RESEARCH

Open Access



Compound heterozygous inheritance of two novel COQ2 variants results in familial coenzyme **Q** deficiency

Aliaa H. Abdelhakim^{1†}, Avinash V. Dharmadhikari^{2†}, Sara D. Ragi¹, Jose Ronaldo Lima de Carvalho Jr.^{1,3}, Christine L. Xu¹, Amanda L. Thomas², Christie M. Buchovecky², Mahesh M. Mansukhani², Ali B. Naini², Jun Liao², Vaidehi Jobanputra², Irene H. Maumenee¹ and Stephen H. Tsang^{1*}

Abstract

Background: Primary coenzyme Q10 deficiency is a rare disease that results in diverse and variable clinical manifestations. Nephropathy, myopathy and neurologic involvement are commonly associated, however retinopathy has also been observed with certain pathogenic variants of genes in the coenzyme Q biosynthesis pathway. In this report, we describe a novel presentation of the disease that includes nephropathy and retinopathy without neurological involvement, and which is the result of a compound heterozygous state arising from the inheritance of two recessive potentially pathogenic variants, previously not described.

Materials and methods: Retrospective report, with complete ophthalmic examination, multimodal imaging, electroretinography, and whole exome sequencing performed on a family with three affected siblings.

Results: We show that affected individuals in the described family inherited two heterozygous variants of the COQ2 gene, resulting in a frameshift variant in one allele, and a predicted deleterious missense variant in the second allele (c.288dupC,p.(Ala97Arqfs*56) and c.376C > G,p.(Arq126Gly) respectively). Electroretinography results were consistent with rod-cone dystrophy in the affected individuals. All affected individuals in the family exhibited the characteristic retinopathy as well as end-stage nephropathy, without evidence of any neurological involvement.

Conclusions: We identified two novel compound heterozygous variants of the COQ2 gene that result in primary coenzyme Q deficiency. Targeted sequencing of coenzyme Q biosynthetic pathway genes may be useful in diagnosing oculorenal clinical presentations syndromes not explained by more well known syndromes (e.g., Senior-Loken and Bardet-Biedl syndromes).

Keywords: Coenzyme Q10, COQ2 gene, Oculorenal syndrome, Hereditary retinopathy

*Correspondence: sht2@cumc.columbia.edu

[†]Aliaa H. Abdelhakim and Avinash V. Dharmadhikari have contributed equally to this work

¹ Edward S. Harkness Eye Institute, Columbia University Irving Medical Center, New York, NY, USA

Full list of author information is available at the end of the article



Introduction

Coenzyme Q10 (CoQ_{10}), a key component of the oxidative phosphorylation pathway, is an essential factor in mitochondrial metabolism, and plays important roles in other cellular processes such as nucleotide synthesis, sulfide metabolism and cellular apoptosis. Endogenous defects in the biosynthesis pathway of coenzyme Q result in mitochondrial respiratory chain deficiency and associated manifestations. Involvement of the disease is systemic, and often includes neurologic manifestations

© The Author(s) 2020. Open Access This article is licensed under a Creative Commons Attribution 4.0 International License, which permits use, sharing, adaptation, distribution and reproduction in any medium or format, as long as you give appropriate credit to the original author(s) and the source, provide a link to the Creative Commons licence, and indicate if changes were made. The images or other third party material in this article are included in the article's Creative Commons licence, unless indicated otherwise in a credit line to the material. If material is not included in the article's Creative Commons licence and your intended use is not permitted by statutory regulation or exceeds the permitted use, you will need to obtain permission directly from the copyright holder. To view a copy of this licence, visit http://creativecommons.org/licenses/by/4.0/. The Creative Commons Public Domain Dedication waiver (http://creativeco mmons.org/publicdomain/zero/1.0/) applies to the data made available in this article, unless otherwise stated in a credit line to the data. (encephalopathy, hypotonia, cerebellar ataxia, progressive multiple-system atrophy like disease, spasticity, seizures, and intellectual dysfunction), kidney manifestations (steroid-resistant nephrotic syndrome and endstage renal disease), hypertrophic cardiomyopathy, myopathy and sensorineural hearing loss [1, 2]. Ocular manifestations such as optic atrophy, nystagmus and retinopathy have also been described [1, 2]. Pathogenic variants in any of the genes involved in the biosynthetic pathway of coenzyme Q10 can result in a primary CoQ10 deficiency phenotype; in particular, the genes *COQ2* and *PDSS2* have been associated with retinopathy and optic atrophy, respectively. Depending on the severity of the variant and the deficiency, manifestations may be severe and appear in infancy or mild and begin in adulthood.

In this study, we identify two novel recessive variants in the *COQ2* gene which, when acting together in the compound heterozygous state, result in primary CoQ10 deficiency with associated nephropathy and retinopathy. In contrast to many other *COQ2* pathogenic variant phenotypes described, the phenotype here does not exhibit neurologic manifestations, but rather severe nephropathy combined with retinopathy in multiple affected members of the family.

Materials and methods Subjects

A family with three affected siblings who presented to the Department of Ophthalmology at Edward S. Harkness Eye Institute, Columbia University was recruited and a complete ophthalmic examination along with multi-modal imaging, electroretinography, and exome sequencing (ES) were conducted. Patient consent was obtained prior to genetic testing. All information in this report has been de-identified in accordance with HIPAA and institutional review board regulations.

Ophthalmic Imaging

After measuring best corrected visual acuity, all patients' eyes were dilated using topic 1% tropicamide and 2.5% phenylephrine followed by complete ophthalmic assessment including funduscopic examination, digital fundus photography, fundus short-wavelength autofluorescence (SW-AF) and spectral domain optical coherence tomography (SD-OCT). SW-AF (488 nm wavelength stimulus, barrier filtered transmitted light from 500 to 680 nm, $55^{\circ} \times 55^{\circ}$ field) and SD-OCT images were taken by the use of a confocal scanning laser ophthalmoscope (cSLO; Spectralis HRA+OCT, Heidelberg Engineering, Heidelberg, Germany). SD-OCT images were acquired as horizontal 9 × 9 mm scans (870 nm light source and 7 μ m axial resolution) positioned through the macula and obtained in high-resolution mode. The scans were

recorded automatically by the use of recorded IR-R (820 nm light source) fundus images.

Electroretinography

Full-field electroretinogram (ffERG) (Diagnosys, LLC, Lowell, Massachusetts, USA) was performed with Dawson Trick Litzkow (DTL) fiber electrodes and Ganzfield stimulation. ERG recordings were obtained for our patient for both eyes in accordance with the International Society for Clinical Electrophysiology of Vision (ISCEV) guidelines in both the scotopic and photopic states [3]. The amplitudes and implicit times recorded for each eye of the patient were compared to control patients with normal values and healthy eyes.

Clinical exome sequencing (ES) and variant analysis

ES was performed on the three affected siblings at the Laboratory of Personalized Genomic Medicine (PGM) at Columbia University Irving Medical Center (CUIMC) on DNA obtained from peripheral blood. Written consent was obtained, exome sequencing libraries were prepared from genomic DNA from the probands using Agilent SureSelectXT (Human All Exon v.5+UTRs) capture kit according to the manufacturers' protocol. Pairedend sequencing was performed on the Illumina HiSeq 2500 platform. The sequence data were aligned and annotated using NextGENe (version 2.3.4.5; Softgenetics LLC, PA) software. Variant filtering and annotation were performed using an in-house developed proprietary analytical pipeline. Variants of interest were further analyzed using VarSome [4] and classified using the current ACMG/AMP guidelines [5].

Results

Clinical presentation

A 25-year old man of Ashkenazi Jewish (AJ) descent presented to the clinic with new-onset difficulty with night vision. His medical history was notable for biopsyproven diffuse glomerulosclerosis and associated hypertension. Immunofluorescence studies of his renal biopsy demonstrated no evidence supporting immune complex type disease, and the majority of glomeruli examined were globally sclerotic. Best-corrected visual acuity was 20/30 in the right eye (OD) and 20/25 in the left eye (OS), with no afferent pupillary defects. Visual fields to confrontation were constricted; anterior segment exam was unremarkable. His dilated fundus exam revealed healthyappearing nerves and a flat normal-appearing macula, and clear vitreous in both eyes. Notably, he had extensive intraretinal pigment migration in the periphery arranged in a ring-like fashion, encircling the posterior poles in both eyes (Fig. 1a). Autofluorescence revealed hypoautofluorescence in a ring pattern mirroring the bone spicule



Fig. 1 Multimodal Imaging of COQ2 Associated Rod-Cone Dystrophy. **a** Wide-field color fundus photographs demonstrating a pale optic nerve, attenuation of the vessels, and bony spicules in the periphery and midperiphery of the retina in the youngest proband. **b** Wide-field fundus autofluorescence images show hypoautofluorescence in the periphery with a ring of hyperautofluorescence in the macular region in the youngest proband. **c** Spectral-domain optical coherence tomography (SD-OCT) unveils retinal thinning due to atrophy of the outer retinal layers with regions of blurred or absent ellipsoid zone. Corresponding wide-field color fundus photographs, wide-field fundus autofluorescence, and SD-OCT images showing similar findings in the affected brother (**d**–**f**), and affected sister (**g**–**I**)

pattern seen on clinical examination and color photography (Fig. 1b). Spectral domain optical coherence tomography (SD-OCT) revealed outer retinal atrophy in an annular pattern around the macula, with preserved outer retinal structure at the fovea (Fig. 1c).

The patient's family history was notable for two of four other siblings with similar ophthalmic and medical histories. The parents showed no manifestation of disease and denied familial consanguinity. His first affected sibling was an older brother of 32 years of age, who was diagnosed with "retinitis pigmentosa" at age 21, and who had received a kidney transplant at age 5 years for a diagnosis of mesangial sclerosis. His medical history was also notable for elevated liver enzymes of unknown etiology and childhood acute lymphoblastic lymphoma that went into remission. On ophthalmologic exam, this sibling demonstrated a best corrected visual acuity of 20/25 OD, 20/30 OS, absence of relative afferent pupillary defects, and constricted visual fields to confrontation. Corneas were clear and the anterior segments were quiet, but he had a P1 posterior subcapsular cataract [6] in both eyes. Dilated fundus examination revealed nerves that were pale and cupped, flat normal-appearing maculae, and similar to the youngest sibling described above, peripheral ringlike patterns of bone speculation and pigment migration encircling the posterior poles in both eyes (Fig. 1d). Autofluorescence imaging showed hypoautofluorescent rings corresponding to the bone spiculated areas on color photography (Fig. 1e), and SD-OCT showed outer retinal atrophy in an annular pattern surrounding the macula, similar to his younger brother (Fig. 1f).

The patient's second affected sibling was a 28-year old sister, also with a history of "retinitis pigmentosa" diagnosed at age 23, and a kidney transplant at age 10 for end-stage renal disease of unknown etiology; her birth history was notable for an extra digit on the right foot. Similar to her two affected siblings, her best-corrected central visual acuity was relatively good at 20/30 in both eyes (OU), without relative afferent pupillary defects. Her visual fields to confrontation were constricted, her corneas were clear and her anterior segments were quiet. Her lenses contained a P1 posterior subcapsular cataract [6] in both eyes. Her dilated fundus examination revealed healthy-appearing nerves, flat unremarkable maculae and clear vitreous. Again, her periphery was notable for extensive bone spiculation in a ring around the posterior pole OU, with mirroring autofluorescence patterns and outer retinal atrophy on SD-OCT (Fig. 1g–i).

Electroretinography (ERG) was performed on all three siblings and revealed a pattern consistent with rodcone dystrophy, with an undetectable scotopic response (Fig. 2). All affected siblings had similar results with this modality of testing. An attempt was made to treat the underlying CoQ10 deficiency with exogenous coenzyme



responses being more severely affected, indicating a rod-cone dysfunction

Q10 supplementation for all affected siblings for a period of 6 months to determine if any improvement could be detected on ERG readings (Tishcon Corp. Liposomal QH 10% ubiquinol solution, 10–30 mg/kg/day with the guidance of nephrology colleagues). Supplementation did not demonstrate any ERG improvement; however, both best corrected visual acuity and areas of retinal atrophy on autofluorescence were noted to be stable on treatment.

Genetic analysis

ES was performed to determine the genetic cause of the three affected siblings' phenotype. Although there was clinical suspicion for entities such as Senior-Loken syndrome 1-9 (MIM# 606996, 613615, 266900, 616629, 609254, 606995, 616307, and 610189) and Bardet-Biedl syndrome 1-21 (MIM# 209900, 615981, 600151, 615982, 615983, 605231, 615984, 615985, 615986, 615987, 615988, 615989, 615990, 615991, 615992, 615993,

615995, 615996, 617119, and 617406), no variants were detected in anyone of the corresponding genes; ES (see "Materials and methods" for detailed description) detected two heterozygous variants in the clinically relevant gene *COQ2* (NM_015697.8) that segregated in all affected individuals (Fig. 3).

The first variant was a duplication c.288dupC, in exon 1 of 7, which results in a frameshift and is predicted to cause premature termination of the protein, p.(Ala97Argfs*56).This variant was detected at an allele frequency of 6.987e-3 in gnomAD v3 exomes and genomes, seen in the heterozygous state in 10/143116 alleles, with no homozygotes. This suggests that it is not a common benign variant in the populations represented in these databases. A literature search revealed no previous patient reports described with this variant. There are conflicting predictions of pathogenicity for this variant in ClinVar (VarID: 631951), partly due to lack of reports



in patients and higher allele frequency in the gnomAD Ashkenazi Jewish population (heterozygous in 8/3322 alleles (allele frequency=0.2408, gnomAD v3). Though pathogenic frameshift variants reported in this gene are rare, there is one frameshift variant p.(Leu234Argfs*14) that terminates the protein downstream of this variant reported in the literature in a patient with isolated nephrotic syndrome (Fig. 4a). Based on current ACMG/AMP sequence variant interpretation guidelines, we classified this variant in our patients as pathogenic.

The second *COQ2* variant found in these patients, c.376C > G, p.(Arg126Gly) substitutes a glycine for an arginine at a conserved amino acid residue 126/422 in the hydroxybenzoate polyprenyl transferase domain of COQ2 (Fig. 4b) (Accession: TIGR01474, TIGRFAMS domain). In silico methods predicted this variant to be deleterious or damaging to protein function (Provean score: -4.34 and SIFT score: 0.004). The variant is absent in gnomAD v3 exomes and genomes, which indicates that it is not a common benign variant in the populations

represented in these databases. This variant is absent from ClinVar and no previously described cases were found in the literature. We classified this variant based on ACMG/AMP guidelines as a likely pathogenic variant.

Although neither parent in this family was tested for these two variants, clinical ES results revealed that the variants were present in different sequencing reads in the proband (Fig. 4c), indicating they came from different alleles and are in compound heterozygous state. This was possible as the variants are very close (88 bases apart) and phase could be determined based on the two variants not co-existing in the same sequence reads (100 bp in length). Similar results were also found in the older siblings, providing further evidence for bi-allelic inheritance of the two *COQ2* variants.

Discussion

Here we describe two novel recessive potentially pathogenic variants: *COQ2*(NM_015697.8):c.288dupC,p. (Ala97Argfs*56) and *COQ2*(NM_015697.8):c.376C>G,p.

(See figure on next page.)

Fig. 4 Genetic Results: Pedigree analysis, localization of *COQ2* variants, and next generation sequencing (NGS) pile-ups. **a** Localization of novel and reported *COQ2* variants in protein (NP_056512.5) and RNA (NM_015697.8) transcripts using the Illustrator for BioSequence (IBS) tool [30] Red boxes represent transmembrane domains per Forsgren et al. [31]; blue box represents an allylic polyprenyl diphosphate substrate-binding site (APDSBS); variants in light blue boxes are frameshift or non-sense variants; missense variants in yellow boxes are reported in patients with severe multisystem disease presentation; missense variants in green boxes are reported in patients with nephrotic syndrome with /without neurological symptoms; missense variants in purple boxes are reported in patients with multiple system atrophy; retinitis pigmentosa, and in the current study. **b** Conservation of p.Arg126Gly variant across multiple species and localization in the 4-hydroxybenzoate polyprenyl transferase domain. **c** NexGENe pile-ups demonstrating that the two *COQ2* variants are *in trans* in the youngest proband indicating compound heterozygous inheritance



(Arg126Gly), which together in the compound heterozygous state led to primary CoQ10 deficiency in three siblings. Due to limitations imposed by the COVID-19 pandemic, we were unable to confirm our genetic findings through measurement of CoQ10 levels from muscle biopsy or fibroblast samples. However, the pathogenicity of these variants is supported by several observations, including manifestation of symptoms in all three affected family members, population database analysis and in silico predictions. The combination of these variants in the compound heterozygous state primarily presented as severe progressive glomerulosclerosis leading to eventual kidney transplant at a young age, as well as progressive retinal degeneration mimicking retinitis pigmentosa.

Pathogenic variants of the *COQ2* gene and clinical manifestations, including neurologic and renal involvement, have previously been reported in the literature (Table 1).COQ2 deficiency has a variable presentation and can be associated with a severe multi organ system disorder [7–13], an intermediate nephrotic syndrome with or without neurological symptoms [9, 14–23], and milder later presentations of multi system atrophy and retinitis pigmentosa [24, 25]. The residual COQ2 function associated with a variant is shown to correlate with the severity of the phenotype [26]. Like previously reported cases, our patients exhibited renal degeneration, however they were unique in that that none of them developed neurologic abnormalities (but developed retinitis pigmentosa).

The COQ2 gene was the first to be identified as a cause of primary CoQ10 deficiency when altered [19]. Although there are several reports of COQ2 variants causing primary CoQ10 deficiency, only three of the reported patients with confirmed variants in the COQ2 gene with associated retinopathy have been reported in the literature. The first patient was one of two affected siblings in a family who presented with nephrotic syndrome, neurological symptoms and rod-cone retinopathy [20]. His affected sister only presented with isolated nephrotic syndrome. Both siblings were homozygous for the variant, *COQ2* (NM_015697.8):c.890A > G,p. (Tyr297Cys). The other two patients were siblings with a homozygous variant, COQ2(NM_015697.8): c.328A>G, p.(Met128Val) that manifested as multiple-system atrophy and retinitis pigmentosa without clinical signs of nephropathy [27]. One of these siblings, similar to the patients in our series, was diagnosed with retinitis pigmentosa at a young adult age (33 years old), and developed neurological symptoms in the seventh decade of life. On post-mortem examination, her retina revealed severe loss of rods and cones and patchy disappearance of pigmentary epithelial cells [24]. The other sibling developed night blindness at the age of 48 and was Page 7 of 12

subsequently diagnosed with retinitis pigmentosa. The siblings in our series differ from these aforementioned cases by presenting with severe nephropathy in addition to clinical retinitis pigmentosa, with absence of neurological involvement. The variants in our family are a combination of a damaging frameshift variant predicted to have no residual protein function and a missense variant. The intermediate phenotype seen in this family suggests that the p.(Arg126Gly) variant, similar to the neighboring p.(Met128Val), could be a mild variant with residual activity. Functional studies in yeast demonstrate that the p.(Met128Val) is a mild variant associated with residual activity [26], and therefore in the homozygous state leads to a late onset phenotype. Interestingly, both the p.(Met128Val) and p.(Arg126Gly) variants are found in patients with retinitis pigmentosa, suggesting that this region of the protein is likely essential for proper functioning of the protein in the eye. Although none of the siblings manifested any neurological abnormalities, we cannot exclude the possibility of neurological involvement later in life as with the previously reported cases.

Interestingly, COQ2 is the only gene in the CoQ10 biosynthesis pathway that has been definitively linked with a retinitis pigmentosa-like presentation in the reported literature. There are two additional reports of patients with severe CoQ10 deficiency and retinopathy without confirmed variants in any of the biosynthetic pathway genes; rather, CoQ10 deficiency in these cases was confirmed biochemically. Severe neurological impairment at a young age was observed in both of these cases. In one of the reported cases of a young patient with severe coenzyme Q10 deficiency, retinopathy was demonstrated by an abnormal electroretinogram and mild pigmentary degeneration of the retina [28]. In the second case, a patient with coenzyme Q10 deficiency was diagnosed with retinitis pigmentosa at 10 years of age with concurrent optic nerve atrophy of the left eye and cataract of the right eye [29]. Sequencing of the COQ1 gene in this latter case revealed no causative variant. Other genes in the biosynthetic pathway have been associated with other ocular abnormalities other than pigmentary retinopathy, such as such as optic atrophy (e.g., PDSS1/2). Why COQ2 is the only gene thus far to definitively cause pigmentary retinopathy remains to be determined.

Attempted treatment with high dose CoQ10 supplementation in this family failed to improve the ERG signal or vision, however vision remained stable, and there was no progression of retinopathy was observed on autofluorescence or in terms of retinal atrophy on optical coherence tomography. This may suggest that CoQ10 supplementation can aid in preservation of viable retina even if there is no improvement in areas that are already affected. Thus, CoQ10 supplementation

ned in the literature	
ants and phenotypes publish	
Table 1 COQ2 vari	

Domedi-Camassel et al [9] C(47)C-A)(47)C-A) Acute rear al failure, splieptic encephs (particity) (packing) WA Mollet et al [12] C(47)C-A)(47)C-A) Nature mark failure, splieptic encephs (particity) (packing) NA Mollet et al [12] Digodenty (packing) Nature mark failure, splieptic encephs (particity) NA Jakos del 17 C(97)C-A)(130C-A) Digodenty generalized defan; (particity) NA Jakos del 13 C(95)C-T1(90C-C-T1) Digodenty generalized defan; (particity) NA Scalais et al [13] C(95)C-T1(90C-C-T1) Digodenty generalized defan; (particity) NA Dinwiddle et al [8] C(57)C-A)(115C-T Nature mark incolaris (particity) NA Dinwiddle et al [8] C(37)C-A)(115C-T Neorabilisers, MA, hypoton; (particity) NA Dinwiddle et al [8] C(37)C-A)(115C-T Neorabilisers, MA, hypoton; (particity) NA Dinwiddle et al [7] C(37)C-A)(115C-T Neorabilisers, MA, hypoton; (particity) NA Distributed C(37)C-A)(115C-T Neorabilisers, MA, hypoton; (particity) NA Distributed C(37)C-A)(115C-T Neorabilisers, MA, hypoton; (particity) NA	References	Genotype:COQ2 (NM_015697.8, NP_056512.5)	Phenotype	Effect of CoQ10 supplementation	Phenotype category
Mollet et al. [1,2] C(I) y3/deT[I] (19/deT] Neurological distres at birty, liver lief*15) NA Jakobs et al. [11] C(I) y3/deT[I] (13/deT] 2/dsys (rel of ad y1.7 form muti- lief*15) Na Jakobs et al. [11] C(I) y3/deT[I] (13/deT] 2/dsys (rel of ad y1.7 form muti- lief*15) Na Scalais et al. [13] C(I) y3/deT[I] (13/deT) Disgosic (rwins; generalized efermi- matury) in one twin-infanite death Na Dinwiddie et al. [13] C(J) SZ/SS /AJ (13/GeT) Nyocionic seizures, HCM, hyotomia, windome (H mO); ded ad as Tommis Na Dinwiddie et al. [13] C(J) SZ/SS /AJ (13/GeT) Nyocionic seizures, HCM, randomegaly, windome (H mO); ded ad as Tommis Na Disposition (ST) C(J) SZ/SS /AJ (13/GET) Nyocionic seizures, HCM, randomegaly, windome (H mO); ded ad as Tommis Na Desbats et al. [7] C(J) SZ/SS /AJ (J) SZ /AJ (J) SZ /AJ (J) SZ /AJ (J) (J) (J) (J) (J) (J) (J) (J) (J) (J	Diomedi-Camassei et al. [9]	c.[437G > A];[437G > A] p.[(Ser146Asn)];[(Ser146Asn)]	Acute renal failure, epileptic encepha- lopathy, hypotonia; died at 6 months	N/A	Severe multisystem
Jakobs et al. [11] CJ905C > TLJ905C > TLJ905C > TL Dizgotic twins; Premature birth, Feed NA Scalais et al. [13] CJ95G > AJJ326G > AJJ327G >	Mollet et al. [12]	c.[1197delT];[1197delT] p.[(Asn4011lefs*15)];[(Asn401 llefs*15)]	Neurological distress at birth; liver failure and nephrotic syndrome at 2 days; died at day 12 from multi- organ failure	N/A	Severe multisystem
Scalais et al. [13] C(3265 - Å)(3266 - Å) Myoclonic seizures, HCM, hypotonia, No improvement and consistences and conservation in the constraints (10 week) negations in the constraints (10 week) negations is stretchild negations in the constraint and constraint in the constraint renal Lubular Gisess, receptal oparthy, additional C(376 - Å)(3765 - Å)(3765 - Å)(3765 - Å) No improvement and constraint and constraints (10 metabolic acidosis, proteinuria, hepatic in (10) No improvement and constraints (10 metabolic acidosis, proteinuria, hepatic in (10) No improvement of ten (11) Eroglu et al. [10] C(3755 - Å)(3755 - Å)(3755 - Å) Lactic acidosis, proteinuria, nepatic in (10) NA Eroglu et al. [10] C(3755 - Å)(3755 - Å) Index case in family, neonatal disbetes, in (10) NA Eroglu et al. [10] C(3755 - Å)(3755 - Å) Index case in family, neonatal disbetes, interrological condition, died at 23 in (10) NA Eroglu et al. [10] C(3755 - Å)(3755 - Å) Index case in family, neonatal disbetes, interrological dispetes, one (10 metabolic acidosis, neurological dispetes, neurological	Jakobs et al. [11]	c.[905C > T];[905C > T] p.[(Ala302Val)];[(Ala302Val)]	Dizygotic twins, Premature birth, feed- ing problems, generalized edema, seizures, apnea; retinopathy of pre- maturity in one twin; infantile death	N/A	Severe multisystem
Dinwiddie et al. [8] c.437G > A(3) I159C>T Neonatal diabetes, HCM, cardiomegaly. NA Retablic standing cardosis, percephalopatity, additional cardosis, percephalopatity, additional cardosis, percephalopatity, additional cardosis, percephalopatity, additional cardosis, percentria, percephalopatity, hepatic N/A Desbats et al. [7] c.[545T > G];[545T > G] Lactic acidosis, porteinuria, hepatic N/A Desbats et al. [7] c.[545T > G];[547 + G] Lactic acidosis, proteinuria, hepatic N/A Desbats et al. [7] c.[545T > G];[567 + 46Asn)];[(561 + 46Asn)	Scalais et al. [13]	c.[326G > A];[326G > A] p.[(Ser109Asn)];[(Ser109Asn)]	Myoclonic seizures, HCM, hypotonia, nystagmus (10 weeks),nephrotic syndrome (4 mo); died at 5 months	No improvement and continued deterioration	Severe multisystem
Desbats et al. [7] C[545 T > G];[545 T > G] Lactic acidosis, proteinuria, hepatic insufficiency, dilation of left ventricle, worsening neurological condition, died at 23 h NA Eroglu et al. [10] C[437G > A];[437G > A] Index case in family, neonatal diabetes, glomenular proteinuria, seizures, death at 45 months No improvement of ren glomenular proteinuria, seizures, death at 45 months No improvement of ren glomenular proteinuria, seizures, death at 45 months Eroglu et al. [10] C[437G > A];[437G > A] Affected sbling; neonatal diabetes, proteinuria, mild metabolic acidosis, seizures, kidney failure, died due to nulti-organ failure at 31 months Normalization of gluccos death at 45 months Eroglu et al. [10] C[437G > A];[437G > A] Affected sbling; neonatal diabetes, proteinuria, mild metabolic acidosis, seizures, died due to nulti-organ failure at 31 months Normalization of gluccos death at 45 months Eroglu et al. [10] C[437G > A];[437G > A] Index case in family; nomiting, protein nulti-organ failure at 31 months Normalization of gluccos seizures, died at 6 months Froglu et al. [10] C[437G > A];[437G > A] Index case in family; nomiting, protein nulti-organ failure at 14 months Proteinuria, mild with encephalopatity Froglu et al. [10] C[437G > A];[437G > A] Index case in family; nomiting, protein seizures, died at 6 months Proteinuria, seizures, died at 6 months Froglu et al. [10] C[437G > A];[437G > A] <t< td=""><td>Dinwiddie et al. [8]</td><td>c.437G > A(;)1159C > T p.(Ser146Asn)(;)(Arg387Ter) Additional c.3754C > A variant in the <i>MT-ND1</i> gene</td><td>Neonatal diabetes, HCM, cardiomegaly, metabolic acidosis, encephalopathy, chronic renal tubular disease, respira- tory failure, died at 2 months</td><td>N/A</td><td>Severe multisystem</td></t<>	Dinwiddie et al. [8]	c.437G > A(;)1159C > T p.(Ser146Asn)(;)(Arg387Ter) Additional c.3754C > A variant in the <i>MT-ND1</i> gene	Neonatal diabetes, HCM, cardiomegaly, metabolic acidosis, encephalopathy, chronic renal tubular disease, respira- tory failure, died at 2 months	N/A	Severe multisystem
Eroglu et al. [10]c.[437G > A];[437G > A]Index case in family; neonatal diabetes, glomerular proteinuria, seizures, encephalopathy, hypotonia, electro- retinography showed no response, death at 4.5 monthsNo improvement of real sizures, cal symptomsEroglu et al. [10]c.[437G > A];[437G > A]Index case in family; neonatal diabetes, death at 4.5 monthsNo improvement of real sizures, deatinor proteinuria, mild metabolic seizures, kidney failure died due to multi-organ failure at 31 monthsNormalization of glucos levels; neurological du seizures; deterioration functionEroglu et al. [10]c.[437G > A];[437G > A]Index case in family; vomiting, protein uria, nephrotic syndrome, refractory seizures, died at 6 monthsNormalization of glucos inctionEroglu et al. [10]c.[437G > A];[437G > A]Index case in family; vomiting, protein uria, nephrotic syndrome, refractory seizures, died at 14 monthsNAKu et al. [23]c.[518G > A];[973A > GAffected sibling; neonatal diabetes, p.[(Argl 173His)];[(Th17325Ala)]NAKu et al. [23]c.[518G > A];[973A > GEdema, mild motor delay, moderate for otherProteinuria, seizures, for otherKu et al. [23]c.[518G > A];[973A > GEdema, mild motor delay, moderate for otherFedema, mild motor delay, motorKu et al. [23]c.[518G > A];[973A > GFedema, mild motor delay, motorProteinuria, seizures, for otherKu et al. [23]c.[518G > A];[973A > GFedema, mild motor delay, moderate for otherProteinuria, seizures, for other	Desbats et al. [7]	c.[545 T > G];[545 T > G] p.[(Met182Arg)];[(Met182Arg)]	Lactic acidosis, proteinuria, hepatic insufficiency, dilation of left ventricle, worsening neurological condition, died at 23 h	N/A	Severe multisystem
Eroglu et al. [10]c.[437G > A];[437G > A]Affected sibling; neonatal diabetes, proteinuria, mild metabolic acidosis, seizures, kidney failure, died due to multi-organ failure at 31 monthsNormalization of glucos levels; neurological do seizures, deterioration functionEroglu et al. [10]c.[437G > A];[437G > A] [Ser146Asn]);[(Ser146Asn)];[(Ser146Asn)];[(Ser146Asn)]; p.[(Ser146Asn)];[(Ser146Asn)];[(Ser146Asn)]; seizures, died at 6 monthsNormalization of glucos levels; neurological do seizures, died at 6 monthsEroglu et al. [10]c.[437G > A];[437G > A] p.[(Ser146Asn)];[(Ser146Asn)]Index case in family; vomiting, protein- nuria, nephrotic syndrome, refractory seizures, died at 6 monthsN/AEroglu et al. [10]c.[437G > A];[437G > A] p.[(Ser146Asn)];[(Ser146Asn)]Affected sibling; neonatal diabetes, nuria, nephrotic sprotome, refractory seizures, died at 6 monthsProteinuria improved; cn neurological deteriorXu et al. [23]c.[518G > A];[973 A > G] p.[(Arg173His)];[(Thri325Ala)]Affected dibling; neonatal diabetes, noteinuria, seizures, diffuse cerebral natrophy; died at 14 monthsProteinuria improved; cn neurological deteriorXu et al. [23]c.[518G > A];[973 A > G] p.[(Arg173His)];[(Thri325Ala)]Edema, mild motor delay, moderate motor developmentEdema, moderateRu et al. [23]p.[(Arg173His)];[(Thri325Ala)]Affected delay nephrotic syndrome motor developmentProteinuria	Eroglu et al. [10]	c.[437G > A];[437G > A] p.[(Ser146Asn)];[[Ser146Asn]]	Index case in family; neonatal diabetes, glomerular proteinuria, seizures, encephalopathy, hypotonia, electro- retinography showed no response, death at 4.5 months	No improvement of renal or neurologi- cal symptoms	Severe multisystem
Eroglu et al. [10]c.[437G > A];[437G > A]Index case in family: vomiting, protein- uria, nephrotic syndrome, refractory seizures, died at 6 monthsN/AEroglu et al. [10]c.[437G > A];[437G > A]Affected sibling; neonatal diabetes, proteinuria, seizures, diffuse cerebral atrophy; died at 14 monthsProteinuria improved; o neurological deterior atrophy; died at 14 monthsXu et al. [23]c.[518G > A];[7hr325Alal]Edema, mild motor delay, moderate speech delay, moderateEdema resolved; improved; o neurological deterior atrophy: died at 14 months	Eroglu et al. [10]	c.[437G > A];[437G > A] p.[(Ser146Asn)];[[Ser146Asn]]	Affected sibling; neonatal diabetes, proteinuria, mild metabolic acidosis, seizures, kidney failure, died due to multi-organ failure at 31 months	Normalization of glucose, lactate levels; neurological deterioration with encephalopathy and refractory seizures; deterioration of kidney function	Severe multisystem
Eroglu et al. [10] c.[437G > A];[437G > A] Affected sibling; neonatal diabetes, p.[(Ser146Asn)];[(Ser146Asn)] Proteinuria, seizures, diffuse cerebral atrophy; died at 14 months Proteinuria improved; creation Xu et al. [23] c.[518G > A];[973A > G] Edema, mild motor delay, moderate speech delay, nephrotic syndrome Edema evolopment months	Eroglu et al. [10]	c.[437G > A];[437G > A] p.[(Ser146Asn)];[(Ser146Asn)]	Index case in family; vomiting, protein- uria, nephrotic syndrome, refractory seizures, died at 6 months	N/A	Severe multisystem
Xu et al. [23] c.[518G > A];[973A > G] Edema, mild motor delay, moderate Edema resolved; improv p.[(Arg173His)];[(Thr325Ala)] speech delay, nephrotic syndrome motor development	Eroglu et al. [10]	c.[437G > A];[437G > A] p.[(Ser146Asn)];[(Ser146Asn)]	Affected sibling; neonatal diabetes, proteinuria, seizures, diffuse cerebral atrophy; died at 14 months	Proteinuria improved; continued neurological deterioration	Severe multisystem
	Xu et al. [23]	c.[518G > A];[973A > G] p.[(Arg173His]];[(Thr325Ala)]	Edema, mild motor delay, moderate speech delay, nephrotic syndrome (11 months)	Edema resolved; improved kidney and motor development	NS + neurological symptoms

References	Genotype:COQ2 (NM_015697.8, NP_056512.5)	Phenotype	Effect of CoQ10 supplementation	Phenotype category
Quinzii et al. [19], Salviati et al. [20], Diomedi-Camassei et al. [9], Lopez-Martin [16], Montini et al. [18]	c.[890A > G];[890A > G] p.[(1]yr297Cys)];[(1]yr297Cys)]	Index case in family: epileptic encepha- lopathy, hypotonia, mild psychomo- tor delay, optic atrophy, nephropathy, rod-cone retinopathy (11 mo), kidney transplant (3 years)	Drastic improvement in neurological manifestations; no improvement in renal dysfunction	NS + neurological symptoms + RP
Quinzii et al. [19], Salviati et al. [20], Diomedi-Camassei et al. [9], Lopez-Martin [16], Montini et al. [18]	c.[890A > G];[890A > G] p.[(1yr297Cys)];[(1yr297Cys)]	Sister affected with only nephrotic syndrome (12 mo); no extra-renal involvement	Resolution of nephrotic syndrome	Isolated NS
Diomedi-Camassei et al. [9]	c.[590G > A];[683A > G] p.[(Arg197His)];[(Asn228Ser)]	Nephrotic syndrome (onset at 18 mo); no extrarenal involvement	Neurological examination continued to be normal after 8-mo of follow up	Isolated NS
McCarthy et al. [17]	c.[683A > G];[701deIT] p.[(Asn228Ser)];[(Leu234Argfs*14)]	Nephrosis, renal failure (2 years)	N/A	Isolated NS
Bezdicka et al. [14]	c.[683A > G];[683A > G] p.[(Asn228Ser)];[(Asn228Ser)]	Two affected siblings; nephrotic syn- drome (3 years)	Kidney function improved in sibling; no supplementation in index case	Isolated NS
Starr et al. [21]	c.[973A > G];[1159C > T] p.[(Thr325Ala)];[(Arg387Ter)]	Nephrotic syndrome (9 months)	Nephrotic syndrome resolved; contin- ued proteinuria	Isolated NS
Starr et al. [21]	c.[1 76dupT];[683A > G] p.[(Ala69Argfs*33)]; [(Asn228Ser)]	Nephrotic syndrome (2 years)	Nephrotic syndrome resolved; contin- ued proteinuria	Isolated NS
Wu et al. [22]	c.[832T>C];832T>C] p.[(Cys278Arg)];[(Cys278Arg)] Also homozygous c.1213 + 1G>A vari- ant in <i>ARSB</i> gene	Nephrotic syndrome, died (6 mo); no extrarenal involvement	N/A	Isolated NS
Starr et al. [21]	c.[683A > G];[881C > T] p.[(Asn228Ser)];[(Thr29411e)]	Nephrotic syndrome (10 years)	Nephrotic syndrome not resolved, progressed to ESRD	Juvenile-onset NS
Gigante et al. [15]	c.[1169G > C];[1169G > C] p.[(Gly390Ala)];[(Gly390Ala)]	Index case in family with juvenile onset nephrotic syndrome and myoclonic epilepsy (16 years), affected cousin also with juvenile onset nephrotic syndrome (16 years), and headaches associated with phono-and photo- phobia	N/A	Juvenile-onset NS + neurological symptoms
Hara et al. [24], Mitsui et al. [25]	c.[382A > G];[382A > G] p.[(Met128Val)];[(Met128Val)] Additionally homozygous p.(Val393Ala) variant in <i>COQ2</i> also detected	Index case in family; multi-system atro- phy (onset 68 years), RP (diagnosed at 33 years)	N/A	MSA+RP
Hara et al. [24], Mitsui et al. [25]	c.[382A > G];[382A > G] p.[(Met128Val)];[(Met128Val)] Additionally homozygous p.(Val393Ala) variant in <i>COO2</i> also detected	Affected sibling; multi-system atrophy (onset 62 years),RP(diagnosed at 51 years)	N/A	MSA+RP

Table 1 (continued)

References	Genotype:COQ2 (NM_015697.8, NP_056512.5)	Phenotype	Effect of CoQ10 supplementation	Phenotype category
Current study	c.[288dupC];[376C > G] p.[(Ala97Argfs*56)]; [(Arg126Gly)]	Three affected siblings with renal abnormalities and RP	No improvement in ERG function	NS + RP

NS, nephrotic syndrome; ESRD, end-stage renal disease; MSA, multiple system atrophy; RP, retinitis pigmentosa

in patients with retinopathy should be considered as a possible treatment to stabilize remaining retinal function in individuals with the ocular complications associated with pathogenic COQ2 variants. The efficaciousness of CoQ10 may be higher in cases where supplementation is started at an earlier age and prior to the onset of irreversible damage to the retina. In previously reported patients with retinitis pigmentosa stemming from variants in the COQ2 gene, treatment with CoQ10 supplementation was not administered [24, 27]. In two reported patients with variants of the COQ2 gene, after CoQ10 supplementation, neurologic manifestations improved significantly, but there were no noted effects on renal function [9, 19]. In one reported case, a patient with corticosteroid-resistant nephrotic syndrome and encephalomyopathy showed improvement in neurologic symptoms in response to CoQ10 supplementation, but renal function remained unchanged [20]. Response to CoQ10 supplementation seems to vary however, as seen in another case of a patient with a homozygous COQ2 variant exhibiting renal failure [9, 18]. Twenty days after initial CoQ10 supplementation (30 mg per kg of body weight per day), this patient responded to treatment and recovered renal function [9, 18].

Variable response to supplementation by CoQ10 may also be due to variable bioavailability of CoQ10 or CoQ10 analog supplementation. Several studies have investigated the use of CoQ10 precursors that may sidestep the defects in the synthesis pathway, and such precursors may be another consideration for treatment in similar families to the study family described. One such precursor is 4-hydroxybenzoic acid (4-HBA), which is a substrate for COQ2. Although this molecule does not bypass the COQ2-mediated enzymatic step in the CoQ10 synthesis pathway, one study showed that supplementation of COQ2-deficient fibroblasts in vitro with 4-HB appeared to overcome the enzymatic defect in a dose-dependent manner, implying that certain defects may be overcome by saturating the defective COQ2 enzyme with this CoQ10 precursor [30]. Other studies have shown that intermediates further downstream of COQ2, such as β -resorcylic acid $(\beta$ -RA) and vanillic acid may be used to bypass synthesis defects in the CoQ10 pathway more effectively than CoQ10 or its analogs, although these intermediates were used to address deficiencies in enzymes further down the synthesis pathway than COQ2 [31–33]. Further clinical studies will be required to assess the efficacy of these intermediates as viable therapeutic alternatives to primary CoQ10 deficiency syndromes.

Conclusions

We present in this report a rare oculorenal syndrome in three related patients associated with novel compound heterozygous variants in the *COQ2* gene, resulting in primary coenzyme Q10 deficiency that manifests primarily as retinitis pigmentosa and renal failure. Unlike other primary CoQ10 deficiency syndromes previously described, these patients do not display neurological dysfunction at presentation. We believe this rare entity should not be missed when considering the wide differential in patients with oculorenal syndromes, as it is potentially treatable if caught early.

Abbreviations

CoQ10: Coenzyme q10; ES: Exome sequencing; SD-OCT: Spectral domain optical coherence tomography; ERG: Electroretinography; OD: Oculus dexter (right eye); OS: Oculus sinister (left eye); OU: Oculus uterque (both eyes); ISCEV: International Society for Clinical Electrophysiology of Vision; ACMG/ AMP: American College of Medical Genetics and Genomics/Association for Molecular Pathology; NS: Nephrotic syndrome; ESRD: End-stage renal disease; MSA: Multiple system atrophy; RP: Retinitis pigmentosa.

Acknowledgements

Not applicable.

Authors' contributions

A.H.A, S.D.R, J.R.L.C, C.L.X., and S.H.T. examined the patients in the clinic, collected multimodal imaging, made determinations regarding diagnostics and management, performed electroretinography, analyzed the clinical data, and contributed to writing and proofreading the manuscript. A.V.D., A.L.T., C.M.B., M.M.M., J.L., V.J. identified the relevant gene variants in the patients through molecular pathology and genomics techniques, analyzed laboratory and clinical data, performed literature review and contributed to writing and proofreading the manuscript. A.B.N and I.H.M. provided critical insight into diagnosis and management, and contributed to the final manuscript. All authors read and approved the final manuscript.

Funding

Research was supported by the Jonas Children's Vision Care and Bernard & Shirlee Brown Glaucoma Laboratory and by the National Institutes of Health [P30EY019007, U01EY030580, U54OD020351, R24EY027285, 5P30EY019007, R01EY018213, R01EY024698, R01EY026682, and R21AG050437], National Cancer Institute Core [5P30CA013696], Foundation Fighting Blindness [TA-NMT-0116-0692-COLU], the Research to Prevent Blindness (RPB) Physician-Scientist Award, and unrestricted funds from RPB, New York, NY, USA. S.H.T. is a member of the RD-CURE Consortium and is supported by Kobi and Nancy Karp, the Crowley Family Fund, the Rosenbaum Family Foundation, the Tistou and Charlotte Kerstan Foundation, the Schneeweiss Stem Cell Fund, New York State [C029572], and the Gebroe Family Foundation. A.H.A is a recipient of the Heed Ophthalmic Foundation fellowship.

Availability of data and materials

Data sharing not applicable to this article.

Ethics approval and consent to participate

Details included in Methods section.

Consent for publication

Not applicable.

Competing interests

The authors declare no competing interests related to this study.

Author details

¹ Edward S. Harkness Eye Institute, Columbia University Irving Medical Center, New York, NY, USA. ² Laboratory of Personalized Genomic Medicine, Department of Pathology and Cell Biology, Columbia University Irving Medical Center, New York, NY, USA. ³ Department of Ophthalmology, Hospital das Clínicas de Pernambuco (HCPE) - Empresa Brasileira de Serviços Hospitalares (EBSERH), Federal University of Pernambuco (UFPE), Recife, Pernambuco, Brazil.

Received: 4 June 2020 Accepted: 26 October 2020 Published online: 13 November 2020

References

- 1. Quinzii CM, Hirano M. Primary and secondary CoQ(10) deficiencies in humans. BioFactors. 2011;37(5):361–5.
- Salviati L, Trevisson E, Doimo M, Navas P, et al. Primary Coenzyme Q10 Deficiency. In: Adam MP, Ardinger HH, Pagon RA, Wallace SE, Bean LJH, Stephens K, et al., editors. GeneReviews ((R)). Seattle: University of Washington; 1993–2020. 2017.
- McCulloch DL, Marmor MF, Brigell MG, Hamilton R, Holder GE, Tzekov R, et al. ISCEV Standard for full-field clinical electroretinography (2015 update). Documenta ophthalmologica Adv Ophthalmol. 2015;130(1):1–12.
- Kopanos C, Tsiolkas V, Kouris A, Chapple CE, Albarca Aguilera M, Meyer R, et al. VarSome: the human genomic variant search engine. Bioinformatics. 2019;35(11):1978–80.
- Richards S, Aziz N, Bale S, Bick D, Das S, Gastier-Foster J, et al. Standards and guidelines for the interpretation of sequence variants: a joint consensus recommendation of the American College of Medical Genetics and Genomics and the Association for Molecular Pathology. Genet Med. 2015;17(5):405–24.
- Chylack LT Jr, Wolfe JK, Singer DM, Leske MC, Bullimore MA, Bailey IL, et al. The lens opacities classification system III. The longitudinal study of cataract study group. Arch Ophthalmol. 1993;111(6):831–6.
- Desbats MA, Vetro A, Limongelli I, Lunardi G, Casarin A, Doimo M, et al. Primary coenzyme Q10 deficiency presenting as fatal neonatal multiorgan failure. Eur J Human Genet EJHG. 2015;23(9):1254–8.
- Dinwiddie DL, Smith LD, Miller NA, Atherton AM, Farrow EG, Strenk ME, et al. Diagnosis of mitochondrial disorders by concomitant next-generation sequencing of the exome and mitochondrial genome. Genomics. 2013;102(3):148–56.
- Diomedi-Camassei F, Di Giandomenico S, Santorelli FM, Caridi G, Piemonte F, Montini G, et al. COQ2 nephropathy: a newly described inherited mitochondriopathy with primary renal involvement. J Am Soc Nephrol JASN. 2007;18(10):2773–80.
- Eroglu FK, Ozaltin F, Gonc N, Nalcacioglu H, Ozcakar ZB, Yalnizoglu D, et al. Response to early coenzyme Q10 supplementation is not sustained in CoQ10 deficiency caused by CoQ2 mutation. Pediatr Neurol. 2018;88:71–4.
- Jakobs BS, van den Heuvel LP, Smeets RJ, de Vries MC, Hien S, Schaible T, et al. A novel mutation in COQ2 leading to fatal infantile multisystem disease. J Neurol Sci. 2013;326(1–2):24–8.
- Mollet J, Giurgea I, Schlemmer D, Dallner G, Chretien D, Delahodde A, et al. Prenyldiphosphate synthase, subunit 1 (PDSS1) and OH-benzoate polyprenyltransferase (COQ2) mutations in ubiquinone deficiency and oxidative phosphorylation disorders. J Clin Investig. 2007;117(3):765–72.
- Scalais E, Chafai R, Van Coster R, Bindl L, Nuttin C, Panagiotaraki C, et al. Early myoclonic epilepsy, hypertrophic cardiomyopathy and subsequently a nephrotic syndrome in a patient with CoQ10 deficiency caused by mutations in para-hydroxybenzoate-polyprenyl transferase (COQ2). Eur J Paediat Neurol. 2013;17(6):625–30.
- Bezdicka M, Stolbova S, Seeman T, Cinek O, Malina M, Simankova N, et al. Genetic diagnosis of steroid-resistant nephrotic syndrome in a longitudinal collection of Czech and Slovak patients: a high proportion of causative variants in NUP93. Pediatric Nephrol. 2018;33(8):1347–63.

- Gigante M, Diella S, Santangelo L, Trevisson E, Acosta MJ, Amatruda M, et al. Further phenotypic heterogeneity of CoQ10 deficiency associated with steroid resistant nephrotic syndrome and novel COQ2 and COQ6 variants. Clin Genet. 2017;92(2):224–6.
- Lopez-Martin JM, Salviati L, Trevisson E, Montini G, DiMauro S, Quinzii C, et al. Missense mutation of the COQ2 gene causes defects of bioenergetics and de novo pyrimidine synthesis. Hum Mol Genet. 2007;16(9):1091–7.
- McCarthy HJ, Bierzynska A, Wherlock M, Ognjanovic M, Kerecuk L, Hegde S, et al. Simultaneous sequencing of 24 genes associated with steroid-resistant nephrotic syndrome. Clin J Am Soc Nephrol CJASN. 2013;8(4):637–48.
- Montini G, Malaventura C, Salviati L. Early coenzyme Q10 supplementation in primary coenzyme Q10 deficiency. N Engl J Med. 2008;358(26):2849–50.
- Quinzii C, Naini A, Salviati L, Trevisson E, Navas P, Dimauro S, et al. A mutation in para-hydroxybenzoate-polyprenyl transferase (COQ2) causes primary coenzyme Q10 deficiency. Am J Hum Genet. 2006;78(2):345–9.
- Salviati L, Sacconi S, Murer L, Zacchello G, Franceschini L, Laverda AM, et al. Infantile encephalomyopathy and nephropathy with CoQ10 deficiency: a CoQ10-responsive condition. Neurology. 2005;65(4):606–8.
- Starr MC, Chang JJ, Finn LS, Sun A, Larson AA, Goebel J, et al. COQ2 nephropathy: a treatable cause of nephrotic syndrome in children. Pediatric Nephrol. 2018;33(7):1257–61.
- 22. Wu X, Wang W, Liu Y, Chen W, Zhao L. A steroid-resistant nephrotic syndrome in an infant resulting from a consanguineous marriage with COQ2 and ARSB gene mutations: a case report. BMC Med Genet. 2019;20(1):165.
- Xu K, Mao XY, Yao Y, Cheng H, Zhang XJ. Clinical analysis of one infantile nephrotic syndrome caused by COQ2 gene mutation and literature review. Zhonghua er ke za zhi = Chin J Pediatr. 2018;56(9):662–6.
- Hara K, Momose Y, Tokiguchi S, Shimohata M, Terajima K, Onodera O, et al. Multiplex families with multiple system atrophy. Arch Neurol. 2007;64(4):545–51.
- Mitsui J, Koguchi K, Momose T, Takahashi M, Matsukawa T, Yasuda T, et al. Three-year follow-up of high-dose ubiquinol supplementation in a case of familial multiple system atrophy with compound heterozygous COQ2 mutations. Cerebellum. 2017;16(3):664–72.
- Desbats MA, Morbidoni V, Silic-Benussi M, Doimo M, Ciminale V, Cassina M, et al. The COQ2 genotype predicts the severity of coenzyme Q10 deficiency. Hum Mol Genet. 2016;25(19):4256–65.
- 27. Multiple-System Atrophy Research C. Mutations in COQ2 in familial and sporadic multiple-system atrophy. N Engl J Med. 2013;369(3):233–44.
- Boitier E, Degoul F, Desguerre I, Charpentier C, Francois D, Ponsot G, et al. A case of mitochondrial encephalomyopathy associated with a muscle coenzyme Q10 deficiency. J Neurol Sci. 1998;156(1):41–6.
- Rotig A, Appelkvist EL, Geromel V, Chretien D, Kadhom N, Edery P, et al. Quinone-responsive multiple respiratory-chain dysfunction due to widespread coenzyme Q10 deficiency. Lancet. 2000;356(9227):391–5.
- Herebian D, Seibt A, Smits SHJ, Rodenburg RJ, Mayatepek E, Distelmaier F. 4-Hydroxybenzoic acid restores CoQ10 biosynthesis in human COQ2 deficiency. Ann Clin Transl Neurol. 2017;4(12):902–8.
- Herebian D, Lopez LC, Distelmaier F. Bypassing human CoQ10 deficiency. Mol Genet Metab. 2018;123(3):289–91.
- Herebian D, Seibt A, Smits SHJ, Bunning G, Freyer C, Prokisch H, et al. Detection of 6-demethoxyubiquinone in CoQ10 deficiency disorders: insights into enzyme interactions and identification of potential therapeutics. Mol Genet Metab. 2017;121(3):216–23.
- Hidalgo-Gutierrez A, Barriocanal-Casado E, Bakkali M, Diaz-Casado ME, Sanchez-Maldonado L, Romero M, et al. β-RA reduces DMQ/CoQ ratio and rescues the encephalopathic phenotype in Coq9^{R239X} mice. EMBO Mol Med. 2019;11(1):e9466.

Publisher's Note

Springer Nature remains neutral with regard to jurisdictional claims in published maps and institutional affiliations.