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# MicroRNAs as potential biopredictors for premenopausal osteoporosis: a biochemical and molecular study

Hadeel A. Al-Rawaf<sup>1</sup>, Sami A. Gabr<sup>2</sup>, Amir Iqbal<sup>2\*</sup> and Ahmad H. Alghadir<sup>2</sup>

## Abstract

**Background** Circulating micro-RNAs have been proposed as a new type of biomarker in several diseases, particularly those related to bone health. They have shown great potential due to their feasibility and simplicity of measurement in all body fluids, especially urine, plasma, and serum.

**Aim** This study aimed to evaluate the expression of a set of mRNAs, namely miR-21, miR-24, miR-100, miR-24a, miR-103-3p, and miR-142-3p. Their proposed roles in the progression of osteoporosis were identified using a real-time polymerase chain reaction (RT-PCR) analysis in premenopausal women. In addition, their correlations with osteocalcin (OC), bone-specific alkaline phosphatase (BAP), and deoxypyridinoline (DPD) bone markers were explored.

**Methods** A total of 85 healthy premenopausal women aged 25–50 years old were included in this study. Based on a DXA scan (Z-score) analysis and calcaneus broadband ultrasound attenuation scores (c-BUAs), measured via quantitative ultrasound (QUS), the subjects were classified into three groups: normal group (n = 25), osteopenia (n = 30), and osteoporosis (n = 30). Real-time-PCR and immunoassay analyses were performed to determine miRNA expression levels and serum OC, s-BAP, and DPD, respectively, as biomarkers of bone health.

**Results** Among the identified miRNAs, only miR-21, miR-24, and miR-100 were significantly upregulated and increased in the serum of patients with osteopenia and osteoporosis, and miR-24a, miR-103-3p, and miR-142-3p were downregulated and significantly decreased in osteoporosis. Both upregulated and downregulated miRNAs were significantly correlated with BMD, c-BUA, OC, s-BAP, and DPD.

**Conclusion** A group of circulating miRNAs was shown to be closely correlated with the parameters BMD, c-BUA, OC, s-BAP, and DPD, which are traditionally used for bone-health measurements. They could be identified as non-invasive biomarkers in premenopausal patients with osteoporosis. More studies with large sample sizes are recommended to estimate the mechanistic role of miRNAs in osteoporosis pathogenesis and to provide evidence for the use of these miRNAs as a non-invasive method of diagnosing clinical osteoporosis, especially in premenopausal patients.

**Keywords** MicroRNAs, Osteoporosis, BMD, QUS, c-BUA, Serum bone markers

\*Correspondence:

Amir Iqbal  
physioamir@gmail.com

<sup>1</sup>Department of Clinical Laboratory Sciences, College of Applied Medical Sciences, King Saud University, Riyadh 11433, Saudi Arabia

<sup>2</sup>Department of Rehabilitation Sciences, College of Applied Medical Sciences, King Saud University, Riyadh 11433, Saudi Arabia



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## Introduction

Bone loss has been recognized as a global health problem [1], and there have been increasing trends of low bone mass. Conditions such as osteoporosis and osteopenia, have been reported to be associated with bone fracture risk [2, 3], age, and many basic metabolic disorders, such as dyslipidemia and obesity. In addition, poor lifestyle factors, such as irregular nutrition, hormonal status, toxin exposure, medications, lower physical activity, and educational levels, are significantly associated with higher ratios of bone mineral density (BMD) loss [4–9]. Low bone mineral density (BMD) mainly results from increased bone resorption, performed by osteoclasts, compared with bone formation, performed by osteoblasts [10].

Several biochemical markers such as deoxypyridinoline (DPD), related to the bone resorption process, and both osteocalcin (OC) and bone-specific alkaline phosphatase (BAP), related to bone formation, have been validated as useful markers of bone metabolism, bone loss, and osteoporosis [11–15]. Their changes are easily estimated in the urine, serum, or plasma. They have been significantly linked to the diagnosis of bone loss or osteoporosis with validated tools that are used in clinical routine such as X-ray- and ultrasound-based techniques, especially DEXA scans and quantitative ultrasounds (QUS) [16–20].

Although these validated biomarkers have been assessed in bone diseases via DXA [21], especially as early markers of drug efficacy, they still have some specific limitations. These include a lack of normative reference in population databases, poor standardized quality control, and poor sample handling [21], as well as having only a moderate association with bone-strength and fracture-risk conditions [22, 23]. Thus, new novel biomarkers should be evaluated to better diagnose bone diseases such as osteoporosis, strength, and fracture risks in patients.

Recently, circulating microRNAs have been reported to be associated with many bone diseases [24]. They are short non-coding RNAs, which are reported to have regulatory roles in gene expression in most cells and tissues, including muscle and bone [25–27], usually by suppressing translation or destabilizing mRNAs [28]. Previous research studies have reported the involvement of mRNAs in the process of osteoblastogenesis [29–31]. Recently, the upregulation of 49 miRNAs and downregulation of 44 miRNAs were estimated, respectively, during the early to late differentiation stages of osteoclastogenesis [32–34].

However, no studies refer to the correlation between miRNAs as biomarkers of bone diseases and previously validated bone markers, as well as routinely used X-ray- and ultrasound-based techniques. Although molecular

markers are more expensive than the usual bone-identification markers, we hypothesize in this study that miRNAs, as biomarkers of bone loss, might correlate with previously reported bone markers as well as X-ray- and ultrasound-based techniques to provide new molecular insights that could aid in the treatment of bone loss.

Thus, this study aimed to evaluate the expression of a set of mRNAs, namely miR-21, miR-24, miR-100, miR-24a, miR-103-3p, and miR-142-3p, and their proposed roles in the progression of osteoporosis were identified via q-RT-PCR analysis in premenopausal women. In addition, their correlations with OC, sBAP, and DPD bone markers were explored.

## Materials and methods

### Subjects

A total of 85 healthy, non-smoking, premenopausal women aged 25–50 years old were included in this study. Patients with mental or physical functional impairments, hormonal disorders, secondary osteoporosis, liver or kidney diseases, or those taking medications such as anti-psychotics, oral anticoagulants, corticoids, calcium, and vitamin D supplements that interfere with bone turnover were excluded from this study. Based on the clinical features and diagnosis evidence from both the DXA scan (Z-score) and quantitative ultrasound (QUS) analysis, subjects were classified into three groups: Group A (control subjects with normal BMD; Z-score;  $> -2.5$ ;  $n=25$ ); Group B (premenopausal patients with low BMD; Z-score;  $-1$  SD to  $-2.5$  SD;  $n=30$ ); Group C (premenopausal subjects with low BMD; Z-score;  $\leq -2.5$ ;  $n=30$ ). For all participants, standard anthropometric measurements, i.e., BMI, WHR, and WC, were estimated as previously reported [48].

Regarding the ethical guidelines of the 1975 Declaration of Helsinki, the study protocol was reviewed and approved by the Ethics Sub-Committee of the King Saud University, Kingdom of Saudi Arabia, under file number ID: RRC-2016-049. All participants signed a written informed consent document before data collection. Blood was collected from all subjects, and serum samples were obtained following centrifugation for 1 min at 1400 rpm. The samples were given a coded study identification number and were shipped frozen at  $20^{\circ}$  C for analysis. The demographic and clinical data of the participants are shown in Table [1].

### Assessment of bone mineral density (BMD)

The BMD ( $\text{g}/\text{cm}^2$ ) of the lumbar spine (L2-L4) and proximal femur of the participants was estimated using the Dual Energy X-ray Absorptiometry (DEXA, UNIGAMMA PLUS AC 230 V 50/60Hz 400w, USA) scan method. Osteoporosis was diagnosed among participants using the DEXA method according to their Z-score:

Group A (control subjects with normal BMD; Z-score;  $> -2.5$ ;  $n=25$ ); Group B (premenopausal patients with mild low BMD; Z-score;  $-1$  SD to  $-2.5$  SD;  $n=30$ ); Group C (premenopausal subjects with moderate low BMD; Z-score;  $\leq -2.5$ ;  $n=30$ ) [35].

#### Calcaneus Broadband Ultrasound attenuation test (C-BUA)

As mentioned previously, a commercially available Achilles ultrasound densitometer (Lunar Corporation, Madison, WI, USA) was used to estimate bone health among all participants [36, 37]. This technique was used briefly to measure the c-BUA values in patients with osteoporosis [16–19]. As suggested in clinical practice, patients with poor bone health are those with a Z-score cut-off of  $\leq -1.5$  SD [38], and the coefficient of variation (CV) for c-BUA values ranges from 0.69 to 1.8% during the day of measurement, as mentioned in the literature [39].

#### Assessment of bone markers

Enzyme immunoassay kits (Metra Biosystems, Mountain View, CA, USA) were used to estimate the levels of DPD as a bone resorption marker in the urine samples of participating patients. Based on the urinary creatinine concentration, the DPD values of all subjects were normalized and expressed as nmol DPD/mmol creatinine. The patients were classified according to the grading scores of DPD into the following categories: normal BMD (DPD; 3.4–7.4), low BMD loss (DPD; 7.4–15), higher BMD loss (DPD; 16–30), and very high BMD loss (DPD;  $>30$ ), whereas subjects with higher DPD values were supposed to have osteoporosis, as previously reported [40]. In addition, both serum BAP (sBAP) and osteocalcin (OC) were measured in the serum samples of patients using an enzyme immunoassay kit (Alka phase, Metra Biosystems) for sBAP and the MicroVue Osteocalcin enzyme immunoassay (QUIDEL Corporation, San Diego, CA) for serum OC, respectively [35, 41].

#### Isolation of miRNAs and RT-PCR

For each subject, the total RNA was extracted from serum samples using the TRIzol LS reagent (Invitrogen, Carlsbad, CA), and subjected to RT-PCR analysis. A ready-made solution containing the primers and probes for human miR-34a, miR-142-3p, miR-103-3p, miR-21, miR-24, and miR-100 (Applied Biosystems, Foster City, CA), and real-time RT-PCR, was estimated using an ABI 7300 system (Applied Biosystems) [42]. RNU43 was used as an endogenous reference control, and all PCR cycles were performed according to the manufacturer's instructions as previously described [43], whereas the relative quantification of miRNAs was performed using the  $2^{-\Delta\text{Ct}}$  method. To avoid errors and to precisely determine the cycle threshold mean values for each sample,

including amplified miRNAs and the endogenous control, all reactions were run in duplicate.

#### Statistical analyses

The power calculations of the selected sample size of 85 subjects gave an estimated power of 95% and a significance level of 0.05 with an expected frequency of 8.6%.

The Shapiro–Wilk test was performed for normal data distribution and was logarithmically subjected to statistical analyses. To measure the differences between the studied groups of subjects, both Student's t-test and ANOVA, followed by Bonferroni's multiple comparison analysis, were used. The microRNA levels were adjusted for comparison via univariate analysis, using a general linear model for the different groups. Multiple stepwise regressions and Pearson's correlation analysis were used to estimate the associations between microRNA levels, obesity, and OC, DPD, and sBAP bone parameters.

#### Results

A total of 85 premenopausal patients were recruited in this study. Based on clinical features and diagnosis evidence from both the DXA scan (Z-score) and quantitative ultrasound (QUS) analysis, about 29.4% of the study subjects ( $n=25$ ) showed normal bone health (DXA Z-score;  $> -1$ ; c-BUA; Z-score;  $> -1.5$ ;  $n=25$ ), and 70.4% of the study population showed abnormal bone health with a low BMD; they were classified into two groups: premenopausal patients with mild low BMD (group B; DXA Z-score;  $-1$  SD to  $-2.5$  SD; c-BUA; Z-score  $\leq -1.5$ ;  $n=30$ ) and premenopausal subjects with moderate low BMD (group C; DXA Z-score;  $\leq -2.5$ ; c-BUA; Z-score  $\leq -1.5$ ;  $n=30$ ), as shown in Table 1]. BMI and WHR, as markers of obesity, showed a significant increase in patients with osteopenia and osteoporosis compared to healthy controls.

Also, the values of c-BUA, SOS, and BMD outcome measures for bone health showed a significant decrease in patients with mild ( $P<0.01$ ) and moderate ( $P<0.001$ ) low BMD, respectively, compared to controls with normal BMD. Conversely, OC, sBAP, and DPD as serum markers of bone metabolism showed a significant increase in both patients with mild BMD ( $P<0.01$ ) and moderate BMD ( $P<0.001$ ) [Table 1]. BMD, c-BUA, and SOS, as outcome variables of bone health, were significantly correlated with obesity and markers of bone metabolism. They correlated negatively with obesity (BMI;  $P<0.001$ ) and positively ( $P<0.001$ ) with OC, s-BAP, and DPD in patients with mild ( $P<0.01$ ) and moderate BMD, respectively, as shown in Table 2.

MicroRNAs as biomarkers for bone health were estimated in all subjects (Fig. 1). A set of miRNAs, including miR-21, miR-24, miR-100, miR-24 a, miR-103-3p, and miR-142-3p, was estimated in all subjects via quantitative

**Table 1** Demographic and clinical data of the participants (N = 85)

Parameters		Group (A) Controls (n = 25; 29.4%)	Group (B) Premenopausal with mild low BMD (n = 30; 35.3%)	Group (C) Premenopausal with moderate low BMD (n = 30; 35.3%)
Age (years)		43.4 ± 2.7	43.8 ± 1.9	43.5 ± 2.6
BMI (kg/m <sup>2</sup> )		24.5 ± 1.6	26.2 ± 3.8 <sup>a</sup>	26.7 ± 2.7 <sup>a</sup>
WHR		0.78 ± 0.38	0.89 ± 0.68 <sup>a</sup>	0.92 ± 0.81 <sup>a</sup>
Waist (cm)		79.3 ± 10.5	86.4 ± 6.5 <sup>a</sup>	91.4 ± 8.4
TBMD		1.4 ± 0.98	0.98 ± 0.86 <sup>a</sup>	0.89 ± 0.058 <sup>b</sup>
BMD (Z Score)	Lumbar spine	1.5 ± 0.75	-1.85 ± 0.85 <sup>a</sup>	-2.15 ± 1.2 <sup>a</sup>
	Femoral neck	1.2 ± 1.3	-1.89 ± 0.8 <sup>a</sup>	-2.089 ± 0.5 <sup>a</sup>
c-BUA (dB/MHz)		59.6 ± 2.3	48.1 ± 1.4 <sup>a</sup>	39.5 ± 1.2 <sup>b</sup>
SOS (m/s)		1645.2 ± 4.5	1638.4 ± 3.1 <sup>a</sup>	1635.9 ± 18.1 <sup>b</sup>
OC (ng/ml)		12.7 ± 3.2	18.3 ± 2.5 <sup>a</sup>	26.1 ± 4.7 <sup>b</sup>
DPD (nmol/mmol creatinine)		5.48 ± 1.05	6.8 ± 3.8 <sup>a</sup>	12.3 ± 1.78 <sup>b</sup>
sBAP (U/l)		14.8 ± 3.1	16.9 ± 4.2 <sup>a</sup>	21.2 ± 3.8 <sup>b</sup>

Data expressed as mean ± SD, median (interquartile range), standard deviation (SD); body mass index (BMI); waist-to-hip ratio (WHR); bone mineral density (BMD); broadband ultrasound attenuation (BUA); speed of sound (SOS); serum bone alkaline phosphatase (s-BAP). DPD: deoxypyridinoline; osteocalcin level (ng/ml). <sup>a</sup> P < 0.05; <sup>b</sup> P < 0.01

**Table 2** Association of BMD and c-BUA as outcome variables of bone health with adiposity, serum levels of bone resorption, and formation markers in premenopausal patients with bone loss (varying ranges of low BMD) (n = 60; 70.6%)

Parameters	Bone-health status {BMD scores <sup>(c)</sup> , c-BUA <sup>(d)</sup> }					
	Control		Mild low BMD		Moderate Low BMD	
	R <sup>2</sup> (β) <sup>a</sup>	95% CI	R <sup>2</sup> (β) <sup>b</sup>	95% CI	R <sup>2</sup> (β) <sup>b</sup>	95% CI
BMI	-2.3(0.21)	91(89–96)	-3.7(0.12)	94(89–100)	-3.8(0.28)	89(75–96)
OC	4.2(0.15)	86(78–97)	5.8(0.31)	94(88–100)	5.8 (0.58)	91(88–100)
DPD	5.1(0.17)	95(87–100)	7.5(0.42)	96(89–100)	9.4(0.68)	94(89–100)
sBAP	3.7(0.32)	90(86–98)	6.8(0.48)	89(84–100)	7.5(0.75)	86 (78–100)
OC	2.7(0.38)	89(78–97)	8.7(0.56)	96(86–100)	10.1(0.86)	96 (87–100)

Beta coefficient (β) and cumulative R<sup>2</sup>\* derived from the stepwise regression analysis model showed additional significant variables that were added to the model via bivariate analysis. BMI: body mass index; OC: osteocalcin; sBAP: bone alkaline phosphatase; BMD: bone mineral density; DPD: deoxypyridinoline; SOS: speed of sound; c-BUA: calcaneus quantitative ultrasound parameter. (c) BMD Z-score; Group A (control subjects with normal BMD; Z-score; > -2.5; n = 25); Group B (premenopausal patients with mild low BMD; Z-score; -1 SD to -2.5 SD; n = 30); Group C (premenopausal subjects with moderate low BMD; Z-score; ≤ -2.5; n = 30); (d) c-BUA; Z-score; normal or healthy bone (> -1.5); poor bone or unhealthy bone (Z-score ≤ -1.5). ΣR<sup>2</sup>=summation of cumulative values of R relating to studied variables. \*P < 0.01; \*\*P < 0.001

RT-PCR analysis. MiR-21, miR-24, and miR-100 were significantly (P < 0.001) upregulated and highly expressed in patients with mild and moderate low BMD compared to control subjects with normal BMD [Fig. 1A, B & C]. Conversely, miR-24 a, miR-103-3p, and miR-142-3p were significantly downregulated and reduced in their expression among patients with mild and moderate low BMD compared to respective control subjects with normal BMD [Fig. 2A, B & C]. The results showed that the varied expression of different microRNAs could have a pathogenic role in the degree of bone loss measured by variations in low BMD.

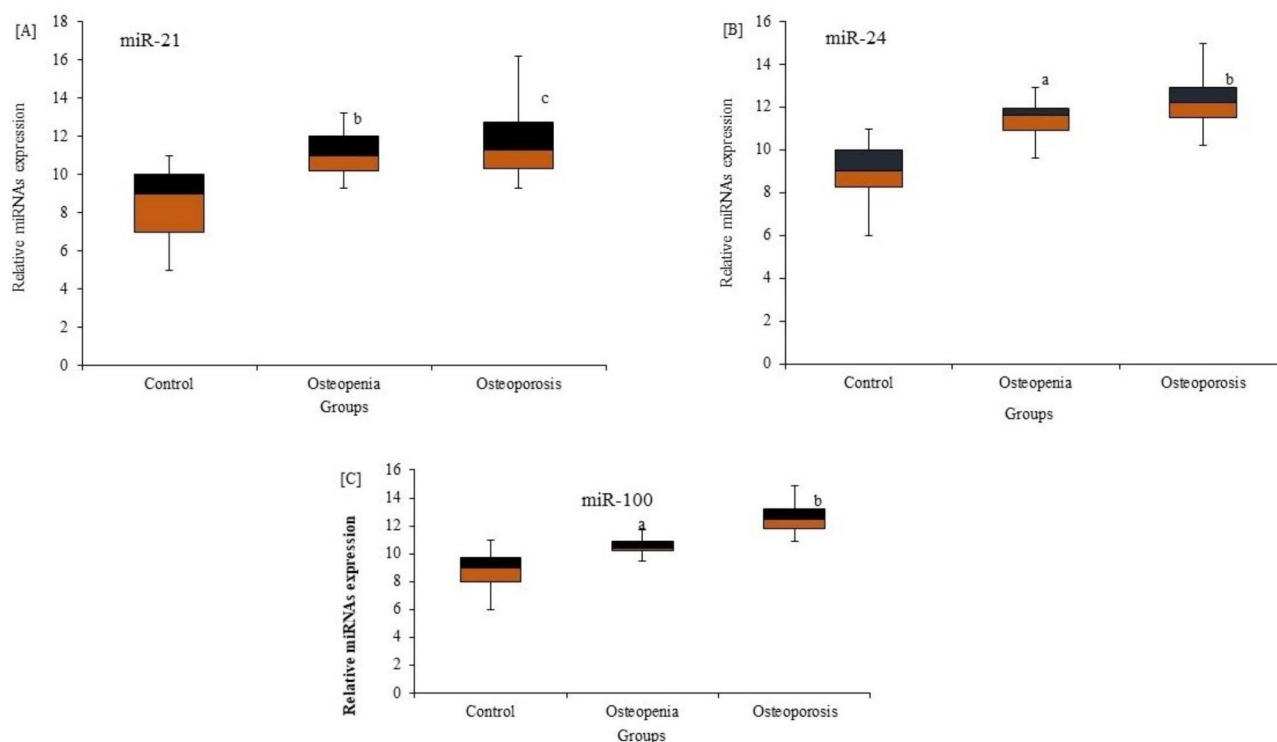
Correlation coefficient analysis showed that both the upregulation and downregulation of the expressed miRNAs were significantly correlated with the serum levels of OC, s-BAP, and DPD as biomarkers of bone metabolism [Table 3]. The bone metabolism markers, c-BUA, and BMD outcome measures for bone health correlated positively with the upregulated miRNAs, miR-21, miR-24,

miR-100, and negatively with the downregulated miRNAs, miR-24 a, miR-103-3p, and miR-142-3p, as shown in Table [3] and Table [4].

## Discussion

Bone health is controlled by a homeostatic dynamic equilibrium between the bone formation and bone resorption processes, which are mediated by osteoblasts and osteoclasts [44, 45]. Any changes or imbalances that occur in this equilibrium result in bone mass reduction, micro-architecture deterioration with an increased probability of bone fragility, and fracture risk [46]. Osteoporosis and osteopenia, as increasing trends related to low bone mass, were significantly reported in association with the probable risks of bone fracture [2, 3].

Bone loss, as measured using BMD and a c-BUA analysis, was the main finding present in this study. It was predicted that 70.6% of premenopausal patients could be classified into two groups: osteopenia and osteoporosis.



**Fig. 1** Differential expression of serum miRNAs in premenopausal females with varying degrees of bone loss (L-BMD) compared to healthy controls. The expression of miRNA-21 (A), miRNA-24 (B), and miRNA-100 (C) was significantly increased (upregulated) in female patients with different bone-loss scores indicated by lower BMD, mild L-BMD ( $P=0.01$ ), and moderate L-BMD ( $p=0.001$ ) compared to healthy control subjects. <sup>a</sup>  $p < 0.05$ , <sup>b</sup>  $p < 0.01$ , <sup>c</sup>  $p < 0.001$  for the comparison indicated by Mann–Whitney U test. Group A: healthy subjects with normal BMD; Group B: premenopausal subjects with mild L-BMD; Group C: premenopausal subjects with moderate L-BMD

Only 29.45% of the study population was characterized by healthy bone parameters. These data were confirmed by a significant rise in the serum levels of OC, s-BAP, and DPD in the urine of patients with mild and moderate bone loss, indicated by a lower BMD, compared with healthy controls. In addition, bone loss, as measured using BMD and c-BUA, correlated positively with OC, s-BAP, and DPD, and negatively with obesity-related markers such as BMI. Both BMD and ultrasound of the calcaneus (c-BUA), as screening tests for osteoporosis and bone loss and the prediction of hip fractures, have been efficiently evaluated in many studies [47–50]. Additionally, the measurements of urinary DPD and serum OC, and s-BAP, have been proposed as inexpensive bone metabolism markers for the screening of hip and spine osteoporosis [11–15, 51]. Also, previous prospective studies indicate c-BUA as a good predictor of bone fractures among older men and women [52, 53]. Although c-BUA, measured using QUS, could be a pre-screening tool for assessing osteoporosis and may reduce the requirement of using the DXA scan, more research is needed to establish its clinical use [54].

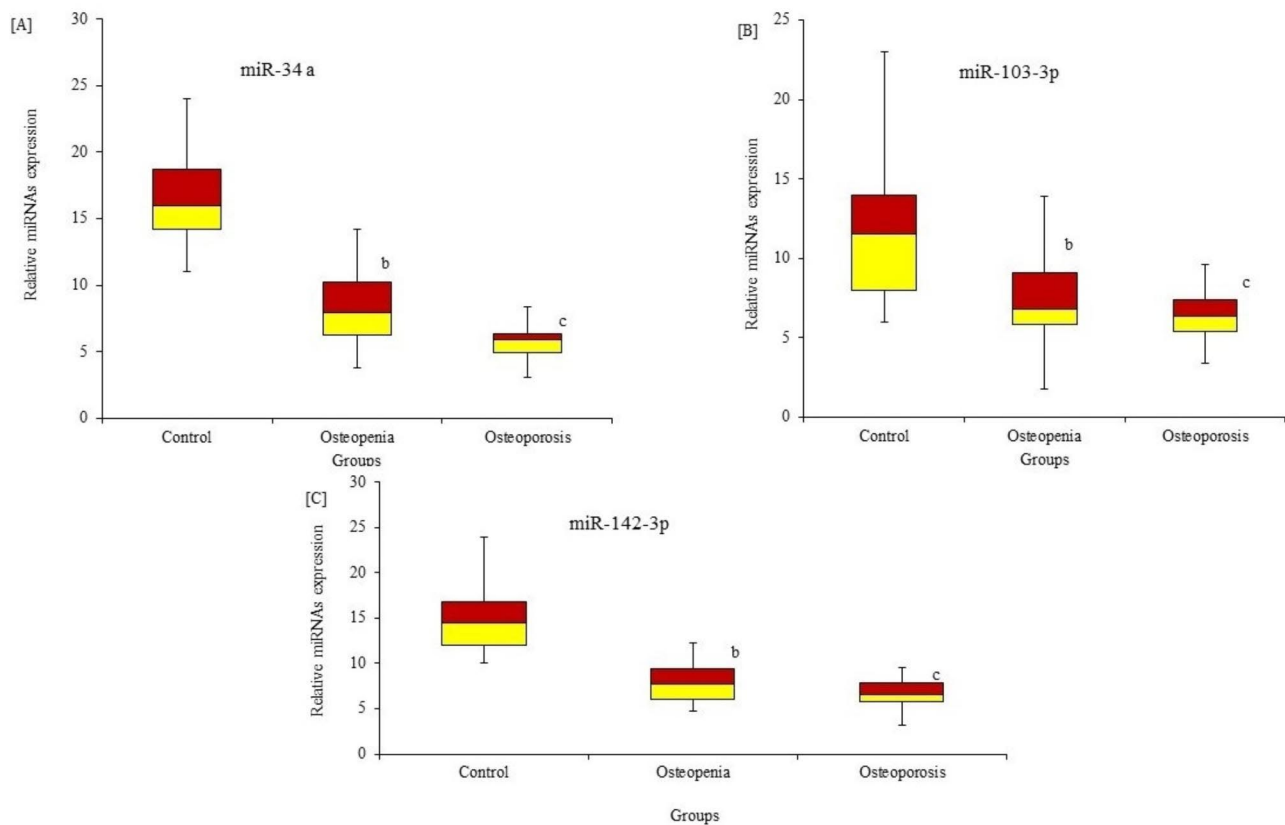
Our study provides suitable combinations of BMD, c-BUA as a static feature of the skeleton, and serum and urine bone biomarkers, such as OC, s-BAP, or DPD,

which provide a dynamic measure of the bone remodeling unit as previously reported [55]; these combinations may be helpful in the assessments of osteoporotic risk fractures [56], and especially in the diagnosis of osteoporosis or bone loss in premenopausal patients.

Although the serum biomarkers of bone formation and resorption still have remarkable roles in the diagnosis of bone loss among osteoporotic patients of both genders [11–15, 51], we still have to identify new specific biomarkers that could either alone, or in combination with BMD and c-BUA analyses, provide a better understanding of bone homeostasis or bone loss in these patients. Thus, identifying the signatures of new specific markers, such as miRNAs in osteoporosis, may provide us with more important cell-based information related to bone-loss mechanisms.

Recently, circulating microRNAs have been reported to be associated with many bone diseases [24]. They are short non-coding RNAs which are reported to have regulatory roles in gene expression in most cells and tissues, including muscle and bone [25–27], usually by suppressing translation or destabilizing mRNAs [28].

In this study, we tried to investigate the expression of a set of miRNAs, miR-21, miR-24, miR-100, miR-24a, miR-103-3p, and miR-142-3p, in premenopausal patients



**Fig. 2** Differential expression of serum miRNAs in premenopausal females with varying degrees of bone loss (L-BMD) compared to healthy controls. The expressions of miRNA-34 a (A), miRNA-103-3p (B), and miRNA-142-3p (C) were significantly reduced (downregulated) in the serum of premenopausal females with mild (P=0.01) and moderate (p=0.001) bone loss (L-BMD) compared to healthy control subjects. <sup>a</sup> p < 0.05, <sup>b</sup> p < 0.01, <sup>c</sup> p < 0.001 for the comparison indicated by Mann–Whitney U test. Group B: premenopausal subjects with mild L-BMD; Group C: premenopausal subjects with moderate L-BMD

**Table 3** Correlation analysis of miRNA expression levels with bone formation and resorption markers in premenopausal patients with varying ranges of low BMD (n = 60; 70.6%)

Parameters	OC		sBAP		DPD	
	R <sup>2</sup> (β) <sup>a</sup>	95% CI	R <sup>2</sup> (β) <sup>b</sup>	95% CI	R <sup>2</sup> (β) <sup>b</sup>	95% CI
<b>Mild low BMD (n = 30)</b>						
Upregulated miRNAs						
miR-21	2.3(0.19)	89 (78–96)	3.5(0.59)	92 (88–96)	7.1(0.65)	91 (84–98)
miR-24	3.6(0.16)	92 (86–98)	5.1(0.35)	97 (87–100)	4.8(0.48)	95 (89–98)
miR-100	4.6(0.27)	85 (75–92)	8.4(0.58)	84 (78–94)	9.1(0.68)	89 (78–96)
Downregulated miRNAs						
miR-34 a	7.2(0.32)	75 (71–89)	8.1(0.65)	76 (81–92)	6.4(0.75)	78 (79–95)
miR-103-3p	6.4(0.28)	96 (85–98)	8.9(0.34)	90 (88–100)	7.1(0.81)	93 (88–100)
miR-142-3p	8.1(0.52)	90 (81–96)	6.2(0.57)	93 (85–98)	5.1(0.49)	96 (85–98)
<b>Moderate low BMD (n = 30)</b>						
Upregulated miRNAs						
miR-21	4.9(0.45)	86 (78–96)	4.7(0.71)	96 (89–100)	5.3(0.81)	95 (86–98)
miR-24	7.5(0.36)	94 (86–98)	8.3(0.78)	86 (78–96)	6.8(0.56)	93 (87–98)
miR-100	5.3(0.34)	81 (75–92)	5.2(0.65)	81 (78–94)	7.1(0.78)	91 (84–96)
Downregulated miRNAs						
miR-34 a	6.2(0.46)	88 (71–96)	9.3(0.78)	78 (81–96)	8.9(0.82)	91 (87–96)
miR-103-3p	3.1(0.31)	91 (88–97)	5.2(0.43)	95 (85–100)	7.5(0.86)	96 (89–100)
miR-142-3p	5.6(0.58)	92 (85–95)	7.2(0.63)	97 (85–100)	8.3(0.67)	90 (86–98)

Beta coefficient (β) and cumulative R2\* derived from the stepwise regression analysis model showed additional significant variables added to the model via bivariate analysis. OC: osteocalcin; sBAP: bone alkaline phosphatase; DPD: deoxypyridinoline. <sup>a</sup>P<0.01; <sup>b</sup>P<0.001

**Table 4** Association of BMD and c-BUA as outcome variables of bone health with miRNA expression levels in premenopausal patients with varying ranges of low BMD (n = 60; 70.6%)

Parameters	{BMD scores <sup>(c)</sup> }		{c-BUA <sup>(d)</sup> }	
	R <sup>2</sup> (β) <sup>a</sup>	95% CI	R <sup>2</sup> (β) <sup>b</sup>	95% CI
<b>Mild low BMD (n = 30)</b>				
Upregulated miRNAs				
miR-21	4.1(0.12)	86 (80–96)	4.8(0.23)	89 (86–96)
miR-24	5.6(0.21)	94 (88–98)	6.1(0.17)	91 (82–100)
miR-100	4.9(0.32)	89 (73–99)	5.7(0.28)	94 (78–99)
Downregulated miRNAs				
miR-34 a	-5.1(0.32)	78 (68–91)	-7.3(0.34)	84 (79–98)
miR-103-3p	-3.8(0.31)	96 (81–98)	-4.8(0.36)	92 (88–100)
miR-142-3p	-6.7(0.58)	99 (87–100)	-6.9(0.38)	94 (85–100)
<b>Moderate low BMD (n = 30)</b>				
Upregulated miRNAs				
miR-21	6.2(0.25)	87 (76–96)	4.7(0.71)	96 (89–100)
miR-24	5.4(0.51)	92 (86–98)	8.3(0.78)	86 (78–96)
miR-100	5.8(0.38)	86 (78–96)	5.2(0.65)	81 (78–94)
Downregulated miRNAs				
miR-34 a	-8.1(0.58)	94(78–100)	-10.1(0.84)	91 (88–98)
miR-103-3p	-6.4(0.71)	89 (75–98)	-8.5(0.91)	89 (81–100)
miR-142-3p	-7.3(0.91)	90 (88–100)	-6.7(0.65)	95 (85–100)

Beta coefficient (β) and cumulative R<sup>2</sup>\* derived from the stepwise regression analysis model showed additional significant variables added to the model via bivariate analysis. BMD: bone mineral density; c-BUA: calcaneus quantitative ultrasound parameter. (c) BMD Z-score; Group A (control subjects with normal BMD; Z-score; > -2.5; n = 25); Group B (premenopausal patients with mild low BMD; Z-score; -1 SD to -2.5 SD; n = 30); Group C (premenopausal subjects with moderate low BMD; Z-score; ≤ -2.5; n = 30); (d) c-BUA; Z-score; normal or healthy bone (> -1.5); poor bone or unhealthy bone (Z-score ≤ -1.5).<sup>a</sup>P < 0.01; <sup>b</sup>P < 0.001

with varying degrees of bone loss, indicated by a low BMD, and evaluated its correlation with the scores of BMD, c-BUA, and the biomarkers of bone metabolism. The data showed that miR-21, miR-24, and miR-100 were significantly upregulated and highly expressed in female patients with mild and moderate low BMD compared with controls with normal BMD and healthy bone parameters. Also, the data showed that during osteoporosis, there was a significant downregulation and lower expression of miR-24a, miR-103-3p, and miR-142-3p in association with bone loss status compared to normal cases.

Previous research studies reported on the involvement of mRNAs in the osteoblast genesis process [29–31]. Recently, the upregulation of 49 miRNAs and downregulation of 44 miRNAs were estimated, respectively, during the early to late differentiation stages of osteoclast genesis [32–34]. The interest in miRNAs has come from their associations with physiological or disease conditions, especially regarding their expression and specific signatures in cancer [57]. Thus, miRNAs in the plasma/serum could be used as prognostic circulating biomarkers in many diseases [57–59].

Also, many research studies indicate the potential role of miRNA expression in the function, differentiation, and development of bone in normal and abnormal bone diseases [60–64]. Consistent with our results, previous studies have showed the upregulation of nine miRNAs, including miR-21, miR-24, and miR-100 [65],

and downregulation of miR-24a, miR-103-3p, and miR-142-3p [66, 67] in the serum of patients with osteoporosis. The data showed that there was a significant increase in upregulated miRNAs, and that downregulated miRNAs were only significantly decreased in osteoporosis, and that they showed a significant association with BMD. Thus, we can conclude that miR-21, miR-24, and miR-100 are potent inhibitors of bone formation in premenopausal patients with osteoporosis, and miR-24a, miR-103-3p, and miR-142-3p are potent activators of bone formation and are thus significantly increased in healthy bone subjects.

Recently, a circulating miRNAs analysis was performed in patients with osteopenia, osteoporosis, and fragility fractures. In that study, downregulated miR-21 and upregulated miR-133a miRNAs were estimated as potential biomarkers for postmenopausal osteoporosis. These markers showed a moderate to strong correlation with BMD [68]. Although many studies have correlated the expression of miRNAs in the plasma and serum of BMD osteoporotic patients, little is known about the correlation between miRNA expression and c-BUA and serum bone markers.

Thus, our study may be the first to evaluate the correlation between circulating miRNAs c-BUA, OC, s-BAP, and DPD in premenopausal patients with varying degrees of bone loss as measured by lower scores of BMDs. The data showed that the expression of both upregulated miR-21, miR-24, miR-100, and downregulated

miR-24a, miR-103-3p, and miR-142-3p correlated positively with bone resorption (DPD) and formation (OC, s-BAP) markers. Similarly, a quantitative estimation of c-BUA and SOS as parameters of BMD in osteoporotic female patients showed a significant correlation with the expressed miRNAs. The values of c-BUA measured using QUS correlated positively with upregulated miR-21, miR-24, and miR-100 and negatively with downregulated miR-24a, miR-103-3p, and miR-142-3p, respectively. The data obtained signify the importance of these miRNAs in osteogenic differentiation in different cell types, which may indicate the homeostatic imbalance between the bone resorption and formation processes that are controlled by osteoclasts and osteoblasts, respectively [60–65].

Based on our results, we propose that miRNAs could have a potential role in the pathogenesis of bone loss. The miRNAs studied were significantly correlated with BMD, c-BUA, and serum bone markers, providing sufficient sensitivity and specificity for distinguishing females with premenopausal osteoporosis.

These new insights into using miRNAs as molecular targets of osteoporosis progression among premenopausal women could be used in the exploration of other related diseases, such as endometrial diseases, and especially precancerous endometrial cancer (EC) lesions involving premenopausal and nulliparous women, or those with pregnancy plans who may prefer more conservative treatment [69]. There are few studies available in the literature that analyze how molecular classification could explain the potential mechanisms related to high EC evolution risk among premenopausal women [70, 71]. It was reported recently that different histopathological or molecular features are present in endometrial diseases with a variety of pathologies [69]. The use of molecular markers has offered the possibility to improve the risk stratification and management of EC [69, 72]. Consequently, detecting and validating the use of different molecular markers like miRNAs in precancerous lesions and associated diseases like premenopausal osteoporosis could change therapeutic strategies, increasing the follow-up of fertility-sparing patients, or tailoring surgical radicality. In addition, molecular markers, such as miRNAs and genomic profiling, might be useful in choosing the most appropriate adjuvant strategies in apparent early-stage EC in pre- and postmenopausal women.

This study has two limitations. Firstly, the recruited bone tissue samples were from female patients; therefore, we were unable to make a sex-comparison analysis of the data. Secondly, although the study is a pilot study with a low sample size, we still need more studies to establish the correlation mechanisms of circulating miRNA biomarkers with high specificity and sensitivity in the pathogenesis of premenopausal osteoporosis.

Nevertheless, our study may be the first to clearly demonstrate the correlation between the profiling of molecular miRNA in premenopausal patients and the scoring rates of bone loss measured using BMD and traditionally used parameters, including c-BUA, OC, s-BAP, and DPD. It also shows that miRNA expression may have potential roles in the detection and classification of osteoporotic diseases.

## Conclusion

A group of circulating miRNAs was shown to be closely correlated with traditionally used parameters, i.e., BMD, c-BUA, OC, s-BAP, and DPD, in bone-health measurement. Additionally, they were identified as non-invasive biomarkers in premenopausal patients with osteoporosis. More studies with large sample sizes are recommended to estimate the mechanistic role of miRNAs in the pathogenesis of osteoporosis and to provide evidence for the use of these miRNAs as a non-invasive method of diagnosing clinical bone loss, especially in premenopausal patients.

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## Authors' contributions

G.S.A. H.A.R. A.H.A. and A.I. proposed the study conception and design. G.S.A. completed the practical work. G.S.A. collected data. A.I. contributed to the data analysis. H.A.R. A.H.A. G.S.A. and A.I. contributed to data interpretation. G.S.A. H.A.R. and A.I. prepared the manuscript's initial draft. H.A.R. and A.H.A. critically reviewed and edited the intellectual content of the manuscript. All authors read, understood, reviewed, and approved the manuscript's final version to be submitted/published and took responsibility for the intellectual content of the same manuscript.

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## Data Availability

All data generated or analyzed during this study are presented in the manuscript. Please contact the corresponding authors for access to the data presented in this study.

## Declarations

### Ethics approval and consent to participate

Regarding the ethical guidelines of the 1975 Declaration of Helsinki, the study protocol was reviewed and approved by the Ethics Sub-Committee of King Saud University, Kingdom of Saudi Arabia, under file number ID: RRC-2016-049. All participants signed a written informed consent document before data collection.

### Consent for publication

Not applicable.

### Competing interests

The authors declare that they have no competing interests, financial or non-financial.

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## References

- Consensus development conference. : diagnosis, prophylaxis, and treatment of osteoporosis (1993) *Am J Med* 94(6):646–650.
- Johnell O, Kanis JA. An estimate of the worldwide prevalence and disability associated with osteoporotic fractures. *Osteoporos Int*. 2006;17:1726–33.
- Jang EJ, Lee YK, Choi HJ, et al. Osteoporotic fracture risk assessment using bone mineral density in Korean: a community-based cohort study. *J Bone Miner Metab*. 2016;23:34–9.
- New SA. Exercise, bone and nutrition. *Proc Nutr Soc*. 2001;60:265–74.
- Espallargues M, Sampietro-Colom L, Estrada MD, et al. Identifying bone-mass-related risk factors for fracture to guide bone densitometry measurements: a systematic review of the literature. *Osteoporos Int*. 2001;12:811–22.
- Tang YJ, Sheu WH, Liu PH, et al. Positive associations of bone mineral density with body mass index, physical activity, and blood triglyceride level in men over 70 years old: a TCVGHAGE study. *J Bone Miner Metab*. 2007;25:54–9.
- Varena M, Binelli L, Zucchi F, et al. Prevalence of osteoporosis by educational level in a cohort of postmenopausal women. *Osteoporos Int*. 1999;9:236–41.
- Kanis JA, Johansson H, Oden A, Johnell O, de Laet C, Melton IL, Tenenhouse A, Reeve J, Silman AJ, Pols HA, Eisman JA, McCloskey EV, Mellstrom D. A meta-analysis of prior corticosteroid use and fracture risk. *J Bone Miner Res*. 2004;19(6):893–9.
- Briot K, Kolta S, Flandre P, Boue F, Ngo Van P, Cohen-Codar I, Norton M, Delraissy JF, Roux C. Prospective one-year bone loss in treatment-naïve HIV + men and women on single or multiple drug HIV therapies. *Bone*. 2011;48(5):1133–9.
- Manolagas SC. Birth and death of bone cells: basic regulatory mechanisms and implications for the pathogenesis and treatment of osteoporosis. *Endocr Rev*. 2000;21:115–1.
- Garnero P, Delmas PD. New developments in biochemical markers for osteoporosis. *Calcif Tissue Int*. 1996;59:2–59.
- Geborek P, Crnkic M, Petersson IF, et al. South Swedish Arthritis Treatment Group: Etanercept, infliximab, and leflunomide in established rheumatoid arthritis: clinical experience using a structured follow up programme in southern Sweden. *Ann Rheum Dis*. 2002;61:793–8.
- van der Woude D, Young A, Jayakumar K, et al. Prevalence of and predictive factors for sustained disease-modifying antirheumatic drug-free remission in rheumatoid arthritis: results from two large early arthritis cohorts. *Arthritis Rheum*. 2009;60:2262–71.
- Vs K, K P, Ramesh M, Venkatesan V. The association of serum osteocalcin with the bone mineral density in post-menopausal women. *J Clin Diagn Res*. 2013;7(5):814–6.
- Singh S, Kumar D, Lal AK. Serum osteocalcin as a diagnostic biomarker for primary osteoporosis in women. *J Clin Diagn Res*. 2015;9(8):RC04–7.
- Albanese CV, De Terlizzi F, Passariello R. Quantitative ultrasound of the phalanges and DXA of the lumbar spine and proximal femur in evaluating the risk of osteoporotic vertebral fracture in postmenopausal women. *Radiol Med*. 2011;116:92–101.
- Pisani P, Renna MD, Conversano F, Casciaro E, Muratore M, Quarta E, Paola MD, Casciaro S. Screening and early diagnosis of osteoporosis through X-ray and ultrasound based techniques. *World J Radiol*. 2013;5(11):398–410.
- Glüer CC, Wu CY, Jergas M, Goldstein SA, Genant HK. Three quantitative ultrasound parameters reflect bone structure. *Calcif Tissue Int*. 1994;55:46–52.
- Mészáros S, Tóth E, Ferencz V, Csopor E, Hosszú E, Horváth C. Calcaneus quantitative ultrasound measurements predicts vertebral fractures in idiopathic male osteoporosis. *Joint Bone Spine*. 2007;74(1):79–84.
- Xu Y, Guo B, Gong J. The correlation between calcaneus stiffness index calculated by QUS and total body BMD assessed by DXA in chinese children and adolescents. *J Bone Miner Metab*. 2014;32:159–66.
- Bauer D, Krege J, Lane N, Leary E, Libanati C, Miller P, Myers G, Silverman S, Vesper HW, Lee D, Payette M, Randall S. National Bone Health Alliance bone turnover marker project: current practices and the need for US harmonization, standardization, and common reference ranges. *Osteoporos Int*. 2012;23:2425–33.
- Ivaska KK, Gerdhem P, Väänänen HK, Akesson K, Obrant KJ. Bone turnover markers and prediction of fracture: a prospective follow-up study of 1040 elderly women for a mean of 9 years. *J Bone Miner Res*. 2010;25:393E403.
- Wheater G, Elshahaly M, Tuck SP, Datta HK, van Laar JM. The clinical utility of bone marker measurements in osteoporosis. *J Transl Med*. 2013;11:201.
- De Guire V, Robitaille R, Tetreault N, Guerin R, Menard C, Bambace N, Sapieha P. Circulating miRNAs as sensitive and specific biomarkers for the diagnosis and monitoring of human diseases: promises and challenges. *Clin Biochem*. 2013;46:846–60.
- Callis TE, Chen J-F, Wang D-Z. MicroRNAs in skeletal and cardiac muscle development. *DNA Cell Biol*. 2007;26:219–25.
- Laxman N, Rubin C, Mallmin H, Nilsson O, Pastinen T, Grundberg E, Kindmark A. 2015. Global miRNA expression and correlation with mRNA levels in primary human bone cells. *2RNA*. 2015;21(8):1433-43. <https://doi.org/10.1261/rna.049148.114>. Epub 2015 Jun 15.
- Lian JB, Stein GS, Wijnen AJ, Van, Stein JL, Hassan MQ, van Wijnen AJ, Gaur T, Zhang Y. MicroRNA control of bone formation and homeostasis. *Nat Rev Endocrinol*. 2013;8:212–27.
- Ebert MS, Sharp PA. Roles for microRNAs in conferring robustness to biological processes. *Cell*. 2012;149:515–24.
- Li Z, Hassan MQ, Volinia S, van Wijnen AJ, Stein JL, et al. A microRNA signature for a BMP2-induced osteoblast lineage commitment program. *Proc Natl Acad Sci U S A*. 2008;105:13906–11.
- Hassan MQ, Gordon JA, Beloti MM, Croce CM, van Wijnen AJ, et al. A network connecting Runx2, SATB2, and the miR-23a,27a,24–2 cluster regulates the osteoblast differentiation program. *Proc Natl Acad Sci U S A*. 2010;107:19879–84.
- Inose H, Ochi H, Kimura A, Fujita K, Xu R, et al. A microRNA regulatory mechanism of osteoblast differentiation. *Proc Natl Acad Sci U S A*. 2009;106:20794–9.
- Franceschetti T, Dole NS, Kessler CB, Lee SK, Delany AM. Pathway analysis of microRNA expression profile during murine osteoclastogenesis. *PLoS ONE*. 2014;9:e107262. <https://doi.org/10.1371/journal.pone.0107262>.
- Tang P, Xiong Q, GeW, Zhang L. The role of microRNAs in osteoclasts and osteoporosis. *RNA Biol*. 2014;11:1355–63. <https://doi.org/10.1080/15476286.2014.996462>.
- Ji X, Chen X, Yu X. MicroRNAs in osteoclastogenesis and function: potential therapeutic targets for osteoporosis. *Int J Mol Sci*. 2016;7:349. <https://doi.org/10.3390/ijms17030349>.
- Alghadir AH, Aly FA, Gabr SA. 2014 Effect of Moderate Aerobic training on bone metabolism indices among adult humans. *Pak J Med Sci*. 30(4):840–4.
- Ramirez-Vélez R, Ojeda-Pardo ML, Correa-Bautista JE et al. 2016 normative data for calcaneal broadband ultrasound attenuation among children and adolescents from Colombia: the FUPRECOL Study. *Arch Osteoporos* 11: 2. <https://doi.org/10.1007/s11657-015-0253-0>.
- Jaworski M, Lebedowski M, Lorenc RS, Trempe J. 1995 Ultrasound bone measurement in pediatric subjects. *Calcif Tissue Int* 56: 368–71.
- Weaver CM, Gordon CM, Janz KF et al. 2016 the national osteoporosis Foundation's position statement on peak bone mass development and lifestyle factors: a systematic review and implementation recommendations. *Osteoporos Int* 27: 1281–386.
- Vignolo M, Brignone A, Mascagni A, Ravera G, Biasotti B, Aicardi G. Influence of age, sex, and growth variables on phalangeal quantitative ultrasound measures: a study in healthy children and adolescents. *Calcif Tissue Int*. 2003;72:681–8.
- Riis Brazier M, Nevi V, et al. Measurement of biochemical markers. *Methods and Limitations J Bone Miner Res*. 1995;10:385–92.
- Alghadir AH, Gabr SA, Al-Eisa ES. 2017 mechanical factors and vitamin D deficiency in schoolchildren with low back pain: biochemical and cross-sectional survey analysis. *J Pain Res* 10:855–65. <https://doi.org/10.2147/JPR.S124859>.
- Mazloom H, Alizadeh S, Pasalar P, Esfahani EN, Meshkani R. Downregulated microRNA-155 expression in peripheral blood mononuclear cells of type 2 diabetic patients is not correlated with increased inflammatory cytokine production. *Cytokine*. 2015;76(2):403–8.
- Kloosterman WP, Plasterk RH. The diverse functions of micro-RNAs in animal development and disease. *Dev Cell*. 2006;11:441–50.
- Bustin SA, Benes V, Garson JA, et al. The MIQE guidelines: minimum information for publication of quantitative real-time PCR experiments. *Clin Chem*. 2009;55(4):611–22.
- Torricelli P, Fini M, Giavaresi G, Giardino R. Human osteoblast cultures from osteoporotic and healthy bone: biochemical markers and cytokine expression in basal conditions and in response to 1,25 (OH)2D3. *Artif Cells Blood Substit Immobil Biotechnol*. 2002;30(3):219–27.
- Kanis JA, et al. European guidance for the diagnosis and management of osteoporosis in postmenopausal women. *Osteoporos Int*. 2013;24:23–57.

47. Marshall D, Johnell O, Wedel H. Metaanalysis of how well measures of bone mineral density predict occurrence of osteoporotic fracture. *Br Med J*. 1996;312:1254–9.
48. Black DM, Cummings SR, Karpf DB, et al. Randomized trial of effect of alendronate on risk of fracture in women with existing vertebral fractures. *Lancet*. 1996;348:1535–41.
49. Hans D, DargentMolina P, Schott AM, et al. Ultrasound heel measurements to predict hip fracture in elderly women: the EPIDOS study. *Lancet*. 1996;348:511–4.
50. Bauer DC, Gluer CC, Cauley JA, et al. Broadband ultrasound attenuation predicts fractures strongly and independently of densitometry in older women. *Arch Intern Med*. 1997;157:629–34.
51. Schneider DL, BarretConner EL. Urinary ntelopeptide levels discriminate normal, osteopenic, and osteoporotic bone mineral density. *Arch Intern Med*. 1997;157:1241–5.
52. Bauer DC, Glüer CC, Cauley JA, et al. Broadband ultrasound attenuation predicts fractures strongly and independently of densitometry in older women. A prospective study. *Arch Intern Med*. 1997;157:629–34.
53. Khaw KT, Reeve J, Luben R, et al. Prediction of total and hip fracture risk in men and women by quantitative ultrasound of the calcaneus: EPIC-Norfolk prospective population study. *Lancet*. 2004;363:197–202.
54. Thomsen K, Jepsen DB, Matzen L, et al. Is calcaneal quantitative ultrasound useful as a prescreen stratification tool for osteoporosis? *Osteoporos Int*. 2015;26:1459–75.
55. Vanitha Jagtap R, Jayashri Ganu V, Nitin Nagane S. BMD and serum intact osteocalcin in post-menopausal osteoporosis women. *Ind J Clin Biochem*. 2011;26(1):70–3.
56. Vs K, Ramesh KP, Venkatesan M. V. The association of serum osteocalcin with the bone mineral density in post-menopausal women. *J Clin Diagn Res*. 2013;7(5):814–6. <https://doi.org/10.7860/JCDR/2013/5370.2946>. Epub 2013 Mar 20.
57. Garzon R, Calin GA, Croce CM. MicroRNAs in Cancer. *Annu Rev Med*. 2009;60:167–79.
58. Kosaka N, Iguchi H, Ochiya T. Circulating microRNA in body fluid: a new potential biomarker for cancer diagnosis and prognosis. *Cancer Sci*. 2010;101(10):2087–92.
59. Wang WT, Zhao YN, Han BW, Hong SJ, Chen YQ. Circulating microRNAs identified in a genome-wide serum microRNA expression analysis as noninvasive biomarkers for endometriosis. *J Clin Endocrinol Metab*. 2013;98(1):281–9.
60. Wang Y, Li L, Moore BT, et al. MiR-133a in human circulating monocytes: a potential biomarker associated with postmenopausal osteoporosis. *PLoS ONE*. 2012;7(4):e34641.
61. Qi Y, Ma N, Yan F et al. The expression of intronic miRNAs, miR-483 and miR-483<sub>+</sub>, and their host gene, Igf2, in murine osteoarthritis cartilage. *Int J Biol Macromol* 2103;61 C:43–9.
62. Eskildsen T, Taipaleenmaki H, Stenvang J, et al. MicroRNA-138 regulates osteogenic differentiation of human stromal (mesenchymal) stem cells in vivo. *Proc Natl Acad Sci USA*. 2011;108(15):6139–44.
63. Hu R, Liu W, Li H, et al. A Runx2/miR-3960/miR-2861 regulatory feedback loop during mouse osteoblast differentiation. *J Biol Chem*. 2011;286(14):12328–39.
64. Goettsch C, Rauner M, Pacyna N, Hempel U, Bornstein SR, Hofbauer LC. miR-125b regulates calcification of vascular smooth muscle cells. *Am J Pathol*. 2011;179(4):1594–600.
65. Seeliger C, Karpinski K, Haug AT, Vester H, Schmitt A, Bauer JS, van Griensven M. Five freely circulating miRNAs and bone tissue miRNAs are associated with osteoporotic fractures. *J Bone Miner Res*. 2014;29(8):1718–28. <https://doi.org/10.1002/jbmr.2175>.
66. Chen J, Li K, Pang Q, et al. Identification of suitable reference gene and biomarkers of serum miRNAs for osteoporosis. *Sci Rep*. 2016;6:36347. <https://doi.org/10.1038/srep36347>.
67. Chen L, Holmström K, Qiu W, Ditzel N, Shi K, Hokland L, Kassem M. MicroRNA-34a inhibits osteoblast differentiation and in vivo bone formation of human stromal stem cells. *Stem Cells*. 2014;32(4):902–12. <https://doi.org/10.1002/stem.1615>.
68. Li H, Wang Z, Fu Q, Zhang J. Plasma miRNA levels correlate with sensitivity to bone mineral density in postmenopausal osteoporosis patients. *Biomarkers*. 2014;19:553–6. <https://doi.org/10.3109/1354750X.2014.935957>.
69. Golia D'Augè, Cuccu T, Santangelo I, Muzii G, Giannini L, Bogani A, Di Donato G. Novel insights into Molecular Mechanisms of Endometrial Diseases. *Biomolecules*. 2023;13(3):499. <https://doi.org/10.3390/biom13030499>. PMID: 36979434; PMCID: PMC10046407.
70. Zhang X, Chen D, Zhao X, Wang C, He Y, Chen Y, Wang J, Shen D. Application of molecular classification to guiding fertility-sparing therapy for patients with endometrial cancer or endometrial intraepithelial neoplasia. *Pathol Res Pract*. 2023;241:154278. <https://doi.org/10.1016/j.prp.2022.154278>.
71. Norimatsu Y, Moriya T, Kobayashi TK, Sakurai T, Shimizu K, Tsukayama C, Ohno E. Immunohistochemical expression of PTEN and be-ta-catenin for endometrial intraepithelial neoplasia in japanese women. *Ann Diagn Pathol*. 2007;11:103–8. <https://doi.org/10.1016/j.anndiagpath.2006.06.009>.
72. Cuccu I, D'Oría O, Sgamba L, De Angelis E, Golia D'Augè T, Turetta C, Di Dio C, Scudo M, Bogani G, Di Donato V, Palaia I, Perniola G, Tomao F, Muzii L, Giannini A. Role of genomic and molecular Biology in the modulation of the treatment of Endometrial Cancer: Narrative Review and Perspectives. *Healthc (Basel)*. 2023;11(4):571. <https://doi.org/10.3390/healthcare11040571>.

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