STUDY PROTOCOL

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Effects of oral *Nigella sativa* oil on the expression levels and serum concentrations of adiponectin, PPAR-γ, and TNF-α in overweight and obese women: a study protocol for a crossover-designed, double-blind, placebo-controlled randomized clinical trial



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Abstract

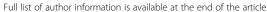
Background: Obesity is a major public health problem in recent decades. The accumulation of excessive fat promotes inflammatory status. Meanwhile, herbal products are marketed for their weight-loss properties, such as *Nigella sativa* (N. *Sativa*) which has been used for centuries to treat rheumatoid arthritis, diabetes, and asthma; recently, the anti-obesity characteristics of N. *sativa* have also been indicated. However, the exact mechanisms and cellular-related pathways are still unclear. Thus, we will aim to assess the effects of oral N. *sativa* on the gene expression of inflammatory and adipogenesis-related factors, including TNF-α, PPAR-γ, and adiponectin as well as assessing their serum concentrations among obese and overweight individuals.

Methods: Obese and overweight women aged 25–55 years with a body mass index (BMI) of 25–35 kg/m² will be recruited from the Obesity Clinic in Shahid Sadoughi University of Medical Sciences and will be assessed for eligibility against inclusion criteria. They will be randomly assigned into two groups to receive either two capsules of *N. sativa* or two capsules of placebo per day for eight weeks (each capsule contains 1000 mg of *N. sativa* or placebo). There will be a four-week wash-out period and then participants will receive the reverse supplements for another eight weeks. Biochemical assessments and gene expressions (using real-time polymerase chain reaction) will be conducted at the beginning and at the end of every intervention period.

Discussion: The present study will investigate the probable cellular pathways for the anti-obesity effects of *N. sativa* in overweight/obese women.

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Trial registration: Iranian Registry of Clinical Trials, IRCT20180528039884N1. Registered on 2nd of July, 2018.

Keywords: Obesity, Overweight, *Nigella sativa*, Tumor necrosis factor-alpha, Adiponectin, Peroxisome proliferator-activated receptors

Background

The epidemic of overweight and obesity presents a major challenge to health around the world. Obesity is defined as the accumulation of abnormal or excessive fat that may interfere with the maintenance of an optimal state of health [1]. The excess of macronutrients in the adipose tissues stimulates them to release inflammatory mediators such as tumor necrosis factor alpha (TNF- α) and reduces production of adiponectin, predisposing to a pro-inflammatory state and oxidative stress [2].

Among the adipokines, adiponectin is widely recognized for its anti-diabetic, anti-inflammatory, anti-atherogenic, and cardio-protective effects and shows protective activity in various processes such as energy metabolism and inflammation [3]. In humans, plasma levels of adiponectin are decreased in cases of insulin resistance and obesity [4]. One of the molecular mechanisms of adiponectin may be direct actions via downregulating of inflammatory responses involving TNF- α [5]. In obesity, alterations in the gene expression of adiponectin and its receptors reduce adiponectin sensitivity leading to insulin resistance [3].

On the other hand, peroxisome proliferator-activated receptors (PPARs) are a group of nuclear receptors with various isoforms, including α , β/δ , and γ , that are involved in transcription regulation of a broad range of genes related to inflammation and energy homeostasis and represent important targets for obesity, obesity-induced inflammation, and metabolic syndrome. PPAR-y is a transcription factor abundant in adipose tissue which has been known to regulate adipocyte differentiation and fatty acid storage [6]. More recently, PPAR-y has been recognized as playing an important role in inhibiting the expression of inflammatory cytokine [7]. Obesity has been reported to induce a decline in the activity and amount of PPAR-γ. This correlation appears to be strongly associated with the pathogenesis of obesity [8]. An association between adiponectin and PPAR-y has been reported by Yadav et al.; they revealed that adiponectin can mediate phosphorylation of adenosine monophosphate-activated protein kinase and also PPARs, after binding to its receptors and thereby increase fatty acid oxidation [9]. An animal study on rats reported that increased adiponectin levels

improvements in insulin sensitivity after a weight-reduction intervention, might be due to an increase in PPAR- γ levels [10].

With regard to the relation between TNF- α and obesity, plasma concentration of TNF-α and its receptors were found to be higher in obese individuals, which may get alleviated with weight loss [11]. Interestingly, TNF- α is reported to have a correlation with PPAR-y activity. Studies revealed that TNF-α might have inhibitory effects on PPAR-y which is involved in the pathogenesis of inflammation [12, 13]. Furthermore, a previous study reported that PPAR-γ activity could be regulated by TNF-α at pretranslational and post-translational levels [13]. More recently, the long-term assessment of anti-TNF-α inhibitor treatment to individuals diagnosed with metabolic syndrome has been shown to increase adiponectin levels, confirming a role for TNF-α in obesity-related insulin resistance in humans [12]. Moreover, TNF- α inhibits the conversion of pre-adipocytes to mature adipocytes, notably through downregulating adipogenic genes such as PPAR-y, allowing further recruitment of uncommitted cells and, thus, possible expansion of adipose tissue mass [14]. TNF-\alpha also downregulates the messenger RNA (mRNA) levels of adiponectin [15] while, conversely, adiponectin suppresses lipopolysaccharide-induced TNF-α production [5].

Studies have shown that losing weight in obese individuals can result in an increase in adiponectin levels and the expression of its receptors [3, 15]. Meanwhile, herbal products are widely marketed for their weight-loss properties [16]. Nigella sativa (N. sativa) is a traditional herbal medicine that has been used for centuries to treat rheumatoid arthritis, diabetes, asthma, and other metabolic disorders. Recently, anti-obesity characteristics of N. sativa have also been indicated [17]. The exact mechanism of anti-obesity effects of N. sativa is unclear; bioactive components of N. sativa including Thymoquinone, inhibitory components of lipase, unsaturated fatty acids, and its appetite-reducing effect are involved in probable mechanisms for anti-obesity effects of N. sativa [18]. Additionally, N. sativa has suggested to act as an agonist of PPAR-y which impacts upon energy homeostasis, thus enhancing the expression of lipogenic genes [19]. Concerning the effects of N. sativa on gene expression, there are only in vitro, in vivo,

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and/or animal trials that evaluated the effects N. sativa or its major active component, Thymoquinone, on the gene expression of various factors such as TNF- α , adiponectin, or PPAR- γ , and reported beneficial improvements in inflammation [20–27]. Nevertheless, the probable mechanisms of N. sativa involving the gene expressions of factors related to obesity and its pathogenesis have not yet been investigated in human studies.

Few clinical trials assessed the effects of *N. sativa* on serum concentrations of inflammatory markers and adipokines [18, 28–30]. Mahdavi et al. investigated the effects of *N. sativa* oil supplementation combined with a low-calorie diet on TNF- α [30] using a parallel-randomized design and reported that *N. sativa* oil may help to better manage weight and inflammatory status in obese women [28].

To the best of our knowledge, there are no clinical trials that assessed the effects of N. sativa or Thymoquinone through the TNF- α , PPAR- γ , or adiponectin pathways in obesity.

Consequently, the present clinical trial aimed to investigate the following objectives:

- To assess the effects of oral *N. sativa* oil on the gene expression of TNF-α, PPAR-γ, and adiponectin along with the changes in serum concentrations of these factors among obese and overweight women.
- To conduct a crossover, placebo-controlled, randomized clinical trial (RCT), to compare the altered gene expression as well as other outcome measures of every individual with herself, as her own control, and with the control group to achieve more accurate results.
- 3. To investigate the alterations in serum levels of liver enzymes including alanine transaminase (ALT), aspartate transaminase (AST), and alkaline phosphatase (ALP) as well as measuring lipid profile including total cholesterol, triglyceride, high density of lipoprotein cholesterol (HDL-C), low density of lipoprotein cholesterol (LDL-C), and atherogenic indices before and after every intervention period.

Materials and methods

Study design

The design of the present study is a crossover, double-blind, placebo-controlled RCT which will be conducted among obese and overweight women for 20 weeks. Participants will receive either a daily dose of *N. sativa* or placebo for eight weeks in two phases separated by a wash-out period of four weeks. Each participant will be reversed for the second phase of the intervention according to the randomized crossover design. This study

will be conducted in the Obesity Clinic of Shahid Sadoughi University of Medical Sciences, Yazd, Iran. The overview of the study is presented in Fig. 1. Moreover, this article will be reported according to the Standard Protocol Items: Recommendations for Interventional Trials (SPIRIT) statement [31]. A SPIRIT diagram detailing the timing of enrolment, interventions and assessments is provided in Fig. 2. A completed SPIRIT checklist is also provided in Additional file 1. Any methodological changes in the study design or sample size, which may potentially affect the participants' safety or study procedures, will be discussed in the committee of ethics before the study initiation.

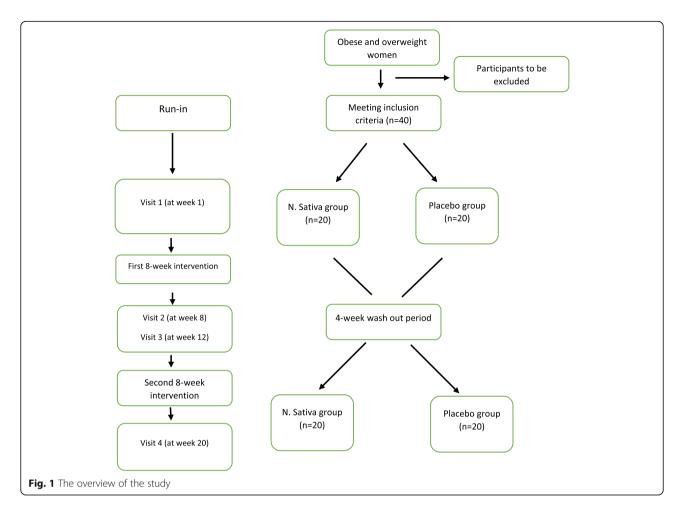
Eligibility of criteria

Obese and overweight women aged 25–55 years with a body mass index (BMI) of 25–35 kg/m² will be recruited from the Obesity Clinic in Shahid Sadoughi University of Medical Sciences. Exclusion criteria will include: cardiovascular diseases; hepatic, pancreatic, thyroid, or renal problems; having allergies to *N. sativa*; or a history of having a weight-reduction diet or program in the six months before the study. Furthermore, taking any antiobesity medications or any herbal and anticoagulant agents including aspirin, vitamin E, vitamin K, and warfarin in the six months before the study as well as being either pregnant, lactating, or having uncontrolled polycystic ovary syndrome will also be considered exclusion criteria.

Randomization and blinding

The present crossover, double-blind, placebo-controlled RCT will be consisted of two intervention periods of eight weeks each. Participants will be randomized 1:1 using stratified block randomization based on age (25-40 years and 40-55 years). One of the main researchers will use computer-generated random numbers to randomly allocate eligible participants into the intervention or control groups using sealed envelopes. Participants and administrators will be blinded to the content of the bottles, study treatment, and allocation until the final analysis. Individuals will be asked about their treatment group at the end of each intervention period. Their prediction will then be compared with the true assignments. Participants will receive four bottles for the entire 20week intervention. Each bottle will contain 56 capsules and every participant will receive two bottles during every intervention period (one bottle at the beginning of week 1 and one bottle at the end of week 4 of every intervention duration).

Participants will be also asked about any possible reactions or different feeling after receiving the treatments at the end of every intervention period. No forms will be Razmpoosh et al. Trials (2019) 20:512 Page 4 of 8



recorded, since the treatment doses were regarded as safe in the previous investigations.

Compliance

Compliance with consumption will be monitored every other day through phone interviews which will be followed by face-to-face interviews once a week. Participants will be asked to consume one capsule before lunch and one capsule before dinner; this will also increase their compliance with consumption. All participants will be requested to bring back their first bottle at the end of the primer intervention and then they will be given the second bottle of supplements. If the remaining capsules of every participant will exceed 10% of the total administered supplements (12 capsules), the participant will be categorized as non-adherent. All individuals, including those who will complete the study or those who will not complete for any reasons, will follow the same schedule.

Sample size

On the basis of sample size formula suggested for a balanced, crossover designed trials (Fig. 3),

considering the non-inferiority margin of $\delta\!=\!0.05$ (clinically meaningful difference), with an expected true difference of 0.02 between the means, and a population variance of 0.01 and considering the significance level of $\alpha\!=\!0.05$, the sample size to achieve 80% power is calculated to be 35. The final sample size was increased to 40 to accommodate the expected 10% of dropout rate.

Intervention

Before the study initiation, two trained researchers will introduce the study to participants. Participants, who will decide whether to join the investigation, will be asked to sign informed consent forms. The questionnaires will be reviewed and approved by the ethical committee of Shahid Sadoughi University of Medical Sciences. Data will be gathered by one of the main investigators. Individuals in the intervention group will receive two capsules of *N. sativa* (produced by Barij Essence Pharmaceutical Co.) and individuals in the placebo group will receive two capsules of paraffin oil per day. Both *N. sativa* and placebo capsules and the packs will be identical in terms of appearance, weight, texture,

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	STUDY PERIOD						
	Enrolment	Allocation	Post-allocation			Close-out	
TIMEPOINT	Month 1 prior to study	0	Week 1	Week 8	Week 12	Week 20	End of week 20
ENROLMENT:							
Eligibility screen	✓						
Informed consent	✓						
Allocation		√					
INTERVENTIONS:							
[first intervention duration (placebo or treatment)]			+	-			
[wash-out period]				+			
[second intervention duration (placebo or treatment)]					+		
ASSESSMENTS:							
[Baseline variables assessment]		√					
[primary and secondary outcome assessments]			>	>	√		✓

SPIRIT figure: Template of the content for the schedule of enrolment, interventions, and assessments.

Fig. 2 SPIRIT figure: Template of the content for the schedule of enrolment, interventions, and assessments

size, and smell and will only be differentiated by a code name ("A" or "B"). A person who is not involved in this project will label the containers "A" or "B." Every capsule of *N. sativa* includes 1000 mg of *N. sativa* oil, which will consist of 0.01 mg of Thymoquinone-active component. The gas chromatography–mass spectrometry (GC/MS)

 $n = \frac{\left(z_{\alpha} + z_{\beta}\right)^2 \sigma^2}{2\left(\mu - \mu_0 - \delta\right)^2}$

Fig. 3 Formula for calculating sample size in crossover studies. δ is the clinically meaningful difference, $\mu 2 - \mu 1$ is an expected true difference between the means, σ^2 is the population variance, α is the significance level, and β is the power of the test

test of every 50 g of N. sativa powder is presented in Table 1.

Matching energy intakes between groups

With regard to the dietary energy intakes of participants, we decided to give every individual an iso-energy dietary program in order to match every participant based on their energy and macronutrient intakes. Hence, an eightweek one-to-one individualized iso-energy dietary program for every intervention period, based on every participant's current weight, will be performed. After determination of required energy, the registered dietician will develop a plan of servings from each food group that will fulfill the macronutrient distribution. The required participants' daily energy intake will be distributed as follows: <30% from fat (mainly monounsaturated and poly-unsaturated fatty acids); 55% from carbohydrates; and 1.2–1.5 g protein per kg of body weight. After baseline measurements, each participant

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Table 1 Fatty acid content in every 50 g of *N. sativa* (GC/MS^a test)

Test	Reference method	Acceptable value	Result
Appearance	Visual	Brown color	Brown color
Odor	Organoleptic	Specific	Specific
Palmitic acid (%)	WR10 ^b	11–14	11.84
Stearic acid (%)	WR10	_	2.00
Oleic acid (%)	WR10	17–26	22.14
Linoleic acid (%)	WR10	52067	59.66
Linolenic acid (%)	WR10	_	1.79
Thymoquinone (%)	EP ^c 0.8 (2.2.23)	_	0.54

^aGas chromatography-mass spectrometry

will meet the registered dietitian three times during each intervention period (at weeks 0, 4, and 8).

Dietary intake will be quantified using three-day food records, including one weekend day and two weekdays as well as 24-h food recalls. Daily macro- and micro-nutrient intakes will be calculated by analyzing food data using nutritionist IV software (First Databank, San Bruno, CA, USA) modified for Iranian foods.

Shahid Sadoughi University of Medical Sciences will be responsible for any related problems; adverse events will be reported to the medical ethics committee from the initiation of the study. Participants may withdraw from the study for any reason at any time.

General information including age, diseases, medications, and supplements will be recorded through interviews at the beginning of the study. Physical activity level will be calculated through a questionnaire using metabolic equivalents (METs) at the beginning and at the end of every intervention period. Results of the assessments will be shared for every participant separately.

Primary outcomes consist of gene expression and serum concentration levels of adiponectin, PPAR- γ , and TNF- α . Secondary outcome measures consist of blood levels of lipid profiles and anthropometric measurements. The assessments of the outcomes are listed below.

Gene expression assay

A total of 2 mL of fresh blood will be transferred into a tube consisting of ethylenediaminetetraacetic acid (EDTA); immediately after, peripheral blood mononuclear cells (PBMCs) will be isolated using density centrifugation with Ficoll-Paque [32]. Total mRNA will then be extracted using a related RNA kit [33]. Total extracted RNA will be then reverse transcribed to complementary DNA (cDNA) using a cDNA synthesis kit [34]. The mRNA gene expression of adiponectin, PPAR- γ , and TNF- α will be applied using the real-time

polymerase chain reaction (PCR) method, after designing primers. The housekeeping gene will be glyceraldehyde phosphate dehydrogenase (GAPDH) in real-time PCR assessments.

Biochemical measurements

A total of 12 mL fasting blood sample of venous blood will be collected from every participant. Circulating adiponectin and TNF-α will be assessed by enzyme-linked immunosorbent assay (ELISA). Blood pressure will be measured with a well-validated automated digital blood pressure monitor (Beurer, Germany, BM 85), following American Heart Association guidelines. Biochemical analyses including total cholesterol, triglycerides, HDL-C, LDL-C as well as liver enzymes, including ALT, AST, and ALP, will be measured using automated enzymatic methods. The values of atherogenic factors, including the ratios of HDL-C/LDL-C and LDL-C/HDL-C, will be calculated. All laboratory data will be identified by an ID number to maintain the participants' confidentiality.

Anthropometric measurements

The following measures will be quantified at the onset and at the end of every intervention period. Actual body weight, visceral fat, body fat, and body muscle percentage will be measured using a commercial body composition monitor and scale (InBody, USA, NO. 770). Body weights will be quantified to the nearest 0.1 kg. Measurements will be taken when participants are without shoes and wearing only light clothing. Their height will be measured to the nearest 0.1 cm with the participants barefoot using a stadiometer (Seca 222). BMI will be calculated as weight in kilograms divided by height in meters squared. Waist circumference measurement will be taken with minimal inspiration at the smallest waist circumference area, rounding up to the nearest 0.1 cm. Waist-to-hip ratio (WHR) will be calculated via standard equations.

Statistical analysis

All statistical analyses will be performed with the Statistical Package for the Social Sciences (SPSS) (IBM SPSS Statistics for Windows, version 23.0; IBM Corp.), with significance set at P < 0.05. Normality of data will be assessed using the Shapiro–Wilk test. The values of continuous variables will be presented as means \pm SEM. Categorical data will be presented as number and percentage in study groups. The general linear model repeated measure procedure will be incorporated to compare the quantitative outcome variables between the intervention periods in crude and multivariable adjusted models taking the rolling method and participants' age, baseline values as between-person factors, and the change in physical activity level, energy intake, and body

^bRectangular waveguide size

^cEuropean Pharmacopoeia

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weight as covariates. The criterion for statistical significance will be defined as P < 0.05. Pearson's correlation coefficient will be applied to show the correlation between biochemical measures and anthropometric indices.

We will analyze the results with intention to treat (ITT) and without participants with non-compliance (study completers). ITT analysis is based on the initial treatment assignment and not on the treatment eventually received. In the ITT method, participant-loss associated bias as well as misallocation or non-adherence of participants is near completely limited. In the case that researchers observe a significant difference between those allocated to receive an intervention and those who actually received that intervention (and adhere to it), per-protocol analysis, which is an additional analysis adjusted for actual treatment, will be performed. The results of the two mentioned analyses will be compared with each other.

Discussion

The epidemic of overweight and obesity presents a major challenge to health around the world.

Adipose tissue predisposes pro-inflammatory status; hence, performing strategies to decrease body weight and the inflammation status would be of interest. *N. sativa* has recently been found to have anti-obesity effects, probably due to anti-inflammatory properties. However, the cellular pathways and mechanisms are not clear. Therefore, the present study will assess the effects of *N. sativa* on the gene expression of some factors involved in adipogenesis and obesity-related inflammation. If the results of the present study are valid, this will broaden the present assumptions on the specific functions of *N. sativa* and the context for performing more trials.

Trial status

The initial participant recruitment has been completed.

Additional file

Additional file 1: SPIRIT 2013 Checklist: Recommended items to address in a clinical trial protocol and related documents*. (DOC 132 kb)

Abbreviations

ALP: alkaline-phosphatase; ALT: alanine transaminase; AST: aspartate transaminase; BMI: body mass index; EDTA: ethylenediaminetetraacetic acid; ELISA: enzyme-linked immunosorbent assay; GAPDH: glyceraldehyde phosphate dehydrogenase; HDL-C: high density of lipoprotein cholesterol; ITT: intention to treat; LDL-C: low-density of lipoprotein cholesterol; METs: metabolic equivalents; *N. sativa: Nigella sativa*; PBMC: peripheral blood mononuclear cell; PCR: Polymerase Chain Reaction; PPAR-y: peroxisome proliferator activated receptor alpha; TNF-a: tumor necrosis factor alpha; WHR: waist to hip ratio

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Authors' contributions

All authors contributed to the idea and design of the study. ER and SF performed the idea of this study, study design, received the grant, and were major contributors in the writing and reviewing of the manuscript. AN and MM reviewed and contributed to manuscript. NA participated in the design and contributed to the manuscript and submission. AA and HF provided statistical expertise in clinical trial design and reviewed and contributed to manuscript. AN and MM edited the final version of the manuscript. All authors approved the final version submitted for publication.

Authors' information

Trial sponsors

1. Shahid Sadoughi University of Medical Sciences, Research Associate, School of Public Health (No. 961020).

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Role of sponsor: The University will control implementation of the study in accordance with the approved protocol based on ethical rules.

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Role of sponsor: This funding source had no role either in the design of this study or in the implementation, analyses, and interpretation of the data.

Roles and responsibilities—committees

"Principal investigator"

- Design and conduct of the study
- Preparation of protocol and revisions
- Recruitment of participants
- Publication of study reports

"Research committee"

- Agreement of final protocol
- Reviewing progress of study and if necessary agreeing changes to the protocol
- Budget administration

"Ethics committee"

- Reviewing progress of study and if necessary agreeing changes to the protocol
- Reporting serious unexpected suspected adverse events

Funding

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Availability of data and materials

The datasets generated during the current study will be available via the corresponding author on reasonable request.

Ethics approval and consent to participate

Written consent form will be obtained from all participants before the study initiation. This protocol is approved by the Ethics Committee of the Shahid Sadoughi University of Medical Sciences (IR.SSU.SPH.REC.1397.007). This study is registered at the Iranian Registry of Clinical Trials (IRCT20180528039884N1).

Consent for publication

Not applicable.

Competing interests

The authors declare that they have no competing interests.

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