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Non-additive effects of ACVR2A in preeclampsia in a Philippine population

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Abstract

Background: Multiple interrelated pathways contribute to the pathogenesis of preeclampsia, and variants in susceptibility genes may play a role among Filipinos, an ethnically distinct group with high prevalence of the disease. The objective of this study was to examine the association between variants in maternal candidate genes and the development of preeclampsia in a Philippine population.

Methods: A case-control study involving 29 single nucleotide polymorphisms (SNPs) in 21 candidate genes was conducted in 150 patients with preeclampsia (cases) and 175 women with uncomplicated normal pregnancies (controls). Genotyping for the *GRK4* and *DRD1* gene variants was carried out using the TaqMan Assay, and all other variants were assayed using the Sequenom MassARRAY Iplex Platform. PLINK was used for SNP association testing. Multilocus association analysis was performed using multifactor dimensionality reduction (MDR) analysis.

Results: Among the clinical factors, older age ($P < 1 \times 10-4$), higher BMI ($P < 1 \times 10-4$), having a new partner (P = 0.006), and increased time interval from previous pregnancy (P = 0.018) associated with preeclampsia. The MDR algorithm identified the genetic variant *ACVR2A* rs1014064 as interacting with age and BMI in association with preeclampsia among Filipino women.

Conclusions: The MDR algorithm identified an interaction between age, BMI and *ACVR2A* rs1014064, indicating that context among genetic variants and demographic/clinical factors may be crucial to understanding the pathogenesis of preeclampsia among Filipino women.

Keywords: Association study, Multifactor dimensionality reduction, Philippines, Preeclampsia, Single nucleotide polymorphism

Background

Hypertensive disorders of pregnancy account for 36.7% of all maternal deaths in the Philippines [1], which is much higher than the worldwide rate of 18% [2]. Included among these hypertensive diseases affecting pregnant women is preeclampsia, a severe and diverse disorder that is associated with life-threatening multiorgan maternal complications and which causes serious feto-placental problems. It accounted for 22.5% of hypertensive patient admissions at the hospital where this study was conducted [3].

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Preeclampsia is a multifactorial disease, with both genetic and environmental factors contributing to its development. Multiple interrelated pathways have been suggested to contribute to its pathogenesis. Previous studies have tested genes with potential biological relevance in specific pathways to ascertain whether certain variants influence the disease process. The biological pathways impacted by preeclampsia include but are not limited to aberrant placental development and dysfunctional hemodynamic and renal functions, impaired immune function, free radical dysregulation and lipid peroxidation, and defects in coagulation and fibrinolysis. We have previously found variants of the VEGF-A and VEGFR1 genes to associate with preeclampsia among Filipinos, an ethnically distinct group with high prevalence [4]. These genes are important in angiogenesis, a critical



© The Author(s). 2019 **Open Access** This article is distributed under the terms of the Creative Commons Attribution 4.0 International License (http://creativecommons.org/licenses/by/4.0/), which permits unrestricted use, distribution, and reproduction in any medium, provided you give appropriate credit to the original author(s) and the source, provide a link to the Creative Commons license, and indicate if changes were made. The Creative Commons Public Domain Dedication waiver (http://creativecommons.org/publicdomain/zero/1.0/) applies to the data made available in this article, unless otherwise stated. process in the establishment of normal pregnancy and in preeclampsia.

The effect of single gene variation will likely be contingent on other genetic variations (gene-gene interaction, or epistasis) and environmental factors (gene-environment interaction). Since many genes and environmental factors interact to cause multifactor and polygenic diseases, including preeclampsia, the effect of any single gene may be too small to be detected using traditional statistical methods, which do not take these interactions into account. The multifactor dimensionality reduction (MDR) algorithm has been designed as an alternative to traditional statistical methods to deal with high-order gene/ factor interactions [5, 6]. MDR has many other advantages over traditional methods. It is model-free, i.e., it does not assume any particular genetic model, and requires only a small sample size that can be used for case-control studies [5–7]. MDR has been successfully applied in detecting gene-gene interactions for a number of clinical phenotypes, which include bronchial asthma [8], autism [9], essential hypertension [10, 11], and type II diabetes [12].

In this study, we evaluated the role of contextual effects in the development of preeclampsia among Filipino women by analyzing 29 previously reported susceptibility SNPs found in 21 genes. The 21 genes are involved in various pathways that regulate the processes implicated in the development of preeclampsia and are as follows: endothelial/angiogenesis (*VEGFA, VEGFC, VEGFR1, VEGFR3*); dopaminergic system (*GRK4, DRD1*); reninangiotensin system (*AGT*); immunity and inflammation (*ERAP2, CTLA4, IL1A, TNSF13B*); lipid metabolism (*LPL*); oxidative stress and detoxification (*eNOS, CYP1A2, PON1, EPHX1, GSTP1*); key signaling proteins (*ACVR2A*), hormone and neurotransmitter regulation (*COMT*) and protein biosynthesis (*MTR* and *MTRR*).

Methods

This is a case-control study that included 381 individuals. Of these, 56 were removed in the final analysis because they had more than 2 genotypes missing. Of the 325 that remained, 150 were patients with preeclampsia (cases) and 175 were women with uncomplicated normal pregnancies (controls). Subjects were recruited upon admission to the hospital and were followed up until 6 weeks after delivery. Subjects included in the normal pregnancy control group had blood pressures $\leq 120/80$ mmHg, consistent with the latest guidelines on hypertension [13]. Blood pressure was measured according to the Seventh Report of the Joint National Committee on Prevention, Detection, Evaluation, and Treatment of High Blood Pressure [14] and verified twice with at least a 4-h interval. Exclusion criteria for the control group were a history of hypertension and pregnancy-induced hypertension, multiple pregnancy, molar pregnancy, personal and family history of diabetes mellitus, ischemic heart disease, cerebrovascular accident, and renal disease.

Included in the preeclampsia group were patients who had a resting systolic blood pressure \geq 140 mmHg and/or a diastolic blood pressure \geq 90 mmHg and had proteinuria after 20 weeks of gestation. Proteinuria was defined as \geq 300 mg protein in a 24-h urine collection or a urine protein dipstick of \geq 2+. Exclusion criteria for the preeclampsia group were a history of hypertension, renal disease, proteinuria before the 20th week of pregnancy, multiple pregnancy, diabetes mellitus, ischemic heart disease, cerebrovascular accident, and renal disease.

This study was undertaken in accordance with the Declaration of Helsinki at the Department of Obstetrics and Gynecology of the University of the Philippines, Philippine General Hospital (UP-PGH) in Manila. All subjects provided informed written consent. The study was approved by the UP-PGH Ethics Review Board.

SNP selection, blood sampling, DNA extraction, and genotyping

The candidate genes and respective variants were selected from different pathophysiological pathways involved in the development of preeclampsia and based on previous association studies. The ACVR2A SNPs in particular have been shown to be associated with preeclampsia in a study involving Brazilian [15] and Norwegian women [16]. Criteria for SNP selection included SNPs for which an association with preeclampsia has been reported in other ethnic populations and those that are also supported by the platform used for genotyping. The gene variants in relevant pathways include: endothelial/angiogenesis (VEGFA [17-19], VEGFC [20], VEGFR1 [20-22]; renin-angiotensin system (AGT [23-25]); immunity and inflammation (ERAP2 [26], CTLA4 [27, 28], IL1A [29, 30], TNSF13B [31]); lipid metabolism (LPL [27, 32]); oxidative stress and detoxification (eNOS [33, 34], CYP1A2 [35], PON1 [36–39], EPHX1 [40, 41], GSTP1 [42]); key signaling proteins (ACVR2A [15, 16]), hormone and neurotransmitter regulation (COMT [43, 44]) and protein biosynthesis (MTR and MTRR) [45-48]. Also included are SNPs in the dopaminergic system (GRK4 [49-52] and DRD1 [52]), which have been established to be associated strongly with hypertension.

Venous blood samples for DNA extraction and genotyping were collected after a definitive diagnosis of preeclampsia or normal pregnancy. Three milliliters of venous blood were extracted at the time of hospital admission by venipuncture at the antecubital fossa, collected in a vacutainer with EDTA, and stored at 4 °C until DNA extraction. DNA was extracted from the peripheral blood mononuclear cells using the QIAamp° DNA Mini Kit. DNA purity and quantity were determined using a Nanodrop 2000 spectrometer (Thermo Scientific, Waltham, MA, USA). The DNA was stored at 4 °C until genetic profiling. Genotyping of the *GRK4* and *DRD1* variants was carried out using the TaqMan Assay at the University of Maryland Biopolymer-Genomics Core Facility. Genotyping of the other genes was carried out using the Sequenom MassARRAY Iplex Platform at the Center for Genomic Sciences of the University of Hong Kong. Repeat genotyping for 16 samples was performed for quality control.

Statistical analysis

Statistical analysis for the clinical parameters was performed using STATA software, version 14.1 [53]. All values were expressed as mean/median ± standard error, while the association of known categorical risk factors was analyzed using Pearson's chi-square test. Odds ratios (OR) were calculated to determine the odds of developing preeclampsia when the individual had the clinical factor of interest. OR was used for binary logistic regression and multinomial logistic regression. Minor allele frequencies and Hardy-Weinberg Equilibrium were calculated for each SNP using PLINK 1.9. Of the 29 SNPs, 6 were removed from further analysis (AGT, LPL, FLT1, ERAP1, FLT4 and TNFSF13B) because these SNPs were monomorphic, or had a minor allele frequency below 5%. Accumulated/average Cross Validation testing, training, consistency and permutation P values were calculated using MDR [6, 54]. The MDR algorithm and ViSEN software [55] were applied to the genetic data to enable the detection and characterization of epistatic SNP-SNP interactions and SNP-clinical factor (age and BMI) interaction. To identify a correct multi-locus model, the Acc. CV testing (Accumulated/average Cross Validation testing) and CV Consistency (Cross Validation Consistency) were calculated for each model.

Results

Normal pregnancies and preeclampsia outcomes differed in several demographic and clinical characteristics. Mothers with normal pregnancies were significantly younger (25.2 vs. 30.3; $P = < 1 \times 10^{-4}$) (Table 1) and had lower BMI (21.75 vs. 23.5; $P = < 1 \times 10^{-4}$) (Table 1). The mean interval from last pregnancy was shorter in normal outcomes (2.03 years vs. 2.87 years; P =0.018). Having a new partner also associated with preeclampsia (P = 0.006). However, neither smoking (P = 0.957) nor gravidity (P = 0.435) associated with preeclampsia (Table 2).

No SNPs associated with preeclampsia in the unadjusted analyses (Table 3). Association of single SNPs was also run adjusting for age, BMI, interval between pregnancies, and new partner (Additional file 1: Table S1).

Table 1 Age, BMI, and interval year are risk factors for preeclampsia

Variable	Category	Mean/Median	P value
Age	NP	Mean: 25.2 ± 0.45 Median: 24	
	PE	Mean: 30.3 ± 0.53 Median: 31	$< 1 \times 10^{-4*}$
BMI ³	NP	Mean: 21.75 ± 0.19 Median: 21.39	
	PE	Mean: 23.5 ± 0.21 Median: 23.31	$< 1 \times 10^{-4}$ *
Interval Year	NP	Mean: 2.03 ± 0.21 Median: 1	
_	PE	Mean: 2.87 ± 0.28 Median: 1	0.018*

Data are expressed as mean $\pm\,\text{SEM}$ and median; odds ratio (OR) used for binary logistic regression

NP Normal pregnancy, PE Preeclampsia, BMI Body mass index

*P values < 0.05 are statistically significant

The table summarizes the demographic, clinical characteristics, and risk factors of preeclampsia patients and controls with normal pregnancy. The preeclampsia patients were generally older, had higher BMI, and longer interval year from previous pregnancy compared with control subjects.

After adjusting for these covariates, one SNP reached nominal statistical significance (*VEGF-A* rs3025039; P = 0.022). This was not significant after adjusting for multiple testing (Bonferroni threshold P = 0.0023).

Context-dependent effects were assessed using MDR. Only one model that included a genetic variant, as well as the significant demographic and clinical variables, was statistically significant; it included maternal age as three categories (18-25; 26-35; 36-Older); maternal BMI as three categories (13-17.97; 18-25; 25.1-Above) and rs1014064 (cross validation testing prediction = 60.95%; permutation P = 0.005 and cross validation consistency = 7/10 (Table 4). Upon examining the model, it was evident that the genetic effect was present primarily in the middle range of BMI (18–25) (Fig. 1). In low BMI ranges (13–17.97), the outcome was dominated by a protective effect and in high BMI ranges (25.1-above) by a risk-increasing effect of body mass. When re-analyzing only the middle range of BMI (18-25), genetics appeared to play a significant non-additive role (cross validation testing prediction = 64.88%; permutation $p = \langle 1 \times 10^{-4}$ and cross validation consistency = 10/10; Fig. 1). There was no statistical epistasis detected using ViSEN.

The genetic data were analyzed for epistasis using MDR and ViSEN statistical software. Table 3 summarizes the different gene variants that were included in the analysis, as well as pertinent information for each, including the gene product and the processes in which it is involved, chromosome location, minor allele frequencies (MAF), and odds ratio (OR). Monomorphic SNPs (VEGFR1 rs7335588, AGT rs41271499, and LPL rs268) and with MAF < 0.05 (VEGFR3 rs307826, ERAP2 rs17408150, and TNFSF13B rs16972194) were excluded from analyses. Genotype frequencies for all the SNPs were in Hardy-Weinberg equilibrium.

Variable	Category	No	Yes	Pearson's correlation	P value	OR
Smoking	NP	181	17			
	PE	167	16	0.003	0.957	1.020077
Alcohol	NP	184	14			
	PE	165	18	1.737	0.333	1.433766
Nulliparous	NP	109	89			
	PE	108	75	0.61	0.435	0.8504994
		Previous	New			
Partner	NP	173	25			
	PE ²	140	43	7.67	0.006*	2.125

Table 2 A new partner is a risk factor for preeclampsia

Data used odds ratio (OR) for binary logistic regression and for multinomial logistic regression

NP Normal pregnancy, PE Preeclampsia

*P values < 0.05 are statistically significant

Among the risk factors analyzed (smoking, alcoholic beverage consumption, nulliparity, and having a new partner) in preeclampsia patients and controls with normal pregnancy, only having a new partner was associated with increased risk of preeclampsia.

Table 3 Gene variants included in the stud
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Chr	SNP	Gene	Product (processes involved in)	MAF	OR	Allelic P value	Genotypic P value
1	rs1051740	EPHX1	Epoxide hydrolase (xenobiotic metabolism)	0.4547	1.039	0.8109	0.6148
1	rs699	AGT	Angiotensinogen (products elicit vasoconstriction)	0.1277	1.098	0.6899	NA
1	rs1805087	MTR	5-Methyltetrahydrofolate-Homocysteine Methyltransferase (methionine biosynthesis)	0.1357	1.013	0.9568	NA
2	rs3783550	IL1A	Interleukin 1-alpha (immunity, inflammation, hematopoiesis)	0.2031	1.003	0.988	0.7627
2	rs1014064	ACVR2A	Activin A Receptor Type 2A (growth and differentiation)	0.4414	0.8699	0.3805	0.556
2	rs2161983	ACVR2A		0.4385	0.8921	0.4717	0.5188
2	rs231775	CTLA-4	Cytotoxic T-Lymphocyte Associated Protein 4 (inhibition of immune responses)	0.4462	1.22	0.216	0.4626
4	rs2960306	GRK4	G protein-coupled receptor kinase 4 (receptor desensitization)	0.0679	1.763	0.0709	NA
4	rs1024323	GRK4		0.1354	1.26	0.3133	NA
4	rs1801058	GRK4		0.4599	1.024	0.8797	0.8192
4	rs7664413	VEGF-C	Vascular endothelial growth factor C (lymphangiogenesis)	0.2436	1.316	0.1412	0.181
5	rs1801394	MTRR	Methionine synthase reductase (methionine biosynthesis)	0.2754	0.9219	0.645	0.8572
5	rs2549782	ERAP 2	Endoplasmic reticulum aminopeptidase 2 (antigen processing)	0.4046	1.097	0.5622	0.8404
5	rs4532	DRD1	Dopamine D1 receptor (sodium transport, blood pressure regulation)	0.2105	1.01	0.9594	0.9148
6	rs2010963	VEGF-A	Vascular endothelial growth factor A (angiogenesis, vasculogenesis,	0.223	0.8585	0.4495	NA
6	rs3025039	VEGF-A	endothelial cell growth & migration)		0.6282	0.0626	NA
7	rs662	PON1	Paraoxonase 1 (inactivation of organophosphates, inhibition of atherosclerosis formation)	0.4306	0.9923	0.9612	0.9841
7	rs1799983	NOS3	Endothelial NOS (vascular relaxation, antioxidant activity)	0.1813	0.9324	0.7342	0.829
11	rs1695	GSTP1	Glutathione S-Transferase Pi 1 (xenobiotic metabolism)	0.2954	1.073	0.6809	0.8688
13	rs12584067	VEGFR-1	Vascular endothelial growth factor receptor 1 (cell proliferation and differentiation	0.0776	1.282	0.3978	NA
13	rs722503	VEGFR-1	Vascular endothelial growth factor receptor 1 (cell proliferation and differentiation	0.1660	0.8019	0.3593	NA
15	rs2470890	CYP1A2	Cytochrome P450 1A2 (xenobiotic metabolism)	0.0923	0.8182	0.4643	NA
22	rs4633	COMT	Catechol-O-methyltransferase (degrades catecholamines)	0.1692	1.056	0.7962	NA

Chr Chromosome, SNP Single nucleotide polymorphism, MAF Minor allele frequencies, OR Odds ratio, NA Not appplicable

The alleles and genotypes of the SNPs are not associated with preeclampsia when analyzed using PLINK. NA corresponds to the inability to determine a

genotypic P value, due to a small number of counts in at least 1 of the 3 genotypes (i.e., AA/AT/TT not present, only AA/AT), thus, only allelic P value is given.

 Table 4 MDR analysis of genetic variants, adjusting for age and BMI

SNP rs1014064	GENE ACVR2A		
INTERACTION	Acc. CV Testing	CV Consistency	
Age	0.6457	10/10	
Age, BMI	06729	10/10	
rs1014064, Age, BMI	0.6095	7/10	P = 0.005

There is an interaction between *ACVR2A* rs1014064, age, and BMI *MDR* Multifactor dimensionality reduction, *SNP* Single nucleotide polymorphism, *BMI* Body mass index

*P value < 0.05 is statistically significant

Discussion

Non-linear interactions among multiple genetic and environmental or clinical factors are now understood to be important components in understanding the underlying pathogenesis, especially when considering the genetic bases of complex diseases such as preeclampsia. The MDR algorithm is a well-known data mining strategy that provides an improved representation of the genotypic and phenotypic data and enables better detection of higher-order interactions, such as epistatic interactions [6]. In the current study, MDR identified a four-locus model that underscores a possible interaction among rs1014064 (ACVR2A), rs7664413 (VEGF-C), rs2549782 (ERAP2), and rs662 (PON1) variants when un-adjusted for age or BMI (Additional file 2: Table S2). However, when age and BMI were adjusted for, these effects disappeared. A significant interaction was found in a model involving the genetic variant rs1014064 (ACVR2A) and the demographic and clinical variables, age and BMI.

The significant gene identified, *ACVR2A*, encodes the receptor for Activin A. ACVR2A expression in the placenta throughout pregnancy indicates its possible role in the regulation of placental development and function [56]. Initial studies have shown a linkage between preeclampsia and various parts of chromosome 2, where the ACVR2A gene is localized. The first reported locus for preeclampsia that met the criteria for genome-wide association significance was seen in chromosome 2p13 and 2q23 in a study involving Icelandic families, representing 343 affected women [57]. Two other genome-wide association studies identified other loci in chromosome 2 distinct from those seen in the initial study, i.e., 2p25 in 15 families with 49 affected women from Finland [58] and 2p11-12 and 2q22 involving 34 families, representing 121 affected women from Australia and New Zealand [59]. With the reported significant linkage to chromosome 2q22, the same group identified the ACVR2A gene as a strong positional candidate gene [60].

The MDR analysis identified the *ACVR2A* rs1014064, an intronic variant (A to G), as the only significant variant. This variant associated with preeclampsia in a large Norwegian population-based study (the HUNT study), together with the other *ACVR2A* variant (rs2161983) [16], which was also evaluated and found not associated with preeclampsia in this study, and with early onset preeclampsia in a Brazilian population [15].

MDR has been used to identify the important role of epistasis in polygenic disorders, such as sporadic breast cancer [6] and essential hypertension. A two-locus model including *ACE* and *GRK4* successfully predicted the blood pressure phenotype 70.5% of the time [10]. A genetic model based on the three common *GRK4* SNPs was 94.4% predictive of salt-sensitive hypertension, while a single-locus model with only the *GRK4* A142V variant was 78.4% predictive. By contrast, for low-renin hypertension, a two-locus model that includes the *GRK4* A142V variant and cytochrome P450 11B2 (*CYP11B2*)





C-344 T was 77.8% predictive [11]. These results reflect the differences in the underlying genetics and the crucial role of epistasis in the development of the different hypertension-related phenotypes. Considering the spectrum of the clinical presentation of preeclampsia, it is conceivable that different phenotypes of the disease may involve specific gene polymorphisms, i.e., locus heterogeniety. In fact, the presence of severe forms of preeclampsia, HELLP syndrome and eclampsia, have been suggested to have their own set of predisposing gene variants. It is therefore important to know which specific genes contribute the most to their development.

We also used ViSEN software, which provides a global interaction map to identify and corroborate risk-associated SNPs, visualize putative gene interactions and generate an interaction or concept map for preeclampsia. Similar to what we observed using MDR, we detected no statistically significant two- and three-way epistatic interactions with ViSEN. Due to its limitation to analyze only up to three-way epistasis, the four-locus model that we observed with MDR was undetected. Moreover, ViSEN as a statistical tool has its own limitations, e.g., the statistical epistasis quantifications in ViSEN only consider discrete traits and cannot incorporate measures on continuous traits like age and BMI [55].

In the analysis of genetic datasets, an important consideration is the power of analytical methods to identify accurate predictive models of disease. The MDR approach overcomes the common setbacks found in other methods. It is non-parametric, model-free, and can identify high-order gene-gene interactions [6, 54, 61]. It retains its power to analyze in the presence of genotyping error and missing data (up to 5%). However, it has its own limitations, including a decrease in power in the presence of phenocopies and genetic heterogeneity [61].

With respect to marginal effects, one of the SNPs we genotyped in VEGF-A, rs3025039, that was previously identified as associating with preeclampsia in the Philippines [4] also showed a marginal association in our data set when adjusting for covariates (p = 0.022). Although this result would not stand after adjusting for multiple testing, as a replication it provided additional evidence that this variant confers pre-eclampsia risk especially since the direction of effect was the same in the present and the previous studies. A second SNP in this gene, rs722503, that was previously reported to be associated with preeclampsia, but only in pregnancies with women over 40, did not show evidence for significance in the current study (Additional file 1: Table S1). Of note, in the age and BMI adjusted model for rs722503 the p value did get smaller as compared to the unadjusted model, which is consistent with an agerelated effect. This, however, was not surprising as very few pregnancies involved women over 40 (4). In addition, this SNP did not appear in our MDR analyses when all other SNPs were included.

A notable limitation of this study is the non-inclusion of other SNPs that have been shown to be associated with preeclampsia in specific ethnic groups, or more importantly in multi-gene meta-analysis studies involving different ethnic groups [25, 27, 62, 63] and in multigene association studies [64]. These include the gene variants of FV, F2, ACE, SERPINE1, AGTR1, MTHFR, and MMP-9. The ACE gene variant is an insertion/deletion polymorphism, which cannot be detected by the method used for genotyping in our studies. The other genes, although included in the initial list for analysis, were eventually dropped from the analysis due to technical problems. These genes, however, should be included in future studies.

Conclusions

Preeclampsia is a multifactorial disease, with both genetic and environmental factors contributing to its development. Genetic variants from multiple, interrelated pathways have been suggested to contribute to the pathogenesis of the disease. The MDR algorithm enabled the analysis of high-order gene/factor interactions and identified *ACVR2A* rs1014064 as important in modulating preeclampsia risk among older Filipino women with a middle-range BMI.

Additional files

Additional file 1: Table S1. Association of SNPs, adjusting for statistically significant risk factors. (DOCX 16 kb)

Additional file 2: Table S2. MDR analysis of genetic variants, without adjusting for age and BMI. (DOCX 14 kb)

Abbreviations

ACVR2A: Activin A Receptor Type 2A; AGT: Angiotensinogen; COMT: Catechol-O-methyltransferase; CTLA4: Cytotoxic T-Lymphocyte Associated Protein 4; CYP1A2: Cytochrome P450 family 1 subfamily A member 2; DRD1: Dopamine D1 receptor; eNOS: Endothelial nitric oxide synthase; EPHX1: Epoxide hydrolase; ERAP2: Endoplasmic reticulum aminopeptidase 2; GRK4: G protein-coupled receptor kinase 4; GSTP1: Glutathione S-Transferase Pi 1; IL1A: Interleukin 1alpha; LPL: Lipoprotein lipase; MDR: Multifactor dimensionality reduction; MTR: 5-Methyltetrahydrofolate-Homocysteine Methyltransferase; MTRR: Methionine synthase reductase; PON1: Paraoxonase 1; SNPs: Single nucleotide polymorphisms; TNSF13B: Tumor necrosis factor ligand superfamily member 13B; VEGFA: Vascular endothelial growth factor A; VEGFC: Vascular endothelial growth factor C; VEGFR1: Vascular endothelial growth factor receptor 1; VEGFR3: Vascular endothelial growth factor receptor 3

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Availability of data and materials

The datasets used and analyzed during the current study are available from the corresponding author on reasonable request.

Authors' contributions

MDA, VMV PAJ, CPS conceived and designed the study; MDA, JAN performed the experiments; LDB clinical input; JAN contributed in the initial analysis, interpretation of results, and manuscript preparation; GRT, SMW analyzed the data; MDA, GRT wrote the final paper; VMV, CPS, PAJ, SMW, review and editing; CPS, PAJ, SMW, VMV, funding acquisition. All authors read and approved the final draft of the manuscript.

Ethics approval and consent to participate

This study was approved by the University of the Philippines, Philippine General Hospital (UP-PGH) Ethics Review Board. All subjects provided informed written consent.

Consent for publication

Not applicable.

Competing interests

The authors declare that they have no competing interests.

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