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Anti-cancer activity of new benzyl isoquinoline alkaloid from Saudi plant *Annona* squamosa

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Abstract

Two alkaloids, belonging to benzylisoquinoline alkaloids, were isolated from *Annona squamosa*. One of these alkaloids, 6, 7-dimethoxy-1- $(\alpha$ -hydroxy-4-methoxybenzyl)-2-methyl-1, 2, 3, 4-tetrahydroisoquinoline, was isolated for the first time from natural sources, while, the alkaloid, Coclaurine was known in *Annona squamosa* L. Moreover, the isolated alkaloids tested for the anti-cancer activities on various cell lines (HepG-2, MCF-7, and HCT-116).

Keywords: *Annona squamosa*, Annonacea, (6, 7-dimethoxy-1-(α-hydroxy-4-methoxybenzyl)-2-methyl-1, 2, 3, 4-tetrahydroisoquinolin, Coclaurine

Introduction

Annona L. belongs to the family Annonaceae which is a large family comprising about 135 genera and more than 2500 species distributed mainly in tropical and subtropical regions [1-3]. Annona genus includes approximately 162 species of trees, shrubs and, rarely lianas [4]. Some species of Annona are of economic importance because of their edible fruits and medicinal properties, like Annona squamosa (sugar apple), Annona muricata (soursop), Annona reticulata (custard-apple) and Annona cherimola (cherimoya) [2]. Only one of Annona genus Annona squamosa was reported in Saudi Arabia [4]. Annona squamosa small tree, 2-3 m tall. Leaves without stipules, petiolate, alternate, lanceolate to elliptic-oblong, $8-13\times3-6$ cm, entire. Flowers solitary or in clusters of 2-4, arising opposite the leaves, borne on a recurved pedicel. Perianth segments in 3 s. Narrowly triangular, green to yellowish brown. Stamens were numerous. Carpels united into a fleshy mass in fruit. Seeds brown, surrounded by white, sweet pulp [4, 5].

Traditionally, all parts of *A. squamosa* are used by different ethnic communities for the treatment of various

chronic diseases such as cancerous tumors, insect bites and other skin complaints [5–8]. However, the seeds powder is toxic and used to kill head lice and fleas [5, 9]. The leaves used for a long time as ant- diabetics, antiulcer, anti-depressants, anti-inflammatory, antimicrobial and antifungal [10–15]. It has also used as Immunomodulatory and hepatoprotective [5, 9, 16]. Also, it used as fertility control [17].

Constituents of *Annona squamosa* have chemical compounds approximately belongs to all natural products compounds steroid, terpenoids, glycoside, alkaloid, flavonoid saponin and phenolic compounds [5, 9, 16].

All previous study done on the anti-cancer activity of *A. squamosa* were dealing with the crude extract that contain all chemical constituents of *A. squamosa* or with non-alkaloidal parts especially acetogenin [18, 19]. Because of there are several natural products have which have anti-cancer activity contain N-atom in there skeleton reported in literature [20–22] and no previous study dealing with the anticancer activity of alkaloids part of *A. squamosa*, so in this research, we isolate some pure alkaloids from this plant also, we study the anti-cancer activity of the isolated alkaloids.

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Results and discussion

Chemical analysis of Annona squamosal

Chemical investigation of alkaloidal part of *A. squamosa* from Saudi origin resulted in the isolation of two benzylisoquinoline alkaloids, namely6, 7-dimethoxy-1- $(\alpha$ -hydroxy-4-methoxybenzyl)-2-methyl-1, 2, 3, 4-tetrahydroisoquinolin.1 and Coclaurine 2 (Fig. 1). The first one isolated for the first time from nature, while coclaurine were isolated before from *A. squamosa*.

Structure elucidated for the isolated compounds 6, 7-dimethoxy-1-(α-hydroxy-4-meth oxybenzyl)-2-methyl-1, 2, 3, 4-tetrahydroisoquinolin (I)

The 1 H-NMR of compound (I) (Table 1, see Additional file 1) shows two doublets in the aromatic region each one integrating for two protons which indicates parasubstituted benzene ring. The first doublet resonates at δ =7.028 ppm (J=10.5 Hz) assigned for 2' and 6' while; another doublet appears at δ =6.838 ppm (J=10.5 Hz) assigned for 3' and 5'. Also, two singlet appears at δ =6.838 ppm and δ =6.649 ppm assigned for H-5 and H-8 respectively. The down-filled shift of H-8 due

Table 1 ¹H and ¹³C NMR spectroscopic data for 1 and 2

No.	1 ^a		2 ^b	
	δ _C type	δ _H mult. (J)	δ _C type	δ _H mult. (J)
1	77.230	5.610 br s	55.157	4.427 m
3	d	3.00-4.00 m	38.95	3.35, 3.25 c
4	29.715	3.00-4.00 m	24.88	2.962, 2.858 m
4a	d	_	122.72	_
5	111.109	6.849 s	115.35	6.727 s
6	147.196	_	147.05	_
7	149.079	_	144.81	_
8	110.966	6.649 s	11.85	6.563 s
8a	d	_	124.72	_
α	d	-	38.79	3.152,3.025dd (18,8)
1'		_	126.31	_
2′	131.342	7.028 d (10.5)	130.566	7.132 d (10)
3'	114.072	6.838 d (10.5)	111.813	6.786 d (10)
4′	158.906	_	156.33	_
5′	114.072	6.838 d (10.5)	111.813	6.786 d (10)
6′	131.342	7.028 d (10.5)	130.566	7.132 d (10)
6-OCH₃	55.455	3.884 s	55.526	3.752 s
7-OCH ₃	55.326	3.798 s	_	_
4'-OCH ₃	55.906	3.468 s	_	_
N-CH ₃	40.046	2.885 s	_	_

^a Data were recorded in CDCI₃ 500 MHz (1H) and 75 MHz (¹³C)

to the inter hydrogen bonding between the hydroxyl group and the tertiary nitrogen which was confirmed by the IR spectroscopy. The hydroxyl methine proton resonates at $\delta = 5.610$ ppm as broad singlet while the proton of hydroxyl group appears as a broad singlet at δ = 5.171 ppm. The protons for C-3 and C-4 overlapped between 3-4 ppm. The spectrum shows two singlets integrating for 3 protons at $\delta = 3.884$ ppm and $\delta = 3.798$ ppm for methoxy group at C-6 and C-4' respectively moreover, the intermolecular hydrogen bonding between the hydroxyl group and the tertiary nitrogen can affect the protons of methoxy group at C-7 which is downfield shifted to $\delta = 3.468$ ppm. The N-methyl protons resonate as a singlet at $\delta\!=\!2.885$ ppm. The $^{13}\text{C-NMR}$ (see Additional file 1) shows 13 peaks, 7 of them for aromatic carbons, 3 peaks resonate at $\delta = 55.455$, 55.326 and 55.906 ppm assigned for methoxy carbon, the N-methyl carbon appears at $\delta = 40.046$ ppm also, the DEPT experiment confirms the presence of 5 methyl groups, four methines in aromatic region and 1 methylene carbon, too, one methine appears in upfield region resonate at $\delta = 77.230 \text{ ppm}.$

The COSY experiment shows a good correlation between the C-3 protons and the C-4 protons, also, good correlations between para-substituted benzene ring protons. The most significant correlation is between C-1 proton and C- α proton (Fig. 2).

The chemical shifts of the different carbons of compound 1 assigned with the help of HMQC and HMBC experiments. The HMBC shows some good correlations between the methoxy protons and the aromatic carbons to which the methoxy groups are attached. Also, shows a good correlation between the protons of C- α and C-1′, C-2′, C-6′and C-8 (Fig. 3).

Coclaurine

The 1 H-NMR of compound (I) (Table 1, see Additional file 1) shows two singlets resonate at δ =6.727 ppm and δ =6.563 ppm assigned for C-5 and C-8 respectively. Also, two doublets each integrating for 2 protons appears at δ =7.132 ppm (J=10 Hz) and δ =6.786 ppm (J=10 Hz) indicating a para disubstituted benzene ring assigned for protons of ring C. The C-1protons resonate at δ =4.427 ppm as multiplet while, the two protons of C- α appears as two doublets at δ =3.152 ppm (J=18, 8 Hz) and at δ =3.025 ppm (J=18,10 Hz). The first coupling due to geminal coupling between the C- α while the second coupling with the C-1 proton.

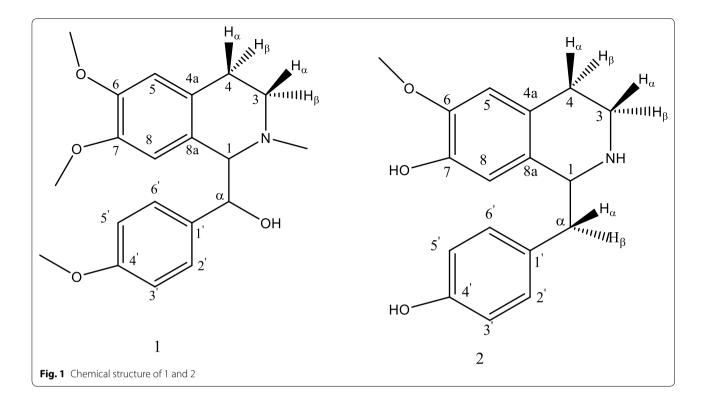
The ¹³C-NMR spectra (see Additional file 1) show 15 peaks for 17 C-atom indicting the presence of para disubstituted benzene ring in the compounds the DEPT experiments revealed this since the DEPT 135 and DEPT 90

^b Data were recorded in DMSO-d₆ at 500 MHz (¹H) and 75 MHz (¹³C)

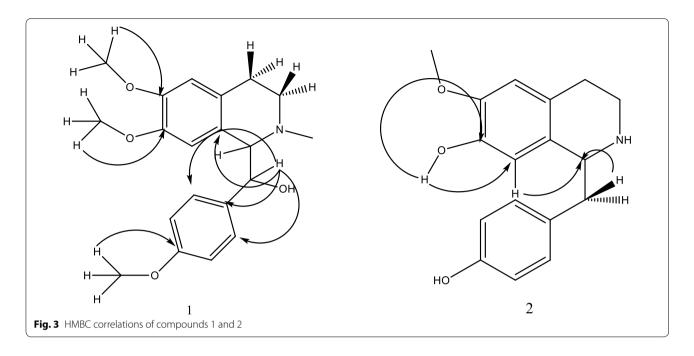
^c Overlapped

^d Not detected but appears in HMBC

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shows four tertiary carbons and six quaternary carbons in aromatic region. Also, shows one methyl, two methylene and one methine carbons in aliphatic region.

The COSY experiments show good correlations between the two $C-\alpha$ protons and C-1 proton (Fig. 2).

The chemical shifts of the different carbons of compound 2 assigned with the help of HMQC and HMBC experiments. The HMBC shows good correlations between O–H proton which resonate at δ =9.447 ppm with C-4′ and both of symmetric carbons C-3′ and C-5′. On the other hand, another O–H proton shows a good correlation with C-7 and C-8 which indicate that the hydroxyl group is attached to C-7 rather than C-6. Also, one of the important correlations seen in HMBC spectra is the correlation between the C-1 carbon and the proton of C-8 and protons of C- α (Fig. 3).

Anti-cancer activity

Anti-cancer activity of *Annona* plants reported in many documents, here, in this research, we study the anti-cancer activity of purely isolated alkaloids. In our study, we use three types of cancer cell line namely: Human Colon cancer cells (HCT116), Human Brest cancer cells (MCF-7) and Human Liver cancer cells (HEPG-2). Table 2 shows the IC50 of Coclaurine and compound 2 against the mentioned cell line. The two isolated compounds gave an excellent activity on the three cell line; also, the two compounds show the most activity against HepG-2, but coclaurine shows a better activity than compound 2 this result on isolated compounds is in confident with structure—activity relationships studies of anti-cancer

Table 2 IC50 of tested compounds

IC50 μg/mL	Tested extract (compound)			
	Colon cancer cells (HCT116)	Human brest cancer cells (MCF-7)	Human liver cancer cells (HEPG-2)	
Coclaurine	8.233	15.345	1.674	
Compound 2	12.344	21.586	5.195	
Doxorubicin	0.8105	1.358	0.777	

activity of benzylisoquinoline alkaloids. The SAR studies show that the increase in the number of hydroxyl groups in the BIQ alkaloids increase the anti-cancer activity, on the other hand, methylation of nitrogen atom decrease the anti-cancer activity [22].

Conclusion

Chemical analysis of the alkaloidal part of *A. squamosa*, afforded two alkaloids belong to simple benzylisoquinoline alkaloid class. One of them is reported from natural sources for the first time.

Isolated alkaloids gave an excellent activity on Colon cancer cells (HCT116) and Human Brest cancer cells (MCF-7), which is confident with the reported structure—activity relationship of activity of benzylisoquino-line alkaloids on a cancer cell.

This result supports using the plant in folk medicine to treat cancer.

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Authors recommended to make total synthesis of the isolated alkaloids.

Materials and methods

Chemicals and materials

All chemicals were purchased from Sigma-Aldrich. cell culture vessels were supplemented from Nunc Co. (Roskilde, Denmark). Human colon (HCT 116), Human liver (HepG-2) and Human breast (MCF-7) cancer cell lines were purchased from Vacsera (Giza, Egypt). Cells were maintained routinely in RPMI 1640 cell culture media supplemented with 1 mM sodium pyruvate, 2 mM L. glutamine, 100 units/mL penicillin–streptomycin and 10% fetal bovine serum. Cells were incubated in a humidified, 5% CO₂ atmosphere at 37 °C.

¹H-NMR spectra were recorded on a Bruker DPX-500 MHz spectrometer with TMS as an internal standard. ¹³C-NMR spectra were recorded at 125.8 MHz using the same instrument.

Plant material

The aerial parts *A. squamosa*, Annonaceae, were collected from Jizan region in Kingdome of Saudi Arabia in January 2017. The plant material was identified in biology department of King Khalid University.

Preparation of plant material:

The aerial parts of the plant were dried in the shade for 15 days then ground to get 6 kg fine powder. The powder was soaked in petroleum ether of 10 days for defatting then extracted thoroughly with ethanol four times each time needs 7 days later the ethanol was evaporated to get a 650 g residue.

Preparation of alkaloidal extract

The ethanol residue was dissolved in 5% HCl until the PH=2 of the solution and filtered, the precipitate which contains neutral material was kept for further fractionation, and the filtrate which provides the basic material was basify using NH4OH solution, and the PH of the solution becomes around 8. After that, the solution was extracted with chloroform 500 mL three times the chloroform layer was evaporated to get 7.35 g of crude alkaloids which represent 0.123% of the dry plant.

Chromatography of crude extract

The crude alkaloids were subjected to silica gel column chromatography using a column packed in chloroform and polarity increasing using methanol till pure methanol was used. The fractions collected (60 fractions, 0.25 L each) were grouped according to their TLC behavior into

six groups. Fraction III gave upon treatment of methanol a yellowish amorphous solid I (30 mg). Fraction V gave a dark brown amorphous solid when treated with methanol this solid were recrystallize by methanol to provide a solid white II (50 mg).

Physical and spectral data of isolated compounds from *Annona sequamosa*

Compound I: Yellowish amorphous solid, IR (KBr) v_{max} (cm $^{-1}$): 3393, 2927, 2852, 1612, 1514. 1 H-NMR and 13 C-NMR data in Table 1.

Coclaurine: Compound II was crystallised from MeOH as white powder, m.p. 254–256d °C, ¹H-NMR and ¹³C-NMR data in Table 1.

Drug dose preparation

0.01 g of each pure compounds was diluted in 1 mL of (DMSO) dimethyl sulfoxide as a stock solution.

Anti-cancer activity of isolated compounds from *Annona* sequamosa

In the present study, SulphoRhodamine-B (SRB) assay had been chosen to detect the anticancer activity of isolated alkaloids. The anticancer activity of isolated alkaloids was tested against Human breast (MCF-7), Human colon (HCT 116), and Human liver (HepG-2) cancer cell lines. Cancer cells were exposed to a range of concentrations (0.01 to 100 μ g/mL) of alkaloids and incubated in 5% CO₂ humidified incubator at 37 °C for 72 h. Doxorubicin was used as a positive control. Cells were treated with the extracts for 72 h then; they were fixed with TCA (10%) for 1 h at 4 °C. To remove TCA cells were washed many times, then 0.4% SRB solution was used to stain cells in a dark place for 10 min. Stained cells were washed with 1% glacial acetic acid. Finally, to dissolve SRB-stained cells, Tris-HCl was used. After drying overnight, the color intensity of remained cells was measured at 540 nm by Elisa.

Statistical analysis

The IC_{50} calculation was performed using Sigma Plot version 12.0

Additional file

Additional file 1: Figure S1. ¹³C-NMR spectra of 6, 7-dimethoxy-1-(a-hydroxy-4-methoxybenzyl)-2-methyl-1, 2, 3, 4-tetrahydroisoquinoline. Figure S2. ¹³C-DEPT90 spectra of 6, 7-dimethoxy-1-(a-hydroxy-4-methoxybenzyl)-2-methyl-1, 2, 3, 4-tetrahydroisoquinoline. Figure S3. ¹³C-DEPT135spectra of 6, 7-dimethoxy-1-(a-hydroxy-4-methoxybenzyl)-2-methyl-1, 2, 3, 4-tetrahydroisoquinoline. Figure S4. ¹H-NMR spectra of 6,

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7-dimethoxy-1-(α-hydroxy-4-methoxybenzyl)-2-methyl-1, 2, 3, 4-tetrahydroisoquinoline. **Figure S5.** ¹³C-NMR spectra of Coclaurine. **Figure S6.** ¹³C-DEPT 90 spectra of Coclaurine. **Figure S7.** ¹³C-DEPT 135 spectra of Coclaurine. **Figure S8.** ¹H-NMR spectra of Coclaurine

Authors' contributions

The author read and approved the final manuscript.

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Competing interests

The author declare no conflicts of interest.

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