RESEARCH Open Access



miR-129-5p as a biomarker for pathology and cognitive decline in Alzheimer's disease

Sang-Won Han¹, Jung-Min Pyun², Paula J. Bice³, David A. Bennett⁴, Andrew J. Saykin³, Sang Yun Kim⁵, Young Ho Park^{5*} and Kwangsik Nho^{3*}

Abstract

Background Alzheimer's dementia (AD) pathogenesis involves complex mechanisms, including microRNA (miRNA) dysregulation. Integrative network and machine learning analysis of miRNA can provide insights into AD pathology and prognostic/diagnostic biomarkers.

Methods We performed co-expression network analysis to identify network modules associated with AD, its neuropathology markers, and cognition using brain tissue miRNA profiles from the Religious Orders Study and Rush Memory and Aging Project (ROS/MAP) (N = 702) as a discovery dataset. We performed association analysis of hub miRNAs with AD, its neuropathology markers, and cognition. After selecting target genes of the hub miRNAs, we performed association analysis of the hub miRNAs with their target genes and then performed pathway-based enrichment analysis. For replication, we performed a consensus miRNA co-expression network analysis using the ROS/MAP dataset and an independent dataset (N = 16) from the Gene Expression Omnibus (GEO). Furthermore, we performed a machine learning approach to assess the performance of hub miRNAs for AD classification.

Results Network analysis identified a glucose metabolism pathway-enriched module (M3) as significantly associated with AD and cognition. Five hub miRNAs (miR-129-5p, miR-433, miR-1260, miR-200a, and miR-221) of M3 had significant associations with AD clinical and/or pathologic traits, with miR129-5p by far the strongest across all phenotypes. Gene-set enrichment analysis of target genes associated with their corresponding hub miRNAs identified significantly enriched biological pathways including ErbB, AMPK, MAPK, and mTOR signaling pathways. Consensus network analysis identified two AD-associated consensus network modules and two hub miRNAs (miR-129-5p and miR-221). Machine learning analysis showed that the AD classification performance (area under the curve (AUC) = 0.807) of age, sex, and APOE &4 carrier status was significantly improved by 6.3% with inclusion of five AD-associated hub miRNAs.

Conclusions Integrative network and machine learning analysis identified miRNA signatures, especially miR-129-5p, as associated with AD, its neuropathology markers, and cognition, enhancing our understanding of AD pathogenesis and leading to better performance of AD classification as potential diagnostic/prognostic biomarkers.

Keywords Alzheimer's disease, Braak, CERAD, Cognition, Machine learning, MicroRNA, miRNA-129-5p, Module, Network

*Correspondence: Young Ho Park kumimesy@snubh.org Kwangsik Nho knho@iupui.edu Full list of author information is available at the end of the article



Background

Alzheimer's disease is a prevalent cause of dementia, accounting for approximately 60% to 80% of cases [1]. It is characterized by the extracellular deposition of amyloid- β (A β) in the form of diffuse and neuritic plaques (NPs) and the presence of intracellular neurofibrillary tangles (NFTs) comprised of aggregated hyperphosphorylated tau protein [2]. However, the exact mechanisms underlying the pathogenesis of Alzheimer's disease remain unclear due to the involvement of complex neurochemical and genetic factors [3]. Dysregulated expression of microRNAs (miRNAs) is a potential mechanism contributing to gene expression changes in Alzheimer's disease [4–6].

miRNAs are endogenous single-stranded RNA molecules approximately 20–23 nucleotides long [7]. They primarily repress the translation of specific messenger RNAs (mRNAs) by binding to their 3'-untranslated regions (3'-UTR) [8]. Several miRNAs have been identified as being dysregulated in Alzheimer's disease, with certain miRNAs being highly expressed in the brain [4, 5, 9, 10]. However, there has been limited consensus among studies conducted on relatively small or modest sample sizes [11].

The construction of network modules based on the correlation of miRNA expression profiles can reveal the global properties of biological organization [12], given the assumption that miRNAs involved in similar functions tend to be co-expressed [13]. The weighted gene co-expression network analysis (WGCNA) approach is a method that focuses on gene co-expression networks and has been useful in describing the system-level correlation structure among transcripts [14]. Additionally, the network-based approach is a dimensionality reduction technique for analyzing high-dimensional omics data, providing insights into the pathogenesis of multifactorial disorders [15]. Therefore, the WGCNA approach has been utilized to enhance our understanding of the pathogenesis of Alzheimer's disease [16].

In this study, using miRNA expression profiles from a large longitudinal study of aging, the Religious Orders Study and Rush Memory and Aging Project (ROS/MAP) (N=702), as a discovery sample, we performed differential expression analysis and co-expression network analysis to identify Alzheimer's dementia (AD)-associated network modules and their hub miRNAs. We also investigated their association with neuropathological markers [2] and cognition. After selecting target genes of the hub miRNAs, we performed association analysis of the hub miRNAs with their target genes and then differential expression analysis of target genes using brain tissue RNA-Seq data. For replication analysis, we performed a consensus miRNA co-expression network analysis to

identify AD-associated consensus network modules and their hub miRNAs using the ROS/MAP dataset and an independent miRNA expression profile dataset (N = 16) from Gene Expression Omnibus (GEO). Finally, we employed a machine learning approach to assess the performance of hub miRNAs for the classification of AD.

Methods

Study samples

Two independent datasets were used: ROS/MAP and GEO. In the ROS/MAP cohort, subjects were categorized as having no cognitive impairment (NCI) or AD [17, 18]. In this study, to achieve a more robust differentiation between AD and NCI by employing both clinical and neuropathology criteria, AD was defined by Braak NFT scores $[19] \ge 4$, Consortium to Establish a Registry for Alzheimer's Disease (CERAD) scores [20] of definite Alzheimer's disease (frequent NPs) or probable Alzheimer's disease (moderate NPs), and cognitive diagnosis of probable Alzheimer's disease with no other causes. NCI was defined by Braak scores ≤ 3, the CERAD scores of possible Alzheimer's disease (sparse NPs) or no Alzheimer's disease, and clinical diagnosis of no cognitive impairment [21]. In the GEO (GSE157239) dataset from the Human Brain Bank of the Brazilian Aging Brain Study Group, AD was defined by Braak NFT scores [19] ≥ 3 and NCI as subjects without neuropathological lesions or neurological signs, as previously described [22].

miRNA profiling data

For the ROS/MAP cohort, miRNA profile data were downloaded from the Accelerating Medicines Partnership for Alzheimer's Disease (AMP-AD) Knowledge Portal on Synapse (syn3387325) (https://www.synapse. org). The miRNAs were extracted from bulk brain tissue in the dorsolateral prefrontal cortex (DLPFC) using the Nanostring nCounter Human miRNA Expression assay kit and annotated using definitions from the miR-Base [23]. These miRNAs were eluted from the miRNeasy spin columns in buffer and tested by Nanodrop and Bioanalyzer RNA 6000 Nano Agilent chips. The dataset consisted of 309 miRNAs from 702 individuals after correcting for the probe-specific backgrounds and performing a three-step filtering of sample and miRNA expression [6, 24]. The miRNA data were normalized using a combination of quantile normalization and Combat [25] to remove batch effects. The miRNAs identified by microarray were validated with specific real-time reverse transcription PCR (qRT-PCR) assays, as previously described in detail [6, 24].

For the GEO dataset, miRNA profiles were down-loaded from the National Center for Biotechnology Information as accession numbers GSE157239 (http://

www.ncbi.nlm.nih.gov/projects/geo/). The miRNAs were extracted from bulk brain tissue in the temporal cortex, profiled using microarray in the Affymetrix miRNA Array, and annotated using definitions from the miRBase [23]. After the isolation and biotin labeling of the miRNAs was performed, the labeled miRNAs were hybridized to the GeneTitan instrument with the Array Strip Hybridization kit [26]. Quality control was performed using the Expression Console software [27], and results were exported for processing in the Transcriptome Analysis Console software [28]. The miRNA identified by microarray were also validated via qRT-PCR, as previously described [22].

RNA-Sequencing for mRNA expression data in the ROS/MAP

RNA-Seq data generated from brain tissue in the DLPFC were downloaded from the AMP-AD Knowledge Portal on Synapse (syn8456638) (https://www.synapse.org). The sequencing was performed on the Illumina HiSeq2000 with 101 base pair paired-end reads, targeting a coverage of 50 million paired reads, as detailed in previous studies [24, 29]. The reads were aligned to the human genome reference (hg19) using Tophat [30] with Bowtie1 as the aligner [31]. The expression levels of transcripts were estimated using the Gencode V14 annotation with the RSEM package [32]. FPKM (fragments per kilobase of transcript per million mapped reads) normalization was applied to the mRNA expression data. The log2 countsper-million (logCPM) values generated in 634 subjects were finally used for further analysis, as previously described [33].

Assessment of CERAD, Braak, and cognition in ROS/MAP

Definite or probable Alzheimer's disease relative to possible or no Alzheimer's disease was based on CERAD, which is a semiquantitative measure of neuritic plaques, as previously reported [20, 34, 35]. Braak scores [19] ≧ 4 relative to \leq 3 were used to dichotomize neurofibrillary tangles as previously reported [34, 35]. We used the term "CERAD positive" for "probable" or "definite" Alzheimer's disease, and "CERAD negative" for "possible" or "no" Alzheimer's disease based on CERAD. We also used the term "Braak positive" for Braak scores ≧ 4, and "Braak negative" for Braak scores \leq 3. Cognitive performance was assessed longitudinally by Z-scores of global cognitive performance averaged across 19 tests spanning five cognitive domains, including episodic memory, working memory, semantic memory, perceptual speed, and visuospatial ability/perceptual orientation, as previously reported [36].

Co-expression miRNA network construction

A scale-free miRNA co-expression network was constructed using the WGCNA package based on miRNA expression profiles [14]. The "pickSoftThreshold" function was used to select an appropriate soft threshold power β that achieves a scale-free topology, and the network adjacency matrix was calculated based on coexpression similarity. Unsupervised hierarchical clustering with the dynamic tree cut procedure was used to identify modules of co-expressed miRNAs. Each module was represented by a module eigengene (ME), which was defined as the first principal component of the expression matrix representing the overall level of miRNA expression within each module.

Identification of hub miRNAs in an AD-associated network module and their target genes

The "softConnectivity" function was used to identify the top 10 hub miRNAs with the highest intramodular connectivity according to the topological overlap matrix (TOM) based on intramodular connectivity measures. The target genes of these hub miRNAs were predicted using the TargetScan [37] and the miRDB [38] databases, which provide both predicted and experimentally verified interaction information between miRNAs and genes. The overlapping target genes between the two databases were selected for further analysis.

Association of AD-associated hub miRNAs with their target genes using RNA-Seq data in the ROS/MAP

After target genes of AD-associated hub miRNAs were selected, brain tissue-based RNA-Seq data analyses were performed to assess whether the predicted target genes were associated with expression levels of their corresponding miRNAs.

Pathway-based enrichment analysis of target genes and miRNAs

Gene-set enrichment analysis was performed using the Database for Annotation, Visualization and Integrated Discovery (DAVID) online tool [39] to identify the biological pathways of target genes significantly associated with the expression levels of AD-associated hub miRNAs. The Kyoto Encyclopedia of Genes and Genomes (KEGG) [40] and Gene ontology (GO) biological processes (BP) [41] pathways were used.

miRNA-set enrichment analysis was also performed using the TAM 2.0 [42] tool to characterize the functional annotations of the miRNA set within each module through overrepresentation analysis. miRNA-set enrichment analysis was restricted to pathways containing 3 or more miRNAs. The most strongly associated biological

processes for the involvement of the central nervous system were selected for each module. Bonferroni correction [43] was applied to adjust for multiple testing.

Consensus network construction

A consensus miRNA co-expression network was constructed across the ROS/MAP and GEO (GSE46579) datasets for replication using the WGCNA package [14]. The "blockwiseConsensusModules" function was used to construct modules in the consensus network based on pairwise miRNA dissimilarity measures. The importance of a miRNA in a network module was determined by kME, defined as the strength of the correlation of expression levels of a miRNA with the ME. Hub miRNAs in a consensus module were defined as those miRNAs with an absolute kME value greater than 0.7.

The "modulePreservation" function was used to construct a preservation network based on the correlation between all pairs of consensus ME values across the two networks from the ROS/MAP and GEO datasets. A density (D) of the eigengene network, defined as the average scaled connectivity, was estimated to discover changes in preservation patterns across each consensus module. The D value close to 1 indicates strong preservation of the correlation patterns between all pairs of eigengenes across the two networks [44, 45].

Statistical analysis

For differential expression analysis, logistic regression models were used to investigate the association of miR-NAs expression levels with AD, CERAD, and Braak stage. For co-expression miRNA network analysis, logistic regression models were used to evaluate the association of ME values of network modules with AD and to identify AD-associated network modules. Linear regression models were used to perform the association analysis of miRNA expression levels and ME values of network modules with global cognitive performance at the last visit, while linear mixed effects models were used to investigate their association with longitudinal changes of global cognitive performance.

For differential expression analysis using RNA-Seq data of the target genes identified as significantly associated with expression levels of AD-associated hub miRNAs, logistic regression models were used to investigate the association between expression levels of the target genes and AD.

Covariates in the association analysis with AD and neuropathological markers included age, sex, study (ROS or MAP), RNA integrity numbers, and post-mortem interval. For the association analysis with cognitive performance, education was included along with the aforementioned covariates. The false discovery rate (FDR)

correction was used to adjust for multiple testing with the Benjamini–Hochberg procedure [46] unless otherwise specified.

The STREAMLINE tool [47], a machine learning approach using penalized logistic regression, was used to investigate the classification performance of ADassociated hub miRNAs in differentiating AD from NCI. This approach was chosen to reduce the effect of multicollinearity on feature selection [48]. The data were randomly divided into 70% used for training the model and 30% used for testing, with a 10-fold cross-validation procedure. The classification performance of five different models was evaluated using the receiver operating characteristic (ROC) curve and the area under the receiver operating characteristic curve (AUC). Training features in Model 1 included age, sex, and apolipoprotein Ε (APOE) ε4 carrier status; training features in Model 2 included five AD-associated hub miRNAs; training features in Model 3 included age, sex, APOE ε4 carrier status, and five AD-associated hub miRNAs; training features in Model 4 included all 309 miRNAs, and training features in Model 5 included age, sex, APOE ε4 carrier status, and all 309 miRNAs. Paired *t*-tests were performed to compare the AUC results for the different models [49].

All statistical analyses were conducted using R version 4.2.0, and statistical significance was set at a *P*-value of 0.05 after adjusting for multiple comparisons. Figure 1 shows a workflow of all analysis steps used in this study.

Results

The ROS/MAP cohorts with miRNA consisted of 702 participants, including 102 NCI and 177 AD, with a median age of 88.5 at death and 64.1% females. The GEO (GSE157239) dataset consisted of 16 participants, including 8 NCI and 8 AD, having miRNA data, with a median age of 81.5 at death and 68.8% females. Table 1 and Table S1 show the demographic characteristics of these individuals.

Differential expression analysis of miRNAs

Differential expression analysis was performed on 309 miRNAs in 177 AD and 102 NCI subjects from ROS/MAP. Fifteen miRNAs were found to be significantly associated with AD (Fig. 2). Notably, expression levels of miR-129-5p and miR-132 were significantly lower in both the CERAD and Braak positive groups (Fig. 2 and Table S2). Additionally, higher expression levels of miR-129-3p, miR-129-5p, miR-132, miR-133b, miR-410, miR-433, and miR-504, as well as lower expression levels of miR-100, miR-19b, miR-29a, miR-335, miR-519a, and miR-99b, were significantly associated with better global cognitive performance at the last visit. Furthermore,

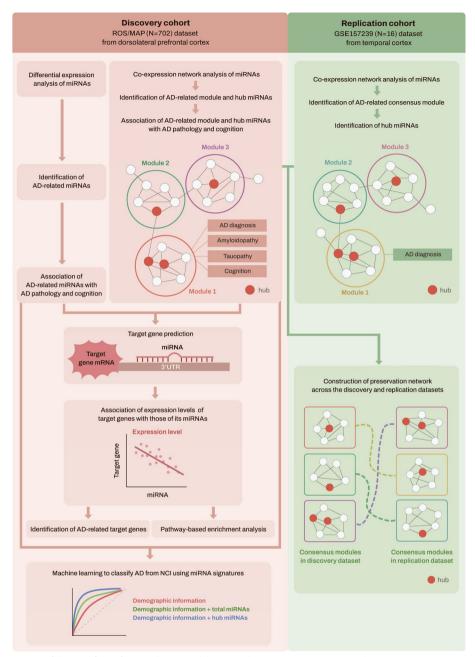


Fig. 1 Schematic overview of the workflow of our analysis. AD Alzheimer's dementia, MAP Memory and Aging Project, miRNAs microRNAs, mRNAs messenger RNAs, NCI no cognitive impairment, ROS Religious Orders Study, 3'-UTR 3'-untranslated region

higher expression levels of miR-129-3p, miR-129-5p, and miR-132 showed significant associations with slower longitudinal decline of global cognition.

Co-expression miRNA network analysis Modules associated with AD, CERAD, Braak, and cognition

A scale-free co-expression network was constructed based on the miRNA expression profiles of 702

subjects using WGCNA. A cutting tree height of 11 was selected to eliminate outliers, and 663 individuals under the cutting tree height were kept for further analysis. A soft thresholding power value of $\beta=4$ was selected, and four network modules were identified (Figure S1). The M0 module consisted of miRNAs not assigned to any other modules and was excluded in further analysis. Lower ME values of the M3 module

Table 1 Demographic information of participants from the ROS/MAP cohort

	NCI (n = 102)	AD (n = 177)	Total (n = 702)	P value ^b
Female	54 (52.9)	126 (71.2)	449 (64.1)	0.002
Age at death in years ^a	85.0 (78.8–89.0)	90.0 (87.4–90.0)	88.5 (84.1-90.0)	< 0.001
Study				
ROS	53 (52.0)	93 (52.5)	381 (54.4)	
MAP	49 (48.0)	84 (47.5)	320 (45.6)	0.926
Global cognitive Z scores at the last visit ^a	0.10 (-0.25-0.36)	-0.35 (-0.93-0.06)	-0.17 (-0.56–0.19)	< 0.001
Slope of global cognitive Z scores ^a	-0.03 (-0.13 - 0.02)	-0.17 (-0.300.08)	-0.09 (-0.220.01)	< 0.001
RNA integrity numbers ^a	7.3 (6.0-8.0)	6.7 (5.4–7.4)	6.9 (5.6–7.7)	0.001
Post-mortem interval in hours ^a	6.3 (4.8–8.6)	5.8 (4.2–8.3)	5.8 (4.3–8.5)	0.124

Values are n (%), unless indicated otherwise. Among a total of 702 participants with miRNA data, 177 subjects met criteria for AD and 102 met criteria for NCI based on neuropathological and clinical data

AD Alzheimer's dementia, MAP Memory and Aging Project, miRNAs microRNAs, NCI no cognitive impairment, ROS Religious Orders Study

^b The Mann–Whitney *U* test or chi-square test was used to determine the *P* value for comparisons between AD and NCI groups, as appropriate

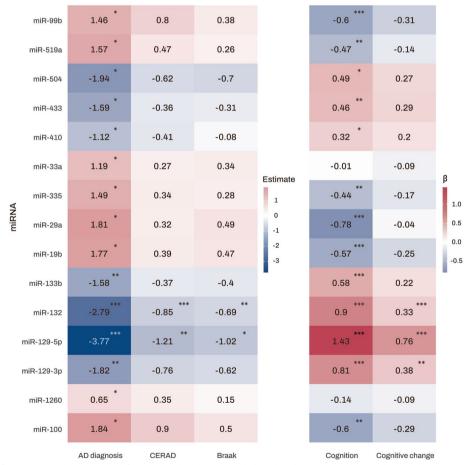


Fig. 2 Heatmap of association analysis results of miRNAs with diagnosis and clinical and pathological traits. The numbers represent the logistic and linear regression coefficients of association between miRNAs and traits. Stars indicate significant associations with FDR-corrected p value < 0.05. Red colors represent positive correlations, while blue colors denote negative correlations. The darker the color, the stronger the association. AD Alzheimer's dementia, CERAD Consortium to Establish a Registry for Alzheimer's Disease, FDR false discovery rate, miRNAs microRNAs. Note: significance stars indicating the p values of the correlations adjusted for multiple comparisons. *p-value < 0.05. *p-value < 0.01. *p-value < 0.001

^a Data are presented as median (interquartile range)

were significantly associated with a greater likelihood of AD and CERAD, but not with Braak stage. Moreover, lower ME values of the M3 module were associated with lower global cognitive performance at the last

visit and faster longitudinal decline of global cognition (Fig. 3 and Table S3). Enrichment analysis of miRNAs revealed that M3 was highly associated with glucose metabolism, and M1 and M2 were strongly linked to

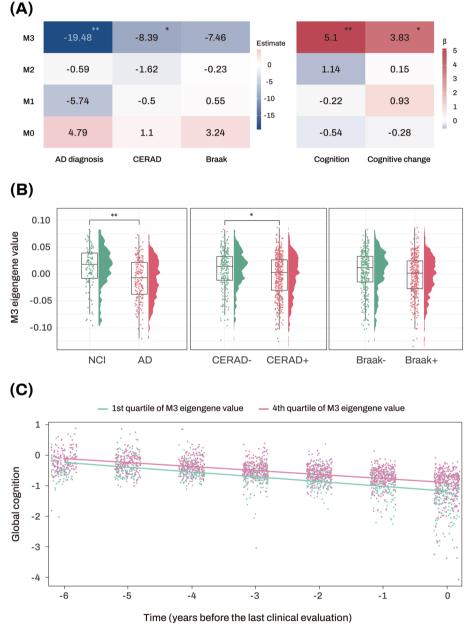


Fig. 3 Association analysis results of module eigengenes with diagnosis and clinical and pathological traits. **A** The numbers represent the logistic and linear regression coefficients of association between module eigengenes and traits. Stars indicate significant associations with FDR-corrected *p* value < 0.05. Red colors represent positive correlations, while blue colors denote negative correlations. The darker the color, the stronger the association. **B** Violin and box plots represent the M3 module eigengene values in the diagnosis, amyloidopathy, and tauopathy groups from the ROS/MAP cohort. **C** The *x*-axis represents years before the last clinical evaluation, while the *y*-axis indicates global cognitive Z scores. The two lines represent the different slopes of global cognitive changes of subjects with 1st quartile (green color) versus 4th quartile (red color) stratified by ME values of AD-associated M3 module. AD Alzheimer's dementia, CERAD Consortium to Establish a Registry for Alzheimer's Disease, FDR false discovery rate, MAP Memory and Aging Project, ME module eigengene, NCI no cognitive impairment, ROS Religious Orders Study. Note: significance stars indicating the *p* values of the correlations adjustment for multiple comparisons. **P*-value < 0.05. ***P*-value < 0.01

innate immunity and embryonic development, respectively (Table S4).

Hub miRNAs associated with AD, CERAD, Braak, and cognition

In the AD-associated glucose metabolism pathwayenriched M3 module, the top 10 miRNAs with the highest TOM-based intramodular connectivity were identified as hub miRNAs (Fig. 4). Among these 10 miRNAs, the strongest finding was for miR-129-5p, which was inversely related to AD, CERAD, and Braak and directly related to level of and change in cognition. Higher expression levels of miR-433 and miR-221 were inversely associated with the likelihood of AD, while only miR-433 was associated with better cognition and slower cognitive decline (Fig. 4 and Table S5). By contrast, higher expression levels of miR-200a and miR-1260 were related to a greater likelihood of AD, higher CERAD and lower cognition, but not with Braak or cognitive decline. Finally, miR-744 was only associated with lower global cognition.

Target genes associated with AD for AD-associated hub miRNAs

Target genes for the five AD-associated hub miRNAs (miR-129-5p, miR-433, miR-1260, miR-200a, and miR-221) were obtained from the miRDB and TargetScan

databases, resulting in 1417, 614, 567, 1526, and 1164 genes from the miRDB database, and 732, 1391, 5239, 4128, and 5878 genes from the TargetScan database; 406, 198, 512, 892, and 769 overlapping target genes for these five miRNAs, respectively, between the two databases were then identified (for reference, target genes for the hub miRNAs identified in the M1, M2, and M3 modules were listed in the Supplementary Additional file).

RNA-Seq data analysis identified 189, 101, 310, 591, and 73 target genes as significantly associated with expression levels of corresponding miRNAs for miR-129-5p, miR-433, miR-1260, miR-200a, and miR-221, respectively. Furthermore, RNA-Seq data analysis identified a total of 22, 3, 12, 30, and 25 target genes as significantly associated with AD for miR-129-5p, miR-433, miR-1260, miR-200a, and miR-221, respectively (Table 2).

Pathway-based enrichment analysis of target genes

The KEGG pathway-based enrichment analysis revealed that the target genes identified through RNA-Seq analysis for the five AD-associated hub miRNAs were mainly involved in the following pathways: axon guidance, erythroblastic leukemia viral oncogene homolog (ErbB) signaling pathway, mitogen-activated protein kinase (MAPK) signaling pathway, gamma-aminobutyric acid-ergic (GABAergic) synapse, autophagy, 5' adenosine monophosphate-activated protein kinase (AMPK)

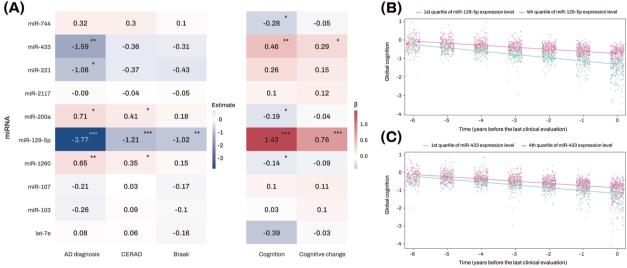


Fig. 4 Association analysis results of top 10 candidate hub miRNAs with diagnosis and clinical and pathological traits. **A** The numbers represent the logistic and linear regression coefficients of association between miRNAs and traits. Stars indicate significant associations with FDR-corrected *p* value < 0.05. Red colors represent positive correlations, while blue colors denote negative correlations. The darker the color, the stronger the association. **B, C** The *x*-axis represents years before the last clinical evaluation, while the *y*-axis indicates global cognitive Z scores. The two lines represent the different slopes of global cognitive changes of subjects with 1st quartile (green color) versus 4th quartile (red color) stratified by expression levels of miR-129-5p (**B**) and miR-433 (**C**), respectively. AD Alzheimer's dementia, CERAD Consortium to Establish a Registry for Alzheimer's Disease, FDR false discovery rate, miRNAs microRNAs. Note: significance stars indicating the *p* values of the correlations adjustment for multiple comparisons. **P*-value < 0.05. ***P*-value < 0.01. ****P*-value < 0.001

Table 2 Results of differential expression analysis of AD-associated target genes of five AD-associated hub miRNAs between NCI and AD. Here we showed only significantly differentially expressed target genes in AD

miR-129-5p		miR-433		miR-1260		miR-200a		miR-221	
Target genes	P value ^a								
PIEZO2	0.001	BRWD1	0.036	SAMD4A	0.005	SLC6A9	< 0.001	DYNC1LI2	0.002
NRN1	0.015	NSMCE1	0.036	NRN1	0.022	C17orf58	0.005	MLIP	0.012
ZNF704	0.015	ST8SIA4	0.036	MID1IP1	0.022	SLFN5	0.025	BRWD1	0.012
COX15	0.030			LGMN	0.023	TNS3	0.025	NRIP2	0.012
CHD7	0.030			TBC1D24	0.023	SLC38A2	0.025	ZFR2	0.012
PCDH8	0.030			LRRC73	0.033	RUFY2	0.027	ZNF652	0.012
TOMM40	0.030			GOLGA8B	0.033	CHP1	0.035	EML6	0.012
SMARCC1	0.030			CCDC113	0.036	BRWD1	0.035	LPAR1	0.012
RAB3B	0.030			RHNO1	0.041	LRRC8B	0.035	KIF1C	0.012
TMEM67	0.030			LIMD2	0.041	STAT4	0.035	RAB3B	0.014
USP13	0.030			BCKDK	0.044	DAP3	0.035	PTP4A2	0.016
SEPHS1	0.030			TRAPPC6A	0.044	TCERG1	0.035	NAP1L5	0.016
NRXN3	0.035					TATDN3	0.035	AKAP5	0.017
PAK3	0.038					MTF2	0.035	NXPH1	0.023
IGSF3	0.039					SLC20A1	0.038	PLXNC1	0.023
TSPYL4	0.039					ZNF652	0.041	PCDHGC5	0.023
SKP1	0.039					SLC4A1AP	0.043	NDFIP1	0.023
ZDHHC23	0.042					BTF3L4	0.043	ATP9B	0.023
GRM7	0.042					FBXL2	0.043	C11orf87	0.023
C2orf88	0.042					PCDH8	0.043	CAMK4	0.024
CKAP4	0.046					GULP1	0.043	COPS7A	0.026
CBX4	0.049					MFN1	0.043	PGAP1	0.030
						GRHL1	0.043	PPP1R14C	0.030
						FOXJ1	0.043	LRFN2	0.036
						KIF1C	0.043	STMN1	0.048
						CHD7	0.043		
						NRIP3	0.047		
						RHEB	0.049		
						MBTPS2	0.049		
						SEPT8	0.049		

AD Alzheimer's dementia, FDR false discovery rate, miRNAs microRNAs, NCI no cognitive impairment

signaling pathway, mammalian target of rapamycin (mTOR) signaling pathway, and glutamatergic synapse (Table 3). The GO-BP pathway-based enrichment analysis showed that target genes identified through RNA-Seq analysis for the five AD-associated hub miRNAs were enriched in the following pathways: protein phosphorylation, nervous system development, chromatin organization, and neuron migration (Table S6).

Consensus network analysis using miRNA profiles from two independent datasets

Consensus modules and a preservation network were constructed to understand changes in preservation

patterns across consensus modules. Additionally, hub miRNAs in consensus modules were selected to assess the replication of the five AD-associated hub miRNAs, identified in the ROS/MAP dataset, within an independent dataset.

Identification of consensus network modules

The construction of miRNA co-expression network modules was performed separately for the discovery and replication datasets, and the consensus modules were identified using the consensus dissimilarity measures in the average linkage hierarchical clustering method. A soft thresholding power value of $\beta=4$ was selected for each

 $^{^{\}rm a}$ Adjusted p value using FDR

Table 3 KEGG pathway analysis of target genes of five AD-associated hub miRNAs (miR-129-5p, miR-1260, miR-200a, miR-433, and miR-221)

KEGG pathway	Number of target genes belonging to the pathway	Fold enrichment	P value ^a	
Axon quidance	31	2.931	< 0.001	
ErbB signaling pathway	20	4.048	< 0.001	
MAPK signaling pathway	35	1.994	0.012	
GABAergic synapse	16	3.093	0.014	
Autophagy - animal	21	2.562	0.015	
AMPK signaling pathway	19	2.702	0.017	
mTOR signaling pathway	22	2.426	0.020	
Glutamatergic synapse	18	2.693	0.032	

AD Alzheimer's dementia, AMPK 5' adenosine monophosphate-activated protein kinase, ErbB erythroblastic leukemia viral oncogene homolog, GABAergic gamma-aminobutyric acid-ergic, KEGG Kyoto Encyclopedia of Genes and Genomes, MAPK mitogen-activated protein kinase, miRNAs microRNAs, mTOR mammalian target of rapamycin

dataset, and four consensus modules were identified (Figure S2). All consensus modules had counterparts in both datasets, indicating that the consensus module structures in two datasets were similar (Figure S3). The consensus CM0 module consisted of miRNAs not assigned to any other modules.

Preservation of consensus modules

A consensus ME network was constructed to investigate whether expression patterns of modules were correlated

with each other (Figure 5A, B). The preservation networks of the correlations of the consensus ME pairs between the discovery and replication datasets were further constructed to understand the changes in preservation patterns of two datasets (Figure 5C). The D value of the preservation networks between all pairs of the consensus ME across the two networks was 0.88 (Figure 5D), indicating that these modules were well preserved in their expression patterns across the two independent datasets.

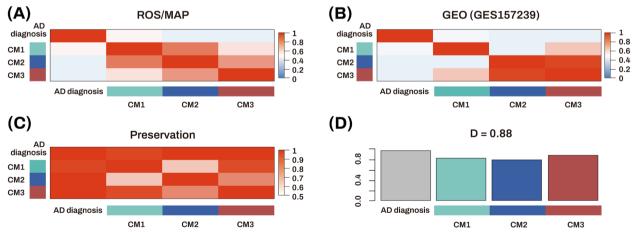


Fig. 5 Construction of consensus module eigengene networks. **A, B** Heatmaps showing the adjacencies of eigengene for each of the eigengene networks (**A**, ROS/MAP cohort; **B**, GSE157239 cohort). Each row and column corresponds to an eigengene labeled by the consensus module or diagnosis. Red colors represent positive correlations, while blue colors denote negative correlations. The darker the color, the stronger the association. **C** Adjacency heatmap displaying the pairwise preservation between the two consensus eigengene networks. Each row and column corresponds to an eigengene labeled by the consensus module or diagnosis. Red colors represent adjacency. The darker the color, the higher the adjacency. **D** Bar plot showing the preservation of correlation of consensus module eigengene between the two networks. Each colored bar corresponds to the eigengene of the associated consensus module. The height of the bar (*y*-axis) represents the eigengene preservation measure. The *D* value indicates the mean preservation of eigengene networks across the datasets. The high density value of *D* (= 0.88) denotes high overall preservation between the two networks. AD Alzheimer's dementia, CM consensus module, M module, MAP Memory and Aging Project, ROS Religious Orders Study

^a Adjusted *p* value from Bonferroni correction

Identification of AD-associated consensus modules across two independent datasets and replication for AD-associated hub miRNAs

Figure 6 shows the module-clinical trait heatmaps, indicating the associations between AD and four consensus modules across two independent datasets. Lower ME values of the consensus CM2 and CM3 modules were significantly associated with AD in the ROS/MAP dataset. In the GEO dataset, although the association between ME values of the consensus CM2 and CM3 modules and AD was not significant, the effect sizes and association directions were consistent with those in the ROS/MAP dataset. Among the five AD-associated hub miRNAs identified in the discovery dataset (ROS/MAP), miR-129-5p, miR-221, and miR-200a were included in the CM2 module (Table S7), but none of the miRNAs were present in the CM3 module (Table S8). Notably, miR-129-5p and miR-221, identified as AD-associated hub miR-NAs in the ROS/MAP cohort, were also hub miRNAs in an independent replication dataset (GEO) because their kME values in the replication dataset (GEO) were higher than 0.7 (Table 4 and Table S8). The correlation analysis showed that the M3 module from the ROS/ MAP dataset and the consensus CM2 module from the combined ROS/MAP and GEO datasets were strongly correlated (correlation coefficient = 0.88) (Figure S4).

Table 4 List of miRNAs with kME > 0.7 for the CM2 module in the replication dataset

kME	P value ^a	
0.946	1.02 × 10 ⁻⁶	
0.892	4.60×10^{-5}	
0.887	4.60×10^{-5}	
0.883	4.60×10^{-5}	
0.881	4.60×10^{-5}	
0.863	9.46×10^{-5}	
0.850	1.51×10^{-4}	
0.833	2.58×10^{-4}	
0.827	2.77×10^{-4}	
0.825	2.77×10^{-4}	
0.823	2.77×10^{-4}	
0.807	4.55×10^{-4}	
0.753	1.99×10^{-3}	
0.716	4.42×10^{-3}	
	0.946 0.892 0.887 0.883 0.881 0.863 0.850 0.833 0.827 0.825 0.825 0.823 0.807 0.753	

kME and p values represent the correlation and significance levels, respectively, between the miRNA expression levels and module eigengene of the consensus CM2 module. The miRNAs with kME > 0.7 in the replication dataset (GEO) are listed for the CM2 module

CM consensus module, FDR false discovery rate, GEO Gene Expression Omnibus, ME module eigengene, miRNAs microRNAs

Enrichment analysis of miRNAs identified glucose metabolism as a significantly enriched biological pathway in both M3 and CM2 (Table S9).

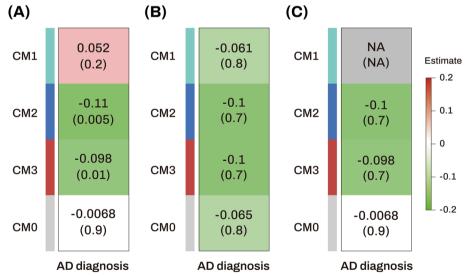


Fig. 6 Association analysis results of consensus module eigengenes with diagnosis in the ROS/MAP cohort, GEO (GSE157239) cohort, and across ROS/MAP and GEO datasets. **A, B** Heatmaps showing module-trait relationships in the ROS/MAP (**A**) and GEO (GSE157239) (**B**) cohorts. **C** Heatmaps showing consensus module-trait relationships across the ROS/MAP and GSE157239 cohorts. The numbers in the table indicate the coefficients (top) and its *p* value (bottom) of association between consensus module eigengenes and diagnosis. Red colors represent positive correlations, while green colors denote negative correlations. The darker the color, the stronger the association. Missing (NA) entry indicates the failure of forming a consensus because the directions of correlations in the ROS/MAP and GSE157239 datasets are opposite. AD Alzheimer's dementia, CM consensus module, GEO Gene Expression Omnibus, M module, MAP Memory and Aging Project, NA not applicable, ROS Religious Orders Study

^a Adjusted *p* value using FDR

Machine learning analysis for AD classification

A machine learning approach using penalized logistic regression for the classification of AD from NCI was used to evaluate five different classification models (Table 5). The results of 10-fold cross-validation are presented in Figure 7. Model 1, including age, sex, and APOE ε4 carrier status, achieved a mean AUC value of 0.807 with a standard deviation of 0.103 (Figure 7A). The mean AUC value of Model 3, obtained by adding five AD-associated hub miRNAs to Model 1, significantly increased to 0.870 with a standard deviation of 0.061 (P value = 0.022) (Figure 7C), which was comparable to that of Model 5, obtained by adding all 309 miRNAs to Model 1 (Figure 7E). The mean AUC values of Model 2, including only five AD-associated hub miRNAs, and Model 4, including all 309 miRNAs, were 0.740 and 0.815 with a standard deviation of 0.068 and 0.052, respectively (Figure 7B, D).

Discussion

In this study, we performed a network-based analysis of miRNAs and identified a network module (glucose metabolism pathway-enriched M3) that showed significant associations with AD, level of and change in cognition, and CERAD and Braak pathologic traits of Alzheimer's disease. In the AD-associated glucose metabolism pathway-enriched M3 module, we identified five hub miRNAs (miR-129-5p, miR-433, miR-1260, miR-200a, and miR-221) as significantly associated with AD, neuropathologic markers, and/or cognition, with miR-129-5p being the strongest and associated with all AD traits. Gene-set enrichment analysis of the target genes of these five AD-associated hub miRNAs revealed enrichment of ErbB, MAPK, AMPK, and mTOR signaling pathways. In the replication analysis using an independent additional dataset, we identified AD-associated CM2 and CM3 modules. Remarkably, miR-129-5p and miR-221, identified as AD-associated hub miRNAs in the discovery cohort, were also found to be hub miRNAs in the replication cohort. Preservation analysis showed consistent expression patterns of the consensus modules across two independent datasets.

This study identified AD-associated miRNAs through network-based analysis, not detected in the miRNAbased differential expression analysis. Moreover, our findings shed light on the association between AD-associated miRNAs and longitudinal changes of cognition.

Among five AD-associated hub miRNAs, we found that greater miR-129-5p was associated with a lower likelihood of AD, better cognition, slower cognition decline, and lower CERAD and Braak pathologic traits of Alzheimer's disease, which aligns with previous studies [6, 50-53]. miR-129-5p has been suggested to play roles in Alzheimer's disease, potentially involving the regulation of autophagy [51], neuroinflammation [52, 54, 55], and neuronal cell death [53-55] through targeting amyloid precursor protein (APP) [51], high-mobility group box 1 (HMGB1) [55], and yes-associated protein 1 (YAP1) [53] genes. In particular, miR-129-5p exhibits specific expression in brain tissues, suggesting a potential role for this miRNA in nervous system function [56]. Within the neocortex, miR-129-5p shows enrichment in glutamatergic projection neurons compared to GABAergic interneurons and GABAergic neurons [57, 58]. For miR-221, we observed decreased expression levels in AD but did not find any associations with neuropathological markers or cognition, which aligns with the study by Manzine et. al. [59]. For miR-200a, we observed increased expression levels in AD, consistent with previous studies [50, 60-64], highlighting its role in the Aβ-induced neuronal apoptosis and cell cycle deregulation [61-64]. Interestingly, we also identified a novel association between higher miR-200a expression levels and worse cognition. For miR-433, we observed decreased expression levels in AD and its association with cognition but not neuropathological markers, which aligns with the study by Wang et al. [65]. Lastly, miR-1260 exhibited increased expression levels in

Table 5 Mean AUC and standard deviation of machine learning models using penalized logistic regression

Model	Training features	Mean AUC	Standard deviation of AUC
1	Age + sex + APOE ε4 carrier status	0.807	0.103
2	Five AD-associated hub miRNAs	0.740	0.068
3	Age + sex + APOE E4 carrier status + five AD-associated hub miRNAs	0.870	0.061
4	All 309 miRNAs	0.815	0.052
5	Age + sex + APOE £4 carrier status + all 309 miRNAs	0.867	0.057

10-fold cross validation was used to investigate and compare the classification performance of five different machine learning models for differentiating AD from NCI. The machine learning model, mean AUC, and standard deviation of the AUC are presented

AD Alzheimer's dementia, APOE apolipoprotein E, AUC area under the curve, miRNAs microRNAs, NCI no cognitive impairment

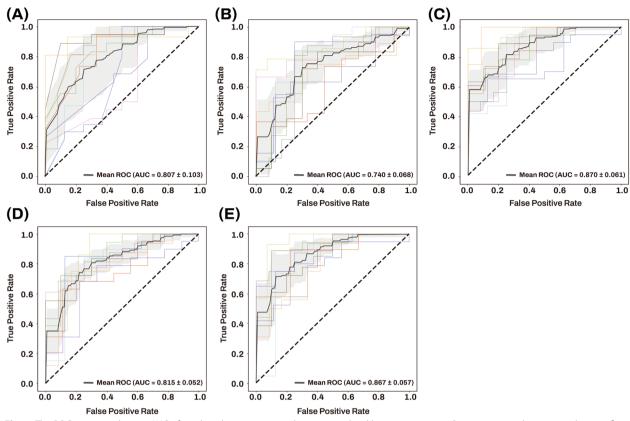


Fig. 7 The ROC curves and mean AUC of machine learning approach using penalized logistic regression. Sensitivity is on the *y*-axis and 1-specificity is on the *x*-axis. 10-fold cross validation was used to investigate and compare the classification performance of five different classification models for differentiating AD from NCI. The gray zone around the mean ROC curve represents ± 1 standard deviation. Five different classification models are shown per following training features. **A** Training features include age, sex, and apolipoprotein E (*APOE*) ε4 carrier status. **B** Training features include five AD-associated hub miRNAs. **C** Training features include age, sex, *APOE* ε4 carrier status, and five AD-associated hub miRNAs. **D** Training features include all 309 miRNAs. **E** Training features include age, sex, *APOE* ε4 carrier status, and all 309 miRNAs. AD Alzheimer's dementia, AUC area under the curve, miRNAs microRNAs, NCI no cognitive impairment, ROC receiver operating characteristic

AD, consistent with previous studies [50, 64]. Notably, increased expression levels of miR-1260 were associated with CERAD pathologic traits and worse cognition, which is another novel finding from our study.

Our pathway-based enrichment analysis revealed that the AD-associated predicted target genes for the five AD-associated hub miRNAs were related to ErbB, AMPK, MAPK, and mTOR signaling pathways. ErbB, involved in various biological processes, such as myelination, neurite outgrowth, cell proliferation, differentiation and protection against apoptosis, is downregulated in Alzheimer's disease due to Aβ-mediated neurotoxicity [66–68]. AMPK, which regulates cell polarity, apoptosis, cell migration, and synaptic plasticity, is activated in Alzheimer's disease, contributing to tauopathy, synaptotoxicity, and dendritic deficits [69–72]. AMPK pathways also play a significant role in regulating glucose balance and metabolism in the brain [73–75], emphasizing their relevance to the glucose metabolism

of the AD-associated miRNAs' module functions in our study. MAPK, a crucial regulator of many cellular biological processes, including autophagy, is significantly upregulated in AD due to A β production and oxidative stress [76–78]. Lastly, mTOR, a conserved serine/ threonine protein kinase, is dysregulated in Alzheimer's disease through A β -induced autophagy impairment, endoplasmic reticulum stress, cell apoptosis, and mitochondrial dysfunction [79–82].

The machine learning analysis for classification of AD from NCI showed that five AD-associated miRNAs significantly improved the performance of demographic information and $APOE\ \epsilon 4$ carrier status for classification of AD from a mean AUC value of 0.807 to that of 0.870.

miRNAs have emerged as promising therapeutic targets in Alzheimer's disease due to their crucial role in regulating the expression levels of target genes involved in Alzheimer's disease pathogenesis [10, 83, 84]. Currently, two types of miRNA targets, miRNA mimics and

anti-miRNAs, are being explored for therapeutic interventions in Alzheimer's disease [10, 77].

Our study has several limitations. Firstly, the limited sample size of the replication cohort might have contributed to the decreased statistical power for replication analysis. Further studies with larger samples are needed to validate our findings. Secondly, the replication dataset lacks neuropathological markers and cognitive information, limiting our investigation for associations with neuropathology and cognition to the ROS/MAP cohort. Thirdly, miRNA expression profiles were generated using different brain regions and different microarray platforms in the discovery and replication cohorts, which may have introduced variability in the results. Lastly, the difference in the definitions of the diagnostic groups (NCI and AD) in the two datasets may have led to a potential confounding factor in the consensus network analysis.

Conclusions

In summary, our network-based approach identified AD pathology and cognition-associated miRNAs. Notably, miR-129-5p and miR-221 were replicated in an independent dataset. The inclusion of AD-associated miR-NAs improved the classification performance of AD from NCI. This integrative network approach can provide insight into AD pathogenesis and highlights these miR-NAs as diagnostic/prognostic biomarkers and potential therapeutic targets for AD. However, further investigations are necessary to elucidate the underlying mechanisms and validate these findings.

Abbreviations

Amyloid-B

AD Alzheimer's dementia

AMP-AD Accelerating Medicines Partnership for Alzheimer's Disease **AMPK** 5' Adenosine monophosphate-activated protein kinase

APOE Apolipoprotein E

APP Amyloid precursor protein

AUC Area under the receiver operating characteristic curve

BP Biological processes

CERAD Consortium to Establish a Registry for Alzheimer's Disease

DAVID Database for Annotation, Visualization and Integrated Discovery

DI PEC Dorsolateral prefrontal cortex

ErbB Erythroblastic leukemia viral oncogene homolog

FDR False discovery rate

GABAergic Gamma-aminobutyric acid-ergic

GEO Gene Expression Omnibus

GO Gene ontology

HMGB1 High-mobility group box 1

Kyoto Encyclopedia of Genes and Genomes KFGG

MAPK Mitogen-activated protein kinase

ME Module eigengene miRNAs MicroRNAs mRNAs Messenger RNAs

mTOR Mammalian target of rapamycin NCI No cognitive impairment NFTs Intracellular neurofibrillary tangles

NPs Neuritic plaques gRT-PCR Real-time reverse transcription PCR ROC Receiver operating characteristic

ROS/MAP Religious Orders Study and Rush Memory and Aging Project TOM

Topological Overlap Matrix

WGCNA Weighted gene co-expression network analysis

YAP1 Yes-associated protein 1 3'-UTR 3'-Untranslated regions

Supplementary Information

The online version contains supplementary material available at https://doi. org/10.1186/s13195-023-01366-8.

Additional file 1. Additional file 2.

Acknowledgements

We would like to thank the databases from the ROS/MAP cohort and GEO for this study. ROSMAP resources can be requested at https://www.radc.rush. edu. This work was supported by the National Research Foundation of Korea grant funded by the Korean government (Ministry of Science and ICT; No. 2020R1C1C1013718), Bio&Medical Technology Development Program of the National Research Foundation (NRF) funded by the Korean government (MSIT) (No. RS-2023-00223501), and Hallym University Medical Center Research Fund. Support for data analysis was provided in part by grants: P30 AG010133, P30 AG072976, R01 AG019771, R01 AG057739, U01 AG024904, R01 LM013463, R01 AG068193, T32 AG071444, U01 AG068057, U01 AG072177, R01 LM012535, U19AG074879, and R01 AG069901. ROSMAP is supported by P30AG10161, P30AG72975, R01AG15819, R01AG17917, U01AG46152, and U01AG61356.

Authors' contributions

SWH contributed to the concept and design of the work, data acquisition, interpretation of data, drafting of the manuscript, statistical analysis, and critical revision of the manuscript for important intellectual content. YHP and KN contributed to the concept and design of the work, data acquisition, interpretation of data, drafting of the manuscript, statistical analysis, critical revision of the manuscript for important intellectual content, and supervision. JMP contributed to the data acquisition, interpretation of data, and statistical analysis. SYK, AJS, and PJB contributed to the interpretation of data and manuscript reviewing. DAB contributed in obtaining funding, study supervision, data acquisition, and manuscript reviewing. All authors read and approved the final manuscript.

Funding

This work was supported by the National Research Foundation of Korea grant funded by the Korean government (Ministry of Science and ICT; No. 2020R1C1C1013718), Bio&Medical Technology Development Program of the National Research Foundation (NRF) funded by the Korean government (MSIT) (No. RS-2023-00223501), and Hallym University Medical Center Research Fund. Support for data analysis was provided in part by grants: P30 AG010133, P30 AG072976, R01 AG019771, R01 AG057739, U01 AG024904, R01 LM013463, R01 AG068193, T32 AG071444, U01 AG068057, U01 AG072177, R01 LM012535, U19AG074879, and R01 AG069901. ROSMAP is supported by P30AG10161, P30AG72975, R01AG15819, R01AG17917, U01AG46152, and U01AG61356.

Availability of data and materials

All data sources mentioned in the study are publicly available summary level information. The ROS/MAP cohort and GEO data will be freely available at the AMP-AD database (https://www.synapse.org) and the National Center for Biotechnology Information as accession numbers GSE157239 (http://www. ncbi.nlm.nih.gov/projects/geo/), respectively.

Declarations

Ethics approval and consent to participate

All data sources mentioned in the study are publicly available summary level information that requires no ethical approval or consent. The Religious Orders Study and Rush Memory and Aging Project (ROS/MAP) studies were each

approved by an Institutional Review Board of Rush University Medical Center. All subjects gave written informed consent, signed a repository consent, and an Anatomic Gift Act. All samples in the GEO dataset were obtained from the Human Brain Bank of the Brazilian Aging Brain Study Group (HBB-BABSG) of the University of São Paulo Medical School, in accordance with the standards of the local ethics committee.

Consent for publication

Not applicable.

Competing interests

The authors declare no competing interests.

Author details

¹Department of Neurology, Chuncheon Sacred Heart Hospital, Hallym University College of Medicine, 77 Sakju-ro, Chuncheon-si, Gangwon-do 24253, Republic of Korea. ²Department of Neurology, Soonchunhyang University Seoul Hospital, 59 Daesagwan-ro, Yongsan-gu, Seoul 03080, Republic of Korea. ³Department of Radiology and Imaging Sciences, Center for Computational Biology and Bioinformatics, Indiana Alzheimer's Disease Research Center, Indiana University School of Medicine, Indianapolis, IN 46202, USA. ⁴Rush Alzheimer's Disease Center, Rush University Medical Center, 1750 W. Harrison St., Suite 1000, Chicago, IL 60612, USA. ⁵Department of Neurology, Seoul National University Bundang Hospital and Seoul National University College of Medicine, 82, Gumi-ro 173 beon-gil, Bundang-gu, Seongnam-si, Gyeonggi-do 13620, Republic of Korea.

Received: 28 October 2023 Accepted: 4 December 2023 Published online: 09 January 2024

References

- Association As. 2012 Alzheimer's disease facts and figures. Alzheimers Dement. 2012;8(2):131–68.
- Long JM, Holtzman DM. Alzheimer disease: an update on pathobiology and treatment strategies. Cell. 2019;179(2):312–39.
- Hampel H, Frank R, Broich K, Teipel SJ, Katz RG, Hardy J, et al. Biomarkers for Alzheimer's disease: academic, industry and regulatory perspectives. Nat Rev Drug Discov. 2010;9(7):560–74.
- Kou X, Chen D, Chen N. The regulation of microRNAs in Alzheimer's disease. Front Neurol. 2020;11:288.
- 5. Wang M, Qin L, Tang B. MicroRNAs in Alzheimer's disease. Front Genet. 2019;10:153.
- Patrick E, Rajagopal S, Wong H-KA, McCabe C, Xu J, Tang A, et al. Dissecting the role of non-coding RNAs in the accumulation of amyloid and tau neuropathologies in Alzheimer's disease. Mol Neurodegener. 2017;12(1):1–13.
- Winter J, Jung S, Keller S, Gregory RI, Diederichs S. Many roads to maturity: microRNA biogenesis pathways and their regulation. Nat Cell Biol. 2009;11(3):228–34.
- 8. Bartel DP. MicroRNAs: genomics, biogenesis, mechanism, and function. cell. 2004;116(2):281–97.
- Arora T, Prashar V, Singh R, Barwal TS, Changotra H, Sharma A, et al. Dysregulated miRNAs in progression and pathogenesis of Alzheimer's disease. Mol Neurobiol. 2022;59(10):1–18.
- Miya Shaik M, Tamargo IA, Abubakar MB, Kamal MA, Greig NH, Gan SH. The role of microRNAs in Alzheimer's disease and their therapeutic potentials. Genes. 2018;9(4):174.
- Lau P, Frigerio CS, De Strooper B. Variance in the identification of microRNAs deregulated in Alzheimer's disease and possible role of lincRNAs in the pathology: the need of larger datasets. Ageing Res Rev. 2014;17:43–53.
- Carter SL, Brechbühler CM, Griffin M, Bond AT. Gene co-expression network topology provides a framework for molecular characterization of cellular state. Bioinformatics. 2004;20(14):2242–50.
- Eisen MB, Spellman PT, Brown PO, Botstein D. Cluster analysis and display of genome-wide expression patterns. Proc Natl Acad Sci U S A. 1998;95(25):14863–8.

- 14. Langfelder P, Horvath S. WGCNA: an R package for weighted correlation network analysis. BMC Bioinformatics. 2008;9(1):1–13.
- Pedersen HK, Forslund SK, Gudmundsdottir V, Petersen AØ, Hildebrand F, Hyötyläinen T, et al. A computational framework to integrate highthroughput '-omics' datasets for the identification of potential mechanistic links. Nat Protoc. 2018;13(12):2781–800.
- Zhang T, Shen Y, Guo Y, Yao J. Identification of key transcriptome biomarkers based on a vital gene module associated with pathological changes in Alzheimer's disease. Aging (Albany NY). 2021;13(11):14940.
- Bennett DA, Schneider JA, Aggarwal NT, Arvanitakis Z, Shah RC, Kelly JF, et al. Decision rules guiding the clinical diagnosis of Alzheimer's disease in two community-based cohort studies compared to standard practice in a clinic-based cohort study. Neuroepidemiology. 2006;27(3):169–76.
- Bennett D, Schneider J, Arvanitakis Z, Kelly J, Aggarwal N, Shah R, et al. Neuropathology of older persons without cognitive impairment from two community-based studies. Neurology. 2006;66(12):1837–44.
- Braak H, Alafuzoff I, Arzberger T, Kretzschmar H, Del Tredici K. Staging of Alzheimer disease-associated neurofibrillary pathology using paraffin sections and immunocytochemistry. Acta Neuropathol. 2006;112(4):389–404.
- Mirra SS, Heyman A, McKeel D, Sumi S, Crain BJ, Brownlee L, et al. The Consortium to Establish a Registry for Alzheimer's Disease (CERAD): part II. Standardization of the neuropathologic assessment of Alzheimer's disease. Neurology. 1991;41(4):479–9.
- Wan Y-W, Al-Ouran R, Mangleburg CG, Perumal TM, Lee TV, Allison K, et al. Meta-analysis of the Alzheimer's disease human brain transcriptome and functional dissection in mouse models. Cell Rep. 2020;32(2):107908.
- Henriques AD, Machado-Silva W, Leite RE, Suemoto CK, Leite KR, Srougi M, et al. Genome-wide profiling and predicted significance of postmortem brain microRNA in Alzheimer's disease. Mech Ageing Dev. 2020;191:111352.
- Kozomara A, Griffiths-Jones S. miRBase: annotating high confidence microRNAs using deep sequencing data. Nucleic Acids Res. 2014;42(D1):D68–73.
- 24. De Jager PL, Ma Y, McCabe C, Xu J, Vardarajan BN, Felsky D, et al. A multiomic atlas of the human frontal cortex for aging and Alzheimer's disease research. Sci Data. 2018;5(1):1–13.
- 25. Zhang Y, Parmigiani G, Johnson WE. ComBat-seq: batch effect adjustment for RNA-seq count data. Genom Bioinform. 2020;2(3):lqaa078.
- Karpenko O, Bahroos N, Chukhman M, Dong X, Kanabar P, Arbieva Z, et al. Galaxy high throughput genotyping pipeline for GeneTitan. AMIA Jt Summits Transl Sci Proc. 2013;2013:102–2.
- 27. Irizarry RA, Hobbs B, Collin F, Beazer-Barclay YD, Antonellis KJ, Scherf U, et al. Exploration, normalization, and summaries of high density oligonucleotide array probe level data. Biostatistics. 2003;4(2):249–64.
- 28. Ringnér M. What is principal component analysis? Nat Biotechnol. 2008;26(3):303–4.
- 29. Wang M, Beckmann ND, Roussos P, Wang E, Zhou X, Wang Q, et al. The Mount Sinai cohort of large-scale genomic, transcriptomic and proteomic data in Alzheimer's disease. Sci Data. 2018;5(1):1–16.
- Trapnell C, Roberts A, Goff L, Pertea G, Kim D, Kelley DR, et al. Differential gene and transcript expression analysis of RNA-seq experiments with TopHat and Cufflinks. Nat Protoc. 2012;7(3):562–78.
- Langmead B, Trapnell C, Pop M, Salzberg SL. Ultrafast and memoryefficient alignment of short DNA sequences to the human genome. Genome Biol. 2009;10(3):1–10.
- 32. Li B, Dewey CN. RSEM: accurate transcript quantification from RNA-Seq data with or without a reference genome. BMC Bioinformatics. 2011;12:1–16.
- 33. Yang H-S, White CC, Klein H-U, Yu L, Gaiteri C, Ma Y, et al. Genetics of gene expression in the aging human brain reveal TDP-43 proteinopathy pathophysiology. Neuron. 2020;107(3):496–508.e496.
- Bennett D, Schneider J, Bienias J, Evans D, Wilson R. Mild cognitive impairment is related to Alzheimer disease pathology and cerebral infarctions. Neurology. 2005;64(5):834–41.
- Schneider JA, Arvanitakis Z, Bang W, Bennett DA. Mixed brain pathologies account for most dementia cases in community-dwelling older persons. Neurology. 2007;69(24):2197–204.
- Wilson RS, Segawa E, Boyle PA, Anagnos SE, Hizel LP, Bennett DA. The natural history of cognitive decline in Alzheimer's disease. Psychol Aging. 2012;27(4):1008.

- Nam J-W, Rissland OS, Koppstein D, Abreu-Goodger C, Jan CH, Agarwal V, et al. Global analyses of the effect of different cellular contexts on microRNA targeting. Mol Cell. 2014;53(6):1031–43.
- 38. Wong N, Wang X. miRDB: an online resource for microRNA target prediction and functional annotations. Nucleic Acids Res. 2015;43(D1):D146–52.
- Huang DW, Sherman BT, Lempicki RA. Systematic and integrative analysis of large gene lists using DAVID bioinformatics resources. Nat Protoc. 2009;4(1):44–57.
- Kanehisa M, Goto S. KEGG: kyoto encyclopedia of genes and genomes. Nucleic Acids Res. 2000;28(1):27–30.
- Botstein D, Cherry JM, Ashburner M, Ball CA, Blake JA, Butler H, et al. Gene Ontology: tool for the unification of biology. Nat genet. 2000;25(1):25–9.
- 42. Li J, Han X, Wan Y, Zhang S, Zhao Y, Fan R, et al. TAM 2.0: tool for MicroRNA set analysis. Nucleic Acids Res. 2018;46(W1):W180–5.
- Neyman J, Pearson ES. On the use and interpretation of certain test criteria for purposes of statistical inference: part I. Biometrika. 1928:20A(1-2):175–240.
- 44. Langfelder P, Horvath S. Eigengene networks for studying the relationships between co-expression modules. BMC Syst Biol. 2007;1:1–17.
- Mueller AJ, Canty-Laird EG, Clegg PD, Tew SR. Cross-species gene modules emerge from a systems biology approach to osteoarthritis. NPJ Syst Biol Appl. 2017;3(1):13.
- Benjamini Y, Hochberg Y. Controlling the false discovery rate: a practical and powerful approach to multiple testing. J R Stat Soc Series B Stat Methodol. 1995;57(1):289–300.
- Urbanowicz RJ, Zhang R, Cui Y, Suri P. STREAMLINE: a simple, transparent, end-to-end automated machine learning pipeline facilitating data analysis and algorithm comparison. ArXiv220612002 2022.
- Teipel SJ, Grothe MJ, Metzger CD, Grimmer T, Sorg C, Ewers M, et al. Robust detection of impaired resting state functional connectivity networks in Alzheimer's disease using elastic net regularized regression. Front Aging Neurosci. 2017;8:318.
- Bouckaert RR, Frank E. Evaluating the replicability of significance tests for comparing learning algorithms. Pacific-Asia conference on knowledge discovery and data mining; 2004: Springer; 2004: 3-12.
- Lau P, Bossers K, Rs J, Salta E, Frigerio CS, Barbash S, et al. Alteration of the micro RNA network during the progression of Alzheimer's disease. EMBO Mol Med. 2013;5(10):1613–34.
- Liu Z, Hu C, Tang D, Luo G. miR-129 Promotes the proliferation of Alzheimer's neuronal cells by binding the 3' untranslated regions (3' UTR) of amyloid precursor protein (APP). J Biomater Tissue Eng. 2022;12(3):551–7.
- Li Z, Chen Q, Liu J, Du Y. Physical exercise ameliorates the cognitive function and attenuates the neuroinflammation of Alzheimer's disease via miR-129–5p. Dement Geriatr Cogn Disord. 2020;49(2):163–9.
- Sun W, Zhao J, Li C. Dexmedetomidine provides protection against hippocampal neuron apoptosis and cognitive impairment in mice with Alzheimer's disease by mediating the miR-129/YAP1/JAG1 axis. Mol Neurobiol. 2020;57(12):5044–55.
- 54. Gupta R, Kumar P. Integrative analysis of OIP5-AS1/miR-129-5p/CREBBP axis as a potential therapeutic candidate in the pathogenesis of metal toxicity-induced Alzheimer's disease. Gene Rep. 2022;26:101442.
- Zhang Y-Y, Bao H-L, Dong L-X, Liu Y, Zhang G-W, An F-M. Silenced IncRNA H19 and up-regulated microRNA-129 accelerates viability and restrains apoptosis of PC12 cells induced by Aβ25-35 in a cellular model of Alzheimer's disease. Cell Cycle. 2021;20(1):112–25.
- Keller A, Gröger L, Tschernig T, Solomon J, Laham O, Schaum N, et al. miRNATissueAtlas2: an update to the human miRNA tissue atlas. Nucleic Acids Res. 2022;50(D1):D211–21.
- He M, Liu Y, Wang X, Zhang MQ, Hannon GJ, Huang ZJ. Cell-typebased analysis of microRNA profiles in the mouse brain. Neuron. 2012;73(1):35–48.
- Pomper N, Liu Y, Hoye ML, Dougherty JD, Miller TM. CNS microRNA profiles: a database for cell type enriched microRNA expression across the mouse central nervous system. Sci Rep. 2020;10(1):4921.
- Manzine PR, Pelucchi S, Horst MA, Vale FA, Pavarini SC, Audano M, et al. microRNA 221 targets ADAM10 mRNA and is downregulated in Alzheimer's disease. J Alzheimers Dis. 2018;61(1):113–23.
- Biglari N, Khalaj-Kondori M, Ghasemi T. Potential of hsa-miR200a-3p and hsa-miR502-3p as blood-based biomarker for Alzheimer's disease. Mol Biol Rep. 2022;49(12):1–8.

- Chanda K, Jana NR, Mukhopadhyay D. Long non-coding RNA MALAT1 protects against Aβ1–42 induced toxicity by regulating the expression of receptor tyrosine kinase EPHA2 via quenching miR-200a/26a/26b in Alzheimer's disease. Life Sci. 2022;302(8):120652.
- Zhang Q-S, Liu W, Lu G-X. miR-200a-3p promotes β-Amyloid-induced neuronal apoptosis through down-regulation of SIRT1 in Alzheimer's disease. J Biosci. 2017;42(3):397–404.
- Schipper HM, Maes OC, Chertkow HM, Wang E. MicroRNA expression in Alzheimer blood mononuclear cells. Gene Regul Syst Bio. 2007;1:GRSB. S361.
- 64. Nagaraj S, Laskowska-Kaszub K, Dębski KJ, Wojsiat J, Dąbrowski M, Gabryelewicz T, et al. Profile of 6 microRNA in blood plasma distinguish early stage Alzheimer's disease patients from non-demented subjects. Oncotarget. 2017;8(10):16122.
- Wang R, Zhang J. Clinical significance of miR-433 in the diagnosis of Alzheimer's disease and its effect on Aβ-induced neurotoxicity by regulating JAK2. Exp Gerontol. 2020;141:111080.
- Jiang Q, Chen S, Hu C, Huang P, Shen H, Zhao W. Neuregulin-1 (Nrg1) signaling has a preventive role and is altered in the frontal cortex under the pathological conditions of Alzheimer's disease. Mol Med Rep. 2016;14(3):2614–24.
- Yang Q, Zhao Q, Yin Y. miR-133b is a potential diagnostic biomarker for Alzheimer's disease and has a neuroprotective role. Exp Ther Med. 2019;18(4):2711–8.
- Chen S, Velardez MO, Warot X, Yu Z-X, Miller SJ, Cros D, et al. Neuregulin 1–erbB signaling is necessary for normal myelination and sensory function. J Neurosci. 2006;26(12):3079–86.
- Liu D, Tang H, Li X-Y, Deng M-F, Wei N, Wang X, et al. Targeting the HDAC2/HNF-4A/miR-101b/AMPK pathway rescues tauopathy and dendritic abnormalities in Alzheimer's disease. Mol Ther. 2017;25(3):752–64.
- Barbato C, Giacovazzo G, Albiero F, Scardigli R, Scopa C, Ciotti M, et al. Cognitive decline and modulation of alzheimer's disease-related genes after inhibition of MicroRNA-101 in mouse hippocampal neurons. Mol Neurobiol. 2020;57:3183–94.
- Chang F, Zhang LH, Xu WP, Jing P, Zhan PY. microRNA-9 attenuates amyloidβ-induced synaptotoxicity by targeting calcium/calmodulindependent protein kinase kinase 2. Mol Med Rep. 2014;9(5):1917–22.
- 72. Li S, Yan Y, Jiao Y, Gao Z, Xia Y, Kong L, et al. Neuroprotective effect of osthole on neuron synapses in an Alzheimer's disease cell model via upregulation of microRNA-9. J Mol Neurosci. 2016;60:71–81.
- 73. Muraleedharan R, Dasgupta B. AMPK in the brain: its roles in glucose and neural metabolism. FEBS J. 2022;289(8):2247–62.
- 74. Zhang S, Lachance BB, Mattson MP, Jia X. Glucose metabolic crosstalk and regulation in brain function and diseases. Prog Neurobiol.
- 75. Lopez M, Nogueiras R, Tena-Sempere M, Dieguez C. Hypothalamic AMPK: a canonical regulator of whole-body energy balance. Nat Rev Endocrinol. 2016;12(7):421–32.
- 76. Li Q, Wang Y, Peng W, Jia Y, Tang J, Li W, et al. MicroRNA-101a regulates autophagy phenomenon via the MAPK pathway to modulate Alzheimer's-associated pathogenesis. Cell Transplant. 2019;28(8):1076–84.
- Hernandez-Rapp J, Rainone S, Goupil C, Dorval V, Smith PY, Saint-Pierre M, et al. microRNA-132/212 deficiency enhances Aβ production and senile plaque deposition in Alzheimer's disease triple transgenic mice. Sci Rep. 2016;6(1):30953.
- Deng Y, Zhang J, Sun X, Ma G, Luo G, Miao Z, et al. miR-132 improves the cognitive function of rats with Alzheimer's disease by inhibiting the MAPK1 signal pathway. Exp Ther Med. 2020;20(6):1–1.
- Zhang W, Yang Y, Xiang Z, Cheng J, Yu Z, Wang W, et al. MRTF-A-mediated protection against amyloid-β-induced neuronal injury correlates with restoring autophagy via miR-1273g-3p/mTOR axis in Alzheimer models. Aging (Albany NY). 2022;14(10):4305.
- Higaki S, Muramatsu M, Matsuda A, Matsumoto K, Satoh J-i, Michikawa M, et al. Defensive effect of microRNA-200b/c against amyloid-beta peptide-induced toxicity in Alzheimer's disease models. PLoS One. 2018;13(5):e0196929.
- Ye X, Luo H, Chen Y, Wu Q, Xiong Y, Zhu J, et al. MicroRNAs 99b-5p/100-5p regulated by endoplasmic reticulum stress are involved in abetainduced pathologies. Front Aging Neurosci. 2015;7:210.

- 82. Liang C, Mu Y, Tian H, Wang D, Zhang S, Wang H, et al. MicroRNA-140 silencing represses the incidence of Alzheimer's disease. Neurosci Lett. 2021;758:135674.
- 83. Angelucci F, Cechova K, Valis M, Kuca K, Zhang B, Hort J. MicroRNAs in Alzheimer's disease: diagnostic markers or therapeutic agents? Front Pharmacol. 2019;10:665.
- 84. Liu S, Fan M, Zheng Q, Hao S, Yang L, Qi C, et al. MicroRNAs in Alzheimer's disease: potential diagnostic markers and therapeutic targets. Biomed Pharmacother. 2022;148:112681.

Publisher's Note

Springer Nature remains neutral with regard to jurisdictional claims in published maps and institutional affiliations.

Ready to submit your research? Choose BMC and benefit from:

- fast, convenient online submission
- $\bullet\,$ thorough peer review by experienced researchers in your field
- rapid publication on acceptance
- support for research data, including large and complex data types
- gold Open Access which fosters wider collaboration and increased citations
- maximum visibility for your research: over 100M website views per year

At BMC, research is always in progress.

Learn more biomedcentral.com/submissions

