

RESEARCH

Open Access



Microfilarial reduction following ProHeart® 6 and ProHeart® SR-12 treatment in dogs experimentally inoculated with a resistant isolate of *Dirofilaria immitis*

Tom L. McTier^{1*}, Aleah Pullins¹, Gregory A. Inskeep¹, Genevieve Gagnon¹, Huihao Fan¹, Adam Schoell¹, Tara Bidgood², Joyce Login² and Patrick Meeus¹

From 15th American Heartworm Society Triennial Symposium
New Orleans, LA, USA. September 11-13, 2016

Abstract

Background: Emerging resistance of heartworms (*Dirofilaria immitis*) to macrocyclic lactone (ML) preventives is an increasing concern for veterinarians, pet owners and animal health companies that supply heartworm preventives, with recent reports of resistant isolates identified from the Mississippi Delta region of the United States. Products that are effective in eliminating microfilariae (MF) in dogs harboring resistant heartworm infections could be important in reducing the spread of heartworm resistance. The current study was conducted to investigate the potential for ProHeart® 6 (PH 6; Zoetis) and ProHeart® SR-12 (PH 12; Zoetis) to reduce MF in dogs experimentally inoculated with an isolate of *D. immitis* (ZoeMo-2012) confirmed to be resistant to MLs.

Methods: Twenty-three dogs with preexisting heartworm infections (via surgical transplantation) were randomly allocated to four groups based on pretreatment (Day -14) MF counts. On Day 0, dogs received a subcutaneous injection of either saline (placebo-treated control, 6 dogs), PH 6 (0.17 mg/kg, 6 dogs), PH 12 (0.5 mg/kg, 5 dogs) or a single oral dose of moxidectin powder in a gelatin capsule (0.25 mg/kg, 6 dogs). All dogs were bled for MF counts (modified Knott's test) on Days 0 (pretreatment), 1, 3, 7, 14, 21, 28, 42, 56, and 84. Dogs in control and PH 6 groups were also bled for MF counts on Days 112, 140, and 168. No adverse events associated with treatment were observed for any dog.

Results: Average reductions in MF counts compared with controls for PH 6 were 9.7% on Day 1, increasing to 75.0% on Day 7, and further to 86.5% on Day 28. On Day 42, average MF reduction increased to 90.3%. Reductions increased further over the next several months with reductions of 91.3, 96.8, 96.6, and 98.9% on Days 56, 84, 112, and 140, respectively. On Day 168, the reduction was 99.3% ($P < 0.0001$). Average reductions in MF counts compared with controls for PH 12 were 20.9% on Day 1, increasing to 78.9% on Day 7, and further to 91.2% on Day 28. On Day 84, the reduction was 96.9%. For dogs receiving a single oral moxidectin (0.25 mg/kg) on Day 0, reductions in MF were 86.3% on Day 1 and fluctuated between 74.4 and 83.6% through Day 28. On Days 42 and 56, percentage reductions were 87.1 and 81.8%, respectively, and 92.6% at the final time point (Day 84).

(Continued on next page)

* Correspondence: tom.mctier@zoetis.com

¹Zoetis, Veterinary Medicine Research and Development, 333 Portage Street, Kalamazoo, MI 49007, USA

Full list of author information is available at the end of the article



(Continued from previous page)

Conclusion: Both PH 6 and PH 12 were highly effective in reducing the MF levels of a confirmed ML-resistant heartworm isolate following a single dose.

Keywords: ProHeart® 6, ProHeart® SR-12, Moxidectin, Macrocytic lactone, Canine heartworm, *Dirofilaria immitis*, Microfilaria, Resistance

Background

Resistance of the canine heartworm, *Dirofilaria immitis*, to macrocyclic lactone (ML) preventive medications is now well documented [1–3]. Much is yet to be learned, however, about the nature and extent of this resistance in natural populations. A better understanding of the epidemiology and transmission dynamics of heartworm in areas where resistant strains of heartworm occur is necessary to estimate how this resistance may spread. Work is being undertaken to gain more information on these factors, but the progress is somewhat slow due to the complicated and multifaceted aspects of this inquiry [4, 5]. Baseline survey data on the current level and geographical distribution of resistance are required, along with factors underlying and contributing to the development and the spread of this resistance.

Reducing the availability of resistant microfilariae in nature for transmission to other competent hosts would be beneficial in reducing the spread of resistant heartworms. Recent publications have shown that several ML-based products that were originally potent microfilaricides failed to substantially reduce or clear some dogs of circulating microfilariae (MF) later identified as resistant [3, 6, 7]. However, there have been no reports on the microfilaricidal activity of ProHeart® 6 (PH 6) (Zoetis) or ProHeart® SR-12 (PH 12) (Zoetis) against resistant heartworm MF. The purpose of the current investigation was to determine the activity of PH 6 and PH 12 in reducing microfilarial levels in dogs surgically implanted with a resistant isolate (ZoeMO-2012) of *D. immitis* (see “*D. immitis* Isolate” following).

Methods

Ethical approval

The study was a masked, negative placebo-controlled, randomized laboratory efficacy study conducted in Michigan, USA. Study procedures were conducted in accordance with the VICH guidelines (GL19) [8]. Masking of the study was assured through the separation of functions. All personnel conducting observations or animal care or performing infestations and counts were masked to treatment allocation. The protocol for this study was approved by the Zoetis Institutional Animal Care and Use Committee (IACUC), and the study was conducted in accordance with state and national/international regulations regarding animal welfare.

Animals

Twenty-five (25) dogs with preexisting adult heartworm infections, established via surgical transplantation (10 pairs of adult heartworms) [9] with an isolate (ZoeMO-2012 isolate), confirmed to be resistant to MLs [10, 11] were available for this study. The dogs had not been treated with a monthly oral preventive dose of a ML for at least 180 days before inoculation with heartworms. Dogs were identified individually by unique numeric ear tattoo or digital microchip. For at least 14 days prior to treatment, dogs were acclimated to the study facilities. For the duration of the study, dogs were housed in individual enclosures, which prevented physical contact with adjacent animals. Dogs were offered water ad libitum and were fed an appropriate standard commercial canine diet. Prior to inclusion in the study, dogs were examined for overall general physical health and study suitability.

D. immitis isolate

The heartworm isolate used in this study was designated ZoeMo-2012. A blood sample was collected from a heartworm-positive dog, originally from Pittsfield, Illinois, USA, on December 4, 2012, which was used to infect *Aedes aegypti* (Liverpool strain) mosquitoes. On December 19, 2012, two dogs were each inoculated with 50 infective larvae (L3) that developed in these mosquitoes. On July 18, 2013, these two dogs were positive for MF on a modified Knott's test, validating passage of this isolate. This isolate was taken from the same parent dog from which the original JYD-34 isolate had been taken 2.5 years earlier (John McCall, personal oral communication, August 2013). The dog had been maintained in mosquito-proof quarters with no additional macrocyclic lactones administered during the intervening time from JYD-34 isolation to ZoeMo-2012 isolation.

Design

On Day -14, 2 weeks prior to treatment, dogs were tested for adult *D. immitis* antigen and were examined by the modified Knott's method for MF. Animals with MF counts >1000 MF/mL and that had positive results on a heartworm antigen (DiroCHEK®; Zoetis) test on Day -14 were included in the study. Two of the 25 dogs available for the study did not meet the minimum requirement for MF on Day -14 and were excluded from the study. The remaining 23 animals were randomly allocated to four treatments based on MF counts as follows: placebo-treated control (six

dogs), ProHeart® 6 (PH 6, six dogs), ProHeart® SR-12 (PH 12, five dogs), and oral moxidectin (six dogs/group).

Treatment

On Day -7, each dog was weighed and given a physical examination. On Day 0, control dogs were administered a single subcutaneous (SC) saline injection (0.05 mL/kg); PH 6 dogs were administered a single SC injection of 0.17 mg/kg body weight (BW) according to label directions; PH 12 dogs were administered a single SC dose of 0.5 mg/kg BW according to label directions; and dogs in the oral moxidectin group were given a single oral dose of 0.25 mg/kg BW of moxidectin in a gelatin capsule (hydroxypropyl methylcellulose moxidectin powder). Without any previous data available on which to select an oral dose of moxidectin for microfilaricidal efficacy against a resistant isolate, the authors chose the highest dose that was estimated could safely be given to MF-positive dogs.

Microfilariae and adult heartworm counts

Dogs in control and PH 6 groups were bled for MF counts on Days 0 (pretreatment), 1, 3, 7, 14, 21, 28, 42, 56, 84, 112, 140, and 168. Dogs treated with PH 12 and oral moxidectin at 0.25 mg/kg were bled for MF counts on Days 0, 1, 3, 7, 14, 21, 28, 42, 56, and 84.

Three control dogs and the dogs in the PH 12 and oral moxidectin groups were necropsied on Day 84 for adult heartworm counts, and the three remaining control dogs and the dogs in the PH 6 group were necropsied for adult worm counts on Day 168. Due to study management constraints and the priority of collecting PH 6 data, data collection for PH 12 was concluded on Day 84. The three control dogs selected to be necropsied on day 84 were randomly selected prior to Day 84.

For recovery of adult heartworms, at the time of euthanasia each dog was given approximately 1 mL of heparin (1000 USP units/mL) intravenously, prior to a lethal-dose euthanasia solution. After euthanasia, the pleural and peritoneal cavities were examined for adult *D. immitis* worms, and the posterior and anterior venae cavae were clamped before removal of the heart and lungs. The precava, right atrium, right ventricle and pulmonary arteries (including

those coursing through the lungs) were dissected and examined for worms. The number and viability of worms recovered from each dog were determined.

Animal observations

Dogs were observed regularly for general health and for adverse events associated with treatment. General health observations (GHOs) were conducted once daily during the acclimation period, twice daily on Days 0–3 (>12 h apart) and for the remainder of the study (>5 h apart). GHOs were conducted when clinical observations (COs) were not performed and included but were not limited to: observations of general physical appearance and behavior, abnormalities of food and water consumption, vomiting/regurgitation and appearance of urine and feces. A suitably experienced veterinarian conducted COs according to the following schedule: Day -14 (± 2 days); Day 0 immediately prior to treatment and 2 to 4 h posttreatment; once daily on Days 1 to 7; once weekly during Days 8 to 84; and once per month during Days 85 to 168. COs included but were not limited to: overall condition, general attitude and cognition, evaluation for vomiting, abnormal feces, abnormal urine, abnormal appetite and any type of hypersensitivity reactions (anaphylaxis, shock, collapse, respiratory distress, depression or fever).

Results

No adverse events associated with treatment with either PH 6, PH 12, or oral moxidectin (0.25 mg/kg) were observed for any dog. Mean MF counts in control dogs were 15,000.0 MF/mL on Day 0, with a somewhat variable but general overall trend toward increasing levels of MF as the study progressed (Table 1). On Day 7, control MF counts dropped to a mean of 12,566.7 MF/mL (the lowest of the study) before rebounding to 14,756.7 MF/mL on Day 14 and further to 23,133.3 MF/mL on Day 112 and finally to 26,633.3 MF/mL on Day 168. It should be noted that all six control dogs were used for percentage reduction calculation for the first 84 days, and for the final 80 days (to Day 168) the remaining three matched control dogs were used for efficacy calculation.

Microfilarial counts for both PH 6 and PH 12 decreased gradually over time compared with those in control

Table 1 Mean microfilariae (MF) counts (per mL) after treatment with a single SC dose of either PH 6 (0.17 mg/kg) or PH 12 (0.5 mg/kg) or a single oral treatment with moxidectin (0.25 mg/kg) on Day 0

Day of Study	0	1	3	7	14	21	28	42	56	84	112	140	168
Control	15,000.0	18,308.3	16,488.3	12,566.7	14,756.7	19,400.0	20,383.3	15,830.0	16,381.7	20,816.7	23,133.3	18,633.3	26,633.3
PH 6	13,916.7	16,531.7	11,796.7	3141.7	4358.3	4318.3	2755.0	1530.0*	1421.7	675.0*	780.0*	203.7*	176.8*
PH 12	11,520.0	14,480.0	6330.0	2656.0	2486.0	2918.0	1786.0	916.2*	997.0*	646.0*	NA	NA	NA
Oral Moxidectin (0.25 mg/kg)	12,033.3	2501.7	3138.3	3213.3	2830.0	3833.3	3333.3	2040.0	2983.3	1531.7	NA	NA	NA

*Significantly different from control mean (P < 0.05)

Table 2 Percentage reduction in mean microfilaria (MF) counts (compared with control) after treatment with a single SC dose of either PH 6 (0.17 mg/kg) or PH 12 (0.5 mg/kg) or a single oral treatment with moxidectin (0.25 mg/kg) on Day 0

Day of Study														
Test Group	0	1	3	7	14	21	28	42	56	84	112	140	168	
PH 6	7.2	9.7	28.5	75.0	70.5	77.7	86.5	90.3	91.3	96.8	96.6	98.9	99.3	
PH 12	23.2	20.9	61.6	78.9	83.2	85.0	91.2	94.2	93.9	96.9	ND	ND	ND	
Oral Moxidectin (0.25 mg/kg)	19.8	86.3	81.0	74.4	80.8	80.2	83.6	87.1	81.8	92.6	ND	ND	ND	

animals (Table 1). Average reductions in MF counts for PH 6 were 9.7% on Day 1, increasing to 28.5 and 75.0% on Day 3 and 7, respectively, and further to 86.5% on Day 28 (Table 2; Fig. 1). On Day 42, average MF reduction increased to >90.3%. Reductions increased further over the next several months with reductions of 91.3, 96.8, 96.6, and 98.9% on Days 56, 84, 112, and 140, respectively. At the final time point (Day 168), the reduction was 99.3%. Mean MF counts decreased from 13,916.7 (range: 800–34,500 MF/mL) on Day 0 to 176.8 on Day 168 (range: 1–490 MF/mL) in PH 6 dogs (Table 1). None of the PH 6 dogs was ever negative for MF, but one dog had a single MF on Day 168. Mean MF counts for PH 6 were significantly lower ($P < 0.05$) than control counts on Days 42, 84, 112, 140, and 168 (Table 2).

Reductions in MF counts were similar for PH 6 and PH 12 (Table 2; Fig. 1). Average reductions in MF counts compared with controls for PH 12 were 20.9% on Day 1, increasing to 61.6 and 78.9% on Days 3 and 7, respectively and further to 91.2% on Day 28. On Days 42 and 56, MF reductions were 94.2 and 93.9%, respectively, increasing slightly to 96.9% on Day 84 (the last time point for this group). For PH 12, MF counts were similar to those for PH 6, with initial mean counts at 11,520 MF/mL and decreasing to 646 MF/mL (675 MF/mL for PH 6) on Day 84. One dog had 0 MF on Days 56 and 84, despite having a high initial MF level (31,000 MF/mL on Day 0). Mean MF counts for PH 12 were significantly lower ($P < 0.05$) than control counts on Days 42, 56, and 84 (Table 2).

For dogs receiving a single oral moxidectin treatment (0.25 mg/kg) on Day 0, MF levels decreased more rapidly immediately after treatment compared to those for PH 6 and PH 12, with a mean reduction of 86.3% on Day 1. However further reductions were not observed during the following 2 months. On Day 84 (the final time point), the reduction was 92.6%. In addition, mean MF counts for oral moxidectin were not significantly lower ($P < 0.05$) than control counts on any of the count days.

The dogs treated with PH 12 and oral moxidectin and three control dogs were necropsied on Day 84 and the dogs treated with PH 6 and three controls were necropsied on Day 168 for recovery of adult heartworms (Table 3). A mean of 15.3 and 16.7 of the 20 worms/dog initially transplanted were recovered from the control and PH 6-treated, respectively, indicating no effect of PH 6 on adult heartworms over the 168-day study period. The numbers of heartworms in the PH 12 and oral moxidectin groups were reduced by 19 and 14%, respectively; and there did appear to be a greater reduction of the female worms (30 and 23%, respectively) compared with controls. However, these reductions were not statistically different ($P > 0.05$).

Discussion

These are the first reported data showing the microfilaricidal effects of ProHeart® (PH 6 and PH 12) on a resistant isolate of *D. immitis* in the dog. Lack of efficacy of approved monthly preventive products containing selamectin, milbemycin oxime and ivermectin against the JYD-34 isolate has

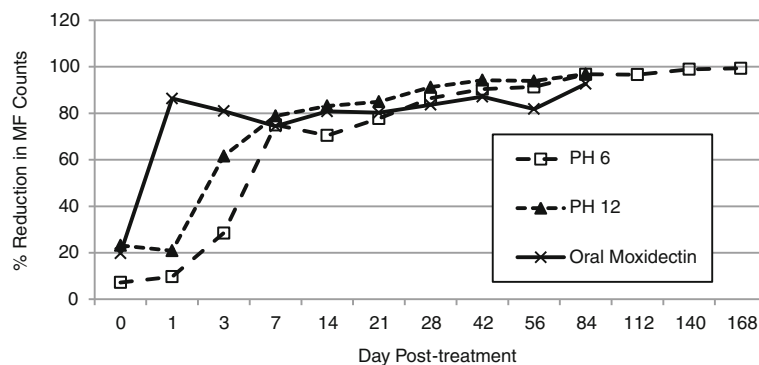


Fig. 1 Percentage reduction in microfilarial counts of a resistant isolate of *Dirofilaria immitis* (ZoeMo-2012) after treatment with a single SC dose of either PH 6 (0.17 mg/kg) or PH 12 (0.5 mg/kg) or a single oral treatment with moxidectin (0.25 mg/kg) compared with controls

Table 3 Mean adult heartworm counts and percentage reductions at necropsy (compared with control) after treatment with a single SC dose of either PH 6 (0.17 mg/kg) or PH 12 (0.5 mg/kg) or a single oral treatment with moxidectin (0.25 mg/kg) on Day 0

Test Group	Values	Live Adult <i>D. immitis</i> Counts ³		
		Males	Females	Total
Control ¹	Mean	7.7	7.7	15.3 ^a
PH 6 ¹	Mean	8.2	8.5	16.7 ^a
	% Reduction	0%	0%	0%
Control ²	Mean	8.7	10.0	18.7 ^a
PH 12 ²	Mean	7.7	6.5	14.9 ^a
	% Reduction	5%	30%	19%
Control ²	Mean	8.7	10.0	18.7 ^a
Oral Moxidectin (0.25 mg/kg) ²	Mean	8.3	7.7	16.0 ^a
	% Reduction	4%	23%	14%

¹Necropsied on Day 168²Necropsied on Day 84³Surgically transplanted with 10 pairs of heartworms ~12 weeks prior to treatment^aMeans with the same superscripts and not statistically different ($P > 0.05$)

been previously demonstrated [2], suggesting that this isolate is resistant to approved doses of some MLs. Additional genetic analysis of markers associated with resistance [1] along with evidence of heritable resistance characteristics [3] have allowed us to confirm this resistance.

Subsequently, we have confirmed that both the JYD-34 and ZoeMo-2012 isolates, as well as several other recent field isolates (ZoeLA and AMAL), are ML-resistant through both phenotypic testing in dogs using an oral preventive dose (3 µg/kg) of moxidectin [10] and by genetic testing of the isolates using genetic marker analysis [11].

All MLs have demonstrated microfilaricidal effects on susceptible microfilariae [12–23], and one of these MLs in a product (Advantage Multi®; Bayer) with the active ingredient moxidectin (2.5%, topical) has a claim for this indication [24]. However, none of these products, except ProHeart® 6 and ProHeart® SR-12 reported here, has demonstrated microfilaricidal potency against a resistant heartworm isolate. A product that has the ability to reduce MF levels of resistant strains substantially could be useful in reducing the overall risk of transmission of these strains in a natural population where resistant heartworm strains occur.

Additional surveillance work on the baseline prevalence of heartworm resistance, along with periodic monitoring across the United States, is needed to understand more completely the risk that populations of animals have to exposure to resistant heartworms; and some of this work has already begun [4].

Conclusions

Both PH 6 and PH 12 reduced microfilarial levels of a confirmed resistant isolate of *D. immitis* (ZoeMO-12) by >92% at 84 days and for PH 6 by >99% at 168 days after a single subcutaneous injection.

Abbreviations

COs: Clinical observations; GHOs: General health observations; MF: Microfilariae; ML: Macrocytic lactone; SC: Subcutaneous

Funding

The work reported herein was funded by and conducted under the direction of Zoetis Inc. The article's publication fee was funded by the American Heartworm Society.

Availability of data and materials

All data generated or analyzed during this study are included in this published article.

About this supplement

This article has been published as part of *Parasites and Vectors* Volume 10 Supplement 2, 2017: Proceedings of the 15th American Heartworm Society Triennial Symposium 2016. The full contents of the supplement are available online at <https://parasitesandvectors.biomedcentral.com/articles/supplements/volume-10-supplement-2>.

Authors' contributions

All authors participated in study and protocol design and reviewed and approved the manuscript. HF conducted the statistical analysis.

Ethics approval

All animal work was performed at Zoetis Inc., and all protocols for these studies were approved by the appropriate animal welfare committees or governing authorities; and studies were conducted in accordance with state and national/international regulations regarding animal welfare.

Consent for publication

Not applicable.

Competing interests

All authors are current employees of Zoetis.

Publisher's Note

Springer Nature remains neutral with regard to jurisdictional claims in published maps and institutional affiliations.

Author details

¹Zoetis, Veterinary Medicine Research and Development, 333 Portage Street, Kalamazoo, MI 49007, USA. ²Veterinary Operations, Zoetis, 10 Sylvan Way, Parsippany, NJ 07054, USA.

Published: 9 November 2017

References

- Bourguinat C, Lee A, Lizunda R, Blagburn B, Liotta J, Kraus M, et al. Macrocytic lactone resistance in *Dirofilaria immitis*: failure of heartworm preventives and investigation of genetic markers for resistance. *Vet Parasitol.* 2015;210:167–78.
- Blagburn BL, Arther RG, Dillon AR, Butler JM, Bowles JV, Newton JC, et al. Efficacy of four commercially available heartworm preventive products against the JYD-34 laboratory strain of *Dirofilaria immitis*. *Parasit Vectors.* 2016;9:191.
- Pulaski CN, Malone JB, Bourguinat C, Prichard R, Geary T, Ward DR, et al. Establishment of macrocytic lactone resistant *Dirofilaria immitis* isolates in experimentally infected laboratory dogs. *Parasit Vectors.* 2014;7:494.
- Geary T, Pulaski C, Ballesteros C, Keller K, Prichard R. Correlating genotype with phenotypic response to a macrocytic lactone in *Dirofilaria immitis*: a status update. In: Abstract presented at the American heartworm society 15th triennial symposium. New Orleans, LA; 2016.
- Pulaski C. Macrocytic lactone resistance in *Dirofilaria immitis*: the next phase in understanding this complex and still contentious issue [published abstract]. *American Association of Veterinary Parasitologists, San Antonio, TX, 2016; August 6–9.*
- Geary TG, Bourguinat C, Prichard RK. Evidence for macrocytic lactone anthelmintic resistance in *Dirofilaria immitis*. *Top Companion Anim Med.* 2011;26:186–92.
- Bourguinat C, Keller K, Bhan A, Peregrine A, Geary T, Prichard R. Macrocytic lactone resistance in *Dirofilaria immitis*. *Vet Parasitol.* 2011;181:388–92.
- Efficacy of anthelmintics: specific recommendations for canines. VICH GL19 (Anthelmintics: Canine). June 2001. International Cooperation on Harmonisation of Technical Requirements for Registration of Veterinary Medicinal Products.
- Rawlings CA, McCall JA. Surgical transplantation of adult *Dirofilaria immitis* to study heartworm infection and disease in dogs. *Am J Vet Res.* 1985;46(1): 221–4.
- McTier TL, Six RS, Pullins A, Chapin S, McCall JW, Rugg D, et al. Efficacy of oral moxidectin against susceptible and resistant isolates of *Dirofilaria immitis* in dogs. *Parasit Vectors.* 2017;10(Suppl 2). doi:10.1186/s13071-017-2429-5.
- Prichard RK, McTier T, Woods DJ, Keller K, Bourguinat K. Genetic profile of seven *Dirofilaria immitis* isolates with variable response profiles to heartworm preventives. *Parasit Vectors.* 2017;10(Suppl 2). doi:10.1186/s13071-017-2428-6.
- Schlotthauer JC, Stromberg BE, Paul AJ, Todd KS, McCall JW, Dzimiński MT, et al. Safety and acceptability of ivermectin in dogs with acquired patent infection of *Dirofilaria immitis*. In: Otto GF, editor. *Proceedings of the Heartworm Symposium.* Washington, DC: American Heartworm Society; 1986. p. 29–35.
- Bradley R. Dose titration and efficacy of milbemycin oxime for prophylaxis against *Dirofilaria immitis* infection in dogs. In: Otto GF, editor. *Proceedings of the Heartworm Symposium.* Washington, DC: American Heartworm Society; 1989. p. 115–20.
- Grieve RB, Frank GR, Stewart VA, Parsons JC, Abraham D, MacWilliams PS, et al. Effect of dosage and dose timing on heartworm (*Dirofilaria immitis*) chemoprophylaxis with milbemycin. In: Otto GF, editor. *Proceedings of the Heartworm Symposium.* Washington, DC: American Heartworm Society; 1989. p. 121–4.
- Blagburn BL, Hendrix CM, Lindsay DS, Vaughan JL, Hepler DI. Post-adulticide milbemycin oxime microfilaricidal activity in dogs naturally infected with *Dirofilaria immitis*. In: Soll MD, editor. *Proceedings of the Heartworm Symposium.* Batavia, IL: American Heartworm Society; 1992. p. 159–64.
- Bowman DD, Johnson RC, Ulrich ME, Neumann N, Lok JB, Zhang Y, et al. Effects of long-term administration of ivermectin and milbemycin oxime on circulating microfilariae in dogs with naturally acquired infections. In: Soll MD, editor. *Proceedings of the Heartworm Symposium.* Batavia, IL: American Heartworm Society; 1992. p. 151–8.
- Bowman DD, Neumann NR, Rawlings C, Stansfield DG, Legg W. Effects of avermectins on microfilariae in dogs with existing and developing heartworm larvae. In: Seward RL, editor. *Recent Advances in Heartworm Disease: Symposium.* Batavia, IL: American Heartworm Society; 2001. p. 173–8.
- Hendrix CM, Blagburn BL, Bowles JV, Spano JS, Aguilar R. The safety of moxidectin in dogs infected with microfilariae and adults of *Dirofilaria immitis*. In: Soll MD, editor. *Proceedings of the Heartworm Symposium.* Batavia, IL: American Heartworm Society; 1992. p. 183–7.
- Lok JB, Knight DH, LaPaugh DA, Zhang Y. Kinetics of microfilaremia suppression in *Dirofilaria immitis*-infected dogs during and after a prophylactic regimen of milbemycin oxime. In: Bradley RE, editor. *Proceedings of the Heartworm Symposium.* Batavia, IL: American Heartworm Society; 1992. p. 143–9.
- McCall JW, Ryan WG, Roberts RE, Dzimiński MT. Heartworm adulticidal activity of prophylactic doses of ivermectin (6 mcg/kg) plus pyrantel administered monthly to dogs. In: Seward RL, editor. *Recent Advances in Heartworm Disease: Symposium.* Batavia, IL: American Heartworm Society; 1998. p. 209–15.
- McCall JW, Supakorndej P, Dzimiński MT, Supakorndej N, Mansour A, Jun JJ, et al. Evaluation of retroactive and adulticidal activity of moxidectin canine SR (sustained release) injectable formulation against *Dirofilaria immitis* in beagles. In: Seward RL, editor. *Recent Advances in Heartworm Disease: Symposium.* Batavia, IL: American Heartworm Society; 2001. p. 165–72.
- McCall JW. The safety-net story about macrocytic lactone heartworm preventives: a review, an update, and recommendations. *Vet Parasitol.* 2005; 133:197–206.
- McTier TL, Shanks DJ, Watson P, McCall JW, Genchi C, Six RH, et al. Prevention of experimentally induced heartworm (*Dirofilaria immitis*) infections in dogs and cats with a single topical application of selamectin. *Vet Parasitol.* 2000;91:259–68.
- McCall JW, Arther R, Davis W, Settje T. Safety and efficacy of 10% imidacloprid + 2.5% moxidectin for the treatment of *Dirofilaria immitis* circulating microfilariae in experimentally infected dogs. *Vet Parasitol.* 2014; 206:86–92.

Submit your next manuscript to BioMed Central and we will help you at every step:

- We accept pre-submission inquiries
- Our selector tool helps you to find the most relevant journal
- We provide round the clock customer support
- Convenient online submission
- Thorough peer review
- Inclusion in PubMed and all major indexing services
- Maximum visibility for your research

Submit your manuscript at
www.biomedcentral.com/submit

