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Saikosaponin D exerts antidepressant effect by regulating Homer1-mGluR5 and mTOR signaling in a rat model of chronic unpredictable mild stress

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Abstract

Background Many studies about depression have focused on the dysfunctional synaptic signaling in the hippocampus that drives the pathophysiology of depression. *Radix Bupleuri* has been used in China for over 2000 years to regulate liver-qi. Extracted from *Radix Bupleuri*, Saikosaponin D (SSD) is a pharmacologically active substance that has antidepressant effects. However, its underlying mechanism remains unknown.

Materials and methods A chronic unpredictable mild stress (CUMS) paradigm was used as a rat model of depression. SD rats were randomly assigned to a normal control (NC) group or one exposed to a CUMS paradigm. Of the latter group, rats were assigned to four subgroups: no treatment (CUMS), fluoxetine-treated (FLU), high-dose and low-dose SSD-treated (SSDH and SSDL). SSD was orally administered of 1.50 mg/kg and 0.75 mg/kg/days for three weeks in the SSDH and SSDL groups, respectively. Fluoxetine was administered at a dose of 2.0 mg/kg/days. SSD's antidepressant effects were assessed using the open field test, forced swim test, and sucrose preference test. Glutamate levels were quantified by ELISA. Western blot and immunochemical analyses were conducted to quantify proteins in the Homer protein homolog 1 (Homer1)-metabotropic glutamate receptor 5 (mGluR5) and mammalian target of rapamycin (mTOR) pathways in the hippocampal CA1 region. To measure related gene expression, RT-qPCR was employed.

Results CUMS-exposed rats treated with SSD exhibited increases in food intake, body weight, and improvements in the time spent in the central area and total distance traveled in the OFT, and less pronounced pleasure-deprivation behaviors. SSD also decreased glutamate levels in CA1. In CA1 region of CUMS-exposed rats, SSD treatment increased mGluR5 expression while decreasing Homer1 expression. SSD also increased expressions of postsynaptic density protein 95 (PSD95) and synapsin I (SYP), and the ratios of p-mTOR/mTOR, p-p70S6k/p70S6k, and p-4E-BP1/4E-BP1 in the CA1 region in CUMS-exposed rats.

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Conclusions SSD treatment reduces glutamate levels in the CA1 region and promotes the expression of the synaptic proteins PSD-95 and SYP via the regulation of the Homer1-mGluR5 and downstream mTOR signaling pathways. These findings suggest that SSD could act as a natural neuroprotective agent in the prevention of depression.

Highlights

- Saikosaponin D, a pharmacologically active substance isolated from the Radix *Bupleuri* has been shown to have antidepressant effects.
- Its effects are mediated through the Homer1-mGluR5 and mTOR pathways.
- The findings support saikosaponin D's actions as a neuroprotective agent.

Keywords Saikosaponin D, Chronic unpredictable mild stress, Homer1-mGluR5, mTOR

Introduction

The effects of depression are widespread, as it disrupts productivity, impairs social interaction and overall health, reduces life expectancy, and increases the burden on the healthcare system [1]. In the last ten years, there has been growing concern regarding the impact of depression on global health, especially in low-income countries. Furthermore, the World Health Organization has predicted that depression will be the most disabling disease in the world by 2030 [2], highlighting the need for more studies to understand the pathophysiology of the disease and develop novel therapeutic regimens for depression that have fewer side effects. Among the studied compounds, Saikosaponin D (SSD) is being increasingly recognized due to its neuroprotective effects [3–5].

Radix Bupleuri, the root of *Bupleurum chinense* DC. or *Bupleurum scorzonerifolium* Willd., is a common component of Chinese herbal prescriptions such as Xiaoyaosan [6] and Chaihu-Shugan-San [7], which are administered clinically to improve symptoms of depression. Saikosaponin D (SSD) is a pharmacologically active ingredient extracted from *Radix Bupleuri* [8, 9], which is capable of 'dispersing stagnated liver qi for relieving qi stagnation' according to traditional Chinese Medicine (TCM) theory. Several researches reported that SSD was founded to have the effect to inhibit neuronal apoptosis and microglia activation as well as neuroinflammation in LPS-induced mice [3, 5]. These studies suggest that SSD could combat depression-like behaviors caused by inflammation. However, there are few studies on the effect of SSD on chronic stress-induced depression, and the underlying mechanism is still unclear. Chronic stress is closely related to neurological disorders. CUMS is a common rodent stress model that is frequently used in modern research investigating depression [10]. Katz and colleagues first reported the CUMS model [11]. Therefore, it is of important meaning to research the underlying

mechanism of SSD in CUMS-induced rats in order to develop further therapeutic options for depression.

Recent clinical as well as basic research studies have linked the pathogenesis of depression with synaptic dysfunction in the prefrontal cortex and hippocampus [12], specifically within the hippocampal CA1 region. Chronic stress affects the hippocampus, a key region containing marginal structures involved in the processing of memories and emotional information. Neurological diseases can be caused by damage to the hippocampal CA1 region [13], which is also one of the brain regions most often studied in rodent models of depression [14, 15]. Furthermore, there is new evidence suggesting that Chinese herbal medicines may have antidepressant benefits resulting from their ability to upregulate the certain synaptic proteins expression [16–18]. Meanwhile, an increasing body of evidence suggests that Homer protein homolog 1 (Homer1)-metabotropic glutamate receptor 5 (mGluR5) and downstream mammalian target of rapamycin (mTOR) signaling pathways affect the expression of proteins involved in the modulation of synaptic plasticity that may drive the pathophysiology of depression. The pathogenesis of and the therapy to treat mood disorders are dependent on changes in mGluRs. For example, antidepressant-like behavior is induced by mGluR5 knockout [19], consistent with the mGluR5^{-/-} mice, an antidepressant-like phenotype mice [20]. Located in the post-synaptic density, Homer1, a postsynaptic scaffold protein serves as a prominent link between group I mGluRs and downstream targets [21, 22]. It is expressed in two main isoforms, namely Homer1a and Homer1b/c isoforms, the latter of which is constitutively expressed, maintains synaptic transmission, and dendritic spine structure [23]. Genome-wide association studies (GWAS) have shown that Homer1 is a key target for mood disorders and antidepressants [24, 25]. The mTOR signaling pathway is a downstream intracellular signaling pathway that regulates synaptic protein synthesis and contributes to

synaptogenesis [26]. Previous studies have suggested that activation of mGluR5 subtypes is required for mTOR phosphorylation in the hippocampal CA1 region [27]. When activated, mTOR complex 1 (mTORC1) causes the phosphorylation and activation of p70S6 kinase (S6K) and eukaryotic translation initiation factor 4E binding protein 1 (4E-BP1), resulting in the upregulation of synaptic proteins [e.g., postsynaptic density protein-95 (PSD-95) and synapsin I (SYP)] and the production of new synapses [28, 29]. Although SSD has been reported to exhibit antidepressant capacity, the underlying mechanism of action remains unclear. Therefore, the aim of this study was to identify the signaling pathways that mediate the antidepressant effects of SSD. We hypothesized that SSD promotes synaptic protein production in CUMS-exposed rats through the modulation of the Homer1-mGluR5 and mTOR signaling pathways.

Materials and methods

CUMS model establishment

Sixty 7-week-old male Sprague–Dawley rats were provided by the Beijing Vital River Laboratory Animal Research Center [license No. SCXK (Beijing) 2016–0006]. All the rats were housed in the Beijing University of Chinese Medicine's specific pathogen-free clean animal facility [license No. SYXK (Jing) 2016–0038, conventional breeding]. All animal experiments were approved by the Institutional Animal Care and Use Committee of Beijing University of Chinese Medicine. The experiments were conducted in strict accordance with the recommendations of the U.S. National Institutes of Health's Guide for the Care and Use of Laboratory Animals.

After a week of adaptation, 48 rats were assigned to the CUMS group randomly. These rats were subjected to a variety of randomly ordered, low-intensity social and environmental stressors for an extended period of time over the course of 6 weeks as previously described [30]. The selected stressors and schedule of exposure are summarized in Table 1. The 12 remaining rats were assigned to the normal control (NC) group and were not exposed to any stressors.

Drug administration

Three weeks after of exposed to CUMS, the CUMS-exposed rats were divided into four subgroups: a CUMS group (no drug treatment), a fluoxetine-treated group (FLU), and SSD high-dose (SSDH) and SSD low-dose (SSDL) groups. SSD was dissolved in dimethyl sulfoxide (DMSO) at a concentration of 0.1 percent. Drug administration was performed from the 4th week to the end. The rats in SSD high and low dose groups were treated with SSD (CAS Registry Number. 20874–52-6, PubChem CID: 107,793, 98% purity, batch number: B20150, Yuanye

Table 1 The schedule of stressors used in CUMS paradigm

Stressors	Days of CUMS experiments
24-h water deprivation	1,15,27
overnight illumination	2,16,25,36
24-h food deprivation	3,19, 33,37
5-min heat exposure at 45 °C	10,20,31,
17-h white noise exposure	4,13,22, 26,41
17-h cage tilt at 45 degrees from the horizontal	5,21,29,34,40
24-h moist padding exposure	6,11,17,38
1-min tail pinch	8,12,18,23,30
3-h physical restraint	9,24,32,39
17-h strange object exposure	7,14,28,35,42

Bio-Technology, Shanghai, China) daily with 1.50 mg/kg and 0.75 mg/kg per day [31] by gavage, respectively. Rats were administered fluoxetine (2 mg/kg) in the FLU group, and rats received equal volumes of 0.1% DMSO by gavage in NC and CUMS groups. Figure 1A shows the SSD's molecular structure. Figure 1B displays the experimental timeline.

Behavioral testing

At the end of the drug treatment period, open field test (OFT), the sucrose preference test (SPT), and the forced swim test (FST) were conducted as previously described [32]. Details can be found in Additional file 1: Materials and Methods.

Tissue sample collection

After a 12-h fasting period at the conclusion of the sixth week, six rats were randomly selected from each group. Rats were anesthetized with one percent sodium pentobarbital i.p., and arterially perfused with 4% paraformaldehyde. Then conventional paraffin embedding was performed. Using an Rat Brain Slicer Matrix (BSRAS001-1, ZIVIC Instrument, USA), the brains of the remaining 30 non-perfused rats were uniformly sectioned into coronal slices. Hippocampal CA1 tissue were isolated under an anatomical microscope.

Enzyme-linked immunosorbent assays (ELISA)

Glutamate levels in each rats' CA1 region were measured using an ELISA kit (Bio-Assay Systems, #EGLT-100).

Quantitative real-time polymerase chain reaction (RT-qPCR)

The mRNA levels of mGluR5, Homer1, PSD-95 and SYP were quantified by RT-qPCR. RT-qPCR were performed as described [33] and detailed described in Additional

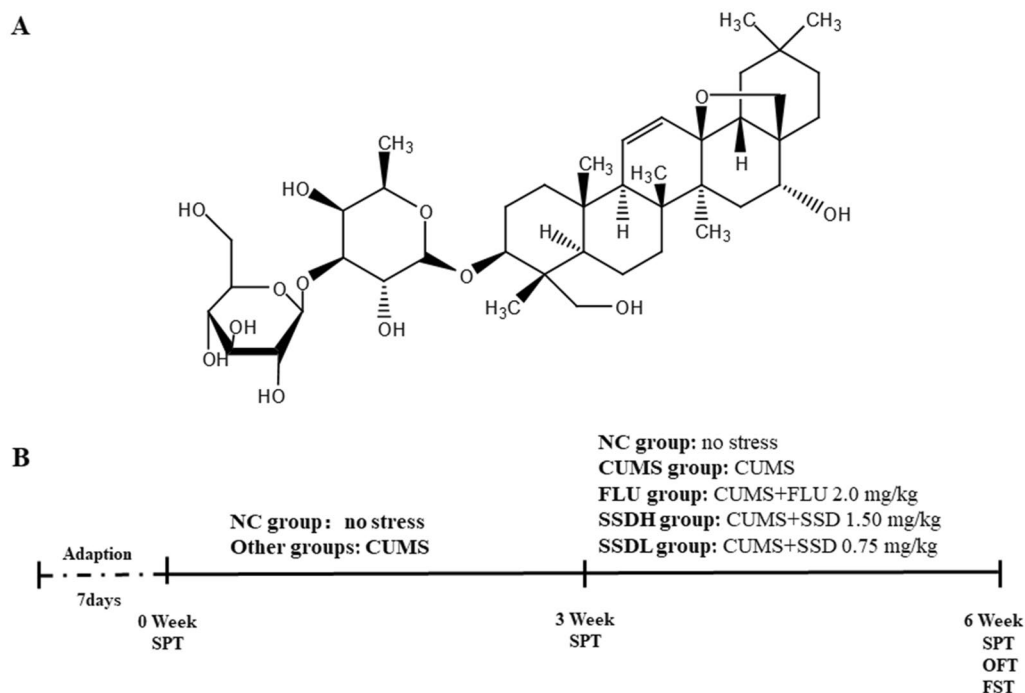


Fig. 1 The molecular structure of SSD is shown in **A**. The experimental timeline is shown in **B**. The animals were provided a 7-day adaptation period prior to the start of the experiment. Except for the rats in the NC group, other rats were subjected to CUMS for six weeks and treated with different drugs from the 4th week to the end

Table 2 Forward and reverse primer sequences

Gene	Sequences
<i>GluR5</i>	F: CATGGAGCAGATCAGCAGCGTAG R: ATCAGGTAGGAGGAGCAGATTGGAG
<i>SYP</i>	F: GCTGTGTTTGCCTTCTACTC R: TGATAATGTTCTCTGGGTCCGTG
<i>Homer1</i>	F: AGGAGAAGTCGCAGGAGAAGATGG R: CCTTGGCTCTGAGTTCTGTGCAC
<i>PSD-95</i>	F: TCCAGTCTGTGCGAGAGGTAGC R: GGACGGATGAAGATGGCGATGG
<i>GAPDH</i>	F: CCATTCTTCCACCTTTGAT R: TGGTCCAGGGTTTCTTACT

file 1: Materials and Methods. The primer sequences shown in Table 2 were synthesized by Sangon Biotech Co., Ltd. (Shanghai, China).

Western blot analysis

Homer1b/c, mGluR5, p-mTOR, mTOR, p70S6 kinase, p-p70S6 kinase, p-4E-BP1, 4E-BP1, PSD-95 and SYP expressions in hippocampal CA1 region of experimental rats were detected by Western blot analysis according

to the protocol [6]. Details are listed at Additional file 1: Materials and Methods.

IHC

The brain tissue was embedded in paraffin and cut into 5 μ m sections for IHC experiment as described [6]. See Additional file 1: Materials and Methods for more details.

Statistical analysis

Statistical analyses were performed using Statistical Package for the Social Sciences (SPSS) software (version 22.0; IBM Corp., Armonk, New York, NY, USA). Statistical significance was defined as $P < 0.05$. See Additional file 1: Materials and Methods for more details.

Results

SSD administration increases the body weight and food intake of CUMS rats

A rat model of CUMS (Fig. 1 and Table 1) was established over a 6-week period, throughout which weekly body weight and food intake were assessed. At week 0, there was no statistically significant differences in body weight among all groups of rats, as shown in Fig. 2A. The weekly body weight of rats in the CUMS group significantly decreased compared with that of the NC group ($P < 0.05$) in the 4th, 5th, and 6th weeks. From week 4 to

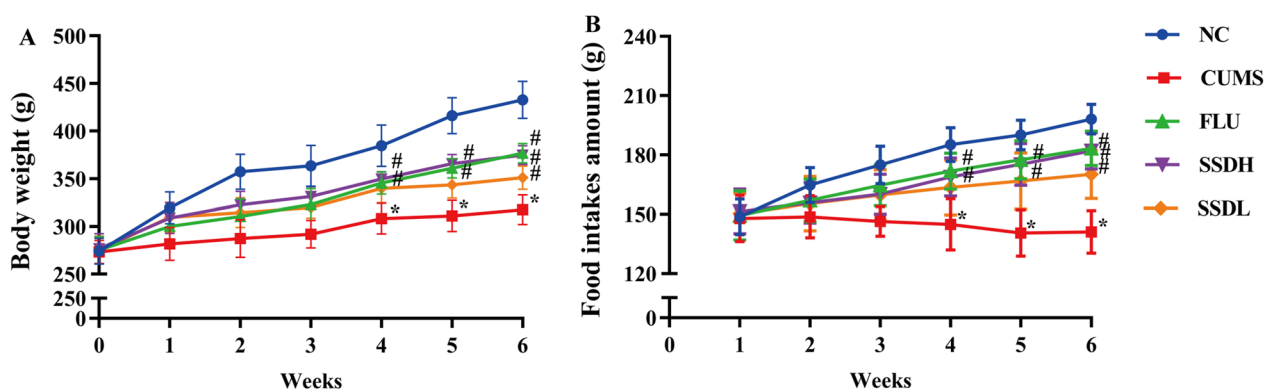


Fig. 2 Changes in body weight and food intake in the CUMS rats. **A** Changes in body weight from week 0 to week 6. **B** Changes in food intake from week 1 to week 6. The data are expressed as the mean \pm SD. * $P < 0.05$ compared to the control group, # $P < 0.05$ compared to the CUMS group; $n = 12$. CUMS, chronic unpredictable mild stress; SD, standard deviation

week 6, FLU, SSDH and SSDL treatment increased the rats' body weight significantly compared with that of rats in the CUMS group ($P < 0.05$). Figure 2B shows that the food intake of the rats in the five different groups did not initially vary. However, from the second to the sixth week, there was a significant difference in food intake between the NC and CUMS groups ($P < 0.05$), and during weeks 4~6, the levels of food intake of the rats in the SSD and FLU groups were higher than that of the rats in the CUMS group ($P < 0.05$).

SSD ameliorates depression-like behaviors of CUMS rats

OFT, FST, and SPT were selected, as these are three classic behavioral tests for assessing depression-like behavior. Figure 3A shows the trajectory of the rats. The amount of time spent in the central region (Fig. 3B) was significantly shorter for the rats in the CUMS groups than in those of the NC group ($P < 0.05$). Rats in the FLU and SSDH groups spent more time in the central area than those of the CUMS group ($P < 0.05$). There was no significant difference in the time spent in the central area between the CUMS group and the SSDL-treated groups. The total distances travelled (Fig. 3C) by rats in the CUMS groups were shorter than those of the NC group ($P < 0.05$). Rats in the FLU, SSDH, and SSDL groups spent more time travelling in the central area than those in the CUMS group ($P < 0.05$). The SPT results are shown in Fig. 3D. The immobility times of rats in the CUMS groups were longer than that of the NC group ($P < 0.05$). Rats in the SSDH and FLU groups exhibited lower immobility times than rats in the CUMS group ($P < 0.05$). There was no significant difference in immobility time between rats in the CUMS and SSDL groups.

The SPT is the gold standard for evaluating decreases in pleasure in rodents. Anhedonia is characterized by a lack of interest in rewarding stimuli, which is a common

symptom of affective disorders such as depression. The SPT was conducted on days 0, 21, and 41 (Fig. 3E). There was no significant difference in sucrose preference among the five groups at baseline. However, three weeks later, rats exposed to the CUMS paradigm (the CUMS, FLU, SSDH, and SSDL groups) exhibited a significant decrease in sucrose preference compared with that of the NC group ($P < 0.05$). This trend was reversed in the FLU and SSDH groups after three weeks of drug administration, with significantly higher sucrose preference rates compared with that of the CUMS group ($P < 0.05$). Therefore, a low dose of SSD is not as effective as a high dose in terms of the drug's ability to improve depression-like behavior.

SSD decreases glutamate levels in the hippocampal CA1 area of CUMS-exposed rats

As shown in Fig. 4, glutamate levels in the CA1 area of the rats in the CUMS group were significantly higher than those in the NC group ($P < 0.05$). This outcome was attenuated in the FLU, SSDH, and SSDL groups ($P < 0.05$).

SSD regulates Homer1b/c and mGluR5 signaling in the hippocampal CA1 region of CUMS-exposed rats

In this study, we evaluated changes in Homer1b/c and mGluR5 expression in the hippocampal CA1 region of CUMS-exposed rats. IHC labeling revealed that the expression of Homer1b/c was significantly higher in the CUMS group than in the NC group, as illustrated in Fig. 5A and B ($P < 0.05$). In the FLU, SSDH, and SSDL groups, the expression of Homer1b/c in the hippocampal CA1 region was significantly higher than that of the CUMS group ($P < 0.05$). The results of the western blot (Fig. 5E) and RT-qPCR (Fig. 5G) experiments were consistent with the IHC results.

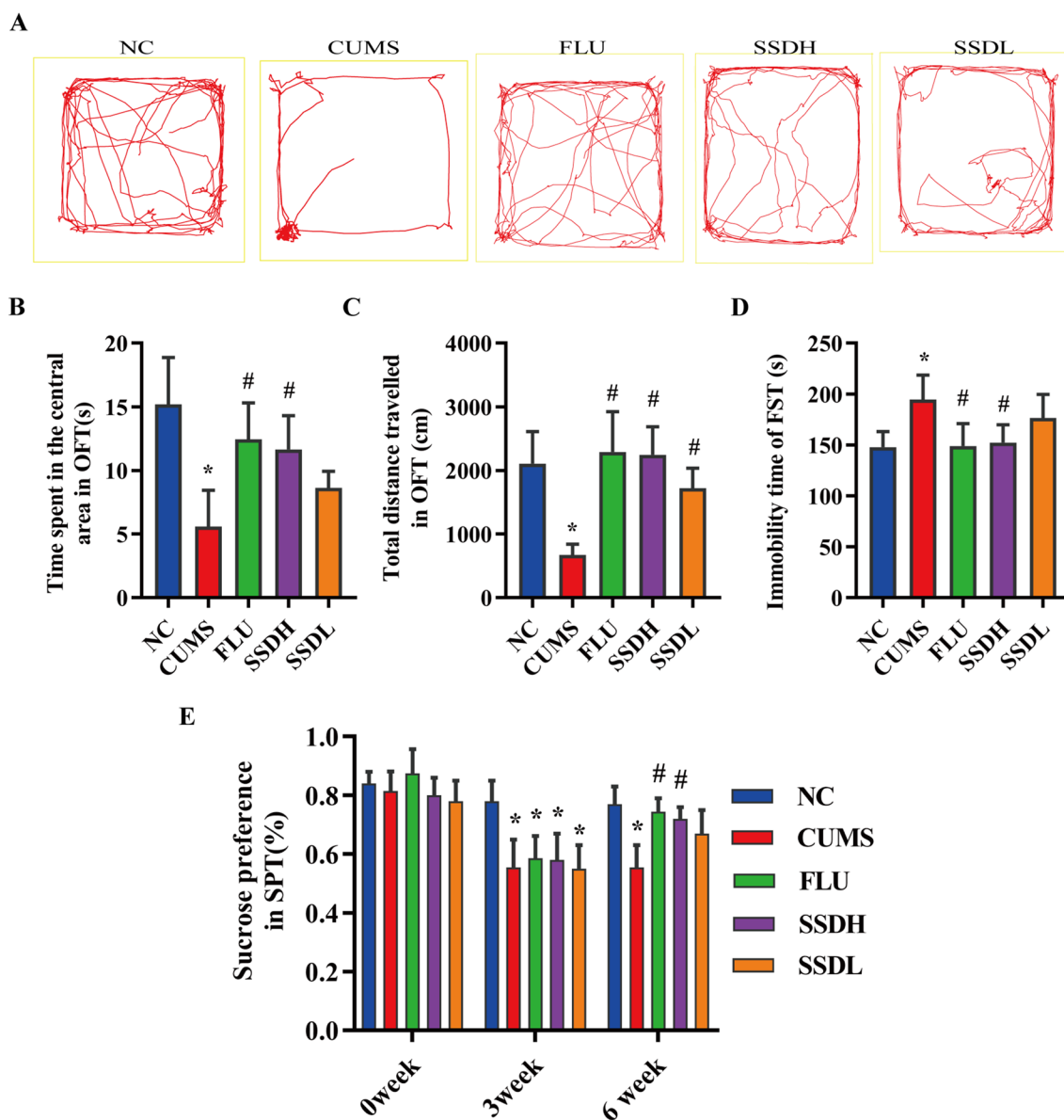


Fig. 3 SSD ameliorates depression-like behaviors in CUMS-exposed rats. **A** Movement trajectory of the rats in each group. **B** The time spent in the central area of the OFT apparatus by each group of rats. **C** The total distance travelled by the rats in each group over a 5-min period in the OFT. **D** Immobility time of the rats in each group in the FST. **E** Sucrose preference as assessed by the SPT of the rats in each group at weeks 0, 3, and 6. All data are expressed as the mean \pm SD. * $P < 0.05$ compared to the control group, # $P < 0.05$ compared to the CUMS group; $n = 12$

As shown in Fig. 5C and D, IHC labeling showed that the expression levels of mGluR5 in the hippocampal CA1 regions of rats in the CUMS group were reduced compared with those in the NC group ($P < 0.05$). This change was significantly attenuated in the FLU, SSDH, and SSDL groups, all of which exhibited higher levels of mGluR5 expression than that of the CUMS group in the hippocampal CA1 region ($P < 0.05$). The results of the western blot (Fig. 5F) and RT-qPCR (Fig. 5H) experiments were consistent with those of the IHC.

SSD regulates the mTOR pathway in CUMS-exposed rats

Proteins and genes in the mTOR pathway were quantified in the hippocampal CA1 area of the rat brain by western blot, IHC labeling, and RT-qPCR to further investigate the probable mechanisms of action of SSD in CUMS-exposed rats. As shown in Fig. 6A, B, the expression of p-mTOR/ mTOR was significantly lower in the hippocampal CA1 area of rats in the CUMS group compared with that of the NC group ($P < 0.05$). Interestingly, after three weeks of SSD administration, the expression

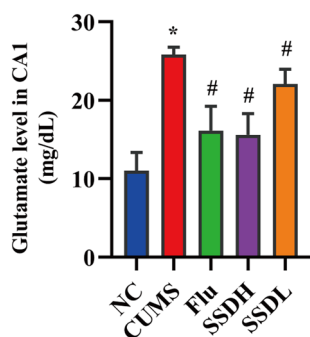


Fig. 4 Glutamate levels in the hippocampus CA1 region of rats in each group. The data are expressed as the mean \pm SD. * $P < 0.05$ compared to the control group, # $P < 0.05$ compared to the CUMS group; $n = 12$

of p-mTOR in the hippocampal CA1 region was significantly upregulated ($P < 0.05$). The results of the western blot experiments showed that the ratio of p-mTOR/mTOR was decreased in rats of the CUMS group compared with that of rats in the NC group ($P < 0.05$). Figure 6C and D show that the level of expression of p-p70s6k was significantly reduced in the hippocampal CA1 area of the CUMS group compared with that of the NC group ($P < 0.05$). However, CUMS rats who were administered SSD exhibited increased p-p70s6k expression in the CA1 region of the hippocampus compared with that of CUMS-exposed rats who were not ($P < 0.05$). As shown in Fig. 6E and F, the level of expression of p-4E-BP1 was significantly lower in the hippocampal CA1 area of the rats in the CUMS group than in those of the NC group ($P < 0.05$). However, the expression of p-4E-BP1 in rats of the SSDH group was higher than that of the CUMS group ($P < 0.05$). The results of the western blot experiments evaluating the ratios of p-mTOR/mTOR (Fig. 6G), p-p70s6k/p70s6k (Fig. 6H) and p-4E-BP1/4E-BP1 (Fig. 6I) were consistent with the IHC results.

SSD regulates PSD-95 and SYP expression in the hippocampal CA1 region of CUMS-exposed rats

We investigated changes in the expression of the synaptic markers PSD-95 and SYP in the hippocampal CA1 region in CUMS-exposed rats. As shown in Fig. 7A and B, the IHC labeling revealed that the expression of PSD-95 in the hippocampal CA1 region of the rats in the CUMS group was significantly lower than that of the NC group ($P < 0.05$). Both FLU and SSDH treatment increased the expression of PSD-95 in the hippocampal CA1 region of rats compared with levels seen in the CUMS group ($P < 0.05$). There was no significant difference in the expression of PSD-95 between the CUMS and SSDL groups. The results of the western blot (Fig. 7E) and

RT-qPCR (Fig. 7G) experiments were consistent with the IHC results. Figure 7C and D show that the expression levels of SYP in the hippocampal CA1 region of the rats in the CUMS group were decreased compared with levels in the NC group ($P < 0.05$). Both FLU and SSD administration significantly increased the expression of SYP in the hippocampal CA1 region of rats compared with that of the CUMS group ($P < 0.05$). The results of the western blot (Fig. 7F) and RT-qPCR (Fig. 7H) experiments were consistent with those of the IHC experiments.

Discussion

In this study, we made the following discoveries: (i) CUMS-exposed rats treated with SSD exhibited increased body weight and food intake, with a reduction in depression-like behaviors; (ii) SSD administration decreased glutamate levels in the rats' CA1 region of CUMS-exposed; (iii) SSD treatment resulted in downregulated Homer1b/c expression as well as upregulated mGluR5, mTOR, PSD-95, and SYP expression, and increased p-mTOR/mTOR, p-p70S6k/p70S6k, and p-4E-BP1/4E-BP1 expression ratios in the CUMS-exposed rats' CA1 region. From these results, we can conclude that synaptic damage caused by the elevated glutamate levels induced by CUMS could be reversed by SSD treatment. This amelioration of glutamate-induced synaptic damage appears to result from regulation of the Homer1-mGluR5 as well as downstream mTOR pathways (Fig. 8).

The CUMS, which is a common rodent stress model, involves the continuous and uninterrupted exposure of rodents to chronic stress environments; the chronic exposure to unpredictable micro-stressors in rats or mice leads to pronounced behavioral changes, including reduced responsivity to rewarding stimuli, and, clinically, the reduction of appetite [34]. There are behavioral correlations with core symptoms of depression [35]. FST, OFT and SPT were the commonly methods used in the research of depressive-like behavior in rodents. The FST it involves the exposure of animals to stress, which was shown to have a role in the tendency for major depression. The OFT can reflect an animal's exploration characteristics and fear of a new environment, which can be used to quantitatively evaluate the animal's spontaneous activity, exploration behavior, and the state of depression. SPT is an anhedonia test that measures sucrose consumption in rodents [36]. For decades, it has been known that stress is closely linked to the development of neurological disorders [11], and a previous study demonstrated that rats exposed to CUMS for 6 weeks developed depression-like behaviors [6]. LPS-induced mice produced depressive-like behaviors, including increased immobility time in FST, reduced movement distance

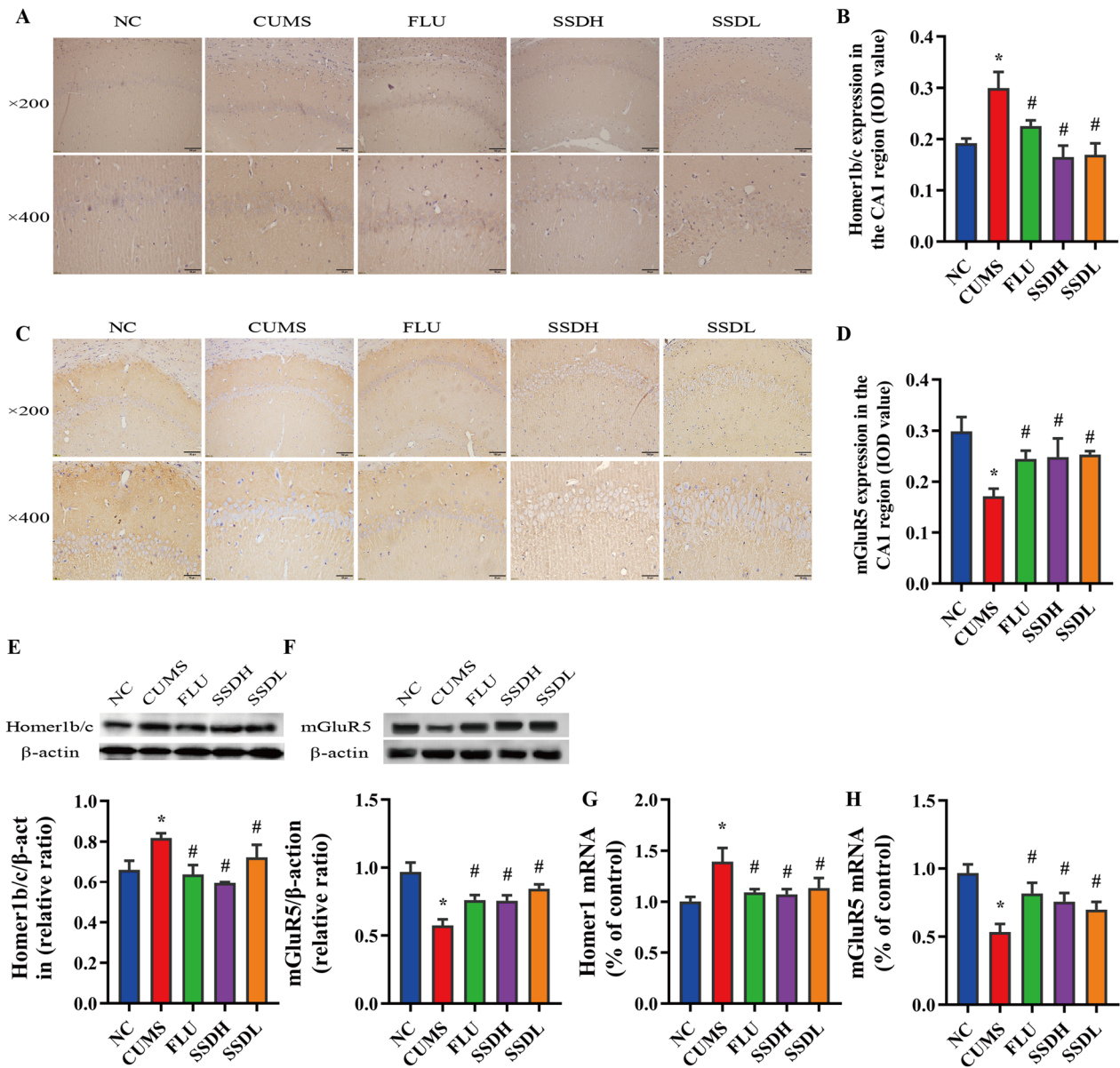


Fig. 5 SSD treatment increases the expression of Homer1 and mGluR5 in the hippocampal CA1 area of the brains of CUMS-exposed rats. Images of IHC labeling at the original magnification ($\times 200$ and $\times 400$) and the respective IOD values of Homer1 (A, B) and mGluR5 (C, D) expression in the hippocampi of CUMS-exposed rats. (E, F) show representative western blot images and the relative ratios of Homer1 and mGluR5 expression in the hippocampal CA1 region of the different groups of rats exposed to the CUMS paradigm. (G, H) show the Homer1 and mGluR5 mRNA levels in the hippocampal CA1 region of the rats in each group. All data are expressed as the mean \pm SD. * $P < 0.05$ compared to the control group, # $P < 0.05$ compared to the CUMS group; $n = 6$. SSD

(See figure on next page.)

Fig. 6 Treatment of CUMS-exposed rats with SSD results in elevated expression of p-mTOR, p-p70s6k, and p-4E-BP1 in the hippocampal CA1 region. IHC labeling at the original magnification ($\times 200$ and $\times 400$) and the respective IOD values of p-mTOR (A, B), p-p70s6k (C, D), and p-4E-BP1 (E, F) expression in the hippocampal CA1 region of CUMS rats. (G-I) show representative western blot images and the relative ratios of p-mTOR, p-p70s6k, and p-4E-BP1 expression in the hippocampal CA1 regions of the different groups of CUMS-exposed rats. All data are expressed as the mean \pm SD. * $P < 0.05$ compared to the control group, # $P < 0.05$ compared to the CUMS group; $n = 6$

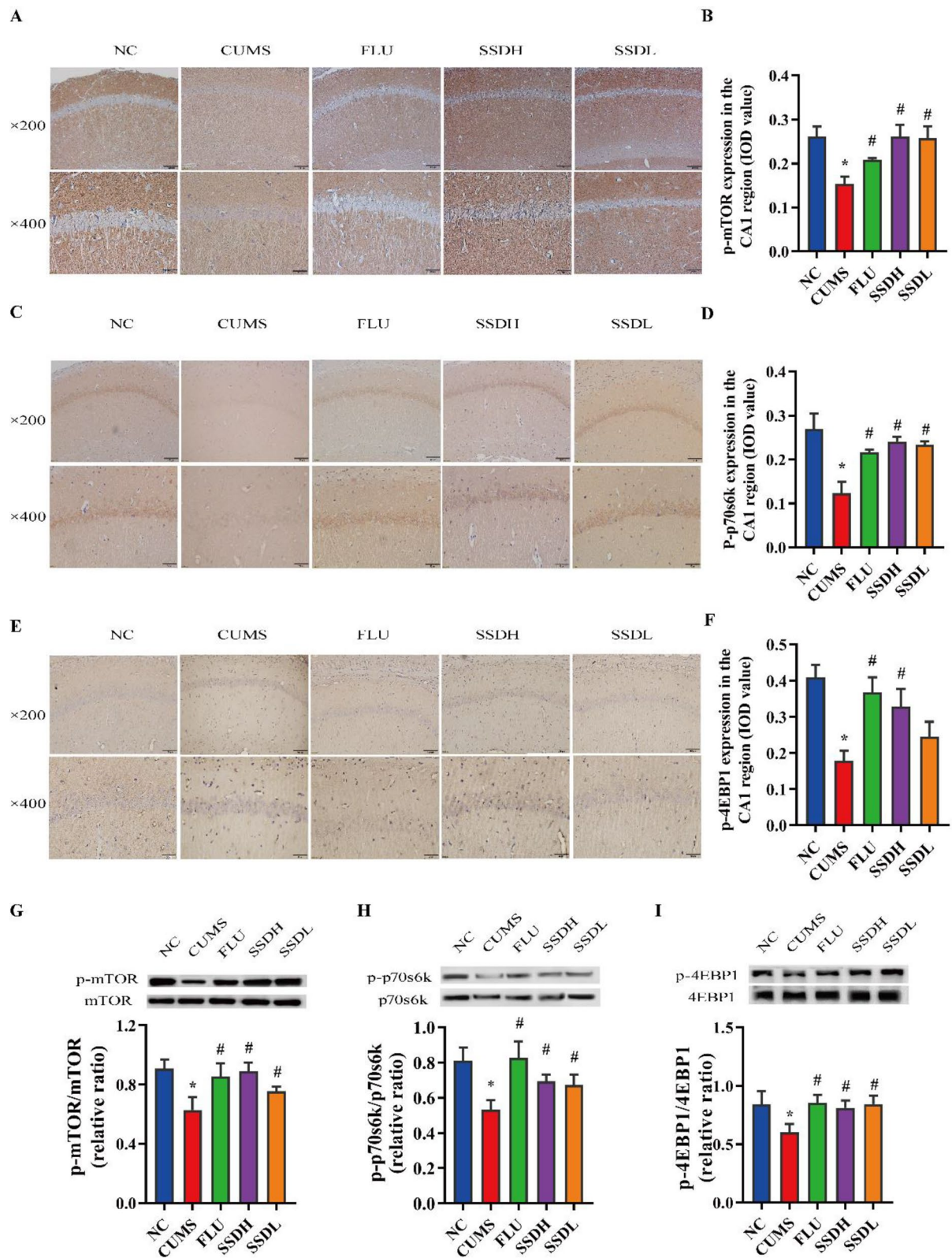


Fig. 6 (See legend on previous page.)

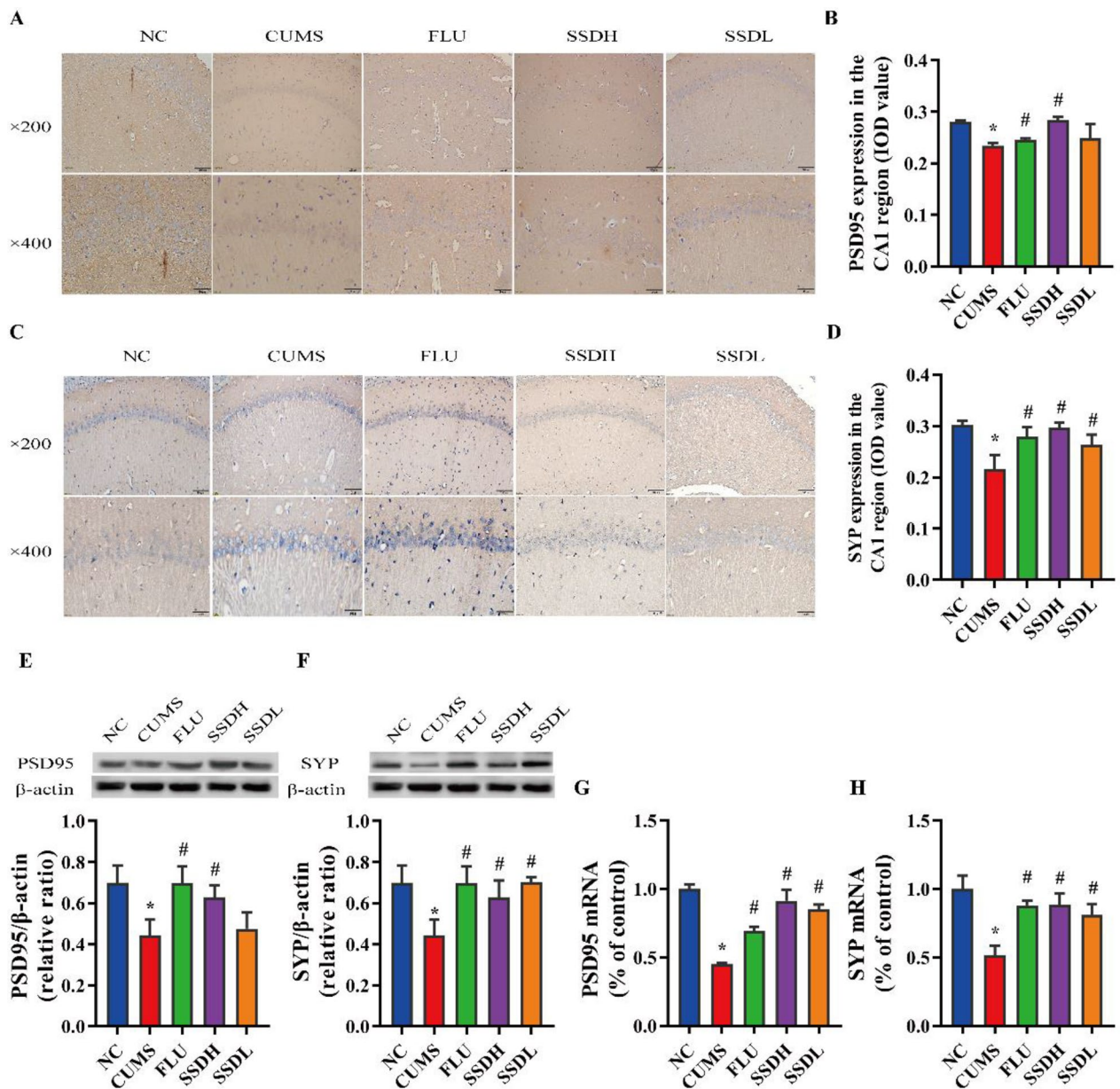


Fig. 7 Treatment of CUMS-exposed rats with SSD results in elevated expression of PSD-95 and SYP in the hippocampal CA1 region. Images of IHC labeling at the original magnification (×200 and ×400) and the respective IOD values of PSD-95 (A, B), and SYP (C, D) expression in the hippocampal tissue of CUMS-exposed rats. (E, F) show representative western blot images and the relative ratios of PSD-95 and SYP expression in the hippocampal CA1 regions of the different groups of CUMS-exposed rats. (G, H) show the PSD-95 and SYP mRNA levels in the hippocampal CA1 region of the rats in each group. All data are expressed as the mean ± SD. **P* < 0.05 compared to the control group, #*P* < 0.05 compared to the CUMS group; n = 6

and shortened stay time in the central region in the OFT, reduced sucrose consumption, which were consistent with depressive-like behaviors shown in this study [3, 5].

Study we explored, SSD administration improved depression-like behavior in rats exposed to CUMS. More specifically, rats in the SSDH and SSDL groups spent more time and travelled more than rats in the CUMS

group in the OFT central area, and exhibited lower immobility time than those in the CUMS group. Loss of appetite is another important indicator of depression [34]. According to reports, 48 percent of individuals suffering from depression lose their appetite [37]. In the present study, SSD treatment to rats elevated both the food intake and body weight compared to that of rats

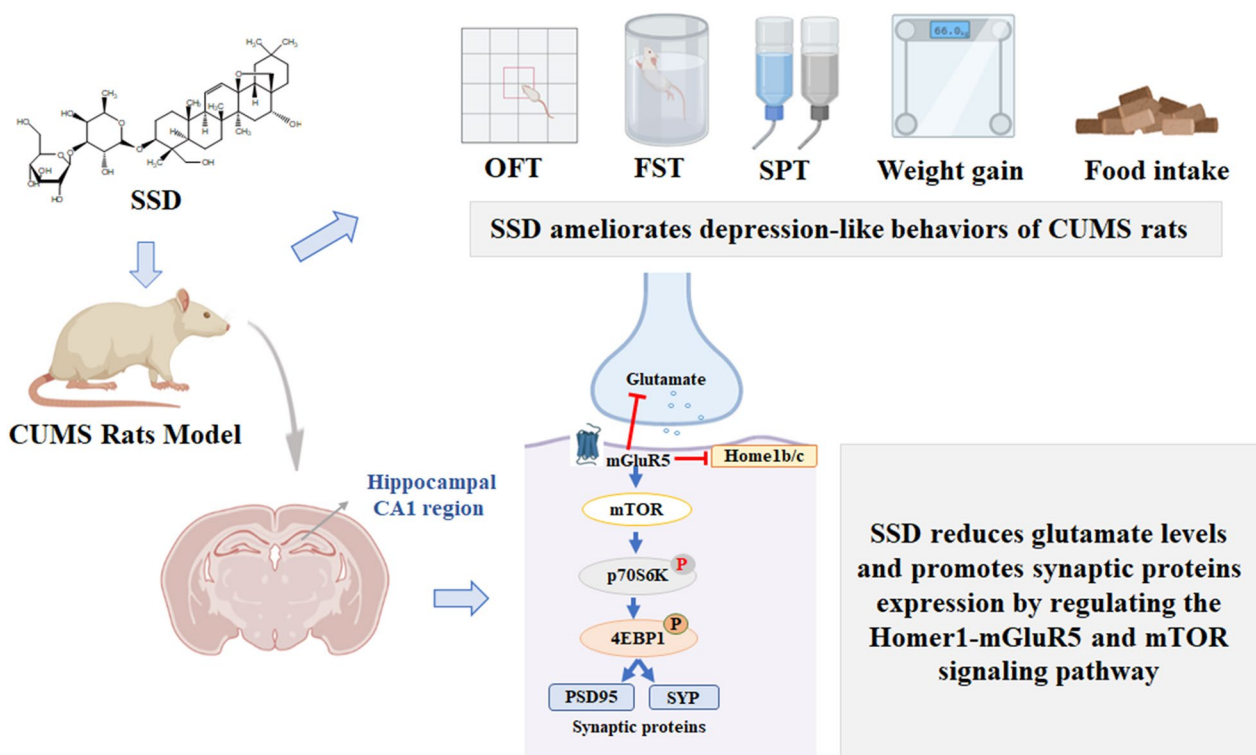


Fig. 8 The potential mechanism of SSD against depression in CUMS rats

who were exposed to the CUMS paradigm but did not receive SSD. Hong-Yan Li et al. as well as Bin Chao et al. found that SSD administration with UCMS (consistent with CUMS)-induced rats could improve depression-like behavior, which is consistent with the results of this study [4, 31].

SSD administration increased glutamate levels in the CUMS rats' CA1 region, according to our current findings. Glutamate is considered to be the main excitatory neurotransmitter in the nervous system, exerting internal as well as external control over the information flow. There is evidence that abnormal glutamate synthesis, metabolism, and reabsorption by neurons in the brain are key components of the depression's pathophysiological mechanism [38]. Furthermore, preclinical investigations have shown that CUMS increases glutamate levels in the extracellular of brain hippocampus region, resulting in glutamate-mediated excitotoxicity [39], which, in turn, leads to synaptic damage [40, 41]. Glutamate exposure also upregulates the expression of mGluR5 in the HT-22 hippocampal neuronal cell line [42]. Researches indicate that the dysregulation of glutamate transmission, primarily via changes in postsynaptic AMPARs, mGluRs and NMDAR, are important in the onset of psychiatric illnesses [43–45]. Thus, regulation of glutamate receptor activity will be a new target for novel antidepressant

drugs [46]. Among these postsynaptic receptors, mGluRs have been shown to be closely related to the pathological mechanism of mood disorders such as depression [47]. For example, antidepressant-like behaviors are induced by conditional knockdown of mGluR5 in gamma-aminobutyric acid- (GABA)ergic neurons, [19], which is congruent with mGluR5^{-/-} mice, an antidepressant phenotype mice [20]. Overall, the results of this study show that mGluR5 is an important mediator of depression. Clinically [48], as well as in preclinical models of depression [49], decreased function of excitatory mGlu5 receptors should reduce the function of the glutamate system, which appears to be enhanced in depression. The results of this experiment are consistent with these studies that showed in depression models, there was a reduction of mGlu5R [50].

Homer1 is a postsynaptic scaffold protein, located in the postsynaptic density [21]. GWAS have revealed that Homer1 plays an important role mood disorders as well as antidepressant treatments [24, 25]. Researches also discovered that in China, rs7713917, the Homer1 gene variant, is a significant correlation to suicide attempts [51]. Homer1 has two distinct isoforms: the short isoform Homer1a and the long isoform Homer1b/c. Homer1b/c, which is constitutively expressed, maintains synaptic transmission and dendritic spine structure

[23]. Homer1b/c has been shown to affect the response to antidepressants [52]. Therefore, in the present study, we concentrated on Homer1b/c. Previous research has found that chronic restraint stress increases Homer1b/c expressions in the hippocampus of rats [53]. Furthermore, the stress of social failure during youth increased the expression mRNA level of Homer1b/c in the dorsal mice's hippocampus [54]. Some antidepressant treatments suppress depression-like behaviors by modulating Homer1 expression [55]. Thus, in this study, we assessed changes in Homer1b/c expression in the CUMS-exposed rats CA1 region.

The mTOR pathway has been shown to be regulated by the increase in Homer1 and the decrease in mGluR5 expression caused by chronic stress. Our findings show that mTOR induces the phosphorylation and activation of p70S6 and 4E-BP1, increasing the expression of synaptic proteins such as PSD-95 and SYP. Studies have also shown that antidepressants function by activating the mTOR pathway [28, 56]. mTOR is a serine/threonine protein kinase with a molecular weight of 289 kDa; it regulates autophagy, protein synthesis and lysosomal biogenesis [57–59]. Several studies have found that decreasing mTOR activation in animal depression models is beneficial. CUMS causes depressive-like behavior in mice and rats, and those changes are associated with decreased phosphorylation of mTOR and its downstream signaling components [60]. This is consistent with our findings. The activities of several receptors, including mGluRs, influence mTOR function [61–63]. Studies have suggested that the activation of mGluR5 subtypes is required for phosphorylation of mTOR in the CA1 area [27]. Once activated, mTORC1 causes the phosphorylation and activation of p70S6K and 4E-BP1, increasing the expression of the synaptic proteins PSD-95 and SYP that drive the formation of new synapses. Our data support these assumptions, as 3 weeks of SSD treatment significantly increased PSD-95 and SYP expression in the CA1 region of rats exposed to the CUMS paradigm. Although we initially confirmed the anti-depression effects and mechanism of SSD on CUMS rats, further researches are needed in vitro. Furthermore, pathway blockers and agonists need to be further validated in experiments. In summary, the findings suggest that chronic stress causes affective changes that can be attenuated by SSD treatment through the modulation of Homer1-mGluR5 and mTOR signaling in the hippocampus.

Conclusion

In this study, we discovered that SSD treatment attenuated the increase in glutamate levels in the hippocampal CA1 region induced by chronic stress, which is

known to lead to synaptic damage and promote the expression of synaptic proteins. Thus, SSD may alleviate depression-like behaviors in CUMS-exposed rats by regulating the Homer1-mGluR5 and mTOR signaling pathways. These findings point to the possibility that SSD could be administered as a natural neuroprotective agent in the prevention of depression.

Abbreviations

4E-BP1	Eukaryotic translation initiation factor 4E-binding protein 1
AMPA	α -Amino-3-hydroxy-5-methyl-4-isoxazolepropionic acid receptor
CUMS	Chronic unpredictable mild stress
DMSO	Dimethyl sulfoxide
GABA	Gamma-aminobutyric acid
GAPDH	Glyceraldehyde-3-phosphate dehydrogenase
FLU	Fluoxetine
FST	Forced swim test
GWAS	Genome-wide association studies
Homer1	Homer protein homolog 1
IHC	Immunohistochemistry
IOD	Integrated optical density
mGluRs	Metabotropic glutamate receptors
mTOR	Mammalian target of rapamycin
mTORC1	MTOR complex 1
NC	Normal control
NMDAR	N-methyl-D-aspartate receptor
OFT	Open field test
RT-qPCR	Real-time quantitative polymerase chain reaction
SPT	Sucrose preference test
PSD-95	Postsynaptic density protein 95
PVDF	Polyvinylidene difluoride
S6K	P70S6 kinase
SSD	Saikosaponin D
SSDH	Saikosaponin D high-dose
SSDL	Saikosaponin D low-dose
SYP	Synapsin I
TBST	Tris-buffered saline containing 0.1% Tween 20

Supplementary Information

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Additional file 1. Describes the Behavioral testing, RT-QPCR, Western blot analysis, Immunohistochemistry experimental procedure, and Statistical analysis in detail.

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Author contributions

CYL, JBC and YYL were responsible for the conception and design of the study and the supervision of the experiments and contributed to revising the manuscript. CYL, JBC, YYL, XMZ, MZ, YMJ, QYM, ZX, ZYZ and XJL performed the experiments. CYL, JBC and JXC analyzed the data. CYL and YYL contributed to the drafting of the manuscript. All authors contributed to manuscript revision. All authors read and approved the final manuscript.

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Availability of data and materials

The data used to support the findings of this study are included in the manuscript.

Declarations

Ethics approval and consent to participate

All animal experiments are approved by the Animal Care and Use Institutional Committee Beijing University of Chinese Medicine abides by animal welfare guidelines to minimize animal suffering and animal use.

Consent for publication

All the authors have agreed that the manuscript published in Chinese Medicine.

Competing interests

The authors declare that they have no known competing financial interest or personal relationships that could have appeared to influence the work reported in this paper.

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References

- Kim J, Farchione T, Potter A, Chen Q, Temple R. Esketamine for Treatment-Resistant Depression - First FDA-approved antidepressant in a new class. *N Engl J Med*. 2019;381(1):1–4.
- Chen C, Luo F, Liu X, Lu L, Xu H, Yang Q, Xue J, Shi L, Li J, Zhang A, et al. NF-kB-regulated exosomal miR-155 promotes the inflammation associated with arsenite carcinogenesis. *Cancer Lett*. 2017;388:21–33.
- Xu L, Su J, Guo L, Wang S, Deng X, Ma S. Modulation of LPA1 receptor-mediated neuronal apoptosis by Saikosaponin-d: A target involved in depression. *Neuropharmacology*. 2019;155:150–61.
- Li HY, Zhao YH, Zeng MJ, Fang F, Li M, Qin TT, Ye LY, Li HW, Qu R, Ma SP. Saikosaponin D relieves unpredictable chronic mild stress induced depressive-like behavior in rats: involvement of HPA axis and hippocampal neurogenesis. *Psychopharmacology*. 2017;234(22):3385–94.
- Su J, Pan YW, Wang SQ, Li XZ, Huang F, Ma SP. Saikosaponin-d attenuated lipopolysaccharide-induced depressive-like behaviors via inhibiting microglia activation and neuroinflammation. *Int Immunopharmacol*. 2020;80: 106181.
- Zhou XM, Liu CY, Liu YY, Ma QY, Zhao X, Jiang YM, Li XJ, Chen JX. Xiaoyaosan Alleviates Hippocampal Glutamate-Induced Toxicity in the CUMS Rats via NR2B and PI3K/Akt Signaling Pathway. *Front Pharmacol*. 2021;12: 586788.
- Wang Y, Fan R, Huang X. Meta-analysis of the clinical effectiveness of traditional Chinese medicine formula Chaihu-Shugan-San in depression. *J Ethnopharmacol*. 2012;141(2):571–7.
- Chang B, Liu Y, Hu J, Tang Z, Qiu Z, Song Z, Jia A, Zhang Y. Bupleurum chinense DC improves CUMS-induced depressive symptoms in rats through upregulation of the cAMP/PKA/CREB signalling pathway. *J Ethnopharmacol*. 2022;289: 115034.
- Li ZY, Guo Z, Liu YM, Liu XM, Chang Q, Liao YH, Pan RL. Neuroprotective effects of total saikosaponins of Bupleurum yinchowense on corticosterone-induced apoptosis in PC12 cells. *J Ethnopharmacol*. 2013;148(3):794–803.
- Willner P, Mitchell PJ. The validity of animal models of predisposition to depression. *Behav Pharmacol*. 2002;13(3):169–88.
- Katz RJ, Roth KA, Schmaltz K. Amphetamine and tranylcypromine in an animal model of depression: pharmacological specificity of the reversal effect. *Neurosci Biobehav Rev*. 1981;5(2):259–64.
- Duman RS, Aghajanian GK. Synaptic dysfunction in depression: potential therapeutic targets. *Science*. 2012;338(6103):68–72.
- Bartsch T, Dohring J, Rohr A, Jansen O, Deuschl G. CA1 neurons in the human hippocampus are critical for autobiographical memory, mental time travel, and autoethic consciousness. *Proc Natl Acad Sci U S A*. 2011;108(42):17562–7.
- Song Q, Fan C, Wang P, Li Y, Yang M, Yu SY. Hippocampal CA1 betaCaMKII mediates neuroinflammatory responses via COX-2/PGE2 signaling pathways in depression. *J Neuroinflammation*. 2018;15(1):338.
- Hu W, Wu J, Ye T, Chen Z, Tao J, Tong L, Ma K, Wen J, Wang H, Huang C. Farnesoid X Receptor-Mediated Cytoplasmic Translocation of CRT2 Disrupts CREB-BDNF Signaling in Hippocampal CA1 and Leads to the Development of Depression-Like Behaviors in Mice. *Int J Neuropsychopharmacol*. 2020;23(10):673–86.
- Zhu Y, Duan X, Cheng X, Cheng X, Li X, Zhang L, Liu P, Su S, Duan JA, Dong TT, et al. Kai-Xin-San, a standardized traditional Chinese medicine formula, up-regulates the expressions of synaptic proteins on hippocampus of chronic mild stress induced depressive rats and primary cultured rat hippocampal neuron. *J Ethnopharmacol*. 2016;193:423–32.
- Pan R, Cai J, Zhan L, Guo Y, Huang RY, Li X, Zhou M, Xu D, Zhan J, Chen H. Buyang Huanwu decoction facilitates neurorehabilitation through an improvement of synaptic plasticity in cerebral ischemic rats. *BMC Complement Altern Med*. 2017;17(1):173.
- Li C, Huang J, Cheng YC, Zhang YW. Traditional Chinese medicine in depression treatment: from molecules to systems. *Front Pharmacol*. 2020;11:586.
- Lee KW, Westin L, Kim J, Chang JC, Oh YS, Amreen B, Gresack J, Flajolet M, Kim D, Aperia A, et al. Alteration by p11 of mGluR5 localization regulates depression-like behaviors. *Mol Psychiatry*. 2015;20(12):1546–56.
- Li X, Need AB, Baez M, Witkin JM. Metabotropic glutamate 5 receptor antagonism is associated with antidepressant-like effects in mice. *J Pharmacol Exp Ther*. 2006;319(1):254–9.
- de Bartolomeis A, Tomassetti C. Calcium-dependent networks in dopamine-glutamate interaction: the role of postsynaptic scaffolding proteins. *Mol Neurobiol*. 2012;46(2):275–96.
- Li MX, Li Q, Sun XJ, Luo C, Li Y, Wang YN, Chen J, Gong CZ, Li YJ, Shi LP, et al. Increased Homer1-mGluR5 mediates chronic stress-induced depressive-like behaviors and glutamatergic dysregulation via activation of PERK-elf2alpha. *Prog Neuropsychopharmacol Biol Psychiatry*. 2019;95: 109682.
- Sala C, Futai K, Yamamoto K, Worley PF, Hayashi Y, Sheng M. Inhibition of dendritic spine morphogenesis and synaptic transmission by activity-inducible protein Homer1a. *J Neurosci*. 2003;23(15):6327–37.
- Ising M, Lucae S, Binder EB, Bettecken T, Uhr M, Ripke S, Kohli MA, Hennings JM, Horstmann S, Kloiber S, et al. A genomewide association study points to multiple loci that predict antidepressant drug treatment outcome in depression. *Arch Gen Psychiatry*. 2009;66(9):966–75.
- Rietschel M, Mattheisen M, Frank J, Treutlein J, Degenhardt F, Breuer R, Steffens M, Mier D, Esslinger C, Walter H, et al. Genome-wide association-, replication-, and neuroimaging study implicates HOMER1 in the etiology of major depression. *Biol Psychiatry*. 2010;68(6):578–85.
- Ignacio ZM, Reus GZ, Arent CO, Abelaira HM, Pitcher MR, Quevedo J. New perspectives on the involvement of mTOR in depression as well as in the action of antidepressant drugs. *Br J Clin Pharmacol*. 2016;82(5):1280–90.
- Hou L, Klann E. Activation of the phosphoinositide 3-kinase-Akt-mammalian target of rapamycin signaling pathway is required for metabotropic glutamate receptor-dependent long-term depression. *J Neurosci*. 2004;24(28):6352–61.
- Duman RS, Li N, Liu RJ, Duric V, Aghajanian G. Signaling pathways underlying the rapid antidepressant actions of ketamine. *Neuropharmacology*. 2012;62(1):35–41.
- Tavares MK, Dos Reis S, Platt N, Heinrich IA, Wolin IAV, Leal RB, Kaster MP, Rodrigues ALS, Freitas AE. Agmatine potentiates neuroprotective effects of subthreshold concentrations of ketamine via mTOR/S6 kinase signaling pathway. *Neurochem Int*. 2018;118:275–85.
- Willner P. Validity, reliability and utility of the chronic mild stress model of depression: a 10-year review and evaluation. *Psychopharmacology*. 1997;134(4):319–29.

31. Chao B, Huang S, Pan J, Zhang Y, Wang Y. Saikosaponin d downregulates microRNA-155 and upregulates FGF2 to improve depression-like behaviors in rats induced by unpredictable chronic mild stress by negatively regulating NF- κ B. *Brain Res Bull.* 2020;157:69–76.
32. Ma Q, Li X, Yan Z, Jiao H, Wang T, Hou Y, Jiang Y, Liu Y, Chen J. Xiaoyaosan Ameliorates Chronic Immobilization Stress-Induced Depression-Like Behaviors and Anorexia in Rats: The Role of the Nesfatin-1-Oxytocin-Proopiomelanocortin Neural Pathway in the Hypothalamus. *Front Psychiatry.* 2019;10:910.
33. Liu C, Zhu R, Liu H, Li L, Chen B, Jia Q, Wang L, Ma R, Tian S, Wang M, et al. Aqueous Extract of Mori Folium Exerts Bone Protective Effect Through Regulation of Calcium and Redox Homeostasis via PTH/VDR/CaBP and AGEs/RAGE/Nox4/NF- κ B Signaling in Diabetic Rats. *Front Pharmacol.* 2018;9:1239.
34. Bersani G, Liberati D, Rasa A, Polli E, Ciuffa M, Limpido L, Russo D, Marconi D, Pacitti F. Premorbid sleep, appetite, energy, and cognitive circadian profile in patients with depressive disorders. *Eur Psychiatry.* 2010;25(8):461–4.
35. Willner P. The chronic mild stress (CMS) model of depression: History, evaluation and usage. *Neurobiol Stress.* 2017;6:78–93.
36. Belovicova K, Bogi E, Csatislova K, Dubovicky M. Animal tests for anxiety-like and depression-like behavior in rats. *Interdiscip Toxicol.* 2017;10(1):40–3.
37. Simmons WK, Burrows K, Avery JA, Kerr KL, Bodurka J, Savage CR, Drevets WC. Depression-Related Increases and Decreases in Appetite: Dissociable Patterns of Aberrant Activity in Reward and Interoceptive Neurocircuitry. *Am J Psychiatry.* 2016;173(4):418–28.
38. Murrrough JW, Abdallah CG, Mathew SJ. Targeting glutamate signaling in depression: progress and prospects. *Nat Rev Drug Discov.* 2017;16(7):472–86.
39. Autry AE, Adachi M, Nosyreva E, Na ES, Los MF, Cheng PF, Kavalali ET, Monteggia LM. NMDA receptor blockade at rest triggers rapid behavioural antidepressant responses. *Nature.* 2011;475(7354):91–5.
40. Wipfel C, Maurer J, Fortsch C, Hupp S, Bohl A, Ma J, Mitchell TJ, Bunkowski S, Bruck W, Nau R, et al. Bacterial cytolysin during meningitis disrupts the regulation of glutamate in the brain, leading to synaptic damage. *PLoS Pathog.* 2013;9(6): e1003380.
41. Sapolsky RM. The possibility of neurotoxicity in the hippocampus in major depression: a primer on neuron death. *Biol Psychiatry.* 2000;48(8):755–65.
42. Sagara Y, Schubert D. The activation of metabotropic glutamate receptors protects nerve cells from oxidative stress. *J Neurosci.* 1998;18(17):6662–71.
43. Kendell SF, Krystal JH, Sanacora G. GABA and glutamate systems as therapeutic targets in depression and mood disorders. *Expert Opin Ther Targets.* 2005;9(1):153–68.
44. Mathews DC, Henter ID, Zarate CA. Targeting the glutamatergic system to treat major depressive disorder: rationale and progress to date. *Drugs.* 2012;72(10):1313–33.
45. Tronson NC, Guzman YF, Guedea AL, Huh KH, Gao C, Schwarz MK, Radulovic J. Metabotropic glutamate receptor 5/Homer interactions underlie stress effects on fear. *Biol Psychiatry.* 2010;68(11):1007–15.
46. Popoli M, Yan Z, McEwen BS, Sanacora G. The stressed synapse: the impact of stress and glucocorticoids on glutamate transmission. *Nat Rev Neurosci.* 2011;13(1):22–37.
47. Chaki S, Fukumoto K. mGlu receptors as potential targets for novel antidepressants. *Curr Opin Pharmacol.* 2018;38:24–30.
48. Deschwanden A, Karolewicz B, Feyissa AM, Treyer V, Ametamey SM, Johayem A, Burger C, Auberson YP, Sovago J, Stockmeier CA, et al. Reduced metabotropic glutamate receptor 5 density in major depression determined by [(11)C]ABP688 PET and postmortem study. *Am J Psychiatry.* 2011;168(7):727–34.
49. Smialowska M, Szewczyk B, Branski P, Wieronska JM, Palucha A, Bajkowska M, Pilc A. Effect of chronic imipramine or electroconvulsive shock on the expression of mGluR1a and mGluR5a immunoreactivity in rat brain hippocampus. *Neuropharmacology.* 2002;42(8):1016–23.
50. Chaki S, Ago Y, Palucha-Paniewiera A, Matriciano F, Pilc A. mGlu2/3 and mGlu5 receptors: potential targets for novel antidepressants. *Neuropharmacology.* 2013;66:40–52.
51. Rao S, Siu CO, Shi M, Zhang J, Lam MHB, Yu M, Wing YK, Wayne MMY. Associations of Homer Scaffolding Protein 1 gene and psychological correlates with suicide attempts in Chinese: A pilot study of multifactorial risk model. *Gene.* 2018;679:382–8.
52. Benedetti F, Poletti S, Locatelli C, Mazza E, Lorenzi C, Vitali A, Riberto M, Brioschi S, Vai B, Bollettini I, et al. A Homer 1 gene variant influences brain structure and function, lithium effects on white matter, and antidepressant response in bipolar disorder: A multimodal genetic imaging study. *Prog Neuropsychopharmacol Biol Psychiatry.* 2018;81:88–95.
53. Kaastrup Muller H, Orłowski D, Reidies Bjarkam C, Wegener G, Elfving B. Potential roles for Homer1 and Spinophilin in the preventive effect of electroconvulsive seizures on stress-induced CA3c dendritic retraction in the hippocampus. *Eur Neuropsychopharmacol.* 2015;25(8):1324–31.
54. Wagner KV, Hartmann J, Labermaier C, Hausl AS, Zhao G, Harbich D, Schmid B, Wang XD, Santarelli S, Kohl C, et al. Homer1/mGluR5 activity moderates vulnerability to chronic social stress. *Neuropsychopharmacology.* 2015;40(5):1222–33.
55. Serchov T, Clement HW, Schwarz MK, Iasevoli F, Tosh DK, Idzko M, Jacobson KA, de Bartolomeis A, Normann C, Biber K, et al. Increased Signaling via Adenosine A1 Receptors, Sleep Deprivation, Imipramine, and Ketamine Inhibit Depressive-like Behavior via Induction of Homer1a. *Neuron.* 2015;87(3):549–62.
56. Li N, Lee B, Liu RJ, Banasr M, Dwyer JM, Iwata M, Li XY, Aghajanian G, Duman RS. mTOR-dependent synapse formation underlies the rapid antidepressant effects of NMDA antagonists. *Science.* 2010;329(5994):959–64.
57. Dazert E, Hall MN. mTOR signaling in disease. *Curr Opin Cell Biol.* 2011;23(6):744–55.
58. Laplante M, Sabatini DM. mTOR signaling in growth control and disease. *Cell.* 2012;149(2):274–93.
59. Bockaert J, Marin P. mTOR in Brain Physiology and Pathologies. *Physiol Rev.* 2015;95(4):1157–87.
60. Zhong P, Wang W, Pan B, Liu X, Zhang Z, Long JZ, Zhang HT, Cravatt BF, Liu QS. Monoacylglycerol lipase inhibition blocks chronic stress-induced depressive-like behaviors via activation of mTOR signaling. *Neuropsychopharmacology.* 2014;39(7):1763–76.
61. Dadalko OI, Siuta M, Poe A, Erreger K, Matthies HJ, Niswender K, Galli A. mTORC2/riCTOR signaling disrupts dopamine-dependent behaviors via defects in striatal dopamine neurotransmission. *J Neurosci.* 2015;35(23):8843–54.
62. Hsu WL, Chung HW, Wu CY, Wu HI, Lee YT, Chen EC, Fang W, Chang YC. Glutamate Stimulates Local Protein Synthesis in the Axons of Rat Cortical Neurons by Activating alpha-Amino-3-hydroxy-5-methyl-4-isoxazolepropionic Acid (AMPA) Receptors and Metabotropic Glutamate Receptors. *J Biol Chem.* 2015;290(34):20748–60.
63. Koskimaki J, Matsui N, Umemori J, Rantamaki T, Castren E. Nimodipine activates TrkB neurotrophin receptors and induces neuroplastic and neuroprotective signaling events in the mouse hippocampus and prefrontal cortex. *Cell Mol Neurobiol.* 2015;35(2):189–96.

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