

RESEARCH

Open Access



Anti-inflammatory effect of fullerene C₆₀ in a mice model of atopic dermatitis

Nadezda Shershakova*, Elena Baraboshkina, Sergey Andreev*, Daria Purgina, Irina Struchkova, Oleg Kamyshnikov, Alexandra Nikonova and Musa Khaitov

Abstract

Background: Water-soluble form of fullerene C₆₀ is a promising tool for the control of ROS-dependent inflammation including allergic diseases. Anti-inflammatory effects of C₆₀ (nC₆₀) aqueous dispersion were evaluated in the mouse models of atopic dermatitis using subcutaneous (SC) and epicutaneous (EC) applications during 50 days period. A highly stable nC₆₀ was prepared by exhaustive dialysis of water-organic C₆₀ solution against water, where the size and ζ-potential of fullerene nanoparticles are about 100 nm and −30 mV, respectively.

Results: To induce skin inflammation, female BALB/c mice were EC sensitized with ovalbumin three times during one-weekly exposures. The nC₆₀ solution was administrated in mice subcutaneously (SC) (0.1 mg/kg) and epicutaneously (EC) (1 mg/kg). Significant suppression of IgE and Th2 cytokines production and a concomitant rise in concentrations of Th1 cytokines were observed in nC₆₀-treated groups. In addition, a significant increase in the levels of Foxp3⁺ and filaggrin mRNA expression was observed at EC application. Histological examination of skin samples indicated that therapeutic effect was achieved by both EC and SC treatment, but it was more effective with EC. Pronounced reduction of the eosinophil and leukocyte infiltration in treated skin samples was observed.

Conclusions: We suppose that nC₆₀ treatment shifts immune response from Th2 to Th1 and restores to some extent the function of the skin barrier. This approach can be a good alternative to the treatment of allergic and other inflammatory diseases.

Keywords: Fullerene C₆₀, Atopic dermatitis, Mouse model, Cytokine

Background

Atopic dermatitis (AD) is a chronic inflammatory skin disease that predominantly affects children and is characterized by skin lesions, persistent erythema, scaling, excoriations, and pruritus. In addition, the disease is commonly associated with allergic rhinitis and asthma. The number of AD patients increased by 10–30 % in children and 2–10 % in adults in the last 30 years worldwide [1–3]. AD is the result of complex interactions among several genetic factors, deficiencies in skin barrier function, exposure to various allergens and infectious agents and features of the immune response [4–6]. In 60 % of patients, AD starts before the age of 6 years, in 18 % the onset is after the age of 20 [7]. Its pathogenesis involves

impairments in the skin barrier, allowing abnormally enhanced dermal presentation of antigens/allergens to the immune system [8, 9]. In many AD patients Th2/IgE-mediated allergic reactions play leading role, however, Th2 cells predominate in the initial stage lesions with a switch to Th1-cells in a chronic phase. As a rule, diseases arising from a dysfunction of immune cells and/or their products often manifest with skin symptoms [10]. Many observations suggest that allergic and inflammatory skin diseases like AD are mediated by oxidative stress [11–13]. Mast cells generate mainly intracellular reactive oxygen species (ROS) following the aggregation of FcεRI; these ROS may act as secondary messengers in the induction of several biological responses [14]. A generation of ROS induces oxidative protein damage in the stratum corneum, which leads to the disruption of barrier function and the exacerbation of AD [15, 16].

*Correspondence: nn.shershakova@nrcii.ru; sm.andreev@nrcii.ru
NRC Institute of Immunology FMBA of Russia, Moscow, Russia

Fullerene is a molecular carbon form with cage spheroid structure, and it possesses strong antioxidant activity. At present, the fullerene C_{60} is produced industrially in large quantities and is commercially available. Its unique properties include strong electron-acceptor activity, high polarizability and hydrophobicity; a large surface of C_{60} molecule allows attachment of various hydrophilic addends, providing prospects for the design of new biomedical products. Fullerene and some of its derivatives as potent antioxidants (in vitro and in vivo) are capable of effective ROS inactivation [17]. An important feature of fullerene C_{60} is its low toxicity [18, 19], a permeability through biological membranes [20, 21], and lack of immunogenicity [22]. Thus, fullerene presents a promising therapeutic agent for the treatment of allergies and other inflammatory diseases such as Parkinson's, Alzheimer's and AD.

Anti-inflammatory activity of fullerene derivatives was recorded by several investigators, and, in particular, researchers from the Luna group (led by Dr. C. L. Kopley) were the pioneers in this field [23–26]. Some C_{60} -driven compounds were able to inhibit an IgE-dependent allergic response. For instance, fullerenol ($C_{60}(OH)_n$) and amino-fullerene ($C_{60}(NHCH_2CH_3)_n$) inhibited in vitro IgE-dependent degranulation of mast cells and secretion of cytokines and prostaglandins in response to allergen and authors suggested that this effect is associated with lower levels of ROS in the stimulated cells [23]. On the other hand, the C_{60} aqueous dispersion obtained by Andrievsky's method [27] induced gene expression of proinflammatory cytokines (IL-1, TNF- α , IL-6) and Th1 cytokines (IL-12, IFN- γ) in mice via aerosol administration to the airways [28]. Aqueous dispersion of C_{60} (nC_{60}) provided inhibitory effect on IgE-mediated histamine release from peripheral blood basophils in vitro and suppressed anaphylaxis in mice caused by administration of ovalbumin [29]. Similar C_{60} dispersion exhibited significant modulator activity on the DTH reaction suppressing Th1 cytokines release [30, 31].

Fullerene C_{60} in aqueous media always exists in form of nanoparticles (clusters), and hence, the degree of their possible undesirable effect is still being discussed [32, 33]. In several studies, nanoparticles are perceived as toxic agents, in others—the damage is not detected. As was shown, the polystyrene nanoparticles aggravated AD-like skin lesions [34] and metal oxide nanoparticles such as ZnO induced systemic production of IgE antibodies [35]. Currently, nanoparticles were used for transcutaneous drugs delivery, but not for the AD therapy [36, 37]. It should be noted, that the use of fullerene at the AD has not been described.

The purpose of this study was to evaluate the therapeutic properties of the nC_{60} obtained by novel

biocompatible method for the treatment of AD induced in a mouse model [38].

Results

Aqueous fullerene solution, nC_{60}

Recently, we have proposed a new simple method for preparation of C_{60} aqueous solution using a “dialysis principle” [39]. Shortly, the protocol design includes the dissolution of crystalline fullerene in N-methylpyrrolidone (NMP), dilution of this solution with distilled water or aqueous solution of an L-amino acid used as a stabilizing agent with subsequent exhaustive dialysis against deionized water. Thus, the protocol excludes the use of toxic organic solvents (toluene, tetrahydrofuran) as well as sonication, heating and durable mixing frequently used in the known methods [27, 40, 41]. This approach provides a high conversion of the fullerene C_{60} from the crystalline state to the solution with concentration of C_{60} up to 1 g/l (at additional vacuum concentration) and hydrodynamic size of C_{60} particles about 80–100 nm (Table 1). Absorption spectrum of nC_{60} is characterized by three intense maxima at 219, 265 and 344 nm and weak broad band between 400 and 500 nm, and it practically did not differ from those of nC_{60} obtained through other methods. FTIR spectra of dried nC_{60} shows vibration bands of free C_{60} molecules at 1182 and 1428 cm^{-1} and additional ones at 3500–3200, 1650–1660, and 1000–1100 cm^{-1} due to obvious presence of residual water molecules and NMP (as a donor–acceptor complex). Calculations on the molar ratio of NMP to fullerene displayed the value 0.7–1.1, while a calculation of the ratio H_2O to C_{60} based on the hydrogen content gave values from 5 to 10. The NMP is known to have low toxicity via oral, dermal and inhalation routes of delivery. In medicine, it has a long track recorded as a constituent in medical devices approved by the European Commission and FDA and thus can be considered as safe [42]. In addition, good heat resistance

Table 1 Physical parameters of the nC_{60}

Parameter	Value
Feasible concentration ^a	~1 mg/ml
Average size (nm)	80–100 nm
PDI (polydispersity index)	0.175 ± 5
Zeta potential (mV)	–27/30
UV–Vis major absorption peaks (nm)	219 (s), 265 (s), 344 (s), 450 (bw)
FTIR spectra (cm^{-1})	525, 1000–1100, 1182, 1428, 1650–1660, 3200–3500
Mol. ratio C_{60} :NMP	~1
Mol. ratio C_{60} : H_2O	5–10

s strong, bw broad weak

^a With optional vacuum evaporation

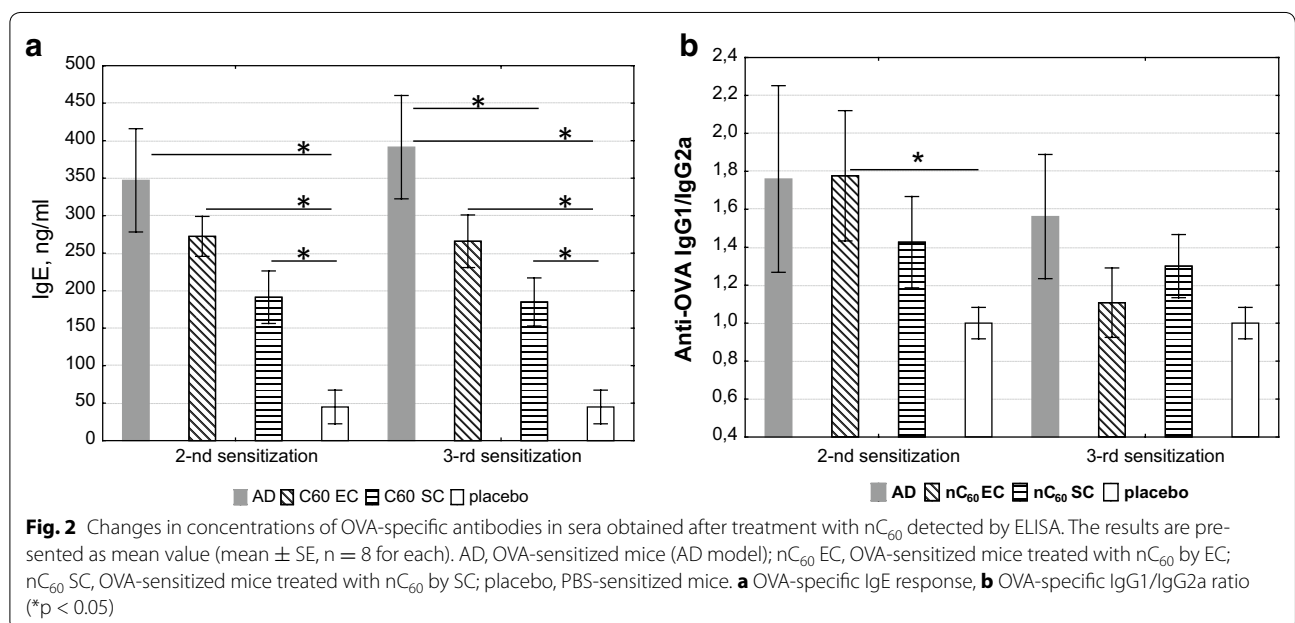
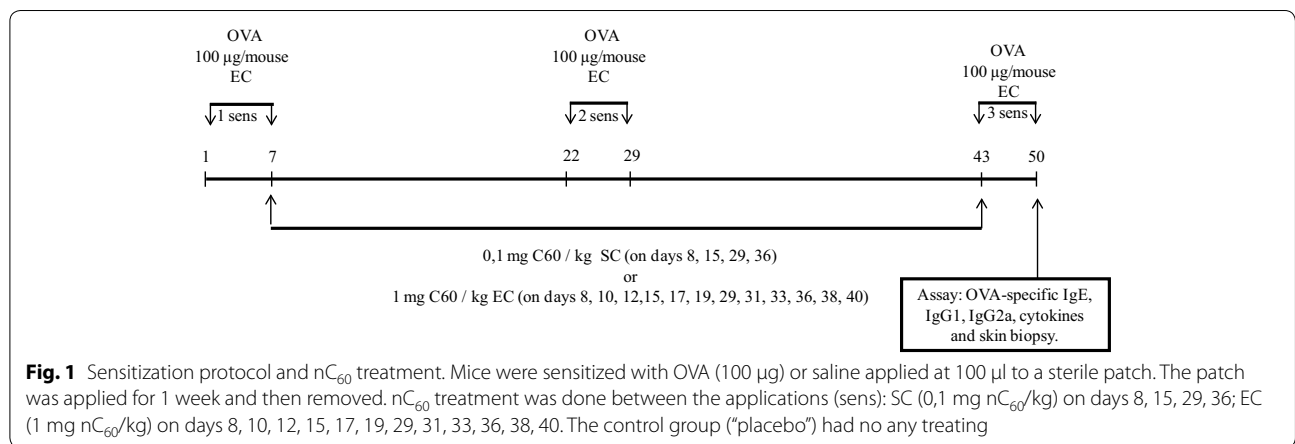
enables to sterilize the C_{60} solution prior to its administration into the body. In this study, concentration of C_{60} in the stock nC_{60} was 120 $\mu\text{g/ml}$.

nC_{60} modulates allergen-specific antibody production

To induce a mouse AD [38, 43, 44], female BALB/c mice were EC sensitized by skin application with 100 μg doses of OVA in PBS (1 mg/ml) during three one-week exposures with two-week intervals as described in “Materials and methods” and showed in Fig. 1 (“AD” group). Mice received nC_{60} by both EC (“ nC_{60} EC”) and SC (“ nC_{60} SC”) administration routes in doses 1 mg/kg and 0.1 mg/kg of nC_{60} , respectively, between OVA-applications. Control group received PBS-application only (“placebo”). Figure 2 shows that OVA-specific IgE and IgG1 levels in sera were elevated in OVA-sensitized mice (“AD” group). The

main question was whether nC_{60} treatment could change OVA-specific antibody level in OVA-sensitized mice.

To assess the effect of nC_{60} -treatment on specific IgE and IgG production sera were collected after 2-nd and 3-rd sensitizations. As can be seen from Fig. 2a, specific IgE concentrations in OVA-sensitized mice were reduced after nC_{60} treatment. SC administration of nC_{60} had stronger inhibition effect on IgE production compared with the EC one. This tendency was observed both after the 2-nd and 3-rd OVA-sensitizations (2 and 3 sens, Fig. 2). After the last sensitization the reduction of IgE concentration was about 50 % (statistically significant). OVA-specific IgG1 and IgG2a levels and IgG1/IgG2a ratio as markers for Th2 and Th1 responses [45] were measured for each group, and results (Fig. 2b) suggest that the nC_{60} treatment had no statistically significant



effect on the specific antibody production (although a decreasing trend, especially after 3-rd sensitization was observed).

Fullerene-induced cytokine profile alterations

Analysis of cytokine expression was carried out after the 3-rd sensitization in supernatants of OVA-stimulated mouse splenocytes (3 sensitization, Fig. 1). It was shown that the nC₆₀ exerts a strong effect as the IL-4 concentration was decreased by 2.5 times compared with “AD” group regardless of the route of administration (Fig. 3a). The same effect we also observed for IL-5. In this case, a slightly stronger effect was observed via EC application (Fig. 3b).

Figure 3c shows that the IL-12 concentration was significantly higher in group “nC₆₀ EC” and “placebo” compared with “AD” and “nC₆₀ SC” group. IFN-γ expression was significantly increased in nC₆₀ group compared with

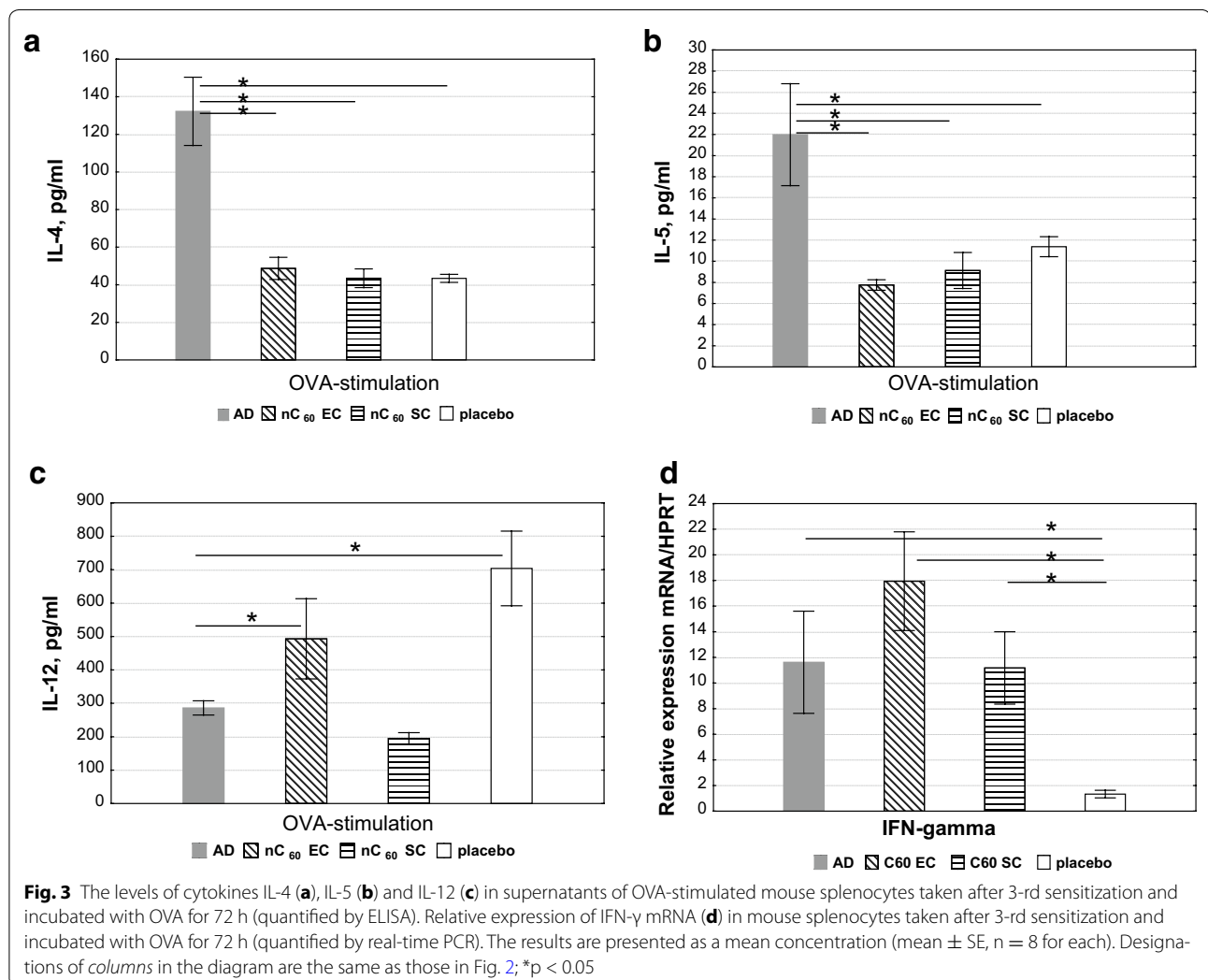
“AD” and normal-control (“placebo”) Fig. 3d. Moreover, the strongest effect was observed after EC/nC₆₀ treatment.

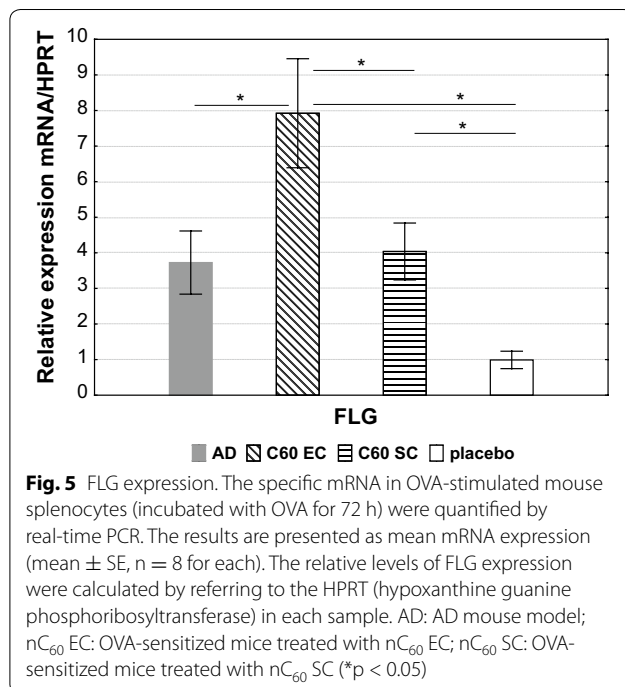
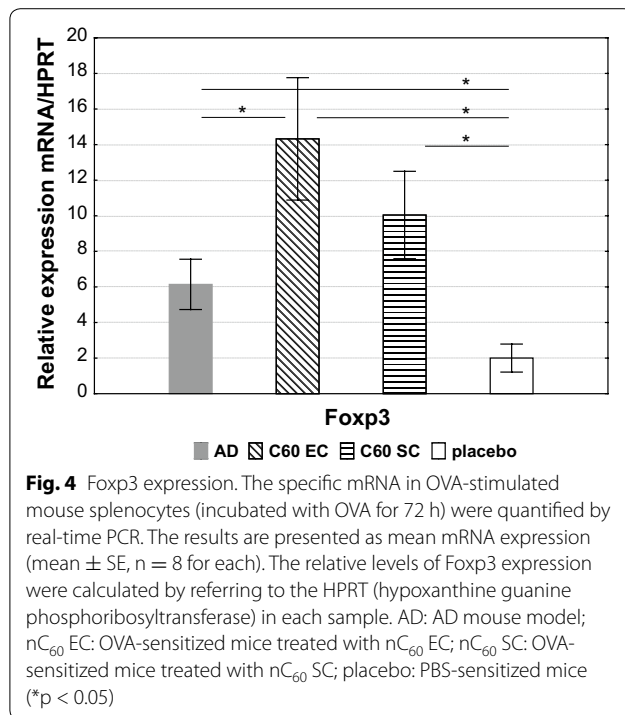
Foxp3 + regulatory cells (Foxp3 + Tregs) induction

In this study, we have measured the Foxp3 expression in OVA-stimulated mouse splenocytes to evaluate the level of Foxp3 + Tregs after the nC₆₀ treatment. As we can see a significant increase in Foxp3 expression was observed in the “nC₆₀ EC” group compared with “AD” and “placebo” (Fig. 4). The Foxp3 level was only slightly higher in group “nC₆₀ SC” than in “AD”.

Analysis of flaggrin expression

We used a quantitative real-time PCR to evaluate the effect of nC₆₀ on expression the FLG in AD-induced mice (Fig. 5). It was shown that the nC₆₀ potently promoted the FLG expression. The most significant increase of FLG





was observed at EC/nC₆₀ application, in this case, FLG expression was increased about 3 times (p < 0.05).

Histological assay

Skin biopsies from sensitized mice were taken after the final 3-rd sensitization. Skin sections were stained with

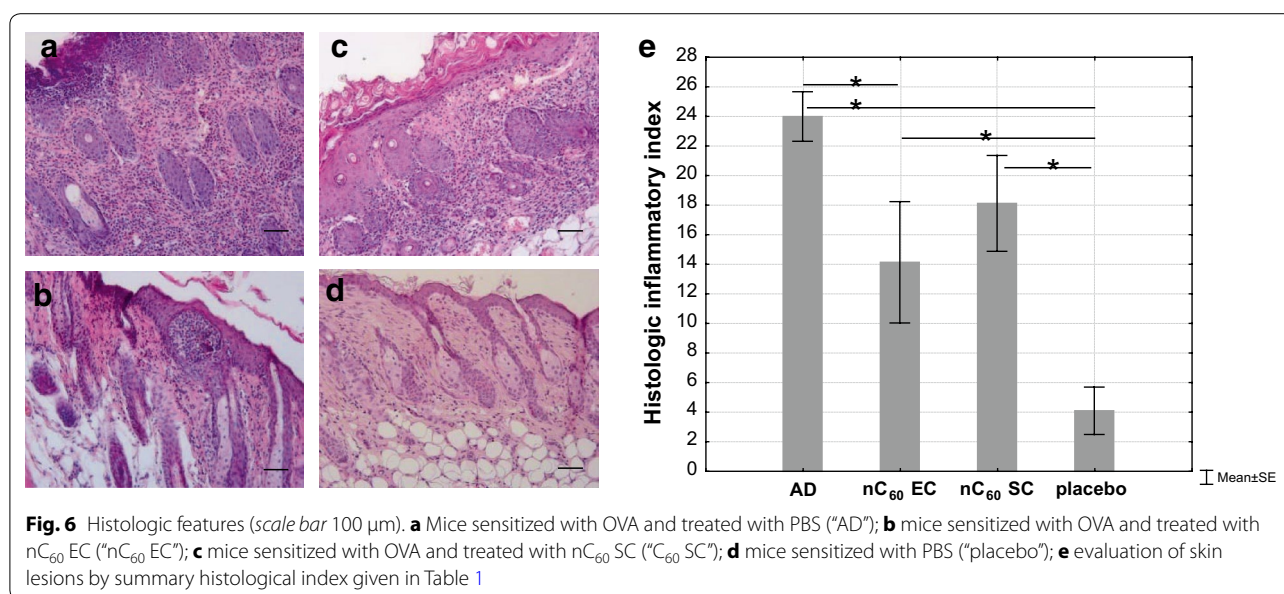
H&E and examined at 100-fold magnification. The skin samples from the “AD” group had dermal and epidermis necrosis from severe to medium degree. There were visible marked epidermis hyperplasia, pronounced diffuse mixed leukocyte infiltrates with eosinophils prevalence both in dermis and epidermis (Fig. 6a). The cellular infiltrate consisted of neutrophils, eosinophils, and lymphocytes, while the group “placebo” did not present any pathologic changes showing almost normal appearance (Fig. 6d). The infiltration degree of leukocytes in dermis and epidermis were decreased in SC/nC₆₀-treated mice (“nC₆₀ SC”) compared with “AD”, however, the eosinophils amount was the same as in the “AD” (Fig. 6c). In this case, we observed decrease in the epidermis necrosis and destructive hemorrhage in derma. There was a moderate epidermal hyperkeratosis obviously as a compensatory protection mechanism against damage. Figure 6b shows that the most pronounced nC₆₀ therapeutic effect was in the case of EC nC₆₀-treated mice (“nC₆₀ EC”). The leukocytes infiltration degree and the eosinophils number were significantly reduced in “nC₆₀ EC” group compared with “AD” and “nC₆₀ SC”. It was shown, that the epidermal necrosis, destructive hemorrhage in dermis and hyperkeratosis either were absent or were mild.

We evaluated the histologic pictures in semi-quantitative histological index (score) comparing nC₆₀-treated and non-treated groups with each other based on the microscopic features. Epidermal thickening, epidermal necrosis, epidermal hyperkeratosis, dermal and subcutaneous fat necrosis, swelling, hemorrhage, and cell infiltration of dermis and subcutaneous fat were the main evaluation criteria for the histologic skin lesions (Table 2). Each parameter had a degree of manifestation, estimated notional value and appropriate score.

Figure 6e shows that the maximum inflammatory index was observed in the “AD” (score = 24) while the “placebo” shows the value of 4. The inflammatory indexes for groups “nC₆₀ EC” and “nC₆₀ SC” were 14 and 18, respectively. Thus, the skin samples histological analysis showed that the nC₆₀ therapy improved histological picture reducing an allergic inflammation by approximately 42 and 25 %, respectively for EC and SC applications compared with the untreated mice.

Discussion

The investigation of fullerene C₆₀ biological effects has attracted increasing attention in recent years. An important issue is that a crystalline fullerene C₆₀ is practically insoluble in an aqueous medium without special processing. In this study, an aqueous solution of the fullerene was prepared by novel dialysis method [46]. Based on the spectral and elemental analyzes, we can speculate that process underlying the C₆₀ solubilization can involve a



formation of the C₆₀ molecule complexes or their clusters with NMP followed by partial hydroxylation of nanoparticle surface that is capable of stabilizing the nC₆₀ aggregates. Apparently, these aggregates are surrounded by firmly bound water envelope; the water is not removed on drying under high vacuum. We suppose, that the bond between C₆₀ and NMP has definitely non-covalent character (donor–acceptor bond) as evidenced by experimental results and theoretical calculations [47]. It should be noted that in course of time (>6 months), we have observed a decrease in the heterogeneity of nanoparticles size, they have become more uniform in size (100 nm). Perhaps, it reflects the establishment of a dynamic equilibrium in a system where a process of fullerene molecules distribution from cluster to cluster takes place.

We suggest that a new method for the nC₆₀ preparing as a biocompatible process is potentially very suitable for medical use. The relationship between basic physicochemical characteristics and toxic effects (in vivo) of nC₆₀ practically have not been covered in scientific literature. The relationship between zeta potentials and size of functionalized C₆₀ aggregates and their influence on the toxicity against bacterial cells was recently described. It was shown, that an increase of surface charge (indifferently, + or –) always leads to a decrease in the size of nanoparticles, but the toxicity is always associated with positively charged C₆₀ aggregates [48]. Nanoparticles in the nC₆₀ always bear a negative charge. In other studies, dimensional effects have been shown to affect the cytotoxicity of nC₆₀. Authors speculate that this effect may be associated with cell-contacting surface of nanoparticles. The size of nanoparticles is negatively correlated with their toxicity [49]. The question

of C₆₀ water-soluble form toxicity is closely linked to the nanoparticle morphology and its surface chemistry, which is also determined by the method of its preparation. The surface of the nC₆₀ particles differs from those described in the literature, in view of the presence of complexed hydrophilic component, NMP, and partial hydroxylation [46]. Our studies on safety of the nC₆₀ upon acute intravenous administration to BALB/c mice demonstrated that no lethal outcomes were observed and the body weight was increased in a pattern similar to the control group (unpublished observations). Exact dose for humans will be defined at the stage of clinical trials only.

Some researchers have found that fullerene derivatives, as fullerenol and amino-fullerene, inhibit in vitro the IgE-dependent degranulation of mast cells, secretion of Th2 cytokines and prostaglandins in response to an allergen stimulation that appears to be, in part, through the cellular ROS levels inhibition [23]. Certain fullerene derivatives have been able to prevent the development of inflammation and edema in mice after administration of phorbol-myristate-acetate (PMA) [24]. The inhibition has been shown to depend on the structure of addends attached to carbon skeleton. For example, fullerene C₇₀, containing 4 glycolic acid molecules, noticeably inhibited artificially induced anaphylaxis, and at the same time, fullerene with 4 attached inositol molecules showed no activity. It was shown that the fullerene material in contact with serum medium or in the cells is capable of binding to albumin and other proteins, including some enzymes. Thus, perhaps it modulates both enzymatic and signaling redox processes in the cell, but available data are very limited [25].

Table 2 The main assessment criteria for histological skin lesions

No.	Criterion	The degree of manifestation	Score
1	Epidermal thickening	Absent	0
		Mild	1
		Moderate	2
		Pronounced	3
2	Epidermal necrosis	Absent	0
		Present	1
3	Epidermal hyperkeratosis	Absent	0
		Present	1
4	Connective tissue like dermal proliferation	Absent	0
		Mild	1
		Moderate	2
		Pronounced	3
5	Dermal necrosis	Absent	0
		Present	1
6	Dermal swelling	Absent	0
		Present	1
7	Dermal hemorrhage	Absent	0
		Present	1
8	Connective tissue like subcutaneous fat proliferation	Absent	0
		Mild	1
		Moderate	2
		Pronounced	3
9	Subcutaneous fat necrosis	Absent	0
		Present	1
10	Subcutaneous fat swelling	Absent	0
		Present	1
11	Subcutaneous fat hemorrhage	Absent	0
		Present	1
12	Cell infiltration of dermis	Absent	0
		Mild	1
		Moderate	2
		Pronounced	3
13	Cell infiltration of dermis (eosinophils)	Absent	0
		Mild	1
		Moderate	2
		Pronounced	3
14	Cell infiltration of dermis (polynuclear leukocytes)	Absent	0
		Mild	1
		Moderate	2
		Pronounced	3
15	Cell infiltration of dermis (lymphocytes)	Absent	0
		Mild	1
		Moderate	2
		Pronounced	3

Table 2 continued

No.	Criterion	The degree of manifestation	Score
16	Cell infiltration of subcutaneous fat	Absent	0
		Mild	1
		Moderate	2
		Pronounced	3
17	Cell infiltration of subcutaneous fat (eosinophils)	Absent	0
		Mild	1
		Moderate	2
18	Cell (polynuclear leukocytes) infiltration of subcutaneous fat	Pronounced	3
		Absent	0
		Mild	1
		Moderate	2
19	Cell (lymphocytes) infiltration of subcutaneous fat	Pronounced	3
		Absent	0
		Mild	1
		Moderate	2
		Pronounced	3

Earlier, based on a DTH mouse model, it was shown that the fullerene treatment significantly attenuated the footpad inflammation compared with DTH-control and switched the cytokine balance towards Th1-dominance [50]. Later we have also shown that Th1 cytokines production was significantly suppressed, however it was intriguing that Th2 cytokines IL-4 and IL-5 were also significantly suppressed by a fullerene treatment [31].

These studies were undertaken in order to evaluate the OVA-induced Th1 and Th2 cytokine secretion and to determine whether the treatment with nC₆₀ can shift Th2 to Th1 response and modulate the proinflammatory cytokine production. Th2 cells produce IL-4 and IL-5 that have a prominent role in immediate-type hypersensitivity and apparently are involved in the initial stages of AD.

The level of Th1 cytokines, including IL-12 and IFN- γ was also examined. IL-12, key factor of T-cell differentiation into Th1 cells, plays a dominant role in many inflammatory diseases like AD and stimulates the IFN- γ production [51]. Thus, we demonstrated that therapeutic treatment of AD mice with nC₆₀ shifts immune response from Th2 to Th1 for example changing the IgG1/IgG2a ratio as markers for Th2 and Th1 responses. We do not know the exact mechanism, but one can assume that this phenomenon is associated with the action of the nC₆₀ on a redox homeostasis. ROS and other redox active molecules fulfill key functions in immunity and Th1/Th2 shift

which appears to be crucially dependent on the activation of redox-sensitive signaling cascades [52].

Immune regulation and tolerance are essential functions of the immune system to prevent and limit harmful immune responses to self- and non-self-antigens. CD4⁺/CD25⁺ regulatory T cells (Tregs) [53] represent a unique lineage of immunoregulatory cells in both humans and animals and play a central role in the maintenance of immunologic self-tolerance and are involved in the release of anti-inflammatory cytokine IL-10 [54]. Lack of Foxp3 + CD4⁺CD25⁺ T cells leads to immune dysregulation and affected patients often have AD-like skin lesions, increased IgE levels, and enhanced Th2 responses. However, conflicting results regarding the numbers and functions of Tregs in AD have been reported. Some investigators [55] demonstrated the absence of Foxp3 + Tregs in patient's skin that suggests a dysregulation in process of inflammation. In contrast, other authors [56] showed the elevated number of circulating CD4⁺CD25⁺ Tregs with a normal suppressive function in patients with AD. However, they used the CD25 molecule expression only (without Foxp3) as a marker for Tregs.

This experiment clearly shows that the EC treatment by fullerene leads to increased expression of Foxp3 and may shed light on the mechanism of nC₆₀ therapeutic effect.

The weakening of the skin barrier function in patients with mutations in filaggrin gene (FLG) probably promotes increased penetration of allergens by transdermal route. There is a direct interrelation between the AD and nonsense mutations in a filaggrin encoding gene [3]. Hence, one of possible therapeutic strategy to regulate AD is upregulating the FLG expression [57]. Recently, Otsuka et al. have screened more than 1000 compounds in a bioactive chemical library to find candidates to stimulate FLG mRNA expression and have revealed the compound JTC801 promoted the FLG mRNA and protein expression in vitro and in vivo. Potential utility of such therapy is indicated by the fact that a modest 20 % increase in filaggrin copy number leads to the 40 % reduction in AD susceptibility [25, 58].

Based on histological data and the cytokine secretion profile switch from Th2 to Th1 pattern we can conclude that fullerene nC₆₀ has significant anti-allergic and anti-inflammatory activity. We observed a significant suppression of IgE and Th2 cytokines (IL-4 and IL-5) production, and with a concomitant increase of Th1 cytokines production: IL-12 (at EC application only) and IFN- γ . However, there was a difference in nC₆₀ effect depended on the administration route. More intense specific IgE and Th2 cytokines suppression were observed at the EC application. In addition, this treatment significantly increased the IL-12 level compared with "AD". Based on this facts, we can hypothesize that nC₆₀ reduces AD

inflammation by activating the IFN- γ production and Th2 response suppression [58]. These results suggest that nC₆₀ might be used as an agent to suppress proinflammatory cytokine production.

Since AD is a chronic disease, apparently the barrier dysfunction is a leading primary cause of AD. Surprisingly, it turned out that the nC₆₀ quite markedly increases the filaggrin expression in vivo (Fig. 5). We could not find any scientific publications on the effect of fullerene on filaggrin gene activation except the data from presentation of "Vitamin C60 BioResearch Corporation", where C₆₀/PVP complex ("Radical Sponge") increased FLG expression in RS-treated cells control cells about a four times (<http://www.novac60.com/wp-content/uploads/2013/10/VC60-Fullerene-skin-barrier-effect.pdf>). However, these data once again point out the potential of fullerene C₆₀ as a stimulator of the filaggrin production.

Histological analysis revealed that positive therapeutic effect was achieved both at EC and SC nC₆₀-treatment, but the former was more effective. The main dermal inflammatory response component was an eosinophilic infiltration and a pronounced reduction of eosinophils number was observed in "nC₆₀ EC" group. It should be noted that this result correlated with the decrease in the IL-5 concentration in the same group combined with the regenerative processes in the skin as opposed to "AD".

Thus, in this study we have demonstrated that nC₆₀ application inhibits the inflammatory process and may represent a perspective therapeutic approach to control allergic inflammation. However further studies are needed to understand the mechanism of fullerene activity.

Conclusions

We have found that the nC₆₀ inhibits significantly specific IgE production in mouse AD model. In addition, it was shown that IL-4, IL-5 levels were significantly decreased after EC and SC C₆₀-treatment. It was observed that EC C₆₀-treatment shifts immune response from Th2 to Th1, markedly increasing the production of IL-12 and IFN- γ . We have also revealed that the use of nC₆₀ in EC route increases Foxp3 and FLG expression. Thus, simultaneous increase of Foxp3 and filaggrin expression leads to reduction in AD susceptibility. The histological analysis of skin samples showed that the nC₆₀ therapy improved histological picture reducing an allergic inflammation via EC as well as SC applications compared with the untreated mice. We do not know exactly why EC application is more effective than SC one. The possible explanation is that EC application with nC₆₀ increases the fullerene availability to the immune system due to the presence of a large amounts of immune cells in the skin with allergic inflammation (AD).

Thus, the use of nC₆₀ in form of EC application is a promising alternative therapeutic approach to control allergy inflammation. Exact dose for humans will be defined at the stage of clinical trials only.

Methods

Reagents

Ovalbumin (OVA) (Grade V, 99 %) and L-alanine were purchased from Sigma-Aldrich (USA). N-methylpyrrolidone (NMP, 99 %) was from Panreac (Spain). Crystalline fullerene C₆₀ was purchased from SES Research (99.9 %, catalog 600–9969, USA).

nC₆₀ preparation

Aqueous fullerene dispersion, nC₆₀, was obtained by method described earlier [39]. Briefly, 20 mg of C₆₀ were dissolved in 25 ml of N-methylpyrrolidone (magnetic stirrer) and resulting dark brown-purple solution was mixed with solution of 40 mg L-alanine in 100 ml of deionized water. The obtained dark-red transparent solution was stirred for 1 h and then subjected to exhaustive dialysis (cut off 10/50 kDa) against deionized water. Final dialysis solution was filtered through 0.45 mm nitrocellulose membrane resulting in a clear transparent solution with brownish-yellow color with concentration 120 µg/ml.

Mice sensitization (AD model) and nC₆₀ treatment

Female BALB/c mice ages 4–6 weeks were purchased from the animal nursery Filial SCBMT “Stolbovaya” (Moscow region, Russia) and kept in a pathogen-free environment with an OVA-free diet. All experimental procedures were carried out according to order no. 708 of the Ministry of Health of the Russian Federation and “Regulations on the ethical attitudes to laboratory animals NRC Institute of Immunology FMBA of Russia (Moscow, Russia)”.

EC sensitization of mice to induce a skin inflammation was carried out as described by Spergel et al. (1998). Briefly, mice were shaved with an electric razor. OVA (100 µg) in PBS (100 µl) or PBS as placebo was placed on a 1 × 1 cm² patch of sterile gauze, which was then secured onto the skin with a transparent biocclusive dressing (Systagenix Wound Management Limited, United Kingdom). The patch was applied thrice over a 1-week period. An inspection at the end of each sensitization period confirmed that the patch remained in place. C₆₀ treatment was carried out by subcutaneous (SC; 0.1 mg/kg) and epicutaneous nC₆₀ (EC; 1 mg/kg) administrations, between OVA-applications as shown in Fig. 1.

Antibody and cytokine assay

The anti-OVA IgE, IgG1, and IgG2a antibodies levels in sera obtained before, during, and after nC₆₀-treatment

were detected by ELISA (ELISA kits from BD, USA) according to the manufacturer’s protocol. OVA was used for coating the plates. Mouse anti-ovalbumin IgE mAb (AbD Serotec, UK) and biotin rat anti-mouse IgE (BD) were used for detection anti-OVA mouse IgE. These components were used to construct the calibration curve and then to analyze sera.

The spleens were taken after the last allergen application, and levels of IL-4, IL-5, and IL-12 (p40) in supernatants of OVA-stimulated splenocytes were determined by using ELISA [Duo-Set from R&D Systems (UK) and ELISA set from BD (USA)] according to the manufacturer’s protocol.

Real-time PCR

The total RNA from OVA-stimulated mice splenocytes was extracted using the RNeasy Mini Kit (Qiagen, Courtaboeuf, France) according to the manufacturer’s instructions. The RNA concentration was determined, and cDNA was synthesized through a reverse transcription reaction (“Reverta-L”, Interlabservice, Russia). Quantitative real-time PCR analysis of mRNA expression was done by the iQ5 system (Bio-Rad, USA) and the PCR-Mix kit (Sintol, Russia).

The results are presented as mRNA expression (Foxp3, ILG). Calculations to determine the relative level of gene expression were made using the comparative C_t method (ΔΔC_t) referring to the mHPRT in each sample; the results are presented as arbitrary units.

Histological analysis

The skin specimens from patch areas were removed for histologic examination immediately after the last EC application with OVA. Skin biopsies were taken from similar body sites, fixed overnight with 10 % paraformaldehyde at 4 °C, and embedded in paraffin. Four-micrometer sections were stained with hematoxylin and eosin (H&E). The histological preparations were analyzed under a light microscope (Leica DM2000, Germany) with 50×, 100×, and 400× lenses.

Statistical analysis

The data are shown as mean ± SE. Statistical analysis was done with the program Statistica 8.0 (StatSoft Inc., USA). The significance of the results was determined by using Student’s t test. Differences were considered significant at p < 0.05. The Quantitative RT-PCR data were calculated by using the comparative C_t method (ΔΔC_t).

Authors’ contributions

NS, EB, IS, AN carried out the molecular genetic studies and the immunoassays. DP have prepared the C₆₀ aqueous solution. OK carried out the histological analysis. NS, SA, MK have made substantial contributions to conception, design, analysis, interpretation of data and given final approval of the version to be published. All authors read and approved the final manuscript.

Acknowledgements

We would like to thank Dr. Svetlana Korobova for critical reading of the manuscript. This work was supported by the Ministry of Science and Education of the Russian Federation, the Federal Target Program "Research and development on priority directions of scientific-technological complex of Russia in 2014–2020" (agreement 14.604.21.0059, the unique identifier RFMEFI60414X0059).

Competing interests

The authors declare that they have no competing interests.

Received: 3 November 2015 Accepted: 13 January 2016

Published online: 25 January 2016

References

- Bieber T. Atopic dermatitis. *N Engl J Med*. 2008;358:1483–94.
- Bieber T, Novak N. Pathogenesis of atopic dermatitis: new developments. *Curr Allergy Asthma Rep*. 2009;9:291–4.
- Palmer CN, Irvine AD, Terron-Kwiatkowski A, Zhao Y, Liao H, Lee SP, et al. Common loss-of-function variants of the epidermal barrier protein filaggrin are a major predisposing factor for atopic dermatitis. *Nat Genet*. 2006;38:441–6.
- Kezic S, Novak N, Jakasa I, Jungersted JM, Simon M, Brandner JM, et al. Skin barrier in atopic dermatitis. *Front Biosci*. 2014;19:542–56.
- Arkwright PD, Motala C, Subramanian H, Spergel J, Schneider LC, Wollenberg A, et al. Management of difficult-to-treat atopic dermatitis. *J Allergy Clin Immunol Pract*. 2013;1:142–51.
- Shershakova NN, Babakhin AA, Elisyutina OG, Khaitov MR. Atopic dermatitis: experimental models for study of pathogenesis and development new methods of treatment. *RAG*. 2011;6:3–11 (in Russian).
- Garmhausen D, Hagemann T, Bieber T, Dimitriou I, Fimmers R, Diepgen T, et al. Characterization of different courses of atopic dermatitis in adolescent and adult patients. *Allergy*. 2013;68:498–506.
- Bieber T. Atopic dermatitis. *Ann Dermatol*. 2010;22:125–37.
- Bonness S, Bieber T. Molecular basis of atopic dermatitis. *Curr Opin Allergy Clin Immunol*. 2007;7:382–6.
- Schlapbach C, Simon D. Update on skin allergy. *Allergy*. 2014;69:1571–81.
- Nakai K, Yoneda K, Maeda R, Munehiro A, Fujita N, Yokoi I, et al. Urinary biomarker of oxidative stress in patients with psoriasis and atopic dermatitis. *J Eur Acad Dermatol Venerol*. 2009;23:1405–8.
- Kapun AP, Salobir J, Levart A, Kotnik T, Svetec AN. Oxidative stress markers in canine atopic dermatitis. *Res Vet Sci*. 2012;92(3):469–70.
- Koren Carmi I, Haj R, Yehuda H, Tamir S, Reznick AZ. The Role of oxidation in FSL-1 induced signaling pathways of an atopic dermatitis model in hHaCaT keratinocytes. *Adv Exp Med Biol*. 2015;849:1–10.
- Okayama Y. Oxidative stress in allergic and inflammatory skin diseases. *Curr Drug Targets Inflamm Allergy*. 2005;4(4):517–9.
- Niwa Y, Sumi H, Kawahira K, Terashima T, Nakamura T, Akamatsu H. Protein oxidative damage in the stratum corneum: evidence for a link between environmental oxidants and the changing prevalence and nature of atopic dermatitis in Japan. *Br J Dermatol*. 2003;149:248–54.
- Bito T, Nishigori C. Impact of reactive oxygen species on keratinocyte signaling pathways. *J Dermatol Sci*. 2012;68(1):3–8.
- Gharbi N, Pressac M, Hadchouel M, Szwarc H, Wilson SR, Moussa F. [60] Fullerene is a powerful antioxidant in vivo with no acute or subacute toxicity. *Nano Lett*. 2005;5:2578–85.
- Hendrickson OD, Morozova OV, Zherdev AV, Yaropolov AI, Klochkov SG, Bachurin SO, Dzantiev BB. Study of distribution and biological effects of fullerene C60 after single and multiple intragastrical administrations to rats. *Fuller Nanotub Carbon Nanostruct*. 2015;23(7):658–68.
- Baati T, Bourasset F, Gharbi N, Njim L, Abderrabba M, Kerkeni A, Szwarc H, Moussa F. The prolongation of the lifespan of rats by repeated oral administration of 60 fullerene. *Biomaterials*. 2012;33:4936–46.
- Li W, Chen C, Ye C, Wei T, Zhao Y, Lao F, Chen Z, Meng H, Gao Y, Yuan H, Xing G, Feng Z, Chai Z, Zhang X, Yang F, Han D, Tang X, Zhang Y. The translocation of fullerene nanoparticles into lysosome via the pathway of clathrin-mediated endocytosis. *Nanotechnology*. 2008;19(14):145102.
- Andreev I, Petrukhina A, Garmanova A, Babakhin A, Andreev S, Romanova V, Troshin P, Troshina O, DuBuske L. Penetration of fullerene C60 derivatives through biological membranes. *Fuller Nanotub Carbon Nanostruct*. 2008;16:89–102.
- Andreev SM, Babakhin AA, Petrukhina AO, Romanova VS, Parnes ZN, Petrov RV. Immunogenic and allergic properties of fullerene conjugates with amino acid and proteins. *Dokl Biochem*. 2000;370:4–7.
- Ryan JJ, Bateman HR, Stover A, Gomez G, Norton SK, Zhao W, Schwartz LB, Lenk R, Kepley CL. Fullerene nanomaterials inhibit the allergic response. *J Immunol*. 2007;179:665–72.
- Dellinger A, Zhou Z, Lenk R, MacFarland D, Kepley CL. Fullerene nanomaterials inhibit phorbol myristate acetate-induced inflammation. *Exp Dermatol*. 2009;18:1079–81.
- Norton SK, Dellinger A, Zhou Z, Lenk R, MacFarland D, Vonakis B, Conrad D, Kepley CL. A new class of human mast cell and peripheral blood basophil stabilizers that differentially control allergic mediator release. *Clin Trans Sci*. 2010;3:158–69.
- Magoulas GE, Garnelis T, Athanassopoulos CM, Papaioannou D, Matheolabakis G, Avgoustakis K, Hadjipavlou-Litina D. Synthesis and antioxidative/anti-inflammatory activity of novel fullerene–polyamine conjugates. *Tetrahedron*. 2012;68(35):7041–9.
- Andrievsky GV, Kosevich MV, Vovk OM, Shelkovsky VS, Vashchenko LA. On the production of an aqueous colloidal solution of fullerenes. *J Chem Soc*. 1995;12:1281–2.
- Bunz H, Plankenhorn S, Klein R. Effect of buckminsterfullerenes on cells of the innate and adaptive immune system: an in vitro study with human peripheral blood mononuclear cells. *Intern J Nanomed*. 2012;7:4571–80.
- Babakhin AA, Andrievsky G, DuBuske LM. Inhibition of systemic and passive cutaneous anaphylaxis by water-soluble fullerene C60. *J Allergy Clin Immunol*. 2009;123(2):118.
- Bashkatova YeN, Andreev SM, Shershakova NN, Babakhin AA, Shilovsky IP, Khaitov MR. Study of modulatory activity of fullerene C60 derivatives on DTH reaction. *Phiziologiya i patologiya immunnoy sistemy*. 2012; 2:17–27 (in Russian).
- Bashkatova E, Shershakova N, Shilovskii I, Babakhin A, Andreev S, Khaitov M. Fullerene adducts attenuate delayed-type hypersensitivity reactions induced in mice. *Allergy*. 2013;68:442.
- Yanagisawa R, Takano H, Inoue KI, Koike E, Sadakane K, Ichinose T. Size effects of polystyrene nanoparticles on atopic dermatitis-like skin lesions in NC/NGA mice. *Int J Immunopathol Pharmacol*. 2010;23(1):131–41.
- Ilves M, Palomäki J, Vippola M, Lehto M, Savolainen K, Savinko T, Alenius H. Topically applied ZnO nanoparticles suppress allergen induced skin inflammation but induce vigorous IgE production in the atopic dermatitis mouse model. *Part Fibre Toxicol*. 2014;11:38. doi:10.1186/s12989-014-0038-4.
- Wiesenthal A, Hunter L, Wang S, Wickliffe J, Wilkerson M. Nanoparticles: small and mighty. *Int J Dermatol*. 2011;50(3):247–54. doi:10.1111/j.1365-4632.2010.04815.x.
- Rancan F, Gao Q, Graf C, Troppens S, Hadam S, Hackbarth S, Kembangan C, Blume-Peytavi U, Rühl E, Lademann J, Vogt A. Skin penetration and cellular uptake of amorphous silica nanoparticles with variable size, surface functionalization, and colloidal stability. *ACS Nano*. 2012;6(8):6829–42. doi:10.1021/nn301622h.
- Hussain Z, Katas H, Mohd Amin MC, Kumolosas E, Sahudin S. Downregulation of immunological mediators in 2,4-dinitrofluorobenzene-induced atopic dermatitis-like skin lesions by hydrocortisone-loaded chitosan nanoparticles. *Int J Nanomed*. 2014;9:5143–56. doi:10.2147/IJN.S71543.
- Kang MJ, Eum JY, Jeong MS, Park SH, Moon KY, Kang MH, Kim MS, Choi SE, Lee MW, Do Lee I, Bang H, Lee CS, Joo SS, Li K, Lee MK, Seo SJ, Choi YW. Tat peptide-admixed elastic liposomal formulation of hirsutenone for the treatment of atopic dermatitis in NC/Nga mice. *Int J Nanomed*. 2011;6:2459–67. doi:10.2147/IJN.S24350.
- Shershakova NN, Babakhin AA, Bashkatova EN, Kamyshnikov OY, Andreev SM, Shilovsky IP, et al. Allergen-specific immunotherapy of experimental atopic dermatitis. *Immunologia*. 2014;35(3):155–60 (in Russian).
- Andreev SM, Purgina DD, Bashkatova EN, Garshev AV, Maerle AV, Khaitov MR. Facile preparation of aqueous fullerene C60 nanodispersions. *Nanotechnol Russ*. 2014;9(7–8):369–79.

40. Deguchi SH, Mukai SA. Top-down preparation of dispersions of C60 nanoparticles in organic solvents. *Chem Lett*. 2006;35(4):396–3974.
41. Mchedlov-Petrosyan NO. Fullerene C₆₀ solutions: colloid aspect. *Chem Phys Technol Surf*. 2010;1(1):19–37.
42. Opinion on NMP. Scientific Committee on Consumer Safety. 2011. SCCS/1413/11. http://ec.europa.eu/health/scientific_committees/consumer_safety/docs/sccs_o_050.pdf.
43. Spergel JM, Mizoguchi E, Brewer JP, Martin TR, Bhan AK, Geha RS. Epicutaneous sensitization with protein antigen induces localized allergic dermatitis and hyperresponsiveness to methacholine after single exposure to aerosolized antigen in mice. *J Clin Invest*. 1998;101:1614–22.
44. Shershakova N, Bashkatova E, Babakhin A, Andreev S, Nikonova A, Shilovsky I, Kamyshnikov O, Buzuk A, Elisyutina O, Fedenko E, Khaïtov M. Allergen-specific immunotherapy with monomeric allergoid in a mouse model of atopic dermatitis. *PLoS ONE*. 2015;10(8):e0135070. doi:10.1371/journal.pone.0135070.
45. Mountford AP, Fisher A, Wilson RA. The profile of IgG1 and IgG2a antibody responses in mice exposed to *Schistosoma mansoni*. *Parasit Immunol*. 1994;16(10):521–7.
46. Andreev S, Purgina D, Bashkatova E, Garshev A, Maerle A, Andreev I, Osipova N, Shershakova N, Khaïtov M. Study of fullerene aqueous dispersion prepared by novel dialysis method: simple way to fullerene aqueous solution. *Fuller Nanotub Carbon Nanostruct*. 2015;23(9):792–800. doi:10.1080/1536383X.2014.998758.
47. Karpenko OB, Trachevskij VV, Filonenko OV, Lobanov VV, Avdeev MV, Tropin TV, Kyzyma OA, Snegir SV. NMR study of non-equilibrium state of fullerene C60 in N-methyl-2-pyrrolidone. *Ukr J Phys*. 2012;8:860–3.
48. Deryabin DG, Efremova LV, Vasilchenko AS, Saidakova EV, Sizova EA, Troshin PA, Zhilenkov AV, Khakina EE. *J Nanobiotechnol*. 2015;13:50. doi:10.1186/s12951-015-0112-6.
49. Song M, Yuan S, Yin J, Wang X, Meng Z, Wang H, Jiang G. Size-dependent toxicity of nano-C₆₀ aggregates: more sensitive indication by apoptosis-related Bax translocation in cultured human cells. *Environ Sci Technol*. 2012;46:3457–64.
50. Yamashita K, Sakai M, Takemoto N, Tsukimoto M, Uchida K, Yajima H, et al. Attenuation of delayed-type hypersensitivity by fullerene treatment. *Toxicology*. 2009;261:19–24.
51. Sun L, He C, Nair L, Yeung J, Egwuagu CE. Interleukin 12 (IL-12) family cytokines: role in immune pathogenesis and treatment of CNS autoimmune disease. *Cytokine*. 2015. doi:10.1016/j.cyto.2015.01.030.
52. Gostner JM, Becker K, Fuchs D, Sucher R. Redox regulation of the immune response. *Redox Rep*. 2013;18(3):88–94.
53. Lu LF, Lind EF, Gondek DC, Bennett KA, Gleeson MW, Pino-Lagos K, et al. Mast cells are essential intermediaries in regulatory T-cell tolerance. *Nature*. 2006;442(7106):997–1002.
54. Moore KW, de WaalMalefyt R, Coffman RL, O'Garra A. Interleukin-10 and the interleukin-10 receptor. *Annu Rev Immunol*. 2001;19:683–765.
55. Verhagen J, Akdis M, Traidl-Hoffmann C, Schmid-Grendelmeier P, Hijnen D, Knol EF, et al. Absence of T regulatory cell expression and function in atopic dermatitis skin. *J Allergy Clin Immunol*. 2006;117:176–83.
56. Ou LS, Goleva E, Hall C, Leung DY. T regulatory cells in atopic dermatitis and subversion of their activity by superantigens. *J Allergy Clin Immunol*. 2004;113:756–63.
57. Otsuka A, Doi H, Egawa G, Maekawa A, Fujita T, Nakamizo S, et al. Possible new therapeutic strategy to regulate atopic dermatitis through upregulating filaggrin expression. *J Allergy Clin Immunol*. 2014;133(1):139–46.
58. Toda M, Leung DY, Molet S, Boguniewicz M, Taha R, Christodoulouopoulos P, et al. Polarized in vivo expression of IL-11 and IL-17 between acute and chronic skin lesions. *J Allergy Clin Immunol*. 2003;111:875–81.

Submit your next manuscript to BioMed Central and we will help you at every step:

- We accept pre-submission inquiries
- Our selector tool helps you to find the most relevant journal
- We provide round the clock customer support
- Convenient online submission
- Thorough peer review
- Inclusion in PubMed and all major indexing services
- Maximum visibility for your research

Submit your manuscript at
www.biomedcentral.com/submit

