

REVIEW

Open Access



Implications of TDP-43 in non-neuronal systems

Hao Ke¹, Kang Liu², Baowei Jiao^{3,4*} and Limin Zhao^{1*}

Abstract

TAR DNA-binding protein 43 (TDP-43) is a versatile RNA/DNA-binding protein with multifaceted processes. While TDP-43 has been extensively studied in the context of degenerative diseases, recent evidence has also highlighted its crucial involvement in diverse life processes beyond neurodegeneration. Here, we mainly reviewed the function of TDP-43 in non-neurodegenerative physiological and pathological processes, including spermatogenesis, embryonic development, mammary gland development, tumor formation, and viral infection, highlighting its importance as a key regulatory factor for the maintenance of normal functions throughout life. TDP-43 exhibits diverse and sometimes opposite functionality across different cell types through various mechanisms, and its roles can shift at distinct stages within the same biological system. Consequently, TDP-43 operates in both a context-dependent and a stage-specific manner in response to a variety of internal and external stimuli.

Keywords Embryonic development, Fat metabolism, Mammary gland development, Spermatogenesis, Stem cell, TDP-43, Tumor, Viral infection

Introduction

Encoded by the *TARDBP* gene, TDP-43 is highly conserved across various species [1]. The protein was initially recognized as a transcription repressor (with a size of 43 kD, hence its name), exhibiting affinity for the TAR DNA sequence of human immunodeficiency virus type 1 (HIV-1) [2]. Subsequent study identified TDP-43 as a constituent of ubiquitinated insoluble aggregates found in the

brains of individuals with frontotemporal lobar dementia (FTLD) [3], with more recent research establishing strong associations between TDP-43 and a range of neurodegenerative disorders.

TDP-43 is a multifunctional protein that operates within a precise regulatory framework. Heterozygous knockout of TDP-43 in mice does not result in successful TDP-43 knockdown due to its own negative feedback regulation, while homozygous knockout of TDP-43 leads to death in both embryonic and adult stages [4–6]. This precise regulation highlights the importance of TDP-43 in fundamental life processes. In addition to its established functions in neurodegenerative diseases, TDP-43 is reported to participate in diverse developmental stages [4, 7–9]. Therefore, in this review, we discuss the important functions of TDP-43 in different tissues and developmental stages in non-neurodegenerative systems. Notably, current research suggests that TDP-43 plays a context-dependent and stage-specific role in different environments and developmental stages. Furthermore, TDP-43 likely serves as an important stress sensor,

*Correspondence:

Baowei Jiao
jiaobaowei@mail.kiz.ac.cn
Limin Zhao
zhaolimin@ncu.edu.cn

¹ Human Aging Research Institute (HARI) and School of Life Science, Nanchang University, and Jiangxi Key Laboratory of Human Aging, Nanchang 330031, China

² Ganzhou People's Hospital, Ganzhou 341000, China

³ National Key Laboratory of Genetic Evolution & Animal Models, Kunming Institute of Zoology, Chinese Academy of Sciences, Kunming, Yunnan 650201, China

⁴ KIZ-CUHK Joint Laboratory of Bioresources and Molecular Research in Common Diseases, Kunming Institute of Zoology, Chinese Academy of Sciences, Kunming 650201, China



© The Author(s) 2023. **Open Access** This article is licensed under a Creative Commons Attribution 4.0 International License, which permits use, sharing, adaptation, distribution and reproduction in any medium or format, as long as you give appropriate credit to the original author(s) and the source, provide a link to the Creative Commons licence, and indicate if changes were made. The images or other third party material in this article are included in the article's Creative Commons licence, unless indicated otherwise in a credit line to the material. If material is not included in the article's Creative Commons licence and your intended use is not permitted by statutory regulation or exceeds the permitted use, you will need to obtain permission directly from the copyright holder. To view a copy of this licence, visit <http://creativecommons.org/licenses/by/4.0/>. The Creative Commons Public Domain Dedication waiver (<http://creativecommons.org/publicdomain/zero/1.0/>) applies to the data made available in this article, unless otherwise stated in a credit line to the data.

responding to intracellular and extracellular stimuli to maintain homeostasis in life processes and activities.

Structure and function of TDP-43

As part of the heterogeneous nuclear ribonucleoprotein (hnRNP) family, TDP-43 is primarily located in the nucleus, although it can also exist within the cytoplasm and mitochondria [10–12]. The structural composition of TDP-43 includes an N-terminal region with a nuclear localization signal (NLS), two RNA recognition motifs (RRM1 and RRM2), a nuclear export signal (NES) located within RRM2, a C-terminal region featuring a glutamine/asparagine-rich (Q/N) domain and a glycine rich region, as well as five putative mitochondrial localization signals (M1–M5) (Fig. 1) [13–17]. The N terminal domain is thermodynamically stable and well-folded and undergoes reversible oligomerization [18, 19]. Several reports have proposed that N-terminal domain-induced dimerization of TDP-43 is necessary for its physiological functions, such as RNA splicing [16, 20]. While the NLS and NES are both implicated in the nucleocytoplasmic shuttling of TDP-43, with the NLS additionally facilitating the nuclear transport of TDP-43 [19, 21], the precise functionalities of the NES remain controversial [15].

TDP-43 participates in many functions (Fig. 2) and can act as a DNA-binding protein. TDP-43 was initially studied as a transcriptional inhibitor that binds to the TAR regulatory element of HIV-1, thereby influencing transcription factor assembly [2]. Subsequent investigations revealed that TDP-43 binds to the TGTGTG domains in the promoter region of the mouse *SP-10* gene, leading to the inhibition of gene transcription and impacting sperm formation [7]. Additional studies have also demonstrated that TDP-43 is involved in DNA damage, DNA replication, and genome stability [22–24].

As a splicing factor, TDP-43 can affect alternative splicing of various genes, such as apolipoprotein A-II [25], *RXRG* [26], *SC35* [27], *SMN* [28], *ETFI* [26], *BRCA1* [26], schizophrenia-associated TNIK gene [29], *PAR3/NUMB*

[30], and cancer stem cell marker *CD44* [31]. TDP-43 also exerts influence on various other RNA processes, including RNA transport and stability [32], RNA translation [33], and microRNA (miRNA) biogenesis [34]. In conjunction with fragile X syndrome protein (FMRP), TDP-43 can cooperatively suppress the translation initiation of *Map1b*, *Rac1*, and *GluR1* mRNAs [33]. In addition, TDP-43 forms associations with 126 proteins to jointly regulate mRNA transport and stability in HEK-293 cells [35], and its overexpression can induce profound RNA destabilization [36]. Interestingly, TDP-43 appears to play a dual role in RNA stability, both promoting mRNA instability, as observed for CDK6 mRNA decay [37, 38] and tau mRNA instability [39], and maintaining mRNA stability, as observed for *G3BP1* [40], *Add2* [41], *RPTOR/RAPTOR* [42], *Btn1a1*, and *Xdh* [8]. Moreover, TDP-43 not only regulates targeted mRNA stability, but also stabilizes mitochondrial transfer RNA (mt-tRNA) in human mitochondria [43]. Thus, these studies collectively demonstrate the involvement of TDP-43 in various RNA metabolic processes.

TDP-43 in degenerative neurological diseases

TDP-43 is a well-recognized neurodegenerative disease-related protein. In 2006, ubiquitinated and hyperphosphorylated TDP-43 was identified in both amyotrophic lateral sclerosis (ALS) and FTLN [3, 44]. Subsequently, abnormal aggregation of TDP-43 has been implicated in various other neurodegenerative diseases. These pathological inclusions of TDP-43, known collectively as “TDP-43 proteinopathy”, are characterized by the accumulation of hyperphosphorylated, ubiquitinated, and cleaved TDP-43 in the cytoplasm, with a simultaneous reduction of TDP-43 levels in the nucleus. Nevertheless, the precise mechanisms underlying TDP-43 proteinopathy in neurodegenerative diseases remain to be fully elucidated.

At present, various theories exist regarding the cytotoxicity caused by the mislocalization of TDP-43 from

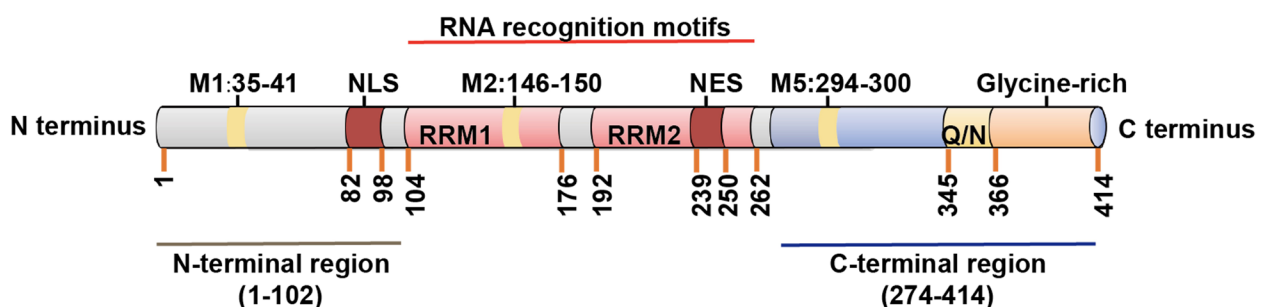


Fig. 1 Structure of TDP-43 protein. Numbers represent amino acid lengths of TDP-43. NLS: nuclear localization signal, RRM: RNA recognition motifs, NES: nuclear export signal, Q/N: glutamine/asparagine-rich domain, M1–M5: five putative mitochondrial localization signals

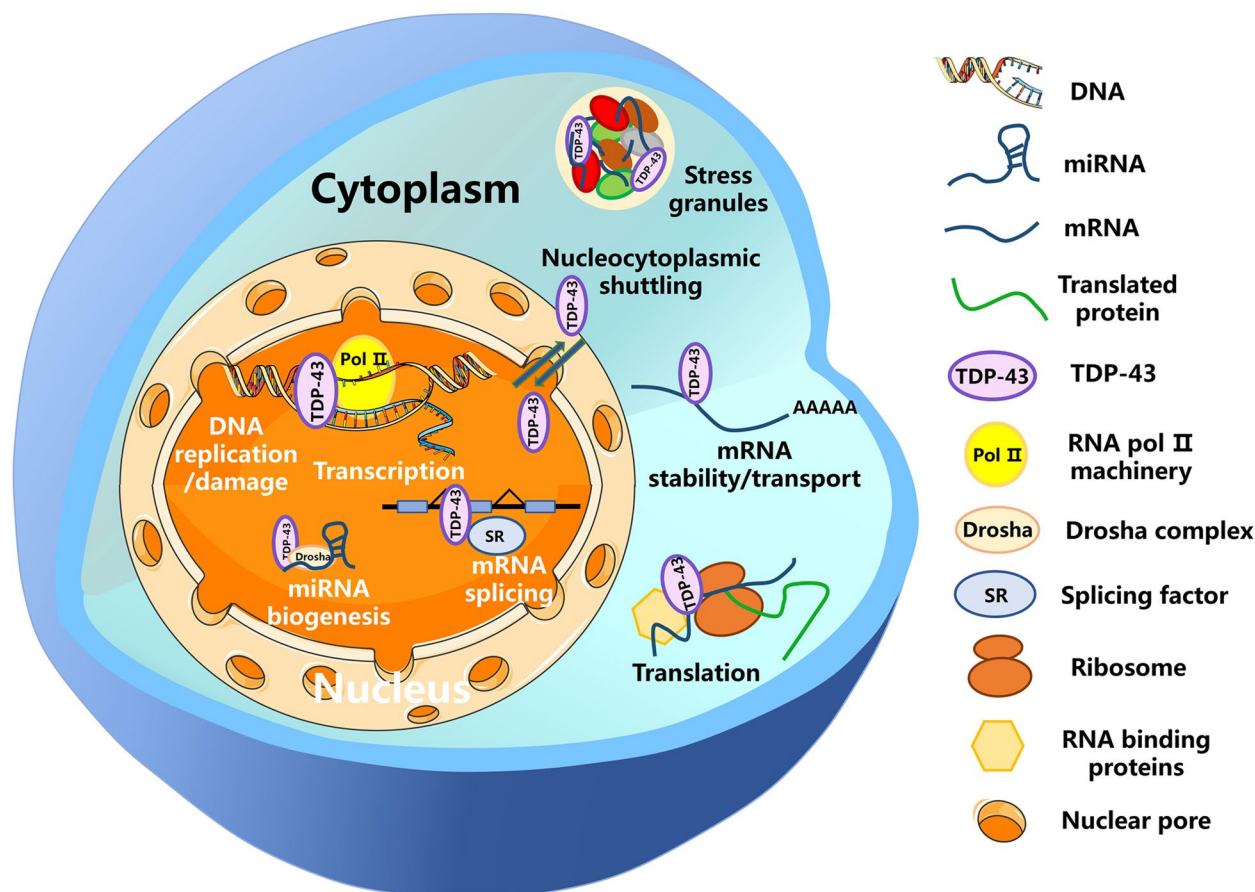


Fig. 2 Functions of TDP-43 protein. TDP-43 functions as a nucleocytoplasmic shuttling protein via nuclear pores, which can regulate cellular functions at multiple levels. At the DNA level, TDP-43 is involved in gene transcription, DNA damage, and DNA replication. At the RNA level, TDP-43 is involved in RNA splicing, RNA transport, and RNA stability. TDP-43 is also involved in miRNA biogenesis, stress granules, and various other cellular processes

the nucleus to the cytoplasm (Fig. 3). The first theory postulates that neuronal exposure to specific stressors triggers the cytoplasmic mislocalization of TDP-43, leading to the formation of phosphorylated pre-inclusion bodies within the cytoplasm, thereby sequestering free TDP-43 protein and depleting normal nuclear TDP-43 (Fig. 3A, B). In response, the nucleus initiates a compensatory mechanism to generate more TDP-43 protein to restore its normal nuclear function (Fig. 3C). However, the increased production of TDP-43 exacerbates its accumulation in the cytoplasm, ultimately resulting in cell death (Fig. 3E). An alternative hypothesis posits that the formation of pre-inclusion bodies by TDP-43 in the cytoplasm triggers an automatic regulation response in the cytoplasm, leading to a reduction in TDP-43 protein synthesis (Fig. 3D). Given the essential role of TDP-43 in neuronal cells, the down-regulation of its protein synthesis can also lead to normal nuclear TDP-43 deprivation and subsequent neuronal death [45, 46] (Fig. 3E).

TDP-43 in non-neurodegenerative diseases

TDP-43 in cancer

The involvement of TDP-43 in cancer was identified as early as two decades ago, with notable associations found between common variants near *TARDBP* and *EGR2* and the incidence of Ewing sarcoma [47]. In more recent years, research has expanded our understanding of the role of TDP-43 in the progression of different cancers, including breast cancer, cervical cancer, lung cancer, hepatocellular carcinoma, glioblastoma, and melanoma.

TDP-43 acts as an oncogenic factor to promote tumor progression

In the context of breast cancer, multiple reports, including our own, have suggested that TDP-43 may act as an oncogenic factor to promote tumor progression. Notably, Fang et al. found that curcumin, a dietary pigment with known anticancer activities, can significantly inhibit TDP-43 expression in the breast cancer cell line MCF7 [48]. In our previous study, we observed elevated levels of

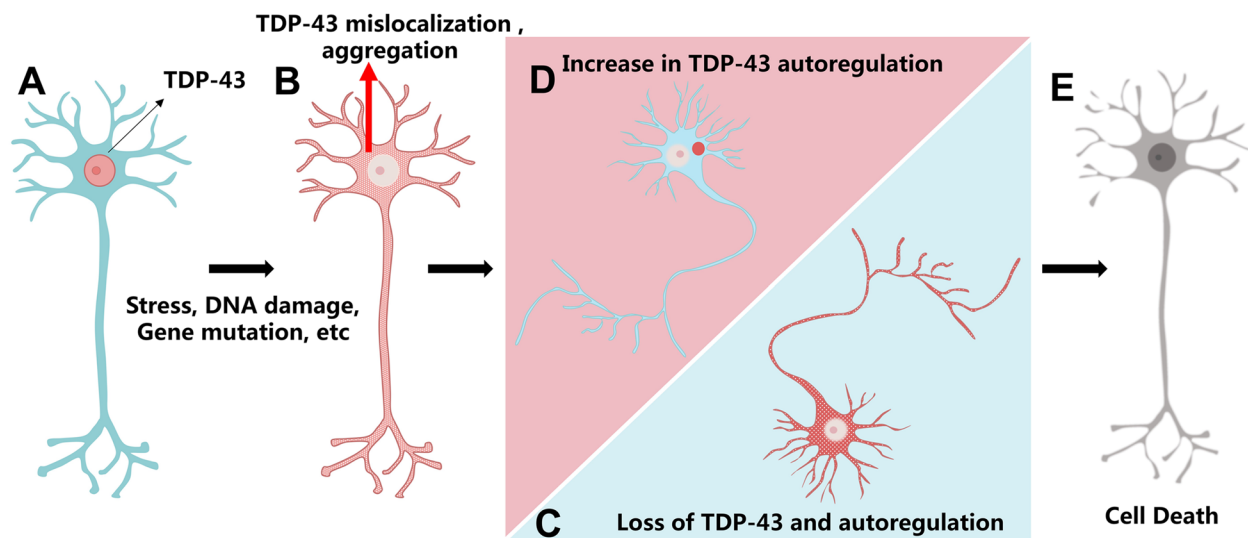


Fig. 3 Models of TDP-43 toxicity. **A** Under normal conditions, TDP-43 expression is under strict regulation and is predominantly located in the nucleus. **B** When subjected to stress, such as DNA damage, the TDP-43 protein moves from the nucleus to cytoplasm, where it forms phosphorylated pre-inclusion bodies. **C** This leads to the loss of normal nuclear TDP-43. If TDP-43 autoregulation occurs within the nucleus, loss of TDP-43 stimulates the cell to produce more TDP-43 protein, thereby aggravating the accumulation of TDP-43 in the cytoplasm. **D** If TDP-43 autoregulation occurs within the cytoplasm, the increase in cytoplasmic TDP-43 can result in an increase in TDP-43 autoregulation and a decrease in the synthesis of new TDP-43 protein. **E** Regardless of whether TDP-43 autoregulation occurs in the nucleus or cytoplasm, it ultimately leads to neuronal death

TDP-43 in triple-negative breast cancer (TNBC), which showed a significant correlation with poor prognosis. Furthermore, we found TDP-43 knockdown led to a significant reduction in tumor progression, including proliferation and metastasis, accompanied by extensive changes in splicing events [9]. In subsequent studies, we demonstrated that TDP-43 can directly bind to the pre-mRNA of CD44, thereby regulating its alternative splicing [31]. Given that CD44 is a well-known marker in breast cancer stem cells (BCSCs), the influence of TDP-43 on CD44 alternative splicing may accelerate cancer progression.

In relation to lung cancer, TDP-43 has been found to enhance the metastasis of non-small cell lung cancer cells (NSCLC) through its regulation of *MALAT1* expression [49]. TDP-43 also promotes the growth and metastasis of NSCLC as a downstream effector of the long noncoding RNA (lncRNA) *MIAT* [50]. In advanced NSCLC patients carrying sensitized epidermal growth factor receptor (EGFR) mutations, the use of EGFR-tyrosine kinase inhibitors (TKIs) in combination with targeted therapy drugs is considered a viable treatment option, but can result in the eventual development of EGFR-TKI resistance in patients. Interestingly, the lncRNA *LCETRL3*, which is located at 4q12 and is associated with resistance to EGFR-TKI [51], can partially diminish the effectiveness of EGFR-TKIs by stabilizing TDP-43 [52], implying

potential involvement of TDP-43 in the development of resistance to EGFR-TKIs.

Park et al. found that TDP-43 can regulate glycolysis by modulating the phosphofructokinase isoform through miRNA 520, with TDP-43 knockdown leading to impaired glucose metabolism and cell proliferation in multiple HCC cell lines [53]. Guo et al. also found that TDP-43 can induce epithelial-mesenchymal transition and promote HCC metastasis by stimulating the Wnt/ β -catenin signaling pathway [54]. Liu et al. further reported that TDP-43 can suppress apoptosis in HCC by up-regulating the lipid metabolism modulator ABHD2 [55].

TDP-43 also plays an important role in various other cancers. Similar to its effects in HCC, TDP-43 promotes the proliferation and migration of melanoma cells, potentially through modulation of glucose metabolism [56]. The TDP-43-HDAC6 signaling axis in glioblastoma multiforme (GBM) acts as a stress-responsive pathway, driving GBM progression and activating autophagy to promote cell survival under nutrient deprivation [57]. Furthermore, using the human bone osteosarcoma (U2OS) cell line, Ayala et al. revealed that loss of TDP-43 can affect nuclear membrane stability and increases apoptosis [37].

TDP-43 plays dual roles in tumor progression

In addition to promoting cancer progression, TDP-43 can also function as a tumor suppressor. For example,

Kim et al. found that a high TDP-43 expression is necessary during TRIM16-induced cancer cell death, with both TRIM16 and TDP-43 serving as good prognostic markers in neuroblastoma and breast cancer [58]. Zaman et al. reported the higher *TARDBP* microarray values are associated with a significant increase in overall survival in pediatric neuroblastoma [59]. Lee et al. provided evidence that elevated levels of TDP-43 in HeLa cells can induce G2/M arrest and cell death via a p53-dependent mechanism [60].

TDP-43 also affects diverse functions of downstream genes. In HeLa cells, TDP-43 inhibits CDK6 by recruiting UG-rich *Cdk6* transcripts [37], whereas in CHO-K1 cells, it activates CDK6 expression [38]. Additionally, TDP-43 exhibits a dual role in the complex regulation of cancer-related miRNAs, acting as a promoter of cancer progression through the regulation of miR-423-3p, while also exerting inhibitory effects on cancer progression through the regulation of miR-500a-3p [61].

These studies underscore the heterogeneity of cancer and highlight the intricate regulatory role of TDP-43 in cancer progression. Although the mechanisms by which TDP-43 regulates cancer are complex and not yet fully understood, they likely depend on the specific cellular context and cancer type.

Potential correlation of TDP-43 between cancer and neurodegenerative diseases

Accumulating evidence suggests the existence of an inverse comorbidity phenomenon between oncological and neurodegenerative conditions. In 2014, Ibanez et al. conducted a transcriptomic meta-analysis of several neurodegenerative diseases and three cancers (lung, prostate, and colorectal cancer). They discovered a significant overlap between up-regulated genes in neurodegenerative diseases and down-regulated genes in cancer, as well as down-regulated genes in neurodegenerative diseases and up-regulated genes in cancer [62]. Subsequent studies have revealed that many genes, proteins, and signaling pathways regulated in both cancer and neurodegenerative diseases display contrasting patterns. Notably, the well-known tumor suppressor p53 is up-regulated in AD, PD, and Huntington's disease (HD), but down-regulated in many cancers [63–65]. Given the multifaceted nature of TDP-43, it is plausible to consider the interconnected roles it plays in both cancer and neurodegenerative diseases.

Furthermore, the intricate mechanisms underlying the effects of TDP-43 in neurodegenerative diseases can be elucidated from various perspectives, including point mutations within the genome, multiple splicing mutants, and phosphorylation proteins. However, it remains to be investigated whether TDP-43 mutations and truncations

exist and hold significant roles in cancer. Additionally, given its role as a splicing factor, the phosphorylation status of TDP-43 directly impacts its splicing function, prompting inquiry into the presence of abnormal phosphorylation of TDP-43 in tumor cells. While exogenous overexpression of TDP-43 can lead to elevated expression of the 35 kD isoform, which may exert cytotoxic effects on cancer cells and induce cell death [66], further in-depth studies are necessary to comprehensively understand the precise function of TDP-43 in tumorigenesis.

TDP-43 in development

TDP-43 is expressed in almost all cell types and is highly conserved among different species. Increasing evidence confirms its crucial regulatory role in various developmental processes, including embryonic development, body fat metabolism, mammary gland development, the reproductive system and so on.

TDP-43 in embryonic development

Various studies utilizing knockout mice have provided compelling evidence for the crucial roles of TDP-43 in embryonic development. Researchers have previously constructed TDP-43 knockout mice by targeting the translation initiation site of exon 2 of *Tardbp* mRNA [4, 67], with such models demonstrating that homozygous deletion of TDP-43 can lead to peri-implantation lethality. Notably, despite morphological normality *in vitro*, blastocysts with homozygous *Tardbp* deletion exhibit impaired outgrowth in the inner cell mass. Conversely, mice with heterozygous TDP-43 deletion display normal development and fertility, with no noticeable abnormalities up to 14 months of age and no changes in TDP-43 protein expression compared to wild-type mice [4]. Sephton et al. [5] also reported that while homozygous *Tardbp* knockout mice died between ED 3.5 and 8.5, heterozygous *Tardbp* mice showed no differences in TDP-43 expression or phenotypic changes, including body weight, growth rate, appearance, and fertility, and further displayed no gross tissue abnormalities up to the age of 6 months compared to control littermates [5]. In another study, Chiang et al. inserted loxp sites on both sides of the third exon of *Tardbp* mRNA and produced a nonfunctional truncated TDP-43 variant in mice after crossing with the CAG-Cre transgenic mouse line [6, 68]. In accordance with the above studies, their mouse model yielded fertile heterozygous knockout mice with normal TDP-43 expression but failed to produce viable homozygous TDP-43 knockout mice due to embryonic death at around day 7.5 [6].

Collectively, these different mouse models demonstrate that homozygous loss of TDP-43 can result in embryo death during blastocyst implantation, confirming the

necessity of TDP-43 expression in embryonic development. In addition, protein levels in heterozygous *Tardbp* knockout mice remained stable across these models, indicating that TDP-43 possesses an inherent regulatory mechanism to mitigate the adverse effects of TDP-43 deletion.

Regarding the mechanism by which TDP-43 safeguards embryonic development, a recent study conducted in mouse embryonic stem cells (mESCs) discovered that TDP-43 protects the embryonic genome by interacting with L1 open reading frame 1 protein (L1 ORF1p) [46].

TDP-43 in fat metabolism

In addition to its vital role in embryonic development, the potential functions of TDP-43 in adult animals have been increasingly investigated. To overcome the issue of embryonic death caused by TDP-43 deletion, researchers have employed conditional knockout mice to study the physiological function of TDP-43. For example, Chiang et al. crossed floxed *Tardbp* mice with *Rosa26-ErCre* mice (which express Cre protein upon tamoxifen induction) to generate inducible *Tardbp* knockout mice. Upon tamoxifen induction, these conditional homozygous *Tardbp* knockout mice unexpectedly died by day 9, characterized by substantial fat loss and increased fatty acid consumption. Mechanistically, deletion of TDP-43 down-regulated the expression of obesity-associated gene *Tbc1d1*, leading to alterations in body fat metabolism [6].

Accumulating evidence supports the involvement of TDP-43 in fat metabolism. Stallings et al. demonstrated that TDP-43 regulates fat homeostasis and glucose metabolism [69]. Coughlan et al. showed that high-fat jelly diets in TDP-43^{A315T} mutant mice (ALS model) can restore bioenergetic balance and extend lifespan [70]. Egawa et al. revealed that TDP-43 can reduce the expression of sterol regulatory element-binding protein 2 (SREBP2), and thus regulate cholesterol biosynthesis [71]. Li et al. demonstrated that TDP-43, acting as a transcription suppressor, alleviates the inhibition of the downstream target gene *Cyp8b1* by binding with the lncRNA lncLSTR, thus regulating systemic lipid metabolism in mice [72]. Studies have also identified the involvement of TDP-43 in obesity pathogenesis [73]. In our previous work, we revealed TDP-43 as a key regulator of milk fat metabolism. Notably, through targeted TDP-43 knockout in the mammary gland from middle pregnancy to the lactation stage, we found that TDP-43 loss leads to abnormal milk fat metabolism, resulting in lactation failure and death of infant mice due to inadequate nutrition [8].

Lipids serve as essential components of biological membranes, not only providing efficient energy storage for organisms, but also acting as signaling molecules/messengers in diverse developmental pathways [74], with

the regulation of fat metabolism by TDP-43 further highlighting its importance during development.

TDP-43 in the reproductive system

Precise regulation of cell type-specific gene transcription is crucial for spermatogenesis, where a multitude of testis-related genes are activated in a programmed spatiotemporal order. For example, *Acrv1*, which codes for the sperm acrosomal protein SP-10, is strictly expressed at the transcriptional level in the testes and round spermatids. Evidence has shown that the proximal promoter of *Acrv1* is sufficient to maintain round spermatid-specific expression and acts as an insulator, preventing ectopic expression of *Acrv1* in somatic cells [75]. TDP-43 was first defined as a putative regulator that binds to the *Acrv1* promoter via two GTGTGT-motifs, eventually inhibiting premature *Acrv1* expression during spermatogenesis [76]. TDP-43 was subsequently identified as a transcriptional repressor in spermatocytes, suppressing *Acrv1* gene transcription in a histone deacetylase-independent manner [77]. In somatic cells, TDP-43 can act as an insulator protein to prevent *Acrv1* ectopic expression [78]. These findings demonstrate that TDP-43 can maintain spermatogenesis through precise and diverse regulatory mechanisms.

Subsequent investigations have revealed that TDP-43 is expressed in both Sertoli cells and germ cells, indicating its potential adoption of multiple conformational states during different stages of spermatogenesis. Studies have shown that TDP-43 expression initiates in type B/intermediate spermatogonia, reaches its peak in preleptotene spermatocytes, disappears in leptotene and zygotene spermatocytes, reappears in pachytene spermatocytes and early round spermatids, and subsequently decreases in later spermatids [79]. The varied expression and localization patterns of TDP-43 suggest that it plays a multifaceted role in spermatogenesis. Indeed, deficiency in TDP-43 is linked with impaired spermatogenesis and male infertility, as observed in the germ cells of fertile and subfertile men [80]. Further studies involving TDP-43 knockout in Sertoli cells and male germ cells have confirmed its crucial role in the male reproductive system [81, 82]. Campbell et al. demonstrated that TDP-43 loss in mouse spermatogonia can initiate meiotic failure, resulting in fewer and more morphologically abnormal sperm accompanied by severely reduced fertility [82]. Zomer et al. also observed that TDP-43 loss in mouse Sertoli cells can induce spermatogenesis failure and male subfertility [81]. These findings collectively establish the importance of TDP-43 during spermatogenesis, suggesting that TDP-43 could potentially serve as an essential indicator of male factor infertility.

TDP-43 in mammary gland development

Lactation, a highly distinctive feature of mammals, provides offspring with sufficient nutrition to ensure their survival. In earlier research, we found that TDP-43 is required for mammary gland development [8, 83].

In our previous studies focused on identifying key post-transcriptional regulatory genes during lactation and exploring the impact of positive selection on species divergence, we conducted an evolutionary analysis of RNA-binding proteins (RBPs) across 15 mammalian species. Our analysis revealed that TDP-43 underwent positive selection within the mammalian lineage, after applying false discovery rate correction. Subsequently, we investigated the expression patterns of TDP-43 during different stages of mammary gland development in mice and observed high expression levels during pregnancy and early lactation. Notably, TDP-43 knockout in the mouse mammary gland resulted in a significant reduction in milk production, with subsequent starvation of the pups within three weeks postpartum. Furthermore, we collected human milk from lactating women and analyzed the expression level of TDP-43 in isolated RNA from the milk lipids. Remarkably, we found a positive correlation between TDP-43 expression and higher milk output, suggesting that TDP-43 mRNA expression may serve as a potential indicator of lactation in parturient women [8]. Furthermore, we observed that conditional knockout of TDP-43 in the pubertal mammary gland had a significant inhibitory effect on mammary epithelial proliferation and mammary gland repopulation [83]. This finding is particularly significant as pubertal mammary gland development serves as the basis for subsequent breast lactation. Thus, these results provide evidence that TDP-43 may promote milk secretion at multiple levels.

TDP-43 in stem cells and other developmental processes

Stem cells are necessary for tissue development, homeostasis, and repair [84]. Abnormal differentiation of stem cells can have profound consequences on the development of individuals and organs [85, 86]. Multiple studies suggest that TDP-43 participates in the regulation of stem cell development. For example, Modic et al. showed that TDP-43 regulates pluripotency-differentiation transition in ESCs by regulating *Sox2* alternative polyadenylation. Additionally, TDP-43 is also reported to regulate the expression of the *Neat1* gene, repressing the formation of paraspeckles. Notably, during differentiation, TDP-43 is recruited to paraspeckles and facilitates the exit of ESCs from pluripotency and embryonic patterning [87]. Moreover, our previous study further revealed the importance of TDP-43 as a critical regulator in mammary stem cells [83].

TDP-43 is also implicated in various other developmental processes. Regarding myogenesis, Militello et al. found that TDP-43 binds to the muscle-enriched lncRNA *Myolinc* and regulates muscle-related gene expression, while knockdown of TDP-43 inhibits myogenic differentiation [88]. Furthermore, Xia et al. verified that CHCHD10 interacts with TDP-43 to promote myofiber regeneration and newly differentiated myotubes during myogenesis [89]. Li et al. revealed that H19 may act as a scaffold to recruit TDP-43 to the promoter region of myogenic differentiation (MYOD) in porcine satellite cells (PSCs), thereby activating the transcription of MYOD, and leading to PSC differentiation [90]. In the context of islets, TDP-43 is associated with early-stage insulin secretion through CaV1.2-induced exocytosis [91]. TDP-43 is also reported to regulate mammalian spinogenesis through translational repression of Rac1 [92].

TDP-43 in viral infection

Initially recognized as a protein associated with HIV, TDP-43 has been increasingly implicated in the entry, replication, and latency of other viruses [93] beyond HIV [2, 94, 95], including hepatitis B virus (HBV) [96], enteroviruses (EVs) [97–99], herpes simplex virus-2 (HSV) [93, 100], Theiler's murine encephalomyelitis virus (TMEV) [101], human endogenous retrovirus K (HERV-K) [102, 103], West Nile virus (WNV) [93, 104], and severe acute respiratory syndrome coronavirus 2 (SARS-CoV-2) [105–107]. However, the function of TDP-43 appears to vary markedly among individuals, depending on the specific viral context.

Multiple studies have provided evidence that TDP-43 acts as a protective factor in individuals during viral infections. Ou et al. first cloned and characterized TDP-43 in 1995, demonstrating its binding to the TAR sequence motifs of HIV-1 and inhibition of *HIV-1* gene expression [2]. Although subsequent reports indicated that TDP-43 expression in human immune cells is unrelated to HIV-1 replication [108], more recent observations have indicated that TDP-43 affects HIV viral envelope glycoprotein complex (Env) fusion, infection capacities, and viral production [94, 95]. Additionally, TDP-43 is also involved in the antiviral innate immune response. Notably, upon viral infection, TDP-43 is released from the lncRNA *Malat1* and undergoes cleavage to form the TDP-35 isoform. This process promotes the production of IRF3-initiated antiviral type I interferons (IFNs) by preventing IRF3 proteasomal degradation, thereby enhancing host defense against viral infection [90].

TDP-43 may also have negative effects on individuals. For example, during HBV infection, TDP-43 acts as a host factor that stimulates gene transcription by binding

to the HBV core promoter, forms complexes with other proteins supporting the HBV life cycle, and inhibits pregenomic HBV RNA splicing, thereby promoting the production of HBV replicative intermediates, mRNAs, proteins, and virions [96].

Conclusions and perspectives

TDP-43 in mammalian life cycle

The above findings highlight the wide-ranging roles of TDP-43 throughout various stages of mammalian life, encompassing early development to aging (Fig. 4).

Mammalian early embryonic development commences with the fusion of egg and sperm, giving rise to a totipotent zygote that develops into a fully formed individual. The production of sperm and eggs depends on germ cell meiosis. TDP-43 has been established as an essential protein for the completion of prophase I during spermatogenesis, and its deletion can lead to meiotic arrest, spermatogenesis failure, and low fertility in male mice [82] (Fig. 4A). However, as meiosis is not exclusive to spermatogenesis, also occurring during oogenesis, it would be worth exploring whether the absence of TDP-43 also impacts oogenesis and female fertility. In addition, as meiosis is not unique to mammals and TDP-43 exhibits a high degree of conservation across different species, it is likely that TDP-43 is essential for the reproductive system

in all sexually reproducing animals. Although the specific involvement of TDP-43 in the fertilization process and zygote formation remains uncertain (Fig. 4B), it is evident that TDP-43 is crucial for embryonic development (Fig. 4C). Various gene knockout models have confirmed that TDP-43 loss results in the death of pre-implantation embryos, emphasizing the significance of TDP-43 as a key regulatory gene in both sperm and egg formation and subsequent embryonic development [4–6, 46].

TDP-43 is involved in many life processes during adult development after birth (Fig. 4D, E). Firstly, systemic deletion of TDP-43 in postnatal mice causes rapid body fat loss and subsequent death, suggesting that TDP-43 is necessary for adult survival [6]. Secondly, TDP-43 is a key regulatory factor in mammary gland development and milk secretion, playing a crucial role in lactation and newborn survival [8, 83]. In addition, tissue-specific knockout or *in vitro* reduction of TDP-43 expression in other organs has confirmed that TDP-43 is also associated with stem cells [83, 87], muscle generation [88, 89], neural development [109–111], insulin secretion [91], and systemic lipid metabolism [6, 69, 72], underscoring the importance of TDP-43 in adult development.

With advancing age, TDP-43 is correlated with certain diseases (Fig. 4F), including neurodegenerative disorders and tumors, often accompanied by abnormal changes in

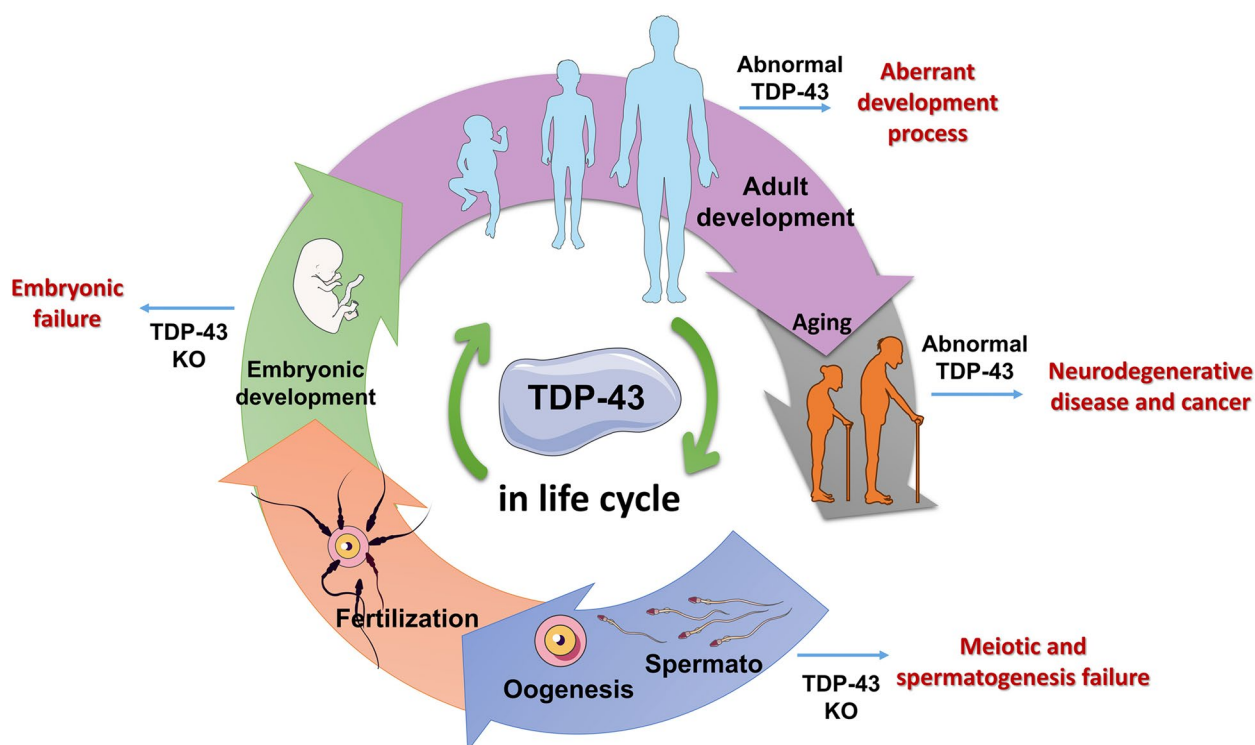


Fig. 4 Functions of TDP-43 in human life cycle. TDP-43 is reported to participate in spermatogenesis, embryonic development, adult development, and aging. Abnormal TDP-43 expression and location can lead to developmental failure and disease

its expression or localization. TDP-43 is a highly conserved gene with strict and precise gene expression regulation mechanisms, and even heterozygous knockout of TDP-43 does not affect its expression. This suggests that the self-regulatory capacity of TDP-43 may diminish with age, and disturbances in TDP-43 protein homeostasis can contribute to the development of various diseases, potentially leading to death. Overall, considering its extensive involvement in different life stages, it is plausible to consider that TDP-43 is a key regulatory gene involved in the entire process of mammalian life, from the very beginning to the very end. Future research should focus on the functions of TDP-43 in other contexts, especially in response to stressful conditions.

Implications from TDP-43 study

The above studies emphasize the diverse and sometimes opposing functions of TDP-43 in various tissues and developmental stages, which may be attributed to the following factors:

- 1) TDP-43 exhibits stage-specific functions. In degenerative diseases, TDP-43 dysfunction typically occurs in middle to old age. Studies have indicated that the absence of TDP-43 can lead to age-related neuronal degeneration [112]. Comparisons among different-aged patients with ALS have revealed that TDP-43 pathology is more severe in elderly ALS patients. TDP-43 deficient mice also exhibit progressive motor dysfunction and neuropathological changes. Similarly, the consequences of TDP-43 deficiency in oligodendrocytes depend on their maturation stage. Early deletion leads to progressive degeneration of mature oligodendrocytes, leading to seizure and premature death. In contrast, late deletions retain oligodendrocytes to a large extent, and mice survive without seizure, indicating that TDP-43 may be dispensable in mature oligodendrocytes [113].
- 2) The function of TDP-43 is context-dependent and influenced by basal expression levels in cells and tissues. Notably, in nerve cells, genome RNA cross-linking immunoprecipitation (HITS-CLIP) technology has revealed that TDP-43 can bind with more than 6 000 mRNA targets, representing about 30% of the entire transcriptome [114, 115], thus suggesting the importance of TDP-43 in the nervous system. Regarding breast lactation, TDP-43 can stabilize the mRNA expression levels of *Btn1a1* and *Xdh*, thereby maintaining normal initiation of lactation. Notably, these two genes are transcriptionally activated exclusively in breast lactation cells during lactation, illustrating that TDP-43 regulates their expression

through post-transcriptional mechanisms [8]. Furthermore, the cellular expression levels of genes can vary among different cell types, resulting in distinct downstream targets regulated by TDP-43 and consequently exhibiting diverse functions.

- 3) TDP-43 serves a critical function in the cellular stress response. Notably, the TDP-43 protein can sense intracellular signals, such as misfolding proteins Sup35, Pab1, and Pub1 [116]. In ovariectomized mice, progesterone treatment can promote TDP-43 expression, suggesting that TDP-43 may be regulated by progesterone [117]. TDP-43 can also sense extracellular signals, such as viral infections [2, 93, 95] and oxygen free radicals [118]. Moreover, TDP-43 is defined as a component of stress granules [119, 120] and participates in cellular liquid-liquid phase separation [121, 122]. Under a certain degree of internal and external stimuli, TDP-43 maintains a relatively stable expression state through self-regulation of mRNA, enabling normal responses to relevant stimuli and maintenance of life processes. However, when cells are subjected to intense stimulation or long-term imbalance, the expression or subcellular localization of TDP-43 may be disrupted, leading to disease manifestation. Neural cells continuously receive internal signals from synapses, and TDP-43 serves as a stress sensor gene, detecting and responding to specific stimuli at precise times and in specific cell types. This stress response mechanism is vital for maintaining a steady state during cellular processes.

Authors' contributions

HK and LZ designed original draft. HK was responsible for acquisition analysis, interpretation of data, prepared all figures and wrote the main manuscript text. KL assisted in collecting materials. BJ and LZ revised and reviewed the manuscript. All authors read and approved the final manuscript.

Funding

This work was supported by the National Natural Science Foundation of China (Nos. 32200679, 82260488 and 32360164), "Double Thousand Plan" of Jiangxi Province (jxsq2023101075), Science Fund for Distinguished Young Scholars of Jiangxi Province (20232ACB215001), China Postdoctoral Science Foundation (Nos. 2021TQ0137 and 2021M701544), Jiangxi Provincial Natural Science Foundation (Nos. 20224BAB205014 and 20224BAB216071), Natural Science Foundation of Chongqing (No. CSTB2022NSCQ-MSX056), Postgraduate Innovation Special Fund Project of Jiangxi (No. YC2022-s047), College Students' Innovative Entrepreneurial Training Plan Program of Nanchang University (Nos. 2022CX024 and 2022CX158), and Scientific Research Training Program of Nanchang University (2023).

Availability of data and materials

Not applicable.

Declarations

Competing interests

The authors declare no competing interests.

Received: 27 July 2023 Accepted: 26 September 2023
Published online: 23 November 2023

References

- Ayala YM, et al. Human, Drosophila, and C.elegans TDP43: nucleic acid binding properties and splicing regulatory function. *J Mol Biol.* 2005;348:575–88.
- Ou SH, Wu F, Harrich D, Garcia-Martinez LF, Gaynor RB. Cloning and characterization of a novel cellular protein, TDP-43, that binds to human immunodeficiency virus type 1 TAR DNA sequence motifs. *J Virol.* 1995;69:3584–96.
- Neumann M, et al. Ubiquitinated TDP-43 in frontotemporal lobar degeneration and amyotrophic lateral sclerosis. *Science (New York, NY).* 2006;314:130–3.
- Wu LS, et al. TDP-43, a neuro-pathosignature factor, is essential for early mouse embryogenesis. *Genesis (New York, NY: 2000).* 2010;48:56–62.
- Sephton CF, et al. TDP-43 is a developmentally regulated protein essential for early embryonic development. *J Biol Chem.* 2010;285:6826–34.
- Chiang PM, et al. Deletion of TDP-43 inhibits progression of triple-negative breast cancer in coordination with SRSF3. *Proc Natl Acad Sci U S A.* 2010;107:16320–4.
- Acharya KK, Govind CK, Shore AN, Stoler MH, Reddi PP. Cis-requirement for the maintenance of round spermatid-specific transcription. *Dev Biol.* 2006;295:781–90.
- Zhao L, et al. TDP-43 facilitates milk lipid secretion by post-transcriptional regulation of Btn1a1 and Xdh. *Nat Commun.* 2020;11:341.
- Ke H, et al. Loss of TDP43 inhibits progression of triple-negative breast cancer in coordination with SRSF3. *Proc Natl Acad Sci U S A.* 2018;115:E3426–35.
- Ayala YM, et al. Structural determinants of the cellular localization and shuttling of TDP-43. *J Cell Sci.* 2008;121:3778–85.
- Rossi C, et al. Cell Stress Induces Mislocalization of Transcription Factors with Mitochondrial Enrichment. *Int J Mol Sci.* 2021;22(16):8853.
- Wang W, et al. The inhibition of TDP-43 mitochondrial localization blocks its neuronal toxicity. *Nat Med.* 2016;22:869–78.
- Loganathan S, Lehmkuhl EM, Eck RJ, Zarnescu DC. To Be or Not To Be... Toxic-Is RNA Association With TDP-43 Complexes Deleterious or Protective in Neurodegeneration? *Front Mol Biosci.* 2019;6:154.
- Huang C, Yan S, Zhang Z. Maintaining the balance of TDP-43, mitochondria, and autophagy: a promising therapeutic strategy for neurodegenerative diseases. *Transl Neurodegener.* 2020;9:40.
- Suk TR, Rousseaux MWC. The role of TDP-43 mislocalization in amyotrophic lateral sclerosis. *Mol Neurodegener.* 2020;15:45.
- Prasad A, Bharathi V, Sivalingam V, Girdhar A, Patel BK. Molecular Mechanisms of TDP-43 Misfolding and Pathology in Amyotrophic Lateral Sclerosis. *Front Mol Neurosci.* 2019;12:25.
- Kuo PH, Doudeva LG, Wang YT, Shen CK, Yuan HS. Structural insights into TDP-43 in nucleic-acid binding and domain interactions. *Nucleic Acids Res.* 2009;37:1799–808.
- Tsoi PS, et al. The N-Terminal Domain of ALS-Linked TDP-43 Assembles without Misfolding. *Angewandte Chemie (International ed in English).* 2017;56:12590–3.
- François-Moutal L, et al. Structural Insights Into TDP-43 and Effects of Post-translational Modifications. *Front Mol Neurosci.* 2019;12:301.
- Zhang YJ, et al. The dual functions of the extreme N-terminus of TDP-43 in regulating its biological activity and inclusion formation. *Hum Mol Genet.* 2013;22:3112–22.
- Winton MJ, et al. Disturbance of nuclear and cytoplasmic TAR DNA-binding protein (TDP-43) induces disease-like redistribution, sequestration, and aggregate formation. *J Biol Chem.* 2008;283:13302–9.
- Giannini M, Bayona-Feliu A. TDP-43 mutations link Amyotrophic Lateral Sclerosis with R-loop homeostasis and R loop-mediated DNA damage. *PLoS Genet.* 2020;16(12):e1009260.
- Wood M, et al. TDP-43 dysfunction results in R-loop accumulation and DNA replication defects. *J Cell Sci.* 2020. p. 133.
- Mitra J, et al. Motor neuron disease-associated loss of nuclear TDP-43 is linked to DNA double-strand break repair defects. *Proc Natl Acad Sci U S A.* 2019;116:4696–705.
- Mercado PA, Ayala YM, Romano M, Buratti E, Baralle FE. Depletion of TDP 43 overrides the need for exonic and intronic splicing enhancers in the human apoA-II gene. *Nucleic acids research.* 2005;33:6000–10.
- Passoni M, De Conti L, Baralle M, Buratti E. UG repeats/TDP-43 interactions near 5' splice sites exert unpredictable effects on splicing modulation. *J Mol Biol.* 2012;415:46–60.
- Dreumont N, et al. Antagonistic factors control the unproductive splicing of SC35 terminal intron. *Nucleic Acids Res.* 2010;38:1353–66.
- Bose JK, Wang IF, Hung L, Tarn WY, Shen CK. TDP-43 overexpression enhances exon 7 inclusion during the survival of motor neuron pre-mRNA splicing. *J Biol Chem.* 2008;283:28852–9.
- Gumina V, et al. TDP-43 and NOVA-1 RNA-binding proteins as competitive splicing regulators of the schizophrenia-associated TNIK gene. *Biochim et Biophys Acta Gene Regul Mech.* 2019;1862:194413.
- Deshaies JE, et al. TDP-43 regulates the alternative splicing of hnRNP A1 to yield an aggregation-prone variant in amyotrophic lateral sclerosis. *Brain.* 2018;141:1320–33.
- Guo L, et al. TDP43 promotes stemness of breast cancer stem cells through CD44 variant splicing isoforms. *Cell Death Dis.* 2022;13(5):428.
- Tejedor AR, Garaizar A, Ramírez J, Espinosa JR. RNA modulation of transport properties and stability in phase-separated condensates. *Biophys J.* 2021;120:5169–86.
- Majumder P, Chu JF, Chatterjee B, Swamy KB, Shen CJ. Co-regulation of mRNA translation by TDP-43 and Fragile X Syndrome protein FMRP. *Acta Neuropathol.* 2016;132:721–38.
- Kawahara Y, Mieda-Sato A. TDP-43 promotes microRNA biogenesis as a component of the Drosha and Dicer complexes. *Proc Natl Acad Sci U S A.* 2012;109:3347–52.
- Ma X, et al. The Regulatory Role of RNA Metabolism Regulator TDP-43 in Human Cancer. *Front Oncol.* 2021;11:755096.
- Tank EM, et al. Abnormal RNA stability in amyotrophic lateral sclerosis. *Nat Commun.* 2018;9:2845.
- Ayala YM, Misteli T, Baralle FE. TDP-43 regulates retinoblastoma protein phosphorylation through the repression of cyclin-dependent kinase 6 expression. *Proc Natl Acad Sci U S A.* 2008;105:3785–9.
- Liu X, Li D, Zhang W, Guo M, Zhan Q. Long non-coding RNA gadd7 interacts with TDP-43 and regulates Cdk6 mRNA decay. *EMBO J.* 2012;31:4415–27.
- Gu J, et al. TDP-43 suppresses tau expression via promoting its mRNA instability. *Nucleic Acids Res.* 2017;45:6177–93.
- Sidibé H, et al. TDP-43 stabilizes G3BP1 mRNA: relevance to amyotrophic lateral sclerosis/frontotemporal dementia. *Brain.* 2021;144:3461–76.
- Costessi L, Porro F, Iaconig A, Muro AF. TDP-43 regulates β -adducin (Add2) transcript stability. *RNA Biol.* 2014;11:1280–90.
- Ying Z, et al. TARDBP/TDP-43 regulates autophagy in both MTORC1-dependent and MTORC1-independent manners. *Autophagy.* 2016;12:707–8.
- Izumikawa K, et al. TDP-43 stabilises the processing intermediates of mitochondrial transcripts. *Sci Rep.* 2017;7:7709.
- Arai T, et al. TDP-43 is a component of ubiquitin-positive tau-negative inclusions in frontotemporal lobar degeneration and amyotrophic lateral sclerosis. *Biochem Biophys Res Commun.* 2006;351:602–11.
- Lee EB, Lee VM, Trojanowski JQ. Gains or losses: molecular mechanisms of TDP43-mediated neurodegeneration. *Nat Rev Neurosci.* 2011;13:38–50.
- Liao YZ, Ma J, Dou JZ. The Role of TDP-43 in Neurodegenerative Disease. *Mol Neurobiol.* 2022;59:4223–41.
- Postel-Vinay S, et al. Common variants near TARDBP and EGR2 are associated with susceptibility to Ewing sarcoma. *Nat Genet.* 2012;44:323–7.
- Fang HY, Chen SB, Guo DJ, Pan SY, Yu ZL. Proteomic identification of differentially expressed proteins in curcumin-treated MCF-7 cells. *Phytomedicine.* 2011;18:697–703.
- Guo F, et al. Regulation of MALAT1 expression by TDP43 controls the migration and invasion of non-small cell lung cancer cells in vitro. *Biochem Biophys Res Commun.* 2015;465:293–8.
- Zhao HL, et al. Long noncoding RNA MIAT promotes the growth and metastasis of non-small cell lung cancer by upregulating TDP43. *Eur Rev Med Pharmacol Sci.* 2020;24:7209.

51. Chang IS, et al. Genetic Modifiers of Progression-Free Survival in Never-Smoking Lung Adenocarcinoma Patients Treated with First-Line Tyrosine Kinase Inhibitors. *Am J Respir Crit Med*. 2017;195:663–73.
52. Li Y, et al. LncRNAs LCCTRL3 and LCCTRL4 at chromosome 4q12 diminish EGFR-TKIs efficiency in NSCLC through stabilizing TDP43 and EIF2S1. *Signal Transduct Target Ther*. 2022;7(1):30.
53. Park YY, et al. Tat-activating regulatory DNA-binding protein regulates glycolysis in hepatocellular carcinoma by regulating the platelet isoform of phosphofruktokinase through microRNA 520. *Hepatology* (Baltimore, Md). 2013;58:182–91.
54. Guo F, et al. TDP-43 induces EMT and promotes hepatocellular carcinoma metastasis via activating Wnt/ β -catenin signaling pathway. *Am J Cancer Res*. 2020;10:3285–301.
55. Liu BW, et al. TDP-43 upregulates lipid metabolism modulator ABHD2 to suppress apoptosis in hepatocellular carcinoma. *Commun Biol*. 2022;5(1):816.
56. Zeng Q, et al. Identification of TDP-43 as an oncogene in melanoma and its function during melanoma pathogenesis. *Cancer Biol Ther*. 2017;18:8–15.
57. Lin TW, et al. TDP-43/HDAC6 axis promoted tumor progression and regulated nutrient deprivation-induced autophagy in glioblastoma. *Oncotarget*. 2017;8:56612–25.
58. Kim PY, et al. High TDP43 expression is required for TRIM16-induced inhibition of cancer cell growth and correlated with good prognosis of neuroblastoma and breast cancer patients. *Cancer Letters*. 2016;374:315–23.
59. Zaman S, Chobrutskiy BI, Blanck G. MAPT (Tau) expression is a biomarker for an increased rate of survival in pediatric neuroblastoma. *Cell cycle* (Georgetown, Tex). 2018;17:2474–83.
60. Lee K, Suzuki H, Aiso S, Matsuoka M. Overexpression of TDP-43 causes partially p53-dependent G2/M arrest and p53-independent cell death in HeLa cells. *Neurosci Lett*. 2012;506:271–6.
61. Chen X, et al. TDP-43 regulates cancer-associated microRNAs. *Protein Cell*. 2018;9:848–66.
62. Ibáñez K, Boullosa C, Tabarés-Seisdedos R, Baudot A, Valencia A. Molecular evidence for the inverse comorbidity between central nervous system disorders and cancers detected by transcriptomic meta-analyses. *PLoS Genet*. 2014;10:e1004173.
63. Maor-Nof M, et al. p53 is a central regulator driving neurodegeneration caused by C9orf72 poly(PR). *Cell*. 2021;184:689–708.e620.
64. Salemi, M. & Mogavero, M.P. Examples of Inverse Comorbidity between Cancer and Neurodegenerative Diseases: A Possible Role for Noncoding RNA. 11(2022).
65. Seo J, Park M. Molecular crosstalk between cancer and neurodegenerative diseases. *Cell Mol Life Sci*. 2020;77:2659–80.
66. Nan Y, Wang S, Jia W. Caspase independent cleavages of TDP-43 generates 35kD fragment that cause apoptosis of breast cancer cells. *Biochem Biophys Res Commun*. 2018;497:51–7.
67. Wu LS, Cheng WC, Shen CK. Targeted depletion of TDP-43 expression in the spinal cord motor neurons leads to the development of amyotrophic lateral sclerosis-like phenotypes in mice. *J Biol Chem*. 2012;287:27335–44.
68. Tsao W, et al. Rodent models of TDP-43: recent advances. *Brain Res*. 2012;1462:26–39.
69. Stallings NR, et al. TDP-43, an ALS linked protein, regulates fat deposition and glucose homeostasis. *PLoS One*. 2013;8:e71793.
70. Coughlan KS, Halang L, Woods I, Prehn JH. A high-fat jelly diet restores bioenergetic balance and extends lifespan in the presence of motor dysfunction and lumbar spinal cord motor neuron loss in TDP-43A315T mutant C57BL6/J mice. *Dis Model Mech*. 2016;9(9):1029–379.
71. Egawa N, et al. TDP-43 regulates cholesterol biosynthesis by inhibiting sterol regulatory element-binding protein 2. *Sci Rep*. 2022;12:7988.
72. Li P, et al. A liver-enriched long non-coding RNA, lncLSTR, regulates systemic lipid metabolism in mice. *Cell Metabol*. 2015;21:455–67.
73. Lee S, Lee TA, Song SJ, Park T, Park B. Hyperproduction of IL-6 caused by aberrant TDP-43 overexpression in high-fat diet-induced obese mice. *FEBS Lett*. 2015;589:1825–31.
74. Yao Y, Ding L, Huang X. Diverse Functions of Lipids and Lipid Metabolism in Development. 2020.
75. Reddi PP, et al. Spermatid-specific promoter of the SP-10 gene functions as an insulator in somatic cells. *Dev Biol*. 2003;262:173–82.
76. Acharya KK, Govind CK, Shore AN, Stoler MH, Reddi PP. cis-requirement for the maintenance of round spermatid-specific transcription. *Dev Biol*. 2006;295:781–90.
77. Lalmansingh AS, Urekar CJ, Reddi PP. TDP-43 is a transcriptional repressor: the testis-specific mouse *acr1* gene is a TDP-43 target in vivo. *J Biol Chem*. 2011;286:10970–82.
78. Abhyankar MM, Urekar C, Reddi PP. A novel CpG-free vertebrate insulator silences the testis-specific SP-10 gene in somatic tissues: role for TDP-43 in insulator function. *J Biol Chem*. 2007;282:36143–54.
79. Osuru HP, et al. Immunolocalization of TAR DNA-binding protein of 43 kDa (TDP-43) in mouse seminiferous epithelium. *Mol Reprod Dev*. 2017;84:675–85.
80. Varghese DS, et al. Aberrant expression of TAR DNA binding protein-43 is associated with spermatogenic disorders in men. *Reprod Fert Dev*. 2016;28:713–22.
81. Zomer HD, et al. Sertoli cells require TDP-43 to support spermatogenesis. *Biol Reprod*. 2022;107:1345–59.
82. Campbell KM, et al. Loss of TDP-43 in male germ cells causes meiotic failure and impairs fertility in mice. *J Biol Chem*. 2021;297:101231.
83. Zhao L, et al. TDP-43 is Required for Mammary Gland Repopulation and Proliferation of Mammary Epithelial Cells. *Stem Cells Dev*. 2019;28:944–53.
84. Royall LN, Jessberger S. How stem cells remember their past. *Curr Opin Cell Biol*. 2021;69:17–22.
85. Wang C, et al. UTX regulates mesoderm differentiation of embryonic stem cells independent of H3K27 demethylase activity. *Proc Natl Acad Sci U S A*. 2012;109:15324–9.
86. Huo Y, Macara IG. The Par3-like polarity protein Par3L is essential for mammary stem cell maintenance. *Nat Cell Biol*. 2014;16:529–37.
87. Modic M, et al. Cross-Regulation between TDP-43 and Paraspeckles Promotes Pluripotency-Differentiation Transition. *Molecular cell*. 2019;74:951–965.e913.
88. Milletello G, et al. A novel long non-coding RNA Myolinc regulates myogenesis through TDP-43 and Filip1. *J Mol Cell Biol*. 2018;10:102–17.
89. Xia W, et al. Chchd10 is dispensable for myogenesis but critical for adipose browning. *Cell Regen*. 2022;11(1):14.
90. Liu W, et al. LncRNA Malat1 inhibition of TDP43 cleavage suppresses IRF3-initiated antiviral innate immunity. *Proc Natl Acad Sci U S A*. 2020;117:23695–706.
91. Araki K, et al. TDP-43 regulates early-phase insulin secretion via CaV1.2-mediated exocytosis in islets. *J Clin Invest*. 2019;129:3578–93.
92. Majumder P, et al. TDP-43 regulates the mammalian spinogenesis through translational repression of Rac1. *Acta Neuropathol*. 2012;124:231–45.
93. Rahic Z, Buratti E, Cappelli S. Reviewing the Potential Links between Viral Infections and TDP-43 Proteinopathies. *Int J Mol Sci*. 2023;24(2):1581.
94. Cabrera-Rodríguez R, et al. Transactive Response DNA-Binding Protein (TARDBP/TDP-43) Regulates Cell Permissivity to HIV-1 Infection by Acting on HDAC6. *Int J Mol Sci*. 2022;23(11):6180.
95. Cabrera-Rodríguez R, et al. TDP-43 Controls HIV-1 Viral Production and Virus Infectiveness. *Int J Mol Sci*. 2023;24(8):7658.
96. Makokha GN, et al. Regulation of the Hepatitis B virus replication and gene expression by the multi-functional protein TARDBP. *Sci Rep*. 2019;9:8462.
97. Fung G, et al. Cytoplasmic translocation, aggregation, and cleavage of TDP-43 by enteroviral proteases modulate viral pathogenesis. *Cell Death Differ*. 2015;22:2087–97.
98. Xue YC, et al. Enteroviral Infection Leads to Transactive Response DNA-Binding Protein 43 Pathology in Vivo. *Am J Pathol*. 2018;188:2853–62.
99. Zhang L, et al. Enterovirus D68 Infection Induces TDP-43 Cleavage, Aggregation, and Neurotoxicity. *J virol*. 2023;97:e0042523.
100. Cabrera JR, Rodríguez-Izquierdo I, Jiménez JL, Muñoz-Fernández M. Analysis of ALS-related proteins during herpes simplex virus-2 latent infection. *Journal of neuroinflammation*. 2020;17:371.
101. Masaki K, Sonobe Y, Ghadge G, Pytel P, Roos RP. TDP-43 proteinopathy in Theiler's murine encephalomyelitis virus infection. *PLoS Pathog*. 2019;15:e1007574.

102. Manghera M, Ferguson-Parry J, Douville RN. TDP-43 regulates endogenous retrovirus-K viral protein accumulation. *Neurobiol Dis.* 2016;94:226–36.
103. Simula ER, Arru G, Zarbo IR, Solla P, Sechi LA. TDP-43 and HERV-K Envelope-Specific Immunogenic Epitopes Are Recognized in ALS Patients. *Viruses.* 2021;13(11):2301.
104. Constant O, Barthelemy J, Nagy A, Salinas S, Simonin Y. West Nile Virus Neuroinfection in Humans: Peripheral Biomarkers of Neuroinflammation and Neuronal Damage. *Viruses.* 2022;14(4):756.
105. Idrees D, Kumar V. SARS-CoV-2 spike protein interactions with amyloidogenic proteins: Potential clues to neurodegeneration. *Biochem Biophys Res Commun.* 2021;554:94–8.
106. Yang J, et al. The SARS-CoV-2 main protease induces neurotoxic TDP-43 cleavage and aggregates. *Signal Transduct Targeted Ther.* 2023;8:109.
107. Dehipawala S, Cheung E, Tremberger G, Cheung T. Entropy and Fractal Dimension Study of the TDP-43 Protein Low Complexity Domain Sequence in ALS Disease Severity and SARS-CoV-2 Gene Sequences in Virulence Variability. *Entropy (Basel, Switzerland).* 2021;23(8):1038.
108. Nehls J, Koppensteiner H, Brack-Werner R, Floss T, Schindler M. HIV-1 replication in human immune cells is independent of TAR DNA binding protein 43 (TDP-43) expression. *PLoS One.* 2014;9:e105478.
109. Ho WY, et al. TDP-43 mediates SREBF2-regulated gene expression required for oligodendrocyte myelination. *The Journal of cell biology.* 2021;220(9):e201910213.
110. Koza P, et al. Neuronal TDP-43 depletion affects activity-dependent plasticity. *Neurobiol Dis.* 2019;130:104499.
111. Sephton CF, Cenik B, Cenik BK, Herz J, Yu G. TDP-43 in central nervous system development and function: clues to TDP-43-associated neurodegeneration. *Biol Chem.* 2012;393:589–94.
112. Iguchi Y, et al. Loss of TDP-43 causes age-dependent progressive motor neuron degeneration. *Brain.* 2013;136:1371–82.
113. Heo D, Ling JP. Stage-specific control of oligodendrocyte survival and morphogenesis by TDP-43. *Elife.* 2022;11:e75230.
114. Tollervey JR, et al. Characterizing the RNA targets and position-dependent splicing regulation by TDP-43. *Nat Neurosci.* 2011;14:452–8.
115. Xiao S, et al. RNA targets of TDP-43 identified by UV-CLIP are deregulated in ALS. *Mol Cell Neurosci.* 2011;47:167–80.
116. Pillai M, Jha SK. Early Metastable Assembly during the Stress-Induced Formation of Worm-like Amyloid Fibrils of Nucleic Acid Binding Domains of TDP-43. *Biochemistry.* 2020;59(3):315–32859.
117. Fernandez-Valdivia R, et al. Transcriptional response of the murine mammary gland to acute progesterone exposure. *Endocrinology.* 2008;149:6236–50.
118. Zuo X, et al. TDP-43 aggregation induced by oxidative stress causes global mitochondrial imbalance in ALS. *Nat Struct Mol Biol.* 2021;28:132–42.
119. Khalfallah Y, et al. TDP-43 regulation of stress granule dynamics in neurodegenerative disease-relevant cell types. *Sci Rep.* 2018;8:7551.
120. Niss F, Piñero-Paez L, Zaidi W, Hallberg E, Ström AL. Key Modulators of the Stress Granule Response TIA1, TDP-43, and G3BP1 Are Altered by Polyglutamine-Expanded ATXN7. *Mol Neurobiol.* 2022;59:5236–51.
121. Conicella AE, et al. TDP-43 α -helical structure tunes liquid-liquid phase separation and function. *Proc Natl Acad Sci U S A.* 2020;117:5883–94.
122. Wang C, et al. Stress Induces Dynamic, Cytotoxicity-Antagonizing TDP-43 Nuclear Bodies via Paraspeckle LncRNA NEAT1-Mediated Liquid-Liquid Phase Separation. *Mol Cell.* 2020;79:443–458.e447.

Publisher's Note

Springer Nature remains neutral with regard to jurisdictional claims in published maps and institutional affiliations.

Ready to submit your research? Choose BMC and benefit from:

- fast, convenient online submission
- thorough peer review by experienced researchers in your field
- rapid publication on acceptance
- support for research data, including large and complex data types
- gold Open Access which fosters wider collaboration and increased citations
- maximum visibility for your research: over 100M website views per year

At BMC, research is always in progress.

Learn more biomedcentral.com/submissions

