

Review

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Lymph node status as a guide to selection of available prognostic markers in breast cancer: the clinical practice of the future?

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Abstract

Prognosticators evaluating survival in breast cancer vary in significance in respect to lymph node status. Studies have shown e.g. that HER2/neu immunohistochemistry or HER2/neu gene amplification analysis do perform well as prognosticators in lymph node positive (LN +) patients but are less valuable in lymph node negative (LN -) patients. We collected data from different studies and tried to evaluate the relative significance of different prognosticators in LN+/LN- patient groups. In LN+ patients HER2/neu and E-cadherin immunohistochemistry were the statistically most significant prognosticators followed by proliferation associated features (mitotic counts by SMI (standardised mitotic index) or MAI (mitotic activity index), or S-phase fraction). Bcl-2 immunohistochemistry was also significant but p53 and cystatin A had no significance as prognosticators. In LN- patients proliferation associated prognosticators (SMI, MAI, Ki-67 index, PCNA immunohistochemistry, S-phase fraction) are especially valuable and also Cathepsin D, cystatin A, and p53 are significant, but HER2/neu or bcl-2, or E-cadherin less significant or without significance. We find that in studies evaluating single prognosticators one should distinguish between prognosticators suitable for LN+ and LN- patients. This will allow the choice of best prognosticators in evaluating the prospects of the patient. The distinction between LN+ and LN- patients in this respect may also be of special value in therapeutic decisions.

Background

Breast cancer is the most common cancer among women and the second leading cause of cancer deaths in women today. For example, 212,920 new cases of invasive breast cancer are expected to be diagnosed among women in the United States in 2006 (1.4 new cases among 1000 women) [1]. In Finland, according to the Finnish Cancer Registry, about 3800 females are diagnosed with breast cancer every year (1.5 new cases among 1000 women)[2]. Improved methods of detection and treatment of breast cancer have had a significant influence on disease out-

come [3]. The main challenge today is to find factors which could predict the patients who have a tumor with aggressive nature. There are a lot of data on clinical prognostic factors such as axillary lymph node (LN) status, tumor size, histological grade, and clinical stage. Breast cancer treatment could be based on these factors [4]. However, additional factors have also been used such as estrogen receptor (ER) positivity, progesterone receptor (PR) positivity, and HER2/neu status. The latter are considered especially useful in identifying patients who benefit from systemic adjuvant therapy. The International

Consensus Panel of St. Gallen determined the standard prognostic factors of breast cancer as follows: LN status, ER and PR, tumor size, histological grade, and age [5]. These prognostic factors have been used to identify the high-risk patients who may benefit from the adjuvant systemic therapy. However, still there is an urgent need for finding biological factors which could help in planning the future therapy of breast cancer.

Here we review several biological prognostic factors and associate their prognostic role with the lymph node status. Such grouping could help clinicians to create tailored and more individualized prognostic models which could help in choosing the right treatment for individual patients. Earlier research has shown that prognosticators vary between LN+ and LN- patients. Table 1 shows the characteristics of a few studies we will refer to in the following.

Differences in the size and other characteristics of the studied populations make comparison between different

reports difficult. Most of the studied populations are Caucasian (white), but in two American studies, there is also a fraction of black patients [37,68]. Because populations may differ in prognostic characteristics, we have decided to stress this point [1,65]. Most studies do not include adjustment for age and there are studies which do not necessarily clearly describe how cutpoints for each prognostic marker were defined. [In writing this review, however, we have expected that the authors have been able to select the most significant cutpoint for each prognosticator studied (see e.g. [123])].

Oncogenes and tumor suppressor genes

C-erbB-2 (HER2) or HER2/neu

HER2/neu proto-oncogene encodes a 185-KDa transmembrane glycosylated epidermal growth factor receptor that contains an extracellular domain and has intracellular tyrosine kinase activity [6]. Amplification of the HER2/neu gene and overexpression of its protein are known to be characteristic of many breast cancers [7-9]. Several

Table 1: Survival associated tissue-section-based prognostic factors and lymph node status in breast cancer.

Prognosticator	N	Age range or median	Country	Methods	Stage/LN status of all patients	Stength of association (survival %)*	P value, type of analysis	Mean or median follow-up (months)	Reference
LN+ patients									
AgNOR	164	N.A.	Italy	MA	N-, N+	DFS (42,61)	0.0093 M	108	Derenzini ⁹⁰
Bcl-2	107	56	Belgium	IHC	N-, N+	DFS (21,62)	<0.001 U	91	Hellemans ⁵⁵
CD44	74	62	Finland	IHC	N-, N+	DSS (45,56)	0.02 U	84	Joensuu ¹⁵⁸
E-cadherin	57	59	Finland	IHC	N-, N+	OS (36,88)	0.0001 U	66.5	Elzagheid ¹⁵⁰
EGFR	404	53.4	Japan	IHC	N-, N+	DFS (50,75)	<0.0001 U	46	Tsutsui ¹¹⁰
ER	705	54	UK	IHC	I-IV	DFS, ReR 0.651	0.004 M	62	Rehim ¹²²
HER2/neu	106	59	Finland	IHC	N-, N+	OS (13,80)	0.001 U	67.2	Jalava ¹⁶
IGF-1	98	57.1	Finland	IHC	N-, N+	OS (16,35)	0.0286 U	169.2	Toropainen ¹¹⁵
Mitoses	131	59	Finland	SMI	N-, N+	RR 3.5	0.0005 U	69	Kronqvist ⁷¹
Mitoses	368	25-81	MC	MAI	N+	OS (57,83)	0.00003 U	60	Simpson ⁶⁸
P21	328	N.A.	USA	IHC	N-, N+	DFS, X ² 74.61	0.054 M	195.6	Thor ³⁷
PgR	106	58	Finland	IHC	N-, N+	N.A.	0.0186	70	Jalava ¹²³
SPF	257	25-81	MC	FCM	N+	OS (70,82)	0.026 U	60	Simpson ⁶⁸
LN- patients									
Ploidy	50	56	UK	FCM	N-	DFS (32,71)	<0.0001 U	>120	Yuan ¹³²
Cathepsin D	262	60	Finland	IHC	N-	DFS (47,68)	<0.0001 U	98.4	Isola ¹⁰⁰
CD44	237	56	Holland	IHC	N-, N+	DFS (50,75)	0.005 U	84	Foekens ¹⁵⁶
EGFR	618	53.4	Japan	IHC	N-, N+	DFS, ReR 2.05	0.0241 M	46	Tsutsui ¹¹⁰
HER2/neu	324	N.A.	USA	FISH	N-	DFS (57,75)	0.0077 U	50	Press ¹⁷
HER2/neu	224	59	Finland	IHC	N-, N+	N.A.	N.S.	67.2	Jalava ¹⁶
Ki-67	89	60.5	Finland	IHC	N-, N+	OS (50,70)	0.0297 U	103.2	Pietilainen ⁷⁸
Mitoses	516	<55	MC	MAI	N-	DFS (62,85)	<0.00001U	118	Baak ⁶⁹
Mitoses	232	59	Finland	SMI	N-, N+	RR 4.2	0.0007 U	69	Kronqvist ⁷¹
P27	75	<65	Canada	IHC	N-, N+	DSS, ReR 0.24	0.03 M	180	Foulkes ⁴⁷
P53	700	<50	USA	IHC	N-	DFS (53,85)	0.0001U	54	Allred ³³
PCNA	205	60.5	Finland	IHC	N-	DFS (56,80)	0.0003 U	141.6	Aaltomaa ⁸⁴
SPF	180	N.A.	China	N.A.	N-	N.A.	<0.001	60	Zhang ⁹⁶

Prognosticators for LN+ patients are presented first followed by those for LN- patients. The database applied was PubMed, the presented papers were published during years 1991-2005. The most significant p-values with defined survival characteristics for each marker are shown. P-values are not comparable with each other in absolute terms because also the size of the sample influences the p value. The value of prognosticators can only be compared reliably by comparing different prognosticators in the same patient material.

* The figures within the parantheses refer to the survival of 2 cutpoint-associated patient groups after median follow up. IHC, immunohistochemistry; N-, node negative; N+, node positive; MA, morphometric analysis; FCM, flow-cytometry; NA, not available; MC, multicenter study; SMI, standardized mitotic index; MAI, mitotic activity index; OS, overall survival; DFS, disease free survival; DSS, disease specific survival; RR, risk ratio; ReR, relative risk; X², Chi square; FISH, fluorescence in situ hybridization; N.S., not significant; M, multivariate; U, univariate.

studies have correlated the overexpression of the HER2/neu oncogene with poor prognosis in breast cancer patients [10,11], and demonstrated that evaluating HER2/neu protein by immunohistochemistry (IHC), fluorescence in situ hybridization (FISH), and chromogenic in situ hybridization (CISH) are important in selecting optimal therapy and predicting prognosis in breast cancer patients [12-14]. Slamon et al. and Jalava et al. [15,16], showed that intensive HER2/neu immunostaining was a highly significant negative prognosticator in LN+ patients (especially postmenopausal), but not a significant prognosticator among LN- patients or patients younger than 52 years. The paper of Jalava et al. [16] suggested that the size of the patient group which absolutely seemed to need the targeted therapy against the amplified HER2/neu receptor was at least 2% of all breast cancer patients. Press et al. [17], however, suggested that HER2/neu is a significant prognosticator in LN- patients. However, there is still no perfect consensus on which method is most predictive of positive patient response to trastuzumab, also known as Herceptin, a monoclonal antibody that selectively targets HER2/neu. Tawfik et al. [18] discussed the results from a study comparing HER2/neu expression and gene amplification in the same patients by IHC using automated cellular imaging system (ACIS) and by FISH. They concluded that HER2/neu assessment by IHC-ACIS correlates highly with results obtained by FISH. The concordance rate between two methods was 94%. Isola et al. [19] showed that CISH could provide an accurate and practical alternative to FISH for clinical diagnosis of HER2/neu oncogene amplification in archival formalin-fixed breast cancer samples. Seidman and associates [20] reported that the overall response rate (for combination paclitaxel and trastuzumab) in patients with HER2/neu overexpression ranged from 67% to 81%, compared with 41% to 46% in patients with tumors having normal Her-2 expression. In agreement with them Joensuu et al., Piccart-Gebhart et al., and Romond et al. [21-23] demonstrated positive response among patients showing HER2/neu amplification or intensive membrane staining by immunohistochemistry. The effect was clear in disease-free survival and overall survival in freedom from distant metastases.

P53

Early observations after the identification of the p53 protein suggested that p53 functioned as an oncogene. In fact it does, but only in the mutated form. In the late 1980s, however, several discoveries proved that the normal function of p53 was anti-oncogenic. One of the several functions of the normal p53 gene is suppression of cell proliferation. When DNA is damaged p53 inhibits the progression of cell cycle from G1 to the S-phase or during S-phase [24-26]. The mutation of the p53 gene has been detected in almost 50% of human cancers including breast cancer [27-29]. Several studies demonstrated that

mutations of p53 or increased nuclear expression of p53 protein is a prognostic factors in breast cancer, and associated with worse prognosis [30-32]. Allred and associates [33] showed that p53 predicted disease free survival in patients with LN- breast cancer. Expression of mutant p53 protein was associated with early disease recurrence and early death in LN- breast cancer. The results of Kuopio et al. [34] clearly support the latter finding. In their series immunohistochemical positivity of p53 was a significant prognosticator among all and LN- patients, but did not show significance among LN+ patients.

P21

p21 (WAF) is an inhibitor of cyclin-dependent kinase. It plays a central role both in the regulation of the cell cycle and in the DNA replication [35]. p21 (WAF) is regarded as a putative tumor suppressor and its role in breast cancer is still unclear. p21 (WAF) might have a functional role in the inhibition of PCNA mediated DNA replication [35]. So far there is not much evidence that p21 (WAF) could be used as a prognostic factor in breast cancer [36]. Thor and co-workers demonstrated marginal independent prognostic significance of the p21 (WAF) in the LN+ set of patients, whereas p21 status was not significantly associated with survival among LN- patients [37]. Ras p21 is a guanine nucleotide-binding protein that is involved in the signal transduction pathway that control cell proliferation [38,39]. The ras gene is rarely mutated in breast tumors, but is overexpressed in 50-60% of breast tumors [40]. Rundle et al. suggested that the presence of ras p21 in blood is associated with a five fold increased risk of breast cancer [41]. Czerniak et al. found that p21 expression was significantly higher in cancer cells than in epithelial cells of control specimens. As a group, tumors with axillary lymph node metastases expressed higher levels of ras p21 than nonmetastasizing tumors [42].

P27

P27 is a cyclin dependent kinase inhibitor, which may act as a potential suppressor gene. Reduction of p27 expression is related to uncontrolled cell proliferation and tumorigenesis. P27 acts in G1 to inhibit cyclin-cdks [43,44]. Several studies have suggested that low expression of p27 is an independent factor weakly associated with poor prognosis [45,46]. Foulkes and his co-workers (2004) [47] demonstrated that the level of p27 was one of independent predictors in LN- patients but not in LN+ patients.

C-myc

Amplification of the c-myc gene is found in 20% of primary breast cancers, and this is more frequent in larger tumors, and in lymph node positive patients [48]. Overexpression and other alterations of c-myc gene may be related to breast cancer progression. It can be expected

that c-myc has a potential as a marker of poor prognosis [49,50]. We did not find c-myc associated prognostic information which was related to the lymph node status.

Bcl-2

Bcl-2 gene encodes for a mitochondrial protein thought to prevent apoptosis in normal cells. Dysregulation of this gene can contribute to tumor progression and increased drug resistance [51,52]. Bcl-2 expression was associated with favourable prognosis in breast cancer and was both estrogen receptor(ER) and progesterone receptor (PR) associated [53,54]. Several studies have demonstrated the independent favourable prognostic impact of Bcl-2 on breast cancer particularly among LN+ patients [55,56]. It is obvious that the prognostic value of Bcl-2 staining among all patients is solely based on prognostic applicability among LN+ patients because the association among LN- patients is clearly non significant [57].

Cell proliferation

Mitotic count

Evaluation of proliferation activity of the neoplastic cells gives important prognostic information, especially in breast cancer [58,59]. There are many available ways to measure cell proliferation, including the determination of the mitotic rate by counting the mitotic figures [60,61]. Mitotic counts are performed by counting the number of mitoses from ten high power fields (Baak et al. ; mitotic activity index; MAI), or by expressing the count by square millimeter, which produces the standardised mitotic index (SMI) [62], or volume fraction corrected mitotic index (M/V_v index) [63,64]. Many studies demonstrated that the MAI is an independent prognostic factor for recurrence free survival [65,66]. SMI is a bit more efficient than MAI as a prognosticator [67]. Simpson et al. [68] showed that determination of mitotic activity is able to identify a group with improved disease free and overall survival among patients with LN+ breast cancer. The mitotic activity is also an independent prognostic marker in LN- breast cancer patients younger than 55 years [69]. Baak and his co-worker also found that the LN- patients with MAI > 10 are at risk for distant metastases, similar to LN+ [70]. SMI was a highly significant prognosticator among all patients but especially among LN- patients [71]. Among LN+ patients this marker was less significant. It is surprising in light of numerous ways to measure proliferative activity how consistently the mitotic count emerges as the most powerful prognosticator. All ways of estimation are prognosticators but the mitotic count is the strongest one in the majority of comparing studies.

Ki-67

Ki-67 antigen is one of several cell-cycle regulating proteins which can be determined by immunohistochemistry [72,73]. It is expressed in proliferating G1-, S- and G2/M-

phase nuclei [74,75]. Because most of the Ki-67 positive cells are in the cell cycle, Ki-67 labelling index (fraction of Ki-67 positive nuclei of all nuclei) reflects the fraction of proliferating cells. Many studies demonstrated the association between a high Ki-67 labelling index, histological grade and large tumor size in breast carcinoma [76,77]. Ki-67 labelling index (determined e.g. by the MIB1 antibody; an IgG monoclonal antibody used for detection Ki-67) has prognostic value in breast cancer, particularly in LN- patients [78,79]. However, the prognostic significance is less than that of mitotic count (as determined with either MAI or SMI). However, there is some evidence that 3-dimensional analysis of Ki-67-positive nuclei may improve the prognostic power of Ki-67 immunohistochemistry [80].

PCNA

Proliferating cell nuclear antigen (PCNA) is a DNA-polymerase-related protein which is expressed in all proliferating cells. The expression is increased during G1 phase, S-phase and declines during G2/M [81]. Many reports observed that PCNA shows prognostic value in breast cancer [82,83]. Aaltomaa and his coworkers stated that the prognostic significance of PCNA can be demonstrated especially among LN- patients [84]. They did not include LN+ patients in their study.

AgNOR

Argyrophilic nucleolar organizer regions (AgNOR) correlate with the proliferative activity of neoplasms. Increased AgNOR counts may reflect increased proliferative activity of cells [85,86]. AgNOR counts have been studied in breast carcinoma, but the results have been conflicting. Some studies have demonstrated that quantitative analysis of AgNORs yields a prognostic factor in breast cancer [87,88]. The combination of MIB-1 immunopositivity and AgNOR measurements in MIB-1 positive nuclei improves prognostication [89]. Derenzini et al. reported that AgNOR has a prognostic value specially among LN+ patients [90]. Others were not able to find prognostic significance in AgNOR counts for breast cancer [91,92].

S-phase fraction (SPF)

The histogram describing the nuclear DNA content determined by flow cytometry can be used as an estimate of the proliferation rate. The measurement estimates the fraction of cells in the S phase (S phase fraction), which reflects proliferative activity. The S-phase fraction (SPF) is a rough estimate of neoplastic growth rate. Several reports observed the prognostic value of SPF in breast cancer [93,94]. Low SPF is associated with an excellent prognosis in LN- breast patients [95-97]. Based on the grading efficiency analysis [98] SPF estimation from isolated nuclei may reach efficiencies comparable to that of mitotic counts.

Cathepsin D (CD)

Cathepsin D is a 52-kd protein, precursor to a lysosomal acidic protease. This proteolytic enzyme can degrade basement membranes. It has mitogenic activity on MCF-7 cells that are estrogen depleted and contributes to proliferation, invasion, and progression in breast [99]. Several studies have demonstrated that Cathepsin D is an important prognostic factor in breast cancer, especially in LN- patients [100,101]. In the report by Rochefort [102] over-expression of cathepsin D was associated with increased risk of metastasis, but there are also studies that failed to find prognostic significance of cathepsin D in breast cancer [103,104].

Growth factors**Epidermal growth factor receptor (EGFR)**

EGFR is a member of the tyrosine kinase growth factor receptor family. EGFR and its ligand, transforming growth factor- α (TGF- α) play an important role in several human cancers [105,106], through the autocrine growth-regulation system in breast cancer, EGFR has been reported to be associated with a poor clinical outcome [107,108]. Rampaul and his co-workers found that the EGFR was significant only in lymph node positive breast cancer patients [109], with high expression reflecting worse outcome. Tsutsui et al. reported that a combination of EGFR and ER was an independent significant factor for both disease free survival (DFR) and overall survival (OS) both in patients with LN- and LN+ breast cancer. Patients with EGFR (+) and ER (-) had worse DFS and OS [110].

Insulin-like growth factor (IGF)

IGF1 and IGF2 are circulating peptide hormones and locally acting growth factors with both paracrine and autocrine functions. Both IGF1 and IGF2 are involved in the regulation of cell proliferation and apoptosis [111]. Many observers suggested that IGFs are involved in the progression of breast cancer [112,113]. The presence of IGF-1 immunoreactivity in breast cancer epithelial cells indicates a lower degree of malignancy than the lack of IGF1 [114]. Toropainen et al. reported that, in a univariate analysis, IGF-1 was significantly related to a high survival probability particularly in LN+ breast cancer patients [115].

Estrogen (ER) and progesterone (PR) receptors

Since breast cancer is one of the hormone dependent tumors much attention has been paid to the relationship between ER and PR and breast cancer. The study by Blanco et al. (1984) [116] showed that ER+ PR+ patients had better prognosis than ER- PR- patients. In late 1970's and early 1980's the measurement of ER as well as PR became standard practice in the prediction of the outcome of breast cancer patients [117-119]. The immunohistochemical analysis of ER and PR has replaced the traditional and

also clinically validated dextran charcoal radioactive ligand binding assay [120]. The prognostic value of ER and PR seems to be greater among axillary LN+ than among LN- patients [121,122]. The study of Jalava et al. [123] showed that immunohistochemical ER score is associated with prognosis. However, cutpoints for defining the groups with good or worse prognosis may differ between LN- and LN+ patients.

DNA content

Cytometric quantitation of nuclear DNA content can assist in the diagnosis and grading of malignant tumors [124]. A great number of studies using flow cytometry and static image cytometry suggested that nuclear DNA content has a significant value in prognosis of breast cancer [125-127]. Tumors with DNA peaks within diploid limits have a more favorable prognosis than those with aneuploid peaks [128,129]. Nuclear DNA content strongly correlated to histopathologic grade of ductal carcinoma. Histologic grade 3 tumors were more likely to be aneuploid than others [130,131]. Many observers stated that aneuploid tumors were associated with a poorer prognosis than diploid tumors in LN- breast cancer patients [132,129]. However, an adverse correlation was stated by Chassevent et al. They confirmed that ploidy status had no prognostic impact in the overall population or in subgroups defined by lymph node status [133]. Moureau-Zabotto et al. suggested that combination of DNA ploidy and SPF predict patients outcome particular in LN- breast cancer patients [134]. DNA ploidy results can be combined with other features in efficient evaluation of prognosis [135].

Nuclear morphometry

Baak and his coworkers (1985) were among the pioneers in introducing morphometry for prognostication of breast cancer [62]. They found that morphologic features are associated with high risk in pre-invasive breast cancer. The independent prognostic value of nuclear variables was established in several studies on infiltrating breast cancer [136]. Nuclear area and diameter were shown to be useful prognostic factors. However, nuclear diameter has failed to separate the primary tumor from its metastasis [137]. Patients with high nuclear area values and high SD of nuclear area values tend to have poor prognosis [138]. High mean nuclear area is related to poor histological grade [139]. Evaluation of nuclear area can be used for morphometric grading of breast cancer [140]. The latter study also showed that the nuclear area was a significant prognosticator among LN+ and all patients, but completely lacked significance among LN- patients.

Adhesion molecules**E-cadherin**

Cell adhesion molecules play an important role in the maintenance of tissue architecture [141]. E-cadherin is a member of a family of transmembrane glycoproteins that mediate homotypic calcium dependent cell-to-cell adhesion in epithelial tissues [142]. Loss of normal function of E-cadherin is known to promote cancer invasion in several human cancers [143]. Several studies have demonstrated that reduced E-cadherin expression is an indicator of increased invasiveness and dedifferentiation in breast cancer [144,145]. A recent study found that the expression of E-cadherin correlates with histological type and grade [146]. The expression of E-cadherin in infiltrative lobular carcinoma was completely negative or weakly positive whereas infiltrating ductal carcinoma showed greater immunoreactivity in grade 1 breast carcinoma than in grade 2 and grade 3 carcinomas [147]. There was an inverse relationship between E-cadherin expression and axillary lymph node involvement. Reduction in E-cadherin expression was associated with the involvement of the axillary lymph nodes [148]. Gamallo and his associates [149], and Lipponen and his co-workers [142] did not agree with this finding. Siitonen et al. [144] showed that reduced E-cadherin expression was independently associated with shorter disease free survival. Other authors stated that the prognostic value of E-cadherin was stronger among patients with LN+ breast cancer [150,151] than among all patients. However, there was no significant association with prognosis among LN- patients [150]. The latter findings can explain the variable findings by different research groups on the prognostic significance of E-cadherin among all breast cancer patients.

Catenins

Intracellular proteins (α - β - γ -catenins), are associated with E-cadherin's cytoplasmic tail and link actin to the microfilament network of the cellular cytoskeleton. This binding is essential for the adhesive function of E-cadherin [152]. Alterations in expression of catenins may lead to the disassembly of cadherin junctions and to the generation of more invasive cells [153]. Many studies have suggested that loss of E-cadherin and catenin expression may be associated with poor prognosis of breast cancer [154,155]. We did not find any reports of catenins associated with prognostic data which is related to the lymph node status.

CD 44

CD44 is a family of cell surface transmembrane glycoproteins which is expressed in a wide variety of tissues and cell types [156,157]. In human breast cancer, the prognostic value of the various isoforms has not been recognized as independent predictors of breast cancer outcome [158,159]. Foekens and co-workers [156] stated that the

expression of exon v6 of CD44 may be a marker for identifying patients with relatively favourable prognosis among LN- patients.

Extracellular matrix**Tenascin**

Tenascin is a glycoprotein of the extracellular matrix and appears in the stroma of benign and malignant tumors [160]. Many studies have stated that loss of tenascin expression in breast cancer cells appears to indicate poor prognosis [161,162]. So far we do not have knowledge of differences in the prognostic value between LN + and LN - patients.

Matrix metalloproteinases (MMPs)

MMPs are a family of proteinases that play an important role in malignant tumor growth, invasion, metastasis and angiogenesis [163,164]. High expression levels of MMPs in tumor tissue are usually associated with poor survival [165]. In breast cancer, Talvensaaari-Mattila and his co-worker [166] demonstrated that overexpression of MMP-2 is associated with poor prognosis. Prognostic data on LN + and LN - patients are not available.

Cystatin A

Cystatin A is a natural cysteine proteinase inhibitor and is found in a wide variety of normal cells. The physiologic role of Cystatin A is not fully known, however. Cystatin A is present in large amounts in follicular dendritic cells in lymphoid tissues [167]. In malignant tissues, cystatin A has been found in many tumors including breast cancer [168]. In breast cancer patients, Kuopio et al. (1998) demonstrated that the expression of cystatin A was associated with poor outcome; the study also showed that cystatin A was a prognostic marker for LN- patients. In LN+ patients, cystatin A was without significance [34].

Grading

Various histological grading systems of breast carcinoma have been described [169-171]. The majority of tumor grading systems combine nuclear grade, tubular formation and mitotic rate. In Europe, Elston and Ellis (1991) stressed the importance of careful grading [172] and corresponding approach has become increasingly popular in the US [173,68]. It was suggested that the histological grade functions best as a prognosticator in LN- patients for making decision when tumor sizes fall between tumor size categories [174]. The applied histological features can be measured morphometrically [71,140,175], making the grading system robust and reproducible. The multivariate grading methods [62,176-178] are comparable or better than grading methods alone and can be expected to be more reproducible. However, the performance in grouping of the patients in low and high grade categories is not necessarily identical because these grading methods were

originally based on different patient materials [179]. It is important to notice that most multivariate methods include lymph node status as a contributing feature and for that reason multivariate methods or indices are applicable to both LN+ and LN- patients. This also applies to the Nottingham prognostic index which combines tumor size, lymph node status, and histological grade [172]. So far, however, there are few studies comparing traditional and multivariate grading in clinical practice or in different patient groups in terms of power to finding patients with bad prognosis. But we know that different individual prognostic factors and grading systems often give parallel results. For example: when DNA ploidy status is diploid, histological grade is generally low, and when DNA ploidy is aneuploid, histological grade is higher [180]. If histological grade is used as the basic classifier of patients into treatment groups, DNA ploidy can be used as a method to confirm the classification especially in situations of uncertainty. In the same way, other biological prognosticators can and probably should be applied in clinical practice.

Conclusion

Mitotic activity indices (SMI or MAI) are the most powerful general prognostic markers and highly associated with survival in LN+ patients and in LN- patients younger than 55 years old.

E-cadherin was the second most important prognostic marker in patients with LN +, followed by HER2/neu. In LN- patients, cathepsin D was the second most important prognostic marker followed by HER2/neu, SPF, and aneuploidy.

Table 1 lists histological prognostic markers in LN- and LN+ breast cancers. Because the presented survival associated significances are based on different studies, they are not directly comparable. However, reliable comparison in term of prognostic value can be done after univariate analysis in the same group of patients. Multivariate analysis is less suitable for comparison of variables because the goal of the multivariate analysis is the removal of the prognostic overlaps. The strength of association can be evaluated either by risk ratio, relative risk, or comparison of the survival curves defined by 2 cutpoint associated patient groups in respect to disease free survival (DFS), overall survival (OS), or disease specific survival (DSS). The difference in the importance of prognosticators is presented in Table 2. Generally, comparison is made difficult by the lack of standardized study protocols. Table 2 shows, however, that prognosticators vary in terms of stage: the survival of LN- and LN+ patients can best be predicted with prognosticators specific to the LN status.

We conclude that studies on prognostic evaluation of new potential markers in breast cancer should not be under-

Table 2: Comparison of different prognosticators in single studies on breast cancer.

LN +	Prognosticator	P value
Kuopio et al. 1998 [34]	1. Bcl-2	0.017
	2. p53	0.121
	3. Cystatin A	0.386
Jalava et al. 2002 [16]	1. ErbB2	0.0002
	2. SMI	0.0014
	3. Bcl-2	0.048
Elzagheid et al. 2002 [150]	1. E-cadherin	0.0006
	2. SMI	0.0133
<hr/>		
LN -		
Kuopio et al. 1998 [34]	1. Cystatin A	0.010
	2. p53	0.021
	3. Bcl-2	0.874
Jalava et al. 2002 [16]	1. SMI	0.0001
	2. ErbB2	N.S.
	3. Bcl-2	N.S.
Elzagheid et al. 2002 [150]	1. SMI	0.0299
	2. E-cadherin	0.5581

P values refer to the results of univariate analysis. Comparison of univariate analyses in a study allows one to conclude on the relative strengths of different prognosticators.

taken without studying the prognosis among all patients, and separately among LN + and LN - patients. Correspondingly the use of prognosticators for clinical decision making should be based on the gathered knowledge of differences between LN + and LN - patients.

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References

- Smigal C, Jemal A, Ward E, Cokkinides V, Smith R, Howe HL, Thun M: **Trends in breast cancer by race and ethnicity: update 2006.** *CA A Cancer J Clin* 2006, **56**:168-183.
- Finnish Cancer Registry: Cancer in Finland. 2002 and 2003. Cancer statistics of the National Research and Development Centre for Welfare and Health. Cancer Society of Finland.** Publication No. 66, Helsinki 2005. ISSN 0585-9603
- Levi F, Lucchini F, Negri E, La Vecchia C: **Worldwide patterns of cancer mortality, 1990-1994.** *Eur J Cancer Prev* 1999, **8(5)**:381-400.
- Fisher ER, Anderson S, Redmond C, Fisher B: **Pathologic findings from the National Surgical adjuvant breast project protocol B-06. 10-year pathologic and clinical prognostic discriminants.** *Cancer* 1993, **71(8)**:2507-14.
- Goldhirsch A, Wood WC, Gelber RD, Coates AS, Thurlimann B, Senn HJ: **Meeting highlights: updated international expert consensus on the primary therapy of early breast cancer.** *J Clin Oncol* 2003, **21(17)**:3357-65.

6. Ali SM, Leitzel K, Chinchilli VM, Engle L, Demers L, Harvey HA, Carney W, Allard JW, Lipton : **Relationship of serum HER-2/neu and serum CA 15-3 in patients with metastatic breast cancer.** *Clin Chem* 2002; **13**:14-20.
7. King CR, Kraus MH, Aaronson SA: **Amplification of a novel v-erbB-related gene in a human mammary carcinoma.** *Science* 1985, **229**(4717):974-6.
8. Tal M, Wetzler M, Joseberg Z, Deutch A, Gutman M, Assaf D, Kris R, Shiloh Y, Givol D, Schlessinger J: **Sporadic amplification of the HER2/neu protooncogene in adenocarcinomas of various tissues.** *Cancer Res* 1988, **48**(6):1517-20.
9. Devilee P, Schuurin E, van de Vijver MJ, Cornelisse CJ: **Recent developments in the molecular genetic understanding of breast cancer.** *Crit Rev Oncog* 1994, **5**(2-3):247-70.
10. Ravdin PM, Chamness GC: **The c-erbB-2 proto-oncogene as a prognostic and predictive marker in breast cancer: a paradigm for the development of other macromolecular markers.** *Gene* 1995, **159**(1):19-27.
11. Naidu R, Yadav M, Nair S, Kutty MK: **Expression of c-erbB3 protein in primary breast carcinomas.** *Br J Cancer* 1998, **78**(10):1385-90.
12. Yeh IT: **Measuring HER-2 in breast cancer. Immunohistochemistry, FISH, or ELISA?** *Am J Clin Pathol* 2002, **117**(Suppl):S26-35. Review
13. Winters ZE, Leek RD, Bradburn MJ, Norbury CJ, Harris AL: **Cytoplasmic p21WAF1/CIP1 expression is correlated with HER-2/neu in breast cancer and is an independent predictor of prognosis.** *Breast Cancer Res* 2003, **5**(6):R242-9.
14. Gu M, Ghafari S, Zhao M: **Fluorescence in situ hybridization for HER-2/neu amplification of breast carcinoma in archival fine needle aspiration biopsy specimens.** *Acta Cytol* 2005, **49**(5):471-6.
15. Slamon DJ, Clark GM, Wong SG, Levin WJ, Ullrich A, McGuire WL: **Human breast cancer: correlation of relapse and survival with amplification of the HER-2/neu oncogene.** *Science* 1987, **235**(4785):177-82.
16. Jalava PJ, Kuopio T, Kortelainen S, Kronqvist P, Collan YU: **Quantitation of erbB2 positivity for evaluation of high-risk patients.** *Ann Med* 2002, **34**(7-8):544-53.
17. Press MF, Bernstein L, Thomas PA, Meisner LF, Zhou JY, Ma Y, Hung G, Robinson RA, Harris C, El-Naggar A, Slamon DJ, Phillips RN, Ross JS, Wolman SR, Flom KJ: **HER-2/neu gene amplification characterized by fluorescence in situ hybridization: poor prognosis in node-negative breast carcinomas.** *J Clin Oncol* 1997, **15**(8):2894-904.
18. Tawfik OW, Kimler BF, Davis M, Donahue JK, Persons DL, Fan F, Hagemester S, Thomas P, Connor C, Jewell W, Fabian CJ: **Comparison of immunohistochemistry by automated cellular imaging system (ACIS) versus fluorescence in-situ hybridization in the evaluation of HER-2/neu expression in primary breast carcinoma.** *Histopathology* 2006, **48**(3):258-67.
19. Isola J, Tanner M, Forsyth A, Cooke TG, Watters AD, Bartlett JM: **Interlaboratory comparison of HER-2 oncogene amplification as detected by chromogenic and fluorescence in situ hybridization.** *Clin Cancer Res* 2004, **10**(14):4793-8.
20. Seidman AD, Fornier MN, Esteva FJ, Tan L, Kaptain S, Bach A, Panageas KS, Arroyo C, Valero V, Currie V, Gilewski T, Theodoulou M, Moynahan ME, Moasser M, Sklarin N, Dickler M, D'Andrea G, Cristofanilli M, Rivera E, Hortobagyi GN, Norton L, Hudis CA: **Weekly trastuzumab and paclitaxel therapy for metastatic breast cancer with analysis of efficacy by HER2 immunophenotype and gene amplification.** *J Clin Oncol* 2001, **19**(10):2587-95.
21. Joensuu H, Kellokumpu-Lehtinen PL, Bono P, Alanko T, Kataja V, Asola R, Utriainen T, Kokko R, Hemminki A, Tarkkanen M, Turpeenniemi-Hujanen T, Jyrkkio S, Flander M, Helle L, Ingalsuo S, Johansson K, Jaaskelainen AS, Pajunen M, Rauhala M, Kaleva-Kerola J, Salminen T, Leinonen M, Elomaa I, Isola J, FinHer Study Investigators: **Adjuvant docetaxel or vinorelbine with or without trastuzumab for breast cancer.** *N Engl J Med* 2006, **354**(8):809-20.
22. Piccart-Gebhart MJ, Procter M, Leyland-Jones B, Goldhirsch A, Untch M, Smith I, Gianni L, Baselga J, Bell R, Jackisch C, Cameron D, Dowsett M, Barrios CH, Steger G, Huang CS, Andersson M, Inbar M, Lichinitser M, Lang I, Nitz U, Iwata H, Thomssen C, Lohrisch C, Suter TM, Ruschoff J, Suto T, Greaetorex V, Ward C, Straehle C, McFadden E, Dolci MS, Gelber RD, Herceptin Adjuvant (HERA) Trial Study Team: **Trastuzumab after adjuvant chemotherapy in HER2-positive breast cancer.** *N Engl J Med* 2005, **353**(16):1659-72.
23. Romond EH, Perez EA, Bryant J, Suman VJ, Geyer CE Jr, Davidson NE, Tan-Chiu E, Martino S, Paik S, Kaufman PA, Swain SM, Pisansky TM, Fehrenbacher L, Kutteh LA, Vogel VG, Visscher DW, Yothers G, Jenkins RB, Brown AM, Dakhil SR, Mamounas EP, Lingle WL, Klein PM, Ingle JN, Wolmark N: **Trastuzumab plus adjuvant chemotherapy for operable HER2-positive breast cancer.** *N Engl J Med* 2005, **353**(16):1673-84.
24. Levine AJ, Momand J, Finlay CA: **The p53 tumour suppressor gene.** *Nature* 1991, **351**(6326):453-6.
25. Kastan MB, Onyekwere O, Sidransky D, Vogelstein B, Craig RW: **Participation of p53 protein in the cellular response to DNA damage.** *Cancer Res* 1991, **51**(23 Pt 1):6304-11.
26. Barnes DM, Camplejohn RS: **P53, apoptosis, and breast cancer.** *J Mammary Gland Biol Neoplasia* 1996, **1**(2):163-75.
27. Hollstein M, Sidransky D, Vogelstein B, Harris CC: **p53 mutations in human cancers.** *Science* 1991, **253**(5015):49-53.
28. Bookstein R, MacGrogan D, Hilsenbeck SG, Sharkey F, Allred DC: **p53 is mutated in a subset of advanced-stage prostate cancers.** *Cancer Res* 1993, **53**(14):3369-73.
29. Temmim L, Baker H, Sinowatz F: **Immunohistochemical detection of p53 protein expression in breast cancer in young Kuwaiti women.** *Anticancer Res* 2001, **21**(1B):743-8.
30. Bergh J: **Clinical studies of p53 in treatment and benefit of breast cancer patients.** *Endocr Relat Cancer* 1999, **6**(1):51-9.
31. Kalogeraki A, Panayiotides J, Tamiolakis D, Tzardi M, Chaniotis V, Chalkiadakis G, Melissas J, Stiftis D, Kanavaros P, Delides GS: **P53 expression in patients with malignant and benign breast diseases.** *Anticancer Res* 2000, **20**(3A):1801-5.
32. Metcalfe S, Wheeler TK, Picken S, Negus S, Jo Milner A: **P53 autoantibodies in 1006 patients followed up for breast cancer.** *Breast Cancer Res* 2000, **2**(6):438-43.
33. Allred DC, Clark GM, Elledge R, Fuqua SA, Brown RW, Chamness GC, Osborne CK, McGuire WL: **Association of p53 protein expression with tumor cell proliferation rate and clinical outcome in node-negative breast cancer.** *J Natl Cancer Inst* 1993, **85**(3):200-6.
34. Kuopio T, Kankaanranta A, Jalava P, Kronqvist P, Kotkansalo T, Weber E, Collan Y: **Cysteine proteinase inhibitor cystatin A in breast cancer.** *Cancer Res* 1998, **58**(3):432-6.
35. Gohring UJ, Bersch A, Becker M, Neuhaus W, Schondorf T: **p21(waf) correlates with DNA replication but not with prognosis in invasive breast cancer.** *J Clin Pathol* 2001, **54**(11):866-70.
36. Elledge RM, Allred DC: **Prognostic and predictive value of p53 and p21 in breast cancer.** *Breast Cancer Res Treat* 1998, **52**(1-3):79-98.
37. Thor AD, Liu S, Moore DH 2nd, Shi Q, Edgerton SM: **p(21WAF1/CIP1) expression in breast cancers: associations with p53 and outcome.** *Breast Cancer Res Treat* 2000, **61**(1):33-43.
38. Wiesmuller L, Wittinghofer F: **Signal transduction pathways involving Ras.** *Cell Signal* 1994, **6**(3):247-67.
39. Medema RH, Bos JL: **The role of p21ras in receptor tyrosine kinase signaling.** *Crit Rev Oncog* 1993, **4**(6):615-61.
40. Gulbis B, Galand P: **Immunodetection of the p21-ras products in human normal and preneoplastic tissues and solid tumors.** *Hum Pathol* 1993, **24**(12):1271-85.
41. Rundle A, Tang D, Brandt-Rauf P, Zhou J, Kelly A, Schnabel F, Perera FP: **Association between the ras p21 oncoprotein in blood samples and breast cancer.** *Cancer Lett* 2002, **185**(1):71-8.
42. Czerniak B, Chen R, Tuziak T, Markiewski M, Kram A, Gorczyca W, Deitch D, Herz F, Koss LG: **Expression of ras oncogene p21 protein in relation to regional spread of human breast carcinomas.** *Cancer* 1989, **63**(10):2008-13.
43. Barbareschi M: **p27 Expression, a cyclin dependent kinase inhibitor in breast carcinoma.** *Adv Clin Path* 1999, **3**(4):119-27.
44. Chiarle R, Pagano M, Inghirami G: **The cyclin dependent kinase inhibitor p27 and its prognostic role in breast cancer.** *Breast Cancer Res* 2001, **3**(2):91-4.
45. Tsuchiya A, Zhang GJ, Kanno M: **Prognostic impact of cyclin-dependent kinase inhibitor p27kip1 in node-positive breast cancer.** *J Surg Oncol* 1999, **70**(4):230-4.
46. Chappuis PO, Kapusta L, Begin LR, Wong N, Brunet JS, Narod SA, Slingerland J, Foulkes WD: **Germline BRCA1/2 mutations and p27(Kip1) protein levels independently predict outcome after breast cancer.** *J Clin Oncol* 2000, **18**(24):4045-52.

47. Foulkes WD, Brunet JS, Stefansson IM, Straume O, Chappuis PO, Begin LR, Hamel N, Goffin JR, Wong N, Trudel M, Kapusta L, Porter P, Akslen LA: **The prognostic implication of the basal-like (cyclin E high/p27 low/p53+/glomeruloid-microvascular-proliferation+) phenotype of BRCA1-related breast cancer.** *Cancer Res* 2004, **64(3)**:830-5.
48. Berns EM, Klijn JG, van Putten WL, van Staveren IL, Portengen H, Foekens JA: **c-myc amplification is a better prognostic factor than HER2/neu amplification in primary breast cancer.** *Cancer Res* 1992, **52(5)**:1107-13.
49. Naidu R, Wahab NA, Yadav M, Kutty MK: **Protein expression and molecular analysis of c-myc gene in primary breast carcinomas using immunohistochemistry and differential polymerase chain reaction.** *Int J Mol Med* 2002, **9(2)**:189-96.
50. Aulmann S, Bentz M, Sinn HP: **C-myc oncogene amplification in ductal carcinoma in situ of the breast.** *Breast Cancer Res Treat* 2002, **74(1)**:25-31.
51. Silvestrini R, Veneroni S, Daidone MG, Benini E, Boracchi P, Mezzetti M, Di Fronzo G, Rilke F, Veronesi U: **The Bcl-2 protein: a prognostic indicator strongly related to p53 protein in lymph node-negative breast cancer patients.** *J Natl Cancer Inst* 1994, **86(7)**:499-504.
52. Olopade OI, Adeyanju MO, Safa AR, Hagos F, Mick R, Thompson CB, Recant WM: **Overexpression of BCL-x protein in primary breast cancer is associated with high tumor grade and nodal metastases.** *Cancer J Sci Am* 1997, **3(4)**:230-7.
53. Knowlton K, Mancini M, Creason S, Morales C, Hockenbery D, Anderson BO: **Bcl-2 slows in vitro breast cancer growth despite its antiapoptotic effect.** *J Surg Res* 1998, **76(1)**:22-6.
54. Park SH, Kim H, Song BJ: **Down regulation of bcl2 expression in invasive ductal carcinomas is both estrogen-and progesterone-receptor dependent and associated with poor prognostic factors.** *Pathol Oncol Res* 2002, **8(1)**:26-30.
55. Hellemans P, van Dam PA, Weyler J, van Oosterom AT, Buytaert P, Van Marck E: **Prognostic value of bcl-2 expression in invasive breast cancer.** *Br J Cancer* 1995, **72(2)**:354-60.
56. Bhatavdekar JM, Patel DD, Shah NG, Vora HH, Suthar TP, Chikhlikar PR, Ghosh N, Trivedi TI: **Prognostic significance of immunohistochemically localized biomarkers in stage II and stage III breast cancer: a multivariate analysis.** *Ann Surg Oncol* 2000, **7(4)**:305-11.
57. Jalava PJ, Collan YU, Kuopio T, Juntti-Patinen L, Kronqvist P: **Bcl-2 immunostaining: a way to finding unresponsive postmenopausal N+ breast cancer patients.** *Anticancer Res* 2000, **20(2B)**:1213-9.
58. Laroye GJ, Minkin S: **The impact of mitotic index on predicting outcome in breast carcinoma: a comparison of different counting methods in patients with different lymph node status.** *Mod Pathol* 1991, **4(4)**:456-60.
59. Aaltomaa S, Lipponen P, Eskelinen M, Kosma VM, Marin S, Alhava E, Syrjanen K: **Mitotic indexes as prognostic predictors in female breast cancer.** *J Cancer Res Clin Oncol* 1992, **118**:75-81.
60. Haapasalo H, Pesonen E, Collan Y: **Volume corrected mitotic index (M/V-INDEX). The standard of mitotic activity in neoplasms.** *Pathol Res Pract* 1989, **185**:551-4.
61. Kujari HP, Collan YU, Atkin NB: **Use of the mitotic counts for the prognosis and grading of breast cancer. Method evaluation study.** *Pathol Res Pract* 1994, **190(6)**:593-9.
62. Baak JP, Van Dop H, Kurver PH, Hermans J: **The value of morphometry to classic prognosticators in breast cancer.** *Cancer* 1985, **56(2)**:374-82.
63. Lipponen P, Papinaho S, Eskelinen M, Klemi PJ, Aaltomaa S, Kosma VM, Marin S, Syrjanen K: **DNA ploidy, S-phase fraction and mitotic indices as prognostic predictors of female breast cancer.** *Anticancer Res* 1992, **12**:1533-8.
64. Collan YU, Kuopio T, Baak JP, Becker R, Bogomoletz WV, Deverell M, van Diest P, van Galen C, Gilchrist K, Javed A, Kosma VM, Kujari H, Luzzi P, Mariuzzi GM, Matze E, Montironi R, Scarpelli M, Sierra D, Sisti S, Toikkanen S, Tosi P, Whimster WF, Wisse E: **Standardized mitotic counts in breast cancer. Evaluation of the method.** *Pathol Res Pract* 1996, **192(9)**:931-41.
65. Ikpat OF, Kuopio T, Collan Y: **Proliferation in African breast cancer: biology and prognostication in Nigerian breast cancer material.** *Mod Pathol* 2002, **15(8)**:783-9.
66. Groenendijk RP, Bult P, Noppen CM, Boetes C, Ruers TJ, Wobbes T: **Mitotic activity index in interval breast cancers.** *Eur J Surg Oncol* 2003, **29(1)**:29-31.
67. Lipponen PK, Collan Y, Eskelinen MJ: **Volume corrected mitotic index (M/V index), mitotic activity index (MAI), and histological grading in breast cancer.** *Int Surg* 1991, **76**:245-9.
68. Simpson JF, Gray R, Dressler LG, Cobau CD, Falkson CI, Gilchrist KW, Pandya KJ, Page DL, Robert NJ: **Prognostic value of histologic grade and proliferative activity in axillary node-positive breast cancer: results from the Eastern Cooperative Oncology Group Companion Study, EST 4189.** *J Clin Oncol* 2000, **18(10)**:2059-69.
69. Baak JP, van Diest PJ, Voorhorst FJ, van der Wall E, Beex LV, Vermorken JB, Janssen EA: **Prospective multicenter validation of the independent prognostic value of the mitotic activity index in lymph node-negative breast cancer patients younger than 55 years.** *J Clin Oncol* 2005, **23(25)**:5993-6001.
70. Baak JP, Colpaert CG, van Diest PJ, Janssen E, van Diermen B, Albernaz E, Vermeulen PB, Van Marck EA: **Multivariate prognostic evaluation of the mitotic activity index and fibrotic focus in node-negative invasive breast cancers.** *Eur J Cancer* 2005, **41(14)**:2093-101.
71. Kronqvist P, Kuopio T, Collan Y: **Morphometric grading in breast cancer: thresholds for mitotic counts.** *Hum Pathol* 1998:1462-8.
72. Gerdes J, Schwab U, Lemke H, Stein H: **Production of a mouse monoclonal antibody reactive with a human nuclear antigen associated with cell proliferation.** *Int J Cancer* 1983, **31(1)**:13-20.
73. Cooper LS, Gillett CE, Smith P, Fentiman IS, Barnes DM: **Cell proliferation measured by MIB1 and timing of surgery for breast cancer.** *Br J Cancer* 1998, **77(9)**:1502-7.
74. Gerdes J, Lemke H, Baisch H, Wacker HH, Schwab U, Stein H: **Cell cycle analysis of a cell proliferation-associated human nuclear antigen defined by the monoclonal antibody Ki-67.** *J Immunol* 1984, **133(4)**:1710-5.
75. Gerdes J, Li L, Schlueter C, Duchrow M, Wohlenberg C, Gerlach C, Stahmer I, Kloth S, Brandt E, Flad HD: **Immunobiochemical and molecular biologic characterization of the cell proliferation-associated nuclear antigen that is defined by monoclonal antibody Ki-67.** *Am J Pathol* 1991, **138(4)**:867-73.
76. Isola JJ, Helin HJ, Helle MJ, Kallioniemi OP: **Evaluation of cell proliferation in breast carcinoma. Comparison of Ki-67 immunohistochemical study, DNA flow cytometric analysis, and mitotic count.** *Cancer* 1990, **65(5)**:1180-4.
77. Railo M, Nordling S, von Boguslawsky K, Leivonen M, Kyllonen L, von Smitten K: **Prognostic value of Ki-67 immunolabelling in primary operable breast cancer.** *Br J Cancer* 1993, **8(3)**:579-83.
78. Pietilainen T, Lipponen P, Aaltomaa S, Eskelinen M, Kosma VM, Syrjanen K: **The important prognostic value of Ki-67 expression as determined by image analysis in breast cancer.** *J Cancer Res Clin Oncol* 1996, **122(11)**:687-92.
79. Jalava PJ, Kuopio T, Juntti-Patinen L, Kotkansalo T, Kronqvist P, Collan YU: **Ki-67 immunohistochemistry: A valuable marker in prognostication but also a risk of misclassification. Proliferation subgroups based on Ki-67 fraction and standardized mitotic index.** *Histopathology* 2006, **48**:674-682.
80. Santamaria L, Martin R, Gomez V, Ingelmo I, Lopez C, Revestido R: **Stereologic estimation of Ki-67, caspase3, and GSTPI positive cells in prostate lesions.** *Image Analysis Stereol* 2005, **24**:77-84.
81. Linden MD, Torres FX, Kubus J, Zarbo RJ: **Clinical application of morphologic and immunocytochemical assessments of cell proliferation.** *Am J Clin Pathol* 1992, **97(5 Suppl 1)**:S4-13.
82. Kesari AL, Chellam VG, Nair PP, Madhavan J, Nair P, Nair MK, Pillai MR: **Tumor proliferative fraction in infiltrating duct carcinoma.** *Gen Diagn Pathol* 1997, **143(4)**:219-24.
83. Jeziorski A, Blonski JZ, Niewiadomska H: **The expression of products of oncogenes c-erbB2 and EGFR and proliferating antigens Ki67 and PCNA in primary invasive ductal cancer of female breast.** *J Exp Clin Cancer Res* 2000, **19(1)**:61-7.
84. Aaltomaa S, Lipponen P, Syrjanen K: **Proliferating cell nuclear antigen (PCNA) immunolabeling as a prognostic factor in axillary lymph node negative breast cancer.** *Anticancer Res* 1993, **13(2)**:533-8.
85. Mourad WA, Erkman-Balis B, Livingston S, Shoukri M, Cox CE, Nicolsia SV, Rowlands DT Jr: **Argyrophilic nucleolar organizer**

- regions in breast carcinoma. Correlation with DNA flow cytometry, histopathology, and lymph node status. *Cancer* 1992, **69(7)**:1739-44.
86. Dasgupta A, Ghosh RN, Sarkar R, Laha RN, Ghosh TK, Mukherjee C: **Argyrophilic nucleolar organiser regions (AgNORs) in breast lesions.** *J Indian Med Assoc* 1997, **95(9)**:492-4.
 87. Aubele M, Auer G, Jutting U, Falkmer U, Gais P: **Prognostic value of quantitatively measured AgNORs in ductal mammary carcinoma.** *Anal Quant Cytol Histol* 1994:211-8.
 88. Rzymowska J: **AgNOR counts and their combination with flow cytometric analyses and clinical parameters as a prognostic indicator in breast carcinoma.** *Tumori* 1997, **83(6)**:938-42.
 89. Biesterfeld S, Farokhzad F, Kluppel D, Schneider S, Hufnagl P: **Improvement of breast cancer prognostication using cell kinetic-based silver-stainable nucleolar organizer region quantification of the MIB-1 positive tumor cell compartment.** *Virchows Arch* 2001, **438(5)**:478-84.
 90. Derenzini M, Ceccarelli C, Santini D, Taffurelli M, Trere D: **The prognostic value of the AgNOR parameter in human breast cancer depends on the pRb and p53 status.** *J Clin Pathol* 2004, **57(7)**:755-61.
 91. Toikkanen S, Joensuu H: **AgNOR counts have no prognostic value in breast cancer.** *J Pathol* 1993, **169(2)**:251-4.
 92. Simha M, Menon M, Doctor V: **Prognostic value of argyrophilic nucleolar organiser regions (AgNORs) in breast lesions.** *Indian J Cancer* 1996, **33(2)**:76-85.
 93. Ferno M, Baldetorp B, Borg A, Olsson H, Sigurdsson H, Killander D: **Flow cytometric DNA index and S-phase fraction in breast cancer in relation to other prognostic variables and to clinical outcome.** *Acta Oncol* 1992, **31(2)**:157-65.
 94. Michels JJ, Duigou F, Marnay J: **Flow cytometry in primary breast carcinomas. Prognostic impact of proliferative activity.** *Breast Cancer Res Treat* 2000, **62(2)**:117-26.
 95. Fietkau R, Iro H, Tulusan AH, Dressel V, Altendorf-Hofmann A, Sauer R: **Prognostic value of S-phase fraction in head and neck squamous cell carcinomas and nodal negative breast carcinomas.** *Strahlenther Onkol* 1994, **170(1)**:13-24.
 96. Zhang S, Yuan Y, Wang X: **[Prognosis prediction of S-phase fraction and p53, c-erbB-2, estrogen receptor, progesterone receptor in axillary node-negative breast cancer].** *Zhonghua Wai Ke Za Zhi* 1997, **35(8)**:475-7.
 97. Jones S, Clark G, Koleszar S, Ethington G, Mennel R, Paulson S, Brooks B, Kerr R, Denham C, Savin M, White C, Blum J, Kirby R, Stone M, Pippen J, Kitchens L, George T, Cooper B, Peters G, Knox S, Grant M, Cheek H, Jones R, Kuhn J, Lieberman Z, Savino D, Rietz C: **Low proliferative rate of invasive node-negative breast cancer predicts for a favorable outcome: a prospective evaluation of 669 patients.** *Clin Breast Cancer* 2001, **1(4)**:310-4.
 98. Collan Y, Klemi P, Kallioniemi OP, Joensuu H, Nordling S, Eskelinen M: **Significance of variation in DNA flow cytometric analyses from paraffin-embedded breast cancers. Evaluation of the grading efficiency of ploidy determination, DNA index, and S-phase fraction.** *Pathol Res Pract* 1992, **188(4-5)**:581-6.
 99. Greco S, Marsigliante S, Leo G, Storelli C: **Co-expression of thymidine kinase and cathepsin D in 200 primary breast carcinomas.** *Cancer Lett* 2000, **160(1)**:13-9.
 100. Isola J, Weitz S, Visakorpi T, Holli K, Shea R, Khabbaz N, Kallioniemi OP: **Cathepsin D expression detected by immunohistochemistry has independent prognostic value in axillary node-negative breast cancer.** *J Clin Oncol* 1993, **11(1)**:36-43.
 101. Billgren AM, Rutqvist LE, Johansson H, Hagerstrom T, Skoog L: **The role of cathepsin D and PAI-1 in primary invasive breast cancer as prognosticators and predictors of treatment benefit with adjuvant tamoxifen.** *Eur J Cancer* 2000, **36(11)**:1374-80.
 102. Rochefort H: **Estrogens, cathepsin D and metastasis in cancers of the breast and ovary: invasion or proliferation?** *CR Seances Soc Biol Fil* 1998, **192(2)**:241-51.
 103. Aaltonen M, Lipponen P, Kosma VM, Aaltomaa S, Syrjanen K: **Prognostic value of cathepsin-D expression in female breast cancer.** *Anticancer Res* 1995, **15(3)**:1033-7.
 104. Ramirez-Ortega MC, Frias-Mendivil M, Delgado-Chavez R, Meneses-Garcia A, Carrillo-Hernandez JF, Ramirez-Ugalde MT, Zeichner-Gancz I: **Expression of cathepsin D in breast cancer and its clinical and histopathological correlations.** *Rev Invest Clin* 1997, **49(5)**:361-8.
 105. Lippman ME, Dickson RB, Gelmann EP, Rosen N, Knabbe C, Bates S, Bronzert D, Huff K, Kasid A: **Growth regulation of human breast carcinoma occurs through regulated growth factor secretion.** *J Cell Biochem* 1987, **35(1)**:1-16.
 106. Umekita Y, Ohi Y, Sagara Y, Yoshida H: **Co-expression of epidermal growth factor receptor and transforming growth factor- α predicts worse prognosis in breast-cancer patients.** *Int J Cancer* 2000, **89(6)**:484-7.
 107. Kumar RR, Meenakshi A, Sivakumar N: **Enzyme immunoassay of human epidermal growth factor receptor (hEGFR).** *Hum Antibodies* 2001, **10(3-4)**:143-7.
 108. Aziz SA, Pervez S, Khan S, Kayani N, Rahbar MH: **Epidermal growth factor receptor (EGFR) as a prognostic marker: an immunohistochemical study on 315 consecutive breast carcinoma patients.** *J Pak Med Assoc* 2002, **52(3)**:104-10.
 109. Rampaul RS, Pinder SE, Wencyk PM, Nicholson RI, Blamey RW, Robertson JF, Ellis IO: **Epidermal growth factor receptor status in operable invasive breast cancer: is it of any prognostic value?** *Clin Cancer Res* 2004, **10(7)**:2578.
 110. Tsutsui S, Ohno S, Murakami S, Hachitanda Y, Oda S: **Prognostic value of epidermal growth factor receptor (EGFR) and its relationship to the estrogen receptor status in 1029 patients with breast cancer.** *Breast Cancer Res Treat* 2002, **71(1)**:67-75.
 111. Ellis MJ, Jenkins S, Hanfelt J, Redington ME, Taylor M, Leek R, Siddle K, Harris A: **Insulin-like growth factors in human breast cancer.** *Breast Cancer Res Treat* 1998, **52(1-3)**:175-84.
 112. Yu H, Levesque MA, Khosravi MJ, Papanastasiou-Diamandi A, Clark GM, Diamandis EP: **Associations between insulin-like growth factors and their binding proteins and other prognostic indicators in breast cancer.** *Br J Cancer* 1996, **74(8)**:1242-7.
 113. Surmacz E, Guvakova MA, Nolan MK, Nicosia RF, Sciacca L: **Type I insulin-like growth factor receptor function in breast cancer.** *Breast Cancer Res Treat* 1998, **47(3)**:255-67.
 114. Eppler E, Zapf J, Bailer N, Falkmer UG, Falkmer S, Reinecke M: **IGF-I in human breast cancer: low differentiation stage is associated with decreased IGF-I content.** *Eur J Endocrinol* 2002, **146(6)**:813-21.
 115. Toropainen EM, Lipponen PK, Syrjanen KJ: **Expression of insulin-like growth factor II in female breast cancer as related to established prognostic factors and long-term prognosis.** *Anticancer Res* 1995, **15(6B)**:2669-74.
 116. Blanco G, Alavaikko M, Ojala A, Collan Y, Heikkinen M, Hietanen T, Aine R, Taskinen PJ: **Estrogen and progesterone receptors in breast cancer: relationships to tumour histopathology and survival of patients.** *Anticancer Res* 1984, **4(6)**:383-9.
 117. Barbareschi M, Doglioni C: **[The immunohistochemical detection of steroid hormone receptors in breast cancer: open problems and new perspectives].** *Pathologica* 2002, **94(3)**:115-20.
 118. DeSombre ER, Kledzik G, Marshall S, Meites J: **Estrogen and prolactin receptor concentrations in rat mammary tumors and response to endocrine ablation.** *Cancer Res* 1976, **36(2 Pt 1)**:354-8.
 119. Silvestrini R, Daidone MG, Luisi A, Boracchi P, Mezzetti M, Di Fronzo G, Andreola S, Salvadori B, Veronesi U: **Biologic and clinicopathologic factors as indicators of specific relapse types in node-negative breast cancer.** *J Clin Oncol* 1995, **13(3)**:697-704.
 120. Talley LI, Grizzle WE, Waterbor JW, Brown D, Weiss H, Frost AR: **Hormone receptors and proliferation in breast carcinomas of equivalent histologic grades in pre- and postmenopausal women.** *Int J Cancer* 2002, **98(1)**:118-27.
 121. Nomura Y, Miura S, Koyama H, Enomoto K, Kasumi F, Yamamoto H, Kimura M, Tominaga T, Iino H, Morimoto T, Tashiro H: **Relative effect of steroid hormone receptors on the prognosis of patients with operable breast cancer. A univariate and multivariate analysis 3089 Japanese patients with breast cancer from the Study Group for the Japanese Breast Cancer Society on Hormone Receptors and Prognosis in Breast Cancer.** *Cancer* 1992, **69(1)**:153-64.
 122. Abd El-Rehim DM, Pinder SE, Paish CE, Bell JA, Rampaul RS, Blamey RW, Robertson JF, Nicholson RI, Ellis IO: **Expression and co-expression of the members of the epidermal growth factor receptor (EGFR) family in invasive breast carcinoma.** *Br J Cancer* 2004, **91(8)**:1532-42.
 123. Jalava P, Kuopio T, Huovinen R, Laine J, Collan Y: **Immunohistochemical staining of estrogen and progesterone receptors:**

- aspects for evaluating positivity and defining the cutpoints. *Anticancer Res* 2005, **25**(3c):2535-42.
124. Bocking A, Giroud F, Reith A: **Consensus report of the European Society for Analytical Cellular Pathology task force on standardization of diagnostic DNA image cytometry.** *Anal Quant Cytol Histol* 1995, **17**(1):1-7.
 125. Auer GU, Caspersson TO, Wallgren AS: **DNA content and survival in mammary carcinoma.** *Anal Quant Cytol* 1980:161-5.
 126. Erhardt K, Auer G: **Mammary carcinoma: DNA analysis in areas showing different histological features in the same tumor.** *Acta Pathol Microbiol Immunol Scand [A]* 1986, **94**(1):21-8.
 127. Bocking A, Chatelain R, Biesterfeld S, Noll E, Biesterfeld D, Wohltmann D, Goecke C: **DNA grading of malignancy in breast cancer. Prognostic validity, reproducibility and comparison with other classifications.** *Anal Quant Cytol Histol* 1989, **11**(2):73-80.
 128. Friedrich K, Scheithauer J, Dimmer V, Meyer W, Theissig F, Haroske G, Kunze KD: **DNA ploidy and chromosomal imbalances in invasive ductal breast cancer. A comparative study of DNA image cytometry and comparative genomic hybridization (CGH).** *Anal Cell Pathol* 2000, **20**(2-3):69-82.
 129. Tsutsui S, Ohno S, Murakami S, Hachitanda Y, Oda S: **Prognostic value of DNA ploidy in 653 Japanese women with node-negative breast cancer.** *Int J Clin Oncol* 2001, **6**(4):177-82.
 130. Wong SW, Rangan AM, Bilous AM, Boyages J, Gebski V, Benson EM: **The value of S-phase and DNA ploidy analysis as prognostic markers for node-negative breast cancer in the Australian setting.** *Pathology* 1999, **31**(2):90-4.
 131. Bracko M, Us-Krasovec M, Cufer T, Lamovec J, Zidar A, Goehde W: **Prognostic significance of DNA ploidy determined by high-resolution flow cytometry in breast carcinoma.** *Anal Quant Cytol Histol* 2001, **23**(1):56-66.
 132. Yuan J, Hennessy C, Corbett IP, Dykin R, Givan AL, Shenton BK, Henry JA, Wright C, Lennard TW: **Node negative breast cancer: the prognostic value of DNA ploidy for long-term survival.** *Br J Surg* 1991, **78**(7):844-8.
 133. Chassevent A, Jourdan ML, Romain S, Descotes F, Colonna M, Martin PM, Bolla M, Spyrtos F: **S-phase fraction and DNA ploidy in 633 T1T2 breast cancers: a standardized flow cytometric study.** *Clin Cancer Res* 2001, **7**(4):909-17.
 134. Moureau-Zabotto L, Bouchet C, Cesari D, Uzan S, Lefranc JP, Antoine M, Genestie C, Deniaud-Alexandre E, Bernaudin JF, Touboul E, Fleury-Feith J: **Combined flow cytometry determination of S-phase fraction and DNA ploidy is an independent prognostic factor in node-negative invasive breast carcinoma: analysis of a series of 271 patients with stage I and II breast cancer.** *Breast Cancer Res Treat* 2005, **91**(1):61-71.
 135. Collan YU, Eskelinen MJ, Nordling SA, Lipponen P, Pesonen E, Kumpusalo LM, Pajarinen P, Kettunen KO: **Prognostic studies in breast cancer. Multivariate combination of nodal status, proliferation index, tumor size, and DNA ploidy.** *Acta Oncol* 1994, **33**(8):873-8.
 136. Aaltomaa S, Lipponen P, Papinaho S, Klemi P, Kosma VM, Marin M, Alhava E, Syrjanen K: **Nuclear morphometry and DNA flow cytometry as prognostic factors in female breast cancer.** *Eur J Surg* 1992, **3**:135-41.
 137. Zajdela A, De LaRiva LS, Ghossein NA: **The relation of prognosis to the nuclear diameter of breast cancer cells obtained by cytologic aspiration.** *Acta Cytologica* 1979, **23**:75-80.
 138. Tosi P, Luzi P, Sforza V, Santopietro R, Bindi M, Tucci E, Barbini P, Baak JPA: **Correlation between morphometrical parameters and disease-free survival in ductal breast cancer treated by surgery.** *Appl Pathol* 1986, **4**:33-42.
 139. Sarker S: **Mean nuclear area of fine needle aspirates of primary preoperative palpable breast carcinoma using image cytometry.** *Anal Quant Cytol Histol* 2002, **24**(2):85-8.
 140. Kronqvist P, Kuopio T, Collan Y: **Morphometric grading of invasive ductal breast cancer. I. Thresholds for nuclear grade.** *Br J Cancer* 1998, **78**(6):800-5.
 141. Gonzalez MA, Pinder SE, Wencyk PM, Bell JA, Elston CW, Nicholson RI, Robertson JF, Blamey RW, Ellis IO: **An immunohistochemical examination of the expression of E-cadherin, alpha- and beta/gamma-catenins, and alpha2- and beta1-integrins in invasive breast cancer.** *J Pathol* 1999, **187**(5):523-9.
 142. Lipponen P, Saarelainen E, Ji H, Aaltomaa S, Syrjanen K: **Expression of E-cadherin (E-CD) as related to other prognostic factors and survival in breast cancer.** *J Pathol* 1994, **174**(2):101-9.
 143. Asgeirsson KS, Jonasson JG, Tryggvadottir L, Olafsdottir K, Sigurgeirsdottir JR, Ingvarsson S, Ogmundsdottir HM: **Altered expression of E-cadherin in breast cancer. patterns, mechanisms and clinical significance.** *Eur J Cancer* 2000:1098-106.
 144. Siitonen SM, Kononen JT, Helin HJ, Rantala IS, Holli KA, Isola JJ: **Reduced E-cadherin expression is associated with invasiveness and unfavorable prognosis in breast cancer.** *Am J Clin Pathol* 1996, **105**(4):394-402.
 145. Asgeirsson KS, Olafsdottir K, Jonasson JG, Ogmundsdottir HM: **The effects of IL-6 on cell adhesion and e-cadherin expression in breast cancer.** *Cytokine* 1998:720-8.
 146. Berx G, Van Roy F: **The E-cadherin/catenin complex: an important gatekeeper in breast cancer tumorigenesis and malignant progression.** *Breast Cancer Res* 2001, **3**(5):289-93.
 147. Charpin C, Garcia S, Bouvier C, Devictor B, Andrac L, Choux R, Lavaut M: **E-cadherin quantitative immunocytochemical assays in breast carcinomas.** *J Pathol* 1997, **181**(3):294-300.
 148. Bankfalvi A, Terpe HJ, Breukelmann D, Bier B, Rempe D, Pschadka G, Krech R, Lelle RJ, Boecker W: **Immunophenotypic and prognostic analysis of E-cadherin and beta-catenin expression during breast carcinogenesis and tumour progression: a comparative study with CD44.** *Histopathology* 1999, **34**(1):25-34.
 149. Gamallo C, Palacios J, Suarez A, Pizarro A, Navarro P, Quintanilla M, Cano A: **Correlation of E-cadherin expression with differentiation grade and histological type in breast carcinoma.** *Am J Pathol* 1993, **142**(4):987-93.
 150. Elzagheid A, Kuopio T, Ilmen M, Collan Y: **Prognostication of invasive ductal breast cancer by quantification of E-cadherin immunostaining: the methodology and clinical relevance.** *Histopathology* 2002, **41**(2):127-33.
 151. Rakha EA, Abd El Rehim D, Pinder SE, Lewis SA, Ellis IO: **E-cadherin expression in invasive non-lobular carcinoma of the breast and its prognostic significance.** *Histopathology* 2005, **46**(6):685-93.
 152. Hirohashi S: **Inactivation of the E-cadherin-mediated cell adhesion system in human cancers.** *Am J Pathol* 1998, **153**(2):333-9.
 153. Ghadimi BM, Behrens J, Hoffmann I, Haensch W, Birchmeier W, Schlag PM: **Immunohistological analysis of E-cadherin, alpha-, beta- and gamma-catenin expression in colorectal cancer: implications for cell adhesion and signaling.** *Eur J Cancer* 1999, **35**(1):60-5.
 154. Yoshida R, Kimura N, Harada Y, Ohuchi N: **The loss of E-cadherin, alpha- and beta-catenin expression is associated with metastasis and poor prognosis in invasive breast cancer.** *Int J Oncol* 2001, **18**(3):513-20.
 155. Lim SC, Lee MS: **Significance of E-cadherin/beta-catenin complex and cyclin D1 in breast cancer.** *Oncol Rep* 2002:915-28.
 156. Foekens JA, Dall P, Klijn JG, Skroch-Angel P, Claassen CJ, Look MP, Ponta H, Van Putten WL, Herrlich P, Henzen-Logmans SC: **Prognostic value of CD44 variant expression in primary breast cancer.** *Int J Cancer* 1999, **84**(3):209-15.
 157. Morris SF, O'Hanlon DM, McLaughlin R, McHale T, Connolly GE, Given HF: **The prognostic significance of CD44s and CD44v6 expression in stage two breast carcinoma: an immunohistochemical study.** *Eur J Surg Oncol* 2001, **27**(6):527-31.
 158. Joensuu H, Klemi P, Toikkanen S, Jalkanen S: **Glycoprotein CD44 expression and its association with survival in breast cancer.** *Am J Pathol* 1993, **143**(3):867-74.
 159. Tempfer C, Losch A, Heinzl H, Hausler G, Hanzal E, Kolbl H, Breitenacker G, Kainz C: **Prognostic value of immunohistochemically detected CD44 isoforms CD44v5, CD44v6 and CD44v7-8 in human breast cancer.** *Eur J Cancer* 1996, **32A**(11):2023-5.
 160. Moch H, Torhorst J, Durmuller U, Feichter GE, Sauter G, Gudat F: **Comparative analysis of the expression of tenascin and established prognostic factors in human breast cancer.** *Pathol Res Pract* 1993, **189**(5):510-4.
 161. Shoji T, Kamiya T, Tsubura A, Hamada Y, Hatano T, Hioki K, Morii S: **Tenascin staining positivity and the survival of patients with invasive breast carcinoma.** *J Surg Res* 1993, **55**(3):295-7.
 162. Yoshida T, Ishihara A, Hirokawa Y, Kusakabe M, Sakakura T: **Tenascin in breast cancer development - is epithelial tenascin a marker for poor prognosis?** *Cancer Lett* 1995, **90**(1):65-73.

163. Hidalgo M, Eckhardt SG: **Development of matrix metalloproteinase inhibitors in cancer therapy.** *J Natl Cancer Inst* 2001, **93(3)**:178-93.
164. Vihinen PP, Pyrhonen SO, Kahari VM: **New prognostic factors and developing therapy of cutaneous melanoma.** *Ann Med* 2003, **35(2)**:66-78.
165. Vihinen P, Kahari VM: **Matrix metalloproteinases in cancer: prognostic markers and therapeutic targets.** *Int J Cancer* 2002, **99(2)**:157-66.
166. Talvensaari-Mattila A, Paakko P, Hoyhtya M, Blanco-Sequeiros G, Turpeenniemi-Hujanen T: **Matrix metalloproteinase-2 immunoreactive protein: a marker of aggressiveness in breast carcinoma.** *Cancer* 1998, **83(6)**:1153-62.
167. Voltersvik P, Bostad L, Dyrhol-Riise AM, Eide GE, Rosok BI, Olofsson J, Asjo B: **Cystatin A and HIV-1 p24 antigen expression in tonsillar lymphoid follicles during HIV-1 infection and during highly active antiretroviral therapy.** *J Acquir Immune Defic Syndr* 2006, **41(3)**:277-84.
168. Lah TT, Kos J, Blejec A, Frkovic-Georgio S, Golouh R, Vrhovec II, Turk VV: **The Expression of Lysosomal Proteinases and Their Inhibitors in Breast Cancer: Possible Relationship to Prognosis of the Disease.** *Pathol Oncol Res* 1997, **3(2)**:89-99.
169. Greenough RB: **Varying degrees of malignancy in cancer of the breast.** *J Cancer Res* 1925, **9**:452-463.
170. Patey DH, Scarff RW: **The position of histology in the prognosis of carcinoma of the breast.** *Lancet* 1928, **1**:801-804.
171. Bloom HJG, Richardson WW: **Histological grading and prognosis in breast cancer. A study of 1409 cases of which 359 have been followed for 15 years.** *Br J Cancer* 1957, **11**:359-377.
172. Elston CW, Ellis IO: **Pathological prognostic factors in breast cancer. I. The value of histological grade in breast cancer: experience from a large study with long-term follow-up.** *Histopathology* 1991, **19(5)**:403-10.
173. Page DL, Jensen RA, Simpson JF: **Premalignant and malignant disease of the breast: the roles of the pathologist.** *Mod Pathol* 1998, **11(2)**:120-8. Review
174. Cianfrocca M, Goldstein LJ: **Prognostic and predictive factors in early-stage breast cancer.** *Oncologist* 2004, **9(6)**:606-16. Review
175. Kronqvist P, Kuopio T, Collan Y: **Morphometric grading of breast cancer: thresholds for tubular differentiation.** *Br J Cancer* 2000, **82(10)**:1656-61.
176. Stenkvist B, Bengtsson E, Dahlqvist B, Eklund G, Eriksson O, Jarkrans T, Nordin B: **Predicting breast cancer recurrence.** *Cancer* 1982, **50(12)**:2884-93.
177. Eskelinen M, Collan Y, Pajarinen P, Pesonen E, Kettunen K, Nordling S: **An improved prognostic index of axillary node involvement in breast cancer incorporating DNA ploidy and tumour size.** *Acta Chir Scand* 1990, **156(8)**:521-7.
178. Collan YU, Eskelinen MJ, Nordling SA, Lipponen P, Pesonen E, Kumpulainen LM, Pajarinen P, Kettunen KO: **Prognostic studies in breast cancer. Multivariate combination of nodal status, proliferation index, tumor size, and DNA ploidy.** *Acta Oncol* 1994, **33(8)**:873-8.
179. Collan Y, Kuopio T, Auranen A, Linna M: **Prognostication of breast cancer by multivariate methods.** *Adv Clin Path* 1997, **1(4)**:269-273.
180. Moureau-Zabotto L, Bouchet C, Cesari D, Uzan S, Lefranc JP, Antoine M, Genestie C, Deniaud-Alexandre E, Bernaudin JF, Touboul E, Fleury-Feith J: **Combined flow cytometry determination of S-phase fraction and DNA ploidy is an independent prognostic factor in node-negative invasive breast carcinoma: analysis of a series of 271 patients with stage I and II breast cancer.** *Breast Cancer Res Treat* 2005, **91(1)**:61-71.

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