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Use of titanium dioxide nanoparticles biosynthesized by *Bacillus mycooides* in quantum dot sensitized solar cells

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Abstract

Background: One of the major challenges of nanotechnology during the last decade has been the development of new procedures to synthesize nanoparticles. In this context, biosynthetic methods have taken hold since they are simple, safe and eco-friendly.

Results: In this study, we report the biosynthesis of TiO₂ nanoparticles by an environmental isolate of *Bacillus mycooides*, a poorly described Gram-positive bacterium able to form colonies with novel morphologies. This isolate was able to produce TiO₂ nanoparticles at 37°C in the presence of titanyl hydroxide. Biosynthesized nanoparticles have anatase polymorphic structure, spherical morphology, polydisperse size (40–60 nm) and an organic shell as determined by UV-vis spectroscopy, TEM, DLS and FTIR, respectively. Also, conversely to chemically produced nanoparticles, biosynthesized TiO₂ do not display phototoxicity. In order to design less expensive and greener solar cells, biosynthesized nanoparticles were evaluated in Quantum Dot Sensitized Solar Cells (QDSSCs) and compared with chemically produced TiO₂ nanoparticles. Solar cell parameters such as short circuit current density (I_{sc}) and open circuit voltage (V_{oc}) revealed that biosynthesized TiO₂ nanoparticles can mobilize electrons in QDSSCs similarly than chemically produced TiO₂.

Conclusions: Our results indicate that bacterial extracellular production of TiO₂ nanoparticles at low temperatures represents a novel alternative for the construction of green solar cells.

Keywords: Titanium dioxide nanoparticles, QDSSC, Phototoxicity, Nanoparticle biosynthesis

Introduction

The rapid advance of nanotechnology and the increasing number of applications involving nanomaterials have prompted the interest in developing simple and environmentally friendly protocols for nanoparticle synthesis.

To date, titanium nanoparticles (NPs) are one of the most required nanomaterials because of its use on different technological applications. Nanoparticulated titanium dioxide is a highly valuable material since it is used as photocatalyst degrading organic molecules in water treatment [1], white pigment in paint manufacturing, additive in food

and personal care products [2], and composite films in biomedical sciences [3], among many other applications.

Since TiO₂ nanoparticles can conduct electrons as a wide-band gap semiconductor, its use as photoanode in the manufacture of Dye- and Quantum Dots- Sensitized Solar Cells (DSSCs and QDSSCs, respectively) has gained importance during the last decade [4,5]. This application has become more attractive during the last years due to the global need for replacing fossil fuels for energy generation. Accordingly, the research and development of non-conventional renewable energies (NCRE), particularly solar energy, which arises as a sustainable and abundant alternative to meet the high world energy demand, has been strongly stimulated [6].

Current methods to produce TiO₂ nanoparticles involve different chemical procedures such as sol-gel [7],

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hydrothermal [8] and solvothermal [9], among others. All these methods involve high temperatures (>200°C) and in some cases elevated pressures, both conditions affecting the safety and costs of the process.

During the last years, NPs biosynthesis methods involving microorganisms [10,11] or plant extracts [12], as well as eco-friendly chemical procedures involving low toxicity reagents and mild conditions of temperature and pressure, have taken hold [13]. This has allowed the

development of nanoparticles displaying novel properties such as composition, size and biocompatibility [14-20]. Biological protocols for the synthesis of TiO₂ nanoparticles have been developed, and the use of bacteria [21], yeasts [22], fungi [23] and plant extracts [24] have been recently reported (Table 1). Most biosynthetic procedures involve the use of titanyl hydroxide as precursor, and generate NPs with similar properties to those obtained by chemical procedures, such as size distribution

Table 1 Biosynthesis of TiO₂ nanoparticles reported to date

Organism used in the biosynthesis	Gram	Precursor	Synthesis temperature	Particle size	Crystal structure (dominant)	Application	Reference
<i>Saccharomyces cerevisiae</i>	Not applicable	Titanyl hydroxide TiO(OH) ₂	60°C	13 nm (TEM) 18 nm (XRD)	Anatase and Rutile	-	[22]
<i>Lactobacillus sp.</i>	(+)	Titanyl hydroxide TiO(OH) ₂	60°C	25 nm (TEM) 30 nm (XRD)	Anatase and Rutile	-	[22]
<i>Bacillus subtilis</i>	(+)	Titanyl hydroxide TiO(OH) ₂	60°C	66-77 nm (SEM)	Anatase	-	[21]
Leaves extract of <i>Nyctanthes arbor-tristis</i>	Not applicable	Titanium tetraisopropoxide	1. 50°C	100 nm (XRD)	Not shown	-	[29]
		Ti(OCH(CH ₃) ₂) ₄	2. 500°C (calcined)	100-150 nm (SEM)			
Aqueous extract of <i>Jatropha curcas L. latex</i>	Not applicable	Titanyl hydroxide TiO(OH) ₂	50°C	25-100 nm	Anatase	-	[30]
<i>Aspergillus flavus</i>	Not applicable	Titanium dioxide TiO ₂	37°C	62-74 nm	Anatase and Rutile	Against pathogenic bacteria	[23]
Leaf aqueous extract of <i>Eclipta prostrata</i>	Not applicable	Titanyl hydroxide TiO(OH) ₂	Room Temperature	49.5 nm (36-68 nm)	Rutile	-	[24]
<i>Annona squamosa</i> peel extract	Not applicable	Titanyl hydroxide	60°C (Optimal)	26 nm (XRD)	Rutile	-	[31]
		TiO(OH) ₂		23 nm (TEM)			
<i>Bacillus subtilis</i>	(+)	Potassium hexafluorotitanate	1. Not shown	10-30 nm	Anatase	Photocatalytic activity on aquatic biofilm	[28]
		K ₂ TiF ₆	2. 500°C (to crystallize particles)				
Leaves extract of <i>Catharanthus roseus</i>	Not applicable	Titanium dioxide	50°C	25-110 nm (SEM)	Anatase and Rutile	Antiparasitic activity	[32]
		TiO ₂ (powder)		65 nm (XRD)			
Flower aqueous extract of <i>Calotropis gigantea</i>	Not applicable	Titanyl hydroxide	90°C	160-220 nm (SEM)	Not shown	Acaricidal activity	[26]
		TiO(OH) ₂		10.52 nm (XRD)			
<i>Planomicrobium sp.</i>	(+)	Titanium dioxide	50°C	100-500 nm (SEM)	Not shown	Antibacterial and antifungal activity	[33]
		TiO ₂		8.89 nm (XRD)			
<i>Propionibacterium jensenii</i>	(+)	Titanyl hydroxide	1. 60°C	65 nm (XRD)	Anatase	Preparation of collagen-TiO ₂ wound dressing	[27]
		TiO(OH) ₂	2. 300°C (annealed)	10-80 nm (FE-SEM)			
<i>Aeromonas hydrophila</i>	(-)	Titanyl hydroxide	30°C	40.5 nm (XRD)	Rutile	Antibacterial activity	[25]
		TiO(OH) ₂		28-54 nm (SEM)			
Leaves extract of <i>Solanum trilobatum</i>	Not applicable	Titanyl hydroxide TiO(OH) ₂	Room Temperature	70 nm (SEM)	Rutile	Antiparasitic activity	[34]

and anatase/rutile crystal structure. In addition, most applications of biosynthesized TiO₂ nanoparticles tested to date are based on their toxic properties against pathogens such as bacteria [23,25] and mites [26]. Furthermore, *in vivo* and *in vitro* biocompatibility studies [27] have been carried out and their biocidal/photocatalytic activity on aquatic biofilms has been evaluated [28] (Table 1).

No reports regarding the application of biosynthesized titanium dioxide nanoparticles in energy devices have been published to date (Table 1). The use of other biologically produced materials in sensitized solar cells has been recently reported, however these reports focused on using plant pigments or channel proteins in the photon harvest process [35,36].

The present work reports for the first time the use of biosynthesized titanium dioxide nanoparticles by *B. mycooides*, as semiconductors in the manufacture of photoanodes for QDSSCs. Along with introducing a new method for the synthesis of TiO₂ nanoparticles, the present manuscript constitutes a first approach for using biosynthesized nanoparticles in solar cells and opens the interest in using other biosynthesized nanoparticles in energy devices as a way to develop greener photovoltaic technologies at low production costs.

Results and discussion

Environmental isolate of *Bacillus mycooides*

A *B. mycooides* strain isolated from a soil sample obtained from a volcanic zone in Chile was used for biosynthesis of TiO₂ NPs. *B. mycooides* is a member of the *Bacillus cereus* group of bacteria, a nonpathogenic soil and saprophyte Gram-positive bacilli. When grown on agar plates, this bacterium has the ability to form chains of cells that define macroscopic colonies with filaments projecting radially and curving in two possible orientations, clockwise (Dextral or DX strains) or counter-clockwise (Sinistral or SIN strains) [37]. The environmental isolate used in this work displays the classical colony morphology of *B. mycooides* with radial filaments in an anti-clockwise direction, indicative of a SIN strain (Figure 1).

Biosynthesis of TiO₂ nanoparticles by the isolated strain of *B. mycooides* was carried out by exposing bacterial cultures to titanyl hydroxide at 37°C, the optimal growth temperature determined for this environmental isolate (not shown). Then, the temperature of the culture was diminished (20–25°C) to stop the reaction and a white precipitate was formed, indicative of TiO₂ nanoparticles synthesis. The precipitate was purified from the culture, washed and resuspended in Mili-Q ultra pure water for subsequent studies.

To date, there are two studies reporting the use of the genus *Bacillus* for biosynthesis of TiO₂ nanoparticles, however, these reports do not use titanyl hydroxide as precursor [28] and require high temperatures for the synthesis

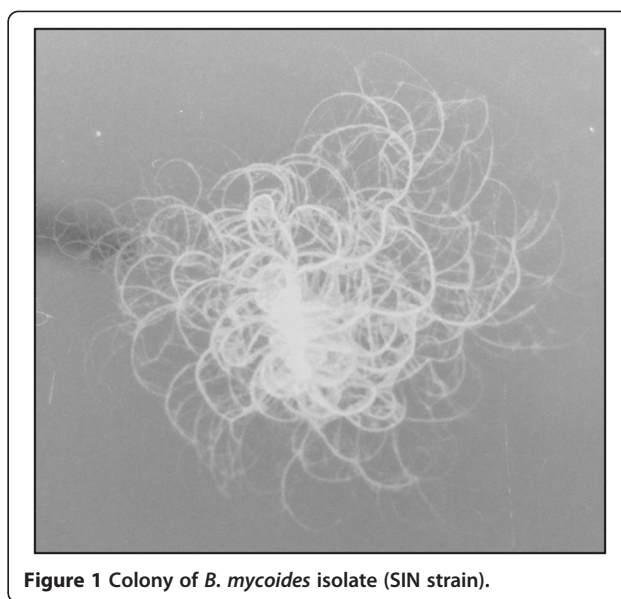


Figure 1 Colony of *B. mycooides* isolate (SIN strain).

of NPs [21] (Table 1). Differences in the biosynthetic process mediated by *B. mycooides* suggest that different biomolecules could be involved in biosynthesis and that the produced NPs could display novel properties.

Transmission electron microscopy (TEM)

A TEM analysis of the nanoparticles produced by *B. mycooides* was performed with the aim to determine their nanometric size and distribution. Biosynthesized TiO₂ nanoparticles display a size between 40–60 nm and spherical morphology (Figure 2). The size distribution histogram indicates a high polydispersity of the sample; which is a common behavior of NPs produced by biosynthetic methods [23,24,27]. This result suggests that nanoparticles are produced by cells at different times after addition of titanyl hydroxide. TEM images of Figure 2a show nanoparticles coated by an organic envelope, probably corresponding to the extracellular matrix produced by *B. mycooides*. This matrix could participate in substrate biotransformation (titanyl hydroxide to titanium dioxide nanoparticles), or maybe could help stabilizing and/or capping the NPs. When this sample was further purified by successive washings steps using ultra pure water, individual nanoparticles were observed (Figure 2b).

Fourier transform infrared spectroscopy (FTIR) analysis

Figure 3 shows the FT-IR spectrum of titanyl hydroxide, biosynthesized titanium dioxide nanoparticles and chemically synthesized titanium dioxide nanoparticles. The FTIR spectrum of TiO(OH)₂ shows the characteristic signals at 3400 cm⁻¹ and 1630 cm⁻¹ attributed to the presence of hydroxyl groups (Ti-OH) and water in their structure. The characteristic signal for TiO₂ nanoparticles due to the vibration of Ti-O-Ti bond is observed at 450–700 cm⁻¹ in

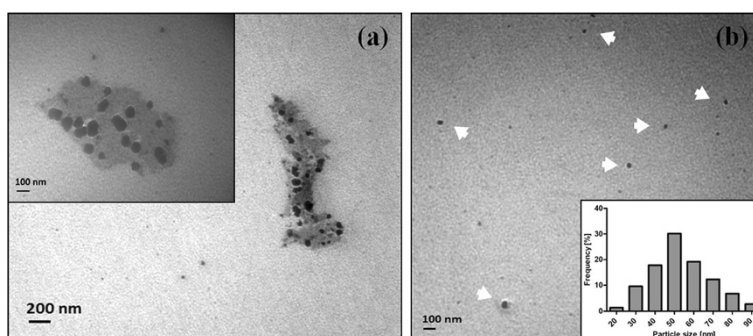


Figure 2 Transmission electron microscopy of TiO_2 nanoparticles synthesized by *B. mycooides*. Biosynthesized nanoparticles before (a) and after successive washings in water (b). The inset in (a) shows the spherical morphology of TiO_2 nanoparticles. The inset in (b) shows the size distribution histogram of nanoparticles. Arrow heads indicate individual NPs. Images were obtained with a 100,000 \times magnification.

both chemical and microbiological NPs. For biosynthesized TiO_2 nanoparticles a broad band at 3431 cm^{-1} is observed. This signal corresponds to the O-H stretching due to the alcoholic group. The peaks around 2985 cm^{-1} are assigned to the symmetric stretch (C-H) of CH_2 and CH_3 groups of aliphatic chains. In 1646 cm^{-1} , 1554 cm^{-1} , 1462 cm^{-1} and 1400 cm^{-1} the characteristic signals of C=O and N-H vibrations due to the presence of amide and amine groups are shown. 1246 cm^{-1} corresponds to C-O stretch vibrations, possibly due to the presence of an alcohol or carboxylic acid group. The band at 1047 cm^{-1} corresponds to the C-N stretching vibrations of aliphatic amines. The peaks at 1554 cm^{-1} and 1400 cm^{-1} might also indicate the C=C ring stretching and bending vibration of CH_2 . All these signals can be attributed to the presence of biomolecules like peptides or carbohydrates

bound to the TiO_2 nanoparticles produced by *B. mycooides*. These biomolecules are part of the cell envelope of this bacterium and can provide support for the nucleation of the nanoparticles, and/or be involved in the biosynthesis process acting as stabilizing and capping agents.

UV-visible spectroscopy and Tauc Plot analysis

In order to determine the UV-visible absorption spectrum of biosynthesized TiO_2 nanoparticles, the product of biosynthesis was washed gently with ethanol and resuspended in ultrapure water to remove organic residues that might interfere with the measurement.

The observed absorption spectrum coincides with those obtained with titanium dioxide nanoparticles produced by chemical methods, with a broad absorption band in the UV range and the cut off wavelength near 380 nm

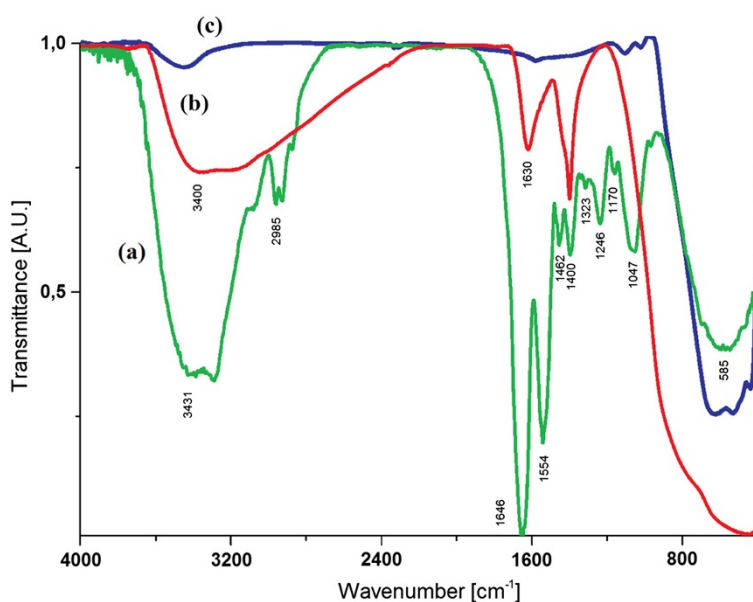


Figure 3 FTIR spectra of TiO_2 nanoparticles biosynthesized by *B. mycooides* (a), the precursor $\text{TiO}(\text{OH})_2$ (b), and TiO_2 nanoparticles produced by chemical synthesis (c).

(Figure 4) [27,28,38,39]. The obtained UV-vis spectra of biosynthesized TiO₂ nanoparticles was used to determine the band gap (E_{bg}) by the Tauc relation [40]. The method for determination of the E_{bg} value involves plotting $(\alpha h\nu)^2$ versus $h\nu$, where α is the absorption coefficient and $h\nu$ is the energy of the incident photons. After making a linear fit to the curve, the value of the band gap is given by the value of the intercept of the line with the X-axis ($h\nu$ -intercept) in this graph (Figure 4, inset) [41]. The band gap determined for biosynthesized TiO₂ nanoparticles was 3.27 eV. This band gap value confirms that *B. mycooides* is producing titanium dioxide in anatase crystalline structure. Due to their wide band gap, TiO₂ nanoparticles in anatase crystalline form are preferably used in sensitized solar cells [42]. This result suggests that biosynthesized nanoparticles are suitable semiconductor materials that can be used in QDSSCs.

Antibacterial activity of biosynthesized TiO₂ nanoparticles

Toxicity of TiO₂ nanoparticles has become a relevant parameter since it can determine its use in different technological applications. Almost all publications related to biosynthesis of TiO₂ NPs report the toxic and/or phototoxic effects of them (Table 1). Toxicity can decrease the number of technological applications in which TiO₂ NPs can be used, but is the base of its use as antimicrobials and photo-reactive compounds. The use of nanoparticles in solar cells is not the exception and in addition to proper size, composition and semiconductor properties, increased biocompatibility will strongly favor their application in harvesting solar energy.

As shown in Figure 5a, almost no toxicity was determined in chemical and biological nanoparticles. *E. coli* cultures were able to reestablish their growth after NPs exposure. To confirm our results, growth inhibition area assays were done, and no toxicity was determined for the biological and chemical nanoparticles evaluated (data not shown). Other studies have reported the biosynthesis of TiO₂ nanoparticles displaying high toxicity for *E. coli*, with minimum inhibitory concentrations (MIC) near 20 and 40 $\mu\text{g/mL}$ [23,25]. In contrast, the nanoparticles produced by *B. mycooides* do not display any toxicity to *E. coli* at these concentrations.

Most of the damage that TiO₂ nanoparticles produce on microorganisms has been associated to phototoxicity (Table 1). When photocatalytic activity against *E. coli* was evaluated, a small decrease in CFU was determined for chemically synthesized TiO₂ (Figure 5b). This is due to lipid peroxidation on bacterial cell membranes produced by reactive oxygen species (OH \cdot , O₂ \cdot^- and H₂O₂) generated when titanium dioxide NPs are irradiated with UV light [43]. When *E. coli* cultures were amended with biologically synthesized TiO₂ NPs and exposed to UV-B light, no effect on cell viability was determined, indicating that biosynthesized NPs do not display phototoxicity under the evaluated conditions (Figure 5b). Based on these results we can speculate that the organic coating of nanoparticles produced by *B. mycooides* protects bacteria from the phototoxic damage by interacting with UV-produced radicals. In this context, additional purification steps decreasing the organic matter of NPs probably increase phototoxicity of biosynthesized TiO₂ NPs.

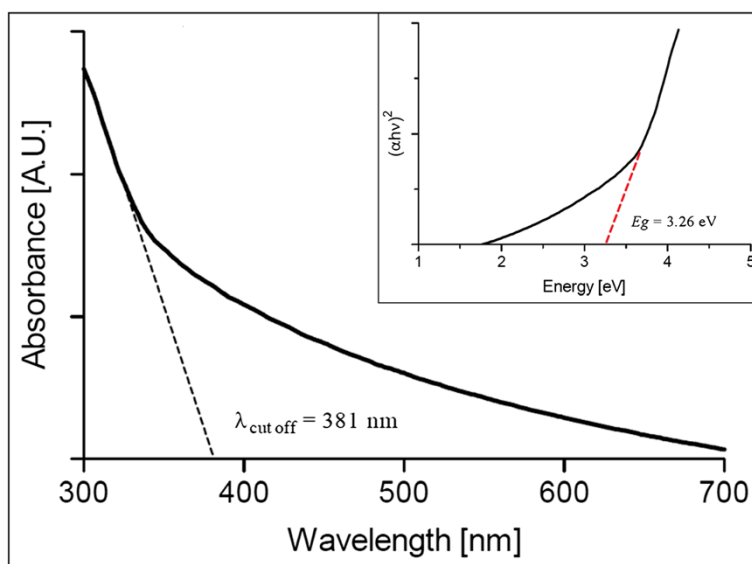


Figure 4 UV-visible absorption spectrum of biosynthesized TiO₂ nanoparticles and Tauc plot used to determine its band gap (inset).

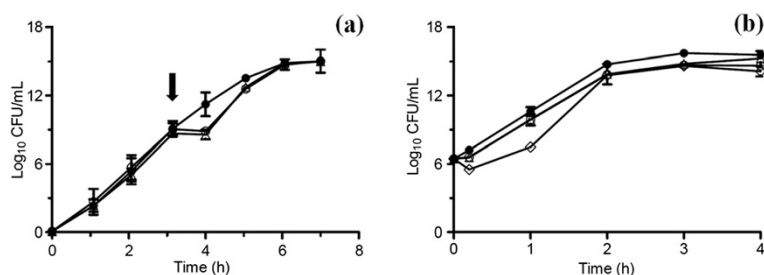


Figure 5 Toxicity and phototoxicity evaluation of TiO₂ nanoparticles against *E. coli*. (a) *E. coli* CFUs after exposure to 200 µg/mL of chemical (○) and biological (▲) TiO₂ nanoparticles (control ●). (b) *E. coli* CFUs after treatment with 200 µg/mL of chemical (◇) and biological (△) TiO₂ nanoparticles exposed to UV light (control exposed to UV ○, control unexposed to UV ●). The arrow in a indicates the time in which cells were treated with TiO₂ nanoparticles.

Characterization and I-V measurement of the quantum dot sensitized solar cells

Based on the favorable properties of biosynthesized TiO₂ nanoparticles as wide-band gap semiconductors with low levels of toxicity, we decided to evaluate their use on solar cells.

A schematic representation of the solar cell used to evaluate the biosynthesized TiO₂ NPs is shown in Figure 6. When QDs absorb light, they inject electrons from their excited levels to the conduction band of the TiO₂ nanoparticles film. The recirculation of the redox electrolyte in its oxidized-reduced state allows to recharge the electrons lost by oxidized QDs while serving as a pathway for electron transfer between the two electrodes [5,44]. Thus, when light shines on the solar cell, the device directly converts sunlight into electricity and the current and voltage data can be recorded in an external circuit.

The results of short circuit current density (I_{SC}) and open circuit voltage (V_{OC}) for the studied solar cells are summarized in Figure 7. The I_{SC} value corresponds to the maximum current (flow of electric charge) through the solar cell when the voltage in the device is zero. Moreover, the V_{OC} is the maximum voltage (electric potential difference) produced by the solar cell when the current flow is zero.

The control solar cells produced with chemical titanium dioxide nanoparticles display the highest values of voltage and current, confirming that the flow of electrons between the CdTe-GSH QDs and chemical TiO₂ NPs works properly. Moreover, the QDSSC that uses biosynthesized TiO₂ nanoparticles shows decreased values of I_{SC} and V_{OC} , attributable to the presence of calcined organic matter on the surface of nanoparticles after sintering of the material; this calcined matter would work as insulation in electrical conduction. However, when the organic coating of TiO₂ nanoparticles was removed, V_{OC} and I_{SC} values are significantly increased and the performance of the solar cell using biosynthesized TiO₂ nanoparticles is similar to that observed in the control. The results obtained indicate that it is possible to use the TiO₂ nanoparticles produced by *B. mycoides* in the development of greener solar cells.

Conclusions

In this work the biosynthesis of titanium dioxide nanoparticles using an environmental isolate of *B. mycoides* was reported. Although the transformation of TiO(OH)₂ to TiO₂ normally occurs in a drying process or sol-gel combustion at temperatures between 150 and 400°C [45-47], in this paper we have shown that a *B. mycoides*

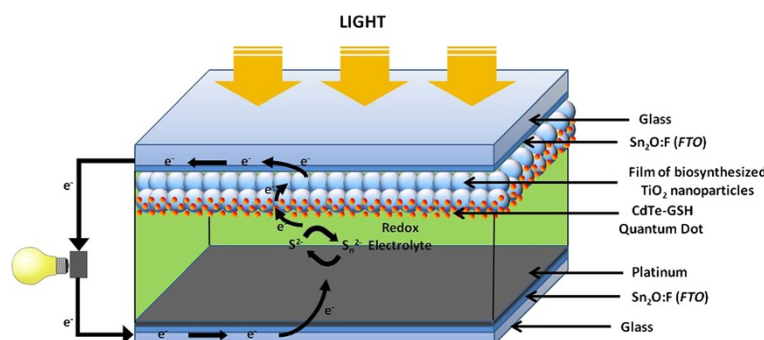


Figure 6 Scheme of a quantum dot sensitized solar cell. The film of biosynthesized TiO₂ nanoparticles on conductive glass is sensitized with CdTe-GSH QDs forming the photoanode, while the cathode or counter electrode is platinum on FTO glass. The redox electrolyte fills the space between electrodes.

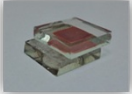
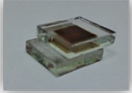
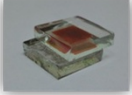
Wide-bandgap semiconductor	Solar Cell	Open circuit voltage V_{oc} [mV]	Short circuit current I_{sc} [$\mu\text{A}\cdot\text{cm}^{-2}$]
Chemical TiO_2 nanoparticles		440	1.66
Biosynthesized TiO_2 nanoparticles Without purification		28	0.10
Biosynthesized TiO_2 nanoparticles SDS 1 %		249	1.25

Figure 7 I-V values of QDSSCs produced using biosynthesized TiO_2 NPs.

isolate is able to biotransform this precursor into its nanostructured form at 37°C . A possible mechanism for TiO_2 biosynthesis using titanyl hydroxide as precursor and a still unknown organic molecule from *B. mycooides* (X) is proposed in Scheme 1. Experiments to determine the identity of the organic molecule(s) involved in biosynthesis are under way in our laboratory.

TiO_2 nanoparticles biosynthesized by *B. mycooides* exhibit low toxicity against *E. coli*, probably as consequence of the organic coating.

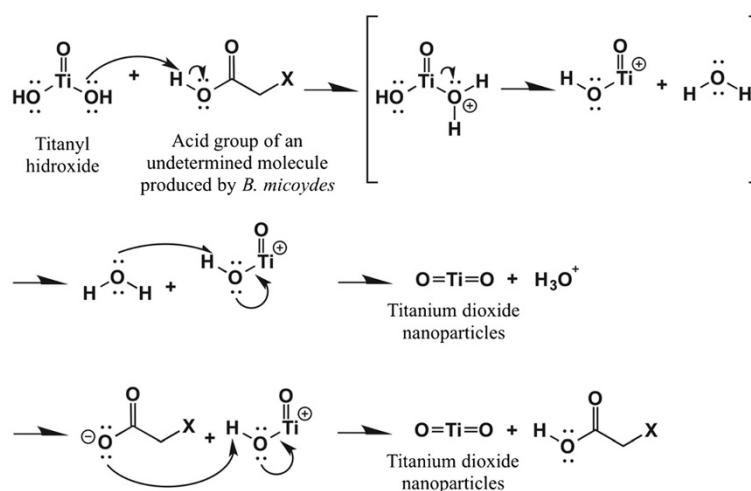
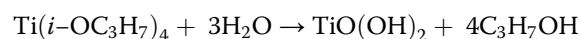
Biosynthesized nanoparticles are able to conduct electrons in QDSSC with values near those determined in a control solar cell produced with chemically synthesized TiO_2 nanoparticles.

The main projection of this work is the use of these and other green nanoparticles in the sustainable manufacturing of solar cells to develop ecologically friendly and less expensive photovoltaic panels.

Methods

Synthesis of titanyl hydroxide precursor

Titanyl hydroxide was obtained by the hydrolysis of titanium isopropoxide [$\text{Ti}(i\text{-OPr})_4$] according to the following reaction [46]:



Scheme 1 Possible mechanism for biotransformation of titanyl hydroxide to titanium dioxide nanoparticles. The dehydration reaction would be mediated by an acidic group present in an "unknown" component of the extracellular matrix of *B. mycooides* that could have a key role in biotransformation.

The reaction mixture was stirred for two hours under ice-cold condition until an opalescent suspension of $\text{TiO}(\text{OH})_2$ was obtained. The isopropyl alcohol (propan-2-ol) was removed by successive centrifugations and washings with Mili-Q ultrapure water. Finally, titanil hydroxide was resuspended in water. The precursor concentration was obtained by determining the dry mass of a 1 mL solution, which was lyophilized (freeze-dried) for 24 h.

Biosynthesis of TiO_2 nanoparticles

A culture of 200 μL of *B. mycooides* grown overnight was used to inoculate 200 mL of LB medium (dilution 1:1000). This culture solution was grown for 12 h at 37°C with constant shaking (150 RPM). Then, 40 mL of a 25 mM titanil hydroxide solution were added and the mixture incubated at 37°C for 24 h with constant shaking. After this time, the solution was incubated at room temperature for 8 h and the appearance of a white precipitate was indicative of the production of titanium dioxide nanoparticles. The precipitate was removed from the culture by centrifuging 15 min at $3820 \times g$. Finally, the biosynthesis product was washed and resuspended by successive centrifugations in Mili-Q ultra pure water.

Characterization of TiO_2 nanoparticles

Biosynthesized TiO_2 nanoparticles were characterized by UV-visible spectroscopy using a Synergy™ H1 Microplate Reader (BioTek Instrument Inc.). Absorbance spectrum between 300–700 nm (2 nm resolution) was performed and used for band gap (E_{bg}) determination using the Tauc relation [41].

For Transmission Electron Microscopy (TEM) studies, a suspension of TiO_2 nanoparticles was deposited on a copper grid and examined using a Low Voltage Transmission Electron Microscope 5 (LVEM5) (DeLong Instruments) operated at 5.1 kV. The size distribution histogram was performed using ImageJ software. Dinamic Light Scattering (DLS) was performed in a Zetasizer Nano ZS (Malvern Instrument Ltd.) equipment using the protocol previously described by our group [10].

For the Fourier Transform Infrared Spectroscopy (FT-IR) characterization samples were lyophilized (freeze-dried) for 24 h and the powder was mixed with KBr to form a thin pellet. FT-IR measurements were carried out using a Spectrum One FT-IR Spectrometer (Perkin Elmer Inc.) in the $400\text{--}4000\text{ cm}^{-1}$ range with a 4 cm^{-1} resolution.

Antibacterial activity of TiO_2 nanoparticles

The antibacterial activity of biosynthesized TiO_2 nanoparticles was evaluated against *E. coli* (BW25113). Bacterial cultures were grown in LB medium at 37°C with constant shaking (150 RPM). After 3 h incubation ($\text{OD}_{600} = 0.3$) cultures were amended with 200 $\mu\text{g}/\text{mL}$ of chemically (TiO_2 nanopowder from Sigma-Aldrich, $\sim 21\text{ nm}$ particle

size) or biologically synthesized TiO_2 nanoparticles. The photocatalytic effect of nanoparticles was evaluated using the same concentrations indicated above, but irradiating the culture with UV-B light for 2 min in the presence of the nanoparticles ($\text{OD}_{600} = 0.3$).

The effect of TiO_2 nanoparticles on bacterial growth was evaluated by determining the number of colony forming units (CFU) over time. Culture aliquots were taken every hour and diluted to obtain 10^{-1} to 10^{-7} serial dilutions. 5 μL of every dilution were plated on LB agar, and incubated at 37°C for 12 h. After this time, CFU were determined.

Fabrication and characterization of quantum dot sensitized solar cells

QDSSCs were produced following the protocols described by Bang *et al.* [48], Giménez *et al.* [49] and Pan *et al.* [50], with some modifications. To fabricate the electrodes of QDSSCs, $10 \times 10 \times 2\text{ mm}$ size fluorine doped tin oxide coated glass (FTO glass) TEC15, with a surface resistivity of 13 [Ω/sq] and 85% transmittance was used. Conductive glasses were cleaned by successive sonication in absolute ethanol and deionized water for approximately 10 min to remove organic contaminants. The anode was prepared using a suspension of biosynthesized TiO_2 nanoparticles that was deposited on the glass through spin-coating at 2000 rpm for 10 sec.

To prepare a uniform titanium dioxide film that facilitates electron transfer process in QDSSCs, it is important to remove the organic coating on the surface of biosynthesized TiO_2 nanoparticles. For this reason, an additional purification step was performed. Nanoparticles were treated with 1% sodium dodecyl sulfate (SDS) and the solution was sonicated gently for a few seconds to allow disaggregation of the nanoparticles. Then, the suspension of TiO_2 nanoparticles was recovered by centrifugation, washed and resuspended in Mili-Q ultra pure water. Titanium(IV) oxide nanopowder from Sigma-Aldrich and biosynthesized TiO_2 nanoparticles were used to manufacture the photoanodes of QDSSCs. The electrodes (TiO_2 films) underwent a sintering process at 450°C for 30 min. Sensitization of TiO_2 film was performed by direct adsorption of CdTe-GSH quantum dots (QDs) [51]. The active area of the cells was 0.16 cm^2 . Moreover, the cathode or counter electrode was prepared from a solution of $\text{H}_2\text{PtCl}_6 \cdot 6\text{H}_2\text{O}$ in isopropanol. 10 μL of the solution were dispensed on a FTO coated glass by spin-coating and heated 20 min at 400°C .

Then, the photoanode and the counter electrode were assembled leaving 127 μm space between them. Before sealing the cell, a drop of electrolyte was added. The electrolyte solution used was sulfide/polysulfide ($\text{S}^{2-}/\text{S}_n^{2-}$) prepared from Na_2S (1.0 M), S (0.1 M) and NaOH (0.1 M) in Mili-Q ultrapure water. Characterization of solar cells was

performed under constant conditions of temperature and irradiance at a one sun intensity as the light source ($\sim 100 \text{ mW} \cdot \text{cm}^{-2}$ and AM1.5).

Competing interests

The authors declare that they have no competing interests.

Authors' contributions

NOA participates in the experimental design, carried out the precursor synthesis, microbiological work, built and characterized the solar cell, and drafted the manuscript. LSA isolated the bacteria and participated in the design of the experiments. JPM designed the experiments and purified the nanoparticles. VDT carried out bacterial toxicity experiments of nanoparticles. DB drafted the manuscript. JMP conceived the study, participated in the experimental design and coordination and writing of the manuscript. All authors have read and approved the final manuscript.

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