

RESEARCH

Open Access



Effect of lactobacillus reuteri-derived probiotic nano-formulation on recurrent aphthous stomatitis: a double-blinded randomized clinical trial

Nazafarin Samiraninezhad¹, Hojat Kazemi¹, Mostafa Rezaee^{2,5*} and Ahmad Gholami^{3,4}

Abstract

Objectives We aimed to assess the therapeutic effects of a topical probiotic nano-formulation derived from *Lactobacillus reuteri* on treating recurrent aphthous stomatitis.

Materials and methods 60 participants were randomly allocated into two groups (control and probiotic). Probiotic group administered topical probiotic nano-formulation three times a day for seven days. The control group administered a standard analgesic oral rinse. The size of ulcer(s) and pain severity were recorded on days 0, 3, 5, and 7 after intervention.

Results Before the intervention, the groups had no significant differences in terms of pain severity (P -value = 0.28) and lesion size (P -value = 0.24). Both groups exhibited significant reductions in pain severity and lesion size over the course of the intervention. After one week, the probiotic group had a notably larger lesion size reduction than the control group (P -value = 0.01). The probiotic group also showed a significantly greater reduction in pain severity than the control group (P -value = 0.04).

Conclusions Applying topical probiotic nano-formulation derived from *Lactobacillus reuteri* three times a day decreased lesion size and pain severity in RAS patients faster than the local analgesic oral rinse.

Clinical relevance *Lactobacillus reuteri*-derived probiotic nano-formulation might be a promising treatment option for RAS.

Keywords *Lactobacillus*, *Limosilactobacillus reuteri*, Probiotics, Stomatitis, Aphthous, Chitosan, Nanogels

*Correspondence:

Mostafa Rezaee

rezaim@sums.ac.ir

¹Student Research Committee, Shiraz University of Medical Sciences, Shiraz, Iran

²Department of Oral and Maxillofacial Medicine, School of Dentistry, Shiraz University of Medical Sciences, Shiraz, Iran

³Pharmaceutical Sciences Research Center, Shiraz University of Medical Sciences, Shiraz, Iran

⁴Department of Medical Nanotechnology, School of Advanced Medical Science and Technology, Shiraz University of Medical Sciences, Shiraz, Iran

⁵Oral and Dental Disease Research Center, School of Dentistry, Shiraz University of Medical Sciences, Shiraz, Iran



© The Author(s) 2023. **Open Access** This article is licensed under a Creative Commons Attribution 4.0 International License, which permits use, sharing, adaptation, distribution and reproduction in any medium or format, as long as you give appropriate credit to the original author(s) and the source, provide a link to the Creative Commons licence, and indicate if changes were made. The images or other third party material in this article are included in the article's Creative Commons licence, unless indicated otherwise in a credit line to the material. If material is not included in the article's Creative Commons licence and your intended use is not permitted by statutory regulation or exceeds the permitted use, you will need to obtain permission directly from the copyright holder. To view a copy of this licence, visit <http://creativecommons.org/licenses/by/4.0/>. The Creative Commons Public Domain Dedication waiver (<http://creativecommons.org/publicdomain/zero/1.0/>) applies to the data made available in this article, unless otherwise stated in a credit line to the data.

Introduction

Recurrent aphthous stomatitis (RAS) is a common type of oral ulcer that can occur in the general population, with a reported prevalence range of 0.7–50% [1]. It is more frequently observed in females and typically affects individuals between the ages of 7 to 44 years old [2, 3]. RAS ulcers can be classified into three forms based on their clinical presentation: minor, central, and herpetiform. Minor RAS lesions are the most common, representing over 80% of cases. They are characterized by small painful ulcers (5–10 mm) with a well-defined border that appears on non-keratinized mucosae such as the buccal mucosa, labial mucosa, tongue, soft palate, and pharynx [4, 5].

The exact cause of RAS remains unclear, but it is believed to involve multiple factors. Local factors such as minor trauma [6], altered microbiota, parafunctional habits, and systemic factors such as genetic susceptibility [7], trace element deficiencies (e.g., ferritin, zinc, and selenium) [8], chronic inflammatory gastrointestinal diseases, food allergy, hematologic conditions, and systemic medications (e.g., captopril, phenobarbital, diclofenac, and piroxicam) have been implicated in previous studies [9]. No specific curative medicine is available due to the unknown etiology of RAS. Therefore, various topical and systemic palliative drugs and pain relief methods have been used, including antiseptics, analgesics, steroids, antibiotics, and immunosuppressive drugs [10–14]. Topical coating agents are generally preferred for their effectiveness and safety, especially for mild to moderate cases [15]. Applying topical nano-formulations as efficient drug delivery to the lesions has also captured much attention [16–18].

In recent years, there has been a shift toward minimally invasive protocols for maintaining a balanced oral microbiota. These protocols include the use of probiotics and Para probiotics to rebalance oral flora, glycine and erythritol-based powders to target specific bacteria, and ozone therapy for its antibacterial and healing properties [19].

Probiotics are highly reproducible living organisms that can protect the mucosa from harmful microorganisms and have shown effectiveness in improving oral health in basic and clinical studies [20–22]. *Lactobacillus reuteri*, a probiotic, is naturally found in the gastrointestinal tract and has been studied for its potential benefits in oral health, including preventing halitosis, candidiasis, periodontitis, and caries. *Lactobacillus reuteri* can enhance oral immunity through two mechanisms: the production of reutroin, an antimicrobial compound that inhibits various opportunistic microorganisms, and the inhibition of TNF alpha and proinflammatory cytokines production [23, 24].

Although numerous studies have been conducted on the systemic effects of probiotics in treating oral lesions

[25–28], there needs to be more research evaluating the therapeutic efficacy of probiotic nano-formulations on oral recurrent aphthous stomatitis. This study aimed to assess the therapeutic effects of a mixture of chitosan nanogel with a probiotic drug derived from *Lactobacillus reuteri* on recurrent aphthous lesions.

Method and materials

Materials

Chitosan Nanogel was purchased from Katokichi Co., Japan. *Lactobacillus reuteri* was generously donated by the Pasteur drug bank of Tehran.

Participants

This randomized controlled trial was conducted at the School of Dentistry of Shiraz University of Medical Sciences, involving 60 adult patients (above 18 years old) diagnosed with minor aphthous lesions by two board-certified oral medicine specialists using the WHO index. This study complied with the ethical principles outlined by the Ethical Committee of Shiraz University of Medical Sciences, with approval granted under the ethical code IR.SUMS.DENTAL.REC.1398.103 and clinical trial code IRCT20110428006322N2 (date of registration: 17/10/2019). Written informed consent was obtained from all participants before their inclusion in the study.

Inclusion criteria included patients between 18 and 50 years old, without any systemic medication use in the past six months, presence of 1 to 3 simultaneous lesions, no more than one day since the appearance of the lesion, and no history of rheumatologic, gastrointestinal, renal diseases, or iron-deficiency anemia. Exclusion criteria were pregnancy and lack of cooperation. Participants were asked to abstain from consuming sources of probiotics, including yogurt and dietary supplements, throughout the duration of the study.

Using stratified block randomization, participants were randomly divided into two groups, the control group, and the probiotic group. The study was double-blinded, with neither the examiner nor the patients aware of the medication. The medication was distributed to each patient by an assistant according to the randomization blocks, and the assistant also provided instructions on how to use the assigned medication.

Sample size and randomization

Based on previous studies, a sample size of 60 participants was determined to be sufficient to meet the power of 80% with a significance level of 0.05.

The patients were randomly allocated to two groups using a stratified block randomization method: the control and probiotic groups. The randomization was conducted with a block size of 4; each block contained four patients. The randomization process was carried out by a

dental assistant who was not involved in the assessments or treatments to maintain blinding of the participants and examiners throughout the study.

Drug preparation

The control group took routine palliative care. They were given an oral rinse, a mixture of 60 ml diphenhydramine, and 60 ml aluminum mg (ADIGEL-S). The control group was told to gargle the oral rinse told three times a day, for 3–4 min every time.

The probiotic group administered a topical Chitosan Nanogel/Probiotic mixture (CNP) thrice daily. They were told not to eat for at least 30 min after applying the oral, topical mixture. The *Lactobacillus reuteri* probiotic suspension was prepared by cultivating *Lactobacillus reuteri* in MRS (DeMan, Rogosa, Sharpe) broth at 37 °C for 48 h. The optical density of the culture was measured at a wavelength of 600 nm using a bio-photometer. When the density reached 0.8 (equivalent to the presence of 1×10^9 bacteria), the culture was centrifuged at 4000 rpm for 20 min. The pellet containing the bacteria was resuspended in an appropriate volume of phosphate-buffered saline (PBS), and the supernatant was discarded. All processes were conducted under aseptic conditions in a hood.

A 10 ml aliquot of the bacterial suspension was removed and added to the MRS agar culture medium, which was then incubated at 37 °C for 24 h to confirm the viability of the *Lactobacillus reuteri*.

Lactobacillus reuteri suspension was concentrated using an ultra-filtration kit with a 30,000 molecular weight cut-off membrane to retain the probiotic bacteria while removing excess buffer. The concentrated probiotic suspension was mixed with the chitosan nanogel at a 1% (v/v) concentration. The mixture was stirred thoroughly using a magnetic stirrer for 24 h to ensure homogeneous distribution of the probiotic within the nanogel. The prepared Chitosan Nanogel/Probiotic mixture (CNP) was stored in containers at four °C for patient use.

Evaluation and indices

The data for this study was obtained through clinical examination and observation. Patients were assessed for pain intensity and lesion size in four consecutive sessions, namely before, on day 3, day 5, and day seven after the treatment. Lesion size was measured using a calibrated caliper and reported in millimeters. Pain severity was assessed using the Visual Analog Scale (VAS).

Statistical analysis

The data were analyzed using SPSS software version 26. Mean \pm standard deviation (SD) was used to report the descriptive statistics of the data. Wilcoxon test was utilized to compare the Visual Analog Scale (VAS) scores

Table 1 Mean lesion size over sessions in control and probiotic group

	Before intervention	Day 3	Day 5	Day 7
Control group	7.14 \pm 2.12	5.26 \pm 1.69	3.26 \pm 1.55	1.20 \pm 0.81
Probiotic group	7.88 \pm 1.44	4.44 \pm 1.47	1.32 \pm 1.18	0.47 \pm 0.60
P-value	0.24	0.00	0.00	0.01

Table 2 Mean VAS over sessions in control and probiotic group

	Before intervention	Day 3	Day 5	Day 7
Control group	6.64 \pm 1.41	5.00 \pm 1.06	2.47 \pm 0.80	0.82 \pm 0.80
Probiotic group	7.17 \pm 1.42	3.70 \pm 1.57	1.55 \pm 1.17	0.5 \pm 0.21
P-value	0.28	0.00	0.00	0.04

at different time points (0, day 3, day 5, and day 7). In contrast, a repeated measurement test was employed to compare the lesion size across the same time points. Additionally, the Mann-Whitney test was used to compare each session's intervention scores. A significance level of $P < 0.05$ was considered statistically significant.

Results

The study comprised two groups, each consisting of 30 participants. Throughout the course of this trial, no patients withdrew or dropped out from the study. None of the participants reported any adverse effects or disturbances during the interventions. There were 18 females in the control group and 16 in the probiotic group. The distribution of males and females did not show a statistically significant difference according to the chi-square test ($P = 0.30$). The age of the patients ranged from 20 to 48 years, with a mean age of 27.82 ± 6.79 in the probiotic group and 28.62 ± 6.35 in the control group. There was no statistically significant difference in the mean age between the two groups according to the independence test ($P = 0.71$). The VAS and lesion size results in the two groups did not show any statistically significant difference, as shown in Tables 1 and 2.

The assessment conducted over four consecutive sessions revealed favorable healing of lesions in both the control and probiotic groups. Table 1 presents the mean size of the lesions, and Table 2 displays the pain intensity assessed using the VAS in both groups. Analysis using the general linear model and repeated measures test, considering the measurement of quantitative variables across multiple sessions, demonstrated that both groups exhibited statistically significant changes in lesion size and VAS over time. Additionally, a significant difference was observed in mean lesion size and VAS between each session and its adjacent sessions.

The study's findings revealed that the probiotic group exhibited a mean reduction of 7.41 ± 1.79 millimeters in lesion size and a mean reduction of 6.54 ± 1.56 scores in

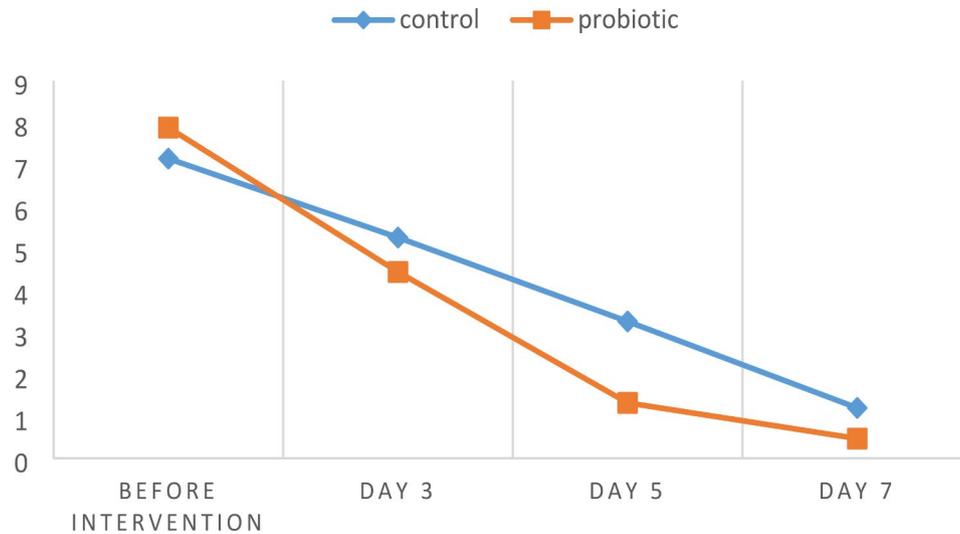


Fig. 1 Mean lesion size over sessions in control and probiotic group

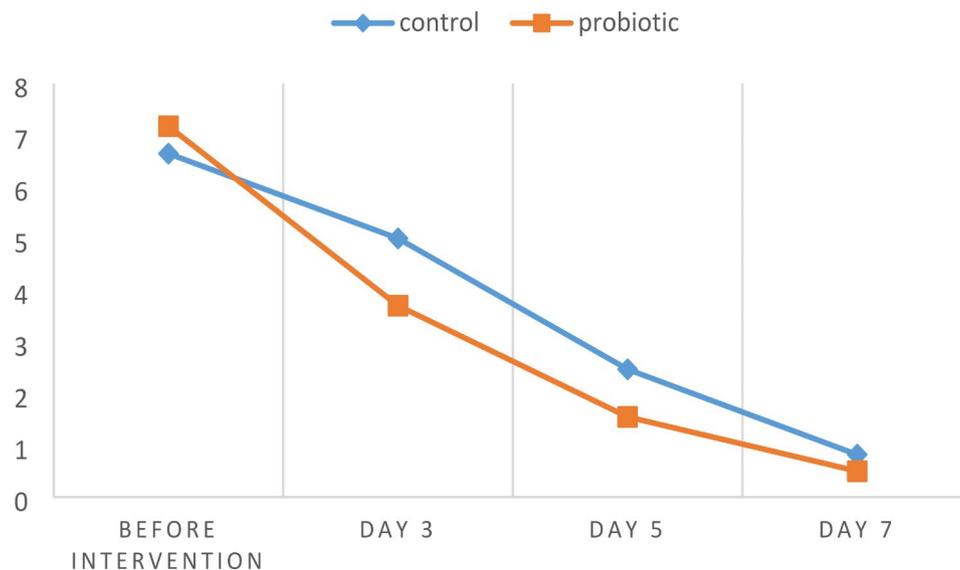


Fig. 2 Mean VAS over sessions in control and probiotic group

pain intensity at the end of the study. The control group exhibited a reduction of 5.94 ± 1.59 millimeters in lesion size and a reduction of 5.82 ± 1.66 scores in pain intensity. (Figs. 1 and 2) In terms of Mauchly's test of sphericity, and the independent t-test analysis, the reductions in lesion size and pain severity were statistically significant in the probiotic group compared to the control group.

Discussion

Given the prevalence of recurrent aphthous stomatitis (RAS), we aimed to investigate the efficacy of a topical *Lactobacillus reuteri*-derived probiotic nano-formulation in reducing lesion size and pain intensity in patients with RAS.

The study results showed that the probiotic drug derived from *Lactobacillus reuteri* exhibited a statistically significant reduction in lesion size and pain intensity compared to the control group. The probiotic group demonstrated faster and more substantial healing than the control group, indicating that the probiotic nano-formulation was more effective than common analgesic care. Notably, there was no significant difference in the effect of the probiotic nano-formulation between genders.

A recent hypothesis suggests it may be due to a local immune system defect triggered by oral bacteria. Studies show differences in oral microbiota between RAS patients and healthy individuals, with imbalances in specific bacteria potentially contributing to RAS symptoms

such as ulcers, delayed healing, and severe pain [29–31]. The findings of this study could be attributed to the potential immunomodulatory and anti-inflammatory properties of *Lactobacillus reuteri* [32]. Probiotics are believed to modulate the immune system, correct dysbiosis, strengthen mucosal barriers and regulate inflammatory responses [33–36]. They also improve the integrity of the mucosal epithelium by inhibiting the production of pro-inflammatory cytokines, such as TNF-, and subsequently, decrease epithelial permeability [33]. The modulation of the oral microbiota by the probiotic formulation may have reduced pain and lesion size in patients with RAS. Using probiotics as a nano-formulation may enhance their stability, bioavailability, and effectiveness in delivering beneficial bacteria to the oral cavity, thereby exerting their therapeutic effects on RAS [37].

This study's findings agree with previous studies that reported probiotics' beneficial effects in managing RAS. Pedersen et al. showed that a probiotic lozenge containing *Lactobacillus reuteri* improved the Ulcer Severity Score and reduced oral pain related to RAS over a 3-month [38]. However, the improvement was not statistically significant compared to the placebo group [38]. Nirmala et al. showed considerable improvement in erythema, pain reduction, decreased oral thrush, and burning sensation in the mouth following topical application of *Bacillus Clausii* probiotics can be used as an adjuvant in treating recurrent aphthous ulcer and oral candidiasis [27]. These studies and the present study suggest that probiotics may be a promising therapeutic option for managing RAS.

However, it is worth noting that some previous studies have reported conflicting results. For example, a systematic review and meta-analysis conducted by Cheng et al. found probiotics effective in relieving oral pain but ineffective in reducing ulcer severity [39]. Aggour et al. reported significant differences in pain reduction and insignificant ulcer size reduction following applying *Lactobacillus acidophilus* probiotic lozenges compared to the control group [40]. These discrepancies in findings could be attributed to differences in study design, probiotic strains, formulations used, dosages, and study populations.

Nanofibers, nanoparticles, and nanostructured materials have shown promise for probiotic delivery due to their efficient encapsulation, site-specific release, stability during manufacturing and storage, biocompatibility and controlled drug release, and improved viability [37]. Overall, nanomaterials-based formulations can enhance the therapeutic effects of probiotic products [37].

Despite the promising results of this study, some limitations need to be considered in future studies. The sample was small, and the follow-up period was limited to four sessions. The study did not investigate the mechanisms

underlying the beneficial effects of the probiotic nano-formulation via in-vitro tests. Our analysis also lacked a separate chitosan nanogel group so that we could investigate the possible positive effect of chitosan as an antibacterial and anti-inflammatory polymer on the study result.

In light of the encouraging results from this study, it is worth considering the potential implications for future research and clinical applications. The concept of para probiotics, probiotics, and postbiotics presents exciting opportunities in the field of oral health. Probiotics are effective in promoting various health benefits, but concerns about their safety have emerged, especially when administered to vulnerable individuals like the elderly and those with weakened immune systems. In response to these concerns, non-viable probiotic products known as para probiotics and postbiotics have been introduced. Para probiotics are inactivated microbial cells that offer health benefits without posing health risks. They can regulate the immune system, combat pathogens, and exhibit anti-inflammatory, antiproliferative, and antioxidant properties [19, 41]. Furthermore, future studies could delve deeper into the specific mechanisms underlying the beneficial effects of probiotics and para probiotics. In-vitro tests and mechanistic research are warranted to elucidate the precise interactions between these microbial formulations and the oral environment, shedding light on the pathways through which they exert their therapeutic effects.

Conclusions

Considering the limitations, the findings of this study contribute to the growing body of evidence on the potential benefits of probiotics in managing RAS and highlight the importance of further research in this field. *Lactobacillus reuteri*-derived probiotic nano-formulation might be a promising treatment option for RAS. Moreover, the study emphasizes the need for further research to explore proactive actions with para probiotics, probiotics, and postbiotics which present exciting opportunities to enhance oral health care.

Acknowledgements

Hereby, we gratefully acknowledge the participants in this study, the vice chancellor for research of Shiraz University of medical sciences and Dr. Vosoughi, who helped us in the statistical analysis of this project.

Author contributions

A. G. and M. R. contributed to the conception, design and critically revised the manuscript at the end. H. K. contributed to the study design, data acquisition, and data interpretation. N.S. wrote the main manuscript and contributed to data interpretation. All authors were aware of all parts of the study, gave their final approval, and agreed to be accountable for all aspects of the work.

Funding

Vice Chancellor of Shiraz University of Medical Sciences funded this study.

Data availability

The datasets generated during and analyzed during the current study are available from the corresponding author upon reasonable request.

Declarations**Ethics approval and consent to participate**

The study protocol conforms to the ethical guidelines of the 1975 Declaration of Helsinki and the Ethical Committees of Shiraz University of Medical Sciences approved the study by ethical code IR.SUMS.DENTAL.REC.1398.103 and clinical trial code IRCT20110428006322N2 (date of registration: 17/10/2019). All methods were carried out in accordance with relevant guidelines and regulations. Besides, Written informed consent was taken from all participants.

Consent for publication

Not applicable.

Competing interests

All authors declare that they have no competing interests, personal or financial relationships that could have affected the results of this study.

Received: 8 June 2023 / Accepted: 11 December 2023

Published online: 19 December 2023

References

- Edgar NR, Saleh D, Miller RA. Recurrent aphthous stomatitis: a review. *J Clin Aesthetic Dermatol*. 2017;10(3):26.
- Raj K, Vadivel JK, Sivaswamy V. Prevalence and age related risk of three clinical variants of Aphthous Stomatitis: a retrospective study. *Indian J Forensic Med Toxicol*. 2020;14(4):5643–9.
- Jahromi NZ, Ghanpanchi J, Pourshahidi S, Zahed M, Ebrahimi H. Clinical evaluation of high and low-level laser treatment (CO₂vslnGaAlp diode laser) for recurrent aphthous stomatitis. *J Dent*. 2017;18(1):17.
- Banu S, Ramakrishnan M. A comprehensive review on oral aphthous Ulcer. *Indian J Public Health Res Dev*, vol. 10, no. 11, 2019.
- Cui RZ, Bruce AJ, Rogers RS. Recurrent aphthous stomatitis. *Clin Dermatol*. 2016;34(4):475–81.
- Gallo Cde B, Mimura MA, Sugaya NN. Psychological stress and recurrent aphthous stomatitis, (in eng), *Clinics (Sao Paulo, Brazil)*, vol. 64, no. 7, pp. 645-8, 2009, <https://doi.org/10.1590/s1807-59322009000700007>.
- Eversole LR. Immunopathology of oral mucosal ulcerative, desquamative, and bullous diseases. Selective review of the literature, (in eng), *Oral surgery, oral medicine, and oral pathology*, vol. 77, no. 6, pp. 555–71, Jun 1994, [https://doi.org/10.1016/0030-4220\(94\)90312-3](https://doi.org/10.1016/0030-4220(94)90312-3).
- Hunter IP, Ferguson MM, Scully C, Galloway AR, Main AN, Russell RI. Effects of dietary gluten elimination in patients with recurrent minor aphthous stomatitis and no detectable gluten enteropathy, (in eng), *Oral surgery, oral medicine, and oral pathology*, vol. 75, no. 5, pp. 595-8, May 1993, [https://doi.org/10.1016/0030-4220\(93\)90232-s](https://doi.org/10.1016/0030-4220(93)90232-s).
- Natah SS, Konttinen YT, Enattah NS, Ashammakhi N, Sharkey KA, Häyriinen-Immonen R. Recurrent aphthous ulcers today: a review of the growing knowledge, (in eng), *International journal of oral and maxillofacial surgery*, vol. 33, no. 3, pp. 221–34, Apr 2004, <https://doi.org/10.1006/ijom.2002.0446>.
- McBride DR. Management of aphthous ulcers, (in eng), *American family physician*, vol. 62, no. 1, pp. 149–54, 160, Jul 1 2000.
- Altenburg A, Zouboulis CC. Current concepts in the treatment of recurrent aphthous stomatitis, (in eng), *Skin therapy letter*, vol. 13, no. 7, pp. 1–4, Sep 2008.
- Scully C, Gorsky M, Lozada-Nur F. The diagnosis and management of recurrent aphthous stomatitis: a consensus approach, (in eng), *Journal of the American Dental Association* (1939), vol. 134, no. 2, pp. 200-7, Feb 2003, <https://doi.org/10.14219/jada.archive.2003.0134>.
- Belenguer-Guallar I, Jiménez-Soriano Y, Claramunt-Lozano A. Treatment of recurrent aphthous stomatitis. A literature review. (in eng) *Journal of Clinical and Experimental Dentistry*. Apr 2014;6(2):e168–74. <https://doi.org/10.4317/jced.51401>.
- Eisen D, Lynch DP. Selecting topical and systemic agents for recurrent aphthous stomatitis, (in eng), *Cutis*, vol. 68, no. 3, pp. 201-6, Sep 2001.
- Choi E, Nahm FS, Han WK, Lee PB, Jo J. Topical agents: a thoughtful choice for multimodal analgesia, (in eng). *Korean J Anesthesiol*. Oct 2020;73(5):384–93. <https://doi.org/10.4097/kja.20357>.
- Karavana SY et al. A new approach to the treatment of recurrent aphthous stomatitis with bioadhesive gels containing cyclosporine A solid lipid nanoparticles: in vivo/in vitro examinations, (in eng), *Int J Nanomedicine*, vol. 7, pp. 5693–704, 2012, <https://doi.org/10.2147/ijn.S36883>.
- Bakhshi M, Mahboubi A, Jaafari MR, Ebrahimi F, Tofangchiha M, Alizadeh A, COMPARATIVE EFFICACY OF 1% CURCUMIN NANOMICELLE GEL AND 2% CURCUMIN GEL FOR TREATMENT OF RECURRENT APHTHOUS STOMATITIS. A DOUBLE-BLIND RANDOMIZED CLINICAL TRIAL, (in eng). *J Evid Based Dent Pract*. Jun 2022;22(2):101708. <https://doi.org/10.1016/j.jebdp.2022.101708>.
- Sadeghian R, Rohani B, Golestannejad Z, Sadeghian S, Mirzaee S. Comparison of therapeutic effect of mucoadhesive nano-triamcinolone gel and conventional triamcinolone gel on oral lichen planus. *Dent Res J*. 2019;16(5):277.
- Scribante A, Butera A, Alovisi M. Customized minimally invasive protocols for the clinical and microbiological management of the oral microbiota. Volume 10. ed: MDPI; 2022. p. 675.
- Caufield PW, Li Y, Dasanayake A, Saxena D. Diversity of lactobacilli in the oral cavities of young women with dental caries, (in eng), *Caries research*, vol. 41, no. 1, pp. 2–8, 2007, <https://doi.org/10.1159/000096099>.
- Busscher HJ, Mulder AF, van der Mei HC. In vitro adhesion to enamel and in vivo colonization of tooth surfaces by Lactobacilli from a bio-yoghurt, (in eng), *Caries research*, vol. 33, no. 5, pp. 403-4, Sep-Oct 1999, <https://doi.org/10.1159/000016541>.
- Lima LM, Motisuki C, Spolidorio DM, Santos-Pinto L. In vitro evaluation of probiotics microorganisms adhesion to an artificial caries model, (in eng), *European journal of clinical nutrition*, vol. 59, no. 7, pp. 884-6, Jul 2005, <https://doi.org/10.1038/sj.ejcn.1602158>.
- Twetman S, Derawi B, Keller M, Ekstrand K, Yucel-Lindberg T, Stecksén-Blicks C. Short-term effect of chewing gums containing probiotic Lactobacillus reuteri on the levels of inflammatory mediators in gingival crevicular fluid, (in eng). *Acta Odontol Scand*. 2009;67(1):19–24. <https://doi.org/10.1080/00016350802516170>.
- Harini PM, Anegundi RT. Efficacy of a probiotic and chlorhexidine mouth rinses: a short-term clinical study, (in eng), *Journal of the Indian Society of Pedodontics and Preventive Dentistry*, vol. 28, no. 3, pp. 179–82, Jul-Sep 2010, <https://doi.org/10.4103/0970-4388.73799>.
- Hasslöf P, Hedberg M, Twetman S, Stecksén-Blicks C. Growth inhibition of oral mutans streptococci and candida by commercial probiotic lactobacilli—an in vitro study, (in eng), *BMC oral health*, vol. 10, p. 18, Jul 2 2010, <https://doi.org/10.1186/1472-6831-10-18>.
- Michalek SM, Hirasawa M, Kiyono H, Ochiai K, McGhee JR. Oral ecology and virulence of Lactobacillus casei and Streptococcus mutans in gnotobiotic rats, (in eng), *Infection and immunity*, vol. 33, no. 3, pp. 690-6, Sep 1981, <https://doi.org/10.1128/iai.33.3.690-696.1981>.
- Nirmala M, Smitha SG, Kamath GJ. A study to assess the efficacy of local application of oral probiotic in treating recurrent aphthous Ulcer and oral candidiasis, (in eng). *Indian J Otolaryngol head neck Surgery: Official Publication Association Otolaryngologists India*. Oct 2019;71:113–7. <https://doi.org/10.1007/s12070-017-1139-9>.
- Pedersen AML, Bukkehave KH, Bennett EP, Twetman S. Effect of Lozenges Containing Lactobacillus reuteri on the Severity of Recurrent Aphthous Ulcers: a Pilot Study, (in eng), *Probiotics and antimicrobial proteins*, vol. 12, no. 3, pp. 819–823, Sep 2020, <https://doi.org/10.1007/s12602-019-09586-x>.
- Ślebioda Z, Szponar E, Kowalska A. Etiopathogenesis of recurrent aphthous stomatitis and the role of immunologic aspects: literature review, *Archivum immunologiae et therapeuticae experimentalis*, vol. 62, no. 3, pp. 205–215, 2014.
- Kim Y-j, Choi YS, Baek KJ, Yoon S-H, Park HK, Choi Y. Mucosal and salivary microbiota associated with recurrent aphthous stomatitis. *BMC Microbiol*. 2016;16(1):1–10.
- Kim YJ, Choi YS, Baek KJ, Yoon SH, Park HK, Choi Y. Mucosal and salivary microbiota associated with recurrent aphthous stomatitis, (in eng), *BMC microbiology*, vol. 16 Suppl 1, p. 57, Apr 1 2016, <https://doi.org/10.1186/s12866-016-0673-z>.
- Cristofori F, Dargenio VN, Dargenio C, Miniello VL, Barone M, Francavilla R. Anti-inflammatory and immunomodulatory effects of probiotics in gut inflammation: a door to the body. *Front Immunol*. 2021;12:578386.
- Hardy H, Harris J, Lyon E, Beal J, Foey AD. Probiotics, prebiotics and immunomodulation of gut mucosal defences: homeostasis and immunopathology, (in eng), *Nutrients*, vol. 5, no. 6, pp. 1869–912, May 29 2013, <https://doi.org/10.3390/nu5061869>.

34. Javanshir N et al. Evaluation of the Function of Probiotics, Emphasizing the Role of their Binding to the Intestinal Epithelium in the Stability and their Effects on the Immune System, (in eng), *Biol Proced Online*, vol. 23, no. 1, p. 23, Dec 1 2021, <https://doi.org/10.1186/s12575-021-00160-w>.
35. Rao RK, Samak G. Protection and Restitution of Gut Barrier by Probiotics: Nutritional and Clinical Implications, (in eng), *Curr Nutr Food Sci*, vol. 9, no. 2, pp. 99–107, May 1 2013, <https://doi.org/10.2174/1573401311309020004>.
36. Lazar V et al. Aspects of Gut Microbiota and Immune System Interactions in Infectious Diseases, Immunopathology, and Cancer, (in eng), *Front Immunol*, vol. 9, p. 1830, 2018, <https://doi.org/10.3389/fimmu.2018.01830>.
37. Baral KC, Bajracharya R, Lee SH, Han HK. Advancements in the Pharmaceutical Applications of Probiotics: dosage forms and Formulation Technology, (in eng). *Int J Nanomedicine*. 2021;16:7535–56. <https://doi.org/10.2147/ijn.S337427>.
38. Pedersen AML, Bukkehave KH, Bennett EP, Twetman S. Effect of lozenges containing *Lactobacillus reuteri* on the severity of recurrent aphthous ulcers: a pilot study. *Probiotics and Antimicrobial Proteins*, vol. 12, no. 3, pp. 819–23, 2020/09/01 2020, <https://doi.org/10.1007/s12602-019-09586-x>.
39. Cheng B, Zeng X, Liu S, Zou J, Wang Y. The efficacy of probiotics in management of recurrent aphthous stomatitis: a systematic review and meta-analysis, *Scientific Reports*, vol. 10, no. 1, p. 21181, 2020/12/03 2020, <https://doi.org/10.1038/s41598-020-78281-7>.
40. Aggour RL, Mahmoud SH, Abdelwhab A. Evaluation of the effect of probiotic lozenges in the treatment of recurrent aphthous stomatitis: a randomized, controlled clinical trial. *Clin Oral Invest*, vol. 25, no. 4, pp. 2151–8, 2021/04/01 2021, <https://doi.org/10.1007/s00784-020-03527-7>.
41. Butera A et al. Paraprobiotics in Non-Surgical Periodontal Therapy: Clinical and Microbiological Aspects in a 6-Month Follow-Up Domiciliary Protocol for Oral Hygiene, (in eng), *Microorganisms*, vol. 10, no. 2, Feb 1 2022, <https://doi.org/10.3390/microorganisms10020337>.

Publisher's Note

Springer Nature remains neutral with regard to jurisdictional claims in published maps and institutional affiliations.