation. J Exp Biol. 1982;100:23–40.
- Milsom WK, Burleson ML. Peripheral arterial chemoreceptors and the evolution of the carotid body. Respir Physiol Neurobiol. 2007;157:4–11.
- Lahiri S, Forster RE. CO₂/H⁺ sensing: peripheral and central chemoreception. Int J Biochem Cell Biol. 2003;35:1413–35.
- Halliwill JR, Morgan BJ, Charkoudian N. Peripheral chemoreflex and baroreflex interactions in cardiovascular regulation in humans. J Physiol. 2003;552:295–302.
- 41. Somers VK, Mark AL, Abboud FM. Interaction of baroreceptor and chemoreceptor reflex control of sympathetic nerve activity in normal humans. J Clin Invest. 1991;87:1953–7.
- Dünser MW, Hasibeder WR. Sympathetic overstimulation during critical illness: adverse effects of adrenergic stress. J Intensive Care Med. 2009;24:293–316.
- 43. Ferreira JA, Bissell BD. Misdirected sympathy: the role of sympatholysis in sepsis and septic shock. J Intensive Care Med. 2018;33:74–86.
- 44. Benedict CR, Rose JA. Arterial norepinephrine changes in patients with septic shock. Circ Shock. 1992;38:165–72.
- Brown SM, Lanspa MJ, Jones JP, Kuttler KG, Li Y, Carlson R, et al. Survival after shock requiring high-dose vasopressor therapy. Chest. 2013;143:664–71.
- Rudiger A, Singer M. Decatecholaminisation during sepsis. Crit Care. 2016;20:309.
- Bucher M, Kees F, Taeger K, Kurtz A. Cytokines down-regulate α1-adrenergic receptor expression during endotoxemia. Crit Care Med. 2003;31:566–71.
- Bernardin G, Strosberg AD, Bernard A, Mattei M, Marullo S. Beta-adrenergic receptor-dependent and -independent stimulation of adenylate cyclase is impaired during severe sepsis in humans. Intensive Care Med. 1998;24:1315–22.
- Cariou A, Pinsky MR, Monchi M, Laurent I, Vinsonneau C, Chiche J-D, et al. Is myocardial adrenergic responsiveness depressed in human septic shock? Intensive Care Med. 2008;34:917–22.

- 50. Rudiger A. Beta-block the septic heart. Crit Care Med. 2010;38:S608–12.
- Schmidt C, Kurt B, Höcherl K, Bucher M. Inhibition of NF-kB activity prevents downregulation of alpha1-adrenergic receptors and circulatory failure during CLP-induced sepsis. Shock. 2009;32:239–46.
- Ackland GL, Kazymov V, Marina N, Singer M, Gourine AV. Peripheral neural detection of danger-associated and pathogen-associated molecular patterns. Crit Care Med. 2013;41:e85-92.
- Schmidt H, Mueller-Werdan U, Nuding S, Hoffmann T, Francis DP, Hoyer D, et al. Impaired chemoreflex sensitivity in adult patients with multiple organ dysfunction syndrome? The potential role of disease severity. Intensive Care Med. 2004;30:665–72.
- Godin PJ, Buchman TG. Uncoupling of biological oscillators. Crit Care Med. 1996;24:1107–16.
- Sharshar T, Gray F, de la Grandmaison GL, HopkInson NS, Ross E, Dorandeu A, et al. Apoptosis of neurons in cardiovascular autonomic centres triggered by inducible nitric oxide synthase after death from septic shock. Lancet. 2003;362:1799–805.
- Shen F-M, Guan Y-F, Xie H-H, Su D-F. Arterial baroreflex function determines the survival time in lipopolysaccharide-induced shock in rats. Shock. 2004;21:556–60.
- Nardocci G, Martin A, Abarzúa S, Rodríguez J, Simon F, Reyes EP, et al. Sepsis progression to multiple organ dysfunction in carotid chemo/ baro-denervated rats treated with lipopolysaccharide. J Neuroimmunol. 2015;278:44–52.
- Li Z, Mao HZ, Abboud FM, Chapleau MW. Oxygen-derived free radicals contribute to baroreceptor dysfunction in atherosclerotic rabbits. Circ Res. 1996;79:802–11.
- Nagano M, Nakamura M, Sato K, Tanaka F, Segawa T, Hiramori K. Association between serum C-reactive protein levels and pulse wave velocity: a population-based cross-sectional study in a general population. Atherosclerosis. 2005;180:189–95.
- Sayk F, Vietheer A, Schaaf B, Wellhoener P, Weitz G, Lehnert H, et al. Endotoxemia causes central downregulation of sympathetic vasomotor tone in healthy humans. Am J Physiol Integr Comp Physiol. 2008;295:R891–8.
- Ramchandra R, Wan L, Hood SG, Frithiof R, Bellomo R, May CN. Septic shock induces distinct changes in sympathetic nerve activity to the heart and kidney in conscious sheep. Am J Physiol Integr Comp Physiol. 2009;297:R1247–53.
- Vayssettes-Courchay C, Bouysset F, Verbeuren TJ. Sympathetic activation and tachycardia in lipopolysaccharide treated rats are temporally correlated and unrelated to the baroreflex. Auton Neurosci. 2005;120:35–45.
- Pålsson J, Ricksten SE, Lundin S. Changes in central hemodynamics during experimental septic shock in conscious rats. Circ Shock. 1987;22:65–72.
- 64. Mills E. Development of the sympathetic nervous system response to endotoxicosis in the rat: importance of non-baroreflex mechanisms in pre-weanlings and adults. J Dev Physiol. 1990;13:99–103.
- 65. Tracey KJ. The inflammatory reflex. Nature. 2002;420:853–9.
- 66. Pavlov VA, Tracey KJ. The cholinergic anti-inflammatory pathway. Brain Behav Immun. 2005;19:493–9.
- Fairchild KD, Srinivasan V, Randall Moorman J, Gaykema RPA, Goehler LE. Pathogen-induced heart rate changes associated with cholinergic nervous system activation. Am J Physiol Integr Comp Physiol. 2011;300:R330–9.
- Borovikova LV, Ivanova S, Zhang M, Yang H, Botchkina GI, Watkins LR, et al. Vagus nerve stimulation attenuates the systemic inflammatory response to endotoxin. Nature. 2000;405:458–62.
- Pavlov VA, Wang H, Czura CJ, Friedman SG, Tracey KJ. The cholinergic anti-inflammatory pathway: a missing link in neuroimmunomodulation. Mol Med. 2003;9:125–34.
- Chen M, Li X, Yang H, Tang J, Zhou S. Hype or hope: vagus nerve stimulation against acute myocardial ischemia-reperfusion injury. Trends Cardiovasc Med. 2020;30:481–8.
- Nuntaphum W, Pongkan W, Wongjaikam S, Thummasorn S, Tanajak P, Khamseekaew J, et al. Vagus nerve stimulation exerts cardioprotection against myocardial ischemia/reperfusion injury predominantly through its efferent vagal fibers. Basic Res Cardiol. 2018;113:22.
- Stauss HM. Identification of blood pressure control mechanisms by power spectral analysis. Clin Exp Pharmacol Physiol. 2007;34:362–8.

- Stauss HM. Heart rate variability. Am J Physiol Integr Comp Physiol. 2003;285:R927–31.
- Haji-Michael PG, Vincent J-L, Degaute JP, van de Borne P. Power spectral analysis of cardiovascular variability in critically ill neurosurgical patients. Crit Care Med. 2000;28:2578–83.
- Bravi A, Longtin A, Seely AJE. Review and classification of variability analysis techniques with clinical applications. Biomed Eng Online. 2011;10:90.
- Malik M, Bigger JT, Camm AJ, Kleiger RE, Malliani A, Moss AJ, et al. Heart rate variability: standards of measurement, physiological interpretation, and clinical use. Eur Heart J. 1996;17:354–81.
- Pontet J, Contreras P, Curbelo A, Medina J, Noveri S, Bentancourt S, et al. Heart rate variability as early marker of multiple organ dysfunction syndrome in septic patients. J Crit Care. 2003;18:156–63.
- Jarkovska D, Valesova L, Chvojka J, Benes J, Sviglerova J, Florova B, et al. Heart rate variability in porcine progressive peritonitis-induced sepsis. Front Physiol. 2016;6:412.
- Godin PJ, Fleisher LA, Eidsath A, Vandivier RW, Preas HL, Banks SM, et al. Experimental human endotoxemia increases cardiac regularity. Crit Care Med. 1996;24:1117–24.
- Fairchild KD, Saucerman JJ, Raynor LL, Sivak JA, Xiao Y, Lake DE, et al. Endotoxin depresses heart rate variability in mice: cytokine and steroid effects. Am J Physiol Integr Comp Physiol. 2009;297:R1019–27.
- Van de Borne P, Montano N, Pagani M, Oren R, Somers V. Absence of low-frequency variability of sympathetic nerve activity in severe heart failure. Circulation. 1997;95:1449–54.
- Carrara M, Herpain A, Baselli G, Ferrario M. Vascular decoupling in septic shock: the combined role of autonomic nervous system, arterial stiffness, and peripheral vascular tone. Front Physiol. 2020;11:594.
- Carrara M, Babini G, Baselli G, Ristagno G, Pastorelli R, Brunelli L, et al. Blood pressure variability, heart functionality, and left ventricular tissue alterations in a protocol of severe hemorrhagic shock and resuscitation. J Appl Physiol. 2018;125:1011–20.
- Carrara M, Bollen Pinto B, Baselli G, Bendjelid K, Ferrario M. Baroreflex sensitivity and blood pressure variability can help in understanding the different response to therapy during acute phase of septic shock. Shock. 2018;50:78–86.
- Parker MM, Shelhamer JH, Natanson C, Alling DW, Parrillo JE. Serial cardiovascular variables in survivors and nonsurvivors of human septic shock. Crit Care Med. 1987;15:923–9.
- Azimi G, Vincent J-L. Ultimate survival from septic shock. Resuscitation. 1986;14:245–53.
- 87. Rudiger A, Singer M. The heart in sepsis: from basic mechanisms to clinical management. Curr Vasc Pharmacol. 2013;11:187–95.
- 88. Suzuki T, Morisaki H, Serita R, Yamamoto M, Kotake Y, Ishizaka A, et al. Infusion of the β -adrenergic blocker esmolol attenuates myocardial dysfunction in septic rats^{*}. Crit Care Med. 2005;33:2294–301.
- Aboab J, Sebille V, Jourdain M, Mangalaboyi J, Gharbi M, Mansart A, et al. Effects of esmolol on systemic and pulmonary hemodynamics and on oxygenation in pigs with hypodynamic endotoxin shock. Intensive Care Med. 2011;37:1344–51.
- Morelli A, Ertmer C, Westphal M, Rehberg S, Kampmeier T, Ligges S, et al. Effect of heart rate control with esmolol on hemodynamic and clinical outcomes in patients with septic shock. JAMA. 2013;310:1683.
- Levy B, Fritz C, Piona C, Duarte K, Morelli A, Guerci P, et al. Hemodynamic and anti-inflammatory effects of early esmolol use in hyperkinetic septic shock: a pilot study. Crit Care. 2021;25:21.
- Hosokawa K, Su F, Taccone FS, Post EH, Pereira AJ, Herpain A, et al. Esmolol administration to control tachycardia in an ovine model of peritonitis. Anesth Analg. 2017;125:1952–9.
- 93. Kakihana Y, Nishida O, Taniguchi T, Okajima M, Morimatsu H, Ogura H, et al. Efficacy and safety of landiolol, an ultra-short-acting β 1-selective antagonist, for treatment of sepsis-related tachyarrhythmia (J-Land 3S): a multicentre, open-label, randomised controlled trial. Lancet Respir Med. 2020;8:863–72.
- Wei C, Al Kattani N, Louis H, Albuisson E, Levy B, Kimmoun A. If channel inhibition with ivabradine does not improve cardiac and vascular function in experimental septic shock. Shock. 2016;46:297–303.
- 95. Chen A, Elia N, Dunaiceva J, Rudiger A, Walder B, Bollen PB. Effect of ivabradine on major adverse cardiovascular events and mortality in critically ill patients: a systematic review and meta-analyses of

randomised controlled trials with trial sequential analyses. Br J Anaesth. 2020;124:726–38.

- Aboab J, Mayaud L, Sebille V, de Oliveira R, Jourdain M, Annane D. Esmolol indirectly stimulates vagal nerve activity in endotoxemic pigs. Intensive Care Med Exp. 2018;6:14.
- 97. Suzuki T, Inoue K, Igarashi T, Kato J, Nagata H, Yamada T, et al. Betablocker therapy preserves normal splenic T-lymphocyte numbers reduced in proportion to sepsis severity in a sepsis model. Crit Care Res Pract Hindawi. 2019;2019:1–5.
- Hagiwara S, Iwasaka H, Maeda H, Noguchi T. Landiolol, an ultrashortacting beta1-adrenoceptor antagonist, has protective effects in an LPS-induced systemic inflammation model. Shock. 2009;31:515–20.
- Geloen A, Chapelier K, Cividjian A, Dantony E, Rabilloud M, May CN, et al. Clonidine and dexmedetomidine increase the pressor response to norepinephrine in experimental sepsis. Crit Care Med. 2013;41:e431–8.
- Pichot C, Géloën A, Ghignone M, Quintin L. Alpha-2 agonists to reduce vasopressor requirements in septic shock? Med Hypotheses. 2010;75:652–6.
- Morelli A, Sanfilippo F, Arnemann P, Hessler M, Kampmeier TG, D'Egidio A, et al. The effect of propofol and dexmedetomidine sedation on norepinephrine requirements in septic shock patients. Crit Care Med. 2019;47:e89-95.
- 102. Cioccari L, Luethi N, Bailey M, Shehabi Y, Howe B, Messmer AS, et al. The effect of dexmedetomidine on vasopressor requirements in patients with septic shock: a subgroup analysis of the Sedation Practice in Intensive Care Evaluation [SPICE III] Trial. Crit Care. 2020;24:441.
- Czura CJ, Schultz A, Kaipel M, Khadem A, Huston JM, Pavlov VA, et al. Vagus nerve stimulation regulates hemostasis in swine. Shock. 2010;33:608–13.

- Liu CY, Mueller MH, Grundy D, Kreis ME. Vagal modulation of intestinal afferent sensitivity to systemic LPS in the rat. Am J Physiol Liver Physiol. 2007;292:G1213–20.
- Kohoutova M, Horak J, Jarkovska D, Martinkova V, Tegl V, Nalos L, et al. Vagus nerve stimulation attenuates multiple organ dysfunction in resuscitated porcine progressive sepsis. Crit Care Med. 2019;47:e461–9.
- Chen S-J, Wu C-C, Yang S-N, Lin C-I, Yen M-H. Abnormal activation of K+ channels in aortic smooth muscle of rats with endotoxic shock: electrophysiological and functional evidence. Br J Pharmacol. 2000;131:213–22.
- 107. Buckley J, Singer MCL. Role of KATP channels in sepsis. Cardiovasc Res. 2006;72:220–30.
- Bangash MN, Kong M-L, Pearse RM. Use of inotropes and vasopressor agents in critically ill patients. Br J Pharmacol. 2012;165:2015–33.
- Aiello EA, Malcolm AT, Walsh MP, Cole WC. β-Adrenoceptor activation and PKA regulate delayed rectifier K+ channels of vascular smooth muscle cells. Am J Physiol Circ Physiol. 1998;275:H448–59.
- Landry DW, Oliver JA. The ATP-sensitive K+ channel mediates hypotension in endotoxemia and hypoxic lactic acidosis in dog. J Clin Invest. 1992;89:2071–4.
- Olofsson PS, Rosas-Ballina M, Levine YA, Tracey KJ. Rethinking inflammation: neural circuits in the regulation of immunity. Immunol Rev. 2012;248:188–204.

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