

REVIEW

Open Access



# Removing the stumbling block of exosome applications in clinical and translational medicine: expand production and improve accuracy

Li Han<sup>1,3†</sup>, Zhirong Zhao<sup>2†</sup>, Chuanshi He<sup>1,3</sup>, Jiami Li<sup>1</sup>, Xiangyu Li<sup>1</sup> and Man Lu<sup>1,3\*</sup>

## Abstract

Although the clinical application and transformation of exosomes are still in the exploration stage, the prospects are promising and have a profound impact on the future transformation medicine of exosomes. However, due to the limitation of production and poor targeting ability of exosomes, the extensive and rich biological functions of exosomes are restricted, and the potential of clinical transformation is limited. The current research is committed to solving the above problems and expanding the clinical application value, but it lacks an extensive, multi-angle, and comprehensive systematic summary and prospect. Therefore, we reviewed the current optimization strategies of exosomes in medical applications, including the exogenous treatment of parent cells and the improvement of extraction methods, and compared their advantages and disadvantages. Subsequently, the targeting ability was improved by carrying drugs and engineering the structure of exosomes to solve the problem of poor targeting ability in clinical transformation. In addition, we discussed other problems that may exist in the application of exosomes. Although the clinical application and transformation of exosomes are still in the exploratory stage, the prospects are promising and have a profound impact on drug delivery, clinical diagnosis and treatment, and regenerative medicine.

**Keywords** Exosome, Optimization, Extraction, Translational medicine, Targeted delivery

## Introduction

Extracellular vesicles (EVs) have received increasing attention as novel biomarkers, therapeutics, and drug delivery vehicles for diseases [1]. EVs can be classified according to their biosynthetic or release pathway, including exosomes with a diameter of 30–150 nm that originate from the endocytic pathway, microvesicles with a diameter of approximately 100–1000 nm that are directly released from the plasma membrane, and 50 nm–2 μm that are generated by apoptosis [2]. Among them, the powerful advantages and functions of exosomes are highlighted in medical research and clinical applications due to their containing complex nucleic acids, RNA, and proteins. A variety of cells can secrete exosomes under normal and pathological conditions.

<sup>†</sup>Li Han and Zhirong Zhao authors contributed equally to this study.

\*Correspondence:

Man Lu

luman@westc.edu.cn

<sup>1</sup> Ultrasound Medical Center, Sichuan Clinical Research Center for Cancer, Sichuan Cancer Hospital & Institute, Sichuan Cancer Center, Affiliated Cancer Hospital of University of Electronic Science and Technology of China, Chengdu 610041, Sichuan, China

<sup>2</sup> College of Medicine, Southwest Jiaotong University, Chengdu 610031, Sichuan, China

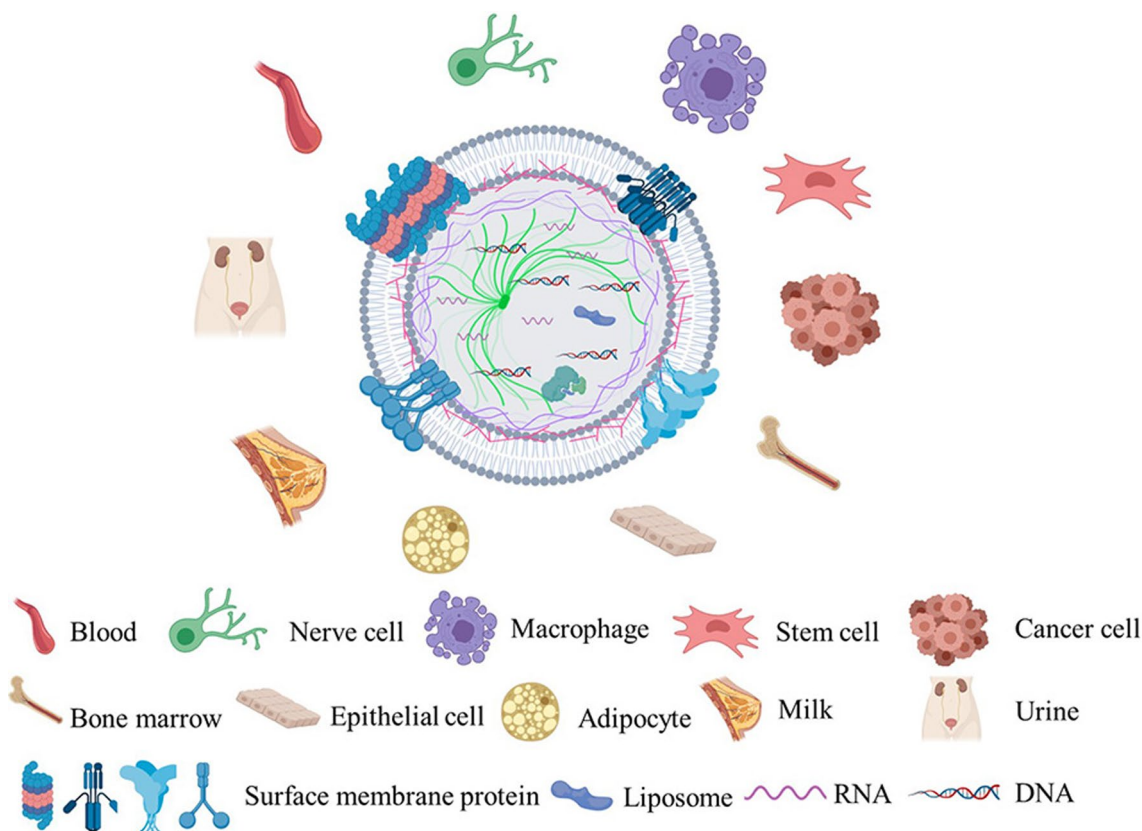
<sup>3</sup> The School of Medicine, University of Electronic Science and Technology of China, Sichuan 611731 Chengdu, China



Exosomes originate from the inward budding of the cell membrane to form an endosome. After undergoing the processes of forming a polycystic complex, directional assembly, and migration, the endosome fused with the cell membrane and excreted out of the cell by exocytosis [3, 4]. Exosomes have a topology similar to that of cells and contain proteins such as transmembrane protein (CD9, CD63, CD81), annexins, and heat shock proteins (HSP) [5–7]. And the exosomes were also enriched in cholesterol and sphingomyelin. Up to now, many studies have found that there are 41,860 kinds of protein, 2838 kinds of microRNA, and 3,408 kinds of mRNA in the exocrine body [8–10] (Fig. 1).

Exosomes not only mediate signal transmission between cells by binding to the plasma membrane, but also they are enriched in a variety of bioactive genes that can transmit messages between cells [11]. Furthermore, exosomes are present in all biological fluids. Exosomes extracted from body fluids were performed multicomponent analysis, which can accurately reflect the source of the cells in which they occur and their physio-pathological status [12]. It may assist in the diagnosis and prognosis of diseases. At present, with the gradual progress of research, exosomes have been paid more and more

attention. For example, umbilical cord mesenchymal stem cell-derived exosomes (ucMSC-Exs) were validated in our previous study to promote pancreatic tissue repair in traumatic pancreatitis injury by inhibiting pancreatic acinar cell apoptosis and controlling the systemic inflammatory response [13]. Kong et al. [14] investigated the effect of cancer cell-derived exosomes linc00313 on M2 macrophage differentiation in non-small cell lung cancer (NSCLC). They found that cancer cell-derived exosomes linc00313 promoted M2 macrophage differentiation in non-small cell lung cancer by upregulating STAT6. Wang et al. investigated the efficacy and mechanism of epidermal stem cell-derived exosomes (ESCs-exo) in improving impaired diabetic wound healing, elucidating that ESCs-exo enhanced the proliferation and migration of diabetic fibroblasts and macrophages and accelerated M2 macrophage polarization to promote diabetic wound healing. Besides cell-derived exosomes, some natural exosomes from body fluids also affect important biological functions. Pan et al. [15] demonstrated that exosomes derived from urine stem cells (USCS) can induce neurogenesis and contribute to recover the function of cerebral ischemia. Mi et al. [16] found that saliva-derived exosomes can induce human umbilical vein endothelial



**Fig. 1** Origin and structural characteristics of exosomes

cell (HUVEC) proliferation, migration, and angiogenesis in vitro and promote skin wound healing in vivo. Zhou et al. [17] found that human milk-derived exosomes ameliorated hyperoxia-induced cell injury to prevent neonatal bronchopulmonary dysplasia by inhibiting the downstream of the IL-17 signaling pathway. However, the bottleneck for exosomes to exert their extensive and abundant biological functions is limited yield and poor targeting ability. Therefore, it is an urgent problem to massively improve the yield of exosomes and their targeted enrichment in target organs [18–20].

Firstly, the multiple biological functions of exosomes depend on the accumulation of a certain amount of exosomes in target organ tissues. The improvement of exosome yield is a big problem for future applications. Extraction of exosomes is difficult due to their differences in size, content, function, and source [21, 22]. The current separation techniques cannot separate it accurately, resulting in relatively low exosome extraction purity and extraction yield [23, 24]. Therefore, how to efficiently enrich exosomes is an important topic at present, which is crucial for analyzing the molecular mechanism of the role of exosomes.

Secondly, another major obstacle to the future clinical application of exosomes is targeted delivery [25]. Improving the enrichment or colonization content of exosomes in target organs or target tissues can more efficiently exert the dominant effect of the exosomes and save the amount of the exosomes used for exerting the same effect, which will relieve the pressure for urgently improving the yield of the exosomes from the side. Natural exosomes are unable to deliver drugs efficiently and be targeted for applications due to several disadvantages such as poor stability and easy destruction [5]. In addition, after the exosomes enter the body, the utilization rate significantly decreases owing to blood circulation, immune clearance and retention of organs [26, 27]. To meet research needs, engineering exosomes by means such as the surface modification or drug loading is a way of practical application that makes exosomes superior in terms of yield and targeted therapy, thereby achieving a precise treatment of exosomes and accelerating the clinical application of exosomes [28, 29].

In this review, we first introduced strategies to solve the mass production problems of exosomes in clinical and research applications: exogenous treatment for parent cells and different extraction methods. Secondly, two methods of improving the targeting ability of exosomes are introduced. One is to improve the targeting ability of exosomes carrying drugs; the other is to improve the targeting ability by engineering modification of the exosome structure. Finally, we summarize the trends and possible challenges in future research and application of

exosomes. Therefore, many researchers have made a lot of efforts how to standardize the isolation, purification, and quantification of exosomes and to improve the targeting ability of exosomes, which has laid the theoretical and technical foundation for the clinical application of exosome therapy.

## **Optimization strategy of exosomes in clinical application and medical research—improving the yield of exosomes**

### **Exogenous treatment of parent cells**

In the clinical application of medical research, the exosomes secreted by cells through paracrine function play a major role [30, 31]. Therefore, the status, vitality, proliferation status and living environment of parent cells are closely related to the content and function of exosomes [32, 33]. In order to produce safer and more effective exosomes for clinical transformation research on a large scale, different measures are taken for parent cells to change and affect the exosomes yield from the source, which is crucial for the development of new therapies for diseases in the future. We summarize recent advances regarding the exogenous management of parental cells.

### **Origin and cell status of parent cells**

A variety of cells in the body can secrete exosomes, and the content of exosomes produced by different cell sources is also different. Some scholars have compared the production and doubling times of exosomes of mesenchymal stem cells from different sources. They have found that umbilical cord mesenchymal stem cells grow faster than mesenchymal stem cells from bone marrow or other tissues under the same culture conditions. Umbilical cord mesenchymal stem cells produce more and larger exosomes than bone marrow or other tissues [34]. Cell state is closely related to cell culture generation. Telomerase activity is continuously decreased during cell division and proliferation, resulting in cell telomere shortening and partial gene loss at the late stage of DNA replication. After multiple passages, the cell senescence occurs, and the differentiation ability of stem cells is gradually reduced. Similarly, the yield and purity of exosomes can be significantly reduced, resulting in an impact on the efficacy of exosome therapy. Some studies have found that there is a positive correlation between the degree of cell aging and cell generation by analyzing the biological characteristics of each generation of bone marrow mesenchymal stem cells. During the culture, the number of apoptotic cells increased significantly from the sixth generation. After the eighth generation, more apoptotic cells appeared and such aging phenomena as cytoplasmic vacuolation and cell body enlargement [35].

In addition, cell inoculation density may affect exosome secretion. Studies have shown that cell-to-cell contact can lead to various physiological changes in cells. This may include cessation of cell growth and changes in differentiation status [36]. One study reported that with the increase in the number of passages of mesenchymal stem cells, the biological activity of exosomes of mesenchymal stem cells was significantly reduced. They further demonstrated that reduced cell seeding density in the culture flask resulted in increased exosome production [37].

Therefore, scholars should pay attention to the following points when obtaining exosomes: First, select appropriate parent cells according to the needs of the experiment. When there are many kinds of parent cells to choose, the cell line with a high yield should be given priority. Secondly, we should select the parent cells with high activity and primitive to improve the ability to derive exosomes. Finally, frequently observe the cell density and growth state in the process of cell culture and select the parent cells in the logarithmic growth period to extract the exosomes.

### **Three-dimensional cell culture**

Three-dimensional cell culture is a technique in which biological cells can be grown in artificially created three-dimensional spaces. Three-dimensional cell culture enables cells to migrate and grow in the three-dimensional tridimensional spatial structure of the vector, constituting three-dimensional cell carrier complexes and allowing cells to grow in all directions in vitro, similar to how they grow in vivo. Three-dimensional culture technology not only preserves the material and structural basis of the in vivo cellular microenvironment, but also exhibits the advantages of intuitiveness and condition controllability of cell culture. Studies have shown that the production of exosomes secreted by umbilical cord-derived mesenchymal stem cells based on three-dimensional culture is 20 times that of two-dimensional (2D) culture [34]. Zhang et al. found that the exosomes produced by three-dimensional cultured mesenchymal stem cells had a stronger anti-inflammatory effect and more stable properties by replacing the traditional two-dimensional culture system with the three-dimensional system [38]. Yang et al. cultured human umbilical cord mesenchymal stem cells using the three-dimensional culture technique and isolated exosomes from the obtained supernatant. They found that three-dimensional cultured exosomes (3D-Exo) can reduce A $\beta$  production in Alzheimer's disease pathological cells and transgenic mice to improve memory and cognitive deficits in Alzheimer's mice [39]. Therefore, making full use of three-dimensional cell culture technology contributes to the large-scale production

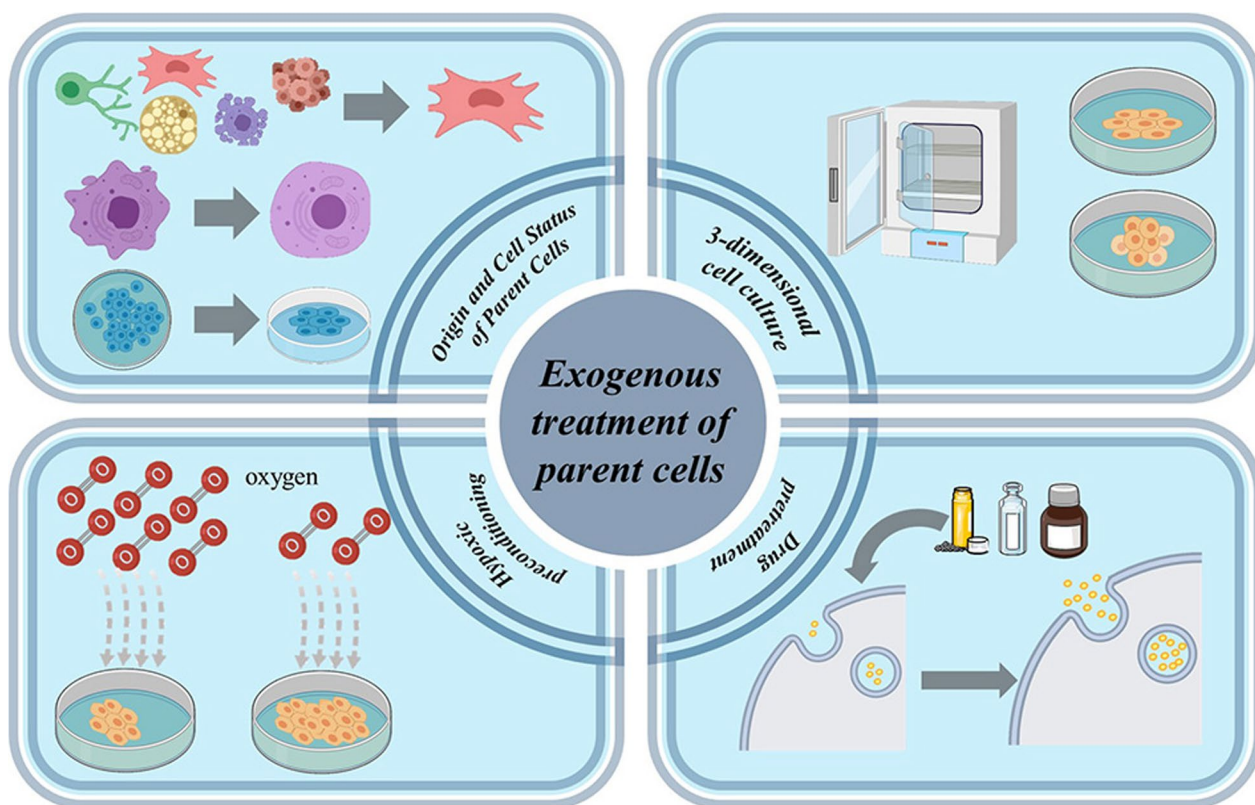
of exosomes and provides greater potential for the clinical transformation of exosomes.

### **Hypoxic preconditioning**

Many diseases or injuries create a hypoxic microenvironment, while stem cells secreting exosomes have a natural tropism to the site of pathological changes such as hypoxia and inflammation and are able to regulate the secretion of corresponding substances depending on the microenvironmental conditions [40, 41]. Stem cell transplantation therapy holds promise for many difficult diseases, but the low number of surviving transplanted cells and their poor secretory function make it difficult to exert therapeutic utility [42]. Hypoxic preconditioning can effectively improve biological characteristics such as proliferative activity, anti-apoptotic ability, and secretory function of implanted cells. Studies have shown that hypoxic preconditions of mesenchymal stem cells (MSCs) can enhance their paracrine effects. This study, using an in vivo fracture model and in vitro experiments, confirms that hypoxic preconditioning can be used as a method to optimize the therapeutic effects of MSC-derived exosomes on fracture healing [43]. Therefore, hypoxic preconditioned stem cells can effectively optimize the yield and function of exosomes to exert better therapeutic effects.

### **Drug pretreatment**

Because of the safety, low toxicity, and immunogenicity of exosomes, the therapeutic outcomes based on exosomes have greatly exceeded initial expectations in many clinically difficult diseases, but the yield of exosomes is a bottleneck for their widespread use. Taking different drug pretreatment to the parental cells could increase the yield of exosomes and optimize the function of exosomes to some extent. Studies have found that melatonin is synthesized from tryptophan and has protective effects under pathological conditions. As a novel preconditioning method, melatonin could effectively enhance the antioxidant, anti-inflammatory, and anti-apoptotic functions of exosomes in chronic kidney disease, diabetic wound healing, and ischemia-reperfusion therapy [44]. Another study pretreated bone marrow-derived mesenchymal stem cells (BMSCs) with traditional Chinese medicine Tongxinluo (TXL) and then collected their derived exosomes, verifying that exosomes secreted by TXL pretreated bone marrow-derived mesenchymal stem cells exhibited enhanced anti-apoptotic and anti-inflammatory effects in acute myocardial infarction (AMI) compared with untreated exosomes [45]. Therefore, proper pretreatment of the source cells combined with a suitable centrifugation method is effective in increasing exosome yield as well as functional properties (Fig. 2).



**Fig. 2** Improving the yield of exosomes from four aspects: origin and cell status of parent cells, three-dimensional cell culture, hypoxic preconditioning, and drug pretreatment

**Improving the yield of exosomes—extraction method**

With the deepening of research in the field of exosomes, in order to better its biological function and further explore its molecular mechanism. Researchers have established many separation techniques to continuously optimize the extraction of exosomes (Table 1). Heretofore, there is still no method that can guarantee the content, purity, and biological activity of exosomes at the same time.

**Traditional methods**

The traditional extraction methods for exosomes are mainly as follows: ultracentrifugation, ultrafiltration centrifugation, chromatography, precipitation, immunoaffinity chromatography, and immunomagnetic beads. The advantages of ultracentrifugation are low cost-effectiveness, no risk of contamination, and the ability to extract larger amounts of exosomes [46]. But this method consumes time due to continuous centrifugation [47]. At the same time, the extracted exosomes may be damaged due to high-speed centrifugation. Ultrafiltration centrifugation uses ultrafiltration membranes with different relative molecular masses for selective separation [48, 49]. Small molecular substances are filtered to the other

side of the membrane, while high relative molecular mass substances larger than the pore size of the membrane is retained on the ultrafiltration membrane. This method is simple, efficient, and does not affect the biological activity of exosomes. However, the disadvantage is that the exosomes may block the filter holes, resulting in the shortened life of the membrane and low separation efficiency. In addition, the exosomes remaining on the membrane may adhere, resulting in decreased yield. The greatest advantages of obtaining exosomes by chromatography are short time, high yield, and no need for special equipment. However, this method has the same problem of low exosome purity as the ultracentrifugation method. Secondly, exosome loss due to possible membrane adhesion during the operation is also a problem faced by this method [49, 50]. The advantage of the precipitation method is that it is economical and does not need additional equipment [51]. However, there are many problems with this separation method: low purity and recovery rate, more foreign proteins, uneven particle size, and destruction of exosomes. The advantage of the immunoaffinity chromatography is that it can accurately separate the specified exosomes with high purity [52]. However, the disadvantages of this method are high

**Table 1** Extraction method of exosomes

Methods	Definition	Advantages	Disadvantages	Years	References
Ultracentrifugation	The method of separating and preparing exosomes by powerful centrifugal force in ultracentrifuges	Low cost-effectiveness, no risk of contamination, and the ability to extract larger amounts of exosomes	Consumes time, the extracted exosomes may be damaged due to high-speed centrifugation	2022	[55, 56]
Ultrafiltration centrifugation	Ultrafiltration membranes with different relative molecular masses for selective separation	Simple, efficient, and does not affect the biological activity of exosomes	Exosomes may block the filter holes, resulting in the shortened life of the membrane and low separation efficiency, decreased yield	2022	[57, 58]
Chromatography	The selective partitioning of different substances in different phase states to elute as a mixture in mobile relative stationary phases	Short time, high yield, and no need for special equipment	Low exosome purity as the ultracentrifugation method, exosome loss due to possible membrane adhesion	2018, 2022	[58, 59]
Precipitation	The product of interest or major impurities in solution are isolated by precipitation in the form of an amorphous solid phase	Economical and does not need additional equipment	Low purity and recovery rate, more foreign proteins, uneven particle size, and destruction of exosome	2021	[60]
Immunoaffinity chromatography	The highly specific affinity properties of antibodies to an antigen to separate a target from a mixture	Accurately separate the specified exosomes with high purity	High economic cost and low yield of exosomes	2021	[61]
Immunomagnetic beads	Incubating magnetic beads coated with anti-marker antibodies with exosome vesicles	High specificity, simple operation, and no influence on the shape integrity of exosomes	Efficiency is low, and the biological activity of exosomes is easily affected by pH and salt concentration	2017, 2019	[6, 62, 63]4
Biochips	Microchip technology based on the principle of specific interaction between molecules	Short time, accurately separate the specified exosomes with high purity	High economic cost, difficult clinical popularization, and low social benefit	2017	[65]
Extraction kit	Exosomes are isolated by sedimentation by centrifugation	Large number of exosomes, convenient operation, simple, and fast	Quality control of extraction kits, high economic cost, and low social benefit	2019	[66]

economic cost and low yield of exosomes. Exosomes have their specific markers (such as CD9, CD81, etc.). The exosomes can be adsorbed and separated by incubating magnetic beads coated with anti-marker antibodies with exosome vesicles. Because the heterogeneity of exosomes is consistent with their origin, the markers on different exosomes are also different. Different types of exosomes can be captured from samples by specific antibody combinations for selective separation. The magnetic bead method has the advantages of high specificity, simple operation, and no influence on the shape integrity of exosomes, but its efficiency is low, and the biological activity of exosomes is easily affected by pH and salt concentration, which is not conducive to downstream experiments and difficult to be widely popularized [53–55].

### **Emerging methods**

In recent years, with the rapid development of science and technology, a number of new methods for the extraction of exosomes have emerged, such as biochips and extraction kits. There are more than 1000 electrodes in an alternating current electrokinetic (ACE) microarray chip device and the surface of which is coated with a thin layer of porous hydrogel. Exosomes are enriched in the high field area. The exosomes enriched at the chip electrode can be directly analyzed and identified by scanning electron microscope and immunofluorescence. The device can quickly separate and obtain exosomes from undiluted human plasma samples, and requires a small number of samples. It can concentrate the exosomes in the high-field area around the microelectrode within 15 min. Separation, washing, and chip fluorescence analysis can be completed within 30 min. However, the disadvantages of this method are high economic cost, difficult clinical popularization, and low social benefit [56]. The extraction kit method can obtain a large number of exosomes with complete structure and function from cell supernatant, serum plasma, or other body fluids by simple mixing and conventional centrifugation. This method has the advantages of convenient operation, simple experimental steps, fast extraction speed, and strong compatibility. It is widely applicable to the isolation of cell supernatant, serum, plasma, or other body fluid secretions. The disadvantages of this method are the same as alternating current electrokinetic (ACE) microarray chip device, and the quality control of the kit should also be considered [57].

Exosomes are widely distributed in various body fluids. Further studies are needed on how to simplify the extraction of exosomes, improve the production of exosomes, and how effectively and accurately separate exosomes. Different separation methods were chosen for different purposes and applications. Maybe a combination of several methods that can be selected simultaneously

provides a more optimal strategy for effective exosome isolation.

## **Optimization strategy of exosomes in clinical application and medical research—improve the targeting ability of exosomes**

### **Drug loading mode**

Exosomes have similar biological structures and dominant physiological functions to those of the parent cells. However, exosomes are smaller and more stable. And exosomes contain abundant bioactive components such as nucleic acids, proteins, and lipids, carrying a lot of biological information. After exosomes are engulfed by target cells, intercellular signaling is completed by delivering these functional molecules to achieve functional regulation on recipient cells [58, 59]. However, the targeting of exosomes is insufficient. Therefore, it is easily cleared quickly and taken up by non-target cells after entering the body, or exosomes interact with the cell membrane, reducing the application of their dominant effect [60]. To solve the problem of limited targeting of exosomes, domestic and foreign scholars have modified the loading of the contents of the exosomes to make the exosomes have specific targeting or reduce the probability of being cleared by the organs, thereby achieving the purpose of improving the targeting and stability of the exosomes [61]. Exosome-targeted drug-loading strategies can be divided into two main categories, endogenous and exogenous.

### **Endogenous**

Endogenous is the loading of various drug molecules (such as nucleic acids, viral proteins, and chemical drugs) in the interior of exosomes [62]. This method co-incubates parental cells with drugs or transfects drug encoding DNA or RNA to parental cells, allowing drugs to enter the cytoplasm. Drugs in the cytoplasm are sorted into exosomes in active or passive ways, and then drug-loaded exosomes can be obtained by suitable extraction methods. This method only involves the treatment and transformation of cells and hardly treats exosomes. Its advantage is that it keeps the integrity and functionality of exosomes, but its disadvantage is that the drug loading efficiency is relatively low. Pascucci et al. increased the uptake capacity of mesenchymal stem cells by passive diffusion of paclitaxel into the cells through the pre-treatment of mesenchymal stem cells with paclitaxel. Exosomes were then extracted from mesenchymal stem cells co-incubated with paclitaxel. Finally, they found that exosomes extracted from mesenchymal stem cells co-incubated with paclitaxel had stronger anti-proliferative activity by comparing pre-treated exosomes with non-treated exosomes [63]. Chen et al. transfected human

adipose mesenchymal stem cells with a lentivirus, enabling them to overexpress miR-375. Exosomes carrying miR375 were then extracted from human adipose mesenchymal stem cells in a rat model of skull defect. They found that the bone regenerative capacity of exosomes carrying miR375 was enhanced by comparison with exosomes extracted from untreated human adipose mesenchymal stem cells [64].

### **Exogenous**

Exogenous loading means that exosomes are extracted first and then loaded with drugs to target specific cells or tissues for exerting effects. The advantages of exogenous are that the method is relatively simple and the drug-loading efficiency is relatively high. However, the drug-loading process may destroy the integrity of exosomes, and additional purification steps are required to remove untrapped drugs. We mainly introduce two common exogenous methods including sonication and electroporation.

Sonication is the sonication of exosomes using a probe sonicator, which deforms the exosome membrane, creating transient small pores that increase the permeability of the membrane, thereby allowing small molecular substances to diffuse into the exosomes. The advantage of this method is the high loading efficiency. The disadvantage of this method is to damage the membrane of exosomes leading to exosome destruction. Haney et al. developed a novel exosome-based delivery system to load catalase into exosomes with the method of sonication. The sonicated exosomes have the advantage of high loading efficiency, sustained release of catalase, and protection from protease degradation. This method not only increases the targeted delivery of drug-loaded exosomes to target cells, but also increases the therapeutic effect of the drug [65]. Electroporation is a method to create temporary hydrophilic pore channels on the exosome membrane and increase the permeability of the exosome membrane so that small molecular substances can cross the pore channels to enter the interior of the exosome. The advantage of this method is the higher efficiency of loading, and the disadvantage is that electroporation causes exosome aggregation and destabilizes the exosome membrane. Wahlgren et al. loaded siRNA into exosomes by using electroporation. Plasma-derived exosomes are used as gene-delivery vehicles to transport exogenous siRNA to cells. Thus, exosomes effectively produce effects by targeted delivery of siRNA into monocytes and lymphocytes [66].

In clinical application and medical research, due to many factors, such as drug properties, disease types, and different cell types, it is necessary to design targeted drug delivery methods according to the different properties of

loaded drugs and the characteristics of exosomes of different selected cells to improve the targeting ability of exosomes.

### **Engineered exosome biomembrane**

Exosomes are able to follow the blood circulation into various parts of the human body and even cross the blood–brain barrier. After exosomes enter the systemic circulation, what cannot avoid is engulfment or excretion by immune cells and the liver [67]. Therefore, their reaching target tissue efficiency depends on the degree of functionalization as well as the strength of interaction with the target cells. To improve the molecular transport capacity of exosomes and targeting properties, the engineering of exosomes is an effective approach. Engineered exosomes are gene fusions of ligands or targeting peptides to transmembrane proteins expressed on the surface of exosomes. Subsequently, donor cells are transfected with plasmids encoding fusion proteins that secrete engineered exosomes bearing targeting ligands on their surface, thereby conferring exosomes with cell and tissue specificity [68]. Methods used for engineering exosomes include chemical ligation of targeting peptides, genetic engineering to modify the exosome membrane, and magnetic nanoparticle technology. Lee et al. prepared sodium azide lipid-containing exosomes and conjugated them with targeting peptides using copper-free click chemistry to enhance targeting efficacy to cancer cells. They engineered the parental cells with membrane fusion liposomes, avoiding the destruction of the exosomes themselves while improving the targeting ability of exosomes in a more efficient and controlled manner [69]. Cheng et al. generated targeted exosomes of synthetic multivalent antibodies by modifying two different types of antibodies on the surface of exosomes. It elicits potent antitumor immunity in vitro and in vivo by expressing monoclonal antibodies specific for T-cell-associated CD3 and cancer cell-associated epidermal growth factor receptor (EGFR). This study achieved the ability to orient and activate cytotoxic T-cells for the targeted killing of cancer cells by engineering modifications to exosome surface membrane proteins [70]. Khongkow et al. utilized gold nanoparticles with multifunctional properties of surface modification combined with exosome membrane proteins for targeted delivery. After symbiosis with brain-targeted exosomes, the unique targeting properties of gold nanoparticles were demonstrated by their binding to brain cells under laminar flow conditions and their enhanced transport through the blood–brain barrier. Synthetic surface modification of gold nanoparticles with brain-targeted exosomes represents a very novel and efficient strategy to provide effective brain targeting [71].



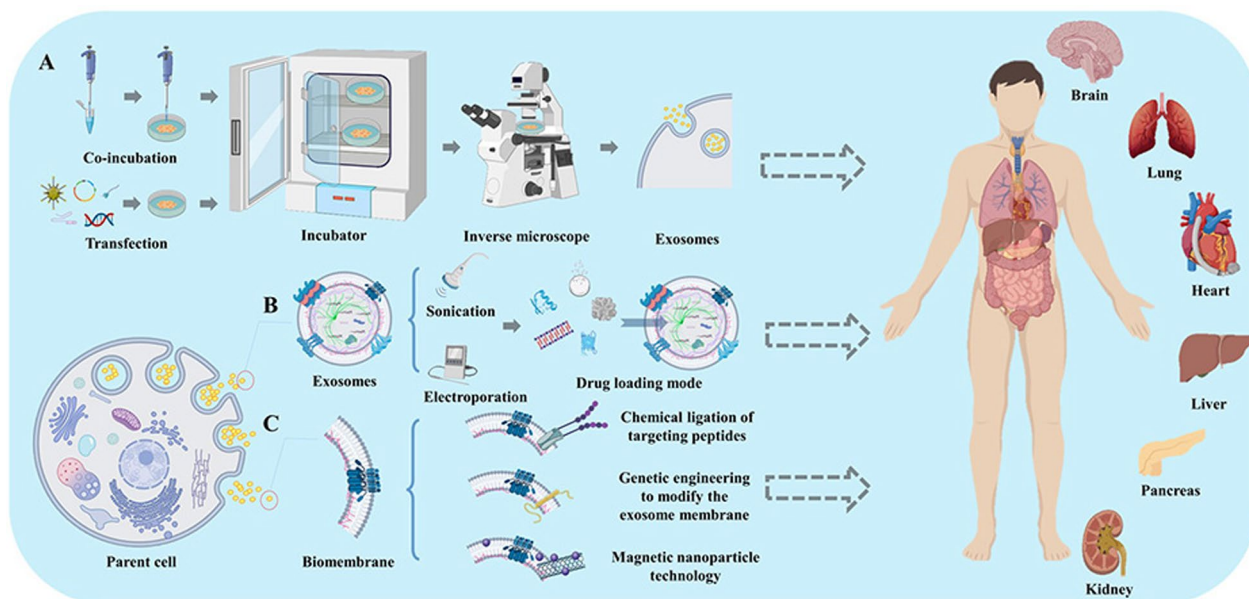
The dominant effects of exosomes are mainly manifested in many processes, such as cell signaling, tissue transformation and re-differentiation, and regulation of signaling pathways. Exosomes are easily manipulated and target active molecules can be encapsulated during the production of exosomes secreted by parental cells and are able to cross multiple biological barriers (Fig. 3 and Table 2). Exosomes have weak targeting ability. This leads to the waste of exosomes in use and affects therapeutic efficacy. In addition, there is increased aggregation in non-target organs or tissues and toxic side effects. Therefore, improving the targeting ability of exosomes is an obligatory way to achieve a large number of future applications of exosomes.

### Summarization and prospect

In recent years, exosomes have been found to serve as a novel kind of information carrier for intercellular information transmission, participating in the regulation of molecular and genetic characteristics of normal or abnormal cells. The presence of exosomes has provided the possibility for cell-free based therapies and sparked a boom in research with strong advantages. In order to obtain higher-quality exosomes for clinical application. It is very important to use appropriate exosome extraction methods, culture conditions, and new engineering transformation technologies [72, 73]. At present, there are many kinds of exosome extraction methods, all of which have their own advantages. However, the exosomes obtained by any extraction methods are heterogeneous,

and a single exosome may contain different amounts and kinds of bioactive substances. Second, the differences between exosomes from different sources are still unclear, and their functional differences have not been fully elucidated. Hence, they cannot meet research and clinical needs. However, both an ideal drug targeting system and the system of the engineered exosome targeting capabilities must ensure that it is nontoxic and nonimmunogenic, and it is also free of adverse effects in vitro and in vivo, which cannot increase its human body burden. Therefore, the application of exosomes still needs further research development.

There are some unsolved problems in clinical application and medical research of exosomes besides the above-mentioned obstacles. For example, the storage of exosomes is time-sensitive and affected by many factors [74]. It should be used as soon as possible after extraction. Therefore, it is of great research value and clinical application prospect to develop a new storage method to maintain the structure and biological function of exosomes. Secondly, most researches mainly focus on the exosomes derived by human stem cells, and the research on the advantages and benefits of plant-derived exosomes should be strengthened. Plant-derived exosomes do not involve ethical issues and are relatively safe. However, there are still many unexplored problems in the mechanism of plant-derived exosomes, which need further study. In summary, although clinically applicable exosome extraction methods are not uniform, engineering is still in the stage of exploration. But the future



**Fig. 3** Improve the targeting ability of exosomes. **A:** two different endogenous loading modes, **B:** two different exogenous loading modes, **C:** three different engineered exosome biomembrane modes

**Table 2** Methods for enhancing the targeting ability of exosomes

Modification strategies	Methods	Main contents	Years	References
Drug-loading strategies	Endogenous	Co-incubates parental cells with drugs	2014	[72]
	Exogenous	Transfects drug encoding DNA or RNA to parental cells	2019	[73]
		Sonication	The method of artificially introducing nucleic acid (DNA or RNA) into cells to change the characteristics of cells	2015
Engineered exosome biomembrane	Chemical ligation of targeting peptides	Sonication is the sonication of exosomes using a probe sonicator, which deforms the exosome membrane, creating transient small pores that increase the permeability of the membrane, thereby allowing small molecular substances to diffuse into the exosomes	2012	[75]
		Electroporation	Electroporation is a method to create temporary hydrophilic pore channels on the exosome membrane and increase the permeability of the exosome membrane so that small molecular substances can cross the pore channels to enter the interior of the exosome	2016
	Genetic engineering to modify the exosome membrane	The chemical linkage of targeting peptides is to fuse relevant targeting peptides with exosomal highly expressed proteins to construct exosomes that specifically target various types of tissues or cells	2018	[79]
Magnetic nanoparticle technology	Magnetic nanoparticle technology	Gene engineering fuses the gene sequence of the guiding protein or polypeptide with the gene sequence of the selected exosome membrane protein, which can effectively display the specific guiding peptide and protein on the exosome surface	2019	[80]
		Magnetic nanoparticle technology uses magnetic nanoparticles with dual targeting function to capture and release endogenous exosomes to target organs		

development of exosomes is still promising. Exploring more methods that can optimize the composition and function of exosomes is a novel and promising direction and has profound implications for the translational medicine of exosomes in the future.

## Conclusion

Exosomes have attracted much attention in recent years as nanoscale biomarkers mediate cellular communication. In order to make better use of multiple advantageous effects, research on improving the yield and targeting ability of exosomes has taken a blowout. This review summarizes the current optimization strategies for exosomes in clinical applications and compares their advantages and disadvantages. In addition, we also propose possible problems and the research prospects in the application of exosomes. Although there are still many challenges in the application of exosomes in disease treatment, however, with the advancement of medical technology, it is just around the corner to further explore the potential of exosomes in translational medicine and provide new ways to create effective clinical diagnoses and treatment strategies.

## Abbreviations

RNA	Ribonucleic acid
Hsp	Heat shock proteins
mRNA	Messenger RNA
ucMSC-Exs	Umbilical cord mesenchymal stem cell-derived exosomes
NSCLC	Non-small cell lung cancer
STAT6	Signal transducer and activator of transcription 6
ESCs-exo	Stem cell-derived exosomes
USCS	Urine stem cells
HUVEC	Human umbilical vein endothelial cell
DNA	Deoxyribonucleic acid
3D	3-Dimensional
3D-Exo	3D-cultured exosomes
MSCs	Mesenchymal stem cells
BMSCs	Bone marrow-derived mesenchymal stem cells
TXL	Tongxinluo
AMI	Acute myocardial infarction
ACE	Alternating current electrokinetic
EGFR	Epidermal growth factor receptor
MCM2	Minichromosome maintenance complex component 2

## Acknowledgements

Not applicable.

## Author contributions

LH and ZRZ participated in the literature search, data interpretation, and writing. CSH, JML, and XYL participated in literature collection, data collation, and article framework construction. ML participated in writing and critical revision. All authors read and approved the final manuscript.

## Funding

This work was supported by the National Natural Science Foundation of China-Ultrasound-Driven Biomimetic Nanosystem Targeting MCM2 to Rescue Imatinib Resistance in Gastrointestinal Stromal Tumor (82272015).

## Availability of data and materials

Not applicable.

## Declarations

### Ethics approval and consent to participate

Not applicable.

### Consent for publication

Not applicable.

### Competing interests

The authors declare that they have no competing interests.

Received: 30 November 2022 Accepted: 16 March 2023

Published online: 01 April 2023

## References

- Fonseka P, Chitti SV, Sanwlani R, Mathivanan S. Sulfoxazole does not inhibit the secretion of small extracellular vesicles. *Nat Commun*. 2021;12(1):977. <https://doi.org/10.1038/s41467-021-21074-x>.
- Meng L, Song K, Li S, Kang Y. Exosomes: small vesicles with important roles in the development, metastasis and treatment of breast cancer. *Membranes* (Basel). 2022. <https://doi.org/10.3390/membranes12080775>.
- Zhu CC, Gong LS, Yang Y. Label-free analysis of exosomes with hairpin structure-mediated multiple signal amplification strategy. *Appl Biochem Biotechnol*. 2022. <https://doi.org/10.1007/s12010-022-03978-6>.
- Ha D, Yang NN, Nadithe V. Exosomes as therapeutic drug carriers and delivery vehicles across biological membranes: current perspectives and future challenges. *Acta Pharm Sin B*. 2016;6(4):287–96. <https://doi.org/10.1016/j.apsb.2016.02.001>.
- Wang WT, Liang XL, Zheng K, Ge GR, Chen X, Xu YZ, et al. Horizon of exosome-mediated bone tissue regeneration: the all-rounder role in biomaterial engineering. *Mater Today Bio*. 2022;16:18. <https://doi.org/10.1016/j.mtbio.2022.100355>.
- Xu K, Jin YL, Li YM, Huang YY, Zhao R. Recent progress of exosome isolation and peptide recognition-guided strategies for exosome research. *Front Chem*. 2022;10:12. <https://doi.org/10.3389/fchem.2022.844124>.
- Prakash A, Crespo-Avilan GE, Hernandez-Resendiz S, Ong S-G, Hausenloy DJ. Extracellular vesicles—mediating and delivering cardioprotection in acute myocardial infarction and heart failure. *Cond Med*. 2020;3(4):227–38.
- Ferguson SW, Nguyen J. Exosomes as therapeutics: the implications of molecular composition and exosomal heterogeneity. *J Control Release*. 2016;228:179–90. <https://doi.org/10.1016/j.jconrel.2016.02.037>.
- Frydrychowicz M, Kolecka-Bednarczyk A, Madejczyk M, Yasar S, Dworacki G. Exosomes—structure, biogenesis and biological role in non-small-cell lung cancer. *Scand J Immunol*. 2015;81(1):2–10. <https://doi.org/10.1111/sji.12247>.
- Rupert DLM, Claudio V, Lasser C, Bally M. Methods for the physical characterization and quantification of extracellular vesicles in biological samples. *BBA-Gen Subj*. 2017;1861(1):3164–79. <https://doi.org/10.1016/j.bbagen.2016.07.028>.
- Batista IA, Quintas ST, Melo SA. The interplay of exosomes and NK cells in cancer biology. *Cancers* (Basel). 2021. <https://doi.org/10.3390/cancers13030473>.
- Kučuk N, Primožič M, Knez Ž, Leitgeb M. Exosomes engineering and their roles as therapy delivery tools, therapeutic targets, and biomarkers. *Int J Mol Sci*. 2021. <https://doi.org/10.3390/ijms22179543>.
- Han L, Zhao Z, Chen X, Yang K, Tan Z, Huang Z, et al. Human umbilical cord mesenchymal stem cells-derived exosomes for treating traumatic pancreatitis in rats. *Stem Cell Res Ther*. 2022;13(1):221. <https://doi.org/10.1186/s13287-022-02893-1>.
- Kong W, Zhang L, Chen Y, Yu Z, Zhao Z. Cancer cell-derived exosomal LINC00313 induces M2 macrophage differentiation in non-small cell lung cancer. *Clin Transl Oncol*. 2022;24(12):2395–408. <https://doi.org/10.1007/s12094-022-02907-7>.
- Wang P, Theocharidis G, Vlachos IS, Kounas K, Lobao A, Shu B, et al. Exosomes derived from epidermal stem cells improve diabetic wound

- healing. *J Invest Dermatol.* 2022;142(9):2508–2517.e13. <https://doi.org/10.1016/j.jid.2022.01.030>.
16. Mi BB, Chen L, Xiong Y, Yan CC, Xue H, Panayi A, et al. Saliva exosomes-derived UBE2O mRNA promotes angiogenesis in cutaneous wounds by targeting SMAD6. *J Nanobiotechnol.* 2020;18(1):14. <https://doi.org/10.1186/s12951-020-00624-3>.
  17. Zhou YH, Liu YW, Xu G, Liu LJ, Li HM, Li YB, et al. Human breast milk-derived exosomes through inhibiting AT II cell apoptosis to prevent bronchopulmonary dysplasia in rat lung. *J Cell Mol Med.* 2022;26(15):4169–82. <https://doi.org/10.1111/jcmm.17334>.
  18. Sattar RSA, Verma R, Nimisha Kumar A, Dar GM, et al. Diagnostic and prognostic biomarkers in colorectal cancer and the potential role of exosomes in drug delivery. *Cell Signal.* 2022;99:110413. <https://doi.org/10.1016/j.cellsig.2022.110413>.
  19. Wu Y, Huang Q, Bu SZ. Cross talk between exosomes and pancreatic beta-cells in diabetes. *Arch Physiol Biochem.* 2022;128(5):1140–9. <https://doi.org/10.1080/13813455.2020.1760303>.
  20. Yao JL, Cai LQ, Chen YR, Zhang J, Zhuang WW, Liang JY, et al. Exosomes: mediators regulating the phenotypic transition of vascular smooth muscle cells in atherosclerosis. *Cell Commun Signal.* 2022. <https://doi.org/10.1186/s12964-022-00949-6>.
  21. Kimiz-Gebologlu I, Oncel SS. Exosomes: large-scale production, isolation, drug loading efficiency, and biodistribution and uptake. *J Control Release.* 2022;347:533–43. <https://doi.org/10.1016/j.jconrel.2022.05.027>.
  22. Burton JB, Carruthers NJ, Stemmer PM. Enriching extracellular vesicles for mass spectrometry. *Mass Spectrom Rev.* 2021. <https://doi.org/10.1002/mas.21738>.
  23. Saad MG, Beyenal H, Dong WJ. Exosomes as powerful engines in cancer: isolation, characterization and detection techniques. *Biosensors-Basel.* 2021. <https://doi.org/10.3390/bios11120518>.
  24. Xu WM, Li A, Chen JJ, Sun EJ. Research development on exosome separation technology. *J Membr Biol.* 2022. <https://doi.org/10.1007/s00232-022-00260-y>.
  25. Fang XN, Wang YQ, Wang SR, Liu BH. Nanomaterials assisted exosomes isolation and analysis towards liquid biopsy. *Mater Today Bio.* 2022;16:18. <https://doi.org/10.1016/j.mtbio.2022.100371>.
  26. Khayambashi P, Iyer J, Pillai S, Upadhyay A, Zhang YL, Tran SD. Hydrogel encapsulation of mesenchymal stem cells and their derived exosomes for tissue engineering. *Int J Mol Sci.* 2021;22(2):15. <https://doi.org/10.3390/ijms22020684>.
  27. Antimisiaris SG, Mourtas S, Marazioti A. Exosomes and exosome-inspired vesicles for targeted drug delivery. *Pharmaceutics.* 2018;10(4):40. <https://doi.org/10.3390/pharmaceutics10040218>.
  28. Lee J, Lee JH, Chakraborty K, Hwang J, Lee YK. Exosome-based drug delivery systems and their therapeutic applications. *RSC Adv.* 2022;12(29):18475–92. <https://doi.org/10.1039/d2ra02351b>.
  29. Cheng J, Sun YX, Ma Y, Ao YF, Hu XQ, Meng QY. Engineering of MSC-derived exosomes: a promising cell-free therapy for osteoarthritis. *Membranes.* 2022;12(8):28. <https://doi.org/10.3390/membranes12080739>.
  30. Lin Y, Zhu W, Chen XM. The involving progress of MSCs based therapy in atherosclerosis. *Stem Cell Res Ther.* 2020;11(1):13. <https://doi.org/10.1186/s13287-020-01728-1>.
  31. Ju C, Liu R, Zhang Y, Zhang F, Sun J, Lv X-B, et al. Exosomes may be the potential new direction of research in osteoarthritis management. *Biomed Res Int.* 2019;2019:7695768. <https://doi.org/10.1155/2019/7695768>.
  32. Li J, Ge ZG, Ji WC, Yuan N, Wang KZ. The proosteogenic and proangiogenic effects of small extracellular vesicles derived from bone marrow mesenchymal stem cells are attenuated in steroid-induced osteonecrosis of the femoral head. *Biomed Res Int.* 2020;2020:11. <https://doi.org/10.1155/2020/4176926>.
  33. Wang YM, Liu JW, Adkins GB, Shen W, Trinh MP, Duan LY, et al. Enhancement of the intrinsic peroxidase-like activity of graphitic carbon nitride nanosheets by ssDNAs and its application for detection of exosomes. *Anal Chem.* 2017;89(22):12327–33. <https://doi.org/10.1021/acs.analchem.7b03335>.
  34. Haraszti RA, Miller R, Stoppato M, Sere YY, Coles A, Didiot MC, et al. Exosomes produced from 3D cultures of MSCs by tangential flow filtration show higher yield and improved activity. *Mol Ther.* 2018;26(12):2838–47. <https://doi.org/10.1016/j.yjthe.2018.09.015>.
  35. He Q, Ye ZY, Zhou Y, Tang WS. Comparative study of mesenchymal stem cells from rat bone marrow and adipose tissue. *Turk J Biol.* 2018;42(6):477–89. <https://doi.org/10.3906/biy-1802-52>.
  36. Lieberman MA, Glaser L. Density-dependent regulation of cell growth: an example of a cell-cell recognition phenomenon. *J Membr Biol.* 1981;63(1–2):1–11. <https://doi.org/10.1007/bf01969440>.
  37. Patel DB, Gray KM, Santharam Y, Lamichhane TN, Stroka KM, Jay SM. Impact of cell culture parameters on production and vascularization bioactivity of mesenchymal stem cell-derived extracellular vesicles. *Bioeng Transl Med.* 2017;2(2):170–9. <https://doi.org/10.1002/btm2.10065>.
  38. Zhang Y, Chen JY, Fu HJ, Kuang SH, He F, Zhang M, et al. Exosomes derived from 3D-cultured MSCs improve therapeutic effects in periodontitis and experimental colitis and restore the Th17 cell/Treg balance in inflamed periodontium. *Int J Oral Sci.* 2021;13(1):15. <https://doi.org/10.1038/s41368-021-00150-4>.
  39. Yang LY, Zhai YX, Hao Y, Zhu ZC, Cheng GS. The regulatory functionality of exosomes derived from hUMSCs in 3D culture for alzheimer's disease therapy. *Small.* 2020;16(3):11. <https://doi.org/10.1002/smll.201906273>.
  40. Rashed MH, Bayraktar E, Helal GK, Abd-Allah MF, Amero P, Chavez-Reyes A, et al. Exosomes: from garbage bins to promising therapeutic targets. *Int J Mol Sci.* 2017;18(3):25. <https://doi.org/10.3390/ijms18030538>.
  41. Zheng HM, Zhan YT, Liu SL, Lu JM, Luo JD, Feng J, et al. The roles of tumor-derived exosomes in non-small cell lung cancer and their clinical implications. *J Exp Clin Cancer Res.* 2018;37:11. <https://doi.org/10.1186/s13046-018-0901-5>.
  42. Song GW, Hu YN, Liu YS, Jiang R. Layer-by-layer heparinization of the cell surface by using heparin-binding peptide functionalized human serum albumin. *Materials.* 2018;11(5):10. <https://doi.org/10.3390/ma11050849>.
  43. Liu W, Li LW, Rong YL, Qian DF, Chen J, Zhou Z, et al. Hypoxic mesenchymal stem cell-derived exosomes promote bone fracture healing by the transfer of miR-126. *Acta Biomater.* 2020;103:196–212. <https://doi.org/10.1016/j.actbio.2019.12.020>.
  44. Zhou ZL, Wang RP, Wang J, Hao YJ, Xie QP, Wang L, et al. Melatonin pretreatment on exosomes: heterogeneity, therapeutic effects, and usage. *Front Immunol.* 2022;13:13. <https://doi.org/10.3389/fimmu.2022.933736>.
  45. Xiong YY, Tang RJ, Xu JY, Jiang WY, Gong ZT, Zhang LL, et al. Tongxinluo-pretreated mesenchymal stem cells facilitate cardiac repair via exosomal transfer of miR-146a-5p targeting IRAK1/NF-kappa B p65 pathway. *Stem Cell Res Ther.* 2022;13(1):18. <https://doi.org/10.1186/s13287-022-02969-y>.
  46. Dzaman K, Czerwaty K. Roles of exosomes in chronic rhinosinusitis: a systematic review. *Int J Mol Sci.* 2022;23(19):31. <https://doi.org/10.3390/ijms231911284>.
  47. Kaminski VD, Ellwanger JH, Chies JAB. Extracellular vesicles in host-pathogen interactions and immune regulation—exosomes as emerging actors in the immunological theater of pregnancy. *Heliyon.* 2019;5(8):17. <https://doi.org/10.1016/j.heliyon.2019.e02355>.
  48. Huang S, Hao XY, Li YJ, Wu JY, Xiang DX, Luo SL. Nonviral delivery systems for antisense oligonucleotide therapeutics. *Biomater Res.* 2022;26(1):23. <https://doi.org/10.1186/s40824-022-00292-4>.
  49. Li M, Fang F, Sun M, Zhang YF, Hu M, Zhang JF. Extracellular vesicles as bioactive nanotherapeutics: an emerging paradigm for regenerative medicine. *Theranostics.* 2022;12(11):4879–903. <https://doi.org/10.7150/thno.72812>.
  50. Yu LL, Zhu J, Liu JX, Jiang F, Ni WK, Qu LS, et al. A comparison of traditional and novel methods for the separation of exosomes from human samples. *Biomed Res Int.* 2018. <https://doi.org/10.1155/2018/3634563>.
  51. Zhao R, Zhao TT, He ZZ, Cai R, Pang WJ. Composition, isolation, identification and function of adipose tissue-derived exosomes. *Adipocyte.* 2021;10(1):587–604. <https://doi.org/10.1080/21623945.2021.1983242>.
  52. Wang J, Ma P, Kim DH, Liu BF, Demirci U. Towards microfluidic-based exosome isolation and detection for tumor therapy. *Nano Today.* 2021;37:27. <https://doi.org/10.1016/j.nantod.2020.101066>.
  53. Cheng H, Fang H, Xu RD, Fu MQ, Chen L, Song XY, et al. Development of a rinsing separation method for exosome isolation and comparison to conventional methods. *Eur Rev Med Pharmacol Sci.* 2019;23(12):5074–83.
  54. Li ZY, Hu CY, Jia J, Xia YY, Xie H, She MJ, et al. Establishment and evaluation of a simple size-selective method for exosome enrichment and

- purification. *J Biomed Nanotechnol.* 2019;15(5):1090–6. <https://doi.org/10.1166/jbn.2019.2768>.
55. Yang F, Liao XZ, Tian Y, Li GY. Exosome separation using microfluidic systems: size-based, immunoaffinity-based and dynamic methodologies. *Biotechnol J.* 2017;12(4):8. <https://doi.org/10.1002/biot.201600699>.
  56. Ibsen SD, Wright J, Lewis JM, Kim S, Ko SY, Ong J, et al. Rapid isolation and detection of exosomes and associated biomarkers from plasma. *ACS Nano.* 2017;11(7):6641–51. <https://doi.org/10.1021/acsnano.7b00549>.
  57. Boriachek K, Masud MK, Palma C, Phan HP, Yamauchi Y, Hossain MSA, et al. Avoiding pre-isolation step in exosome analysis: direct isolation and sensitive detection of exosomes using gold-loaded nanoporous ferric oxide nanozymes. *Anal Chem.* 2019;91(6):3827–34. <https://doi.org/10.1021/acs.analchem.8b03619>.
  58. Fonseca P, Vardaki I, Occhionero A, Panaretakis T. Metabolic and signaling functions of cancer cell-derived extracellular vesicles. In: Jeon KW, Galluzzi L, editors. *International review of cell and molecular biology*, vol. 326. San Diego: Elsevier Academic Press Inc; 2016. p. 175–99.
  59. He D, Zhao Z, Fu B, Li XF, Zhao L, Chen YB, et al. Exosomes participate in the radiotherapy resistance of cancers. *Radiat Res.* 2022;197(5):559–65. <https://doi.org/10.1667/rade-21-00115.1>.
  60. Zhang Y, Yu M, Tian WD. Physiological and pathological impact of exosomes of adipose tissue. *Cell Prolif.* 2016;49(1):3–13. <https://doi.org/10.1111/cpr.12233>.
  61. Zhang Y, Liu YF, Liu HY, Tang WH. Exosomes: biogenesis, biologic function and clinical potential. *Cell Biosci.* 2019;9:18. <https://doi.org/10.1186/s13578-019-0282-2>.
  62. Wang XY, Zhang HY, Yang HO, Bai M, Ning T, Li S, et al. Cell-derived exosomes as promising carriers for drug delivery and targeted therapy. *Curr Cancer Drug Targets.* 2018;18(4):347–54. <https://doi.org/10.2174/1568009617666170710120311>.
  63. Pascucci L, Cocce V, Bonomi A, Ami D, Ceccarelli P, Ciusani E, et al. Paclitaxel is incorporated by mesenchymal stromal cells and released in exosomes that inhibit in vitro tumor growth: a new approach for drug delivery. *J Control Release.* 2014;192:262–70. <https://doi.org/10.1016/j.jconrel.2014.07.042>.
  64. Chen S, Tang YM, Liu YS, Zhang P, Lv LW, Zhang X, et al. Exosomes derived from miR-375-overexpressing human adipose mesenchymal stem cells promote bone regeneration. *Cell Prolif.* 2019;52(5):14. <https://doi.org/10.1111/cpr.12669>.
  65. Haney MJ, Klyachko NL, Zhaoa YL, Gupta R, Plotnikova EG, He ZJ, et al. Exosomes as drug delivery vehicles for Parkinson's disease therapy. *J Control Release.* 2015;207:18–30. <https://doi.org/10.1016/j.jconrel.2015.03.033>.
  66. Wahlgren J, Karlson TD, Brisslert M, Sani FV, Telemo E, Sunnerhagen P, et al. Plasma exosomes can deliver exogenous short interfering RNA to monocytes and lymphocytes. *Nucleic Acids Res.* 2012;40(17):12. <https://doi.org/10.1093/nar/gks463>.
  67. Alptekin A, Parvin M, Chowdhury HI, Rashid MH, Arbab AS. Engineered exosomes for studies in tumor immunology. *Immunol Rev.* 2022;312(1):76–102. <https://doi.org/10.1111/imr.13107>.
  68. Mishra A, Singh P, Qayoom I, Prasad A, Kumar A. Current strategies in tailoring methods for engineered exosomes and future avenues in biomedical applications. *J Mater Chem B.* 2021;9(32):6281–309. <https://doi.org/10.1039/d1tb01088c>.
  69. Lee J, Lee H, Goh U, Kim J, Jeong M, Lee J, et al. Cellular engineering with membrane fusogenic liposomes to produce functionalized extracellular vesicles. *ACS Appl Mater Interfaces.* 2016;8(11):6790–5. <https://doi.org/10.1021/acsmi.6b01315>.
  70. Cheng Q, Shi XJ, Han ML, Smbatyan G, Lenz HJ, Zhang Y. Reprogramming exosomes as nanoscale controllers of cellular immunity. *J Am Chem Soc.* 2018;140(48):16413–7. <https://doi.org/10.1021/jacs.8b10047>.
  71. Khongkow M, Yata T, Boonrungsiman S, Ruktanonchai UR, Graham D, Namdeel K. Surface modification of gold nanoparticles with neuron-targeted exosome for enhanced blood-brain barrier penetration. *Sci Rep.* 2019;9:9. <https://doi.org/10.1038/s41598-019-44569-6>.
  72. Huang LY, Song JX, Cai H, Wang PP, Yin QL, Zhang YD, et al. Healthy serum-derived exosomes improve neurological outcomes and protect blood-brain barrier by inhibiting endothelial cell apoptosis and reversing autophagy-mediated tight junction protein reduction in rat stroke model. *Front Cell Neurosci.* 2022;16:14. <https://doi.org/10.3389/fncel.2022.841544>.
  73. Lv LL, Wu WJ, Feng Y, Li ZL, Tang TT, Liu BC. Therapeutic application of extracellular vesicles in kidney disease: promises and challenges. *J Cell Mol Med.* 2018;22(2):728–37. <https://doi.org/10.1111/jcmm.13407>.
  74. Petrouskova P, Hudakova N, Maloveska M, Humenik F, Cizkova D. Non-exosomal and exosome-derived miRNAs as promising biomarkers in canine mammary cancer. *Life-Basel.* 2022;12(4):34. <https://doi.org/10.3390/life12040524>.

## Publisher's Note

Springer Nature remains neutral with regard to jurisdictional claims in published maps and institutional affiliations.

Ready to submit your research? Choose BMC and benefit from:

- fast, convenient online submission
- thorough peer review by experienced researchers in your field
- rapid publication on acceptance
- support for research data, including large and complex data types
- gold Open Access which fosters wider collaboration and increased citations
- maximum visibility for your research: over 100M website views per year

At BMC, research is always in progress.

Learn more [biomedcentral.com/submissions](https://biomedcentral.com/submissions)

