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# Blood parasite infections in a wild population of ravens (*Corvus corax*) in Bulgaria

Peter Shurulinkov<sup>1</sup>, Lachezar Spasov<sup>2</sup>, Georgi Stoyanov<sup>2</sup> and Nayden Chakarov<sup>3,4\*</sup> 

## Abstract

**Background:** Blood parasites have been studied intensely in many families of avian hosts, but corvids, a particularly cosmopolitan family, remain underexplored. Haemosporidian parasites of the common raven (*Corvus corax*) have not been studied, although it is the largest, most adaptable, and widespread corvid. Genetic sequence data from parasites of ravens can enhance the understanding of speciation patterns and specificity of haemosporidian parasites in corvids, and shed light how these hosts cope with parasite pressure.

**Methods:** A baited cage trap was used to catch 86 ravens and a nested PCR protocol was used to amplify a 479 bp fragment of the haemosporidian cytochrome b gene from the samples. The obtained sequences were compared with the MalAvi database of all published haemosporidian lineages and a phylogenetic tree including all detected raven parasites was constructed. An examination of blood smears was performed for assessment of infection intensity.

**Results:** Twenty blood parasite lineages were recovered from ravens caught in a wild population in Bulgaria. The prevalence of generalist *Plasmodium* lineages was 49%, and the prevalence of *Leucocytozoon* lineages was 31%. Out of 13 detected *Leucocytozoon* lineages six were known from different corvids, while seven others seem to be specific to ravens. A phylogenetic reconstruction suggests that *Leucocytozoon* lineages of ravens and other corvids are not monophyletic, with some groups appearing closely related to parasites of other host families.

**Conclusions:** Several different, morphologically cryptic groups of *Leucocytozoon* parasites appear to infect corvids. Ravens harbour both generalist corvid *Leucocytozoon* as well as apparently species-specific lineages. The extraordinary breeding ecology and scavenging lifestyle possibly allow ravens to evade vectors and have relatively low blood parasite prevalence compared to other corvids.

**Keywords:** Avian malaria, *Leucocytozoon*, Host specificity, Corvids, Passerines, Prevalence, Tolerance, Resistance

## Background

Haemosporidian parasites of the genera *Plasmodium*, *Haemoproteus* and *Leucocytozoon* are widely distributed among birds worldwide and can cause substantial health and fitness reduction to infected hosts [1–4]. Blood parasite prevalence varies greatly among the different families of songbirds *Passeriformes*, and among different species of the same family [5, 6]. Inside the family of warblers (*Sylviidae*), the highly-parasitized members of the genus *Acrocephalus* share similar habitats with their relatives of

the genera *Locustella* and *Cettia* in which haemosporidian infections are of very low prevalence or even absent in some populations [7–9]. Such differences among the bird taxa are difficult to explain. In some cases, they can correlate with different capacities of the host immunity to defend itself against specific parasite infections [10, 11] or with the ecology, habitat preferences and behaviour of the hosts [12–14]. According to studies on different corvid species, such as crows, magpies and jays, the avian family of *Corvidae* seems to harbour a great amount of haemosporidian parasites [15–18]. However, ravens may be less infected than their relatives due to their specific ecological niche.

The common raven (*Corvus corax*) is the largest songbird and most widely distributed corvid with a range

\*Correspondence: nayden.chakarov@uni-bielefeld.de

<sup>3</sup> Department of Animal Behaviour, Bielefeld University, 33501 Bielefeld, Germany

Full list of author information is available at the end of the article

covering nearly the entire Holarctic [19, 20]. Ravens are typical scavengers, often feeding on carrion, with breeding biology resembling vultures and other raptors [21–23]. They breed very early in spring, nesting mostly on rocks in river canyons and mountains, or on trees in lowland and low mountain forests [19]. Particularly in the latter environment they are well exposed to blood parasite vectors—various groups of dipterans. It is hypothesized that scavenging birds are subject to strong parasite-mediated selection on immune defenses, which is supported by the finding that they have larger spleens and higher blood total leukocyte concentrations than non-scavengers [24].

A bird host can utilize two main strategies to cope with an infection—parasite suppression and clearance (resistance), or withstanding the infection, while paying a low fitness cost (tolerance) [25, 26]. In scavenger hosts with particularly potent immunity, parasites may not be able to develop at all. In such species, a very low prevalence of parasites or even their complete absence can be expected. Accordingly, some scavenging birds, such as griffon vultures (*Gyps fulvus*) have very low prevalence of blood parasites [27; own unpublished data]. Other species, such as corvids, harbour a high number of parasites without developing visible illness, corresponding more to a tolerance strategy. Several recent studies have detected high prevalence and high diversity of *Leucocytozoon* parasites in corvids [17, 18, 28]. These studies were completed on close relatives of ravens—crows, jays and magpies, which despite the common incorporation of carrion in their diet are less specialized scavengers compared to ravens. Nevertheless, the phylogenetic relationship between these species allows us to expect similarly high *Leucocytozoon* prevalence in raven populations. Thus, ravens may be expected to have high infection prevalence of blood parasites and low infection intensity.

Until now no haemosporidian genetic lineages from ravens have been studied or are included in MalAvi, a database that includes all published lineages of avian malaria and related blood parasites [29]. The present work, aims to study the prevalence of haemosporidian parasites and their lineage diversity in ravens from a wild population and infer the strategy of ravens to avoid detrimental fitness effects of these parasites.

## Methods

A total of 86 fully grown, adult and immature ravens were sampled in Bulgaria (Dolno Ozirovo, Vratsa district, 43.2393 N, 23.3513 E) between March and September of the years 2013–2016. Ravens were captured in big cage nets baited with carrion, measured and ringed with an ornithological steel ring and a colour ring in order to prevent double sampling. Blood samples were taken from

the brachial vein and stored in absolute ethanol. Two blood smears were prepared for each bird and fixed in methanol for 5 min. Smears were stained with Giemsa, 10,000 blood cells scanned and infection intensity score noted according to Valkiūnas [1].

DNA was extracted following a standard phenol–chloroform protocol. For the detection of the blood parasites nested PCR and light microscopy were used. Molecular identification of infections was performed with nested PCR with the primer pairs NaemNFI/NR3, followed by a second reaction with the primers HaemF/R2 and HaemFL/R2L, following the protocol of Hellgren et al. [30]. The PCR amplicons were visualized on a 2% agarose gel with ethidium bromide under UV light. All amplicons from positive samples were purified with ExoSAP and bidirectionally sequenced on an ABI 3730 Analyzer with BigDye Terminator. The cytochrome b sequences were edited and aligned with Geneious 8.1.9 and blasted against the MalAvi database, state 16.07.2017 [29].

Phylogenetic analyses was performed on a dataset of 40 lineages, 20 detected in ravens and close BLAST hits downloaded from the MalAvi dataset. The alignment was trimmed to 464 bp. Trees were constructed using a MrBayes v3.2.6 [31] with a GTR+G model and two Markov chains, run simultaneously for 10 million generations with a subsampling every 200 generations resulting in 50,000 trees. Posterior probabilities were calculated after discarding 25% of burn-in trees. Bootstrap support for individual branches was calculated using 1000 replicates in RAXML [32]. Any sequence differing by one or more nucleotides from other sequences was considered to be distinct. Distinct lineages should not be automatically interpreted as species [18]. Names of the already described parasite lineages are taken from the MalAvi database, while new lineages were designated names CCORAX1–7 (GenBank accession nos. MG209762–MG209768).

## Results

The total prevalence of haemosporidian infections in fully grown ravens determined by molecular methods was 62.8% (Table 1). The prevalence of *Plasmodium*, *Leucocytozoon* and *Haemoproteus* was 48.8% (42/86), 31.4% (27/86) and 2.3% (2/86), respectively. Mixed infections were detected in 16 occasions (29.6% of all infections found; 18.6% of all sampled birds), 15 of them were *Plasmodium/Leucocytozoon* and one was *Haemoproteus/Leucocytozoon*. A total of 20 haematozoan lineages were detected—*Plasmodium*—5, *Leucocytozoon*—13, and *Haemoproteus*—2. Seven of the *Leucocytozoon* lineages have not been reported until now and have close similarity to other corvid lineages. These lineages have been designated CCORAX1–7 (Table 1).

**Table 1 Summary of haemosporidian parasite lineages detected in ravens in Bulgaria (n = 86)**

GenBank no.	Lineage name	Nr infected	% prevalence	Closest BLAST hit	% match
<i>Plasmodium</i> lineages					
AF495571	SGS1	30	34.8		
DQ368381	GRW06	4	4.7		
AY831748	GRW11	3	3.5		
AY831749	SYAT24	3	3.5		
DQ847258	PBPIP	1	1.2		
Total <i>Plasmodium</i>		41	47.7		
<i>Haemoproteus</i> lineages					
CIRCUM05		1	1.2		
GAGLA05		1	1.2		
Total <i>Haemoproteus</i>		2	2.3		
<i>Leucocytozoon</i> lineages					
MG209762	CCORAX1	8	9.3	COCOR16	99.6
KJ128991	COCOR13	7	8.1		
MG209765	CCORAX4	2	2.3	COCOR16	99.4
JX867111	COCOR02	1	1.2		
JX867112	COCOR03	1	1.2		
KY768841	PICPIC01	1	1.2		
KJ488810	GAGLA06	1	1.2		
JX507218	EUSE1	1	1.2		
MG209763	CCORAX2	1	1.2	GAGLA06	99.8
MG209764	CCORAX3	1	1.2	COCOR02	97.9
MG209766	CCORAX5	1	1.2	COCOR09	99.8
MG209767	CCORAX6	1	1.2	COCOR16	99.0
MG209768	CCORAX7	1	1.2	CORMAC06	99.8
Total <i>Leucocytozoon</i>		27	31.4		
Total infected		54	62.8		
Not infected		32	37.2		

For lineages described for the first time in this study closest BLAST hit to any lineage from the MalAvi database and % similarity are indicated

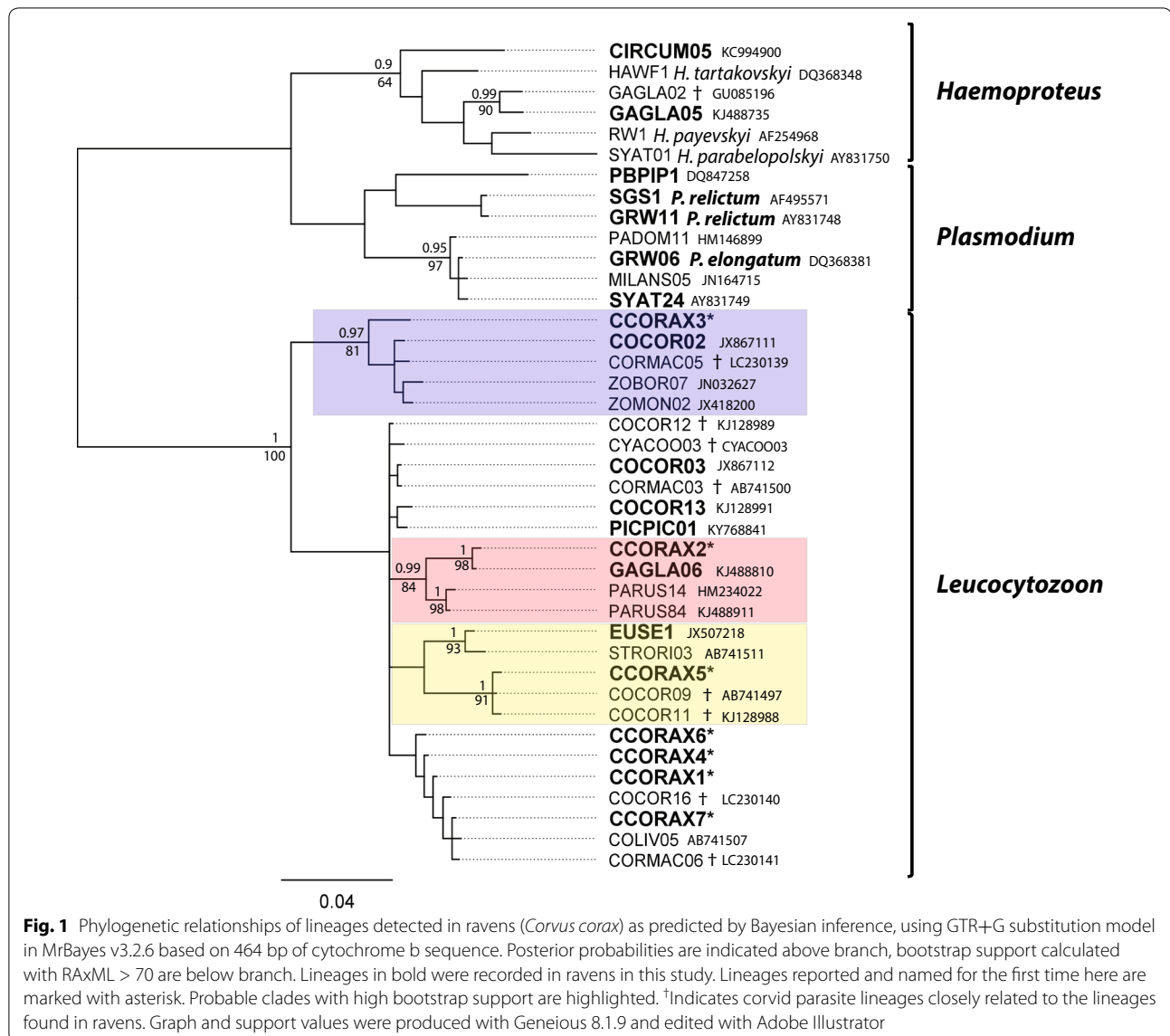
Phylogenetic relationships (Fig. 1) show two main clades inside *Leucocytozoon*—one of them including COCOR02 and CCORAX3, as well as some non-corvid lineages. A second clade contains CCORAX2 and GAGLA06, as well as two *Leucocytozoon* lineages known from blue tits. A third and fourth tentative clades, include CCORAX5, COCOR09, COCOR11, and EUSE1 as well as the lineage STRORI03, known from Oriental turtle doves, respectively. These clades may represent different species of *Leucocytozoon* infecting mostly corvids but also other passerines and doves. A branch including CCORAX1, CCORAX4, CCORAX6 and CCORAX7, recorded in the ravens, as well as COLIV05, CORMAC03, COCOR16 recorded by other authors had low support (0.81 posterior probability and 55% bootstrap support) and could not be verified as a separate clade on the bases of the sequenced cytochrome b region. Several lineages represented in ravens and other corvids such as COCOR12, COCOR13, CORMAC03, CYACOO03 and

PICPIC01 do not clearly cluster within any of the distinct clades.

All microscopically detected infections showed very low intensity, below 0.5 parasites/10,000 erythrocytes. *Leucocytozoon* infections were more common during the summer (June–September, 44.9%, 22/49) compared to the spring period (March–May, 18.5%, 5/27,  $\chi^2 = 5.289$ ,  $p = 0.021$ ). *Plasmodium* prevalence did not show such seasonal variation.

## Discussion

Ravens in Bulgaria showed relatively low prevalence of blood parasites compared to other studied corvid populations [17, 18, 33, 34]. Although all corvids are scavengers to a different degree, ravens have a very strong expression of this trait. This suggests that ravens may have a particularly potent immunity. Scavengers may have a stronger humoral immune response, potentially effective against malaria-like parasites, but not a stronger cell-mediated



immune response [24, 35]. A scavenging lifestyle has also enabled ravens to breed much earlier in spring than crows and magpies, which may allow them to evade the peak of vector emergence [19]. Additionally, raven nests on cliffs are very common at the sampling site and may be less accessible to vectors compared to the nests of other corvids.

Compared to other passerine families, ravens and other corvids have high blood parasite prevalence and are hosts of a number of specific *Leucocytozoon* lineages. This is in accordance with the data presented for three species of North American corvids (*Corvus brachyrhynchos*, *Pica nuttalli* and *Cyanocitta stelleri*) in which high infection prevalence of *Leucocytozoon* infections was reported, and 19 different lineages were found [18].

Almost all microscopically observed infections were of very low intensity, possibly chronic or even latent infections. This suggests that even immature ravens may be past the acute stage of high infection intensity, which most probably occurs in nestlings before fledging. This is the common case in raptors with similar breeding biology, where parental feeding may partly alleviate the risk of starvation during peak infection [1, 36, 37]. The low parasitaemia also suggests that the development of a strong immune response suppresses the blood parasites as ravens get older. Low intensity of infection has been shown to indicate good immune control in several bird species, such as Hawaiian amakihi and Seychelles warblers [38, 39]. During the breeding season the parasite suppression may weaken, enabling a relapse, and

transmission to new raven hosts [1]. Therefore, both resistance and tolerance are probably dynamic in time.

Among the *Plasmodium* lineages found in the studied ravens, SGS1 (lineage of *Plasmodium relictum*) and GRW6 (lineage of *Plasmodium elongatum*) are wide generalists found in large numbers of hosts [29, Malavi Database 2017]. GRW11 (lineage of *P. relictum*) is also a generalist and can infect a rather wide spectrum of hosts—mostly passerines from different families [6]. It has been found to infect corvids in Asia—Eurasian jays (*Garrulus glandarius*) in Armenia [40], jungle crows (*Corvus macrorhynchos*) in Japan [41] and hooded crows (*Corvus cornix*) in Israel [42]. In Bulgaria GRW11 has been detected in passerines of four families—Ploceidae, Fringillidae, Sylviidae and Laniidae [43–45]. The lineage SYAT24 until now was found only in blackcaps (*Sylvia atricapilla*) migrating through Spain [46, 47]. Blackcap is a common breeding and migrating bird in the area of the raven's samples origin for the present study (Dolno Ozirovo). Probably SYAT24 has been transmitted from blackcaps to the local ravens in Bulgaria. Similarly, the lineage PBPIP1 occurs mostly in African birds (local residents) and European long distance migrants—tree pipits (*Anthus trivialis*) and collared flycatchers (*Ficedula albicollis*), wintering in Africa (Malavi Database 2017). Since these migratory species regularly pass through the study area, most likely the lineage has been transmitted locally in Bulgaria to the ravens. Thus, some local mosquitoes are probable competent vectors for this *Plasmodium* lineage.

*Plasmodium* prevalence in corvids appears to follow a latitudinal gradient. This can be expected for parasites of non-migratory species, which depend on suitable climatic conditions for their vectors [1, 48]. *Plasmodium* was not found in crows of two species (*Corvus corone*, *C. macrorhynchos*) in Hokkaido, Japan [28]. Most probably this northern island lacks suitable conditions for competent vectors. However, another study from southern Japan

showed 17% prevalence of *Plasmodium/Haemoproteus* in the same two species [33]. A population of carrion crows in Germany had a *Plasmodium* prevalence of 29.5% [17]. A high prevalence of 59.6% *Plasmodium/Haemoproteus* infections has been reported for hooded crows (*C. cornix*) in Italy [34], which is even higher than the 48.8% *Plasmodium* prevalence for ravens in Bulgaria.

The *Haemoproteus* lineages GAGLA05 and CIR-CUM05, which were detected during the present study in one raven each, have been reported from jays (*G. glandarius*) sampled in Portugal and Morocco [6]. In this case, despite not appearing closely related, both haemoproteid lineages infect birds belonging to two different genera of corvids, a pattern strongly expressed by *Leucocytozoon*.

The prevalence of *Leucocytozoon* found in ravens from Bulgaria was much lower compared to the other studied corvid species (Table 2). The phylogeny of raven-infecting *Leucocytozoon* suggests at least two distinct clades. The parasite lineages CCORAX3 and COCOR02 form a clade with the non-corvid lineages ZOMON02 and ZOBOR07 of white-eyes (*Zosterops*). The majority of corvid *Leucocytozoon* lineages do not cluster significantly in a clade. The lineage CCORAX2 is closely related to GAGLA06 which was reported from Eurasian jays (*G. glandarius*) sampled in Portugal [6]. Both lineages seem to be closely related to the lineages PARUS14 and PARUS84 infecting blue tits (*Cyanistes caeruleus*). Obviously, ravens share some haemosporean lineages with other European *Corvus* species but also with species of other corvid genera. However, a phylogeny based on this cytochrome b fragment does not suggest a single clade of purely corvid-infecting *Leucocytozoon*. Although it was previously known only from blackfly vectors, the results of this study show that the lineage EUSE1 can infect ravens [49]. This lineage forms a clade with the *Leucocytozoon* lineage STRORIO3, known from doves, rather than with other corvid lineages. The lineages COCOR3 and COCOR13 were found in hooded

**Table 2** *Leucocytozoon* parasites prevalence in different studied birds of family Corvidae (only results of molecular studies included)

Corvid species	Location	Prevalence	N	Source
<i>Corvus corone</i>	Hokkaido, Japan	93.2	117	Yoshimura et al. [28]
<i>Corvus cornix</i>	Italy	97.8	46	Scaglione et al. [34]
<i>Corvus macrorhynchos</i>	Hokkaido, Japan	95.8	24	Yoshimura et al. [28]
<i>Corvus corone</i>	Germany	85.3	85	Schmid et al. [17]
<i>Corvus brachyrhynchos</i>	California, USA	57.4	258	Freund et al. [18]
<i>Pica nuttalli</i>	California, USA	54.5	44	Freund et al. [18]
<i>Cyanocitta stelleri</i>	California, USA	100.0	20	Freund et al. [18]
<i>Corvus corax</i>	Dolno Ozirovo, Bulgaria	31.4	88	Present study

crows (*C. cornix*) and domestic pigeons (*Columba livia f. domestica*) in Italy [16, 34]. In this case, the same leucocytozoid lineage was found in birds of different orders living in similar habitat. Both latter cases suggest that corvids and doves may have similar host characteristics, predisposing for host switching. Indeed, *Leucocytozoon* have recently been shown to have high rates of co-speciation with their hosts but also high rates of host switching [50].

The present study shows that even in widely distributed scavenger hosts with relatively low prevalence, such as ravens, the variety of *Leucocytozoon* lineages is quite high. The results also show that although most *Leucocytozoon* lineages in corvids are closely related, as pointed out by Freund et al. [18], they are represented by at least two distinct clades, both of which contain lineages probably specialized on corvids and other host groups, along with lineages tending toward a broader host spectrum.

## Conclusion

Several distinct clades of *Leucocytozoon* parasites appear to infect corvids. Ravens harbour both generalist corvid *Leucocytozoon* as well as apparently species-specific lineages. The extraordinary breeding ecology and scavenging lifestyle possibly allows ravens to evade vectors and have relatively low blood parasite prevalence compared to other corvids.

## Authors' contributions

All authors conceived the idea, gathered the samples for the study and drafted the manuscript. NC performed the molecular and phylogenetic analyses. PS performed the microscopic analyses. All authors read and approved the final manuscript.

## Author details

<sup>1</sup> National Museum of Natural History, Bulgarian Academy of Sciences, Sofia, Bulgaria. <sup>2</sup> Birds of Prey Protection Society, Sofia, Bulgaria. <sup>3</sup> Department of Animal Behaviour, Bielefeld University, 33501 Bielefeld, Germany. <sup>4</sup> Molecular Ecology and Evolution Lab, Department of Biology, Lund University, Lund, Sweden.

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## Competing interests

The authors declare that they have no competing interests.

## Availability of data and materials

The dataset used and analysed in the current study are available from the corresponding author on reasonable request. Sequences of newly described lineages have been deposited in GenBank and MalAvi databases.

## Consent for publication

Not applicable.

## Ethics approval and consent to participate

All procedures in the field were performed with a permit issued to the authors by the Bulgarian Ministry of Environment.

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## References

- Valkiūnas G. Avian malarial parasites and other haemosporidia. Boca Raton: CRC Press; 2005.
- Asghar M, Hasselquist D, Bensch S. Are chronic avian haemosporidian infections costly in wild birds? *J Avian Biol.* 2011;42:530–7.
- Asghar M, Hasselquist D, Hansson B, Zehntindjiev P, Westerdaal H, Bensch S. Hidden costs of infection: chronic malaria accelerates telomere degradation and senescence in wild birds. *Science.* 2015;347:436–8.
- Clark NJ, Clegg SM, Lima MR. A review of global diversity in avian haemosporidians (*Plasmodium* and *Haemoproteus*: Haemosporida): new insights from molecular data. *Int J Parasitol.* 2014;44:329–38.
- Beadell JS, Gering E, Austin J, Dumbacher JP, Peirce MA, Pratt TK, et al. Prevalence and differential host-specificity of two avian blood parasite genera in the Australo-Papuan region. *Mol Ecol.* 2004;13:3829–44.
- Mata VA, da Silva LP, Lopes RJ, Drovetski SV. The Strait of Gibraltar poses an effective barrier to host-specialised but not to host-generalised lineages of avian Haemosporidia. *Int J Parasitol.* 2015;45:711–9.
- Bishop MA, Bennett GF. Host-parasite catalogue of the avian haematozoa supplement 1; and bibliography of the avian blood-inhabiting haematozoa, supplement 2. St. John's: Dept. of Biology, Memorial University of Newfoundland; 1992.
- Merino S, Seoane J, De La Puente J, Bermejo A. Low prevalence of infection by haemoparasites in Cetti's Warblers (*Cettia cetti*) from central Spain. *Ardeola.* 2000;47:269–71.
- Shurulinkov P, Golemsky V. *Plasmodium* and *Leucocytozoon* (Sporozoa: Haemosporida) of wild birds in Bulgaria. *Acta Protozool.* 2003;42:205–14.
- Ellis VA, Kunkel MR, Ricklefs RE. The ecology of host immune responses to chronic avian haemosporidian infection. *Oecologia.* 2014;176:729–37.
- Chakarov N, Pauli M, Krüger O. Immune responses link parasite genetic diversity, prevalence and plumage morphs in common buzzards. *Evol Ecol.* 2017;31:51–62.
- Arriero E, Moller AP. Host ecology and life-history traits associated with blood parasite species richness in birds. *J Evol Biol.* 2008;21:1504–13.
- García-Longoria L, Gáramszegi LZ, Moller AP. Host escape behavior and blood parasite infections in birds. *Behav Ecol.* 2014;25:890–900.
- Lutz HL, Hochachka WM, Engel JJ, Bell JA, Tkach VV, Bates JM, et al. Parasite prevalence corresponds to host life history in a diverse assemblage of Afrotropical birds and haemosporidian parasites. *PLoS ONE.* 2015;10:e0128851.
- Bennett GF, Peirce MA. Leucocytozoids of 7 Old World passeriform families. *J Nat Hist.* 1992;26:693–707.
- Scaglione FE, Pregel P, Cannizzo FT, Perez-Rodriguez AD, Ferroglia E, Bollo E. Prevalence of new and known species of haemoparasites in feral pigeons in northwest Italy. *Malar J.* 2015;14:99.
- Schmid S, Fächet K, Dinkel A, Mackenstedt U, Woog F. Carrion crows (*Corvus corone*) of southwest Germany: important hosts for haemosporidian parasites. *Malar J.* 2017;16:369.
- Freund D, Wheeler SS, Townsend AK, Boyce WM, Ernest HB, Cicero C, et al. Genetic sequence data reveals widespread sharing of *Leucocytozoon* lineages in corvids. *Parasitol Res.* 2016;115:3557–65.
- Cramp S, Simmons KEL. Handbook of the birds of Europe, the Middle East and North Africa. Oxford: Oxford University Press; 1980.
- Glutz von Blotzheim U, Bauer K, Bezzel E. Handbuch der Vögel Mitteleuropas. IV ed. Wiesbaden: Aula-Verlag; 1971.
- Grünkorn T, Potiek A, Looft V, Jonker RM, Chakarov N, Krüger O. Territory quality affects the relative importance of habitat heterogeneity and

- interference competition in a long-lived territorial songbird. *J Avian Biol.* 2014;45:15–21.
22. Krüger O, Chakarov N, Nielsen JT, Looft V, Grünkorn T, Struwe-Juhl B, et al. Population regulation by habitat heterogeneity or individual adjustment? *J Anim Ecol.* 2012;81:330–40.
  23. Janicke T, Chakarov N. Effect of weather conditions on the communal roosting behaviour of common ravens *Corvus corax* with unlimited food resources. *J Ethol.* 2007;25:71–8.
  24. Blount JD, Houston DC, Møller AP, Wright J. Do individual branches of immune defence correlate? A comparative case study of scavenging and non-scavenging birds. *Oikos.* 2003;102:340–50.
  25. Sorci G. Immunity, resistance and tolerance in bird-parasite interactions. *Parasite Immunol.* 2013;35:350–61.
  26. Råberg L, Graham AL, Read AF. Decomposing health: tolerance and resistance to parasites in animals. *Philos Trans R Soc Lond B Biol Sci.* 2009;364:37–49.
  27. Blanco G, Gajon A, Doval G, Martinez F. Absence of blood parasites in Griffon vultures from Spain. *J Wildl Dis.* 1998;34:640–3.
  28. Yoshimura A, Koketsu M, Bando H, Saiki E, Suzuki M, Watanabe Y, et al. Phylogenetic comparison of avian haemosporidian parasites from resident and migratory birds in northern Japan. *J Wildl Dis.* 2014;50:235–42.
  29. Bensch S, Hellgren O, Pérez-Tris J. MalAvi: a public database of malaria parasites and related haemosporidians in avian hosts based on mitochondrial cytochrome b lineages. *Mol Ecol Resour.* 2009;9:1353–8.
  30. Hellgren O, Waldenström J, Bensch S. A new PCR assay for simultaneous studies of *Leucocytozoon*, *Plasmodium*, and *Haemoproteus* from avian blood. *J Parasitol.* 2004;90:797–802.
  31. Ronquist F, Huelsenbeck JP. MrBayes 3: Bayesian phylogenetic inference under mixed models. *Bioinformatics.* 2003;19:1572–4.
  32. Stamatakis A. RAxML-VI-HPC: maximum likelihood-based phylogenetic analyses with thousands of taxa and mixed models. *Bioinformatics.* 2006;22:2688–90.
  33. Murata K. Prevalence of blood parasites in Japanese wild birds. *J Vet Med Sci.* 2002;64:785–90.
  34. Scaglione FE, Cannizzo FT, Pregel P, Perez-Rodriguez AD, Bollo E. Blood parasites in hooded crows (*Corvus corone cornix*) in northwest Italy. *Veterinaria Italiana.* 2016;52:1111–6.
  35. Crompton PD, Moebius J, Portugal S, Waisberg M, Hart G, Garver LS, et al. Malaria immunity in man and mosquito: insights into unsolved mysteries of a deadly infectious disease. *Annu Rev Immunol.* 2014;32:157–87.
  36. Chakarov N, Boerner M, Krüger O. Fitness in common buzzards at the cross-point of opposite melanin–parasite interactions. *Funct Ecol.* 2008;22:1062–9.
  37. Chakarov N, Linke B, Boerner M, Goesmann A, Kruger O, Hoffman JI. Apparent vector-mediated parent-to-offspring transmission in an avian malaria-like parasite. *Mol Ecol.* 2015;24:1355–63.
  38. Atkinson CT, Dusek RJ, Lease JK. Serological responses and immunity to superinfection with avian malaria in experimentally-infected Hawaii Amakihi. *J Wildl Dis.* 2001;37:20–7.
  39. Hammers M, Komdeur J, Kingma SA, Hutchings K, Fairfield EA, Gilroy DL, et al. Age-specific haemosporidian infection dynamics and survival in Seychelles warblers. *Sci Rep.* 2016;6:29720.
  40. Drovetski SV, Aghayan SA, Mata VA, Lopes RJ, Mode NA, Harvey JA, et al. Does the niche breadth or trade-off hypothesis explain the abundance-occupancy relationship in avian Haemosporidia? *Mol Ecol.* 2014;23:3322–9.
  41. Kim KS, Tsuda Y. Seasonal changes in the feeding pattern of *Culex pipiens pallens* govern the transmission dynamics of multiple lineages of avian malaria parasites in Japanese wild bird community. *Mol Ecol.* 2010;19:5545–54.
  42. Martinsen ES, Paperna I, Schall JJ. Morphological versus molecular identification of avian Haemosporidia: an exploration of three species concepts. *Parasitology.* 2006;133:279–88.
  43. Zehtindjiev P, Ilieva M, Westerdahl H, Hansson B, Valkiunas G, Bensch S. Dynamics of parasitemia of malaria parasites in a naturally and experimentally infected migratory songbird, the great reed warbler *Acrocephalus arundinaceus*. *Exp Parasitol.* 2008;119:99–110.
  44. Zehtindjiev P, Ilieva M, Krizanauskiene A, Oparina O, Oparin M, Bensch S. Occurrence of haemosporidian parasites in the paddyfield warbler, *Acrocephalus agricola* (Passeriformes, Sylviidae). *Acta Parasitol.* 2009;54:295–300.
  45. Dimitrov D, Zehtindjiev P, Bensch S. Genetic diversity of avian blood parasites in SE Europe: cytochrome b lineages of the genera *Plasmodium* and *Haemoproteus* (Haemosporida) from Bulgaria. *Acta Parasitol.* 2010;55:201–9.
  46. Perez-Rodriguez A, de la Hera I, Bensch S, Perez-Tris J. Evolution of seasonal transmission patterns in avian blood-borne parasites. *Int J Parasitol.* 2015;45:605–11.
  47. Hellgren O, Waldenström J, Pérez-Tris J, Szöllösi E, Hasselquist D, Krizanauskiene A, et al. Detecting shifts of transmission areas in avian blood parasites—a phylogenetic approach. *Mol Ecol.* 2007;16:1281–90.
  48. Scheuerlein A, Ricklefs RE. Prevalence of blood parasites in European passeriform birds. *Proc Biol Sci.* 2004;271:1363–70.
  49. Synek P, Munclinger P, Albrecht T, Votýpka J. Avian haemosporidians in haematophagous insects in the Czech Republic. *Parasitol Res.* 2013;112:839–45.
  50. Alcalá N, Jenkins T, Christie P, Vuilleumier S. Host shift and cospeciation rate estimation from co-phylogenies. *Ecol Lett.* 2017;20:1014–24.

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