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Update on peripheral mechanisms of pain: beyond prostaglandins and cytokines

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Abstract

The peripheral nociceptor is an important target of pain therapy because many pathological conditions such as inflammation excite and sensitize peripheral nociceptors. Numerous ion channels and receptors for inflammatory mediators were identified in nociceptors that are involved in neuronal excitation and sensitization, and new targets, beyond prostaglandins and cytokines, emerged for pain therapy. This review addresses mechanisms of nociception and focuses on molecules that are currently favored as new targets in drug development or that are already targeted by new compounds at the stage of clinical trials - namely the transient receptor potential V1 receptor, nerve growth factor, and voltage-gated sodium channels - or both.

Introduction

Pain research substantially amplifies our understanding of the nature and mechanisms of pain, and this area is still expanding. We have learned that different forms of pain can be distinguished according to their pathogenesis (for example, nociceptive versus neuropathic pain) (see the following section), and we are beginning to decipher the molecular mechanisms involved in different types of pain. Importantly, this research provided new targets for pain treatment, and different classes of compounds that are effective against pain in animal models and that are now being tested in humans in different phases of clinical trials have emerged. The two main challenges are the efficacy (are the new drugs better than the old ones?) and the side effects (are they less dangerous than the old ones?). The need for better treatment of pain is evident because worldwide about 20% of adults suffer from chronic pain [1,2]. In these patients, the current pain therapy is either not applicable for different reasons (for example, side effects) or not sufficient [1]. In this review, the focus will be on molecular targets which are currently favored in drug development and for which respective compounds are at different stages of clinical trials.

The nature of pain

It is important to note that pain has different facets. In normal tissue, pain is elicited by high-energy stimuli

that potentially or actually damage the tissue (noxious stimuli). This 'physiological nociceptive pain' is a warning sensation and absolutely essential for survival because it triggers adequate avoidance reactions. Pain treatment must not impair this type of pain. On the other hand, pain is an important symptom of disease, and this pain has to be treated because it impairs normal life [1]. The pain elicited by inflammation or injury of an organ is called 'pathophysiological nociceptive pain'. This pain is typically elicited by low-energy stimuli that are normally innocuous and not painful. It appears as allodynia (occurrence of pain upon an innocuous stimulus) or hyperalgesia (more pain during the application of noxious stimuli) or both, and resting pain (in the absence of any intentional stimulation) may also occur. Pathophysiological nociceptive pain results from the sensitization of the pain system (see the following section), and the interference with the mechanisms of sensitization provides the opportunity of treatment without eliminating the physiological nociceptive pain [2,3]. Initially, this pain is beneficial because it indicates the presence of disease and triggers measures for the support of healing (cancer is so dangerous because it does not cause pain in its early stage). However, chronic pain, in particular, creates severe suffering [1].

Whereas 'nociceptive pain' results from noxious stimulation or inflammation/injury of tissue, 'neuropathic pain' is evoked by damage to the neurons of the nociceptive system themselves. Causes include nerve damage, metabolic diseases (for example, diabetes mellitus), and herpes

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zoster. Neuropathic pain does not primarily signal noxious tissue stimulation, often feels abnormal (burning or electrical character), and can be persistent or occur in short episodes (for example, trigeminal neuralgia). It may be combined with hyperalgesia and allodynia or with sensory loss [2,3]. This pain requires treatment that is, however, often not sufficient. Its mechanisms differ in part from those of nociceptive pain.

Neurophysiology of peripheral nociceptors in health and disease

Noxious stimuli are detected by nociceptors that are present in almost all organs. These specialized sensory neurons have Aδ- and C-fibers in the peripheral nerve and sensory non-corpuscular 'free nerve endings' in the innervated organs. Most nociceptors are polymodal, responding to noxious mechanical stimuli (painful pressure, squeezing the tissue), noxious thermal stimuli (heat or cold), and noxious chemical stimuli. They are equipped with sensor molecules in the sensory endings which transduce these mechanical, thermal, and chemical stimuli into a depolarizing sensor potential [2-5] (Figure 1). When this depolarization is sufficiently large, it opens voltage-gated Na^+ channels and triggers the generation of action potentials that are conducted to the dorsal horn of the spinal cord or the brainstem (Figure 1). From their sensory endings, the peptidergic nociceptors can release the neuropeptides substance P and calcitonin gene-related peptide (CGRP), which induce vasodilatation, plasma extravasation, and other effects, thus producing a 'neurogenic inflammation' [6].

Inflammation sensitizes polymodal nociceptors. Their excitation threshold drops such that even light, normally innocuous stimuli activate the fibers, and noxious stimuli evoke stronger responses than in the non-sensitized state. In addition, inflammation recruits so-called silent nociceptors for activation [2,3,7]. These C-fibers are inexcitable by noxious mechanical or thermal stimuli in normal tissue but inflammation sensitizes them to become responsive to stimuli. This 'peripheral sensitization' induces a hyper-excitability of nociceptive neurons in the central nervous system ('central sensitization'), and the two together generate the features of pathophysiological nociceptive pain, namely allodynia and hyperalgesia [2,8,9]. Peripheral sensitization is induced by inflammatory mediators that change the response properties of ion channels through the activation of second messenger systems (Figure 1).

The pathophysiology of neuropathic pain is different. While in healthy sensory nerve fibers action potentials are generated in the sensory endings upon stimulation of the receptive field, damaged nerve fibers (including non-nociceptive Aβ-fibers [10]) often show pathological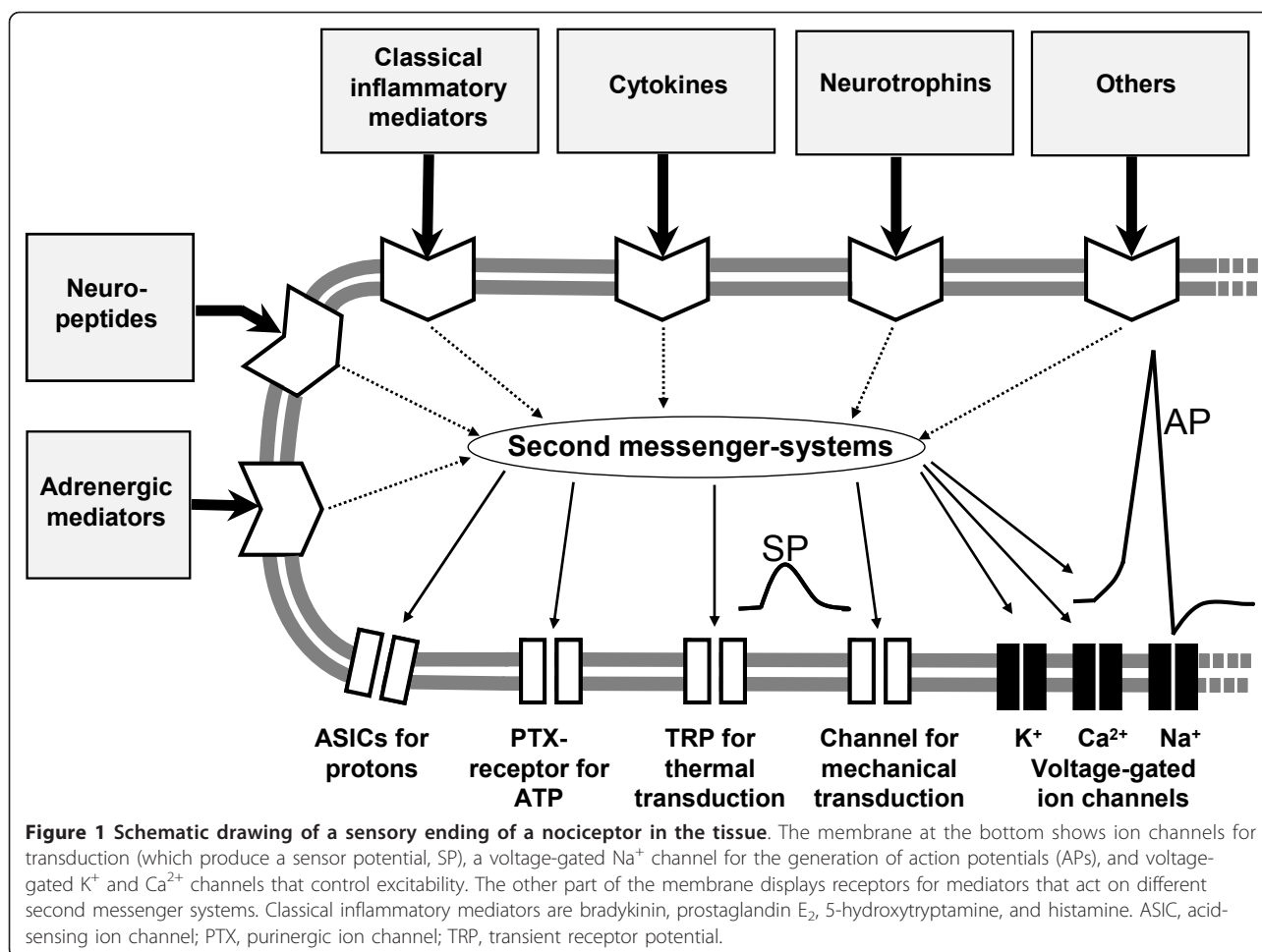 ectopic discharges that are generated at the site of nerve injury or in the cell body in dorsal root ganglia (DRGs) [3,11]. Neuropathic pain may also be generated by intact nerve fibers in the vicinity of injured nerve fibers [12]. Ectopic discharges are thought to be produced by changes in the expression of ion channels [13], pathological activation of axons by inflammatory mediators [14], and pathological activation of injured nerve fibers by the sympathetic nervous system [15].

However, the question of whether there is always a strict distinction between these types of pain under clinical conditions arises. For example, osteoarthritic pain is considered mainly a pathophysiological nociceptive pain because the joints may show severe damage and inflammation [16]. However, changes that are typical for neuropathic conditions may occur in the nervous system [17]. Possibly, pain types become less discernible with time.

Another emerging aspect of pain is that significant differences between the nociceptive innervation and processes in different tissues exist. First, the skin is innervated by two large populations of nociceptors, namely peptidergic (containing substance P and CGRP) and non-peptidergic IB4 (isolectin B4)-positive nociceptive aerents [2,5], whereas the vast majority of aerents in the musculoskeletal system are peptidergic [18,19]. Second, nociceptors in skin and musculoskeletal system have relatively high mechanical thresholds that clearly distinguish them from non-nociceptive low-threshold sensory fibers [20,21], whereas most putative nociceptors in the viscera have their excitation threshold in the innocuous range [22]. Third, during inflammation, nociceptors in the muscle and joint are strongly sensitized to mechanical stimuli whereas cutaneous nociceptors are rather sensitized to thermal stimuli [20,21,23]. These differences, often ignored, may have a significant impact on drug development. Although there is an overlap of nociceptive mechanisms in different tissues, the awareness of tissue-specific nociceptive mechanisms may become more important in the future [2]. Notably, most chronic pain occurs during diseases of the musculoskeletal system whereas skin diseases are rarely painful [1].

Principles of molecular mechanisms of nociception and sensitization

The identification of these mechanisms is based on different experimental approaches. Electrophysiological and behavioral experiments in the intact organism are used to study nociceptive processes *in situ*. However, the sensory free nerve endings themselves cannot be accessed *in situ* for recordings, because they are extremely small and embedded in the tissue. Fortunately, the more accessible somata of the nociceptors in the DRGs also express the relevant molecules. Therefore, patch clamp recordings are usually carried out in cultured DRG

Figure 1 Schematic drawing of a sensory ending of a nociceptor in the tissue. The membrane at the bottom shows ion channels for transduction (which produce a sensor potential, SP), a voltage-gated Na$^+$ channel for the generation of action potentials (APs), and voltage-gated K$^+$ and Ca^{2+} channels that control excitability. The other part of the membrane displays receptors for mediators that act on different second messenger systems. Classical inflammatory mediators are bradykinin, prostaglandin E$_2$, 5-hydroxytryptamine, and histamine. ASIC, acid-sensing ion channel; PTX, purinergic ion channel; TRP, transient receptor potential.

neurons, although subtle differences between the cell body and the endings in the tissue might exist [23]. Pain research aims to achieve convergent information from behavioral experiments *in vivo* (for example, in wild-type and knockout animals), invasive experiments *in situ*, and experiments in cultured neurons.

As mentioned, the excitation of nociceptor endings is produced by ion channels in the membrane which open upon the impact of physical stimuli to the membrane. The cation influx through these channels produces a local depolarization, the sensor potential. Once the ending is sufficiently depolarized by the influx of cations, voltage-gated Na$^+$ channels are opened and action potentials are triggered (Figure 1). K$^+$ and Ca^{2+} channels control excitability of the neurons. During the process of sensitization, the gating properties of ion channels are modified by inflammatory mediators that act on metabotropic membrane receptors and activate second messenger systems [24]. Inflammatory mediators may also evoke neuropathic ectopic discharges [25]. Depending on the mediator, the sensitization of nociceptors can be induced within minutes (for example, by phosphorylation of ion channels in the

membrane) or hours [23,26,27]. In the long term, the expression of receptors and ion channels may be regulated such that more molecules are available for stimulation [2]. These changes may be induced by inflammatory mediators such as cytokines [28,29] and nerve growth factor (NGF) [30]. The following paragraphs address ion channels and receptors that were and are a major focus of pain research and some of them are the focus of drug development.

The TRPV1 receptor

A lot of effort is put into the development of antagonists against the transient receptor potential V1 (TRPV1) receptor, and it is hoped that these antagonists become new analgesics. In fact, clinical trials are under way. Why is there so much interest in the TRPV1 receptor? In the last 10 to 15 years, the TRPV1 receptor was one of the hot topics in pain research and is now considered one of the key molecules in nociception [2,5]. The TRPV1 receptor is a ligand-gated ion channel. Upon opening, cations (in particular, Ca^{2+}) flow into the cell and depolarize it. Importantly, the TRPV1 receptor is expressed in nociceptors but not in other peripheral

neurons [30]; that is, antagonists may selectively act on nociceptive neurons and not on other ones. Stimuli that open the TRPV1 receptor are manifold. First, it is opened by temperatures of higher than 43°C, which are felt as painful heat by humans. TRPV1-deficient mice show attenuated responses to heat stimuli [5] but other molecules seem to contribute to noxious heat sensations as well [23]. Second, TRPV1 is opened by chemicals (such as capsaicin and ethanol applied to a wound) that elicit burning pain [30,31]. However, the main interest for analgesic therapy comes from the findings that the channel characteristics are modified by metabolites that occur in inflamed tissue. In fact, the TRPV1 is opened by low pH (<5.9) occurring in inflamed tissue and is activated by arachidonic acid metabolites produced by lipoxygenases such as 12-hydroperoxyeicosaenoic acid (12-HPETE) and by endocannabinoids such as anandamide and NADA (N-arachidonyl-dopamine) [30]. Further-more, TRPV1 is indirectly sensitized, via second messengers, by the inflammatory mediators bradykinin, prosta-glandin E_2, extracellular ATP, glutamate, proteases, and NGF [25,27,30,31]. Cellularly, sensitization may be produced by the increase of TRPV1 expression level in the membrane, the phosphorylation by protein kinases, and the release of inhibition of TRPV1 by phos-phatidyl-inositol-4,5-biphosphate [25]. The important consequence of this sensitization is that the threshold for opening by temperature stimuli drops such that normal body temperature may be sufficient to activate nociceptors and to elicit action potentials. This form of hyperalgesia is tested by thermal stimuli; in fact, TRPV1-deficient mice do not show the typical inflammation-evoked thermal hyperalgesia in acute carrageenan- or complete Freund's adjuvant (CFA)-induced inflammation [30,31].

However, there are some caveats that may limit the success of TRPV1 antagonists. First, in humans, inflammatory pain appears mainly as mechanical hyperalgesia (that is, the sensitization of the nociceptive system toward mechanical stimulation) [2,3]. The initial data showed that TRPV1-deficient mice exhibit normal responses to noxious mechanical stimuli and that inflammation-evoked mechanical hypersensitivity is not reduced [31]. Thus, TRPV1 was classified as a key molecule for thermal hyperalgesia but not for mechanical hyperalgesia [25]. Only recently, an involvement of TRPV1 in mechanical hyperalgesia was reported in adjuvant-induced chronic arthritis [32]. It is still unclear, therefore, whether TRPV1 receptor antagonists will be efficient against significant mechanical hyperalgesia (for example, during osteoarthritis). Second, the role of TRPV1 receptors in neuropathic pain is not well understood. While it was proposed that TRPV1 receptors are important in the generation of chemical and thermal hyperalgesia in neuropathy, even a protective role of TRPV1 against the development of mechanical hyperalgesia was observed [31]. Thus, the presence of a neuropathic pain component could spoil the success of TRPV1 receptor antagonists.

Other TRPV receptors

Sensory neurons also express other TRP receptors that can be colocalized with TRPV1. The functional significance of other TRPV receptors is much more uncertain or even confusing. Therefore, they are currently not considered targets for antinociceptive therapy, although they might be involved in aspects of nociception [25,31]. The TRPV2 receptor is activated by high temperature (threshold of about 52°C) and may be the heat sensor in high-threshold-temperature Aδ nociceptors. TRPV2 and TRPV1 are colocalized in a very small percentage of DRG neurons [31]. TRPV3 is activated by innocuous temperature stimuli in the range of 31°C to 39°C but shows increased responses to noxious thermal stimuli and may contribute to pain hypersensitivity during inflammation. It can be colocalized with the TRPV1 receptor [31]. TRPV4 is activated by innocuous warmth with a threshold of greater than 27°C. Some data suggest that TRPV4 may, in fact, play a role in the transduction of mechanical stimuli and in mechanical hyperalgesia following exposure to inflammatory mediators [5]. The transient receptor potential A1 (TRPA1) was initially considered the transduction molecule for noxious cold stimuli because its activation threshold is below 18°C [31] (this skin temperature may feel painful), but there is no consensus whether TRPA1 is really a noxious cold sensor [5,33]. The transient receptor potential M8 (TRPM8) receptor is activated in the range of 23°C to 28°C and is also activated by cooling compounds such as menthol, eucalyptol, and icilin [31]. It is assumed that this particular TRP receptor might be the receptor for the sensation 'cool' (not painful) in non-nociceptive nerve fibers; in fact, TRPM8 is rarely colocalized with the other TRP receptors.

The involvement of other TRP receptors in nociception is suggested by their sensitivity to irritant mediators or mediators involved in inflammation or by the fact that they are upregulated under inflammatory conditions or by both. TRPV2 is upregulated in DRG neurons after intra-plantar injection of CFA [31]. TRPV3 shows responses to camphor and other irritants, nitric oxide (NO), and arachidonic acid and other fatty acids [31]. TRPV4 is activated by phorbolester, low pH, citrate, endocannabinoids, arachidonic acid metabolites, and NO [31]. TRPA1 is activated by bradykinin and by pungent ingredients of mustard oil, garlic, and others, all of which induce an acute painful burning or pricking sensation [31].

It should be noted that TRP receptors are not confined to peripheral sensory neurons. They are also detected in the central nervous system and even in non-neuronal tissue. For example, TRPV4 is expressed in cochlear hair cells, and both TRPV3 and TRPV4 are expressed in keratinocytes [30,31]. Whether this restricts or prevents the usage of TRP antagonists as analgesic remains to be seen.

Acid-sensing ion channels

Acid-sensing ion channels (ASICs) are members of the ENa/degenerin family of highly selective Na^+ channels. They are expressed in numerous DRG neurons [5]. ASICs are activated by low extracellular pH and thus are important sensors of tissue acidosis, which often occurs during inflammation. Several isoforms of ASICs were reported [25]. Perhaps ASICs are most important in the skeletal muscle and heart, in which impaired circulation causes immediate pain [5,34].

ATP and purinergic ion channels

In some tissues, extracellular ATP is thought to act as a pain mediator. ATP opens purinergic receptors (P2X2 and P2X3) which are ligand-gated Ca^{2+} channels. ATP can be released from damaged cells or from keratinocytes of the inflamed skin [25]. Like protons, ATP may be particularly important in contracting muscles [34]. The Ca^{2+} influx depolarizes cells and causes secondary Ca^{2+} influx through voltage-gated Ca^{2+} channels. P2X activation was implicated in the development of inflammatory hyperalgesia [25].

Voltage-gated Na^+ channels and other voltage-gated ion channels

Voltage-gated Na^+ channels are essential for the generation and conductance of action potentials. The analgesic effect of local anesthetics is due to their ability to block voltage-gated Na^+ channels and thus to prevent the conduction of action potentials. Because local anesthetics block Na^+ channels in all nerve fibers (including motoneurons, touch receptors, and thermoreceptors), their long-term use for pain treatment is not feasible. However, recent research revealed that different types of Na^+ channels exist and that some of them are particularly expressed in nociceptive neurons. Therefore, a lot of effort is put into research on Na^+ channels and their putative utilization for pain treatment.

Nociceptive neurons express mainly $Na_v1.7$, $Na_v1.8$, and $Na_v1.9$, whereas large-sized non-nociceptive DRG neurons express mainly $Na_v1.1$, $Na_v1.6$, and $Na_v1.7$ and some $Na_v1.8$ [2,35]. $Na_v1.1$, $Na_v1.6$, and $Na_v1.7$ can be blocked by tetrodotoxin (TTX) and are called TTX-sensitive (TTX-S), whereas $Na_v1.8$ and $Na_v1.9$ are TTX-resistant (TTX-R). Na^+ influx into the neuron through TTX-S Na^+ channels activates and inactivates very quickly, and Na^+ influx through TTX-R Na^+ channels activates and inactivates more slowly. Most likely, conduction of action potentials in the axons of primary afferents rests mainly on TTX-S Na^+ channels, whereas TTX-R Na^+ currents are important in the action potential generation in the sensory endings and the cell bodies [36]. The channels are up- or downregulated by second messenger pathways involving protein kinase A, protein kinase C, sphingomyelinase, calmodulin, and p38 mitogen-activated protein kinase [13].

The resting potential of DRG neurons is about -60 mV. $Na_v1.7$ opens after small depolarizations (at -50 to -40 mV), and this initial Na^+ influx brings the neuron closer to the membrane potential for elicitation of an action potential [35]. $Na_v1.8$, which is expressed only in sensory neurons and largely restricted to nociceptive neurons, opens at -30 to -20 mV - that is, when the cell has been pre-depolarized (for example, by $Na_v1.7$) - and provides about 80% of the inward current of the upstroke of the action potentials in DRG neurons. In particular, this Na^+ channel is targeted primarily to terminals and the cell body, suggesting a role in action potential initiation at the sensory terminal of nociceptors [13]. It also mediates repetitive action potentials during persistent membrane depolarization (for example, in the presence of inflammatory mediators) [35]. While $Na_v1.7$ and $Na_v1.8$ are directly involved in the generation of the action potential, $Na_v1.9$ influences the threshold for action potentials. The channel opens around -60 mV and conducts persistent Na^+ currents at subthreshold voltages for action potential firing and thus regulates the distance between membrane potential and threshold; it does not contribute to the upstroke of the action potential [35].

The critical importance of these Na^+ channels in nociception is obvious from numerous findings. A gain-of-function mutation and a loss-of-function mutation of $Na_v1.7$ channel in humans directly demonstrated the involvement of $Na_v1.7$ current in pain perception [37]. During inflammation, $Na_v1.7$ channel mRNA and protein are increased, and in mice with $Na_v1.7$ deletion, inflammatory hypersensitivity is either absent or dramatically attenuated [13]. $Na_v1.8$ current is increased by proinflammatory mediators such as adenosine, endothelin, NGF, prostaglandin E_2, serotonin, and tumor necrosis factor-alpha (TNF-α) [13]. Antisense knockdown of $Na_v1.8$ attenuates the development and maintenance of inflammatory hyperalgesia and can also prevent and reverse hypersensitivity observed following traumatic nerve injury [13]. $Na_v1.9$ is also potentiated by inflammatory mediators [25].

The excitability of neurons may also be controlled by K^+ channels (for example, of the potassium voltage-gated channel subfamily KQT [KCNQ] family) and Ca^{2+}

channels. Excitability is increased when voltage-gated K^+ channels are inhibited (this evokes sustained depolarization of neurons) or when Ca^{2+} flows into the neuron through voltage-gated T-type channels [25].

Mediators activating, sensitizing, and altering nociceptive neurons

Figure 1 schematically shows receptors for classes of inflammatory mediators without addressing all mediators and receptor subtypes in detail (this is beyond the scope of this review). It should be noted that not all of the receptors are located in all neurons. Instead, only subpopulations of neurons respond to certain mediators, but the nociceptive system as a whole is responsive to all of the mediators indicated.

'Classical inflammatory mediators' such as bradykinin and prostaglandins [5,27,38] activate or sensitize neurons (or both) within minutes. For example, prostaglandin E_2 acts on G protein-coupled prostaglandin E receptors that increase cAMP. This activates protein kinase A, which finally leads to a phosphorylation of TRPV1 receptors and voltage-gated Na^+ currents. The importance of this sensitizing mechanism is underscored by the ability of prostaglandin synthesis inhibitors to reduce sensitization. More recent research showed that cytokines can induce long-lasting effects on the excitability and also produce prolonged effects such as regulation of receptor expression [28,29]. In this context, it may be important that, in the acute phase of inflammation, macrophages invade the DRGs of the segments that innervate the inflamed organ; that is, macrophages may directly influence the cell bodies [39]. The blockade of TNF-α signaling was found to reduce inflammatory hyperalgesia, at least in part by a neuronal target [28,39,40].

A highly favored target for new drugs is NGF, and recent clinical studies using an antibody against NGF entered phase III [41] (with a set back, however, because the US Food and Drug Administration put a halt to some studies). Why is NGF a hot target? First, NGF is an essential growth factor for the development of normal nociceptors. In the adult, a large proportion of nociceptors remain dependent on the trophic effect of NGF. These neurons express tyrosine receptor kinase A (TrkA) receptors (specific receptor for NGF), and NGF is required for their structural and functional integrity [42]. Neutralization of NGF may thus disturb the nociceptive function of these neurons. Second, several cell types produce substantial amounts of NGF at inflammatory sites, and NGF can directly affect neurons. NGF enhances currents through TRPV1 channels and reduces the threshold of thermal excitation [30]. *In vivo*, the application of NGF generates hyperalgesia, and pretreatment with antibodies to NGF or a fusion protein attached to a modified TrkA receptor attenuates inflammation-induced hypersensitivity. Third, long-term exposure to NGF increases the expression of TRPV1, bradykinin receptors, P2X receptors, Na^+ channels, and the synthesis of putative nociceptive transmitters such as substance P and CGRP [30]. Fourth, NGF stimulates inflammatory cells to release inflammatory compounds [30]. Thus, NGF is a key molecule for nociceptor biology, and its neutralization proved to be highly analgesic in humans [41]. It remains to be seen whether NGF neutralization will become a therapeutical option or whether NGF neutralization may impair the function of nociceptors too much.

Mediators with inhibitory actions

The drugs addressed so far reduce excitation/sensitization by neutralizing pronociceptive mediators and by interfering with ion channels of nociception. The multitude of mediators and ion channels involved in the generation of pain may limit the success of all efforts to treat pain by interfering with single molecules. Whether neutralization of NGF as a mediator with 'integrative functions' is feasible will be shown. Another principle of pain treatment is to employ drugs that stimulate endogenous inhibitory mechanisms. In fact, sensory neurons exhibit receptors that mediate inhibition of neuronal activity. Recently, peripheral opioid receptors were shown to produce profound antinociception experimentally as well as clinically [30]. Further receptors with inhibitory actions are receptors for somatostatin [43] and cannabinoids [44]. Experimentally, the use of specific agonists at these receptors produces antinociception [44-46]. However, the cannabinoid anandamid also activates TRPV1 receptors that may counteract antinociception [30,45].

Conclusions

Although peripheral mechanisms of nociception significantly contribute to the generation of pain, the contribution of spinal, as well as supraspinal, thalamocortical mechanisms to pain generation is essential [47,48]. As mentioned, peripheral sensitization often causes central sensitization [8,9]. This is even induced by a single intradermal injection of capsaicin, which activates TRPV1 receptors [49], and is quite prominent and long-lasting during persistent inflammation in joints [3,21]. Thus, both peripheral and central nociceptive mechanisms are options for the improvement of pain therapy.

Note

This article is part of the series Evolving understanding of the biology of pain and its application to patient care, edited by Daniel Clauw and Anthony Jones. Other articles in this series can be found at http://arthritis-research.com/series/pain

Abbreviations

ASIC: acid-sensing ion channel; CFA: complete Freund's adjuvant; CGRP: calcitonin gene-related peptide; DRG: dorsal root ganglion; Na_vx.x channel: voltage-gated sodium channel; NGF: nerve growth factor; NO: nitric oxide; P2X: purinergic receptor; TNF-α: tumor necrosis factor-alpha; TrkA: tyrosine receptor kinase A; TRP: transient receptor potential; TRPA1: transient receptor potential A1; TRPM8: transient receptor potential M8; TRPV1: transient receptor potential V1; TRPV2: transient receptor potential V2; TRPV3: transient receptor potential V3; TRPV4: transient receptor potential V4; TTX: tetrodotoxin; TTX-R: tetrodotoxin-resistant; TTX-S: tetrodotoxin-sensitive.

Competing interests

The authors declare that they have no competing interests.

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References

1. Breivik H, Beverly C, Ventafridda V, Cohen R, Gallacher D: **Survey of chronic pain in Europe: prevalence, impact on daily life, and treatment.** *Eur J Pain* 2006, **10**:287-333.
2. Gold MS, Gebhart GF: **Nociceptor sensitization in pain pathogenesis.** *Nat Med* 2010, **16**:1248-1257.
3. Schaible H-G, Richter F: **Pathophysiology of pain.** *Langenbecks Arch Surg* 2004, **389**:237-243.
4. Belmonte C, Cervero E, (Eds): *Neurobiology of Nociceptors* Oxford: Oxford University Press; 1996.
5. Basbaum AI, Bautista DM, Scherrer G, Julius D: **Cellular and molecular mechanisms of pain.** *Cell* 2009, **139**:267-284.
6. Schaible H-G, Del Rosso A, Matucci-Cerinic M: **Neurogenic aspects of inflammation.** *Rheum Dis Clin North Am* 2005, **31**:77-101.
7. Schmidt R, Schmelz M, Forster C, Ringkamp M, Torebjork E, Handwerker H: **Novel classes of responsive and unresponsive C nociceptors in human skin.** *J Neurosci* 1995, **15**:333-341.
8. Cervero F: **Spinal cord hyperexcitablity and its role in pain and hyperalgesia.** *Exp Brain Res* 2009, **196**:129-137.
9. Sandkuhler J: **Models and mechanisms of hyperalgesia and allodynia.** *Physiol Rev* 2009, **89**:707-758.
10. Devor M: **Ectopic discharge in Aβ afferents as a source of neuropathic pain.** *Exp Brain Res* 2009, **196**:115-128.
11. Janig W, Grossmann L, Gorodetskaya N: **Mechano- and thermosensitivity of regenerating cutaneous afferent nerve fibers.** *Exp Brain Res* 2009, **196**:101-114.
12. Wu G, Ringkamp M, Hartke TV, Murinson BB, Campbell JN, Griffin JW, Meyer RA: **Early onset of spontaneous activity in uninjured C-fiber nociceptors after injury to neighbouring nerve fibers.** *J Neurosci* 2001, **21**:RC140, 1-5.
13. Gold MS: **Na$^+$ channel blockers for the treatment of pain; context is everything, almost.** *Exp Neurol* 2008, **210**:1-6.
14. Sommer C, Kress M: **Recent findings on how proinflammatory cytokines cause pain: peripheral mechanisms in inflammatory and neuropathic hyperalgesia.** *Neurosci Lett* 2004, **361**:184-187.
15. Janig W, Levine JD, Michaelis M: **Interactions of sympathetic and primary afferent neurons following nerve injury and tissue trauma.** In *The Polymodal Receptor: A Gateway to Pathological Pain. Progress in Brain Research. Volume 113.* Edited by: Kumazawa T, Kruger L, Mizumura K. Amsterdam: Elsevier Science; 1996:161-184.
16. Bondeson J, Blom AB, Wainwright S: **The role of synovial macrophages and macrophage-produced mediators in driving inflammatory and destructive responses in osteoarthritis.** *Arthritis Rheum* 2010, **62**:647-657.
17. Im H-J, Kim J-S, Li X, Kotwal N, Sumner DR, van Wijnen AJ, Davis FJ, Yan D, Levine B, Henry JL, Desevre J, Krin JS: **Alteration of sensory neurons and spinal response to an experimental osteoarthritis pain model.** *Arthritis Rheum* 2010, **62**:2995-3005.
18. Nakajima T, Ohtori S, Inoue G, Koshi T, Yamamoto J, Takahashi K, Harada Y: **The characteristics of dorsal-root-ganglia and sensory innervation of the hip in rats.** *J Bone Joint Surg Br* 2008, **90**:254-257.
19. Jimenez-Andrade JM, Mantyh WG, Bloom AP, Xu H, Ferng AS, Dussor G, Vanderah TW, Mantyh PW: **A phenotypically restricted set of primary afferent nerve fibres innervate the bone versus skin: therapeutic opportunity for treating skeletal pain.** *Bone* 2010, **46**:306-313.
20. McDougall J: **Arthritis and pain: neurogenic origin of joint pain.** *Arthritis Res Ther* 2006, **8**:220-229.
21. Schaible H-G, Richter F, Ebersberger A, Boettger MK, Vanegas H, Natura G, Vazquez E, Segond von Banchet G: **Joint pain.** *Exp Brain Res* 2009, **196**:153-162.
22. Bielefeldt K, Gebhart GF: **Visceral pain: basic mechanisms.** In *Wall and Melzack's Textbook of Pain.* Edited by: McMahon SB, Koltzenburg M. Edinburgh: Elsevier Churchill Livingstone; 2006:721-736.
23. St Pierre M, Reeh PW, Zimmermann K: **Differential role of TRPV channel block on polymodal activation of rat cutaneous nociceptors in vitro.** *Exp Brain Res* 2009, **196**:31-44.
24. Hucho T, Levine JD: **Signaling pathways in sensitization: toward a nociceptor cell biology.** *Neuron* 2007, **55**:365-376.
25. Linley JE, Rose K, Ooi L, Gamper N: **Understanding inflammatory pain: ion channels contributing to acute and chronic nociception.** *Pflügers Arch* 2010, **459**:657-669.
26. McCleskey EW, Gold MS: **Ion channels of nociception.** *Annu Rev Physiol* 1999, **61**:835-856.
27. Mizumura K, Sugiura T, Katanosaka K, Banik RK, Kozaki Y: **Excitation and sensitization of nociceptors by bradykinin: what do we know?** *Exp Brain Res* 2009, **196**:53-65.
28. Schaible H-G, Segond von Banchet G, Boettger MK, Bräuer R, Gajda M, Richter F, Hensellek S, Brenn D, Natura G: **The role of proinflammatory cytokines in the generation and maintenance of joint pain.** *Ann N Y Acad Sci* 2010, **1193**:60-69.
29. Üceyler N, Schäfers M, Sommer C: **Mode of action of cytokines on nociceptive neurons.** *Exp Brain Res* 2009, **196**:67-78.
30. Stein C, Clark JD, Oh U, Vasko MR, Wilcox GL, Overland AC, Vanderah TW, Spencer RH: **Peripheral mechanisms of pain and analgesia.** *Brain Res Rev* 2009, **60**:90-113.
31. Levine JD, Alessandri-Haber N: **TRP channels: targets for the relief of pain.** *Biochim Biophys Acta* 2007, **1772**:989-1003.
32. Szabo A, Helyes z, Sandor K, Bite A, Pinter E, Nemeth J, Banvolgyi A, Bolcskei K, Elekes K, Szolcsanyi J: **Role of transient receptor potential vanilloid 1 receptors in adjuvant-induced chronic arthritis: in vivo study using genedeficient mice.** *J Pharmacol Exp Ther* 2005, **314**:111-119.
33. Belmonte C, Brock JA, Viana F: **Converting cold into pain.** *Exp Brain Res* 2009, **196**:13-30.
34. Mense S: **Algesic agents exciting muscle nociceptors.** *Exp Brain Res* 2009, **196**:89-100.
35. Rush AM, Cummins TR, Waxman SG: **Multiple sodium channels and their roles in electrogenesis within dorsal root ganglion neurons.** *J Physiol* 2007, **579**:1-14.
36. Pinto V, Derkach VA, Safronov BV: **Role of TTX-sensitive and TTX-resistant sodium channels in Aδ and C-fiber conduction and synaptic transmission.** *J Neurophysiol* 2008, **99**:617-628.
37. Wada A, Wanke E, Gullo F, Schiavon E: **Voltage-dependent Na$_v$1.7 sodium channels: multiple roles in adrenal chromaffin cells and peripheral nervous system.** *Acta Physiol (Oxf)* 2008, **192**:221-231.
38. McMahon SB, Bennet DLH, Bevan S: **Inflammatory mediators and modulators.** In *Wall and Melzack's Textbook of Pain.* Edited by: McMahon SB, Koltzenburg M. Edinburgh: Elsevier Churchill Livingstone; 2006:49-72.
39. Segond von Banchet G, Boettger MK, Fischer N, Bräuer R, Schaible H-G: **Experimental arthritis causes tumor necrosis factor-α dependent infiltration of macrophages in rat dorsal root ganglia which correlates with pain-related behavior.** *Pain* 2009, **145**:151-159.
40. Inglis JJ, Notley CA, Essex D, Wilson AW, Feldmann M, Anand P, Williams R: **Collagen-induced arthritis as a model of hyperalgesia: functional and cellular analysis of the analgesic actions of tumor necrosis factor blockade.** *Arthritis Rheum* 2007, **56**:4015-4023.
41. Lane NE, Schnitzer TJ, Birbara CA, Mokhtarani M, Shelton DL, Smith MD, Brown MT: **Tanezumab for the treatment of pain from osteoarthritis of the knee.** *N Engl J Med* 2010, **363**:1521-1531.
42. Bennett D: **NGF, sensitization of nociceptors.** In *Encyclopedia of Pain. Volume 2.* Edited by: Schmidt RF, Willis WD. Berlin: Springer; 2007:1338-1342.
43. Bär K-J, Schurigt U, Scholze A, Segond von Banchet G, Stopfel N, Bräuer R, Halbhuber K-J, Schaible H-G: **The expression and localisation of somatostatin receptors in dorsal root ganglion neurons of normal and monoarthritic rats.** *Neuroscience* 2004, **127**:197-206.

44. Agarwal N, Pacher P, Tegeder I, Amaya F, Constantin CE, Brenner GJ, Rubino T, Michalski CW, Marsicano G, Monory K, Mackie K, Marian C, Batkai S, Parolaro D, Fischer MJ, Reeh P, Kunos G, Kress M, Lutz B, Woolf CJ, Kuner R: **Cannabinoids mediate analgesia largely via peripheral type 1 cannabinoid receptors in nociceptors.** *Nat Neurosci* 2007, **10**:870-879.

45. Kress M, Kuner R: **Mode of action of cannabinoids on nociceptive nerve endings.** *Exp Brain Res* 2009, **196**:79-88.

46. Helyes Z, Szabó A, Németh J, Jakab B, Pintér E, Bánvölgyi Á, Kereskai L, Kéri G, Szolcsányi J: **Anti-inflammatory and analgesic effects of somatostatin released from capsaicin-sensitive sensory nerve terminals in a Freund's adjuvant-induced chronic arthritis model in the rat.** *Arthritis Rheum* 2004, **50**:1677-1685.

47. Treede RD, Kenshalo DR, Gracely RH, Jones A: **The cortical representation of pain.** *Pain* 1999, **79**:105-111.

48. Vogt BA: **Pain and emotion. Interactions in subregions of the cingulated cortex.** *Nat Rev Neurosci* 2005, **6**:533-544.

49. Willis WD: **The role of TRPV1 receptors in pain evoked by noxious thermal and chemical stimuli.** *Exp Brain Res* 2009, **196**:5-11.