

Research

Open Access

Single dose of morphine produced a prolonged effect on dopamine neuron activities

Die Zhang, Hai Zhang, Guo-zhang Jin, Kehong Zhang and Xuechu Zhen*

Address: State Key Laboratory of Drug Research, Department of Neuropharmacology, Shanghai Institute of Material Medica, Chinese Academy of Sciences, Shanghai, PR China

Email: Die Zhang - drizzt214@hotmail.com; Hai Zhang - zhangsea81@126.com; Guo-zhang Jin - gzjin@mail.shcnc.ac.cn; Kehong Zhang - Lab309@mail.shcnc.ac.cn; Xuechu Zhen* - xczhen@mail.shcnc.ac.cn

* Corresponding author

Published: 17 November 2008

Received: 17 August 2008

Molecular Pain 2008, 4:57 doi:10.1186/1744-8069-4-57

Accepted: 17 November 2008

This article is available from: <http://www.molecularpain.com/content/4/1/57>

© 2008 Zhang et al; licensee BioMed Central Ltd.

This is an Open Access article distributed under the terms of the Creative Commons Attribution License (<http://creativecommons.org/licenses/by/2.0>), which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.

Abstract

Background: Clinical observation and experimental studies have indicated that a single exposure to morphine could induce tolerance and dependence. It has become a concern in clinical antinociceptive practice. However, the underlying mechanism remains unknown. This study was designed to explore the changes of dopamine (DA) neuron activities in the ventral tegmental area (VTA) by employing a spectral analysis followed single morphine treatment.

Results: Acute morphine treatment significantly increased not only the firing rate and firing population but also the power of slow oscillation of DA neurons in naïve rats. These changes lasted at least for 3 days following the morphine treatment. During this period of time, responses of the DA neurons to subsequent morphine challenge were diminished. We further demonstrated a transient desensitization of opiate receptors as monitored by GTP γ S binding to G-proteins. The present study provided first direct evidence for the temporal changes in the VTA DA neuron activities and opiate receptors desensitization.

Conclusion: Prolonged VTA DA neuron activation and opiate receptors desensitization followed single morphine exposure may underlie the development of dependence and tolerance that may associate with the acute analgesic tolerance and acute addiction of morphine.

Introduction

Dopamine (DA) neurons in the ventral tegmental area (VTA) project primarily to the nucleus accumbens and the prefrontal cortex, forming the mesocorticolimbic system [1-3]. Like other drugs of abuse, morphine influence DA neurotransmission directly or indirectly [4,5]. Positive reinforcement of drugs of abuse is believed to be mediated through the activation of the mesocorticolimbic DA system [6]. Repeated exposure to morphine induce substantial adaptive changes in cellular and synaptic functions in the mesocorticolimbic DA system [7,8]. These adapta-

tions are believed to play a critical role in the development of tolerance and dependence [9,10].

Clinical and experimental evidences have revealed that a single exposure to morphine could induce rapid tolerance and dependence that may associate with acute addiction and analgesic tolerance [11-14]. Increased firing activity of VTA DA neurons in response to acute morphine administration has been reported [15-17]. However, the long-lasting effect and the time course (onset, duration, and dissipation) on the activity of VTA DA neurons and how

this change may contribute to the single morphine exposure-induced dependence or tolerance that may associate with decreased analgesic potency for morphine remains inadequately defined.

In the present study, we examined the firing activities of the VTA DA neurons at various time points after a single morphine injection. In addition to the traditional electrophysiological parameters such as firing rate and bursting, we also included slow oscillation (SO), SO represents a novel and major firing pattern of DA neurons in the VTA [18]. The SO DA neurons is characterized by a low frequency oscillation at 0.5–1.5 Hz [18,19]. Previous works in this laboratory and others have demonstrated that nearly 50 percentage of VTA DA neurons exhibits SO in basal condition and psychostimulants were found to alter SO [20,21]. Very recently, we demonstrated that SO in the VTA DA neurons is largely dependent on the input from the prefrontal cortex (PFC)[22], further suggesting the importance of SO in VTA DA neuron functional regulation.

The results from the current study indicated that the firing rate, bursting, SO, and the firing population of the VTA DA neurons in rats remain increased for at least 3 days after a single morphine exposure. Within this limited window of time, the VTA DA neurons failed to respond to additional morphine challenge. Indicating a transient morphine tolerance in VTA DA neuron activity in rats was developed with a single dose of morphine treatment. We further demonstrated that the acute morphine tolerance was associated with impairment of opiate receptor-G protein coupling, indicating that down regulation of G-protein activation may contribute to acute morphine tolerance.

Methods

Materials

Morphine hydrochloride was purchased from Qinghai Pharmaceutical Factory (Qinghai, China) and dissolved in saline. [³⁵S] GTPγS (1050 Ci/mmol) was purchased from Amersham Biosciences (Piscataway, USA). Other chemicals were obtained from standard suppliers.

Animal Preparations

All procedures were in compliance with the Guidelines for the Care and Use of Laboratory Animals (National Research Council, People's Republic of China, 1996). Male Sprague-Dawley (SD) rats, with an initial body weight of 270 to 300 g, were maintained on a 12 h light (8:00 am-8:00 pm)/12 h dark schedule at a temperature of 22 ± 2 °C and 65% humidity. Access to standard food and water was unlimited. Rats were acclimated to the animal facility for at least 7 days before experiments. All experiments were conducted between 10:00 am and 4:00

pm. For the initial exposure, morphine was given subcutaneously (s.c.) at a dose of 20 mg/kg in a volume of 2 ml/kg. For subsequent challenge, morphine was given intravenously (i.v.) at a dose of 5 mg/kg in a volume of 1 ml/kg[23,24].

Single Unit Recording

Rats were anesthetized with choral hydrate (400 mg/kg, i.p.) and mounted in a stereotaxic apparatus (Narishige, Japan). Body temperature was maintained at 36–37 °C with a homeothermic blanket system (Harvard, USA). Glass microelectrodes were filled with 2 M NaCl and 0.2% pontamine Sky Blue as a tracer. Resistance ranged from 6 to 8 MΩ. Electrode was placed into the VTA through a small burr hole in the skull (3.0 mm anterior to the lambda and 0.5–0.9 mm lateral to the midline) by an electro-microdriver. DA neurons, usually at 6.5–8.5 mm below the cortical surface, were identified according to the published criteria[19,21,25]: a biphasic (positive-negative) or triphasic (positive-negative-positive) spike waveform with a duration of greater than 3.0 ms, a broad initial positive phase (>1 ms, measured from the start of action potential to the negative trough), an initial segment-somatodendritic break in the initial positive phase, and a slow firing rate (<10 spikes/s). In addition, DA neurons can also be distinguished from non-DA neurons by the characteristic low-pitched sound that they produce on an audio monitor. Data were collected and analyzed using an acquisition system by the Fudan University (Shanghai, China) and stored in a computer for off-line analysis. After recording, rats were killed and brains were kept for histological exam to verify the location.

Population Recording (cells-per-track)

The number of spontaneously firing DA neurons within the VTA was determined by lowering the electrode through a area of tissue (400 × 600 μm). Twelve passes separated by 200 μm were made in each rat. The sequence of the passes and anatomical boundary of recorded area were identical among experimental subjects[19,21]. Only neurons with electrophysiological characteristics matching those previously established for midbrain DA neurons were included in the count. Each neuron was recorded and analyzed online for 2 to 5 minutes. The average number of spontaneously firing DA neurons encountered per electrode track (cells-per-track) was taken as an index for population activity of the VTA DA neurons.

Membrane preparation

Rats were sacrificed by decapitation. The brain was removed and immediately homogenized in 20 volumes of ice-cold 300 mM sucrose solution using a teflon/glass homogenizer. The homogenate was centrifuged at 1,000 g for 10 min at 4 °C. The supernatant was then centrifuged at 36,000 g for 30 min. The pellet was washed in 50 mM

Tris (pH 7.5) and centrifuged again at 36,000 g for 30 min. The final pellet was suspended in 50 mM Tris (pH 7.5). Protein concentration was determined using a BCA method.

GTP γ S-binding assay

GTP γ S-binding was performed in a reaction buffer containing: 50 mM Tris (pH7.5), 5 mM MgCl₂, 1 mM EDTA, 100 mM NaCl, and 1 mM DTT, 0.1 nM [³⁵S]GTP γ S, and 30 μ M GDP. Basal binding was determined in the absence of morphine. 10 μ M of morphine was used to determine the morphine-stimulated GTP γ S-binding. The assay was started by adding membrane preparation containing 15 μ g protein to 100 μ l reaction buffer. After 60 min of incubation at 30°C, the reaction was stopped with 3 ml ice-cold buffer. Samples were filtered rapidly through GF/C filters. Radioactivity in the filters was measured by liquid scintillation spectrometry. Nonspecific binding was determined using 10 μ M unlabeled GTP γ S.

Data Analyses

Data was analyzed off-line with a software package from BSPS Mathlib (Fudan University; Shanghai, China). The firing rate, bursting, and coefficient of variation (CV) were analyzed every 10 sec as described previously [19,22]. SO was analyzed using a program developed by this laboratory [21]. Data were analyzed by using either independent *t*-test or ANOVA. Spectral data were log-transformed before statistical analysis. All measures were based on a 2-min recording for each subject. All numerical results are expressed as mean \pm SEM.

Results

Acute morphine alters firing rate and firing pattern of the VTA DA neurons in naïve rats

Intravenous injection of morphine at a dose of 5 mg/kg significantly increased the firing rate of the VTA DA neurons (+21.1 \pm 8.6%, *n* = 8, paired *t*-test, *p* < 0.01; Figure. 1). Bursting (percentage of spikes firing in burst) and the coefficient of variation (CV) of interspike intervals (ISIs) were not affected. Moreover, the spectral analysis showed that acute morphine injection greatly enhanced VTA DA neurons slow oscillation (SO) power (+115.7 \pm 58.8%, Paired *t* test, *p* < 0.05). This indicates that acute morphine not only enhanced the firing rate of VTA DA neurons, but also altered the firing patterns.

Single morphine exposure induces prolonged changes of VTA DA neuron activity

As depicted in Figure. 2 & 3, at 24 hours after morphine exposure (20 mg/kg, s.c.), firing rate of the VTA DA neurons was significantly higher than that in the control rats receiving saline (3.99 \pm 0.14 spikes/sec, *t*-test, *p* < 0.05, compared to saline control). Bursting (20.96 \pm 2.12%, *p* < 0.001), CV of ISIs (69.66 \pm 2.40%, *p* < 0.001) and the

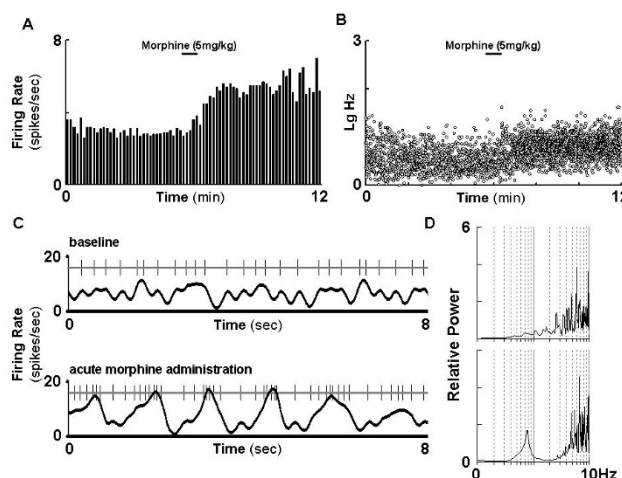


Figure 1
Morphine alters the firing rate and pattern of the VTA DA neurons in naïve rats. The firing activities were recorded before and after morphine injection (5 mg/kg, i.v.). A: firing rate of a representative DA neuron. B: distribution of firing rate in the same neuron. C: representative segment of firing spike train recorded from a DA neuron before and after morphine treatment. D: spectral chart before (upper panel) and after (lower panel) morphine treatment. SO: slow oscillation.

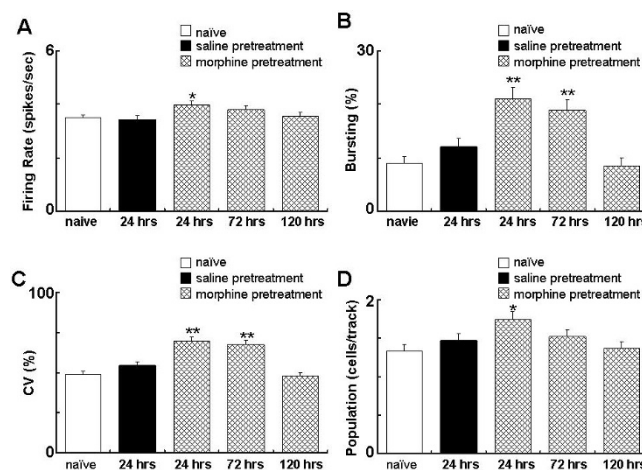


Figure 2
Single morphine exposure produces a prolonged change on the firing activities of the VTA DA neurons. Rats received morphine (20 mg/kg, s.c.) or saline. The firing activities were recorded at 24, 72 and 120 hours after morphine treatment. Naïve rats were used to indicate the basal activity. A: firing rate. B: bursting. C: CV of ISIs. D: cells/track. **P* < 0.05, ***P* < 0.01, compared to saline control. Independent group *t*-test.

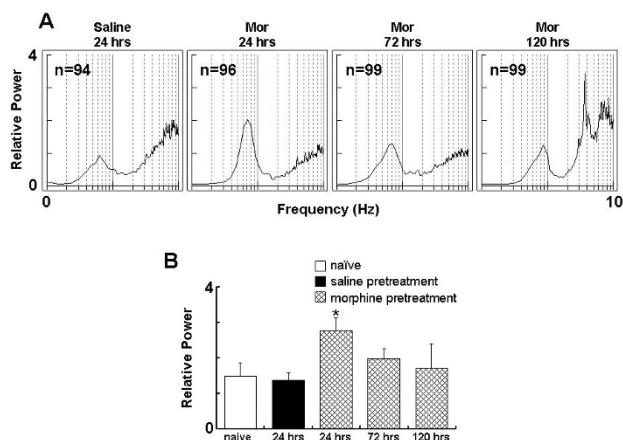


Figure 3
Single morphine exposure alters firing pattern of VTA DA neurons. Rats received morphine (20 mg/kg, s.c.) or saline. A: spectral charts at 24, 72, 120 hours after the treatment. B: group summary. *P < 0.05, compared to saline control. Mor: morphine.

number of firing VTA DA cells/track (1.74 ± 0.10 cells/track, $p < 0.05$). SO power (1.39 ± 0.17 , $p < 0.05$, compared to control) were also significantly higher than the corresponding measures in the control rats receiving saline.

At 72 hours after the exposure, the firing rate, SO power and the number of firing DA neurons were no longer significantly different from the saline control. However, the bursting level and CV of ISIs were still significantly higher than that in the saline control (bursting: $18.87 \pm 2.05\%$, $p < 0.001$; CV: $67.63 \pm 2.34\%$, $p < 0.001$). At 120 hours, all recorded parameters of the VTA DA neurons returned to a level comparable to that in the saline control group and are significantly lower than that of corresponding measurement obtained at 24 hours after morphine treatment. This result indicates that single dose of morphine induced a prolonged response in VTA DA neuron activity which lasted at least 72 hours.

VTA DA neurons fails to response to subsequent challenge after the initial exposure to morphine

We next tested how VTA DA neuron responses to subsequent morphine challenge at different time points after initial exposure. In contrast to the activation of VTA DA neurons in response to morphine challenge in naïve rats (Figure. 1), morphine challenge (5 mg/kg, i.v.) at 24 hours after receiving the initial morphine exposure failed to alter the firing rate ($-3.5 \pm 3.3\%$, t -test, $p = 0.322$) or CV of ISIs ($-13.0 \pm 7.9\%$, t -test, $p = 0.141$) of the VTA DA neu-

rons (Figure. 4 &5). Interestingly, the bursting ($-36.4 \pm 16.0\%$, t test, $p < 0.05$) was significantly decreased.

The failure of the VTA DA neurons in response to morphine challenge lasted at least 72 hours as depicted in firing rate ($+0.9 \pm 5.1\%$, t -test, $p = 0.867$), bursting ($+6.0 \pm 36.4\%$, t test, $p = 0.875$), CV of ISIs ($-11.6 \pm 8.5\%$, t -test, $p = 0.209$) and SO power. This indicated that VTA DA neurons developed a tolerance followed the single dose of morphine pretreatment. However, the response of DA neurons to morphine challenge was restored at 120 hours (Figure. 5): the firing rate was increased by $34.1 \pm 10.6\%$ (t -test, $p < 0.05$); The SO power, as shown in Fig. 6, was significantly decreased ($-41.8 \pm 13.2\%$, t -test, $p < 0.05$) at 24 h after initial morphine exposure, and was started to recover at 72 h ($+24.6 \pm 23.7\%$, t -test, $p = 0.331$), restored at 120 h ($118.5 \pm 62.0\%$, t -test, $p < 0.05$). Taken together, the present data demonstrated that single morphine administration results in prolonged unresponsiveness of VTA DA neurons to subsequent morphine challenge which lasted at least three days.

Inhibition on μ -Opioid-stimulated brain $[^{35}S]GTP\gamma S$ binding followed single morphine administration in rats

Since desensitization of opioid receptor is the important mechanism for morphine tolerance, we next studied whether the tolerance/dependence developed by single dose of morphine treatment is also associated with the

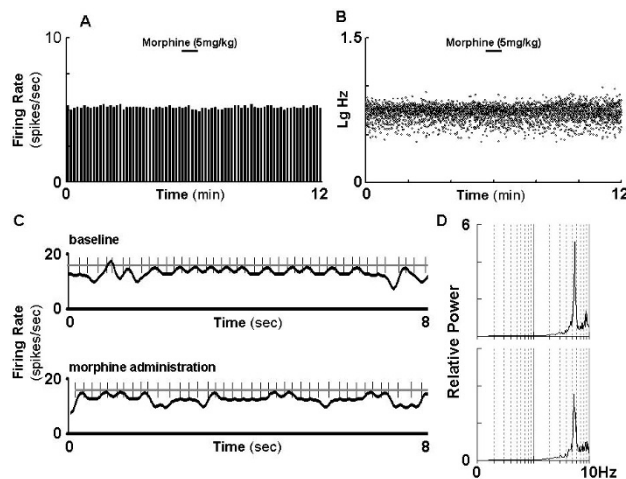


Figure 4
VTA DA neurons fail to response to subsequent challenge following a single morphine exposure. Rats received morphine (20 mg/kg, s.c.) or saline, and then challenged with morphine (5 mg/kg, i.v.) at 24 hours. A: firing rate. B: firing rate distribution in same neuron. C: representative segments of firing spike train before (upper panel) and after (lower panel) the challenge. D: spectral charts before (upper panel) and after (lower panel) the challenge.

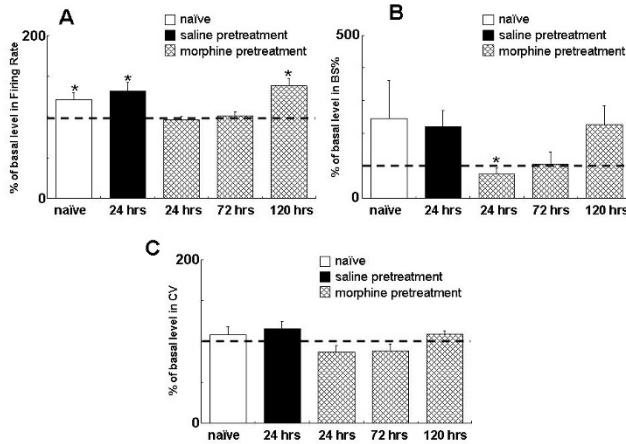


Figure 5
The firing rate of VTA DA is time-dependently changed in response to subsequent challenge after a single morphine exposure. Rats received morphine (20 mg/kg, s.c.) or saline, and then challenged with morphine (5 mg/kg, i.v.) 24, 72 or 120 hours later. Data are summary based on at least 5 subjects at each time point in each group and are expressed as % change induced by the challenge. A: firing rate. B: bursting level. C: CV. * $p < 0.05$, t -test.

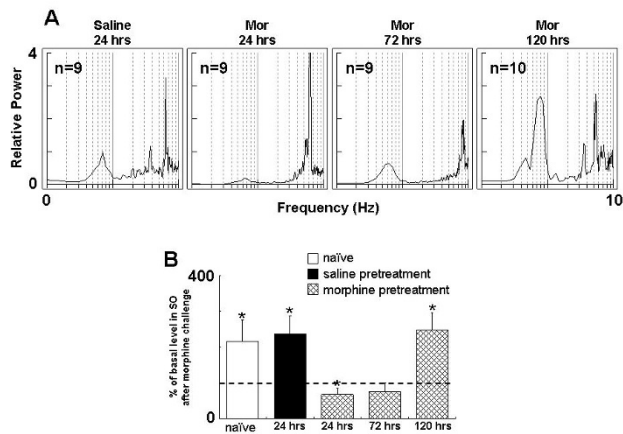


Figure 6
Slow oscillation (SO) of VTA DA neuron is time-dependently changed in response to morphine challenge after a single morphine exposure. Rats received morphine (20 mg/kg, s.c.) or saline, and then challenged with morphine (5 mg/kg, i.v.) 24, 72 or 120 hours later. A: representative spectral chart. B: group summary. * $P < 0.05$, Paired t test, compared to respective control.

change in opiate receptor-stimulated G protein coupling. We employed [³⁵S]GTP γ S binding assay to monitor the opioid receptor activation. As shown in Figure 7, the time-course of morphine-stimulated brain [³⁵S]GTP γ S binding in rats received saline or single dose of morphine was examined at 24, 72, 120 hours after drug or saline administration, respectively. We found that the morphine-stimulated [³⁵S]GTP γ S binding was increased to $127.4 \pm 1.6\%$ over the non-stimulated control ($n = 3$, $p < 0.001$) in membrane preparation obtained from saline-pretreated rats. In contrast, morphine failed to stimulate [³⁵S]GTP γ S binding in brain membrane preparation from rats at 24 hours after initial morphine treatment ($101.3 \pm 4.8\%$, $n = 3$). However, this stimulation was restored at 72 ($128.3 \pm 3.8\%$, $p < 0.05$) and 120 hours ($131.8 \pm 2.2\%$, $n = 3$; $p < 0.05$), respectively (Figure. 7). This indicated that opioid receptor failed to response to morphine stimulation for a prolonged period of time followed the single dose of morphine administration. This result suggested that development of uncoupling between the opioid receptor and related G-protein may contribute to the tolerance and dependence induced by single morphine administration.

Discussion

Morphine alters DA neurotransmission that is associated with rewarding circuit. Microdialysis assay has demonstrated an increased DA release in response to acute morphine administration which is generally believed to associate with the positive reinforcing effect of drug[23,26-29]. The underlying pathways for morphine-

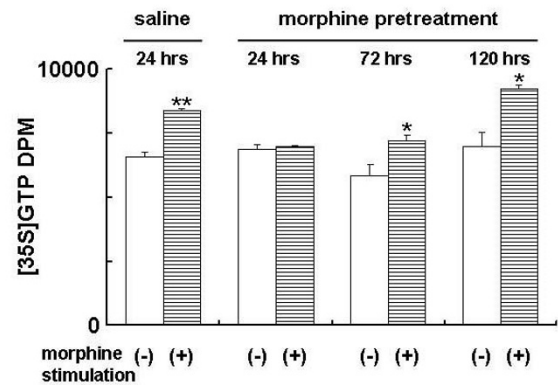


Figure 7
Morphine-stimulated GTP γ S binding is changed after a single morphine exposure. Rats received morphine (20 mg/kg, s.c.) or saline, and sacrificed 24, 72, or 120 hours later for morphine-stimulated GTP γ S binding assay (10 μ M of morphine or vehicle in incubation buffer). Data are expressed as mean \pm SEM obtained from at least 3 rats. ANOVA, * $p < 0.05$, ** $p < 0.01$, compared to respective control. (-): no morphine; (+): morphine.

modulated DA neuron function is suggested to associate with the suppression of GABA inhibitory input to DA neurons in the VTA[30] and an increase in glutamate-mediated excitatory inputs[31]. The direct association between DA neuron firing activities and DA release has been well documented. In agreement with previous observation [15-17], The traditional parameters of the VTA DA neuron activities such as firing rate and bursting were significantly increased in response to acute morphine in naïve rats. In addition, by employing a spectra analysis and cell-per-track technique, we found that the firing pattern and firing population of DA neurons were also changed by acute morphine administration. The present study also provided the first evidence that morphine increases SO of the VTA DA neurons. SO, a recently identified major pattern of DA neuron activity [18], was significantly enhanced upon the acute morphine administration. It is believed that the firing pattern is more important than the firing rate in the regulation of DA release, and oscillatory firing results in oscillatory release of DA from DA neurons[18]. It is thus conceivable that the alteration of SO in VTA DA neuron by acute morphine administration may play an important role in the development of abnormal DA neurotransmission associated with prolonged single morphine-mediated change.

Repeated morphine treatment results in a diminished response of the VTA DA neurons to subsequent morphine challenge over extended period of time [24], which is believed to associate with the development of repeated morphine-induced dependence and tolerance. Likewise, rapid tolerance and dependence induced by a single morphine exposure have also been noticed both in clinical observation and experimental studies[11,14,32]. The underlying mechanisms, however, remain largely unknown. The striking change in the present study is the demonstration of prolonged failure in response to morphine challenge for the VTA DA neuron activities followed initial single morphine exposure which at least sustained for 3 days. The electrophysiological parameters recovered until day 5 followed the single morphine treatment. This failure of VTA DA neurons in response to subsequent morphine challenge after single morphine injection may provide an underlying mechanism for the development of tolerance and dependence followed single morphine treatment at cellular level. However, it is also interested to note that it was reported that in escalation of cocaine self-administration is independent of dopamine levels of nucleus accumbens[33], this indicates that there are mechanistic differences in the development of dependence/tolerance and escalation between morphine vs cocaine [34].

In addition to the prolonged changes in VTA DA neuron activity, at receptor levels, we detected an impairment of

opiate receptor-G-protein coupling in rat brains with single morphine administration, indicating a desensitization of opioid receptor was also developed in response to single morphine injection (Fig. 7). Desensitization of the opioid receptors followed the repeated morphine or opioid receptor agonist has been repeatedly demonstrated and is linked to the development of tolerance and dependence [24,35-38]. Likewise, the present data provided the first data that opioid receptor also develops a prolonged desensitization with single morphine exposure. It is well known that phosphorylation of opioid receptors is critical for receptor desensitization, the desensitized receptor then internalized via endocytosis, where the receptor either recycles back to membrane or degradation. Altered trafficking of opioid receptor is believed to play an essential role in repeated morphine-elicited tolerance and dependence [37,38]. It appears that a prolonged desensitization of the opioid receptors may underlie the single dose morphine-induced tolerance. It would be interested to study the detail mechanism for how the desensitization alters the receptor recycling which, in turn, contributes to the development of dependence and tolerance followed single dose of morphine.

In summary, the present study demonstrated a prolonged failure in both VTA DA neuron activity and opioid receptors to subsequent morphine challenge followed a single morphine treatment. The further work should focus on the detail molecular events that underlie this change.

Conclusion

Prolonged VTA DA neuron activation and opiate receptors desensitization followed single morphine exposure may underlie the development of dependence and tolerance which may associate with the alteration of analgesic response and acute addiction of morphine.

Competing interests

The authors declare that they have no competing interests.

Authors' contributions

DZ performed the electrophysiological experiments, HZ conducted biochemical assays, GZJ participated experimental design, KHZ and X CZ instructed the project and wrote the manuscript. All authors read and approved the manuscript.

Acknowledgements

This work was supported by a 863 and 973-plan of the Ministry of Science and Technology of China (2007AA02z163, CB522000), and grants from Natural Science Foundation of China (30825042 and 30770662).

References

1. Carr DB, Sesack SR: **Projections from the Rat Prefrontal Cortex to the Ventral Tegmental Area: Target Specificity in the Synaptic Associations with Mesoaccumbens and Mesocortical Neurons.** *J Neurosci* 2000, **20**:3864-3873.

2. Onn SP, West AR, Grace AA: **Dopamine-mediated regulation of striatal neuronal and network interactions.** *Trends Neurosci* 2000, **23**:548-56.
3. O'Donnell P: **Dopamine gating of forebrain neural ensembles.** *Eur J Neurosci* 2003, **17**:429-435.
4. Spanagel R, Weiss F: **The dopamine hypothesis of reward: past and current status.** *Trends Neurosci* 1999, **22**:521-527.
5. Marinelli M, Cooper D, Baker L, White F: **Impulse activity of mid-brain dopamine neurons modulates drug-seeking behavior.** *Psychopharmacol* 2003, **168**:84-98.
6. Di Chiara G: **Drug addiction as dopamine-dependent associative learning disorder.** *Eur J Pharmacol* 1999, **375**:13-30.
7. Ammon-Treiber S, Holtt V: **Morphine-induced changes of gene expression in the brain.** *Addict Biol* 1993, **10**:81-89.
8. Zachariou V, Bolanos CA, Selley DE, Theobald D, Cassidy MP, Kelz MB, Shaw-Lutchman T, Berton O, Sim-Selley LJ, Dileone RJ, Kumar A, Nestler EJ: **An essential role for DeltaFosB in the nucleus accumbens in morphine action.** *Nat Neurosci* 2006, **9**:205-11.
9. Harris GC, Aston-Jones G: **Altered motivation and learning following opiate withdrawal: evidence for prolonged dysregulation of reward processing.** *Neuropsychopharmacol* 2003, **28**:865-871.
10. Melis M, Spiga S, Diana M: **The dopamine hypothesis of drug addiction: hypodopaminergic state.** *Int Rev Neurobiol* 2005, **63**:101-54.
11. Cochin J, Kornetsky C: **Development and Loss of Tolerance to Morphine in the Rat after Single and Multiple Injections.** *J Pharmacol & Exp Ther* 1964, **145**:1-10.
12. McQuay HJ, Bullingham RES, Moore RA: **Acute opiate tolerance in man.** *Life Sci* 1981, **28**:2513-2517.
13. Kim DH, Fields HL, Barbaro NM: **Morphine analgesia and acute physical dependence: rapid onset of two opposing, dose-related processes.** *Brain Res* 1990, **516**:37-40.
14. Kissin I, Brown PT, Robinson CA, Bradley EL Jr: **Acute tolerance in morphine analgesia: continuous infusion and single injection in rats.** *Anesthesiology* 1991, **74**(1):166-171.
15. Gysling K, Wang RY: **Morphine-induced activation of A10 dopamine neurons in the rat.** *Brain Res* 1983, **277**:119-127.
16. Matthews RT, German DC: **Electrophysiological evidence for excitation of rat ventral tegmental area dopamine neurons by morphine.** *Neurosci* 1984, **11**:617-625.
17. Williams JT, Christie MJ, Manzoni O: **Cellular and Synaptic Adaptations Mediating Opioid Dependence.** *Physiol Rev* 2001, **81**:299-343.
18. Shi WX: **Slow oscillatory firing: a major firing pattern of dopamine neurons in the ventral tegmental area.** *J Neurophysiol* 2005, **94**:3516-3522.
19. Zhang D, Yang S, Jin G-Z, Bunney BS, Shi W-X: **Oscillatory Firing of Dopamine Neurons: Differences between Cells in the Substantia Nigra and Ventral Tegmental Area.** *Synapse* 2007, **62**:169-75.
20. Shi WX, Pun CL, Zhou Y: **Psychostimulants induce low-frequency oscillations in the firing activity of dopamine neurons.** *Neuropsychopharmacol* 2004, **29**:2160-2167.
21. Zhang D, Yang S, Yang CH, Jin GZ, Zhen X: **Estrogen regulates responses of dopamine neurons in the ventral tegmental area to cocaine.** *Psychopharmacol* 2008, **199**:625-35.
22. Gao M, Liu CL, Yang S, Jin GZ, Bunney BS, Shi WX: **Functional coupling between the prefrontal cortex and dopamine neurons in the ventral tegmental area.** *J Neurosci* 2007, **27**:5414-5421.
23. Pothos E, Rada P, Mark GP, Hoebel BG: **Dopamine microdialysis in the nucleus accumbens during acute and chronic morphine, naloxone-precipitated withdrawal and clonidine treatment.** *Brain Res* 1991, **566**:348-350.
24. Georges F, Moine CL, Aston-Jones G: **No Effect of Morphine on Ventral Tegmental Dopamine Neurons during Withdrawal.** *J Neurosci* 2006, **26**:5720-5726.
25. Grace AA, Bunney BS: **Intracellular and extracellular electrophysiology of nigral dopaminergic neurons – I. Identification and characterization.** *Neurosci* 1983, **10**:301-315.
26. Wikler A: **Present status of the concept of drug dependence.** *Psychol Med* 1971, **1**:377-80.
27. Acquas E, Carboni E, Di Chiara G: **Profound depression of mesolimbic dopamine release after morphine withdrawal in dependent rats.** *Eur J Pharmacol* 1991, **193**:133-134.
28. Pontieri FE, Tanda G, Di Chiara G: **Intravenous cocaine, morphine, and amphetamine preferentially increase extracellular dopamine in the "shell" as compared with the "core" of the rat nucleus accumbens.** *Proc Natl Acad Sci USA* 1995, **92**:12304-12308.
29. Crespo JA, Stöckl P, Zorn K, Saria A, Zernig G: **Nucleus Accumbens Core Acetylcholine is Preferentially Activated During Acquisition of Drug- vs Food-Reinforced Behavior.** *Neuropsychopharmacology* 2008, **33**(13):3213-3220.
30. Johnson SW, North RA: **Opioids excite dopamine neurons by hyperpolarization of local interneurons.** *J Neurosci* 1992, **12**:483-488.
31. Saal D, Dong Y, Bonci A, Malenka RC: **Drugs of Abuse and Stress Trigger a Common Synaptic Adaptation in Dopamine Neurons.** *Neuron* 2003, **37**:577-582.
32. Horvath G, Kekesi G, Dobos I, Klimscha W, Benedek G: **Long-term changes in the antinociceptive potency of morphine or dexmedetomidine after a single treatment.** *Anesth Analg* 2005, **101**:812-8.
33. Ahmed SH, Lin D, Koob GF, Parsons LH: **Escalation of cocaine self-administration does not depend on altered cocaine-induced nucleus accumbens dopamine levels.** *J Neurochem* 2003, **86**:102-13.
34. Zernig G, Ahmed SH, Cardinal RN, Morgan D, Acquas E, Foltin RW, Vezina P, Negus SS, Crespo JA, Stöckl P, Grubinger P, Madlung E, Harling C, Kurz M, Saria A: **Explaining the escalation of drug use in substance dependence: models and appropriate animal laboratory tests.** *Pharmacol* 2007, **80**:65-119.
35. Sim LJ, Selley DE, Dworkin SI, Childers SR: **Effects of chronic morphine administration on mu opioid receptor-stimulated [35S]GTPgammaS autoradiography in rat brain.** *J Neurosci* 1998, **16**:2684-2692.
36. Chakrabarti S, Regec A, Gintzler AR: **Biochemical demonstration of mu-opioid receptor association with Galpha: enhancement following morphine exposure.** *Brain Res- Mol Brain Res* 2005, **135**:217-224.
37. Martini L, Whistler JL: **The role of mu opioid receptor desensitization and endocytosis in morphine tolerance and dependence.** *Curr Opin Neurobiol* 2007, **17**:556-64.
38. Koch T, Höllt V: **Role of receptor internalization in opioid tolerance and dependence.** *Pharmacol Ther* 2008, **117**:199-206.

Publish with **BioMed Central** and every scientist can read your work free of charge

"BioMed Central will be the most significant development for disseminating the results of biomedical research in our lifetime."

Sir Paul Nurse, Cancer Research UK

Your research papers will be:

- available free of charge to the entire biomedical community
- peer reviewed and published immediately upon acceptance
- cited in PubMed and archived on PubMed Central
- yours — you keep the copyright

Submit your manuscript here:
http://www.biomedcentral.com/info/publishing_adv.asp

