

REVIEW Open Access

# Premotor biomarkers for Parkinson's disease - a promising direction of research

Brian R Haas, Tessandra H Stewart and Jing Zhang\*

#### **Abstract**

The second most serious neurodegenerative disease is Parkinson's disease (PD). Over the past several decades, a strong body of evidence suggests that PD can begin years before the hallmark clinical motor symptoms appear. Biomarkers for PD are urgently needed to differentiate between neurodegenerative disorders, screen novel therapeutics, and predict eventual clinical PD before the onset of symptoms. Some clinical evaluations and neuroimaging techniques have been developed in the last several years with some success in this area. Moreover, other strategies have been utilized to identify biochemical and genetic markers associated with PD leading to the examination of PD progression and pathogenesis in cerebrospinal fluid, blood, or saliva. Finally, interesting results are surfacing from preliminary studies using known PD-associated genetic mutations to assess potential premotor PD biomarkers. The current review highlights recent advances and underscores areas of potential advancement.

**Keywords:** Parkinson's disease, Biochemical markers, *LRRK2*, *GBA*, α-synuclein, DJ-1, Clinical biomarkers, Premotor, Neuroimaging, *parkin*, CSF

# Introduction

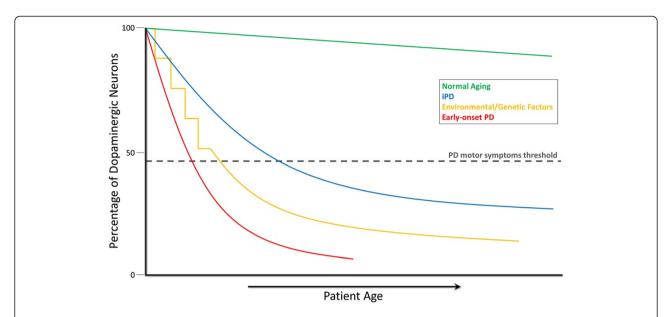
Parkinson's Disease (PD) is a common and incapacitating neurodegenerative disease which affects the human population worldwide [1-3]. It generally presents clinically in adults over 60 years old, and major symptoms include akinesia, tremor, bradykinesia, and postural instability. Currently, its diagnosis relies largely on assessment of clinical symptoms. Unfortunately, this purely clinical diagnosis, combined with the similarities of PD symptoms with those of other neurodegenerative and movement disorders, results in frequent misdiagnosis: up to 50% of the time in primary care settings, while movement specialists fare somewhat better with an early PD misdiagnosis rate around 10% [4-6]. PD can currently only be confirmed through its pathological hallmark of Lewy bodies and Lewy neurites located in residual neurons or axons, respecitively, upon postmortem analysis [7]. To complicate matters, diagnosis is generally not made until an advanced stage of neurodegeneration: by some estimates, 50-60% of dopaminergic neurons in the substatia nigra are lost prior to the time of clinical diageffectiveness nosis [8-13],limiting

neuroprotective therapies due to the low number of remaining neurons (Figure 1).

Thus, introduction of new treatments and interventions for PD will depend on improved diagnosis, preferably at early, premotor stages at which a larger population of dopaminergic cells remains. Furthermore, the progression of the disease must be accurately assessed, in order to monitor efficacy of treatments and improve individualized therapeutic strategies. Much work toward this end has focused on identifying sensitive, specific biomarkers for PD. According to the NIH biomarker working group, a biomarker is "a characteristic objectively measured and evaluated as an indicator of normal biological processes, pathogenic processes or pharmacological responses to a therapeutic intervention"[14,15]. In other words, an ideal biomarker for PD must be able to identify almost all cases of PD in its premotor stages, change concomitantly with disease stages, and differentiate between PD and other neurodegenerative diseases. In practice, however, a combination of several biomarkers will likely be needed to correctly diagnose and track PD progression, and this review focuses on some of the most promising premotor PD biomarkers which could be most effective.

<sup>\*</sup> Correspondence: zhangj@u.washington.edu Department of Pathology, University of Washington School of Medicine, HMC Box 359635, 325 9th Avenue, Seattle, WA98104, USA





**Figure 1 Evolution of PD.** During the course of normal aging (green line), small but slow dopaminergic degeneration occurs without any motor symptoms. Typically idiopathic PD (iPD) (blue line) is of unknown origin but is thought to develop gradually over time with a slow degeneration of dopaminergic neurons leading to the emergence of the classic PD motor symptoms later in life. Another model of dopamingeric neurodegeneration leading to PD motor symptoms involves repeated exposure to environmental toxicants over time in combination with a genetic predisposition to dopaminergic neuron loss (yellow line). Early-onset PD (red line), as caused by mutations in the *parkin* gene, involves a precipitous decline in dopaminergic neuron number where PD motor symptoms can present decades prior to those in iPD. One more scenario (not shown) of PD-motor symptom development involves possible *in utero* environmental toxicants or genetic factors leading to an atypically low number of dopaminergic neurons at birth and increased susceptibility to PD development.

# Potential premotor PD biomarker candidates Clinical biomarkers

It has been recognized recently that a variety of nonmotor symptoms associated with PD may manifest years before the onset of motor symptoms. Indeed, these premotor symptoms have been exploited in the last few years as possible premotor biomarkers for PD. Though not an exhaustive list, we focus here on some of the most promising potential clinical premotor PD biomarkers, namely, rapid eye movement (REM) behavior disorder (RBD), bowel dysfunction, olfactory deficits, and mood disorders.

It has been long observed that PD patients can have a variety of sleep disturbances, most frequently manifesting as RBD, which can precede the clinical motor symptoms associated with PD by several years [4,16-20]. In fact, recent findings indicate that RBD is one of the symptoms most strongly correlated with the presentation of synucleinopathy (including multiple system atrophy (MSA), PD, and dementia with Lewy bodies) later in life [19,21].

Disruptions in bowel movements are another known co-morbidity to PD. Much like RBD, constipation can arise many years prior to the motor symptoms of PD [22] and affects most PD patients [23]. Indeed, the frequency of bowel movements was correlated inversely with PD risk, and constipation may be one of the first

symptoms of PD [22]. These findings were supported by an additional epidemiological study involving women in Olsmsted County, Minnesota, where an association between earlier life constipation and PD risk was well documented [24]. Together, these reports document the association of PD diagnosis with constipation symptoms emerging as much as 20 years earlier.

Olfactory disruption is also a common symptom associated with PD. Difficulties in detecting, discriminating, and identifying odors are observed in up to 90% of PD patients [25-27]. However, cross-sectional and longitudinal studies of PD patients have frequently found olfactory deficits to be largely uncorrelated with disease stage or severity [28-31], leading to the hypothesis that olfactory decline may occur largely prior to the onset of nonmotor symptoms. Indeed, as with constipation and sleep disturbances, olfactory deficits can occur in the several years leading up to motor impairments and clinical PD diagnosis [25]. This idea is further supported by the finding that, in recently diagnosed PD patients with mild symptoms, severity of olfactory deficits has been found to correlate with dopaminergic dysfunction as measured by SPECT, despite a lack of correlation between either measurement and severity of motor dysfunction [32-34]. To determine whether these early olfactory symptoms might prove useful as a premotor biomarker for PD, a number of studies have examined olfaction in early or

asymptomatic patients. A recent prospective study has shown that patients with poor olfaction were more likely to develop PD in the 4-year follow up period than those with normal olfaction [35]. Additionally, another study found that when assessing asymptomatic first-degree relatives of PD patients, performance levels of odor discrimination robustly correlated with future PD risk [36]. Couple these studies with an initial report revealing that olfactory identification dysfunction on the University of Pennsylvania Smell Identification Test was able to distinguish PD from other movement disorders [26], and a strong candidate for a clinical premotor PD biomarker emerges.

Another potential clinical premotor PD biomarker is the presence of mood alterations. Some of the most common psychological symptoms associated with PD are depression, anxiety, and apathy [37]. Depression specifically is thought to be biological component of the disease (as opposed to a psychological reaction to the disease) as it is detected in nearly a quarter of patients in the early stages of PD [38,39]. Several groups have indicated that in the later stages of PD, depression rates are approximately 40%, but that this rate is probably grossly underestimated [40,41]. Further, examination of historical medical records at least 20 years before PD diagnosis led to the discovery that anxiety was associated with the later incidence of PD [42]. A few different groups have also speculated about a possible parkinsonian personality that involved such traits as inflexibility, neuroticism, obsessive-compulsivity, uneasiness, and anxiety, among others [43-47]. These characteristics are often present from childhood, leading to the possibility of long lasting biological changes preceding PD motor symptoms, and possibly implying that the observed motor symptoms of PD are really characteristic of endstage disease. Supporting the idea that these symptoms are a component of the disease are findings of alterations in the serotonergic signaling pathways, long known to interact with the classic dopaminergic neuronal degeneration associated with PD and major target of anti-depressant medications. For example, a major serotonergic nucleus, the dorsal raphe, has enhanced cell loss in depressed PD patients compared to non-depressed PD patients [48]. Moreover, recent studies have shown a reduction of gray matter densities in the orbitofrontal cortex which are known to be involved in serotonergic pathways responsible for mood disorders [49,50]. Lewy body-related lesions also have been found in raphe nucleus of early stage PD patients and in asymptomatic individuals [51,52], further implicating a serotonergic pathology in PD depression. However, as with RBD, constipation, and anosmia, anxiety and depression are very nonspecific, and do not stand alone as clinical premotor PD biomarkers.

In short, a number of clinical symptoms appear years before the motor symptoms of PD. However, while each of the biomarkers occurs during premotor stages of PD, can be measured, and tracked, none is specific enough to be used alone as a premotor PD biomarker. As research progresses, the most likely role of these clinical premotor biomarkers will be a supportive one, serving to confirm the sensitivity and specificity of other premotor PD biomarkers. Future work should also focus on using a combination of clinical premotor biomarkers to enrich patient populations for neuroimaging and biochemical screening and assess in tandem multiple clinical symptoms for correlation for developing PD later in life, along with coupling premotor clinical signs with biochemical markers, neuroimaging, and genetic testing (see below for more discussion on the topic).

#### **Biochemical** markers

Examination of various tissues and body fluids for the relative levels of proteins and other molecules provides information about the state of the local system, including its metabolic state, disease condition, and response to therapeutic treatments. Therefore, measurement of a number of known small proteins and other molecules along with unbiased profiling (recently reviewed elsewhere [53,54]) is currently one of the most promising avenues for potential premotor biomarkers of PD (Table 1).

**a.** α-Synuclein α-synuclein has long been known to form protein aggregates found in Lewy bodies, and has been a major area of inquiry for PD treatments and biomarker discovery. Indeed, a consensus result of measuring the different forms of the protein that accumulate in cerebrospinal fluid (CSF), a body fluid in close proximity and interaction with the central nervous system (CNS), has started to emerge. Using both ELISA and Luminex assays, several groups have examined α-synuclein in CSF samples. These studies, which included large cohorts of patients, have shown a reduction of  $\alpha$ -synuclein levels in CSF of PD patients offering a reasonable differential sensitivity and specificity compared to healthy controls [55,56]. However, there was no correlation between disease severity/duration and α-synuclein levels in these investigations, in contrast to a previous report which also detected reduced α-synuclein in the CSF of PD patients [57]. Notably, in smaller scale investigations, while several groups have also confirmed these observations [56,58,59], others reported no discernible differences in α-synuclein levels between control and PD patients [60,61]. Many issues are potentially responsible for these discrepancies, including sampling protocols, control for blood contamination (see below) and differences in detection/capture antibodies [62].

Because collection of CSF is somewhat invasive, multiple groups have investigated  $\alpha$ -synuclein levels in other tissues as PD biomarkers. Due to its ready accessibility,

Table 1 Summary of possible premotor PD biomarkers

	Existing Biomakers			Potential Biomakers			Candidate biomakers	
	Uric acid	a-sysnuclein	DJ-1	SPECT	fMRI	PET	Αβ42	tau
Blood	74–76	63	82					
CSF		53, 54, 56, 59	53,79				130	130
Saliva		69,71						
Skin		67						
Glucose Binding						92		
Iron					100			
Structural Brain Elements					97–99			
DAT Binding				88-90, 93, 94				

blood has been the subject of a number of studies, but the results have been largely contradictory and inconclusive [55,63-66]. A recent bright spot appears to be the role of the concentrations of oligomeric α-synuclein in plasma samples. El-Agnaf et al. discovered that the concentration of oligomeric forms was increased in PD patients compared to controls [65]. They achieved high specificity (85%) for the differentiation of PD patients versus controls despite significant overlap of concentrations with the control group [65]. Another group has recently demonstrated significantly higher levels of autoantibodies towards monomeric  $\alpha$ -synuclein in the blood sera of PD patients compared to those from the control group [67]. However, measurement of α-synuclein in blood remains a significant problem, since >98% blood based α-synuclein is contained in red blood cells, making hemolysis, in vitro or in vivo, a major confounder in these studies [55,68]. Therefore, potential biomarkers based on blood α-synuclein remain either disappointing or too preliminary to be clinically useful.

Several investigators are pursuing  $\alpha$ -synuclein levels in other tissue types as potential premotor PD biomarkers. For example, one initial report has detected  $\alpha$ -synuclein in skin fibroblasts [69]. This group demonstrated that  $\alpha$ -synuclein levels are elevated in the skin fibroblasts of PD patients compared to controls [69]. Although this study involved a small number of subjects and has yet to be replicated, using skin as a premotor biomarker has potentially high value and utility as it is highly accessible and can be procured without more invasive procedures.

Another promising line of evidence has emerged for using  $\alpha$ -synuclein as a premotor PD biomarker in human saliva. Abnormal saliva production has been a long shown to correlate with PD severity and known to be a prevalent nonmotor symptom [70]. Del Tredici *et al.* recently examined post-mortem PD, MSA, and control submandibular glands for the presence of aggregated  $\alpha$ -synuclein [71]. They discovered the presence of Lewy Body pathology by immunohistochemistry in the submandibular glands of all the PD cases examined, but not

the control or MSA cases [71] . Because the submandibular glands respond to parasympathetic innervations from the medulla [72], the cells responsible for saliva secretion possess direct contact with the CNS. Therefore, any saliva secreted from these glands may provide indirect access to α-synuclein levels of the CNS. Saliva, as a biological fluid, is completely accessible, can be obtained without any invasive procedures, is readily available in large quantities, and can be sampled virtually an unlimited number of times from any individual. For these reasons, we are currently investigating  $\alpha$ -synuclein in saliva as a biomarker. We are able to detect  $\alpha$ -synuclein levels in human saliva [73] and are investigating whether levels of salivary α-synuclein differ between PD and healthy controls. Clearly, this promising line of research will require independent validation from several groups before its adaptation as a clinically useful biomarker.

**b.** Anti-oxidative stress makers Besides α-synuclein, another major biochemical pathway dysregulated in PD is the elevation of oxidative stress [74]. Accordingly, several groups have explored oxidative stress metabolites, such as uric acid [75], and proteins as PD biomarkers. Three independent, large studies have indicated that higher levels of uric acid in serum confer lower risk of developing PD [76-78]. Additionally, the subset of diagnosed PD patients with higher uric acid levels progressed more slowly [79]. Higher uric acid levels however, though correlated with decreased risk [79], do not provide enough information to be pursued further as a premotor PD biomarker. This is mostly due to the largely non-specific action of uric acid as an anti-oxidant. Uric acid levels are tightly controlled by the body and, most importantly, while decreased risk may correlate with higher levels of uric acid, PD can still develop despite higher uric acid levels.

Similarly, the protein DJ-1 is expressed at high concentrations in cells and is thought to act as an antioxidant [80,81] and interacts with many of the same proteins as  $\alpha$ -synuclein [82]. An initial study showed that DJ-1 is

secreted into the plasma of both normal and PD patients [83]. However, no significant differences between PD and control groups were observed, nor was any correlation with age. This initial study set the stage for future studies examining the levels of DJ-1 in CSF and blood plasma. Waragai et al. demonstrated several years ago that DJ-1 levels are increased in the plasma of PD patients [84]. The same group also showed an increase of DJ-1 in the CSF of PD patients compared to control patients [85]. However, a more recent study has examined the levels of DJ-1 in PD patient CSF and has come to the opposite conclusion. It found that DJ-1 levels actually significantly decreased in PD patient CSF using a highly sensitive Luminex assay while at the same time controlling for possible blood contamination of CSF, which can confound interpretation and reliability of results [55]. The same group of investigators also concluded that in the same cohort of patients and controls, while CSF DJ-1 levels decreased, plasma DJ-1 was largely unaltered [55]. Much like α-synuclein, the discrepancy among different investigations is attributable to many issues, particularly hemolysis [68]. As oxidized isoforms of DJ-1 are indicative of increased oxidative stress [86-88], moving forward, they should be explored along with total DJ-1 and other promising biomarkers, in various body fluids, especially in the saliva of PD patients [73] or those with premotor signs, thereby facilitating the development of premotor biomarkers.

# Neuroimaging

As technology has advanced in the past several decades, it has allowed clinicians and researchers to acquire more reliable and higher resolution data. Several techniques are being investigated for their use as potential premotor biomarkers [89]. Here, we will examine single-photon emission computed tomography (SPECT), positron emission tomography (PET), and functional magnetic resonance imaging (fMRI) (Table 1).

SPECT has been used for some time to image the dopamine transporter (DAT). It has routinely established a reduced binding in patients with PD [90] and can aid clinicians in diagnosis and monitoring, as the decreased ligand binding to the DAT correlates with disease progression and clinical scoring [91,92]. Although long term pre-symptomatic screening with DAT SPECT for PD is not particularly feasible due to high costs, limited access, and general difficulty of working with radioactive tracers, one group has shown it is useful for distinguishing between PD and other parkinsonian-like conditions [93]. Additionally, some PET imaging has shown reduced binding of glucose in posterior parietal and cortical gray matter of PD patients with dementia [94]. PET can also be used as a highly sensitive measure to assess dopamine uptake and binding to the DAT. Accordingly, PET

imaging, which assesses radiolabeled ligands for DAT, can be utilized to distinguish premotor PD [95,96]. Additionally, fMRI is another potential neuroimaging modality for use as a premotor biomarker for PD. It has been known almost a decade that whole brain atrophy is precipitated in PD versus controls and slightly less than in Alzheimer's disease [97]. More recent work has demonstrated that irregularities in the substantia nigra of early PD patients correlate with an increase in iron, which is a known to be related to neuronal loss [98]. In addition, one group has been able to provide 100% sensitivity and specificity for differentiating PD patients from healthy controls when imaging the caudal substantia nigra [99]. Another recent work has arrived at a 95% accuracy in distinguishing PD from healthy controls using multimodal MRI of subcortical gray matter [100]. While fMRI can successfully separate PD patients from controls, further volumetric fMRI characterization of PD and related disorders is currently underway and needed to provide insight into whether fMRI can serve as an effective premotor biomarker [101].

The neuroimaging markers reviewed all are able distinguish diseased PD patients from normal controls and have the ability in some situations to identify probable PD in premotor stages. However, their usefulness is limited due to issues of cost and availability, and, while they may detect abnormalities associated with disease state, they do not provide insight into molecular mechanism. Their cost alone limits the ability of these tests to be used widespread in a clinical diagnosis. Finally, the time involved to acquire these neuroimages for both clinical staff and patients is significant and cannot be overlooked when considering potential premotor PD biomarkers. Thus, it is likely that the structural/functional neuroimaging methods will be largely a research tool in the area of biomarkers in the near future.

## Genetic markers

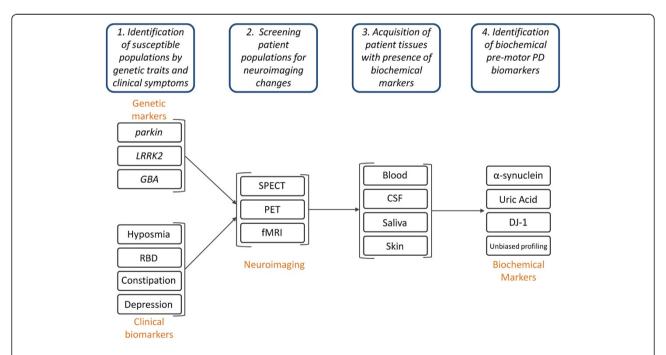
Clearly, identification of premotor biomarkers first requires identification of the population at risk. One obvious choice is those with premotor signs, e.g. RBD and constipation, an option which is, as previously discussed, limited by the non-specific nature of these symptoms prior to the onset of motor symptoms. Another possibility is to follow those with genetic mutations that lead to familial PD. Thus, in the following section, we will address the genetic mutations with a reasonable prevalence for the purpose of premotor biomarker studies: *parkin*, glucocerebrosidase (*GBA*), and leucine-rich repeat kinase-2 (*LRRK2*).

While most idiopathic PD arises *de novo* and without any known genetic mutation, the known genetic aberrations which cause familial PD are invaluable in aiding investigators to elucidate the pathways contributing to

clinical diagnosis and the study of preclinical PD biomarkers. Mutations in the *parkin* gene, which encodes a ubiquitin E3 ligase, results in enzymatic activity loss and misfolding [102] and is responsible for the majority of juvenile, early-onset PD [103]. *Parkin* gene mutations may serve as potential premotor biomarker as detecting this mutation in the pre-symptomatic stages of the disease will allow for the evaluation of relevant biomarkers in relation to early-onset PD [104].

Another genetic mutation that has been associated with PD is the gene encoding GBA. Mutations in GBA cause Gaucher disease, which is the most common autosomal recessive lysosomal storage disease [105], and its relationship with parkinsonism has been reviewed extensively elsewhere [106]. PD and PD-like syndromes associated with GBA mutations were first described in 1996 [107], and have subsequently been demonstrated to occur at much higher frequencies in PD patients than controls in pan-ethnic populations from Israel [108,109], Italy [110], China [111,112], North Africa [113], Great Britain [114], Europe [115], and the United States [116,117]. In fact, a large 16-center, multi-country analysis, 15% of Ashkenazi Jewish PD patients had one of two of the most common GBA mutations compared to 3% of controls [118]. Furthermore, in almost 2000 nonJewish PD patients, 7% tested positive for GBA mutations after full sequencing of the gene and concluded that patients with GBA mutations present with an earlier onset of PD symptoms [118]. Most promisingly, these pan-ethnic population studies are just beginning to be supported by mechanistic studies. For example, one group recently demonstrated that in PD patients with GBA mutations, an average of 75% of Lewy Bodies examined were positive for GBA [119]. Others have shown that GBA mutants can raise the levels of  $\alpha$ -synuclein in vitro and that the amassing of  $\alpha$ -synuclein was reversed by activation of autophagy pathways [120]. Moreover, α-synuclein and *GBA's* interaction takes place optimally under pH conditions like those in the lysosome [121], thereby further linking PD pathology with to GBA function. Thus, GBA mutations represent an increased risk for PD development and add additional evidence to a genetic component to the disease.

Mutations in another gene, *LRRK2*, are probably among the most studied. In contrast to *parkin* mutations, *LRRK2* mutations result in an autosomal dominant form of PD that closely resembles idiopathic PD. *LRRK2* mutations account for about 1-2% of sporadic PD cases worldwide, but in genetically isolated populations, such as Ashkenazi Jews and North African Arabs, the



**Figure 2 The Search for Premotor PD Biomarkers.** 1. In order to discover premotor PD biomarkers, it is necessary to first enrich for patient population more likely to develop PD. Patients with genetic susceptibility markers, for example, *parkin* and *LRRK2* mutations, and clinical biomarkers, hyposmia, RBD, constipation, and depression. 2. Patients with genetic markers and clinical phenotypes will undergo screening for neuroimaging changes known to correlate with PD. 3. After verification with neuroimaging, various patient tissues where the presence of biochemical markers has already been verified will be acquired (blood, CSF, saliva, and skin). 4. Premotor PD biomarker assessment and identification will use changes in the levels of known biochemical markers, i.e., α-synuclein, uric acid, DJ-1, in addition to others identified through unbiased profiling.

mutations can account for upwards of 30-40% of sporadic and familial PD cases [122-127]. The most common mutation is G2019S, which produces a constitutively active kinase [128]. Recent evidence points to LRRK2 involvement in dopaminergic neuron transmission further linking the gene to PD [129]. Additionally, another recent study examined the downstream targets of LRRK2 and found that  $\alpha$ -synuclein is phosphorylated by *LRRK2*, further implicating α-synuclein in both familial and sporadic PD [130]. This led our group to examine a cohort of symptomatic and asymptomatic LRRK2 mutation carriers. Because of the high conversion rate of LRRK2 mutation carriers to clinical disease, this patient population presents a unique subset of PD patients in which evaluation of symptoms and biomarkers can occur decades before clinical signs appear. One of our recent studies attempted to correlate dopaminergic function with both DJ-1 and α-synuclein levels in LRRK2 mutation carrier CSF. The major finding of this study was that no apparent relationship existed between dopaminergic loss and levels of DJ-1 and α-synuclein in patient CSF [131]. Another recent publication resulting from the same patient cohort [131] investigated LRRK2 mutation carriers for the levels of amyloid  $\beta$ -42 (A $\beta$ <sub>42</sub>) and tau in patient CSF [132]. We found that LRRK2 mutation carriers showed reduced CSF  $A\beta_{42}$  and tau levels that correlated with lower dopaminergic neuron function as determined by PET tracer [132]. These findings elucidate the possible role DJ-1,  $\alpha$ -synuclein, A $\beta_{42}$ , and tau in premotor PD and provide insight into which direction future premotor PD biomarker studies might take.

Other researchers have been able to differentiate between LRRK2 PD patients and their healthy family members with or without LRRK2 mutations using metabolomic profiling of LRRK2-PD, LRRK2-mutation carriers, control, and idiopathic PD patients [133]. Another group was also able to distinguish between asymptomatic carriers of LRRK2 mutations and healthy carriers by measuring gait alterations [134]. They found that there were significant differences in gait variability measures under three different gait conditions [134] leaving open the possibility that motor symptoms might appear earlier than previously thought. Taken together, these genetic populations that have a high conversion rate from asymptomatic to PD provide a unique cohort of patients to help validate other biomarker discoveries. As these patients can easily be followed in a longitudinal fashion, examining clinical presentation along with biological fluids and assessing them for PD biomarkers will be key in developing our understanding of biomarkers, disease progression, and possible disease interventions (Figure 2). Structural and functional neuroimaging methods will be certainly helpful in determining whether an asymptomatic subject is at premotor stage or not.

In addition to LRRK2 and parkin mutations, the usefulness of having a genetic marker as an a priori identifier of at-risk populations has led to attempts to identify other loci associated with PD. Several groups have completed independent genome-wide association studies for PD but have had inconsistent results [135-137]. However, two loci previously associated with neurodegenerative disease, SNCA (encoding α-synuclein) and MAPT (encoding tau), have been reproducibly implicated in several studies, strongly suggesting a role for both in PD pathogenesis [138-143]. However, PD generation appears to be a complex process requiring the interactions of multiple genetic and environmental factors, so the identification of high-risk groups, prior to the emergence of motor symptoms, remains a significant problem, until a robust premotor biomarker can be found in an easily accessible sample source.

#### **Conclusions**

The need to develop premotor PD biomarkers is increasingly apparent as the aging population grows and considering the limited effectiveness of neuroprotective treatments in altering disease course at the advanced stages. The clinical biomarkers have been well-characterized and correlated with PD outcomes and severity but tend to lack specificity and sensitivity. Neuroimaging, however, can be highly sensitive and specific for premotor PD but significant cost and access barriers exist. The most promising avenue for future work involves both genetic markers and biochemical markers, and currently, efforts are underway at several sites worldwide, including the Michael J. Fox Foundation (http:// www.michaelifox.org/research MJFFresearchTools.cfm) and the Parkinson's UK Brain Bank (www.parkinsons.org.uk/), to collect and utilize PD tissues to this end [144]. Because some genetically susceptible populations, such as LRRK2 or GBA mutation carriers, possess known PD-associated mutations, they can be employed to validate and screen potential biochemical markers underutilized biological samples, such as saliva. In short, the best outlook for premotor PD biomarkers relies heavily on the rapid and reliable identification of premotor biomarkers with independent validation of biochemical markers the clinical cohort most likely to develop PD.

# **Competing interests**

The authors declare no competing interests.

#### Acknowledgements

This work was supported by NIH grants AG033398, ES004696-5897, ES007033-6364, ES016873, ES019277, NS057567, and NS062684-6221.

### Authors' contributions

BRH carried out literature searches, reviewed and synthesized current literature, and organized, drafted, and revised the manuscript. THS helped draft and critically revise the manuscript. JZ helped critically revise and gave final approval to the manuscript.

Received: 10 February 2012 Accepted: 31 May 2012 Published: 31 May 2012

#### References

- Thomas B, Beal MF: Parkinson's disease. Hum Mol Genet 2007, 16:R183–194. Spec No. 2. England.
- Cookson MR: The biochemistry of Parkinson's disease. Annu Rev Biochem 2005, 74:29–52.
- Robinson P: Understanding the molecular basis of Parkinson's disease, identification of biomarkers and routes to therapy. Expert Rev Proteomics 2010. 7:565–578.
- Morgan JC, Mehta SH, Sethi KD: Biomarkers in Parkinson's disease. Curr Neurol Neurosci Rep 2010, 10:423–430.
- Fahn S, Oakes D, Shoulson I, Kieburtz K, Rudolph A, Lang A, Olanow CW, Tanner C, Marek K: Levodopa and the progression of Parkinson's disease. In N Engl J Med 2004, 351:2498–2508. United States: 2004 Massachusetts Medical Society.
- Meara J, Bhowmick BK, Hobson P: Accuracy of diagnosis in patients with presumed Parkinson's disease. Age Ageing 1999, 28:99–102.
- 7. Shults CW: Lewy bodies. Proc Natl Acad Sci USA 2006, 103:1661–1668.
- Fearnley JM, Lees AJ: Ageing and Parkinson's disease: substantia nigra regional selectivity. Brain 1991, 114(Pt 5):2283–2301.
- Bohnen NI, Albin RL, Koeppe RA, Wernette KA, Kilbourn MR, Minoshima S, Frey KA: Positron emission tomography of monoaminergic vesicular binding in aging and Parkinson disease. J Cereb Blood Flow Metab 2006, 26:1198–1212.
- Ferrer I, Martinez A, Blanco R, Dalfó E, Carmona M: Neuropathology of sporadic Parkinson disease before the appearance of parkinsonism: preclinical Parkinson disease. J Neural Transm 2011, 118:821–839.
- Bernheimer H, Birkmayer W, Hornykiewicz O, Jellinger K, Seitelberger F: Brain dopamine and the syndromes of Parkinson and Huntington. Clinical, morphological and neurochemical correlations. J Neurol Sci 1973, 20:415–455.
- Kish SJ, Shannak K, Hornykiewicz O: Uneven pattern of dopamine loss in the striatum of patients with idiopathic Parkinson's disease. Pathophysiologic and clinical implications. N Engl J Med 1988, 318:876–880.
- Hornykiewicz O: Biochemical aspects of Parkinson's disease. Neurology 1998, 51:S2–9.
- 14. GJDowningNational Institutes of Health (U.S.), United States2000Food and Drug Administration: Biomarkers and surrogate endpoints: clinical research and applications: proceedings of the NIH-FDA conference held on 15-16 April 1999 in Bethesda, Maryland, USA1ElsevierAmsterdamDowning GJ, National Institutes of Health (U.S.), United States: Food and Drug Administration: Biomarkers and surrogate endpoints: clinical research and applications: proceedings of the NIH-FDA conference held on 15-16 April 1999 in Bethesda, Maryland, USA. 1st edition. Amsterdam: Elsevier, 2000.
- Breen DP, Michell AW, Barker RA: Parkinson's disease the continuing search for biomarkers. Clin Chem Lab Med 2011, 49:393–401.
- Olson EJ, Boeve BF, Silber MH: Rapid eye movement sleep behaviour disorder: demographic, clinical and laboratory findings in 93 cases. *Brain* 2000, 123(Pt 2):331–339.
- Savica R, Rocca WA, Ahlskog JE: When does Parkinson disease start? Arch Neurol 2010, 67:798–801.
- Monderer R, Thorpy M: Sleep disorders and daytime sleepiness in Parkinson's disease. Curr Neurol Neurosci Rep 2009, 9:173–180.
- Postuma RB, Gagnon JF, Vendette M, Fantini ML, Massicotte-Marquez J, Montplaisir J: Quantifying the risk of neurodegenerative disease in idiopathic REM sleep behavior disorder. Neurology 2009, 72:1296–1300.
- Wu Y, Le W, Jankovic J: Preclinical biomarkers of Parkinson disease. Arch Neurol 2011, 68:22–30.
- Postuma RB, Gagnon JF, Rompré S, Montplaisir JY: Severity of REM atonia loss in idiopathic REM sleep behavior disorder predicts Parkinson disease. Neurology 2010, 74:239–244.
- Mostile G, Jankovic J: Treatment of dysautonomia associated with Parkinson's disease. Parkinsonism Relat Disord 2009, 15(Suppl 3):S224–232.
- Abbott RD, Petrovitch H, White LR, Masaki KH, Tanner CM, Curb JD, Grandinetti A, Blanchette PL, Popper JS, Ross GW: Frequency of bowel movements and the future risk of Parkinson's disease. Neurology 2001, 57:456–462.

- Savica R, Carlin JM, Grossardt BR, Bower JH, Ahlskog JE, Maraganore DM, Bharucha AE, Rocca WA: Medical records documentation of constipation preceding Parkinson disease: A case-control study. Neurology 2009, 73:1752–1758
- Ponsen MM, Stoffers D, Booij J, van Eck-Smit BL, Wolters ECh, Berendse HW: Idiopathic hyposmia as a preclinical sign of Parkinson's disease. Ann Neurol 2004, 56:173–181.
- Doty RL, Bromley SM, Stern MB: Olfactory testing as an aid in the diagnosis of Parkinson's disease: development of optimal discrimination criteria. Neurodegeneration 1995, 4:93–97.
- Silveira-Moriyama L, Petrie A, Williams DR, Evans A, Katzenschlager R, Barbosa ER, Lees AJ: The use of a color coded probability scale to interpret smell tests in suspected parkinsonism. Mov Disord 2009, 24:1144–1153.
- Müller A, Reichmann H, Livermore A, Hummel T: Olfactory function in idiopathic Parkinson's disease (IPD): results from cross-sectional studies in IPD patients and long-term follow-up of de-novo IPD patients. J Neural Transm 2002, 109:805–811.
- Herting B, Schulze S, Reichmann H, Haehner A, Hummel T: A longitudinal study of olfactory function in patients with idiopathic Parkinson's disease. J Neurol 2008. 255:367–370.
- McKinnon J, Evidente V, Driver-Dunckley E, Premkumar A, Hentz J, Shill H, Sabbagh M, Caviness J, Connor D, Adler C: Olfaction in the elderly: a crosssectional analysis comparing Parkinson's disease with controls and other disorders. Int J Neurosci 2010, 120:36–39.
- Oka H, Toyoda C, Yogo M, Mochio S: Olfactory dysfunction and cardiovascular dysautonomia in Parkinson's disease. J Neurol 2010, 257:969–976.
- Ponsen MM, Stoffers D, Wolters ECh, Booij J, Berendse HW: Olfactory testing combined with dopamine transporter imaging as a method to detect prodromal Parkinson's disease. J Neurol Neurosurg Psychiatry 2010, 81:396–399
- Siderowf A, Newberg A, Chou KL, Lloyd M, Colcher A, Hurtig HI, Stern MB, Doty RL, Mozley PD, Wintering N, et al: [99mTc]TRODAT-1 SPECT imaging correlates with odor identification in early Parkinson disease. Neurology 2005, 64:1716–1720.
- Deeb J, Shah M, Muhammed N, Gunasekera R, Gannon K, Findley LJ, Hawkes CH: A basic smell test is as sensitive as a dopamine transporter scan: comparison of olfaction, taste and DaTSCAN in the diagnosis of Parkinson's disease. QJM 2010, 103:941–952.
- Ross GW, Petrovitch H, Abbott RD, Tanner CM, Popper J, Masaki K, Launer L, White LR: Association of olfactory dysfunction with risk for future Parkinson's disease. Ann Neurol 2008, 63:167–173.
- Berendse HW, Ponsen MM: Diagnosing premotor Parkinson's disease using a two-step approach combining olfactory testing and DAT SPECT imaging. Parkinsonism Relat Disord 2009, 15(Suppl 3):S26–30.
- Aarsland D, Brønnick K, Alves G, Tysnes OB, Pedersen KF, Ehrt U, Larsen JP: The spectrum of neuropsychiatric symptoms in patients with early untreated Parkinson's disease. J Neurol Neurosurg Psychiatry 2009, 80:928–930.
- Ravina B, Camicioli R, Como PG, Marsh L, Jankovic J, Weintraub D, Elm J: The impact of depressive symptoms in early Parkinson disease. *Neurology* 2007, 69:342–347.
- Ravina B, Elm J, Camicioli R, Como PG, Marsh L, Jankovic J, Weintraub D: The course of depressive symptoms in early Parkinson's disease. Mov Disord 2009, 24:1306–1311.
- Dooneief G, Mirabello E, Bell K, Marder K, Stern Y, Mayeux R: An estimate of the incidence of depression in idiopathic Parkinson's disease. Arch Neurol 1992, 49:305–307.
- Tandberg E, Larsen JP, Aarsland D, Cummings JL: The occurrence of depression in Parkinson's disease. A community-based study. Arch Neurol 1996. 53:175–179.
- Shiba M, Bower JH, Maraganore DM, McDonnell SK, Peterson BJ, Ahlskog JE, Schaid DJ, Rocca WA: Anxiety disorders and depressive disorders preceding Parkinson's disease: a case-control study. Mov Disord 2000, 15:669–677.
- Bower JH, Grossardt BR, Maraganore DM, Ahlskog JE, Colligan RC, Geda YE, Therneau TM, Rocca WA: Anxious personality predicts an increased risk of Parkinson's disease. Mov Disord 2010, 25:2105–2113.
- 44. Abe N, Fujii T, Hirayama K, Takeda A, Hosokai Y, Ishioka T, Nishio Y, Suzuki K, Itoyama Y, Takahashi S, *et al*: **Do parkinsonian patients have trouble telling**

- lies? The neurobiological basis of deceptive behaviour. *Brain* 2009, **132**:1386–1395.
- 45. Ishihara L, Brayne C: What is the evidence for a premorbid parkinsonian personality: a systematic review. *Mov Disord* 2006, **21**:1066–1072.
- Macías Y, Benito-León J, Louis ED, Cano-Vindel A: Anger in Parkinson's disease: a case-control study. Mov Disord 2008, 23:195–199.
- Evans AH, Lawrence AD, Potts J, MacGregor L, Katzenschlager R, Shaw K, Zijlmans J, Lees AJ: Relationship between impulsive sensation seeking traits, smoking, alcohol and caffeine intake, and Parkinson's disease. J Neurol Neurosurg Psychiatry 2006, 77:317–321.
- Jellinger KA, Paulus W: Clinico-pathological correlations in Parkinson's disease. Clin Neurol Neurosurg 1992, 94(Suppl):586–88.
- Feldmann A, Illes Z, Kosztolanyi P, Illes E, Mike A, Kover F, Balas I, Kovacs N, Nagy F: Morphometric changes of gray matter in Parkinson's disease with depression: a voxel-based morphometry study. Mov Disord 2008, 23:42–46.
- Blonder LX, Slevin JT: Emotional dysfunction in Parkinson's disease. Behav Neurol 2011. 24:201–217.
- Parkkinen L, Pirttilä T, Alafuzoff I: Applicability of current staging/ categorization of alpha-synuclein pathology and their clinical relevance. Acta Neuropathol 2008, 115:399–407.
- Tan SK, Hartung H, Sharp T, Temel Y: Serotonin-dependent depression in Parkinson's disease: a role for the subthalamic nucleus? Neuropharmacology 2011, 61:387–399.
- Shi M, Caudle WM, Zhang J: Biomarker discovery in neurodegenerative diseases: a proteomic approach. Neurobiol Dis 2009, 35:157–164.
- Caudle WM, Bammler TK, Lin Y, Pan S, Zhang J: Using 'omics' to define pathogenesis and biomarkers of Parkinson's disease. Expert Rev Neurother 2010, 10:925–942.
- Hong Z, Shi M, Chung KA, Quinn JF, Peskind ER, Galasko D, Jankovic J, Zabetian CP, Leverenz JB, Baird G, et al: DJ-1 and alpha-synuclein in human cerebrospinal fluid as biomarkers of Parkinson's disease. Brain 2010, 133:713–726.
- Mollenhauer B, Locascio JJ, Schulz-Schaeffer W, Sixel-Döring F, Trenkwalder C, Schlossmacher MG: α-Synuclein and tau concentrations in cerebrospinal fluid of patients presenting with parkinsonism: a cohort study. Lancet Neurol 2011, 10:230–240.
- Tokuda T, Salem SA, Allsop D, Mizuno T, Nakagawa M, Qureshi MM, Locascio JJ, Schlossmacher MG, El-Agnaf OM: Decreased alpha-synuclein in cerebrospinal fluid of aged individuals and subjects with Parkinson's disease. Biochem Biophys Res Commun 2006, 349:162–166.
- Mollenhauer B, Cullen V, Kahn I, Krastins B, Outeiro TF, Pepivani I, Ng J, Schulz-Schaeffer W, Kretzschmar HA, McLean PJ, et al: Direct quantification of CSF alpha-synuclein by ELISA and first cross-sectional study in patients with neurodegeneration. Exp. Neurol 2008, 213:315–325.
- Shi M, Bradner J, Hancock AM, Chung KA, Quinn JF, Peskind ER, Galasko D, Jankovic J, Zabetian CP, Kim HM, et al: Cerebrospinal fluid biomarkers for Parkinson disease diagnosis and progression. Ann Neurol 2011, 69:570–580.
- Ohrfelt A, Grognet P, Andreasen N, Wallin A, Vanmechelen E, Blennow K, Zetterberg H: Cerebrospinal fluid alpha-synuclein in neurodegenerative disorders-a marker of synapse loss? Neurosci Lett 2009, 450:332–335.
- Borghi R, Marchese R, Negro A, Marinelli L, Forloni G, Zaccheo D, Abbruzzese G, Tabaton M: Full length alpha-synuclein is present in cerebrospinal fluid from Parkinson's disease and normal subjects. Neurosci Lett 2000, 287:65–67.
- Mollenhauer B, El-Agnaf OM, Marcus K, Trenkwalder C, Schlossmacher MG: Quantification of α-synuclein in cerebrospinal fluid as a biomarker candidate: review of the literature and considerations for future studies. Biomark Med 2010, 4:683–699.
- Li QX, Mok SS, Laughton KM, McLean CA, Cappai R, Masters CL, Culvenor JG, Horne MK: Plasma alpha-synuclein is decreased in subjects with Parkinson's disease. Exp Neurol 2007, 204:583–588.
- Lee PH, Lee G, Park HJ, Bang OY, Joo IS, Huh K: The plasma alpha-synuclein levels in patients with Parkinson's disease and multiple system atrophy. J Neural Transm 2006, 113:1435–1439.
- El-Agnaf OM, Salem SA, Paleologou KE, Curran MD, Gibson MJ, Court JA, Schlossmacher MG, Allsop D: Detection of oligomeric forms of alphasynuclein protein in human plasma as a potential biomarker for Parkinson's disease. FASEB J 2006, 20:419

  –425.

- Duran R, Barrero FJ, Morales B, Luna JD, Ramirez M, Vives F: Plasma alphasynuclein in patients with Parkinson's disease with and without treatment. Mov Disord 2010, 25:489

  –493.
- Yanamandra K, Gruden MA, Casaite V, Meskys R, Forsgren L, Morozova-Roche LA: α-synuclein reactive antibodies as diagnostic biomarkers in blood sera of Parkinson's disease patients. PLoS One 2011, 6:e18513.
- Shi M, Zabetian CP, Hancock AM, Ginghina C, Hong Z, Yearout D, Chung KA, Quinn JF, Peskind ER, Galasko D, et al: Significance and confounders of peripheral DJ-1 and alpha-synuclein in Parkinson's disease. Neurosci Lett 2010, 480:78–82.
- Hoepken HH, Gispert S, Azizov M, Klinkenberg M, Ricciardi F, Kurz A, Morales-Gordo B, Bonin M, Riess O, Gasser T, et al: Parkinson patient fibroblasts show increased alpha-synuclein expression. Exp Neurol 2008, 212:307–313.
- Edwards LL, Pfeiffer RF, Quigley EM, Hofman R, Balluff M: Gastrointestinal symptoms in Parkinson's disease. Mov Disord 1991, 6:151–156.
- Del Tredici K, Hawkes CH, Ghebremedhin E, Braak H: Lewy pathology in the submandibular gland of individuals with incidental Lewy body disease and sporadic Parkinson's disease. Acta Neuropathol 2010, 119:703–713.
- Proctor GB, Carpenter GH: Regulation of salivary gland function by autonomic nerves. Auton Neurosci 2007, 133:3–18.
- Devic I, Hwang H, Edgar JS, Izutsu K, Presland R, Pan C, Goodlett DR, Wang Y, Armaly J, Tumas V, et al: Salivary α-synuclein and DJ-1: potential biomarkers for Parkinson's disease. Brain 2011, 134:e178.
- Sato S, Mizuno Y, Hattori N: Urinary 8-hydroxydeoxyguanosine levels as a biomarker for progression of Parkinson disease. Neurology 2005, 64:1081– 1083.
- Cipriani S, Chen X, Schwarzschild MA: Urate: a novel biomarker of Parkinson's disease risk, diagnosis and prognosis. *Biomark Med* 2010, 4:701–712
- de Lau LM, Koudstaal PJ, Hofman A, Breteler MM: Serum uric acid levels and the risk of Parkinson disease. Ann Neurol 2005, 58:797–800.
- Weisskopf MG, O'Reilly E, Chen H, Schwarzschild MA, Ascherio A: Plasma urate and risk of Parkinson's disease. Am J Epidemiol 2007, 166:561–567.
- Chen H, Mosley TH, Alonso A, Huang X: Plasma urate and Parkinson's disease in the Atherosclerosis Risk in Communities (ARIC) study. Am J Epidemiol 2009, 169:1064–1069.
- Schwarzschild MA, Schwid SR, Marek K, Watts A, Lang AE, Oakes D, Shoulson I, Ascherio A, Hyson C, Gorbold E, et al: Serum urate as a predictor of clinical and radiographic progression in Parkinson disease. Arch Neurol 2008, 65:716–723.
- Aleyasin H, Rousseaux MW, Marcogliese PC, Hewitt SJ, Irrcher I, Joselin AP, Parsanejad M, Kim RH, Rizzu P, Callaghan SM, et al: DJ-1 protects the nigrostriatal axis from the neurotoxin MPTP by modulation of the AKT pathway. Proc Natl Acad Sci USA 2010, 107:3186–3191.
- Duan X, Kelsen SG, Merali S: Proteomic analysis of oxidative stressresponsive proteins in human pneumocytes: insight into the regulation of DJ-1 expression. J Proteome Res 2008, 7:4955–4961.
- Jin J, Li GJ, Davis J, Zhu D, Wang Y, Pan C, Zhang J: Identification of novel proteins associated with both alpha-synuclein and DJ-1. Mol Cell Proteomics 2007, 6:845–859.
- Maita C, Tsuji S, Yabe I, Hamada S, Ogata A, Maita H, Iguchi-Ariga SM, Sasaki H, Ariga H: Secretion of DJ-1 into the serum of patients with Parkinson's disease. Neurosci Lett 2008, 431:86–89.
- Waragai M, Nakai M, Wei J, Fujita M, Mizuno H, Ho G, Masliah E, Akatsu H, Yokochi F, Hashimoto M: Plasma levels of DJ-1 as a possible marker for progression of sporadic Parkinson's disease. Neurosci Lett 2007, 425:18–22.
- Waragai M, Wei J, Fujita M, Nakai M, Ho GJ, Masliah E, Akatsu H, Yamada T, Hashimoto M: Increased level of DJ-1 in the cerebrospinal fluids of sporadic Parkinson's disease. Biochem Biophys Res Commun 2006, 345:967–972.
- Modulator of the Oxidized Form of DJ-1, a Wild-type of Familial Parkinson's Disease-Linked PARK7. J Pharmacol Sci 2011, 117:189–203.
- 87. Saito Y, Hamakubo T, Yoshida Y, Ogawa Y, Hara Y, Fujimura H, Imai Y, Iwanari H, Mochizuki Y, Shichiri M, et al: Preparation and application of monoclonal antibodies against oxidized DJ-1. Significant elevation of oxidized DJ-1 in erythrocytes of early-stage Parkinson disease patients.

- Neurosci Lett 2009, 465:1-5.
- Ren H, Fu K, Wang D, Mu C, Wang G: Oxidized DJ-1 interacts with the mitochondrial protein BCL-XL. J Biol Chem 2011, 286:35308–35317.
- Mehta SH, Morgan JC, Sethi KD: Neuroimaging and transcranial ultrasonography in Parkinson's disease. Curr Neurol Neurosci Rep 2008, 8:297–303.
- Booij J, Korn P, Linszen DH, van Royen EA: Assessment of endogenous dopamine release by methylphenidate challenge using iodine-123 iodobenzamide single-photon emission tomography. Eur J Nucl Med 1997, 24:674–677.
- 91. Marek K, Jennings D: Can we image premotor Parkinson disease? *Neurology* 2009, **72**:S21–26.
- Panzacchi A, Moresco RM, Garibotto V, Antonini A, Gobbo C, Isaias IU, Goldwurm S, Bonaldi L, Carpinelli A, Pezzoli G, et al: A voxel-based PET study of dopamine transporters in Parkinson's disease: relevance of age at onset. Neurobiol Dis 2008, 31:102–109.
- Schwingenschuh P, Ruge D, Edwards MJ, Terranova C, Katschnig P, Carrillo F, Silveira-Moriyama L, Schneider SA, Kägi G, Palomar FJ, et al: Distinguishing SWEDDs patients with asymmetric resting tremor from Parkinson's disease: a clinical and electrophysiological study. Mov Disord 2010, 25:560–569.
- Hu MT, Taylor-Robinson SD, Chaudhuri KR, Bell JD, Labbé C, Cunningham VJ, Koepp MJ, Hammers A, Morris RG, Turjanski N, Brooks DJ: Cortical dysfunction in non-demented Parkinson's disease patients: a combined (31)P-MRS and (18)FDG-PET study. Brain 2000, 123(Pt 2):340–352.
- Hilker R, Schweitzer K, Coburger S, Ghaemi M, Weisenbach S, Jacobs AH, Rudolf J, Herholz K, Heiss WD: Nonlinear progression of Parkinson disease as determined by serial positron emission tomographic imaging of striatal fluorodopa F 18 activity. Arch Neurol 2005, 62:378–382.
- Stoessl AJ: Positron emission tomography in premotor Parkinson's disease. Parkinsonism Relat Disord 2007, 13(Suppl 3):S421–424.
- Hu MT, White SJ, Chaudhuri KR, Morris RG, Bydder GM, Brooks DJ: Correlating rates of cerebral atrophy in Parkinson's disease with measures of cognitive decline. J Neural Transm 2001, 108:571–580.
- Martin WR, Wieler M, Gee M: Midbrain iron content in early Parkinson disease: a potential biomarker of disease status. *Neurology* 2008, 70:1411–1417.
- Vaillancourt DE, Spraker MB, Prodoehl J, Abraham I, Corcos DM, Zhou XJ, Comella CL, Little DM: High-resolution diffusion tensor imaging in the substantia nigra of de novo Parkinson disease. *Neurology* 2009, 72:1378–1384.
- Péran P, Cherubini A, Assogna F, Piras F, Quattrocchi C, Peppe A, Celsis P, Rascol O, Démonet JF, Stefani A, et al: Magnetic resonance imaging markers of Parkinson's disease nigrostriatal signature. Brain 2010, 133:3423–3433.
- Wild EJ, Fox NC: Serial volumetric MRI in Parkinsonian disorders. Mov Disord 2009, 24(Suppl 2):S691–698.
- 102. Hampe C, Ardila-Osorio H, Fournier M, Brice A, Corti O: Biochemical analysis of Parkinson's disease-causing variants of Parkin, an E3 ubiquitin-protein ligase with monoubiquitylation capacity. Hum Mol Genet 2006, 15:2059–2075.
- 103. Veeriah S, Taylor BS, Meng S, Fang F, Yilmaz E, Vivanco I, Janakiraman M, Schultz N, Hanrahan AJ, Pao W, et al: Somatic mutations of the Parkinson's disease-associated gene PARK2 in glioblastoma and other human malignancies. Nat Genet 2010, 42:77–82.
- 104. Sutherland G, Mellick G, Sue C, Chan DK, Rowe D, Silburn P, Halliday G: A functional polymorphism in the parkin gene promoter affects the age of onset of Parkinson's disease. Neurosci Lett 2007, 414:170–173.
- Hruska KS, LaMarca ME, Scott CR, Sidransky E: Gaucher disease: mutation and polymorphism spectrum in the glucocerebrosidase gene (GBA). Hum Mutat 2008, 29:567–583.
- Westbroek W, Gustafson AM, Sidransky E: Exploring the link between glucocerebrosidase mutations and parkinsonism. Trends Mol Med 2011, 17:485–493.
- Neudorfer O, Giladi N, Elstein D, Abrahamov A, Turezkite T, Aghai E, Reches A, Bembi B, Zimran A: Occurrence of Parkinson's syndrome in type I Gaucher disease. OJM 1996. 89:691–694.
- Aharon-Peretz J, Rosenbaum H, Gershoni-Baruch R: Mutations in the glucocerebrosidase gene and Parkinson's disease in Ashkenazi Jews. N Engl J Med 2004, 351:1972–1977.

- 109. Zimran A, Neudorfer O, Elstein D: The glucocerebrosidase gene and Parkinson's disease in Ashkenazi Jews. N Engl J Med 2005, 352:728–731. author reply 728-731.
- 110. De Marco EV, Annesi G, Tarantino P, Rocca FE, Provenzano G, Civitelli D, Cirò Candiano IC, Annesi F, Carrideo S, Condino F, et al: Glucocerebrosidase gene mutations are associated with Parkinson's disease in southern Italy. Mov Disord 2008, 23:460–463.
- 111. Mao XY, Burgunder JM, Zhang ZJ, An XK, Zhang JH, Yang Y, Li T, Wang YC, Chang XL, Peng R: Association between GBA L444P mutation and sporadic Parkinson's disease from Mainland China. Neurosci Lett 2010, 469:756–759
- 112. Sun QY, Guo JF, Wang L, Yu RH, Zuo X, Yao LY, Pan Q, Xia K, Tang BS: Glucocerebrosidase gene L444P mutation is a risk factor for Parkinson's disease in Chinese population. Mov Disord 2010, 25:1005–1011.
- 113. Lesage S, Condroyer C, Hecham N, Anheim M, Belarbi S, Lohman E, Viallet F, Pollak P, Abada M, Dürr A, et al: Mutations in the glucocerebrosidase gene confer a risk for Parkinson disease in North Africa. Neurology 2011, 76:301–303
- 114. Neumann J, Bras J, Deas E, O'Sullivan SS, Parkkinen L, Lachmann RH, Li A, Holton J, Guerreiro R, Paudel R, et al: Glucocerebrosidase mutations in clinical and pathologically proven Parkinson's disease. Brain 2009, 132:1783–1794.
- 115. Lesage S, Anheim M, Condroyer C, Pollak P, Durif F, Dupuits C, Viallet F, Lohmann E, Corvol JC, Honoré A, et al: Large-scale screening of the Gaucher's disease-related glucocerebrosidase gene in Europeans with Parkinson's disease. Hum Mol Genet 2011. 20:202–210.
- Goker-Alpan O, Lopez G, Vithayathil J, Davis J, Hallett M, Sidransky E: The spectrum of parkinsonian manifestations associated with glucocerebrosidase mutations. Arch Neurol 2008, 65:1353–1357.
- 117. Mata IF, Samii A, Schneer SH, Roberts JW, Griffith A, Leis BC, Schellenberg GD, Sidransky E, Bird TD, Leverenz JB, et al: Glucocerebrosidase gene mutations: a risk factor for Lewy body disorders. Arch Neurol 2008, 65:379–382.
- 118. Sidransky E, Nalls MA, Aasly JO, Aharon-Peretz J, Annesi G, Barbosa ER, BarShira A, Berg D, Bras J, Brice A, et al: Multicenter analysis of glucocerebrosidase mutations in Parkinson's disease. N Engl J Med 2009, 361:1651–1661.
- 119. Goker-Alpan O, Stubblefield BK, Giasson BI, Sidransky E: Glucocerebrosidase is present in α-synuclein inclusions in Lewy body disorders. Acta Neuropathol 2010, 120:641–649.
- 120. Cullen V, Sardi SP, Ng J, Xu YH, Sun Y, Tomlinson JJ, Kolodziej P, Kahn I, Saftig P, Woulfe J, et al: Acid β-glucosidase mutants linked to Gaucher disease, Parkinson disease, and Lewy body dementia alter α-synuclein processing. Ann Neurol 2011, 69:940–953.
- 121. Yap TL, Gruschus JM, Velayati A, Westbroek W, Goldin E, Moaven N, Sidransky E, Lee JC: Alpha-synuclein interacts with Glucocerebrosidase providing a molecular link between Parkinson and Gaucher diseases. J Biol Chem 2011, 286:28080–28088.
- 122. Lesage S, Brice A: Parkinson's disease: from monogenic forms to genetic susceptibility factors. *Hum Mol Genet* 2009, **18**:R48–59.
- Dächsel JC, Farrer MJ: LRRK2 and Parkinson disease. Arch Neurol 2010, 67:542–547.
- Kumari U, Tan EK: LRRK2 in Parkinson's disease: genetic and clinical studies from patients. FEBS J 2009. 276:6455–6463.
- 125. Westermann B, Wattendorf E, Schwerdtfeger U, Husner A, Fuhr P, Gratzl O, Hummel T, Bilecen D, Welge-Lüssen A: Functional imaging of the cerebral olfactory system in patients with Parkinson's disease. J Neurol Neurosurg Psychiatry 2008, 79:19–24.
- 126. Ozelius LJ, Senthil G, Saunders-Pullman R, Ohmann E, Deligtisch A, Tagliati M, Hunt AL, Klein C, Henick B, Hailpern SM, et al: LRRK2 G2019S as a cause of Parkinson's disease in Ashkenazi Jews. N Engl J Med 2006, 354:424–425.
- 127. Lesage S, Dürr A, Tazir M, Lohmann E, Leutenegger AL, Janin S, Pollak P, Brice A: Group FPsDGS: LRRK2 G2019S as a cause of Parkinson's disease in North African Arabs. N Engl J Med 2006, 354:422–423.
- 128. Anand VS, Reichling LJ, Lipinski K, Stochaj W, Duan W, Kelleher K, Pungaliya P, Brown EL, Reinhart PH, Somberg R, et al: Investigation of leucine-rich repeat kinase 2: enzymological properties and novel assays. FEBS J 2009, 276:466–478.
- 129. Li X, Patel JC, Wang J, Avshalumov MV, Nicholson C, Buxbaum JD, Elder GA, Rice ME, Yue Z: Enhanced striatal dopamine transmission and motor

- performance with LRRK2 overexpression in mice is eliminated by familial Parkinson's disease mutation G2019S. J Neurosci 2010, 30:1788–1797.
- Qing H, Wong W, McGeer EG, McGeer PL: Lrrk2 phosphorylates alpha synuclein at serine 129: Parkinson disease implications. Biochem Biophys Res Commun 2009, 387:149–152.
- 131. Shi M, Furay AR, Sossi V, Aasly JO, Armaly J, Wang Y, Wszolek ZK, Uitti RJ, Hasegawa K, Yokoyama T, et al: DJ-1 and αSYN in LRRK2 CSF do not correlate with striatal dopaminergic function. Neurobiol Aging 2011, :.
- 132. Aasly JO, Shi M, Sossi V, Stewart T, Johansen KK, Wszolek ZK, Uitti RJ, Hasegawa K, Yokoyama T, Zabetian CP, et al: Cerebrospinal fluid amyloid β and tau in LRRK2 mutation carriers. Neurology 2011, :.
- Johansen KK, Wang L, Aasly JO, White LR, Matson WR, Henchcliffe C, Beal MF, Bogdanov M: Metabolomic profiling in LRRK2-related Parkinson's disease. PLoS One 2009, 4:e7551.
- Mirelman A, Gurevich T, Giladi N, Bar-Shira A, Orr-Urtreger A, Hausdorff JM: Gait alterations in healthy carriers of the LRRK2 G2019S mutation. Ann Neurol 2011. 69:193–197.
- 135. Satake W, Nakabayashi Y, Mizuta I, Hirota Y, Ito C, Kubo M, Kawaguchi T, Tsunoda T, Watanabe M, Takeda A, et al: Genome-wide association study identifies common variants at four loci as genetic risk factors for Parkinson's disease. Nat Genet 2009, 41:1303–1307.
- 136. Do CB, Tung JY, Dorfman E, Kiefer AK, Drabant EM, Francke U, Mountain JL, Goldman SM, Tanner CM, Langston JW, et al: Web-based genome-wide association study identifies two novel loci and a substantial genetic component for Parkinson's disease. PLoS Genet 2011, 7:e1002141.
- 137. Nalls MA, Plagnol V, Hernandez DG, Sharma M, Sheerin UM, Saad M, Simón-Sánchez J, Schulte C, Lesage S, Sveinbjörnsdóttir S, et al: Imputation of sequence variants for identification of genetic risks for Parkinson's disease: a meta-analysis of genome-wide association studies. Lancet 2011, 377:641–649.
- 138. Edwards TL, Scott WK, Almonte C, Burt A, Powell EH, Beecham GW, Wang L, Züchner S, Konidari I, Wang G, et al: Genome-wide association study confirms SNPs in SNCA and the MAPT region as common risk factors for Parkinson disease. Ann Hum Genet 2010, 74:97–109.
- 139. Maraganore DM, de Andrade M, Lesnick TG, Strain KJ, Farrer MJ, Rocca WA, Pant PV, Frazer KA, Cox DR, Ballinger DG: High-resolution whole-genome association study of Parkinson disease. Am J Hum Genet 2005, 77:685–693.
- 140. Fung HC, Scholz S, Matarin M, Simón-Sánchez J, Hernandez D, Britton A, Gibbs JR, Langefeld C, Stiegert ML, Schymick J, et al: Genome-wide genotyping in Parkinson's disease and neurologically normal controls: first stage analysis and public release of data. Lancet Neurol 2006, 5:911–916.
- 141. Pankratz N, Wilk JB, Latourelle JC, DeStefano AL, Halter C, Pugh EW, Doheny KF, Gusella JF, Nichols WC, Foroud T, et al: Genomewide association study for susceptibility genes contributing to familial Parkinson disease. Hum Genet 2009, 124:593–605.
- Rhodes SL, Sinsheimer JS, Bordelon Y, Bronstein JM, Ritz B: Replication of GWAS associations for GAK and MAPT in Parkinson's disease. Ann Hum Genet 2011, 75:195–200.
- 143. Trotta L, Guella I, Soldà G, Sironi F, Tesei S, Canesi M, Pezzoli G, Goldwurm S, Duga S, Asselta R: SNCA and MAPT genes: Independent and joint effects in Parkinson disease in the Italian population. Parkinsonism Relat Disord 2011
- 144. Toledo JB, Brettschneider J, Grossman M, Arnold SE, Hu WT, Xie SX, Lee VM, Shaw LM, Trojanowski JQ: CSF biomarkers cutoffs: the importance of coincident neuropathological diseases. Acta Neuropathol 2012, :.

#### doi:10.1186/2047-9158-1-11

Cite this article as: Haas *et al.*: Premotor biomarkers for Parkinson's disease - a promising direction of research. *Translational Neurodegeneration* 2012 1:11.

# Submit your next manuscript to BioMed Central and take full advantage of:

- Convenient online submission
- Thorough peer review
- No space constraints or color figure charges
- Immediate publication on acceptance
- Inclusion in PubMed, CAS, Scopus and Google Scholar
- Research which is freely available for redistribution

Submit your manuscript at www.biomedcentral.com/submit

