

REVIEW

Open Access

# Regenerative medicine for the kidney: stem cell prospects & challenges

Yue Li and Rebecca A Wingert\*

## Abstract

The kidney has key roles in maintaining human health. There is an escalating medical crisis in nephrology as growing numbers of patients suffer from kidney diseases that culminate in organ failure. While dialysis and transplantation provide life-saving treatments, these therapies are rife with limitations and place significant burdens on patients and healthcare systems. It has become imperative to find alternative ways to treat existing kidney conditions and preemptive means to stave off renal dysfunction. The creation of innovative medical approaches that utilize stem cells has received growing research attention. In this review, we discuss the regenerative and maladaptive cellular responses that occur during acute and chronic kidney disease, the emerging evidence about renal stem cells, and some of the issues that lie ahead in bridging the gap between basic stem cell biology and regenerative medicine for the kidney.

**Keywords:** Renal stem cell, Renal progenitor, Kidney regeneration, Acute kidney injury, Chronic kidney disease

## Introduction

The kidney performs essential physiological jobs ranging from metabolic waste excretion to homeostatic functions like osmoregulation. Kidney diseases are currently a global public health problem, with an incidence that has reached epidemic proportions and continues to climb in the U.S. and worldwide [1]. These trends correlate with the global rise in the aged population and the increasing prevalence of conditions that cause renal complications, namely cardiovascular disease, hypertension and diabetes [2]. Kidney diseases arise from congenital defects as well as acquired conditions that result from acute kidney injury (AKI) or chronic kidney disease (CKD) [3-7]. AKI involves a rapid loss of kidney function from sudden renal cell damage, which can be triggered by ischemia, toxins, or sepsis [4-6]. CKD is typified by the progressive loss of kidney function over time due to fibrosis and the erosion of healthy tissue [7]. Kidney disease leads to organ failure, known as end-stage renal disease (ESRD), which requires renal replacement therapy with dialysis or transplantation. Although renal failure can be managed clinically, it has high mortality rates and necessitates intensive, long-term care. This places a considerable burden

on patients and their families, and a tremendous socioeconomic strain on healthcare systems [2].

The need for new methods to alleviate, cure, or prevent renal disease has fueled great interest in the topic of kidney stem cell biology. For over a decade now, the use of stem cells for regenerative medicine has been broadly heralded as the coming of a new age in healthcare [8]. Unfortunately, the sheer magnitude of the excitement, hope, and promise surrounding the notion of stem cell therapies has been rivaled by the enormity of the challenges in making such approaches a reality. A major hurdle facing nephrology researchers is that the human kidney has been classically defined as a non-proliferative and non-regenerative organ. However, with the discovery of adult stem cells in organs that were once thought to be non-regenerative (like the brain), the cellular make-up of the kidney has come to be reevaluated. There is emerging evidence that human kidneys possess innate regenerative abilities. For example, diabetic patients with CKD exhibited reversion of fibrotic lesions in their kidneys ten years after receiving a pancreas transplant [9,10]. Recent research efforts have focused on better understanding how the kidney can heal after injury, and determining if the kidney contains stem cells that mediate tissue replacement. Here we define the roles of stem cells in kidney development, discuss how the adult kidney responds to

\* Correspondence: rwingert@nd.edu  
Department of Biological Sciences, University of Notre Dame, Notre Dame, IN 46556, USA

damage, and explore the current knowledge about the existence of adult renal stem cells.

## Review

### Stem cells and their roles in development and regeneration

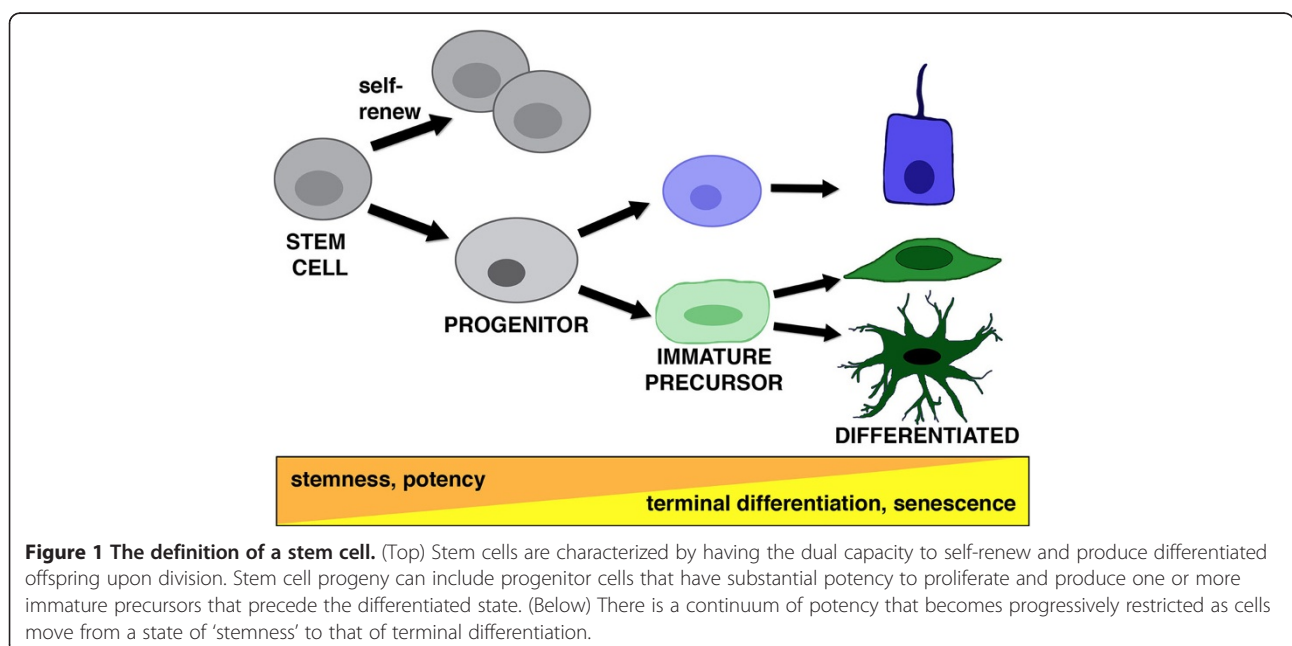
Stem cells are unique cells that are able to replenish themselves through self-renewal and can divide to produce differentiated (i.e. specialized) cell types (Figure 1) [8]. Stem cells can self-renew through a symmetric cell division, where both the newly produced offspring cells maintain the characteristics of the parent stem cell. Stem cells can also self-renew through asymmetric division, in which they produce a stem cell and another offspring that has a different potency and lineage-potential, such as a committed progenitor that transiently amplifies to make several progeny (Figure 1). Interestingly, stem cell division does not always have to involve renewal, and can instead generate assorted combinations of offspring—like two progenitors or a progenitor and a differentiated cell. The signals that trigger stem cell division and progeny identity are influenced by a complex interplay of intrinsic (cell autonomous) factors as well as extrinsic factors such as those present at the niche, which is the local microenvironment where the stem cell resides.

Stem cells drive normal development and play many roles in adult tissue regeneration [11]. Pluripotent stem cells capable of giving rise to any cell lineage can be isolated from early stage mammalian embryos. These embryonic stem (ES) cells maintain pluripotency when cultured in the right conditions [12]. As development

progresses, lineage-restricted stem cells produce the tissues and organs of the body. Development does not necessarily exhaust stem cell pools, and often leads to the formation of tissue-specific or so-called adult stem cells that are maintained throughout life. Adult stem cells typically show more restricted potency (e.g. they are multi-, bi, or unipotent), and have been discovered in diverse creatures from simple metazoans like flies to mammals like mice and humans [13]. In fact, adult stem cells have been reported in a substantial list of human tissues and organs. Adult stem cells can act to replace cells that have a naturally limited lifespan, thus serving to maintain the integrity of the soma. Further, many adult stem cells can respond dynamically to injury and fuel substantial regeneration of damaged tissues. For these reasons, adult stem cells are thought to have profound impacts on the etiology of disease, malignancy, and aging [14,15]. Research efforts to identify regenerative therapeutics have encompassed the study of ES cells, adult stem cells, and the pursuit of reprogramming methods to manipulate differentiated cells and obtain induced pluripotent stem (iPS) cells that have broad lineage potential similar to the ES cell [12].

### Renal complexity: the kidney arises from stem cells during development that produce diverse differentiated cell types

The kidney develops from several stem cell pools during organogenesis. As in other mammals, the human kidney derives from intermediate mesoderm (IM) and proceeds through three progressive phases, each marked by the formation of a more advanced kidney: the pronephros,

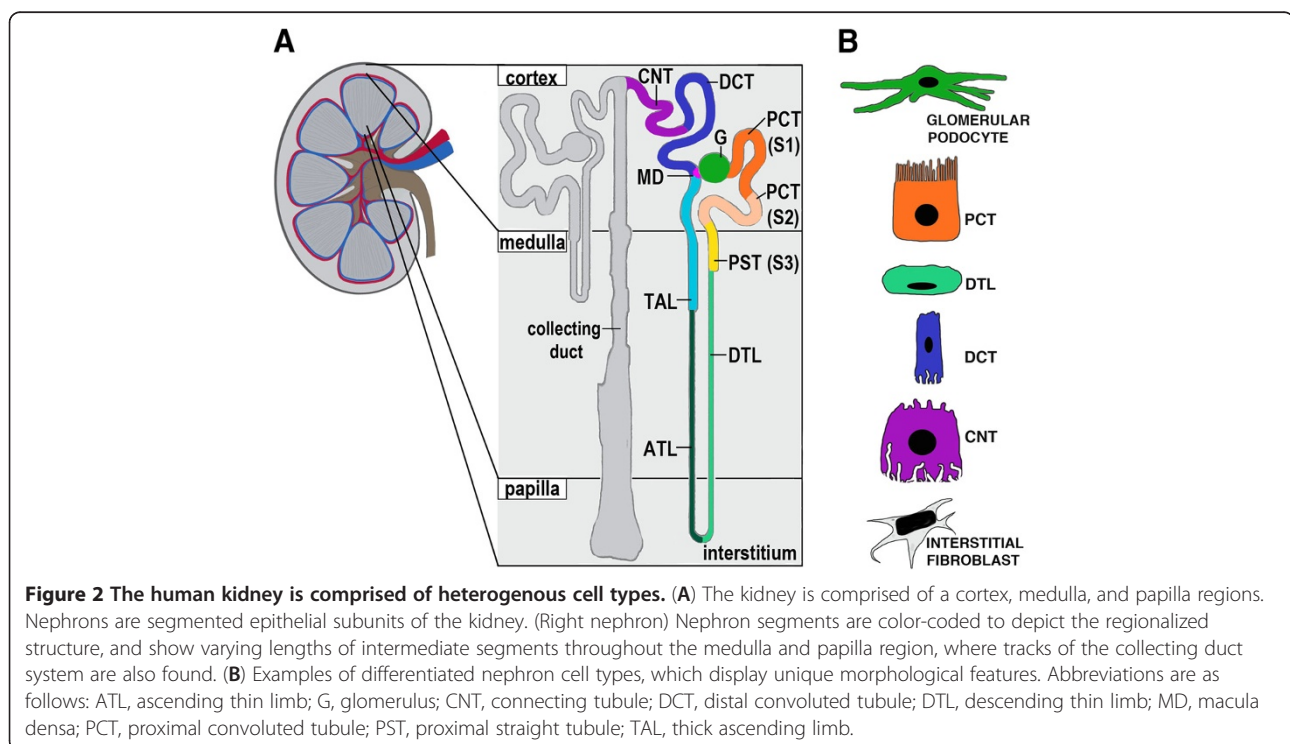


**Figure 1** The definition of a stem cell. (Top) Stem cells are characterized by having the dual capacity to self-renew and produce differentiated offspring upon division. Stem cell progeny can include progenitor cells that have substantial potency to proliferate and produce one or more immature precursors that precede the differentiated state. (Below) There is a continuum of potency that becomes progressively restricted as cells move from a state of 'stemness' to that of terminal differentiation.

which is a rudimentary and non-functional organ, the mesonephros, which only functions a short time during embryonic development, and the metanephros, which becomes the definitive form of the adult kidney [16]. The basic structural unit of each kidney form is the nephron, an epithelial tube that accomplishes waste excretion. Nephrons have three major parts: (1) a glomerulus that filters the blood, (2) a tubule that modifies the filtrate to reabsorb and secrete solutes as the fluid passes through proximal, intermediate, and distal segments, and (3) a duct that carries the urine into a centralized collecting system [17]. The pronephros and mesonephros are made from IM that develops into simple nephrons that are connected to a pair of nephric (Wolffian) ducts, and these tissues degenerate in succession as the metanephros forms. The metanephric kidney is produced when a localized region of the nephric duct forms an outgrowth known as the ureteric bud (UB). The UB invades the adjacent IM, which at this stage is termed the metanephric mesenchyme (MM), and undergoes reiterative branching morphogenesis to create a complex duct network. The MM aggregates and condenses to form the cap mesenchyme (CM), a self-renewing stem cell population that makes nephrons around the branching UB [18-21]. Nephrogenesis proceeds when a cluster of CM undergoes a mesenchymal to epithelial transition (MET) into a renal vesicle. Each renal vesicle will proliferate and undergo morphogenesis to make one nephron. The loosely packed surrounding MM

produces interstitial stroma. There is ongoing work to characterize the UB and MM derivatives, and to profile genes expressed by subsequent renal cell lineages, with a large focus on the CM because of its self-renewal property [18-21]. Although the CM exhibits self-renewal, the CM is transient and the entire CM population is triggered to undergo a final round of nephrogenesis at the end of metanephros formation. There is no evidence that a CM population is maintained to become an adult renal stem cell population. It is currently unknown if self-renewal is a short-lived property of CM renal stem cells and/or if developmental signaling induces all of the CM cells to become renal vesicles that undergo nephrogenesis.

When metanephros development is complete, this kidney has an elaborate, arborized architecture and diverse cellular makeup due to the combined contributions of CM, MM and UB progenitors. For example, over twenty differentiated cell types have been annotated in the human kidney [17] (Figure 2). Kidney composition is conserved between mammals and even with lower vertebrates like fish [17,22]. Nephrons alone contain more than a dozen epithelial cell types with specific functions (examples in Figure 2B)—which attests to the multipotency of CM stem cells during development [17]. The absolute number of nephrons and their architecture around the collecting system also renders complexity to the metanephric kidney. For example, human nephron endowment ranges from several hundred thousand to over one million nephrons per kidney, being quite



variable between individuals [23,24]. Nephrons are found throughout the cortex, or outer zone of the kidney, while their tubule loops and the collecting duct system inhabit the inner zones of the medulla and papilla. Located between nephrons and the collecting system is a heterogeneous interstitial stroma inhabited by several types of fibroblasts (Figure 2B), as well as immune cells such as dendritic cells [17]. The kidney is also highly vascularized, with extensive capillary networks of endothelial cells and pericytes [17]. Finally, lymphatics are present throughout the cortex, and the kidney is innervated by fibers of the sympathetic and parasympathetic nervous systems [17].

The identification of stem cells in these adult renal structures has been riddled with controversy. Several locations in nephrons, the interstitium, and the collecting system have been cited as housing cells with proliferative potential. In some cases, there is evidence that proliferative capacity may exist in differentiated cells. Other experimental data supports the notion of 'stemness' in particular renal cells. In the following sections we discuss several relevant phenomena in kidney biology: first, the adolescent growth of the kidney and nephron hypertrophy, followed by sections exploring the renal response to damage and the evidence concerning the role(s) of putative kidney stem cells and differentiated cells.

#### Renal cell dynamics in healthy kidneys: cell turnover and hypertrophy

During juvenile life, the mammalian kidney undergoes substantial normal growth even though organogenesis is complete just before or immediately following birth.

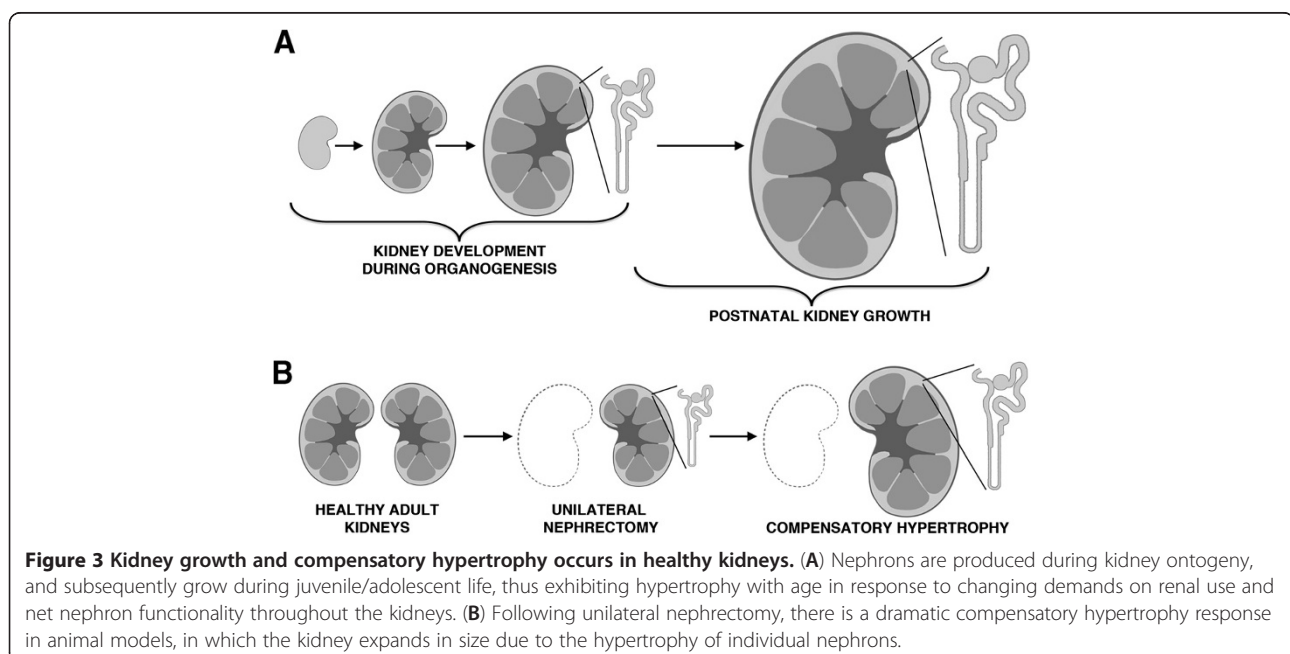
Further, kidneys have been documented to undergo a dramatic compensatory response to the sudden removal of renal tissue.

#### *Nephrogenesis in humans is limited to early life*

Nephron production, or nephrogenesis, is not the stratagem for kidney maintenance in mammals. Rather, nephrogenesis is limited to gestation or early post-natal life. Human nephrogenesis ceases around week 36 of gestation [25], while nephrogenesis subsides soon after birth in rats and mice [26,27]. Although nephron endowment is set early, mammalian kidneys grow remarkably in size and functionality during juvenile stages: as body mass increases, overall kidney size and blood filtration rates increase (Figure 3A) [28]. Nephrons grow by hyperplasia and hypertrophy, leading to enlargements in glomerular size and tubule length that correlate with the elevated capacity of the kidney to filter the blood [28-30].

#### *Cell turnover and hyperplasia during juvenile kidney growth and adult homeostasis*

Tissues throughout the mammalian body exhibit varying rates of cellular turnover during adult life [11]. Epithelia that are faced with high degrees of environmental stress use virtually constant turnover rates of adult stem cells and/or their transiently amplifying progeny as a strategy to maintain tissue integrity, as seen in the skin [31] and lining of the gastrointestinal tract [32] where millions of cells are replaced daily. Kidney nephrons and collecting duct epithelia are exposed to continual passage of filtrate, and thousands of living cells from the healthy human



**Figure 3** Kidney growth and compensatory hypertrophy occurs in healthy kidneys. (A) Nephrons are produced during kidney ontogeny, and subsequently grow during juvenile/adolescent life, thus exhibiting hypertrophy with age in response to changing demands on renal use and net nephron functionality throughout the kidneys. (B) Following unilateral nephrectomy, there is a dramatic compensatory hypertrophy response in animal models, in which the kidney expands in size due to the hypertrophy of individual nephrons.



urinary tract are excreted each day. For example, counts of exfoliated nephron tubular cells numbered ~78,000 cells per hour in men and ~68,000 cells per hour in women [33]. Cells from this so-called urinary sediment can be isolated and cultured, and include epithelial cells shed from the kidney, ureters, bladder, and urethra [33-36]. While the magnitude of renal cell turnover is lower than other organs, homeostatic mechanism(s) are still needed to maintain kidney functionality.

In adult kidneys, cell proliferation continues, albeit at a reduced rate. Renal cell division has been documented in several locales with pulse-chase labeling studies in rodents. After providing a pulse of a nucleotide analog (such as tritiated thymidine [thymidine- $H^3$ ] or 5'-bromo-2'-deoxyuridine [BrdU]), its incorporation into DNA enables the assessment of nuclear replication in preparation for mitosis. Further, the duration of time that the analog is maintained can be used to extrapolate the cycling rate of the label retaining cells (LRCs). A classic study using adult rats reported thymidine- $H^3$  throughout nephron glomeruli, tubules, and the collecting system after a short chase of 8, 24, or 72 hours [37]. More recent BrdU pulse-chase studies in adult rats found LRCs in glomeruli, tubules and collecting ducts after 7 days of BrdU administration, and scattered LRCs in proximal and distal nephron tubules after a 2 week chase [38,39]. Comparisons of proximal tubule proliferation in juvenile and adult rats using BrdU pulse-chase and immunohistochemistry with mitosis markers showed that juveniles had division rates that were ~10 fold higher [40-42]. Healthy human kidneys also have dividing cells in nephron tubules based on staining for the cell division markers Ki67 and proliferating cell nuclear antigen (PCNA) [43]. Interestingly, both healthy rat and human kidneys have tubule cells positive for cyclin D1—suggesting they are in the G1 phase of the cell cycle [42]. This finding is the basis of a hypothesis that a pool of renal cells is poised for division and may serve to bastion against intermittent single cell loss or even more widespread, catastrophic insults [42].

#### **Compensatory renal hypertrophy**

Healthy mammalian kidneys can also undergo remarkable adaptive changes in response to the elimination of renal tissue. After unilateral nephrectomy, or the removal of one kidney, the remaining kidney increases in size through a process of compensatory renal hypertrophy (Figure 3B) [44,45]. The individual nephron cells in the residual kidney enlarge. For example, nephron tubular cell volume increases in proximal and distal segments [44]. The nephrons also show functional adaptations, dramatically increasing their filtration and re-absorption rates [44,45]. Thus, a kidney can respond to shifts in physiological demands posed by marked, sudden organ loss by mechanisms that include cellular

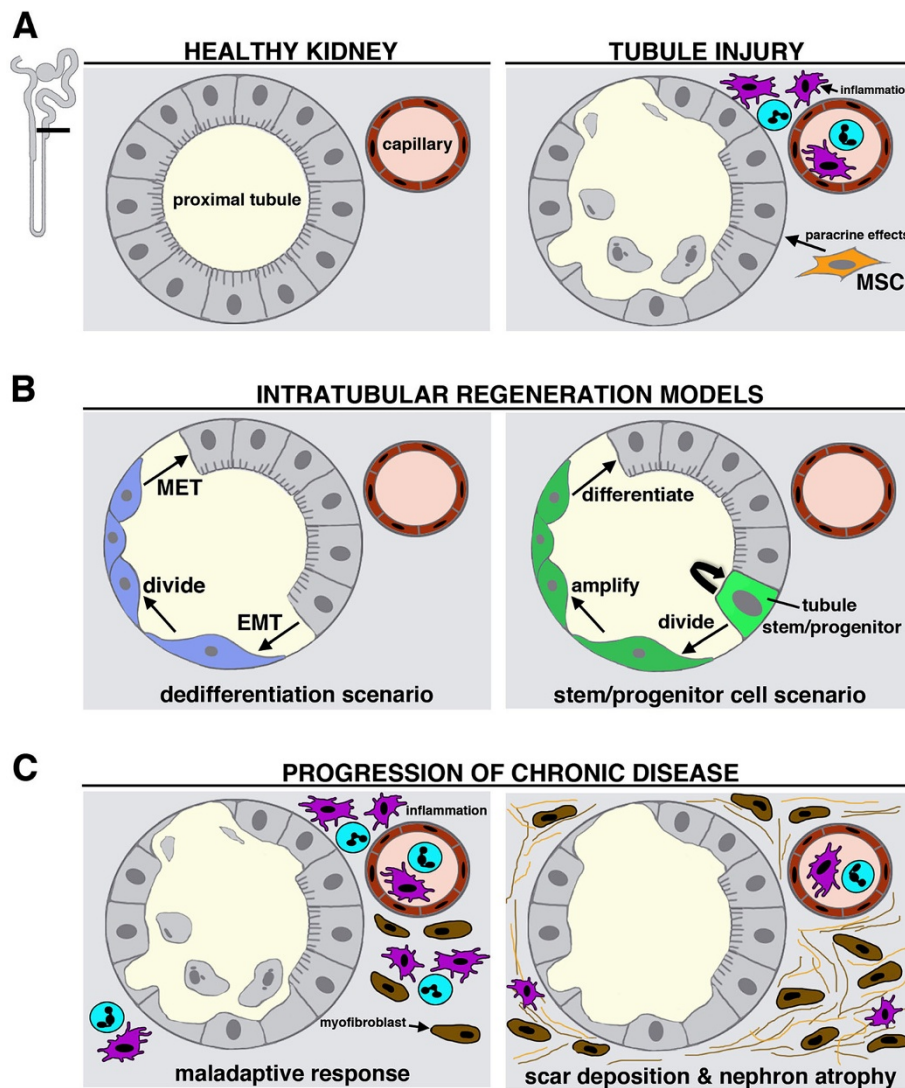
proliferation and/or hypertrophy. However, the ability of the kidney to regenerate varies widely following extensive direct injuries to cells within nephrons or the interstitial environment, as discussed in the next several sections that explore regeneration phenomena in the nephron tubule and glomerulus.

#### **Nephron tubule epithelium can be regenerated after AKI**

Studies in animal models dating back several decades have documented that AKI causes nephron tubule cell death and local inflammation, followed by high cell proliferation that restores tubule structure and function (Figure 4A). Proximal tubule cells are particularly sensitive to AKI from toxins, as they are intensely exposed to blood-borne compounds that enter the tubule after filtration, and from ischemic insults due to their high metabolism. In fact, AKI research has largely focused on the proximal S3 segment (see Figure 2A), which shows the highest rate of death subsequent to these types of acute insults. Seminal investigations using rat kidneys documented a massive rise in the tubular mitotic index and incorporation of thymidine- $H^3$  several days after mercuric chloride exposure [46]. Tubule repopulation was also observed by histology after toxic exposure to the antibiotic gentamicin [47]. Today, similar observations have been made in AKI models of ischemia reperfusion injury (IRI). Extensive proliferation in rat proximal tubules after IRI was reported based on elevated numbers of PCNA positive cells [48]. In BrdU pulse-chase studies after IRI in adult rats, nephron tubules showed increased numbers of scattered LRCs that were mostly PCNA positive [38]. Pathology studies of kidney samples from human AKI patients have also documented prominent numbers of intratubular PCNA positive cells [49], consistent with the notion that a sizeable proliferation response can occur in people after nephron damage. The explanation as to why patients differ widely in their ability to regain renal function after a bout of AKI is a resounding mystery, likely complicated by an intricate host of genetic factors, the environment, and age. Nevertheless, these studies indicate that mammalian nephron tubules (in principle) can harbor cells with a robust proliferation capacity that are sufficient in the right context to accomplish epithelial replacement.

#### **An intratubular cell source fuels nephron tubule epithelium regeneration after AKI**

The origin of tubular dividing cells after AKI has been debated between several scenarios (Figure 4B). In the first model, surviving tubular cells dedifferentiate and undergo an epithelial to mesenchymal transition (EMT) to become a proliferating, migratory populace [4]. The tubular cells proposed to undergo an EMT switch have included those tubular cells detected in G1 in uninjured nephrons [42]. Following sufficient division, the mesenchymal cells are



**Figure 4** Composition of nephron tubules in health and disease. (A) (Left) Healthy tubule with an intact epithelium. (Right) After injury, tubules show the loss of epithelial cells into the luminal space, which denudes the basement membrane and is accompanied by local inflammation. Mesenchymal stem cells (MSCs) provide restorative paracrine effects to damaged tubules. (B) Models of intratubular regeneration in AKI include (left) the dedifferentiation of endogenous differentiated tubule epithelium and (right) the activation of renal stem/progenitor cells. (C) (Left) In CKD, tubular damage is not restored and this is accompanied by a prolonged inflammatory response and the activation of myofibroblasts. (Right) Over time, lesions from myofibroblasts replace once-healthy renal tissue and this fibrosis is associated with permanent nephron erosion.

then proposed to differentiate into epithelium (MET) [4]. This model was based on the observations that surviving tubule cells had a flattened, squamous appearance [46,47], expressed prototypical mesenchymal markers [48], and lost polarity [50-52]. The idea of dedifferentiation was based on the expression of developmental genes by these squamous cells, including the transcription factor Paired box 2 (Pax2) [53], the neural cell adhesion molecule (NCAM) [54], and other nephrogenic genes [55]. Later, a second model was proposed in which the kidney received outside contributions from the bone marrow, which is home to multipotent hematopoietic stem cells

and mesenchymal stem cells (MSCs) [56,57]. In this scenario, bone marrow cells migrated to the damaged tubules, where they engrafted and transdifferentiated to make tubule epithelium [56,57].

Subsequent lineage tracing in murine IRI models has provided compelling evidence that nephron epithelial regeneration originates from an intratubular cell source [58-60]. Chimeric mice in which the bone marrow was labeled with either a transgenic marker (e.g. LacZ, eGFP) or harbored a Y-chromosome showed no contribution of cells with these labels to regenerating tubules [58]. Reciprocally, studies that labeled tubular cells by transgenic

methods showed that regenerated tubules contained only offspring with the marker and were not diluted with unmarked cells [59,60]. While these studies ruled out the scenario that bone marrow-derived cells directly contribute to nephrons, bone marrow cell contributions were confirmed in the renal interstitium [58,59]. There is now evidence that the MSCs promote nephron recovery after AKI through paracrine effects that prevent apoptosis and enhance proliferation (Figure 4A) [61-65]. This suggests that there may be a therapeutic benefit for MSC administration (see conclusions section).

#### **The nature of regenerating intratubular cells in AKI: differentiated, 'progenitor' or stem?**

Further studies of nephron tubule regeneration have interrogated whether the new cells are truly the progeny of surviving epithelia that dedifferentiated, or are instead the offspring of adult renal stem cells that reside within nephrons (Figure 4B). Adult stem cells are typically rare and possess features that unequivocally distinguish them from their differentiated descendants—usually, a combination of discrete morphological and molecular traits. Historically, the existence of tubular stem cells was greeted with considerable skepticism due to the lack of any histological or expression data that suggested nephrons had unique subsets of cells among the differentiated epithelial populace. The existence of tubular stem cells has been revisited in recent studies. Researchers have reexamined the morphological features of tubule cells, reassessed the origins of tubular cells after damage, and interrogated whether subsets of tubule cells expressed genes that mark adult stem cells in other tissues.

Conflicting information about the proliferation capacities of tubular cells in animal models has been collected with label retention and lineage tracing experiments. Through long pulse-chase experiments, rare tubular LRCs were seen in cortical nephrons of 2-month old rat kidneys when BrdU was administered just three days after birth—suggesting that healthy tubules contain cells that divide infrequently [66]. Others have captured a quite different proliferation dynamic following injury. When sequential pulses of two different DNA analogs were administered after IRI in mice, epithelial cells in damaged nephrons showed co-labeling frequencies consistent with stochastic division of many tubular cells [67]. Since high numbers of tubular cells were not co-labeled, the authors concluded that the repair was unlikely to involve a tubular stem cell [67]. However, it is possible that this conclusion is difficult to extrapolate because the timing parameters of this double pulse-chase did not rule out the possibility that a subset of intratubular cells proliferated quite rapidly just after injury. Indeed, another research group reported a specific subset of proximal tubule cells in mice that

expressed nuclear factor of activated T-cells cytoplasmic 1 (NFATc1) and proliferated extensively following epithelial injury from mercuric chloride exposure [68]. The discovery of the NFATc1+ proliferative tubule population is consistent with the notion that nephrons contain a particular subset of cells that divide after injury.

A recent body of evidence suggests human nephrons contain rare proliferative cells with stem cell features (Table 1) [69-73]. Stem cells express unique antigens and can be identified with antibodies that detect these markers. Cell surface antigens used for isolating renal stem cells are mostly based on known markers for other adult stem cells, and further studies are still needed to elucidate whether other markers could be used to identify renal stem cells correctly and efficiently [69-73]. The use of antigen strategies have led several groups to isolate tubule cells that exhibit impressive proliferation capacities *in vitro* and when administered to mice with AKI, which they termed 'renal tubular progenitors' [69-73].

Renal tubule progenitors were first reported based on a study that isolated tubule cells expressing the antigen CD133 (a marker of hematopoietic stem cells) and Pax2 from human renal cortex samples [69]. The CD133+ Pax2+ cells self-renewed and differentiated into tubule-like structures *in vitro*, and incorporated into tubules when injected into severe combined immunodeficiency (SCID) mice with glycerol-induced tubulonecrosis [69]. Another group isolated CD133+CD24+ cells from human cortical tissue and found that this fraction could expand, differentiate into renal epithelial cells, and make tubule-like structures *in vitro* [70]. The criterion of aldehyde dehydrogenase (ALDH) activity, which labels several adult stem cell types, has also been used to assess the kidney [71]. An ALDH<sup>high</sup> population was isolated from proximal tubules, and shown to express CD133+CD24+ and mesenchymal markers like vimentin (VIM) [71]. A recent study reported that a human CD133+CD24+CD106- tubular progenitor population, present in both proximal and distal segments, became the predominant regenerating population in both acute and chronic tubular damage patients [72]. When isolated and injected intravenously into SCID mice with rhabdomyolysis-induced AKI, human CD133+CD24+CD106- cells engrafted into the nephrons, generating tubular cells [72]. A panel of markers that may give further insight into tubular progenitor fractions has been collected, and several unique morphological traits of CD133+CD24+VIM+ human proximal tubule cells were also noted: namely, an absence of the brush border, reduced mitochondria number and reduced cytoplasm [73]. Analogous cells were not found in healthy rat kidneys, but were detected after unilateral ureteral obstruction (UUO), a damage model for renal fibrosis [73].

More research is needed to reconcile between the conflicting data concerning how nephrons are repaired:

**Table 1 Characterization of renal progenitors from adult human kidneys**

Origin & characteristics of human renal progenitor	<i>In vitro</i> capacity	<i>In vivo</i> renal injury model transplantation outcome	Reference
CD133+, Pax2+ from renal cortex	Self-renew, make tubular structures	Engrafted into tubules of SCID mice with glycerol-induced tubulonecrosis	[69]
CD133+, CD24+ in proximal & distal tubules	Differentiate into renal epithelium & make tubular structures	N.D.	[70]
Aldh <sup>high</sup> , CD133+, CD24+ from proximal tubule	N.D.	N.D.	[71]
CD133+, CD24+, CD106- in proximal & distal tubules	High proliferative capacity; differentiate into podocyte & tubular lineages	Engrafted into tubules of SCID mice with rhabdomyolysis induced AKI	[72]
CD133+, CD24+, VIM+, observed in proximal tubule	N.D.	N.D.	[73]

Discrete antigen combinations have isolated renal progenitors that show remarkable potency in both *in vitro* culture and transplantation settings as indicated.

namely whether differentiated tubular cells or renal progenitors are the source. This controversy has not been conclusively resolved. The evidence of renal tubule progenitors in human kidneys is quite compelling at present, but nevertheless remains to be substantiated in other mammalian kidneys. Additional work is needed to elucidate if the renal tubular progenitors are *bona fide* stem cells. Stem cells are operationally defined based on data indicating that they self-renew and make differentiated progeny *in vivo* during tissue homeostasis and/or injury. Lineage tracing to assess the clonogenicity of renal progenitors is needed to track their progeny in the kidney. Such studies will determine the most appropriate nomenclature for these cells. A current challenge is the need to identify specific promoters that can be used to make the necessary transgenic strains in the murine or other models. This will necessitate determining markers of these cells in animal models, though attractive candidates, such as NFATc1, already exist.

Further studies are also needed to delineate the relationship(s) and functional distinctions (if any) between renal progenitors that are positive for different antigen combinations. In addition, it will be interesting to reconcile whether or not these cells are poised in G1, or if this attribute is indicative of some other tubular fraction(s). As renal tubular progenitor subsets were found in both the proximal and distal tubule [72], it will also be interesting to determine the relationship and proliferation abilities of these cells across different nephron segments.

#### Nephron tubule damage and maladaptive regeneration in the setting of CKD

Although AKI research highlights the inherent reparative ability of the nephron tubule, this attribute is unable to counteract tubular damage in the CKD fibrotic micro-environment. CKD includes a heterogenous spectrum of clinical conditions. Nevertheless, CKD pathogenesis shares the characteristic of escalating fibrosis, often initiating with a primary injury (cell loss or abnormal behavior) located at the glomerulus or tubule (Figure 4C).

Fibrosis can proceed at the glomerulus (further discussed in the next section) or in the tubulointerstitial space. These lesions can propagate over many years, causing nephron dysfunction, atrophy, and collapse, coincident with damage to the vasculature that magnifies fibrogenesis and propagates a vicious damage cycle [74]. However, diabetic patients with pancreas transplants evinced amelioration of glomerular and interstitial CKD lesions after ten years [9,10]. Thus, renal scar regression was possible when patients were normoglycemic over a long period of time [9,10]. The mechanism(s) responsible for this remodeling remain unknown. Further, a recent clinical trial documented improved kidney function in ~3% of patients with hypertensive CKD over an extended time period (up to twelve years)—raising more questions about CKD prognosis [75]. Despite these reports, the most prevalent outcome for CKD patients is worsening renal function and ongoing nephron loss.

The sequence of kidney fibrosis has been likened to a wound healing process that goes terribly awry [76]. Renal cell damage is known to trigger the release of cytokines and other signals that trigger inflammation events from which myofibroblasts emerge. The origin of myofibroblasts has been quite contentious. Past candidates have included interstitial fibroblasts [76], tubular cells that undergo EMT [77-79], and endothelial cells [80,81]. Recent lineage tracing has indicated a pericyte/perivascular fibroblast origin [82].

Whether there are disease situations where myofibroblasts emerge from more than one cell compartment remains speculative. Elucidating the local cells and signals that influence myofibroblasts will be important to appreciate the cross talk that governs the progression of fibrosis in different settings. Understanding more about nephron tubular regeneration may provide viable clues as to how nephron loss can be diminished in CKD. In fact, abnormal tubular repair after AKI is thought to be one way to initiate CKD, and the severity of AKI is a robust predictor for the development of CKD [83]. There is also data from animal studies showing that tubular



regeneration capacity diminishes with repeated insults: mice subjected to a single round of targeted destruction of tubular cells in the S1 and S2 proximal segments could regenerate through extensive tubular proliferation, but three rounds of targeted injury led to varying degrees of interstitial fibrosis [84]. It is unknown if this is the case in the S3 and other tubular segments. These findings suggest that work is needed to address how tubular regenerative capacity changes over time, and how it is influenced by past kidney health and the general progression of aging. It remains puzzling why some renal insults are robustly countered with successful regeneration while other injuries lead to futile or actually more damaging cellular responses. While discovering ways to enhance diminished tubule regeneration may provide a viable avenue to protect against nephron loss, the amelioration of myofibroblast activity is still likely to be a keystone for CKD treatment.

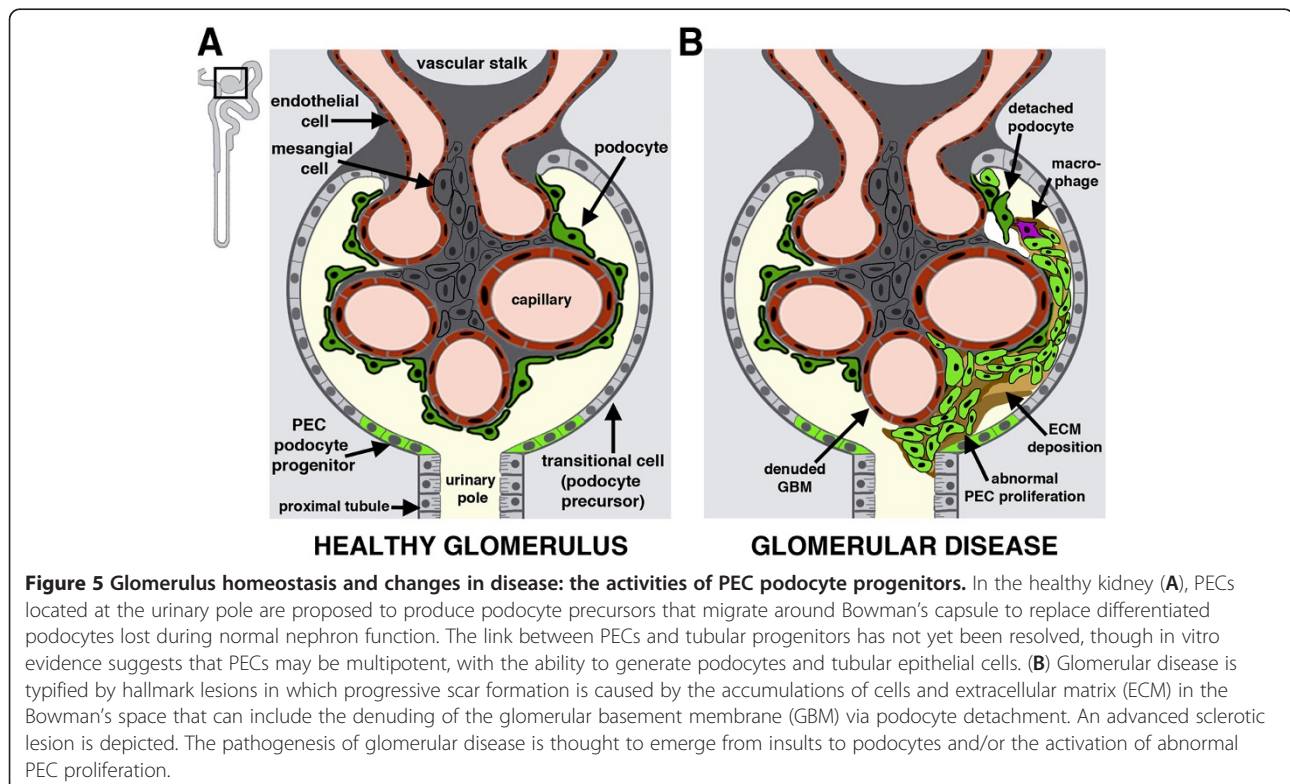
#### Glomerular activities of parietal epithelial (stem?) cells: roles in growth, podocyte turnover, and maladaptation in CKD

##### *The discovery of glomerular parietal epithelial cells with proliferative potential*

The glomerulus is comprised of at least four cell types: a perimeter of parietal epithelial cells (PECs) that form the extra-glomerular tuft, and a central glomerular tuft consisting of podocytes, endothelial cells, and mesangial

cells (Figure 5A). Recent studies have suggested that some PECs produce podocytes and exhibit stem cell features in several contexts [85-88]. Distinct subsets of PECs were first reported based on differential expression of CD24 and CD133 in human kidneys [85]. PECs expressing CD24+ and CD133+ were found in close proximity to the urinary pole, which is the junction between the glomerulus and tubule [85]. When isolated and grown in culture, CD24+CD133+ PECs exhibited high colony-forming capacity and self-renewal potential [85]. They could differentiate into cells that acquired markers associated with a variety of cell types (e.g. proximal or distal tubule cells, adipocytes, osteogenic cells or neurons) when grown in appropriate conditions [85]. Further, CD24+CD133+ PECs contributed to damaged tubules and restored kidney function when injected into SCID mice with rhabdomyolysis-induced AKI [85]. Next, a CD24+CD133+ PEC population was found at the urinary pole in developing human kidneys [86]. The embryonic CD24+CD133+ PECs had similar proliferative capacities as their adult counterparts with *in vitro* culture tests [86]. Interestingly, when they were tested with an *in vivo* xenograft administration assay, they displayed higher rates of tubular engraftment possibly reflecting enhanced regenerative potential [86].

Further scrutiny of the CD24+CD133+ PEC population based on expression of the podocyte marker podocalyxin (PDX) suggested a hierarchy of lineage



restriction: in culture, CD24+CD133+ PDX- PECs produced cells that expressed tubule or podocyte markers, while CD24+CD133+PDX+ PECs produced just podocytes [87]. Further, only CD24+CD133+PDX- PECs improved kidney function when administered to mice with adriamycin nephropathy, engrafting into glomerular and tubular structures [87]. Interestingly, a discrete spatial arrangement of these cells was noted in the glomerulus, with CD24+CD133+PDX- PECs at the urinary pole, CD24-CD133-PDX+ podocytes at the vascular pole, and another epithelial population situated in between that were CD24+CD133+PDX+ [87].

#### **The new working model of podocyte 'stem' cells**

These findings led to the proposal of a largely compelling model that PECs located at the urinary pole are in fact podocyte stem cells that upon division produce progenitors that migrate slowly around the capsule and eventually replace lost or damaged podocytes [87]. It is known that humans shed podocytes into the urine when the kidney is healthy, and patients with glomerular diseases excrete elevated numbers of podocytes (up to 400 fold more) [35]. Podocytes from the mouse and rat can undergo hypertrophy in culture in response to mechanical stress [88,89] and high glucose media [90], and human podocytes undergo hypertrophy in disease states such as diabetes [91,92]. However, hypertrophy is not thought to account for podocyte shedding in healthy kidneys, and podocytes themselves are post-mitotic based on pulse-chase studies [93]. Thus, the PEC stem cell model proffers an explanation that accounts for podocyte homeostasis over life. In support of the PEC stem cell model, genetic lineage data from mice tracked the ability of PECs located at the glomerular urinary pole to differentiate into podocytes during and after post-natal nephrogenesis [94]. These findings implicate PECs as the means for normal glomerular growth, and are consistent with the idea that PECs serve to maintain podocyte numbers. Additional lineage tracing in adults is needed to further support the model. Other studies that isolated self-renewing cell lines from human and mouse glomeruli reported that these cells express various developmental genes [95,96]. Uncovering the roles of these genes could provide useful mechanistic insights into PEC behavior.

There is experimental evidence that abnormal PEC proliferation after glomerular injury can lead to maladaptive glomerular lesions that elicit CKD [97,98] (Figure 5B). Pathologic changes can include hyperplastic lesions like glomerular crescents, which are accumulations of proliferating cells in the extracapillary space [97]. Over time, crescents can obstruct glomerular outflow to the tubule, leading to tubule degeneration and consequent nephron destruction. In addition, lineage tracing has shown that

activated PECs can participate in sclerotic lesions in which cells invade the capsule and deposit extracellular matrix leading to progressive glomerular scarring [98], which destroys the glomerulus and causes proteinuria that ultimately damages the nephron. Further work to discover the signals leading to abnormal PEC proliferation will be crucial for understanding and obviating maladaptive PEC behavior. Toward this end, some factors that influence PEC cell fate decisions during development have been identified, such as Wnt and BMP [99], and the modulation of the Notch pathway has been implicated in balancing glomerular cell proliferation and regeneration after injury [100].

The precedence that glomerular lesions can regress comes from biopsy analysis of diabetic patients who received pancreas transplants [9,10]. Clinical studies have shown that administration of angiotensin-converting enzyme (ACE) inhibitor reversed proteinuria and CKD progression in patients with non-diabetic chronic nephropathy [101,102] and in diabetics [103]. The mechanism for how ACE inhibition treatment triggers regression in humans is hard to address due to the difficulty in obtaining repeat biopsies. However, work in animal models showed that ACE inhibition correlated with structural changes in the glomerulus including podocyte repopulation [104] and reduced proliferation in sclerotic lesions [105]. These findings provide hope that it may be feasible to reverse, halt, or slow glomerular scarring. In turn, lessened or obviated glomerular scarring may prevent nephron atrophy.

#### **The renal papilla as a putative site for renal stem cells**

The renal papilla, or inner medulla, is a central region of the adult kidney that contains epithelial tracks of collecting ducts and nephron tubule intermediate segments (the loops of Henle) that concentrate urine (Figure 2). Whether the papilla serves as a niche for renal stem cells has been highly controversial. Evidence that the papilla may harbor renal stem cells originated from BrdU pulse-chase experiments: LRCs were found in papilla tubules and interstitial cells, and their number was reduced following ischemia, leading to the hypothesis that the LRCs might be involved in renal repair [66]. Plasticity of the papilla cells was tested *in vitro*, where some papilla cells proliferated to form spherical cell aggregates that co-expressed epithelial and mesenchymal markers [66]. The proliferation and migration of LRCs was also observed during normal homeostasis, and their number decreased with age, suggesting they may represent a renal stem cell pool responding to IRI [106]. Conflicting labeling results were also reported—namely that the papilla LRCs became quiescent early during renal organogenesis [107].

Meanwhile, the renal papilla was proposed to be a stem cell niche when marker-based approaches were

employed. Nestin is expressed in multi-lineage stem cells in brain and mesonephric mesenchyme [108]. By examining Nestin-GFP expression in transgenic mice, a large number of Nestin-expressing cells were detected as clusters in the renal papilla [108]. Nestin expression has been correlated with CD133, and Nestin+CD133+ cells were found extensively in the renal papilla [109]. The plasticity of these cells was tested through *in vitro* culture assays, where the cells showed the capacity for tubulogenesis [109].

Despite this body of evidence, not all approaches have supported the notion that the papillary region harbors renal stem cells that migrate to injured regions of the kidney. For example, the migration and proliferation of papillary LRCs after renal IRI was refuted by other BrdU pulse-chase experiments that failed to observe similar events [67]. In another report, the expression of telomerase reverse transcriptase (mTert) expression was surveyed after kidney injury because this gene marks embryonic and adult stem cells [110]. Expression of mTert could be detected in a subset of papillary epithelial cells, but this population neither divided nor migrated out of the renal papilla during repairing phases, suggesting they were not a progenitor-cell population [110]. While the existence of stem cells in the papilla remains an area of research filled with contradictory reports, it is reasonable to speculate that there must be some molecular mechanism(s) to support cell turnover and damage in this kidney region.

#### **The problem of resolving disparate reports of putative renal stem cell populations**

Many laboratories have reported finding kidney cells with stem cell characteristics that may correspond to one, several or even none of the cell types already discussed. Therefore, a significant and unresolved issue is whether the cells described by different researchers are distinct or identical to each other. In the following subsections we provide a short discussion of the information gathered about various putative renal stem cells based on gene expression data and/or assays of proliferative potential.

#### ***The establishment of renal stem cell lines from adult kidneys***

*In vitro* studies have been performed with LRCs isolated from adult ischemic rat kidneys after BrdU labeling, by virtue of their low Hoechst staining in the presence of BrdU [39]. In this damage context, LRCs are enriched in proximal tubules. The Hoechst<sup>low</sup>/BrdU<sup>+</sup> LRCs proliferated in culture, forming tubule-like structures in conditions that enabled tubulogenesis, and incorporating into nephrons and interstitium when cultured with developing rat metanephric kidneys [39]. After isolating single

cells from a healthy rat nephron, another group established a cell line that could expand greatly in culture and generate differentiated tubule cells in different culture conditions; this cell line could also engraft into rat kidney tubules after IRI [111]. However, due to the complex cellular composition of the nephron, the origin of this cell line is unclear. Another caveat in interpreting both of these studies is that the cells of origin are likely from the nephron, but could nonetheless be from other locales as well, such as the interstitium.

A multipotent population of nontubular cells was isolated from the mouse kidney based on the expression of stem cell antigen-1 (Sca-1) and the absence of lineage (Lin) markers of bone-marrow-derived stem cells [112]. Global gene expression analysis found enriched expression of mesodermal lineage genes, and that the Sca-1+ Lin- population could potentially contribute to renal repair after IRI [112]. Multipotent renal progenitor cells (MRPCs) isolated by culturing rat kidney cells showed high self-renewal capacity and expression of genes involved in kidney development, like Pax2 [113]. MRPC potency was demonstrated *in vitro*, as the cells were able to differentiate into myogenic, adipogenic, neural or osteogenic lineages; further, MRPCs differentiated into renal tubular cells after they were injected into the parenchyma of damaged murine kidneys [113]. Another group isolated what they termed mouse kidney progenitor cells (MKPC) from the interstitium of the medulla and papilla that exhibited self-renewal capacity and expressed early nephrogenesis genes [114]. Injection of MKPC rescued renal damage when administered to IRI damaged murine kidneys, and incorporation of MKPCs into some renal tubules was observed [114].

Interestingly, MSCs have been cultured from the renal capsule, which is a thin sheet of connective tissue that envelops the kidney and contains fibroblasts, adipocytes, and blood vessels [115]. After IRI, capsule cells migrated into the renal parenchyma, where they are postulated to participate in the injury response [115]. There have been other reports of MSCs found in adult kidneys, such as the isolation of MSCs able to differentiate into endothelial and smooth muscle cells that could promote vasculogenesis in IRI [116,117]. A limitation in interpreting all of these renal stem cell line studies is that culture conditions can have a significant impact on cellular behaviors that makes understanding the identity and potency of the cells somewhat convoluted in the absence of data to irrefutably track them *in vivo* during health or injury.

#### ***Kidney side population studies***

Stem cells can be isolated through flow cytometry due to their ability to efflux Hoechst 33342 dye and are characterized as a so-called side population (SP). Several SP

cells have been identified in various organs, such as heart, lung, and skeletal muscle. Kidney SP cells from the mouse [118,119] and rat [120] have been characterized, and expression of particular genes like *Musculin/MyoR* has been annotated [119]. Another group isolated a renal SP population located in the proximal tubule, supporting a 'tubular niche' hypothesis [121]. These SP cells were highly heterogeneous, although they showed a humoral role during renal repair without tubular integration [121]. The existence of SP cells in adult human kidneys has also been reported [122,123]. However, *in vitro* characterization demonstrated the heterogeneity of human renal SP population as well, indicating SP cells cannot be equally considered as stem cells [122]. The ability of SP cells to improve renal regeneration has been reported, and the mechanism behind this ameliorative role may be due to their generation of bone morphogenic protein 7 (BMP7), which reversed chronic renal injury when given exogenously and could be induced by treatment with a histone deacetylase inhibitor [124]. Taken together, whether kidney SP cells represent a particular stem cell population still remains controversial because renal SP cells show considerable heterogeneity. To date, the SP assay has not served to unequivocally identify kidney stem cells, and future work is needed to assess if podocyte or tubular progenitors exhibit the SP phenotype, or can be enriched using this parameter.

#### Prospects for future research

It is clear that there are many gaps in our understanding about tubular, glomerular, and other speculated stem cell fractions in the kidney. However, there are several promising research venues that may provide novel insights into how regenerative kidney cells work. One such area is the study of renal development. Developmental studies are applicable for understanding regeneration because information about how kidney lineages are formed can provide gene expression data that could help pinpoint adult renal stem cells and the signaling pathways that modulate the mobilization and behavior of reparative cells. Further, knowledge from studying embryonic progenitors during kidney development may be directly relevant to adult cell types, as multipotent renal progenitors isolated from adult kidneys could represent a subset of embryonic progenitors that persist from early nephrogenesis.

Another promising topic concerns how kidney cells sense injury and interact with their environment. Understanding how kidney cells perceive and react to local damage may provide critical information about the early events in kidney disease. A fundamental player is likely to be the primary cilium [125]. Cilia are organelles that play crucial roles in how cells interact with their environment. Two types of cilia have been documented in

vertebrates, motile cilia and the non-motile primary cilium [126]. Primary cilia are chemical and mechanosensors between the cell and its milieu [126]. In healthy adult human kidneys, the epithelial cells lining the lumen of nephrons and collecting ducts have primary cilia with a 9+0 microtubule structure that interact with urinary flow [127]. Cilia defects cause cystic kidney diseases, characterized by abnormalities in nephron tubular epithelial cells that lead to their overproliferation and development of fluid-filled cysts. Since changes in urinary flow during AKI and CKD are likely sensed by tubule residents through the primary cilium, this signal transduction may have important ramifications for how tubular cells recognize injury. Thus, primary cilium signaling may hold a prominent position in the course of events that lead to nephron regeneration or maladaptive responses.

#### Conclusions

New therapeutic options are needed to mitigate kidney disease. The intricate cellular composition of the kidney poses challenges both to understanding and treating kidney disease. The presence of many renal cell types likely means that the microenvironments in the kidney are manifold and diverse. Nevertheless, the discovery of regenerative renal cells in adult kidneys—especially those in the nephron tubule and PECs in the glomerulus—provides hope that there are already cell templates and conducive environs where kidney integrity can be restored by promoting regenerative mechanisms. Improvements in the detection of specific kidney diseases with diagnostic biomarkers that can reliably predict tissue alterations are crucial in the years ahead. Emerging evidence supports the notion that renal progenitors are a conserved evolutionary phenomenon, and novel future insights are likely to be gleaned from the analysis of renal progenitors across the animal kingdom [128].

One important element of creating regenerative therapeutics will involve the discovery of how to deliver and control such interventions in time and space, as cautioned by the ability of unchecked PEC proliferation to initiate glomerular lesions. Phenotypic improvements triggered by alterations in the microenvironment, as exemplified by the paracrine effects of MSCs when they lodge in renal parenchyma, may be a viable place to start. Many clinical trials with stem cell therapies are currently underway, both with MSCs and other cell types [129]. There have been encouraging outcomes with the safety of using autologous transplants, which provides preliminary reassurance that the efficacy of administering mobilized MSCs (or other cells) to ameliorate human kidney disease may be testable in the near future. Further, in experimental models of CKD, various studies have shown that BMP7 can inhibit or even reverse



fibrosis, which raises the possibility that targeting the BMP7 pathway may provide one avenue to help patients with CKD [130].

Considerations of the appropriate target patient population will also need to weigh more than the kidney disease diagnosis: variables like age and the burden of co-morbidities will also be factors that come into play. Like other body organs, the aging kidney shows dramatic alterations [131]. For example, it has been appreciated for decades that proliferative capacity of the mammalian kidney declines with age [132]. However, recent experimental studies have documented that young bone marrow cells from mice could alleviate aspects of renal aging when administered to old mice [133]. This suggests that therapeutic options may be enhanced as parameters like cellular age are considered and explored.

Taken together, recent research has uncovered enticing data that several cell types within the kidney have innate regenerative capacity. It will be essential to understand how to enhance or induce regeneration in a controlled fashion that can prevent maladaptive outcomes arising from perturbed cell behavior. Although this may turn out to be a complicated line to walk, the existence of several stem cells in the kidney provides encouragement that regenerative approaches might be harnessed to combat renal disease in the not-so-distant future.

#### Abbreviations

ACE: Angiotensin-converting-enzyme; AKI: Acute kidney injury; ALDH: Aldehyde dehydrogenase; BMP7: Bone morphogenic protein 7; BrdU: 5'-bromo-2'-deoxyuridine; CKD: Chronic kidney disease; CM: Cap mesenchyme; ECM: Extracellular matrix; EMT: Epithelial to mesenchymal transition; ES cell: Embryonic stem cell; ESRD: End-stage renal disease; GBM: Glomerular basement membrane; IM: Intermediate mesoderm; iPS cell: Induced pluripotent stem cell; IRI: Ischemia reperfusion injury; Lin: Lineage; LRC: Label retaining cell; MET: Mesenchymal to epithelial transition; MKPC: Mouse kidney progenitor cell; MM: Metanephric mesenchyme; MRPC: Multipotent renal progenitor cell; MSC: Mesenchymal stem cell; NCAM: Neural cell adhesion molecule; NFATc1: Nuclear factor of activated T-cells cytoplasmic 1; Pax2: Paired box 2; PCNA: Proliferating cell nuclear antigen; PDX: Podocalyxin; PEC: Parietal epithelial cell; Sca-1: Stem cell antigen-1; SCID: Severe combined immunodeficiency; SP: Side population; mTERT: Telomerase reverse transcriptase; UB: Ureteric bud; UUU: Unilateral ureteral obstruction; VIM: Vimentin.

#### Competing interest

The authors declare that they have no commercial or other competing interests to disclose.

#### Authors' contributions

The authors YL and RAW contributed equally to the research, writing, and revisions of the manuscript. RAW created and revised the figures. Both authors read and approved the final manuscript.

#### Acknowledgements

The work of the Wingert laboratory is supported by the following: NIH-NIDDK grant K01DK083512, NIH New Innovator award grant DP2OD008470, the March of Dimes Basil O'Connor Starter Scholar grant award #5-FY12-75, start up funds from the University of Notre Dame College of Science and Department of Biological Sciences, and a generous gift to the University of Notre Dame from Elizabeth and Michael Gallagher on behalf of the Gallagher Family to support adult stem cell research. We are grateful to Christina Cheng, Gary Gerlach, Kristen McCampbell, and Robert McKee for

their critical review and suggestions on the manuscript. We thank the staffs of the Department of Biological Sciences and the Center for Zebrafish Research for their ongoing support. Finally, we wish to thank all the members of our wonderful research lab for engaging in stimulating conversations about this topic.

Received: 25 February 2013 Accepted: 14 May 2013

Published: 21 May 2013

#### References

1. United States Renal Data System: *2012 Annual Data Report*. Atlas of Chronic Kidney Disease and End-Stage Renal Disease in the United States; 2012. <http://www.usrds.org/adr.aspx>.
2. Weiner DE: **Public health consequences of chronic kidney disease**. *Clin Pharmacol Ther* 2009, **86**:566–569.
3. Schedl A: **Renal abnormalities and their developmental origin**. *Nat Rev Genet* 2007, **8**:791–802.
4. Bonventre JV, Yang L: **Cellular pathophysiology of ischemic acute kidney injury**. *J Clin Invest* 2011, **121**:4210–4221.
5. Murugan R, Kellum JA: **Acute kidney injury: what's the prognosis?** *Nat Rev Nephrol* 2011, **7**:209–217.
6. Venkatachalam MA, Griffin KA, Lan R, Geng H, Saikumar P, Bidani AK: **Acute kidney injury: a springboard for progression in chronic kidney disease**. *Am J Physiol Renal Physiol* 2010, **298**:F178–F1094.
7. El Nahas AM, Bello AK: **Chronic kidney disease: the global challenge**. *Lancet* 2005, **365**:331–340.
8. Stocum DL: **Stem cells in regenerative biology and medicine**. *Wound Rep Reg* 2001, **9**:429–442.
9. Fioretto P, Steffes MW, Sutherland DER, Goetz FC, Mauer M: **Reversal of lesions of diabetic nephropathy after pancreas transplantation**. *N Engl J Med* 1998, **339**:69–75.
10. Fioretto P, Sutherland DER, Najafian B, Mauer M: **Remodeling of renal interstitial and tubular lesions in pancreas transplant recipients**. *Kidney Int* 2006, **69**:907–910.
11. Weissman IL: **Stem cells: units of development, units of regeneration, and units in evolution**. *Cell* 2000, **100**:157–168.
12. Yamanaka S, Blau HM: **Nuclear reprogramming to a pluripotent state by three approaches**. *Nature* 2010, **465**:704–712.
13. Hsu YC, Fuchs E: **A family business: stem cell progeny join the niche to regulate homeostasis**. *Nat Rev Mol Cell Biol* 2012, **13**:103–114.
14. Burness ML, Sipkins DA: **The stem cell niche in health and malignancy**. *Semin Cancer Biol* 2010, **20**:107–115.
15. Sharpless NE, DePinho RA: **How stem cells age and why this makes us grow old**. *Nat Rev Mol Cell Biol* 2007, **8**:703–713.
16. McCampbell KK, Wingert RA: **Renal stem cells: fact or science fiction?** *Biochem J* 2012, **444**:153–168.
17. Reilly RF, Bulger RE, Kriz W, In Diseases of the Kidney and Urinary Tract, Eighth Edition: *Structural-functional relationships in the kidney*. Philadelphia: Lippincott Williams & Wilkins; Edited by Schrier RW; 2007:2–53.
18. Little MH, Brennan J, Georgas K, Davies JA, Davidson DR, Baldock RA, et al: **A high-resolution anatomical ontology of the developing murine genitourinary tract**. *Gene Expr Patterns* 2007, **7**:680–699.
19. Brunskill EW, Aronow BJ, Georgas K, Rumballe B, Valerius MT, Aronow J, et al: **Atlas of gene expression in the developing kidney at microanatomic resolution**. *Dev Cell* 2008, **15**:781–791.
20. Mugford JW, Yu J, Kobayashi A, McMahon AP: **High-resolution gene expression analysis of the developing mouse kidney defines novel cellular compartments within the nephron progenitor population**. *Dev Biol* 2009, **333**:312–323.
21. Yu J, Valerius MT, Duah M, Staser K, Hansard JK, Guo JJ, et al: **Identification of molecular compartments and genetic circuitry in the developing mammalian kidney**. *Development* 2012, **139**:1863–1873.
22. Wingert RA, Davidson AJ: **The zebrafish pronephros: a model to study nephron segmentation**. *Kidney Int* 2008, **73**:1120–1127.
23. Nyengaard JR, Bendtsen TF: **Glomerular number and size in relation to age, kidney weight, and body surface in normal man**. *Anat Rec* 1992, **232**:194–201.
24. Hughson M, Farris III: **AB, Douglas-Denton R, Yoy WE, Bertram JF: Glomerular number and size in autopsy kidneys: the relationship to birth weight**. *Kidney Int* 2003, **63**:2113–2122.

25. Potter EL, Thierstein ST: Glomerular development in the kidney as an index of fetal maturity. *J Pediatr* 1943, **22**:695–706.
26. Hartman HA, Lai HL, Patterson LT: Cessation of renal morphogenesis in mice. *Dev Biol* 2007, **301**:379–387.
27. Solomon S: Developmental changes in nephron number, proximal tubule length and superficial nephron glomerular filtration rate of rats. *J Physiol* 1977, **272**:573–589.
28. Wesson LG: Compensatory growth and other growth responses of the kidney. *Nephron* 1989, **51**:149–184.
29. de Rouffignac C, Monnens L: Functional and morphologic maturation of superficial and juxtamedullary nephrons in the rat. *J Physiol* 1976, **262**:119–129.
30. Spitzer A, Brandis M: Functional and morphologic maturation of the superficial nephrons. *J Clin Invest* 1974, **53**:279–287.
31. Blanpain C, Fuchs E: Epidermal homeostasis: a balancing act of stem cells in the skin. *Nat Rev Mol Cell Bio* 2009, **10**:207–217.
32. van der Flier LG, Clevers H: Stem cells, self-renewal, and differentiation in the intestinal epithelium. *Annu Rev Physiol* 2009, **71**:241–260.
33. Prescott LF: The normal urinary excretion rates of renal tubular cells, leucocytes and red blood cells. *Clin Sci* 1966, **31**:425–435.
34. Dörrenhaus A, Müller JIF, Golka K, Jedrusik P, Schulze H, Föllmann W: Cultures of exfoliated epithelial cells from different locations of the human urinary tract and the renal tubular system. *Arch Toxicol* 2000, **74**:618–626.
35. Vogelmann SU, Nelson WJ, Myers BD, Lemley KV: Urinary excretion of viable podocytes in health and renal disease. *Am J Physiol Renal Physiol* 2003, **285**:F40–F48.
36. Rahmoune H, Thompson PW, Ward JM, Smith CD, Hong G, Brown J: Glucose transporters in human renal proximal tubular cells isolated from the urine in patients with non-insulin-dependent diabetes. *Diabetes* 2005, **54**:3427–3434.
37. Messier B, Leblond CP: Cell proliferation and migration as revealed by radioautography after injection of thymidine-H3 into male rats and mice. *Amer J Anat* 1960, **106**:247–285.
38. Maeshima A, Yamashita S, Nojima Y: Identification of renal progenitor-like tubular cells that participate in the regeneration processes of the kidney. *J Am Soc Nephrol* 2003, **14**:3138–3146.
39. Maeshima A, Sakurai H, Nigam SK: Adult kidney tubular cell population showing phenotypic plasticity, tubulogenic capacity, and integration capability into developing kidney. *J Am Soc Nephrol* 2006, **17**:188–198.
40. Vogetseder A, Karadeniz A, Kaissling B, Le Hir M: Tubular cell proliferation in the healthy rat kidney. *Histochem Cell Biol* 2005, **124**:97–104.
41. Vogetseder A, Palan T, Bacic D, Kaissling B, Le Hir M: Proximal tubular epithelial cells are generated by division of differentiated cells in the healthy kidney. *Am J Physiol Cell Physiol* 2007, **292**:C807–C813.
42. Vogetseder A, Picard N, Gaspert A, Walch M, Kaissling B, Le Hir M: Proliferation capacity of the renal proximal tubule involves the bulk of differentiated epithelial cells. *Am J Physiol Cell Physiol* 2008, **294**:C22–C28.
43. Nadasdy T, Laszik Z, Blick KE, Johnson LD, Silva FG: Proliferative activity of intrinsic cell populations in the normal human kidney. *J Am Soc Nephrol* 1994, **4**:2032–2039.
44. Hayslett JP, Kashgarian M, Epstein FH: Functional correlates of compensatory renal hypertrophy. *J Clin Invest* 1968, **47**:774–781.
45. Hostetter TH: Progression of renal disease and renal hypertrophy. *Annu Rev Physiol* 1995, **57**:263–278.
46. Cuppage FE, Tate A: Repair of the nephron following injury with mercuric chloride. *Am J Pathol* 1967, **51**:405–429.
47. Houghton DC, Hartnett M, Campbell-Boswell M, Porter G, Bennett W: A light and electron microscopic analysis of gentamicin nephrotoxicity in rats. *Am J Pathol* 1976, **82**:589–612.
48. Witzgall R, Brown D, Schwarz C, Bonventre JV: Localization of proliferating cell nuclear antigen, vimentin, c-Fos and clusterin in the postischemic kidney. Evidence for a heterogenous genetic response among nephron segments, and a large pool of mitotically active and dedifferentiated cells. *J Clin Invest* 1994, **93**:2175–2188.
49. Nadasdy T, Laszik Z, Blick KE, Johnson DL, Burst-Singer K, Nast C, et al: Human acute tubular necrosis: a lectin and immunohistochemical study. *Hum Pathol* 1995, **26**:230–239.
50. Molitoris BA, Wilson PD, Schrier RW, Simon FR: Ischemia induces partial loss of surface membrane polarity and accumulation of putative calcium ionophores. *J Clin Invest* 1985, **76**:2097–2105.
51. Molitoris BA, Hoilien CA, Ahnen DJ, Wilson PD, Kim J: Characterization of ischemia-induced loss of epithelial polarity. *J Membr Biol* 1988, **106**:233–242.
52. Zuk A, Bonventre JV, Brown D, Matlin KS: Polarity, integrin, and extracellular matrix dynamics in the postischemic rat kidney. *Am J Physiol* 1998, **273**:C711–C731.
53. Imgrund M, Gröne E, Gröne HJ, Kretzler M, Holzman L, Schlöndorff D, et al: Re-expression of the developmental gene Pax-2 during experimental acute tubular necrosis in mice. *Kidney Int* 1999, **56**:1423–1431.
54. Abbate M, Brown D, Bonventre JV: Expression of NCAM recapitulates tubulogenic development in kidneys recovering from acute ischemia. *Am J Physiol* 1999, **277**:F454–F463.
55. Villanueva S, Cespedes C, Vio CP: Ischemic acute renal failure induces the expression of a wide range of nephrogenic proteins. *Am J Physiol Regul Integr Comp Physiol* 2006, **290**:R861–870.
56. Lin F, Cordes K, Li L, Hood L, Couser WG, Shankland SJ, et al: Hematopoietic stem cells contribute to the regeneration of renal tubules after renal ischemia-reperfusion injury in mice. *J Am Soc Nephrol* 2003, **14**:1188–1199.
57. Kale S, Karihaloo A, Clark PR, Kashgarian M, Krause DS, Cantley LG: Bone marrow stem cells contribute to repair of the ischemically injured renal tubule. *J Clin Invest* 2003, **112**:42–49.
58. Duffield JS, Park KM, Hsiao LL, Kelley VR, Scadden DT, Ichimura T, et al: Restoration of tubular epithelial cells during repair of the postischemic kidney occurs independently of bone marrow-derived stem cells. *J Clin Invest* 2005, **115**:1743–1755.
59. Lin F, Moran A, Igarashi P: Intrarenal cells, not bone marrow-derived cells, are the major source for regeneration in the postischemic kidney. *J Clin Invest* 2005, **115**:1756–1764.
60. Humphreys BD, Valerius MT, Kobayashi A, Mugford JW, Soeung S, Duffield JS, et al: Intrinsic epithelial cells repair the kidney after injury. *Cell Stem Cell* 2008, **2**:284–291.
61. Morigi M, Imberti B, Zoja C, Corna D, Tomasoni S, Abbate M, et al: Mesenchymal stem cells are renotropic, helping to repair the kidney and improve function in acute renal failure. *J Am Soc Nephrol* 2004, **15**:1794–1804.
62. Herrera MB, Bussolati B, Bruno S, Fonsato V, Romanazzi GM, Camussi G: Mesenchymal stem cells contribute to the renal repair of acute tubular epithelial injury. *Int J Mol Med* 1994, **14**:1035–1041.
63. Tögel F, Hu Z, Weiss K, Isaac J, Lange C, Westenfelder C: Administered mesenchymal stem cells protect against ischemic acute renal failure through differentiation-independent mechanisms. *Am J Physiol Renal Physiol* 2005, **289**:F31–F42.
64. Lange C, Tögel F, Itrich H, Clayton F, Nolte-Ernsting C, Zander AR, et al: Administered mesenchymal stem cells enhance recovery from ischemia/reperfusion-induced acute renal failure in rats. *Kidney Int* 2005, **68**:1613–1617.
65. Bi B, Schmitt E, Israilova M, Nishio H, Cantley LG: Stromal cells protect against acute tubular injury via an endocrine effect. *J Am Soc Nephrol* 2007, **18**:2486–2496.
66. Oliver JA, Maarouf O, Cheema FH, Martens TP, Al-Awqati Q: The renal papilla is a niche for adult kidney stem cells. *J Clin Invest* 2004, **114**:795–804.
67. Humphreys BD, Czerniak S, DiRocco DP, Hsain W, Cheema R, Bonventre JV: Repair of injured proximal tubule does not involve specialized progenitors. *Proc Natl Acad Sci, USA* 2011, **108**:9226–9231.
68. Langworthy M, Zhou B, de Caestecker M, Moeckel G, Baldwin HS: NFATc1 identifies a population of proximal tubule cell progenitors. *J Am Soc Nephrol* 2009, **20**:311–321.
69. Bussolati B, Bruno S, Grange C, Buttiglieri S, Deregibus MC, Cantino D, et al: Isolation of renal progenitor cells from adult human kidney. *Am J Pathol* 2005, **166**:545–555.
70. Sallustio F, De Benedictis L, Castellano G, Zaza G, Loverre A, Constantino V, et al: TLR2 plays a role in the activation of human resident renal stem/progenitor cells. *FASEB J* 2010, **24**:514–525.
71. Lindgren D, Boström AK, Nilsson K: Hansson, Sjölund J, Möller C, et al: Isolation and characterization of progenitor-like cells from human renal proximal tubules. *Am J Pathol* 2011, **178**:828–837.
72. Angelotti ML, Ronconi E, Ballerini L, Peired A, Mazzinghi B, Sagrinati C, et al: Characterization of renal progenitors committed toward tubular lineage and their regenerative potential in renal tubular injury. *Stem Cells* 2012, **30**:1714–1725.
73. Smeets B, Boor P, Jijkman H, Sharma SV, Jirak P, Mooren F, et al: Proximal tubular cells contain a phenotypically distinct, scattered cell population involved in tubular regeneration. *J Pathol*. 2013, **229**:645–659.

74. Liu Y: Cellular and molecular mechanisms of renal fibrosis. *Nat Rev Nephrol* 2011, **7**:684–696.
75. Hu B, Gadegbeku C, Lipkowitz MS, Rostand S, Lewis J, Wright JT, et al: Kidney function can improve in patients with hypertensive CKD. *J Am Soc Nephrol* 2012, **23**:706–713.
76. Hirschberg R: Wound healing in the kidney: complex interactions in renal interstitial fibrogenesis. *J Am Soc Nephrol* 2005, **16**:9–11.
77. Yang J, Liu Y: Dissection of key events in tubular epithelial to myofibroblast transition and its implications in renal interstitial fibrosis. *Am J Pathol* 2001, **159**:1465–1475.
78. Iwano M, Plieth D, Danoff TM, Xiu C, Okada H, Neilson EG: Evidence that fibroblasts derive from epithelium during tissue fibrosis. *J Clin Invest* 2002, **110**:341–350.
79. Yamashita S, Maeshima A, Nojima Y: Involvement of renal progenitor tubular cells in epithelial-to-mesenchymal transition in fibrotic rat kidneys. *J Am Soc Nephrol* 2005, **16**:2044–2051.
80. Zeisberg EM, Potenta SE, Sugimoto H, Zeisberg M, Kalluri R: Fibroblasts in kidney fibrosis emerge via endothelial-to-mesenchymal transition. *J Am Soc Nephrol* 2008, **19**:2282–2287.
81. Li J, Qu X, Bertram JF: Endothelial-myofibroblast transition contributes to the early development of diabetic renal interstitial fibrosis in streptozotocin-induced diabetic mice. *Am J Pathol* 2009, **175**:1380–1388.
82. Humphreys BD, Lin SL, Kobayashi A, Hudson TE, Nowlin BT, Bonventre JV, et al: Fate tracing reveals the pericyte and not epithelial origin of myofibroblasts in kidney fibrosis. *Am J Pathol* 2010, **176**:85–97.
83. Chawla LS, Amdur RL, Amodeo S, Kimmel PL, Palant CE: The severity of acute kidney injury predicts progression to chronic kidney disease. *Kidney Int* 2011, **79**:1361–1369.
84. Grgic I, Campanholle G, Bijol V, Wang C, Sabbisetti VS, Ichimura T, et al: Targeted proximal tubule injury triggers interstitial fibrosis and glomerulosclerosis. *Kidney Int* 2012, **82**:172–183.
85. Sagrinati C, Netti GS, Mazzinghi B, Lazzeri E, Liotta F, Frosali F, et al: Isolation and characterization of multipotent progenitor cells from the Bowman's capsule of adult human kidneys. *J Am Soc Nephrol* 2006, **17**:2443–2456.
86. Lazzeri E, Crescioli C, Ronconi E, Mazzinghi B, Sagrinati C, Netti GS, et al: Regenerative potential of embryonic renal multipotent progenitors in acute renal failure. *J Am Soc Nephrol* 2007, **18**:3128–3138.
87. Ronconi E, Sagrinati C, Angelotti ML, Lazzeri E, Mazzinghi B, Ballerini L, et al: Regeneration of glomerular podocytes by human renal progenitors. *J Am Soc Nephrol* 2009, **20**:322–332.
88. Kriz W: Podocyte hypertrophy mismatch and glomerular disease. *Nat Rev Nephrol* 2012, **8**:618–619.
89. Petermann AT, Pippin J, Durvasula R, Pichler R, Hiromura K, Monkawa T, et al: Mechanical stretch induces podocyte hypertrophy in vitro. *Kidney Int* 2005, **67**:157–166.
90. Xu ZG, Yoo TH, Ryu DR, Park HC, Ha SK, Han DS, et al: Angiotensin II receptor blocker inhibits p27<sup>Kip1</sup> expression in glucose-stimulated podocytes and diabetic glomeruli. *Kidney Int* 2005, **67**:944–952.
91. Osterby R, Gundersen HJ: Glomerular size and structure in diabetes mellitus. I. Early abnormalities. *Diabetologia* 1975, **11**:225–229.
92. Gundersen HJ, Osterby R: Glomerular size and structure in diabetes mellitus. II. Late abnormalities. *Diabetologia* 1977, **13**:43–38.
93. Pabst R, Sterzel RB: Cell renewal of glomerular cell types in normal rats. An autoradiographic analysis. *Kidney Int* 1983, **24**:626–631.
94. Appel D, Kershaw DB, Smeets B, Yuan G, Fuss A, Frye B, et al: Recruitment of podocytes from glomerular parietal epithelial cells. *J Am Soc Nephrol* 2009, **20**:333–343.
95. Bruno S, Bussolati B, Grange C, Collino F, de Cantogno LV, Herrera MB, et al: Isolation and characterization of resident mesenchymal stem cells in human glomeruli. *Stem Cells Dev* 2009, **18**:867–879.
96. Swetha G, Chandra V, Phadnis S, Bhonde R: Glomerular parietal epithelial cells of adult murine kidney undergo EMT to generate cells with traits of renal progenitors. *J Cell Mol Med* 2011, **15**:396–413.
97. Smeets B, Angelotti ML, Rizzo P, Dijkman H, Lazzeri E, Mooren F, et al: Renal progenitor cells contribute to hyperplastic lesions of podocytopathies and crescentic glomerulonephritis. *J Am Soc Nephrol* 2009, **20**:2593–2603.
98. Smeets B, Uhlig S, Fuss A, Mooren F, Wetzels JFM, Floege J, et al: Tracing the origin of glomerular extracapillary lesions from parietal epithelial cells. *J Am Soc Nephrol* 2009, **20**:2604–2615.
99. Smeets B, Moeller MJ: Parietal epithelial cells and podocytes in glomerular diseases. *Semin Nephrol* 2012, **32**:357–367.
100. Lasagni L, Ballerini L, Angelotti ML, Parente E, Sagrinati C, Mazzinghi B, et al: Notch activation differentially regulates renal progenitors proliferation and differentiation toward the podocyte lineage in glomerular disorders. *Stem Cells* 2010, **28**:1673–1685.
101. Ruggenenti P, Perna A, Gherardi G, Gaspari F, Benini R, Remuzzi G, et al: Renal function and requirement for dialysis in chronic nephropathy patients on long-term ramipril: REIN follow-up trial. *Lancet* 1998, **352**:1252–1256.
102. Ruggenenti P, Perna A, Benini R, Bertani T, Zoccali C, Maggiore Q, et al: In chronic nephropathies prolonged ACE inhibition can induce remission: dynamics of time-dependent changes in GFR. *J Am Soc Nephrol* 1999, **10**:997–1006.
103. Wilmer WA, Hebert LA, Lewis EJ, Rohde RD, Whittier F, Cattran D, et al: Remission of nephrotic syndrome in type 1 diabetes: long-term follow-up of patients in the captopril study. *Am J Kidney Dis* 1999, **34**:308–314.
104. Macconi D, Sangalli F, Bonomelli M, Conti S, Condorelli L, Gagliardini E, et al: Podocyte repopulation contributes to regression of glomerular injury induced by ACE inhibition. *Am J Pathol* 2009, **174**:797–807.
105. Benigni A, Morigi M, Rizzo P, Gagliardini E, Rota C, Abbate M, et al: Inhibiting angiotensin-converting enzyme promotes renal repair by limiting progenitor cell proliferation and restoring glomerular architecture. *Am J Pathol* 2011, **179**:628–638.
106. Oliver JA, Lkinakis A, Cheema FH, Friedlander J, Sampogna RV, Martens TP, et al: Proliferation and migration of label-retaining cells of the kidney papilla. *J Am Soc Nephrol* 2009, **20**:2315–2327.
107. Adams DC, Oxburgh L: The long-term label retaining population of the renal papilla arises through divergent regional growth of the kidney. *Am J Physiol Renal Physiol* 2009, **297**:F809–F815.
108. Patschan D, Michurina T, Shi HK, Dolff S, Brodsky SV, Cohen-Gould L, et al: Normal distribution and medullary-to-cortical shift of Nestin-expressing cells in acute renal ischemia. *Kidney Int* 2007, **71**:744–754.
109. Ward HH, Romero E, Welford A, Pickett G, Bacallao R, Gattone VH II, et al: Adult human CD133/1+ kidney cells isolated from renal papilla integrate into developing kidney tubules. *Biochim Biophys Acta* 1812, **2011**:1344–1357.
110. Song J, Czerniak S, Wang T, Ying W, Carlone DL, Breault DT, et al: Characterization and fate of telomerase-expressing epithelial during kidney repair. *J Am Soc Nephrol* 2011, **22**:2256–2265.
111. Kitamura S, Yamasaki Y, Kinomura M, Sugaya T, Sugiyama H, Maeshima Y, et al: Establishment and characterization of renal progenitor like cells from S3 segment of nephron in rat adult kidney. *FASEB J* 2005, **19**:1789–1797.
112. Dekel B, Zangi L, Shezen E, Reich-Zeliger S, Eventov-Friedman S, Katchman H, et al: Isolation and characterization of non-tubular Sca-1+Lin- multipotent stem/progenitor cells from adult mouse kidney. *J Am Soc Nephrol* 2006, **17**:3300–3314.
113. Gupta S, Verfaillie C, Chmielewski D, Kren S, Eidman K, Connaire J, et al: Isolation and characterization of kidney-derived stem cells. *J Am Soc Nephrol* 2006, **17**:3028–3040.
114. Lee PT, Lin HH, Jiang ST, Lu PJ, Chou KJ, Fang HC, et al: Mouse kidney progenitor cells accelerate renal regeneration and prolong survival after ischemic injury. *Stem Cells* 2010, **28**:573–584.
115. Park HC, Yasuda K, Kuo MC, Ni J, Ratliff B, Chander P, et al: Renal capsule as a stem cell niche. *Am J Physiol Renal Physiol* 2010, **298**:F1254–F1262.
116. Plotkin MD, Goligorski MS: Mesenchymal cells from adult kidney support angiogenesis and differentiate into multiple interstitial cell types including erythropoietin-producing fibroblasts. *Am J Renal Physiol* 2006, **291**:F902–F912.
117. Chen J, Park HC, Addabbo F, Ni J, Pelger E, Li H, Plotkin M, et al: Kidney-derived mesenchymal stem cells contribute to vasculogenesis, angiogenesis and endothelial repair. *Kidney Int* 2008, **74**:879–889.
118. Asakura A, Rudnicki MA: Side population cells from diverse adult tissues are capable of in vitro hematopoietic differentiation. *Exp Hematol* 2002, **30**:1339–1345.
119. Hishikawa K, Marumo T, Miura S, Nakanishi A, Matsuzaki Y, Shibata K, et al: Musculin/MyoR is expressed in kidney side population cells and can regulate their function. *J Cell Biol* 2005, **169**:921–928.
120. Iwatani H, Ito T, Imai E, Matsuzaki Y, Suzuki A, Yamato M, et al: Hematopoietic and nonhematopoietic potentials of Hoechstlow/side population cells isolated from adult rat kidney. *Kidney Int* 2004, **65**:1604–1614.
121. Challen GA, Bertoncello I, Deane JA, Ricardo SD, Little MH: Kidney side population reveals multilineage potential and renal functional capacity but also cellular heterogeneity. *J Am Soc Nephrol* 2006, **17**:1896–1912.

122. Addla SK, Brown MD, Hart CA, Ramani VAC, Clarke NW: **Characterization of the Hoechst 33342 side population from normal and malignant human renal epithelial cells.** *Am J Physiol Renal Physiol* 2008, **295**:F680–F687.
123. Inowa T, Hishikawa K, Takeuchi T, Kitamura T, Fujita T: **Isolation and potential existence of side population cells in adult human kidney.** *Int J Urol* 2008, **15**:272–275.
124. Imai N, Hishikawa K, Marumo T, Hirahashi J, Inowa T, Matsuzaki Y, et al: **Inhibition of histone deacetylase activates side population cells in kidney and partially reverses chronic renal injury.** *Stem Cells* 2007, **25**:2469–2475.
125. Deane JA, Ricardo SD: **Emerging roles for primary cilia in epithelial repair.** *Int Rev Cell Mol Biol* 2012, **293**:169–193.
126. Rodat-Despoix L, Delmas P: **Ciliary functions in the nephron.** *Pflugers Arch* 2009, **458**:179–87.
127. Saraga-Babic M, Vukojevic K, Bocina I, Drnasin K, Saraga M: **Ciliogenesis in normal human kidney development and post-natal life.** *Pediatr Nephrol* 2012, **27**:55–63.
128. Romagnani P, Lasagni L, Remuzzi G: **Renal progenitors: an evolutionary conserved strategy for kidney regeneration.** *Nat Rev Nephrol* 2013, **9**:137–146.
129. Trounson A, Thakar RG, Lomax G, Gibbons D: **Clinical trials for stem cell therapies.** *BMC Medicine* 2011, **9**:52.
130. Zeisberg M, Kalluri R: **Reversal of experimental renal fibrosis by BMP7 provides insights into novel therapeutic strategies for chronic kidney disease.** *Pediatr Nephrol* 2008, **23**:1395–1398.
131. Choudhury D, Levi M: **Kidney aging—inevitable or preventable?** *Nat Rev Nephrol* 2011, **7**:706–717.
132. McCreight CE, Sulkin NM: **Cellular proliferation in the kidneys of young and senile rats following unilateral nephrectomy.** *J Gerontol* 1959, **14**:440–443.
133. Yang HC, Rossini M, Ma LJ, Zuo Y, Ma J, Fogo AB: **Cells derived from young bone marrow alleviate renal aging.** *J Am Soc Nephrol* 2011, **22**:2028–2036.

doi:10.1186/2001-1326-2-11

**Cite this article as:** Li and Wingert: Regenerative medicine for the kidney: stem cell prospects & challenges. *Clinical and Translational Medicine* 2013 **2**:11.

**Submit your manuscript to a SpringerOpen<sup>®</sup> journal and benefit from:**

- ▶ Convenient online submission
- ▶ Rigorous peer review
- ▶ Immediate publication on acceptance
- ▶ Open access: articles freely available online
- ▶ High visibility within the field
- ▶ Retaining the copyright to your article

---

Submit your next manuscript at ▶ [springeropen.com](http://springeropen.com)

---