

ORIGINAL ARTICLE

Open Access

Study of bactericidal properties of carbohydrate-stabilized platinum oxide nanoparticles

Saeed Rezaei-Zarchi¹, Saber Imani^{2*}, Ali mohammad Zand², Mojtaba Saadati^{3,4} and Zahra Zaghari⁵

Abstract

Platinum oxide nanoparticles were prepared by a simple hydrothermal route and chemical reduction using carbohydrates (fructose and sucrose) as the reducing and stabilizing agents. In comparison with other metals, platinum oxide has less environmental pollution. Therefore, Pt is considered an appropriate candidate to deal with environmental pathogens. The crystallite size of these nanoparticles was evaluated from X-ray diffraction, atomic force microscopy, and transmission electron microscopy (TEM) and was found to be 10 nm, which is the demonstration of EM bright field and transmission electron microscopy. The effect of carbohydrates on the morphology of the nanoparticles was studied using TEM. The nanoparticles were administered to the *Pseudomonas stutzeri* and *Lactobacillus* cultures, and the incubation was done at 37°C for 24 h. The nanocomposites exhibited interesting inhibitory as well as bactericidal activity against *P. stutzeri* and *Lactobacillus* species. Incorporation of nanoparticles also increased the thermal stability of the carbohydrates. The results of this paper showed that carbohydrates can serve as a carrier for platinum oxide nanoparticles, and nanocomposites can have potential biological applications.

Keywords: Platinum oxide, *P. stutzeri*, *Lactobacillus*, Bactericidal effect

Background

Metal nanoparticles have different properties from those of bulk metal because of their small sizes, and thus, these materials can be employed in various photoelectric, catalytic, magnetic, sensor, and biomedical applications [1-3]. In particular, Pt is known to inactivate microbes by interacting with their enzymes, proteins, or DNA to restrain cell proliferation or cell division [4]. Among all, capping ligands play a vital role in transforming the spherical or triangular-shaped nanoparticles to nanodevices or nanosensors of any desired shape [5]. Much of the recent research efforts have focused on developing new strategies to fabricate nanoconstructs with carbohydrates [6,7] as capping ligands because of their potential applications in the design and development of nanoscale devices and nanosensors for

biomedical applications [8]. Carbohydrates contain many hydroxyl and carbonyl groups; these groups offer sugar-coated nanoparticle, a unique H-bonding capabilities in constructing supramolecular architecture. Upon surface coating with nanoparticles, they provide attractive nanoconstruction abilities for building smart nanomaterials. For example, nanowires of platinum or tellurium have been constructed from glucose-stabilized platinum nanoparticles or starch-stabilized tellurium nanoparticles [9-12]. Current strategies for carbohydrate functionalization of Pt nanoparticles utilize thiol-tailored sugars as synthons [12-15]. Quinti and coworkers have developed a 'green' method to synthesize and stabilize carbohydrate-Pt nanocomposites in starch matrix using glucose as the reducing agent [16].

In this work, we developed an easy method to produce platinum oxide composites with homogeneous size distribution and then these materials were stabilized with sucrose. These stabilized nanoparticles were then used as the antimicrobial agents against *Pseudomonas stutzeri* and *Lactobacillus* cultures in further experiments.

* Correspondence: imani.saber@yahoo.com

²Department of Chemical Injury Research Center, Medical Science Institute, Sheikh Bahaei Street, Baqiyatallah Medical Science University, Tehran 14359-44711, Iran

Full list of author information is available at the end of the article

Methods

The sucrose was supplied by Fibrocel - Produtos Biotecnológicos Ltda. (Ibiporã, Brazil). Platinum oxide and other reagents were commercially purchased from Sigma-Aldrich Corporation (St. Louis, MO, USA) and used as received. Aqueous solutions and bacterial suspensions were prepared using distilled deionized sterile water. Agar, tryptone, and yeast extract of Bacto™, used for the microbiological culture media, were purchased from Himedia (Mumbai, India). The *P. stutzeri* and *Lactobacillus* strains were taken from the Central Laboratory of Pathology, Yazd. All reagents were of analytical grade and used without further purification.

Syntheses of carbohydrate-stabilized platinum oxide nanoparticles

In a typical experiment, 1.7 g (1.0×10^{-2} mol) of platinum oxide (Sigma-Aldrich, 99.9+ %) was dissolved in 100 mL of deionized water. Then, the solution platinum oxide was precipitated with 0.62 g (1.55×10^{-2} mol) of sodium hydroxide (Sigma-Aldrich, 99+ %). An obtained precipitate of platinum oxide was filtered and dissolved in 100 mL of aqueous ammonia (0.4% w/w, 2.3×10^{-2} mol) until a transparent solution of platinum-ammonium complex was formed. Next, 2.5 g (8.9×10^{-3} mol) of oleic acid (Sigma-Aldrich, 99+ %) was added dropwise into the obtained complex, and the resulting solution was gently stirred for 2 h at room temperature until the complete homogeneity of the reaction mixture was achieved. Finally, 2 g (1.11×10^{-2} mol) of glucose was added to the mixture at room temperature with gentle stirring. The reduction process of silver complex solution (in quartz glass) was initiated with UV-irradiation (UV MILTON ROY Company, Ivyland, PA, USA; Spectrum 21). UV treatment was carried out for 8 h under vigorous stirring without additional heating. An UV lamp ($k = 365$ nm, 35 W) was used as light source to stimulate the reduction process. After 8 h of irradiation, the transparent dispersion obtained oleic acid-stabilized platinum oxide nanoparticles (platinum oxide concentration = 10 mg mL⁻¹). The synthesis of platinum oxide NPs was also successfully conducted with the final Pt concentrations in the range of 0.1% to 2%.

The TEM (H987 - Philips, Amsterdam, The Netherlands) images of the Pt oxide nanoparticles were taken by the following method: at first, a solution of Pt oxide was prepared and diluted 100 times with 50% ethanol. A drop of this diluted solution was added to formvar/carbon-coated grids (400 meshes) and, after drying, viewed under the TEM (Brazilian Synchrotron Light Laboratory, Sao Paulo, Brazil), operating at 80 kV.

XRD (U234 - Philips) patterns were obtained in a Siemens Kristalloflex diffractometer (Siemens, Knoxville,

TN, USA) using nickel filtered CuK α radiation from 4° to 70° (2θ angle) as shown in Figure 1. TG (STA 409 - Netzsch (Selb, Germany)) was conducted using dried samples in SDT 2960 device from TA Instruments (New Castle, DE, USA). Samples were heated in open alumina pans from 40°C to 600°C, under an oxidant atmosphere (O₂), using flux of 50 mL/min, at a heating rate of 10°C/min. The Pt oxide content of the BC/Pt composites was estimated from the residue at 600°C.

Antibacterial properties of carbohydrate-stabilized platinum oxide nanoparticles

The antimicrobial activity of sucrose-modified Pt oxide nanoparticles was investigated against *P. stutzeri* and *Lactobacillus*. The following solution and media were prepared to study the antimicrobial activity: (a) NaCl aqueous solution (2.7 g of NaCl dissolved in 300 mL of distilled water), sterilized by autoclaving at 120°C for 20 min; (b) culture medium (Luria-Bertani (LB)) containing 1.0-g NaCl, 1.0-g tryptone, 0.5-g yeast extract of Bacto™, and 100-mL distilled water, sterilized by autoclaving at 120°C for 20 min; and (c) solid LB medium, used to prepare petri dishes containing 3-g NaCl, 3-g tryptone, 1.5-g yeast extract of Bacto™, 4.5-g agar, and 300-mL distilled water. The solid LB medium was sterilized by autoclaving at 120°C for 20 min.

An aliquot (100 μ L) of the bacterial suspension prepared previously was transferred to a test tube with 9.9 mL of lysogenic broth medium (LB) and homogenized. This tube contained a concentration of 10^7 cells mL⁻¹. This bacterial suspension was diluted in a saline solution to obtain a suspension containing about 10^5 cells mL⁻¹. Aliquots of 1,000 μ L of the suspension containing about 10^5 cells mL⁻¹ were transferred to sterile test tubes containing the samples. The test tubes were incubated in a stirring incubator at 37°C and stirred at 120 rpm for 24 h. After incubation, the content of the tubes was transferred to four Erlenmeyer flasks filled with 50 mL of sterile NaCl aqueous solution at 0.9%.

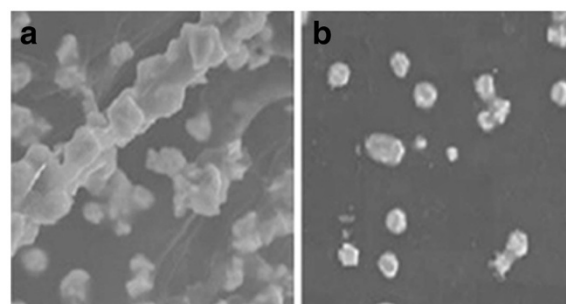


Figure 1 TEM images. (a) TEM bright field image of Pt nanoparticles. (b) TEM image of Pt nanoparticles obtained using sucrose as a reducing agent.

An aliquot (100 μL) of the content of each Erlenmeyer flask was transferred to a microtube filled with 900 μL of sterile NaCl aqueous solution. The microtubes were then vortexed. This dilution sequence was repeated four times to obtain four dilutions. Aliquots (100 μL) of these dilutions were then spread on a nutrient agar plate and incubated at 37°C for 24 h; after which, the colony-forming units were counted, and the average was taken from the three plates corresponding to a particular sample. The antibacterial activity of the carbohydrate-Pt nanocomposites impregnated in bacterial cellulose was determined at different concentrations (10^3 , 10^4 , 10^5 , 10^6 , and 10^7 cells mL^{-1}). The experiments were carried out in triplicate. Statistical analysis was performed using Student's *t*-test, and the percentage reduction in bacterial count was calculated by the following equation:

$$\text{Percentage reduction in bacterial count(\%)} = \left[\frac{\text{viable CFU in an hour} - \text{viable CFU in 24h}}{\text{viable CFU in an hour}} \right] \times 100$$

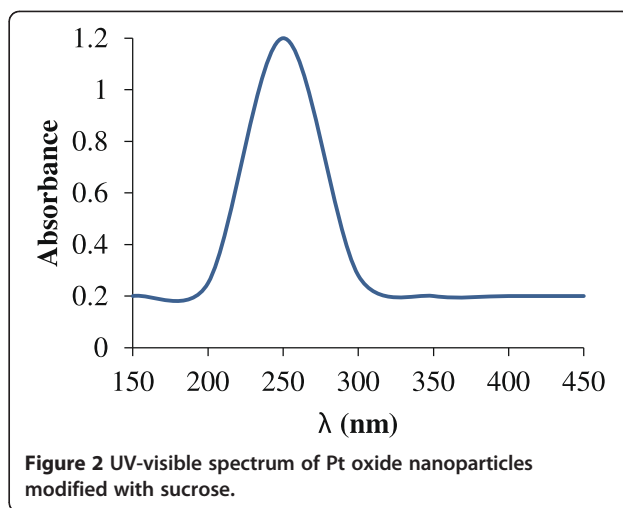
Results and discussion

Pt oxide nanoparticles could be successfully synthesized in the sucrose matrix. The reduction was fastest in the case of sucrose followed by waxy corn and soluble starch while stability of nanoparticles followed the order soluble starch > waxy corn starch > sucrose (data not shown). Thus, the size and molecular weight were important for stabilization of the nanoparticles.

Figure 1a shows the TEM image of Pt oxide nanoparticles alone. Figure 1b shows the TEM image of Pt oxide modified with sucrose. The sucrose reduction occurred only after heating at 80°C for 4 h. This is required because the more hydrolyzed the carbohydrate the better its ability to act as a reducing agent. UV-visible illumination was used to enhance the reduction capacity of sucrose where it is hydrolyzed, and the hydrolyzed products then reduce the metal. The time taken for reduction is also very short (30 s).

UV-vis absorption spectrum of carbohydrate-stabilized Pt oxide nanoparticles in aqueous solution was recorded after sufficient dilution as demonstrated in Figure 2. The characteristic absorption peak due to the surface-plasmon resonance of Pt colloids was observed within the range of 415 to 425 nm.

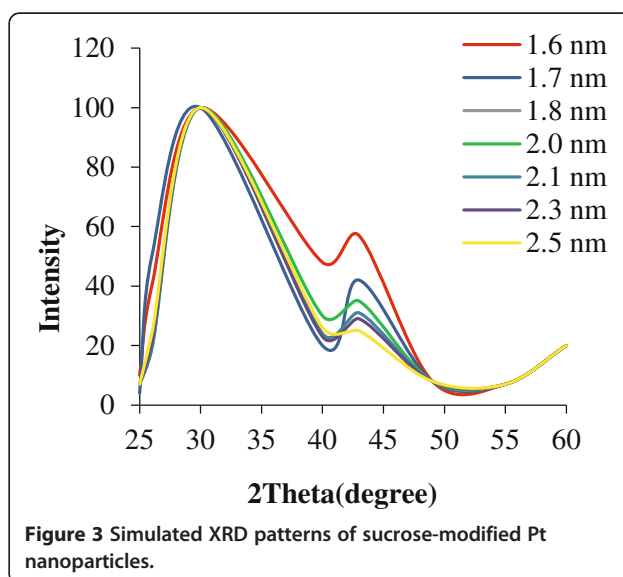
Although the exact mechanism of the formation of the nanostructures is difficult to know, we think that the simple structure of sucrose could serve as a directing template for the growth of Pt nanoparticles. Figure 3 demonstrates the simulated XRD patterns of sucrose-stabilized Pt oxide nanoparticles. All Bragg's reflections representing b111N, b200N, b220N, and b311N planes



of FCC crystal structures due to metallic Pt are observed. About 7% of Pt is present in nanocomposites, and oxygen comes from the surrounding organic matter.

Figure 4 shows the atomic force microscopic image of the sucrose-stabilized Pt oxide nanoparticles. The carbohydrate-stabilized aqueous solution of Pt oxide nanoparticles exhibited antibacterial activity against *P. stutzeri* and *Lactobacillus* bacteria even at concentrations as low as 0.049 mg/L. The photographs in Figures 5 and 6 clearly show the zone of inhibition for carbohydrate-Pt nanocomposites against *P. stutzeri* and *Lactobacillus*, respectively. Similar results were obtained against *Staphylococcus aureus* (data not shown).

The digital photographs in Figure 5a,b represents MIC and MBC tests in Pt oxide-sucrose nanocomposite against *P. stutzeri*, respectively. This figure specifies that MIC/MBC rate is about 0/5.



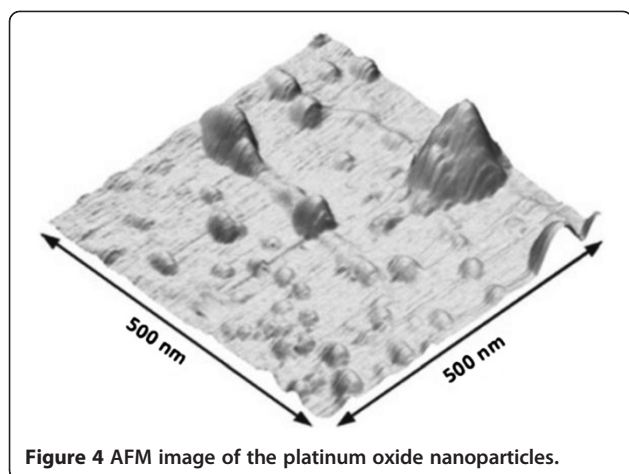


Figure 4 AFM image of the platinum oxide nanoparticles.

On the other hand, Figure 6a,b shows the same parameters against *Lactobacillus* strain. The effectiveness of sucrose nanocomposites was proven to be highly effective. Therefore, it could be concluded that Pt oxide nanoparticles could be released through aqueous carbohydrate solutions owing to the stable dispersion at molecular level and the slow diffusion from the stabilizing medium.

Platinum oxide nanoparticles were obtained by a green approach using sucrose as a reducing as well as stabilizing agent [17-19]. The nanoparticles exhibited interesting morphology when synthesized under hydrothermal conditions in the matrix of sucrose-rich medium. These nanocomposites exhibited high thermal stability as well as bactericidal effect against *P. stutzeri* and *Lactobacillus*. Thus, carbohydrates can serve as a carrier for platinum oxide nanoparticles, and nanocomposites can have potential biological applications [20-22].

Conclusion

The present study reveals the characteristics of antibacterial effect of carbohydrate-stabilized platinum oxide nanoparticles. Given the fact that the nanocomposition of platinum oxide can be used in biomedicine product,

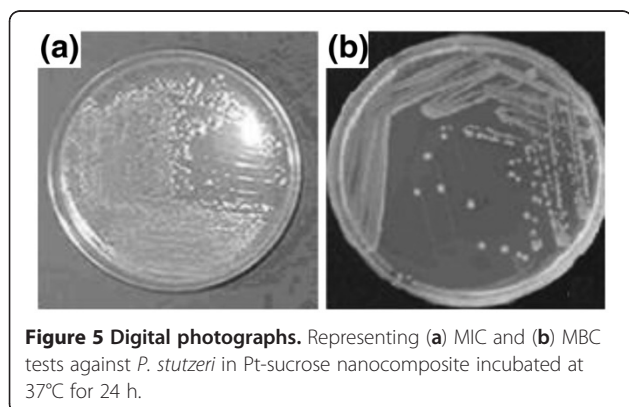


Figure 5 Digital photographs. Representing (a) MIC and (b) MBC tests against *P. stutzeri* in Pt-sucrose nanocomposite incubated at 37°C for 24 h.

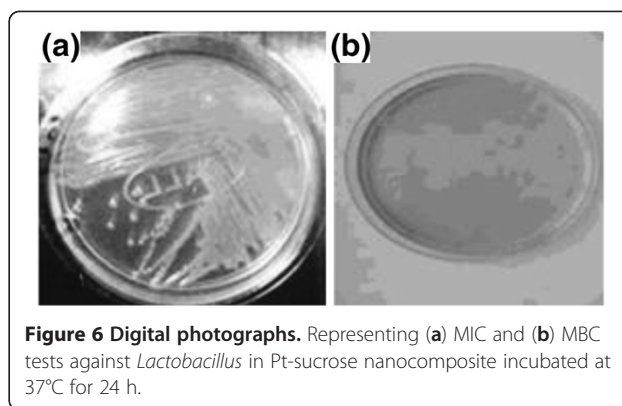


Figure 6 Digital photographs. Representing (a) MIC and (b) MBC tests against *Lactobacillus* in Pt-sucrose nanocomposite incubated at 37°C for 24 h.

nanofilters consequently contained more antibacterial active sites than other antibacterial agents.

Competing interests

The authors declare that they have no competing interests.

Authors' contributions

SRZ and ZZ designed and synthesized the carbohydrate-stabilized platinum oxide nanoparticles, guided the contact angle analysis, and prepared the manuscript. AMZ and SI facilitated and made the antibacterial and biological samples, guided the contact angle analysis, and prepared the manuscript. MS participated in the SEM analysis and prepared the manuscript. All authors read and approved the final manuscript.

Acknowledgments

The authors greatly appreciate the financial support from Payame Noor University, Yazd and Imam Houssein University for running the present project.

Author details

¹Department of Biology, Basic Science Institute, University Street, Payam-e-Noor University, Yazd 89418-57417, Iran. ²Department of Chemical Injury Research Center, Medical Science Institute, Sheikh Bahaei Street, Baqiyatallah Medical Science University, Tehran 14359-44711, Iran. ³Department of Biology, Basic Science Institute, Shahid Bahaei Street, Imam Houssein University, Tehran 479231-98662, Iran. ⁴Department of Applied Biotechnology and Environmental Research Center, Sheikh Bahaei Street, Baqiyatallah Medical Science University, Tehran 14359-44711, Iran. ⁵Department of Biology, Basic Science Institute, Nakhli Street, Payam-e-Noor University, Tehran 19395-4697, Iran.

Received: 3 April 2011 Accepted: 28 February 2012

Published: 24 September 2012

References

1. Shevchenko, EV, Talapin, DV, Kotov, NA, O'Brien, S, Murray, CB: Structural diversity in binary nanoparticle superlattices. *Nature* **439**, 55-59 (2006)
2. Wang, X, Zhuang, J, Peng, Q, Li, YD: A general strategy for nanocrystal synthesis. *Nature* **437**, 121-124 (2005)
3. Portney, NG, Singh, K, Chaudhary, S, Destito, G, Schneemann, A, Manchester, M, Ozkan, M: Organic and inorganic nanoparticle hybrids. *Langmuir* **21**, 2098-2103 (2005)
4. Maye, MM, Lim, IIS, Luo, J, Rab, Z, Rabinovich, D, Liu, TB, Zhong, CJ: Mediator-template assembly of nanoparticles. *J. Am. Chem. Soc.* **127**, 1519-1529 (2005)
5. Chane-Ching, JY, Cobo, F, Aubert, D, Harvey, HG, Airiau, M, Corma, A: A general method for the synthesis of nanostructured large-surface-area materials through the self-assembly of functionalized nanoparticles. *Chem. Eur. J.* **11**, 979-987 (2005)
6. Kim, BS, Hong, DJ, Bae, J, Lee, M: Controlled self-assembly of carbohydrate conjugate rod-coil amphiphiles for supramolecular multivalent ligands. *J. Am. Chem. Soc.* **127**, 16333-16337 (2005)

7. Barrientos, AG, de la Fuente, JM, Rojas, TC, Fernandez, A, Penades, S: Gold glyconanoparticles: synthetic polyvalent ligands mimicking glycocalyx-like surfaces as tools for glycobiological studies. *Chem. Eur. J.* **9**, 1909–1921 (2003)
8. Rojas, TC, de la Fuente, JM, Barrientos, AG, Penades, S, Ponsonnet, L, Fernandez, A: Gold glyconanoparticles as building blocks for nanomaterials design. *Adv. Mater.* **14**, 585–588 (2002)
9. Zand, AM, H Khani, P, Imani, S, Rezaei, M, Rezaei-Zarchi, S: Determining the antibacterial effect of ZnO nanoparticle against the pathogenic bacterium, *Shigella dysenteriae* (type 1). *International Journal of Nano Dimension (IJND)* **1**(4), 279–285 (2010)
10. Reynolds, AJ, Haines, AH, Russell, DA: Gold glyconanoparticles for mimics and measurement of metal ion mediated carbohydrate-carbohydrate interactions. *Langmuir* **22**, 1156–1163 (2006)
11. Halkes, KM, de Carvalho, SA, Elizabeth, C, Maliaars, P, Gerwig, GJ, Kamerling, JP: Facile method for the preparation of gold glyconanoparticles from free oligosaccharides and their applicability in carbohydrate-protein interaction studies. *Eur. J. Org. Chem.* **12**, 3650–3659 (2005)
12. De la Fuente, JM, Eaton, P, Barrientos, AG, Menendez, M, Penades, S: Thermodynamic evidence for Ca²⁺-mediated self-aggregation of Lewis X gold glyconanoparticles. A model for cell adhesion via carbohydrate-carbohydrate interaction. *J. Am. Chem. Soc.* **127**, 6192–6197 (2005)
13. de la Fuente, JM, Barrientos, AG, Rojas, TC, Rojo, J, Canada, J, Fernandez, A, Penades, S: Gold glyconanoparticles as water-soluble polyvalent models to study carbohydrate interactions. *Angew. Chem. Int. Ed.* **40**, 2258–2261 (2001)
14. Ipe, BI, Yoosaf, K, Thomas, KG: Functionalized gold nanoparticles as phosphorescent nanomaterials and sensors. *J. Am. Chem. Soc.* **128**, 1907–1913 (2006)
15. Perez, JM, Josephson, L, Weissleder, R: Use of magnetic nanoparticles as nanosensors to probe for molecular interactions. *Chem. Bio. Chem.* **5**, 261–264 (2004)
16. Quinti, L, Weissleder, R, Tung, CH: A fluorescent nanosensor for apoptotic cells. *Nano. Lett.* **6**, 488–490 (2006)
17. Imani, S, Zagari, Z, Rezaei-Zarchi, S, Zand, AM, Dorodiyani, Bariabarghoyi, H, Lotfi, F: Antibacterial effect of CrO and CoFe₂O₄ nanoparticles upon *Staphylococcus aureus*. **3**, 119–125 (2011)
18. de la Fuente, JM, Penades, S: Types, synthesis and applications in glycoscience, biomedicine and material science. *Biochim. Biophys. Acta* **1760**, 636–651 (2006)
19. Lu, QY, Gao, F, Komarneni, S: A green chemical approach to the synthesis of tellurium nanowires. *Langmuir* **21**, 6002–6005 (2005)
20. Liu, JC, Qin, G, Ravindran, P, Ikushima, Y: Facile “green” synthesis, characterization, and catalytic function of β-d-glucose-stabilized Au nanocrystals. *Chem. Eur. J.* **12**, 2131 (2006)
21. Raghuraman, K, Katti, KK, Barbour, LJ, Pillarsetty, N, Barnes, CL, Katti, KV: Characterization of supramolecular (H₂O)₁₈ water morphology and water-methanol (H₂O)₁₅(CH₃OH)₃ clusters in a novel phosphorus functionalized trimeric amino acid host. *J. Am. Chem. Soc.* **125**, 6955–6961 (2003)
22. Raghuraman, K, Pillarsetty, N, Volkert, WA, Barnes, C, Jurisson, S, Katti, KV: Exceptional kinetic propensity of hydroxymethyl phosphanes toward Rh(III) stabilization in water. *J. Am. Chem. Soc.* **124**, 7276–7277 (2002)

doi:10.1186/2228-5326-2-21

Cite this article as: Rezaei-Zarchi *et al.*: Study of bactericidal properties of carbohydrate-stabilized platinum oxide nanoparticles. *International Nano Letters* 2012 **2**:21.

Submit your manuscript to a SpringerOpen[®] journal and benefit from:

- Convenient online submission
- Rigorous peer review
- Immediate publication on acceptance
- Open access: articles freely available online
- High visibility within the field
- Retaining the copyright to your article

Submit your next manuscript at ► springeropen.com