

Expressed sequence tag analysis in *Cycas*, the most primitive living seed plant

Eric D Brenner^{*}, Dennis W Stevenson^{*}, Richard W McCombie[†],
Manpreet S Katari[†], Stephen A Rudd[‡], Klaus FX Mayer[‡],
Peter M Palenchar[§], Suzan J Runko^{*}, Richard W Twigg^{*}, Guangwei Dai[¶],
Rob A Martienssen[¥], Phillip N Benfey[#] and Gloria M Coruzzi[§]

Addresses: ^{*}The New York Botanical Garden, 200th Street and Kazimiroff, Bronx, NY 10458-5126, USA. [†]Genome Research Center, Cold Spring Harbor Laboratory, 500 Sunnyside Blvd, Woodbury, NY 11797, USA. [‡]Institut für Bioinformatik (IBI), GSF National Research Center for Environment and Health, Ingolstädter Landstrasse 1, 85764 Neuherberg, Germany. [§]New York University, Department of Biology 1009 Main Building, New York, NY 10003, USA. [¶]Department of Computer Science, Courant Institute of Mathematical Sciences, New York University, 251 Mercer Street, New York, NY 10012-1185. [¥]Cold Spring Harbor Laboratory, 1 Bungtown Road, Cold Spring Harbor, NY 11724, USA. [#]Biology Department, Duke University, Box 91000, Durham, NC 27708, USA.

Correspondence: Eric D Brenner. E-mail: ebrenner@nybg.org

Published: 18 November 2003

Genome Biology 2003, **4**:R78

The electronic version of this article is the complete one and can be found online at <http://genomebiology.com/2003/4/12/R78>

Received: 30 June 2003

Revised: 3 October 2003

Accepted: 23 October 2003

© 2003 Brenner *et al.*; licensee BioMed Central Ltd. This is an Open Access article: verbatim copying and redistribution of this article are permitted in all media for any purpose, provided this notice is preserved along with the article's original URL.

Abstract

Background: Cycads are ancient seed plants (living fossils) with origins in the Paleozoic. Cycads are sometimes considered a 'missing link' as they exhibit characteristics intermediate between vascular non-seed plants and the more derived seed plants. Cycads have also been implicated as the source of 'Guam's dementia', possibly due to the production of S(+)-beta-methyl-alpha, beta-diaminopropionic acid (BMAA), which is an agonist of animal glutamate receptors.

Results: A total of 4,200 expressed sequence tags (ESTs) were created from *Cycas rumphii* and clustered into 2,458 contigs, of which 1,764 had low-stringency BLAST similarity to other plant genes. Among those cycad contigs with similarity to plant genes, 1,718 cycad 'hits' are to angiosperms, 1,310 match genes in gymnosperms and 734 match lower (non-seed) plants. Forty-six contigs were found that matched only genes in lower plants and gymnosperms. Upon obtaining the complete sequence from the clones of 37/46 contigs, 14 still matched only gymnosperms. Among those cycad contigs common to higher plants, ESTs were discovered that correspond to those involved in development and signaling in present-day flowering plants. We purified a cycad EST for a glutamate receptor (GLR)-like gene, as well as ESTs potentially involved in the synthesis of the GLR agonist BMAA.

Conclusions: Analysis of cycad ESTs uncovered conserved and potentially novel genes. Furthermore, the presence of a glutamate receptor agonist, as well as a glutamate receptor-like gene in cycads, supports the hypothesis that such neuroactive plant products are not merely herbivore deterrents but may also serve a role in plant signaling.

Background

The Cycadales (cycads) are the most primitive living seed plants and have endured over 270-280 million years since their origins in the Lower Permian [1,2]. Cycads have a fern or palm-like appearance, largely due to their pinnately compound leaves (Figure 1a,b). Unlike ferns or palms, however, cycads belong to the gymnosperms, or non-flowering seed plants. Of the four orders that comprise the gymnosperms, the Cycadales are considered to be the most ancestral compared to Ginkgoales, Gnetales and Coniferales (Figure 2) [3,4]. Cycads (non-flowering seed plants) exhibit a number of characteristics that reflect their evolutionary position between ferns (non-seed plants) and angiosperms (flowering seed plants). Such characteristics include pollen tubes, which release motile sperm before fertilization; dichotomous branching (versus axillary branching in higher plants); and ovules, which contain a large, free-nuclear megagametophytic stage, that are borne on the margins of leaf-like megasporophylls [5-7]. These characteristics, among others, place cycads at a key node in plant evolution.

In addition to their evolutionary importance, cycads have also been studied in the field of medicine, because they produce neurotoxic compounds. In particular, cycads produce a secondary compound, BMAA (S(+)-beta-methyl-alpha, beta-diaminopropionic acid), which has been implicated as the possible cause of Guam's dementia [8]. This disorder occurs among the indigenous Chomorro people, who ate cycads as food, and now suffer from Alzheimer's and Parkinson's dementia [9-11]. BMAA production is unique to cycads, where it has been used as a monophyletic character in plant classification [7]. It is present in both seeds and leaves of all genera of the Cycadaceae [12]. BMAA is neurotoxic in mammals [9,13] because of its excitotoxic action as an agonist of glutamate receptors (GLRs) [14]. The discovery of GLR-like genes in *Arabidopsis* suggests that plant-derived GLR agonists, as well as acting as potential deterrents to herbivores, might also operate in signaling during plant growth and development, by interacting with native plant GLRs [15]. In partial support of this hypothesis, BMAA was shown to affect the development of *Arabidopsis* and consequently was used in a pharmacologically-based genetic screen to isolate mutants in a putative GLR pathway in *Arabidopsis* [16].

Despite the importance of cycads in the study of plant evolution, and their role in neurological disorders in humans, nothing is known about the genes responsible for these traits - primarily because cycads are recalcitrant to genetic analysis. Unlike genetically tractable plants such as tomato, maize and *Arabidopsis*, cycads are dioecious (male and female organs on separate plants), produce a limited number of seeds and take up to 30 years to become reproductive. Furthermore, cycad genomes are large (20,000-30,000 million base-pairs (Mbp)) [17,18] compared to *Arabidopsis* (125 Mbp) [19]. Consequently, cycads have remained outside the realm of both traditional genetic studies and modern genome-sequencing

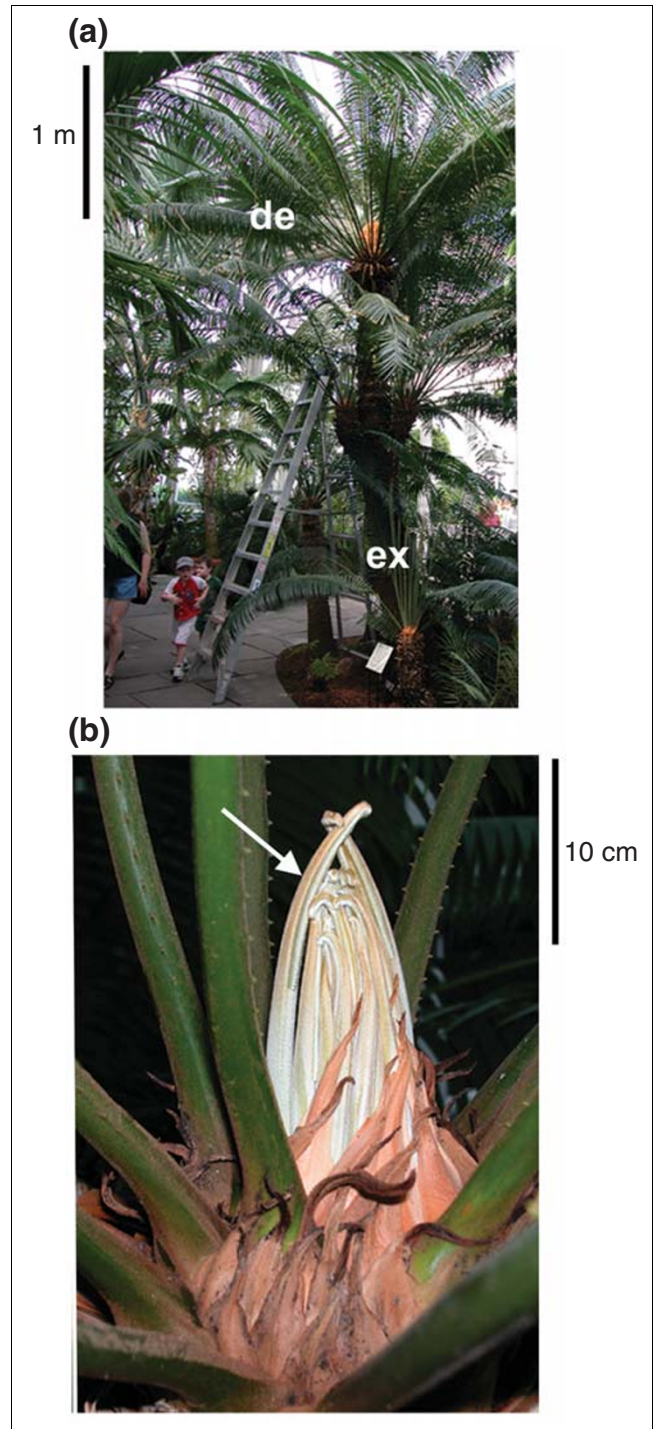


Figure 1
Cycas rumphii used for cDNA library construction. **(a)** Mature cycad trunk with developed (de) leaves and young, expanding (ex) leaves. **(b)** Young emergent leaves (arrow) at the crown, which were used to generate a cDNA library database.

initiatives. Fortunately, recent advances in plant genomics [20,21], provide new tools to study genetically complex species such as cycads. In particular, the availability of the

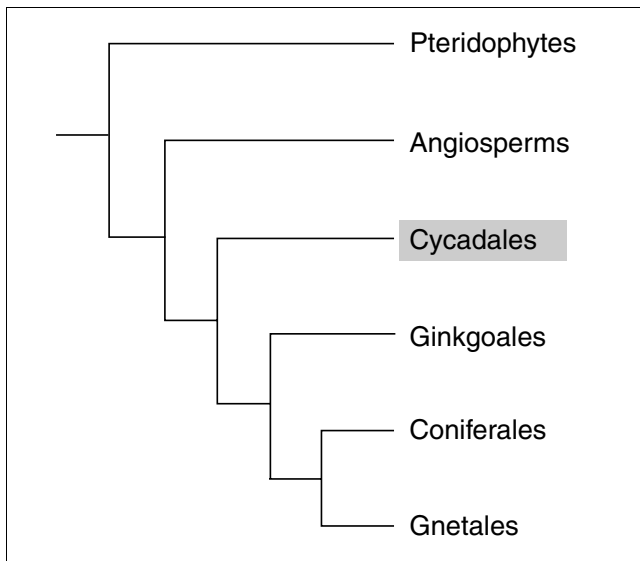


Figure 2
Cycads are the sister group to the seed plants. A phylogenetic tree shows that cycads (highlighted) are the least derived of the seed plants. Cycads are believed to be the oldest extant seed plants.

complete, annotated sequence of two angiosperm genomes - the dicot *Arabidopsis thaliana* [19,22] and the monocot rice (*Oryza sativa*) [23,24] - now makes it possible to study the genomes of evolutionarily important plants by comparing the expressed genes of cycads (ESTs) to the complete genomes of higher plants.

To begin a survey of expressed genes of cycads, the genus *Cycas* was chosen for expressed sequence tag (EST) analysis because *Cycas* is at the basal node - that is, the sister taxon to the rest of the Cycadales [25-27]. Furthermore, the species *Cycas rumphii* Miq. was selected for this analysis as it is suspected to be the dietary cause of Guam's dementia. It has been established that in *C. rumphii*, from which the EST library was made, BMAA levels are nearly 0.1 mg/g tissue [28]. Because of its evolutionary position as a key node within the plant kingdom, as well as its medicinal significance to humans, *Cycas* is ideally suited for genomic prospecting [29].

Here, we describe the construction of a cycad EST database from RNA of young *C. rumphii* leaves. Using this database, our comparison revealed conserved genes, including those involved in development and signaling in present-day flowering plants. Our analysis defined a set of cycad clones that have no similarity to any known angiosperm genes, but possess similarity only to genes of other gymnosperms. Furthermore, as a first step to understanding the function of neurotoxins produced in cycads, we defined a number of candidate genes that encode putative enzymes involved in the biosynthesis of BMAA, as well as a cycad GLR-like gene, the suspected target of BMAA action in animal brains. These cDNA tools will be useful to test whether BMAA, which has

been postulated to serve as an herbivore deterrent [5], also acts to regulate GLR function in plants.

Results

Construction of a cDNA library from *Cycas rumphii*

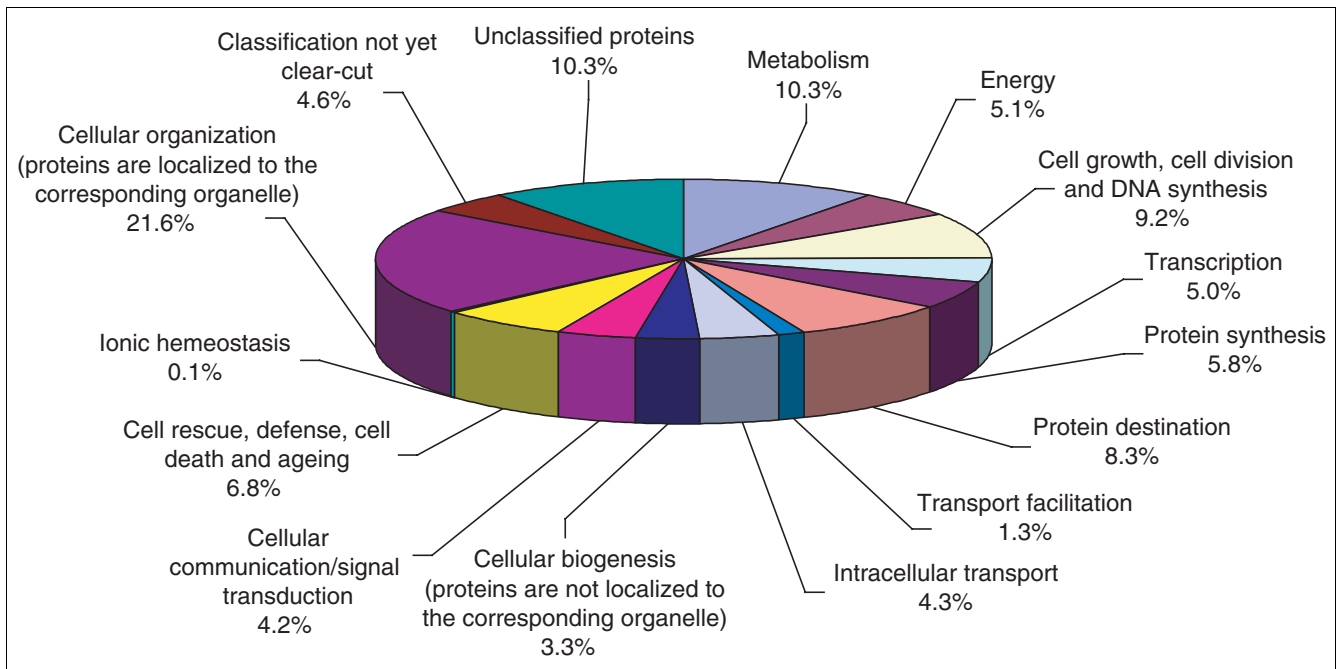
At maturity, *C. rumphii* leaves can reach up to 3 meters in length (Figure 1a). The tissue used in this study consisted of 10 to 40 cm of the immature leaf terminus protruding from the crown collected shortly after emergence (Figure 1b). Immature leaves consist of a petiole, a central rachis and circinate leaflets composed of both expanding and meristematic cells [30]. RNA extracted from this tissue was used to construct a cDNA library from *C. rumphii*. Size fractionation was used to enrich for full-length cDNAs during library construction. It was determined that 53% of the cDNA clones were over 500 bp long. From this cDNA library, 4,210 sequence reads (ESTs) were generated. The majority of these reads (3,917) were generated from the 5' end of the cDNA; however, a small subgroup (293) were sequenced from the 3' end. Cluster analysis performed at the Munich Information Center for Protein Sequences (MIPS) of the entire EST dataset produced a UniGene set of 2,458 contigs consisting of 1,917 singletons and 541 assemblies. Of the clustered ESTs, the longest contig was 1,836 bp. The entire UniGene set can be viewed on the MIPS Sputnik website [31], which features sequence annotations and peptide sequence predictions. At the MIPS Sputnik site there are links to download the complete cycad sequences as an EST fasta file, a cluster fasta file or as the derived peptide fasta file.

Classification of *C. rumphii* ESTs by functional categories

Each contig from the database was automatically assigned to a functional category on the basis of its top match against the complete genomic sequence of *Saccharomyces cerevisiae* and *A. thaliana* databases using BLASTP. A non-stringent expect value (E-value) of $<1e-10$ was chosen as the threshold. The pie chart in Figure 3 illustrates the relative fraction that each functional category comprises within the entire UniGene set. The four largest predominant categories of cycad ESTs according to this functional categorization are: 'cellular organization' (22%), 'metabolism' (10%), 'unclassified proteins' (10%), and 'cell growth, cell division/DNA synthesis' (9%).

Cycad contig matches to genes in angiosperms, gymnosperms and lower plants

Using TBLASTX, a comparison was made between the *C. rumphii* UniGene set versus all available ESTs from GenBank and predicted *Arabidopsis* genes from The *Arabidopsis* Information Resource (TAIR). Both EST and predicted genes were grouped into three subcategories: angiosperms, gymnosperms, and lower plants. The angiosperm database encompasses all annotated rice and *Arabidopsis* genes identified from their respective genomic sequences, as well as all higher

**Figure 3**

Functional gene categories of cycad ESTs. Clustered cycad ESTs were assigned to a functional category based on top BLASTP similarity scores. An expect value (E-value) of $> 1e-10$ was chosen as the cut-off threshold. The analysis was performed at the Munich Information Center for Protein Sequences.

plant ESTs. The gymnosperm database contains ESTs from all gymnosperms, the majority of which came from the *Pinus taeda* EST sequencing project [32,33]. The lower plant databases included genes from all remaining plant ESTs including ferns, fern allies, bryophytes and algae available in GenBank. The angiosperm subgroup consisted of 84.5%, the gymnosperms 6.5% and lower plants 9.0% of the total genes used in this analysis.

The Venn diagram shown in Figure 4 displays the total number of cycad contigs shared between one or more of the plant gene datasets at very low BLAST stringency values (expect $< 1e-5$). The majority of cycad contigs (1,764/2,458) have counterparts in other plants, leaving 694 with no match to other plant genes. As one would expect, most *Cycas* hits (1,718) are to angiosperms, because of the predominance of angiosperm accessions in GenBank. Many of the cycad matches to angiosperms also match gymnosperms and/or lower plants (1,416). There are 1,310 cycad contigs that match gymnosperm genes and 734 that match genes from lower plants.

Full-length sequencing of cycad clones that match only gymnosperm genes

As shown in Figure 4, 44 *Cycas* ESTs specifically match only genes in the gymnosperm subgroup. Two additional *Cycas* ESTs match genes from gymnosperms and lower plants, but not angiosperms. To further analyze these 46 contigs that match only gymnosperms and/or lower plants, we next sequenced these *Cycas* cDNAs in their entirety to determine

whether this 'gymnosperm/lower plant' specific grouping held up when the remaining portions of the cDNA were sequenced. Because ESTs, even when clustered into contigs, usually represent only a portion of the actual gene (particularly for genes poorly represented in the library) 37 of the 46 *Cycas* cDNAs were sequenced in their entirety (the remaining nine clones were not successfully recovered for sequencing), and this sequence can be downloaded from the Internet [34]. Of these 37 fully sequenced cDNAs, 14 clones still showed no similarity to any known angiosperm genes, even at this low stringency cut-off. The insert size for each clone ranges from 586 bp to 1,899 bp, with predicted open reading frames (ORFs) varying from 69 to 527 residues (Table 1). None of these 14 *Cycas* cDNA clones is homologous to any known genes outside the plant kingdom, although Interpro analysis identified a small number of conserved motifs, which are listed in Table 1. To confirm that these genes were indeed derived from *C. rumphii*, gene-specific primers designed to each of the 14 genes were able to amplify a fragment from genomic DNA isolated from a different *C. rumphii* specimen and different tissue (sporophyll) from the source tissue of the cDNA library (data not shown). This distinct *C. rumphii* specimen was cultivated in a geographically separate location (Florida) from the cDNA source *C. rumphii* specimen used for cDNA library construction (New York).

Cycad genes similar to developmental regulators

A survey of the cycad EST dataset reveals a surprisingly large number of genes with highest similarity (BLASTP score $< e-5$) to genes with defined roles in growth and development in

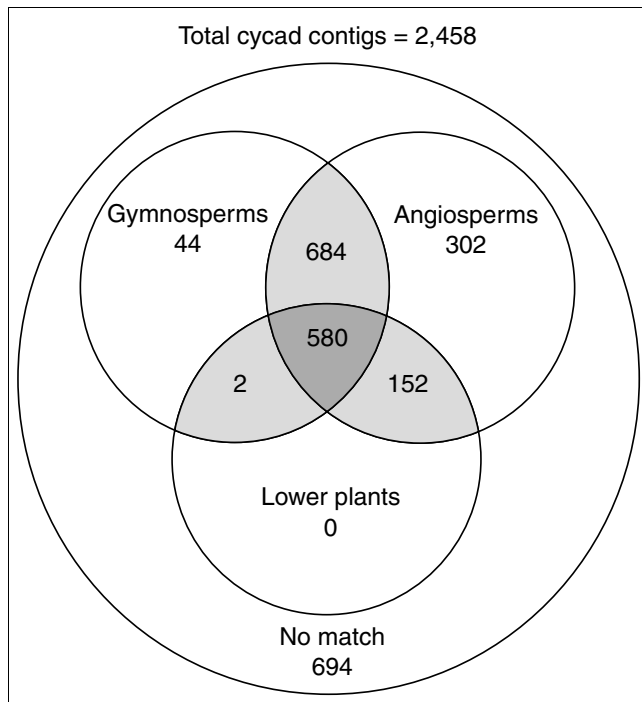


Figure 4
A Venn diagram reveals shared gene sets between cycad contigs versus lower plants, gymnosperms and/or angiosperms. BLASTX (cut-off E value > 1e-5) was used to compare the cycad contigs against all angiosperm ESTs and annotated genes from the full *Arabidopsis* and rice genome sequence from GenBank. Genes that do not have a match to angiosperm genes were then compared to available ESTs from all gymnosperms or lower-plant ESTs available in GenBank. Genes that are common to cycads and more than one group are shown in the intersecting (shaded) regions.

angiosperms (Table 2). Some of these *Cycas* genes have similarity to *Arabidopsis* transcription factors, including *CONSTANS* [35,36], two distinct homeobox genes [37] and a *YABBY* gene [38,39]. Other cycad ESTs have similarity to other regulators of *Arabidopsis* development, including *ARGONAUT* [40] and *COP9* [41,42].

Cycas genes with similarity to Arabidopsis genes involved in signaling

A number of genes in our cycad EST library showed similarity to components of signaling pathways found in higher plants (Table 2). These genes include a photolyase blue-light receptor, genes involved in secondary signaling (including those for calmodulin, kinases, and phosphatases), a 14-3-3 protein, and genes involved in phytohormonal responses, including auxin (IAA-9 and IAA-13) pathways as reviewed in Chory and Wu [43]. Surprisingly, a *Cycas* EST with high similarity to plant GLR-like genes was also found (Table 2) [15,44]. The presence of a GLR-like gene in cycads is of particular interest as it relates to BMAA, as described below.

A predicted pathway for BMAA synthesis in *Cycas* is supported by EST analysis

BMAA, an agonist of mammalian GLRs, is a suspect causative agent of neurological disorders [9,13]. However, nothing is known about the genes and enzymes involved in the biosynthesis of BMAA. Because the structure of BMAA is similar to other beta-substituted alanines [45,46], it is likely that BMAA biosynthesis utilizes phosphoserine, cysteine, o-acetylserine or cyanoalanine as a beginning substrate. On this basis, a likely BMAA biosynthetic pathway is shown in Figure 5. This would require a two-step reaction initiated with the transfer of NH_3 at the beta-carbon of the substituted alanine (Figure 5a), followed by an addition of CH_3 (Figure 5b) to produce BMAA (Figure 5c). NH_3 transfer would require a nucleophilic reaction catalyzed by a cysteine synthase-like protein. A preliminary survey of genes in the cycad EST library identified candidate genes for both of these enzymatic steps (Table 2). The cycad leaf EST library contains two ESTs, which each encode a cysteine synthase. To catalyze the second step of BMAA synthesis, the EST library contains two potential methyltransferases (caffeic acid O-methyltransferase II and caffeoyl-CoA 3-O-methyltransferase). The second step would require a methyl donor, the most likely candidate being S-adenosylmethionine (SAdM). Consumption of SAdM would require the presence of enzymes to regenerate SAdM. A number of cycad ESTs can be implicated in SAdM recycling including: adenosylhomocysteinase, S-adenosylmethionine synthetase and homocysteine methyltransferase. Taken together, the cycad EST library contains candidate genes for all of the enzymes predicted to be present during the biosynthesis of BMAA.

Discussion

Cycads can be regarded as living fossils

Extant genera, such as *Cycas*, have changed little in morphology from their extinct relatives, such as *Crossozamia*, which existed during the Permian [1,2]. The study of cycads has proved to be useful in reconstructing plant evolution, in particular in understanding the rise of important plant structural innovations such as the evolution of seeds [47]. Cycads also produce a variety of neuroactive compounds, some of which are suspected to be the source of Guam's dementia [11,48]. However, despite their scientific importance in plant biology and medicine, virtually nothing is known regarding gene expression, development and signaling in the Cycadales. As a first step in this direction, a cDNA library was made from young, developing *C. rumphii* leaves to produce a cycad EST database.

A cycad EST database: a foundation to study the evolution of early seed plants

One advantage of a genomics approach is that it provides rapid access to genes important for evolutionary studies. The more traditional homology-based gene-cloning approach is limited by tedious gene-by-gene purification. It is also limited

Table 1**Fully sequenced cycad clones from contigs that match only genes in gymnosperms**

Contig	GenBank accession number	Transcript length (bp)	Peptide (residues)	InterPro result
gy79c04_704	CB090702	586	72	No matches found
gy78g12_244	CB090673	627	84	No matches found
te82h02_741	CB093328	647	107	No matches found
he95e08_721	CB091708	651	114	No matches found
hf04g07_288	CB092366	684	141	ASP_RICH (unintegrated)
hk42a07_743	CB093061	790	142	No matches found
gp23c01_369	CB089407	791	189	No matches found
gp26f08_297	CB089628	827	69	No matches found
gy82g05_181	CB090964	840	118	No matches found
he92f06_688	CB091462	935	170	No matches found
gy81e11_544	CB090877	948	211	ASP_RICH (unintegrated)
he97c12_740	CB091858	965	140	No matches found
gp32b03_724	CB089926	1311	335	Peptidoglycan-binding LysM
te83a03_729	CB093338	1899	527	No matches found
Average		893	173	

All available ESTs and annotated genes from GenBank were divided into three categories (angiosperms, gymnosperms and lower plants) and compared with the *C. rumpfii* UniGene set. Forty-six cycad ESTs that had no similarity to angiosperm genes, but matched gymnosperm and lower plant genes, were fully sequenced, of which 14 clones (listed) still have no similarity to angiosperms. To confirm that these genes were of cycad origin, all 14 were successfully amplified from the DNA of a *C. rumpfii* specimen other than the one used to construct the cDNA library. The transcript length, as well as the predicted translation product size, is shown. Interpro analysis identified conserved motifs in three of these cycad ESTs as shown.

in that it may miss related genes if the degeneracy is too great or if nonconserved regions of the protein are chosen during primer design. Finally, the targeted gene approach can never be used to discover new genes.

Sequence analysis of contigs with BLAST similarity to gymnosperms but not angiosperms

An EST project in *Pinus taeda* (loblolly pine) sampled 59,797 transcripts from wood-forming tissues [32]. In this analysis, 66 *P. taeda* contigs showed BLAST similarity at low stringency only to other gymnosperms. Similarly, in our analysis, we found 46 cycad contigs that only matched gymnosperms (including *P. taeda*) and/or lower plant ESTs, but were not found in the genomes of higher plants or non-plants. Complete sequencing of 37 of these cycad cDNA clones showed that 14 clones, ranging in length from 586 to 1,899 bp, were still found only in other gymnosperms. Having no homology to the completely sequenced genomes of two different angiosperm species - *Arabidopsis* [19] (a dicot) and rice [23,24] (a monocot) - suggests that these 14 genes are found only in gymnosperms or lower plants, in which genomic studies have only just begun. However, because ESTs as well as contigs usually represent only a portion of the full-length gene sequence, these results are preliminary. For instance, in *P. taeda*, larger contigs have a higher BLAST match rate to other plant genes than do shorter contigs [32]. Thus, these preliminary results of clade specificity are tenuous and pre-

sumably will change as more ESTs, as well as full-length gene sequences, from cycads and other species are generated in the future.

Genes with potential developmental roles in cycads

As in higher plants, cycad leaves are derived from the shoot apical meristem (SAM) [30]. In *Cycas* leaflet primordia, meristematic growth ceases at the apex, while proceeding basipetally where it becomes localized to the leaflet margins [30]. The presence of these marginal meristems may explain why a surprising number of developmental genes were identified in a relatively small number of ESTs from young cycad leaves (Table 2).

A gene with identity to the *YABBY* gene family was among the cycad ESTs. *YABBY* genes encode transcription factors expressed on the abaxial side of all lateral organs that promote abaxial cell fate [38]. In *Arabidopsis*, mutations in the *YABBY* gene *INO* (*INNER-NO-OUTER*), lead to the loss of the outer integument [49] reminiscent of gymnosperm (and cycad) unitegmy (the presence of a single integument). Unitegmy is considered to be the ancestral condition in seed plants [5,47]. An analysis of *YABBY* gene expression in cycads may help to explain the origin of the integument in gymnosperms, and/or possibly the second integument in angiosperms. One cycad EST from the library has highest similarity to *COP9*. *COP9* encodes a subunit of the COP9

Table 2**Genes in *Cycas rumphii* with potential roles in signaling, development and biosynthesis of BMAA**

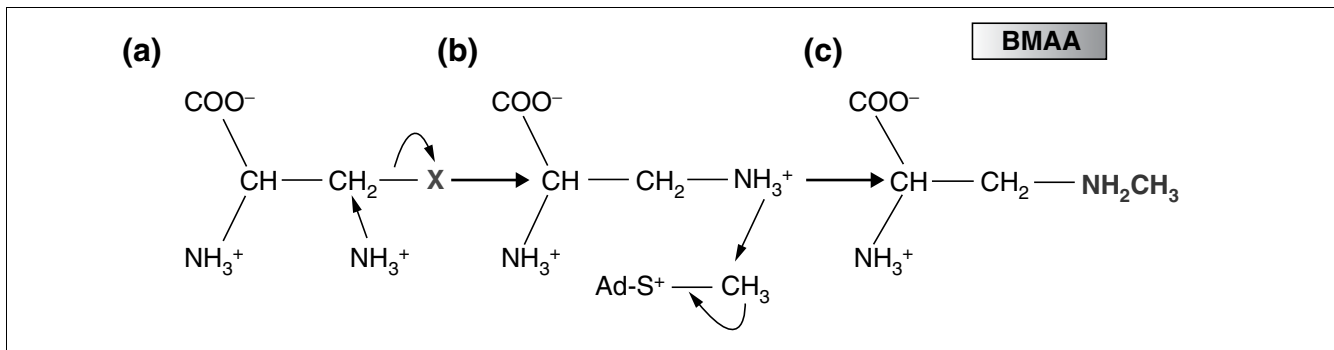
	GenBank accession number	Subject description	E-value	% ID	Score	
Development	CB092871	Argonaute-like protein 1 (AGO1) <i>Arabidopsis thaliana</i>	8.00e-10	0.85	34	
	CB090033	YABBY2 <i>A. thaliana</i>	2.00e-36	0.58	151	
	CB089539	Multisubunit regulator protein COP9 - spinach COP9 <i>Spinacia oleracea</i>	2.00e-31	0.62	98	
	CB092157	CONSTANS B-box zinc finger family protein <i>A. thaliana</i>	1.00e-47	0.48	221	
	CB092462	CRHB3 homeoprotein <i>Ceratopteris richardii</i>	3.00e-44	0.70	131	
	CB089344	Homeodomain protein HB2 <i>Picea abies</i>	3.00e-29	0.62	117	
Signaling	CB089945	Photolyase/blue-light receptor PHR2	8.00e-76	0.69	197	
	CB091652	Putative glutamate receptor protein GLR3.4b	2.00e-45	0.54	161	
	CB093220	Calmodulin-like protein; protein ids At5g44460.1 <i>A. thaliana</i>	3.00e-07	0.58	45	
	CB089469	14-3-3 protein <i>Fritillaria cirrhosa</i>	8.00e-38	0.80	94	
	CB091066	Ser/Thr protein kinase isolog; protein ids, supported by cDNAs <i>Arabidopsis</i>	1.00e-10	0.28	185	
	CB090652	Ser/Thr specific protein phosphatase 2A B regulatory subunit beta <i>Medicago</i>	4.00e-86	0.94	162	
	CB093099	Auxin regulated protein (IAA13) <i>A. thaliana</i>	1.00e-34	0.63	125	
	CB089385	Auxin-induced protein IAA9 <i>A. thaliana</i>	8.00e-29	0.55	111	
	Biosynthetic enzymes of cycad specific phytochemicals (BMAA)					
	Cysteine synthase	CB089577	Cysteine synthase (O-acetylserine sulfhydrylase)	3.00e-50	0.75	128
CB092214		Plastid cysteine synthase 2 <i>Solanum tuberosum</i>	5.00e-27	0.64	83	
Methyl transferases	CB091906	Caffeic acid O-methyltransferase II <i>Nicotiana tabacum</i>	3.00e-35	0.56	122	
	CB090738	Caffeoyl-CoA 3-O-methyltransferase <i>Oryza sativa</i>	1.00e-37	0.47	188	
SAdM metabolism						
Adenosylhomocysteinase (S-adenosyl-L-homocysteine hydrolase)	CB091477	Adenosylhomocysteinase <i>Phalaenopsis</i>	1.00e-87	0.84	185	
	CB091821	Adenosylhomocysteinase <i>Triticum aestivum</i>	3.00e-78	0.90	156	
	CB090818	Adenosylhomocysteinase <i>Medicago sativa</i>	2.00e-18	0.68	66	
S-adenosylmethionine synthase	CB091682	S-adenosylmethionine synthetase <i>Brassica juncea</i>	4.00e-90	0.94	167	
	CB090997	S-adenosylmethionine synthetase (methionine adenosyltransferase) <i>Petunia</i>	1.00e-69	0.94	133	
Homocysteine methyltransferase	CB090407	S-adenosyl-L-methionine synthetase <i>Elaeagnus umbellata</i>	1.00e-93	0.88	191	
	CB092344	Methionine synthase protein <i>Sorghum bicolor</i>	4.00e-94	0.90	190	
	CB091647	5-methyltetrahydropteroyltriglutamate - homocysteine S-methyltransferase	3.00e-79	0.76	205	

C. rumphii ESTs were compared to GenBank with a BLASTP score < e-5. The top match produced from the BLAST search to the cycad EST is listed under subject description.

signalosome complex, which controls multiple signaling pathways that regulate development in all eukaryotes [42,50]. In *Arabidopsis*, the *cop9* mutant is constitutively photomorphogenic in dark-grown seedlings [51]. Some gymnosperms, (in particular the Coniferales) are constitutively photomorphogenic when grown in the dark [52,53]. As yet, the phenotype of dark-grown cycad seedlings has not been fully evaluated. The discovery of a gene encoding a putative subunit of the COP9 complex in cycads could be a first step to

define the ancestral, developmental role of the signalosome in gymnosperms, particularly with regard to its role in photomorphogenesis.

Another gene potentially involved in cycad development has highest similarity to the *CONSTANS* gene family, which are regulators of flowering time that follow internal and external (environmental) inputs in *Arabidopsis* [35]. Because cycads predate the evolution of flowers, it would be of interest to

**Figure 5**

Predicted two-step pathway for the biosynthesis for BMAA in cycads. A postulated route for BMAA biosynthesis supported by cycad EST analysis is shown. In this simple, two-step scheme, BMAA synthesis begins with (a) the transfer of NH_3 to β -substituted alanine, where X = phosphoserine, cysteine, o-acetylserine or cyanoalanine, to form (b) an intermediate. The reaction is catalyzed by a cysteine synthase-like enzyme. This step is followed by transfer of a methyl group from S-adenosylmethionine (Ad-S-CH₃) to the new amine group by a methyltransferase, which would lead to the formation of (c) BMAA. Candidate cycad genes encoding probable cysteine synthase-like enzymes and methyltransferase, as well as S-adenosylmethionine-regenerating enzymes that were identified in the cycad EST collection are listed in Table 2.

determine if *CONSTANS* genes in cycads temporally regulate sporophyll and cone induction, which typically follows a yearly cycle [5,6].

A cycad GLR-like gene expressed in tissue producing the GLR agonist BMAA

An unexpected finding of the *Arabidopsis* EST genome project was the discovery of GLR-like genes, or 'neural' receptor genes, in plants [15]. In *Arabidopsis*, the GLR-like gene family comprises 20 members [54]. Pharmacological evidence has linked *Arabidopsis* GLRs to light and/or growth signaling pathways [15,16]. Supplying exogenous BMAA to growing *Arabidopsis* seedlings was shown to block light-induced hypocotyl shortening and cotyledon expansion [16]. Because BMAA has such profound effects on *Arabidopsis* development, we have previously proposed that BMAA, or glutamate, the natural agonist of GLRs in humans, plays a physiological role in *Arabidopsis* [15,16]. Continuing genetic studies in *Arabidopsis* aim to identify the endogenous components of the BMAA-targeted pathway in plants [16].

Cycads produce BMAA [8,9]. One EST uncovered in the *C. rumphii* leaf cDNA library has a high degree of similarity to plant GLR genes (Table 2). This discovery is intriguing, because it suggests that BMAA might be interacting with native GLR gene products in cycads. To further investigate the relationship between cycad GLR genes and BMAA, we sought to identify cycad genes potentially involved in BMAA synthesis.

From the structure of BMAA, we hypothesized that cycads produce BMAA in a simple two-step pathway, beginning with a β -substituted alanine. To enhance the probability of finding genes involved in BMAA synthesis, we made our cDNA library from tissues that produce relatively large quantities of BMAA (nearly 0.1 mg/g tissue) [28]. According to Ohlrogge

and Benning, there is a 95% chance of finding the gene for a specified enzyme when it is expressed at 0.1% mRNA/protein by sampling only 3,000 ESTs from an unnormalized library [55]. Considering the prevalence of BMAA in *Cycas*, it is not surprising that we discovered cognate genes for the predicted enzymes for this BMAA biosynthetic pathway in the cycad EST database (Figure 5, Table 2). Future biochemical and molecular studies will determine if these genes play a part in BMAA synthesis.

The discovery of GLR-like genes in *C. rumphii* raises the intriguing possibility that endogenous BMAA may interact with native cycad GLRs as a regulatory molecule. Future studies aim to understand the role of GLRs in plants, as well as the role of BMAA in herbivore defense versus endogenous signaling. The production of additional ESTs from cycads will increase the variety of genes available for study, so that a detailed expression profile can be evaluated during cycad development. Complementation studies of these genes in orthologous *Arabidopsis* mutations will help define their roles in cycads. This combined approach to studying cycad gene structure and function will help reveal molecular changes in genes involved in signaling, metabolic and developmental pathways that led to the rise of the seed plants.

Materials and methods

Tissue collection and library construction and DNA purification

Newly emerged immature leaves from the crown of a *C. rumphii* tree, accession 808/59 A, were collected from the New York Botanical Garden Conservatory. Leaves collected ranged from 5 to 30 cm in length. Tissue was frozen in liquid nitrogen. RNA was extracted from pulverized, frozen tissue in a mortar and pestle with the RNeasy maxi kit (Qiagen, Valencia, CA) according to the manufacturer's protocol. Purified

Cycas RNA was precipitated in 2 M LiCl, washed twice with 70% ethanol, and resuspended in 50 μ l water. Poly(A) RNA was subsequently purified from total RNA with the Oligotex Maxi kit (Qiagen). A cDNA library was constructed using the Lambda ZAP-CMV cDNA synthesis kit (Stratagene, La Jolla, CA) using 10 μ g poly(A) RNA. Before cloning, cDNA was size fractionated over a Sepharose CL-6b column. The first five fractions containing a total of around 100 ng cDNA were collected, pooled and precipitated in 70% ethanol/0.3 M sodium acetate and resuspended in 3.5 μ l water. cDNA (0.5 μ l) was then directionally subcloned into the vector at the *Eco*RI and *Xho*I sites.

DNA was collected from unemerged *C. rumphii* sporophylls using the DNeasy purification kit (Qiagen).

EST sequencing

Plasmid DNA was collected as described in the manual (Stratagene) catalog number 200450 in the *in vivo* mass excision section. Sequence analysis was performed at Cold Spring Harbor Laboratory using an ABI 3700 capillary sequencer (Applied Biosystems, Foster City, CA) for separation and nucleotide detection. Reactions were performed using a 1/16 Big Dye Terminator. Sequencing was performed with either the -21 M13 forward and/or reverse primer.

EST clustering and assignment into functional categories

The EST sequences were clustered and assembled using the HarvESTer application (Biomax informatics, Martinsried, Germany). The default HarvESTer settings were optimized to screen for vector against the UniVec nonredundant database of vector and polylinker sequences [56]. The Hashed Position Tree (HPT) clustering used a similarity link threshold of 0.7 and a maximum distance of six steps was required to define a cluster from the similarity network, thus encouraging the separation of likely paralogs. Cluster consensus sequences and concomitant alignments were derived from the HPT clusters using the CAP3 application with default settings. The HarvESTer assemblies and coordinate alignments were imported into the Sputnik EST and cluster analysis application [57].

Peptide extraction

BLASTX [58] was performed against a nonredundant protein database for each of the cluster consensus sequences. Likely coding sequences were derived for each cluster consensus sequence by parsing the best BLASTX match and filtering the results using the arbitrary expect value $<1e-10$. Diconon usage frequencies and probabilities were extracted using tools from the ESTate package [59]. A peptide sequence was predicted for each of the cluster consensus sequences using the Framefinder application from the ESTate package with the cycad-specific codon usage statistics. Framefinder was run using the default parameters. The derived peptide sequences were used as the basic scaffold for peptide-based annotation in Sputnik.

Sequence annotation

Sequence annotation on each of the cycad cluster consensus sequences and derived peptides were performed within the Sputnik application. Results were assessed for possible contamination by searching for homology to the *Escherichia coli* and human genomes and were scored for homology to a wide range of noncoding RNAs and plant chloroplast and mitochondrial genomes. Similarity searches were performed using the BLAST application [58] and results were filtered using the expectation value $<1e-10$. Functional assignment was performed on both cluster consensus sequence and the peptide sequence. Assignments were made using BLASTX and BLASTP respectively against the MIPS catalog of functionally assigned proteins (funcat) [60,61]; tentative functional assignments were filtered using the expectation value $<1e-10$.

Categorization of cycad contigs

All cycad contigs sequences were aligned against the PlantEST database using TblastX [58] and BlastX against the NR(aa) database. The PlantEST database was created by downloading all plant ESTs in GenBank and assembling them using Phrap [60,61]. Todd Wood from Clemson University provided the PERL script that creates the PlantEST databases as described above. The NR(aa) database is a nonredundant database of protein sequences from GenBank.

Determination of gymnosperm-specific genes

All available plant ESTs were downloaded from GenBank and separated into three datasets consisting of angiosperms (monocots and dicots), gymnosperms, or lower plants (ferns, mosses and algae). Downloaded ESTs were assembled using Phrap [60,61]. All matches with an expect value $<1e-5$ were considered significant.

Acknowledgements

We thank Francisco Coelho, Javier Francisco Ortega and the Montgomery Botanical Center, Florida for providing plant tissue; Dan Chamovitz and Trevor Stokes for reviewing the manuscript; Vivekanand Balija and Neilay Dedhia for sequence generation and curation; Eduardo de la Torre and Eugene Mueller for helpful discussions; and Alex Clark and Ayelet Levy for technical help. Funding for this work comes from the Plant Genomics Consortium. The Plant Genomics Consortium is made possible by the generosity of the Altria Group, The Mary Flagler Cary Charitable Trust, The Eppley Foundation for Research, The Leon Lowenstein Foundation, The Ambrose Monell Foundation, The Wallace Genetic Foundation and the National Institutes of Health, grant number GM-32877 to G.C. and an NIH postdoctoral fellowship to E.B.

References

1. Mamay SH: **Cycads: fossil evidence of late paleozoic origin.** *Science* 1969, **164**:295-296.
2. Gao Z and Thomas BA: **A review of fossil cycad megasporophylls, with new evidence of *Crossozamia pomel* and its associated leaves from the lower Permian of Taiyuan, China.** *Rev Palaeobot Palynol* 1989, **60**:205-223.
3. Nixon K, Crepet W, Stevenson DW and Friis E: **A reevaluation of seed plant phylogeny.** *Ann Missouri Bot Garden* 1994, **81**:484-583.
4. Soltis DE, Soltis PS and Zanis MJ: **Phylogeny of seed plants based on evidence from eight genes.** *Am J Bot* 2002, **89**:1670-1681.

5. Norstog KJ and Nicholls TJ: *The Biology of the Cycads* Ithaca, NY: Cornell University Press; 1997.
6. Chamberlain C: *The Living Cycads* Chicago: University of Chicago Press; 1919.
7. Loconte H and Stevenson DW: **Cladistics of the Spermatophyta.** *Brittonia* 1990, **42**:197-211.
8. Vega A and Bell EA: **Alpha-amino-beta-methylaminopropionic acid, a new amino acid from seeds of *Cycas circinalis*.** *Phytochemistry* 1967, **6**:759-762.
9. Spencer PS, Hunn PB, Nugon J, Ludolph AC, Ross SM, Roy DH and Robertson RC: **Guam amyotrophic lateral sclerosis-Parkinsonism-dementia linked to a plant excitant neurotoxin.** *Science* 1987, **237**:517-522.
10. Whiting MG: **Toxicity of cycads.** *Econ Bot* 1963, **17**:271-302.
11. Kurland LT: **An appraisal of the neurotoxicity of cycad and the etiology of amotrophic lateral sclerosis on Guam.** *Fed Proc* 1972, **31**:1540-1543.
12. Charlton TS, Marini AM, Markey SP, Norstog K and Duncan MW: **Quantification of the neurotoxin 2-amino-3-(methylamino)propanoic acid (BMAA) in Cycadalea.** *Phytochemistry* 1992, **31**:3429-3432.
13. Seawright AA, Ng JC, Oelrichs PB, Sani Y, Nolan CC, Lister AT, Holton J, Ray DE and Osborne R: **Recent toxicity studies in animals using chemicals derived from cycads.** In *Biology and Conservation of Cycads - Proceedings of the Fourth International Conference on Cycad Biology* 1996 Beijing: International Academic Publishers; 1999.
14. Brownson D, Mabry T and Leslie S: **The cycad neurotoxic amino acid, beta-N-methylamino-L-alanine (BMAA), elevates intracellular calcium levels in dissociated rat brain cells.** *J Ethnopharmacol* 2002, **82**:159-167.
15. Lam HM, Chiu J, Hsieh MH, Meisel L, Oliveira IC, Shin M and Coruzzi G: **Glutamate-receptor genes in plants.** *Nature* 1998, **396**:125-126.
16. Brenner ED, Martinez-Barboza N, Clark AP, Liang QS, Stevenson DW and Coruzzi GM: ***Arabidopsis* mutants resistant to S(+)-beta-methyl-alpha, beta-diaminopropionic acid, a cycad-derived glutamate receptor agonist.** *Plant Physiol* 2000, **124**:1615-1624.
17. Ohri D and Khoshoo T: **Genome size in gymnosperms.** *Plant Syst Evol* 1986, **153**:119-132.
18. Murray B: **Nuclear DNA amounts in gymnosperms.** *Ann Bot* 1998, **Suppl A**:3-15.
19. The Arabidopsis Genome Initiative: **Analysis of the genome sequence of the flowering plant *Arabidopsis thaliana*.** *Nature* 2000, **408**:796-815.
20. Mayer K and Mewes HW: **How can we deliver the large plant genomes? Strategies and perspectives.** *Curr Opin Plant Biol* 2002, **5**:173-177.
21. Daly DC, Cameron KM and Stevenson DW: **Plant systematics in the age of genomics.** *Plant Physiol* 2001, **127**:1328-1333.
22. Martienssen R and McCombie WR: **The first plant genome.** *Cell* 2001, **105**:571-574.
23. Goff SA, Ricke D, Lan TH, Presting G, Wang R, Dunn M, Glazebrook J, Sessions A, Oeller P, Varma H et al.: **A draft sequence of the rice genome (*Oryza sativa* L. ssp. *japonica*).** *Science* 2002, **296**:92-100.
24. Yu J, Hu S, Wang J, Wong GK, Li S, Liu B, Deng Y, Dai L, Zhou Y, Zhang X et al.: **A draft sequence of the rice genome (*Oryza sativa* L. ssp. *indica*).** *Science* 2002, **296**:79-92.
25. Treutlein J and Wink M: **Molecular phylogeny of cycads inferred from *rbcl* sequences.** *Naturwissenschaften* 2002, **89**:221-225.
26. Stevenson D: **Morphology and systematics of the Cycadales.** *Mem NY Bot Garden* 1990, **57**:8-55.
27. Crane PR: **Phylogenetic analysis of seed plants and the origin of angiosperms.** *Annls Missouri Bot Gardens* 1985, **72**:716-793.
28. Duncan MW, Kopin JJ, Crowley JS, Jones SM and Markey SP: **Quantification of the putative neurotoxin 2-amino-3-(methylamino)propanoic acid (BMAA) in Cycadales: analysis of the seeds of some members of the family Cycadaceae.** *J Anal Toxicol* 1989, **13**:suppl A-G.
29. Brenner ED, Stevenson DW and Twigg RW: **Cycads: evolutionary innovations and the role of plant-derived neurotoxins.** *Trends Plant Sci* 2003, **8**:446-452.
30. Stevenson DW: **Observations on ptyxis, phenology, and trichomes in the Cycadales and their systematic implications.** *Am J Bot* 1981, **68**:1104-1114.
31. **Sputnik *Cycas rumphii*** [<http://mips.gsf.de/proj/sputnik/cycad/>]
32. Kirst M, Johnson AF, Baucom C, Ulrich E, Hubbard K, Staggs R, Paule C, Retzel E, Whetten R and Sederoff R: **Apparent homology of expressed genes from wood-forming tissues of loblolly pine (*Pinus taeda* L.) with *Arabidopsis thaliana*.** *Proc Natl Acad Sci USA* 2003, **100**:7383-7388.
33. Whetten R, Sun YH, Zhang Y and Sederoff R: **Functional genomics and cell wall biosynthesis in loblolly pine.** *Plant Mol Biol* 2001, **47**:275-291.
34. **Index of full-length sequences** [http://genomics.nybg.org/sequences/full_length/]
35. Suarez-Lopez P, Wheatley K, Robson F, Onouchi H, Valverde F and Coupland G: **CONSTANS mediates between the circadian clock and the control of flowering in *Arabidopsis*.** *Nature* 2001, **410**:1116-1120.
36. Putterill J, Robson F, Lee K, Simon R and Coupland G: **The CONSTANS gene of *Arabidopsis* promotes flowering and encodes a protein showing similarities to zinc finger transcription factors.** *Cell* 1995, **80**:847-857.
37. Chan RL, Gago GM, Palena CM and Gonzalez DH: **Homeoboxes in plant development.** *Biochim Biophys Acta* 1998, **1442**:1-19.
38. Eshed Y, Baum SF and Bowman JL: **Distinct mechanisms promote polarity establishment in carpels of *Arabidopsis*.** *Cell* 1999, **99**:199-209.
39. Eshed Y, Baum SF, Perea JV and Bowman JL: **Establishment of polarity in lateral organs of plants.** *Curr Biol* 2001, **11**:1251-1260.
40. Bohmert K, Camus I, Bellini C, Bouchez D, Caboche M and Benning C: **AGO1 defines a novel locus of *Arabidopsis* controlling leaf development.** *EMBO J* 1998, **17**:170-180.
41. Schwecheimer C and Deng XW: **COP9 signalosome revisited: a novel mediator of protein degradation.** *Trends Cell Biol* 2001, **11**:420-426.
42. Chamovitz DA and Glickman M: **The COP9 signalosome.** *Curr Biol* 2002, **12**:R232.
43. Chory J and Wu D: **Weaving the complex web of signal transduction.** *Plant Physiol* 2001, **125**:77-80.
44. Chiu JC, Brenner ED, DeSalle R, Nitabach MN, Holmes TC and Coruzzi GM: **Phylogenetic and expression analysis of the glutamate-receptor-like gene family in *Arabidopsis thaliana*.** *Mol Biol Evol* 2002, **19**:1066-1082.
45. Warrilow AG and Hawkesford MJ: **Cysteine synthase (O-acetylserine (thiol) lyase) substrate specificities classify the mitochondrial isoform as a cyanoalanine synthase.** *J Exp Bot* 2000, **51**:985-993.
46. Warrilow AG and Hawkesford MJ: **Modulation of cyanoalanine synthase and O-acetylserine (thiol) lyases A and B activity by beta-substituted alanyl and anion inhibitors.** *J Exp Bot* 2002, **53**:439-445.
47. Foster AS and Gifford EM: *Comparative Morphology of Vascular Plants* 2nd edition. San Francisco: WH Freeman; 1974.
48. Khabazian I, Bains JS, Williams DE, Cheung J, Wilson JM, Pasqualotto BA, Pelech SL, Andersen RJ, Wang YT, Liu L et al.: **Isolation of various forms of sterol beta-D-glucoside from the seed of *Cycas circinalis*: neurotoxicity and implications for ALS-parkinsonism dementia complex.** *J Neurochem* 2002, **82**:516-528.
49. Villanueva JM, Broadhvest J, Hauser BA, Meister RJ, Schneitz K and Gasser CS: **INNER NO OUTER regulates abaxial-adaxial patterning in *Arabidopsis* ovules.** *Genes Dev* 1999, **13**:3160-3169.
50. Hellmann H and Estelle M: **Plant development: regulation by protein degradation.** *Science* 2002, **297**:793-797.
51. Wei N and Deng XW: **COP9: a new genetic locus involved in light-regulated development and gene expression in *Arabidopsis*.** *Plant Cell* 1992, **4**:1507-1518.
52. Bogdanovic M: **Chlorophyll formation in the dark.** *Physiol Plant* 1973, **29**:17-18.
53. Peer WV, Silverthorne J and Peters JL: **Developmental and light-regulated expression of individual members of the light-harvesting complex b gene family in *Pinus palustris*.** *Plant Physiol* 1996, **111**:627-634.
54. Lacombe B, Becker D, Hedrich R, DeSalle R, Hollmann M, Kwak JM, Schroeder JI, Le Novere N, Nam HG, Spalding EP et al.: **The identity of plant glutamate receptors.** *Science* 2001, **292**:1486-1487.
55. Ohlrogge J and Benning C: **Unravelling plant metabolism by EST analysis.** *Curr Opin Plant Biol* 2000, **3**:224-228.
56. **VecScreen** [<http://www.ncbi.nlm.nih.gov/VecScreen/UniVec.html>]
57. Rudd S, Mewes HW and Mayer KF: **Sputnik: a database platform for comparative plant genomics.** *Nucleic Acids Res* 2003, **31**:128-132.
58. Altschul SF, Gish W, Miller W, Myers EW and Lipman DJ: **Basic local alignment search tool.** *J Mol Biol* 1990, **215**:403-410.

59. Slater GSC: **Algorithms for the Analysis of ESTs**. *PhD thesis* University of Cambridge; 2000.
60. Ewing B and Green P: **Base-calling of automated sequencer traces using phred. II. Error probabilities**. *Genome Res* 1998, **8**:186-194.
61. Ewing B, Hillier L, Wendl MC and Green P: **Base-calling of automated sequencer traces using phred. I. Accuracy assessment**. *Genome Res* 1998, **8**:175-185.

comment

reviews

reports

deposited research

refereed research

interactions

information