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## Long-term potentiation at C-fibre synapses by low-level presynaptic activity *in vivo*

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### Abstract

Inflammation, trauma or nerve injury trigger low-level activity in C-fibres and may cause long-lasting hyperalgesia. Long-term potentiation (LTP) at synapses of primary afferent C-fibres is considered to underlie some forms of hyperalgesia. In previous studies, high- but not low-frequency conditioning stimulation of C-fibres has, however, been used to induce LTP in pain pathways. Recently we could show that also conditioning low-frequency stimulation (LFS) at C-fibre intensity induces LTP *in vitro* as well as in the intact animal, i.e. with tonic descending inhibition fully active. In the slice preparation, this form of LTP requires a rise in postsynaptic  $\text{Ca}^{2+}$ -concentration and activation of  $\text{Ca}^{2+}$ -dependent signalling pathways. Here, we investigated the signalling mechanisms underlying this novel form of LTP *in vivo*. We found that the signal transduction pathways causing LFS-induced LTP *in vivo* include activation of neurokinin I and N-methyl-D-aspartate receptors, rise of  $[\text{Ca}^{2+}]_i$  from intracellular stores and via T-type voltage-dependent  $\text{Ca}^{2+}$  channels, activation of phospholipase C, protein kinase C and  $\text{Ca}^{2+}$ -calmodulin dependent kinase II. These pathways match those leading to hyperalgesia in behaving animals and humans. We thus propose that LTP induced by low-level activity in C-fibres may underlie some forms of hyperalgesia.

### Background

LTP at the first synapse in pain pathways is considered to underlie some forms of pain amplification e.g. after trauma, inflammation or nerve injury [1]. A strong rise in postsynaptic calcium ion concentration triggering  $\text{Ca}^{2+}$ -dependent signal transduction pathways is required for LTP induction [2-4]. Consequently, high-frequency ( $\sim 100$  Hz), burst-like stimulation protocols were previously used to induce LTP at virtually all synapses studied so far.

Low-level activity between  $1-10 \text{ imp} \cdot \text{s}^{-1}$  rather than high frequency bursts are, however, typical for C-fibre discharges during inflammation, trauma or wound healing. Presynaptic activity at these low frequencies is considered

inadequate to cause a sufficiently strong rise in postsynaptic  $[\text{Ca}^{2+}]_i$  for potentiation of synaptic strength. In fact, low-level presynaptic activity was either ineffective or induced synaptic long-term depression (LTD) rather than LTP in previous studies.

We have recently discovered that in a spinal cord slice preparation with long dorsal roots intact LFS of dorsal roots at C-fibre intensity induces LTP which involves a rise in postsynaptic  $[\text{Ca}^{2+}]_i$  and  $\text{Ca}^{2+}$ -dependent signal transduction pathways [4]. In the intact animal spinal dorsal neurons are, however, under a powerful tonic inhibition arising from supraspinal, descending pathways [5,6]. This inhibition is inevitably lost in the *in vitro* situation and

could thereby facilitate LTP-induction. And indeed, removal of descending, putatively inhibitory pathways by spinalisation is required for the induction of LTP by either pinching or noxious heating of the skin [7]. On the other hand, we have shown recently that in the intact animal, LTP can be induced by LFS as well as by subcutaneous capsaicin or formalin injections also [4]. Here, we further characterised this novel LFS-induced LTP at C-fibre synapses in the intact animal.

## Methods

### Preparation of the animals

After obtaining approval from the Institutional Animal Care Comitee (Austrian Federal Ministry for Education, Science and Culture), experiments were performed on adult male Sprague Dawley rats (200 – 300 g) obtained from a local breeding facility.

Animals were intubated using a 16 G cannula and mechanically ventilated (Siemens Servoventilator 900C). Isoflurane in two thirds N<sub>2</sub>O and one third O<sub>2</sub> was used to induce (4 vol% inspiratory) and maintain (1.5 vol% expiratory) anaesthesia. Surgical level of anaesthesia was verified by stable arterial blood pressure and the absence of paw withdrawal reflexes after pinching the paw. Surgical procedures have been described in more detail elsewhere [4]. Briefly, the right femoral vein and artery were cannulated to allow i.v. infusions and to monitor arterial blood pressure. Muscle relaxation was achieved by 2 µg·kg<sup>-1</sup>·h<sup>-1</sup> pancuronium bromide i.v. The left sciatic nerve was dissected free for bipolar electrical stimulation using a silver hook electrode. Lumbar segments L4 and L5 were exposed by laminectomy. The dura mater was incised and reflected. Two metal clamps were used for fixation of the vertebral column in a stereotactic frame. An agarose pool was formed around the exposed spinal segments. The spinal segments were covered with artificial cerebrospinal fluid which was carefully removed before application of drugs (see below). The exposed sciatic nerve was covered with warm paraffin oil. At the end of the experiment animals were decapitated in deep anaesthesia and the spinal cord removed to locate the recording site (see next paragraph).

### Electrophysiological recordings

Electrophysiological recordings were performed as described elsewhere [4]. Briefly, C-fibre-evoked field potentials were recorded with glass electrodes (2–3 MΩ) in laminae I and II of the spinal cord dorsal horn in response to stimulation of the ipsilateral sciatic nerve. Electrodes were driven by a microstepping motor (Narishige Scientific Instrument, Japan). Recordings were made with an ISO-DAM-amplifier (World Precision Instruments Inc., Sarasota, FL, USA) using a band width filter of 0.1 – 1000 Hz. Signals were monitored on a dig-

ital oscilloscope (Tektronix TDS 210) and digitized at a sampling rate of 5 kHz by an A/D converter (DigiData 1200 Series Interface). Field potentials may be evoked by A- or C-fibres. The different components can easily be distinguished by their thresholds and latencies. C-fibre evoked field potentials have high thresholds (> 5–6 V) and long latencies (90 – 150 ms, corresponding to the conduction velocity of < 1.2 m·s<sup>-1</sup>). The potentials were not abolished by spinalisation or muscle relaxation, strongly suggesting that these signals are evoked by activation of primary afferent C-fibres. This is in line with a report by Schouenborg [8], suggesting that field potentials are generated monosynaptically by synapses between primary afferent fibres and second order neurons. A polysynaptic contribution to the signals can, however, not be fully excluded. As test stimuli, single pulses (0.5 ms, 25 V, which is suprathreshold for the activation of C-fibres) were delivered to the sciatic nerve every 5 minutes using an electrical stimulator (Iso-Stim 01D, npi electronic, Germany). Stable responses for 1 hour served as control. LTP of C-fibre evoked field potentials was induced by low frequency stimulation (LFS, 2 Hz, 0.1 ms pulses, 60 V, 2 min). After conditioning stimulation, test pulses were applied again to the sciatic nerve at 5 min intervals for up to 7 hours.

Rhodamine B (0.2%, Sigma) was added to the pipette solution which consisted of (in mM) NaCl 135, KCl 5.4, CaCl<sub>2</sub> 1.8, Hepes 10 and MgCl<sub>2</sub> 1. For identification of the recording position, pressure was applied to the electrode (300 mbar, 1 min) at the end of each electrophysiological experiment. The spinal cord was removed, cryo-fixed and the rhodamine B spot was localised under a fluorescence microscope. Only those experiments where the tip of the recording electrode was located in lamina I or II were included into the study.

### Drugs

Pancuronium bromide (Pancuronium-ratiopharm®, Ratiopharm GmbH, Ulm, Germany) was given as an i.v. infusion (2 µg·kg<sup>-1</sup>·h<sup>-1</sup>). The competitive N-methyl-D-aspartate (NMDA) receptor antagonist D(-)-2-Amino-5-phosphonopentanoic acid (D-AP5, 100 µM, Sigma) and the T-type voltage-dependent Ca<sup>2+</sup> channel (VDCC) blocker mibefradil dihydrochloride hydrate (5 mM, Sigma) were dissolved in 0.9% NaCl. The protein kinase C (PKC) blocker chelerythrine chloride (800 µM, Sigma) was dissolved in ddH<sub>2</sub>O. The Ca<sup>2+</sup>-calmodulin dependent kinase II (CaMKII) blocker 2-[N-(2-hydroxyethyl)]-N-(4-methoxybenzenesulfonyl)]amino-N-(4-chlorocinnamyl)-N-methylbenzylamine (KN-93, 400 µM, Sigma), the phospholipase C (PLC) blocker 1-[6-(((17β)-3-Methoxyestra-1,3,5[10]-trien-17-yl)amino)hexyl]-1H-pyrrole-2,5-dione (U-73122, 500 µM, Sigma), the neurokinin receptor 1 (NK1 receptor) blocker cis-2-(Diphenylme-

thyl)-N- [(2-iodophenyl)methyl]-1azabicyclo [2.2.2]octan-3-amine oxalate salt (L-703,606, 5 mM, Sigma) and the ryanodine receptor blocker dantrolene (500  $\mu$ M, Tocris) were dissolved in DMSO. Dantrolene was further dissolved in standard solution titrated to a pH of 9.5. All other drugs were further dissolved in artificial cerebrospinal fluid to obtain desired concentrations as indicated and applied directly onto the spinal cord [9], starting 30 min prior LFS.

#### Data analysis and statistics

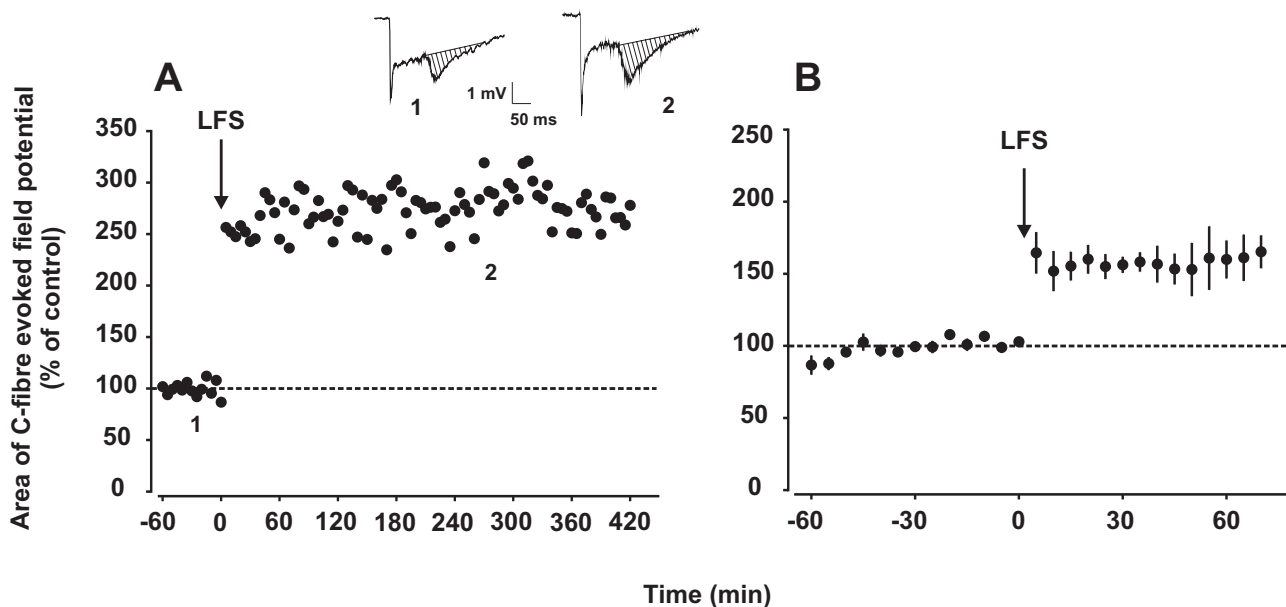
The area under the curve of C-fibre evoked field potentials was determined offline using the software Clampfit 9.0 (Molecular Devices Inc.). For the electrophysiological experiments, responses were normalised for each rat. A one way repeated measures analysis of variance (One way RM-ANOVA) was performed to compare the different experimental protocols and treatments. ANOVA was corrected by the Bonferroni adjustment. A p-value < 0.05 was considered statistically significant. Values are expressed as mean  $\pm$  standard error of mean (SEM).

#### Results and discussion

Intact, adult rats were used in the present study to evaluate LTP induction *in vivo* by low-level activity in C-fibres. Elec-

trical LFS (2 Hz, 2 min, C-fibre strength) delivered to the sciatic nerve induced stable LTP of C-fibre evoked field potentials always lasting until the end of the recording period of up to 7 hours (Fig. 1A). LFS-induced mean LTP of C-fibre evoked field potentials to  $160\% \pm 13\%$  of control ( $p < 0.05$ ,  $n = 5$ ) at 60 min after conditioning stimulation (Fig 1B; Table 1). A-fibre evoked field potentials were never affected (data not shown). In some subsequent experiments DMSO was used as a solvent for drugs applied directly onto the spinal cord surface. Superfusion of the spinal cord at the recording segment with this solvent had no effect on LTP induction in any of the 3 animals tested (Table 1).

We have shown previously that *in vitro* LFS-induced LTP requires a rise in  $[Ca^{2+}]_i$  [4]. This may be achieved by  $Ca^{2+}$ -influx from the extracellular space or by release from intracellular compartments [10]. During LFS, voltage-dependent  $Ca^{2+}$  channels (VDCCs) which open already by moderate depolarization, may be activated. To test if low-voltage activated T-type VDCCs play a role, we applied the blocker mibefradil (5 mM) directly onto the spinal cord 30 min prior to LFS. Topical mibefradil completely prevented LTP induction (Fig. 2A; Table 1), suggesting that LFS of sciatic C-fibres is sufficient to activate T-type



**Figure 1**

**LTP in the spinal dorsal horn is induced by LFS of C-fibres in the sciatic nerve.** Electrical LFS at C-fibre strength delivered to the sciatic nerve induced LTP of C-fibre evoked field potentials in laminae I/II of the spinal cord dorsal horn, lasting undiminished for several hours. A representative example of LFS-induced LTP and two original recordings prior to (1) or after LTP-induction (2) are shown in (A). Areas of C-fibre evoked field potentials were used to quantify synaptic strength (dashed areas in 1 and 2). Mean time course of five experiments is shown in (B). In both graphs, area of C-fibre evoked field potentials (% of control) is plotted against time (min). The arrows represent the time point of conditioning LFS.

**Table 1: Summary of results**

Superfusate	Concentration	C-fibre response (% control)	n
ACSF	-	160% ± 13%*	5
DMSO	25%	150% ± 14%*	3
Mibefradil	5 mM	108% ± 14% n.s.	5
L-703,606	5 mM	88% ± 15%*	5
Dantrolene	500 µM	103% ± 14% n.s.	5
U-73122	500 µM	107% ± 10% n.s.	5
Chelerythrine	800 µM	103% ± 7% n.s.	5
D-AP5	100 µM	107% ± 8% n.s.	5
KN-93	400 µM	99% ± 5% n.s.	5

Mean (± SEM) C-fibre evoked field potentials are expressed as % of control 60 min after LFS. One-way repeated measure-ANOVA was used to compare areas of C-fibre evoked field potentials before and after LFS. The asterisk denotes significant differences from baseline (\* =  $p < 0.05$ ). Drugs in spinal superfusate and their respective concentrations are indicated.

VDCCs in spinal cord, and that this contributes to LTP induction. Our finding is in line with recent studies showing that spinal T-type VDCCs play a role both for normal nociception, and for thermal and mechanical hyperalgesia in models of neuropathic pain [11,12].

Upon stimulation substance P is released from C-fibre afferents, leading to activation of neurokinin-1 (NK1) receptors, mainly in laminae I/II neurons of the spinal dorsal horn [13]. Most NK1-expressing lamina I neurons have a supraspinal projection [14]. Blockade of spinal NK1 receptors attenuates thermal and mechanical hyperalgesia [15-17]. In the present study, blocking spinal NK1 receptors with L-703,606 (5 mM) not only fully blocked LTP induction but rather led to LFS-induced LTD of C-fibre evoked field potentials (Fig 2B; Table 1).

The thresholds for the induction of LTP and LTD have been suggested to be tuned narrowly. Changes in the level of elevation of  $[Ca^{2+}]_i$  might shift the threshold from LTP towards LTD [18]. The polarity of synaptic plasticity further depends upon the magnitude [19], the temporal pattern [20] and the mode of postsynaptic  $Ca^{2+}$  elevation [21]. These parameters are all relevant for activation of distinct  $Ca^{2+}$ -dependent signal transduction pathways involving protein phosphatases and kinases. Interestingly, blockade of NK1 receptors but not of VDCCs or NMDA receptors (see below) might critically change at least one of these characteristics of the rise in  $[Ca^{2+}]_i$ , shifting synaptic plasticity from LTP towards LTD. This illustrates that blocking different routes of  $[Ca^{2+}]_i$  rise may have differential effects on synaptic plasticity.

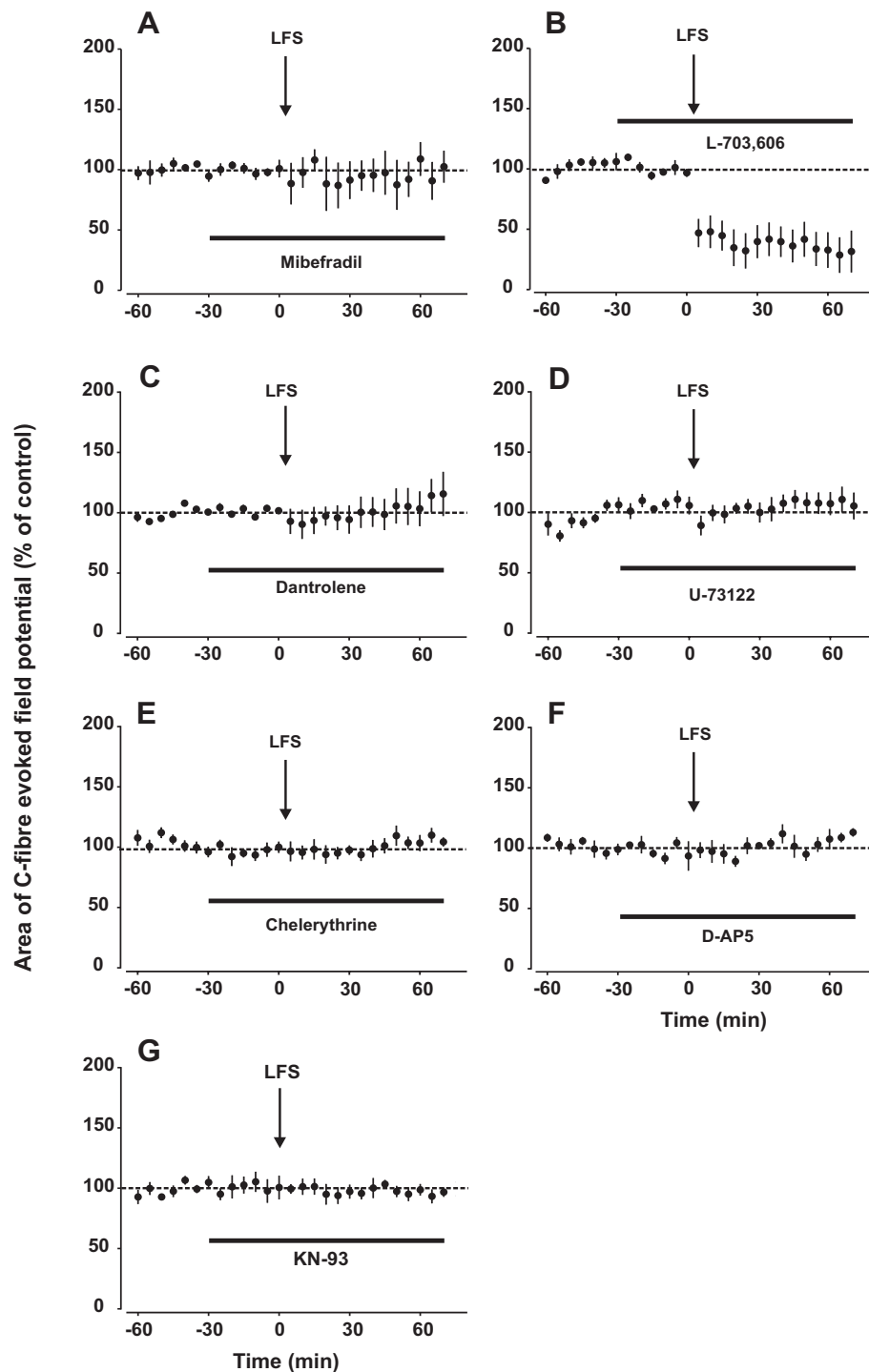
In superficial spinal dorsal horn neurons  $[Ca^{2+}]_i$  may be increased by the release of  $Ca^{2+}$  from intracellular stores [10] via activation of either ryanodine or inositol 1,4,5-trisphosphate ( $IP_3$ ) receptors. Ryanodine receptors may be

activated by  $Ca^{2+}$  which may, for example, enter the cell through VDCCs, leading to  $Ca^{2+}$ -induced  $Ca^{2+}$  release from ryanodine sensitive stores [10]. In the present study, spinal ryanodine receptors were blocked by dantrolene (500 µM), a clinically used ryanodine receptor antagonist. This abolished LTP induction in all animals tested (Fig 2C; Table 1). Intrathecal injection of dantrolene has previously been shown to reduce nociceptive behaviour caused by spinal application of a NK1 receptor agonist [22].

Another trigger for  $Ca^{2+}$  release from intracellular stores is the activation of  $IP_3$  receptors.  $IP_3$  receptors are activated by  $IP_3$ , a product formed by hydrolysis of phosphatidylinositol 4,5 bisphosphate upon activation of the phosphoinositide system. It starts with activation of phospholipase C (PLC) by the alpha subunit of a  $G_q$ -protein [23]. Activation of PLC in the spinal cord is essential for hyperalgesia. For example, mechanical hyperalgesia induced by intraplantar injection of endothelin 1, a peptide produced by many different cell types, requires activation of spinal PLC [24]. Furthermore, intrathecal pre-treatment with a PLC inhibitor reduced the second-phase in the formalin test [25]. In the present study, inhibition of PLC in spinal cord by the selective blocker U-73122 (500 µM) completely prevented induction of LTP by LFS (Fig. 2D; Table 1).

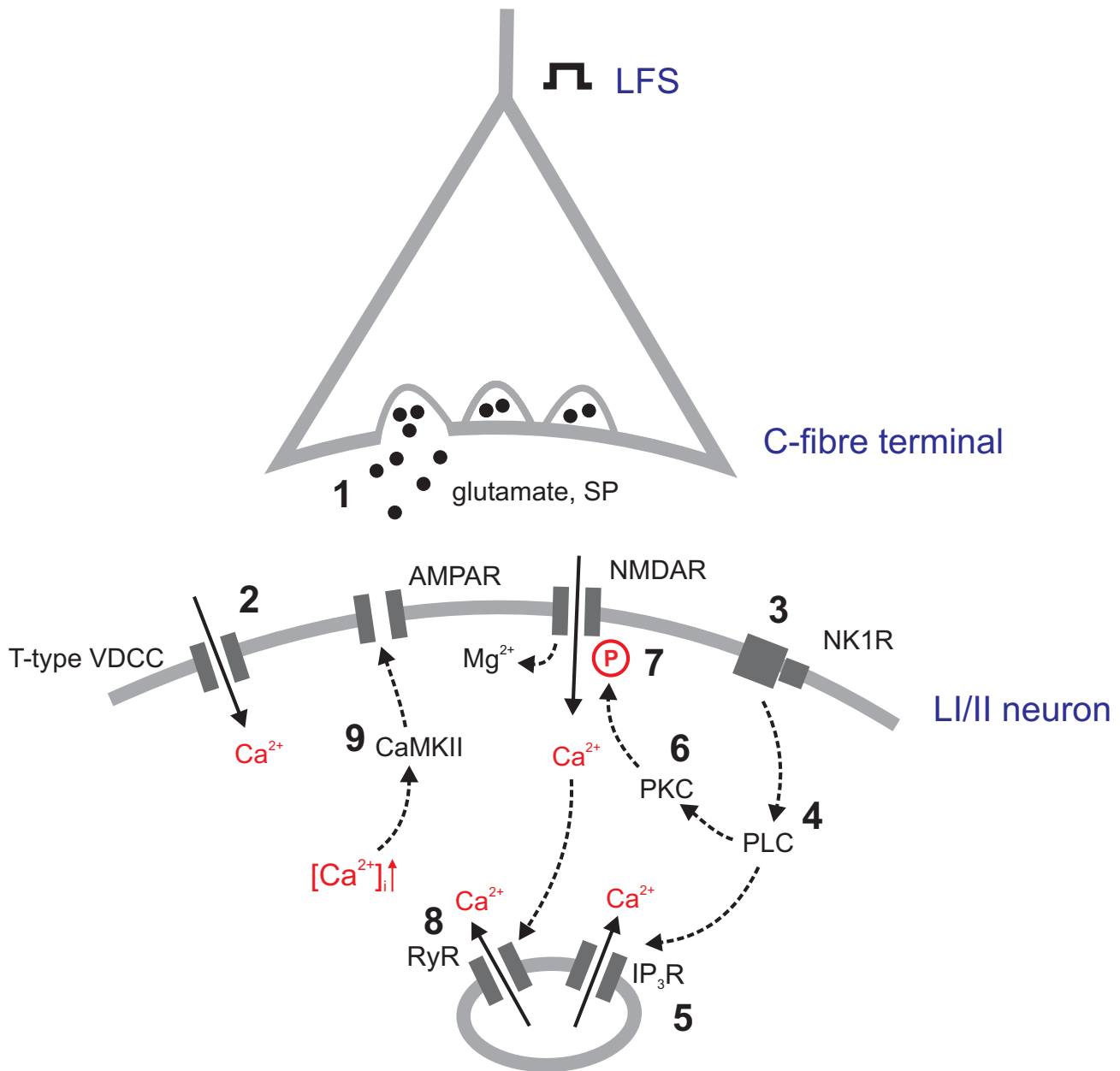
PLC activation also leads to the production diacylglycerol (DAG) which in turn activates protein kinase C (PKC). At least 12 isoforms of this enzyme can be distinguished, many of those are present in superficial laminae of the spinal cord dorsal horn [26]. PKC plays a crucial role in the development of hyperalgesia. Activation of spinal PKC by phorbol esters evokes thermal hyperalgesia and mechanical allodynia in awake rats [27]. An involvement of PKC in inflammatory pain has also been shown. For example, PKC inhibitors attenuate hyperalgesia after intradermal injection of capsaicin or formalin [28,29]. PKC may phosphorylate the NMDA receptor at distinct phosphorylation sites, thereby increasing channel activity [30,31] or removing the  $Mg^{2+}$  block [32]. Here, blockade of spinal PKC by chelerythrine (800 µM) abolished induction of LTP by LFS in all animals tested (Fig. 2E; Table 1).

Activation of  $Ca^{2+}$ -permeable NMDA receptors in spinal dorsal horn is an essential step for the induction of several forms of hyperalgesia and allodynia [33-35]. However, NMDA receptors are normally blocked near the resting membrane potential by  $Mg^{2+}$ . One might speculate that LFS is not sufficient to activate NMDA receptors, unless the  $Mg^{2+}$  block is removed e.g. by phosphorylation via PKC. And indeed, LFS-induced LTP required activation of NMDA receptors, as blockade of spinal NMDA receptors with the competitive NMDA receptor antagonist D-AP5 (100 µM) prevented LTP induction (Fig 2F; Table 1). This



**Figure 2**

**Signalling pathways underlying the induction of LFS-induced LTP *in vivo*.** In all graphs, area of C-fibre evoked field potentials (% of control) is plotted against time (min). Data are expressed as mean  $\pm$  SEM ( $n = 5$  for all groups). Spinal superfusion at the recording segment with the T-type VDCC blocker mibefradil (5 mM, A), the ryanodine receptor blocker dantrolene (500  $\mu$ M, C), the PLC inhibitor U-73122 (500  $\mu$ M, D), the PKC blocker chelerythrine (800  $\mu$ M, E), the NMDA receptor blocker D-AP5 (100  $\mu$ M, F) or the CaMKII blocker KN-93 (400  $\mu$ M, G) fully blocked LTP induction by LFS. Blockade of spinal NK1 receptors with L-703,606 led to induction of LTD rather than LTP by LFS (B). See also Table 1.

**Figure 3**

**Potential mechanisms of LTP in spinal dorsal horn *in vivo*.** At synapses between C-fibres and laminae I/II second order neurons, LFS triggers the release of the excitatory neurotransmitters glutamate and substance P leading to depolarization of the postsynaptic cell (1). T-type VDCCs are activated (2), leading to  $\text{Ca}^{2+}$  influx. The activation of NK1 receptors (3) activates the PLC-pathway (4), triggering the release of  $\text{Ca}^{2+}$  from intracellular stores (5) and the activation of PKC (6). This kinase phosphorylates NMDA receptors, thereby loosening their  $\text{Mg}^{2+}$  block (7).  $\text{Ca}^{2+}$ -induced  $\text{Ca}^{2+}$  release activates ryanodine receptors, thereby further increasing  $[\text{Ca}^{2+}]_i$  (8). The increased  $[\text{Ca}^{2+}]_i$  is sensed by CaMKII that phosphorylates AMPA receptors, thereby potentiating synaptic strength. LFS: low frequency stimulation; SP: substance P; VDCC: voltage-dependent  $\text{Ca}^{2+}$  channel; AMPAR: alpha-amino-3-hydroxy-5-methyl-4-isoxazolepropionic acid receptor; NMDAR: N-methyl-D-aspartate receptor; NK1R: neurokinin receptor 1; PLC: phospholipase C; PKC: protein kinase C;  $\text{IP}_3\text{R}$ : inositol 1,4,5-trisphosphate receptor; RyR: ryanodine receptor; CaMKII:  $\text{Ca}^{2+}$ -calmodulin dependent kinase II;

is similar to findings of previous studies showing that NMDA receptor activation is also required for high frequency stimulation (HFS)-induced LTP in the spinal cord dorsal horn [36-38,3].

A rise in  $[Ca^{2+}]_i$  may lead to  $Ca^{2+}$  binding to calmodulin, which in turn activates  $Ca^{2+}$ -calmodulin dependent kinase II (CaMKII). CaMKII undergoes a  $Ca^{2+}$ -dependent autophosphorylation, resulting in  $Ca^{2+}$ -independent activity [39]. One of its main targets is the GluR1 subunit of the AMPA receptor, which gets phosphorylated at Ser831 [40]. This leads to an increased channel conductance and thus synaptic strength [41]. Activation of spinal CaMKII is required for some forms of hyperalgesia [42-44]. For example, intrathecal pre-treatment with a CaMKII inhibitor prevents thermal hyperalgesia and mechanical allodynia after injection of complete Freund's adjuvant into a hindpaw [45]. Inhibition of CaMKII blocks induction of HFS-induced LTP in the spinal cord dorsal horn [46].

Here, the CaMKII inhibitor KN-93 (400  $\mu$ M) abolished induction of LFS-induced LTP in all animals tested (Fig 2G; Table 1).

## Conclusion

In the present study, we investigated signal transduction mechanisms underlying LFS-induced LTP in the spinal cord *in vivo*. We could show that pathways required for LTP induction match those involved in hyperalgesia. This suggests that LFS-induced LTP at spinal synapses of C-fibres may be a mechanism of pain amplification in behaving animals and perhaps human subjects [47].

Conditioning electrical stimulation of C-fibres at low frequencies induces LTP at the first nociceptive synapses in the spinal cord dorsal horn in slice preparations as well as in intact animals [[4], present study]. The signal transduction pathways underlying this novel form of LTP have been previously investigated *in vitro* only [4]. Here we demonstrate that pathways underlying the induction of LFS-induced LTP *in vivo* fully match those in the *in vitro* slice preparation suggesting that the same cellular elements are involved despite the very different experimental conditions. The absence or presence of descending modulatory pathways or general anaesthesia does apparently not affect the principle mechanisms of LFS-induced synaptic plasticity in superficial spinal dorsal horn.

Based on the results of the present study, the following sequence of events leading to the induction of LTP at synapses of primary afferent C-fibres can be suggested (Fig. 3): LFS of the sciatic nerve triggers the release of excitatory neurotransmitters including glutamate and substance P (1.), which leads to a moderate depolarization of postsynaptic neurons.

This is sufficient to activate low-voltage activated T-type  $Ca^{2+}$  channels and  $Ca^{2+}$  influx into the cell (2.). In addition, NK1 receptors are activated (3.). Downstream to this receptor activation, the PLC-pathway (4.) leads to the release of  $Ca^{2+}$  from intracellular stores (5.) as well as to activation of PKC (6.). This kinase phosphorylates the NMDA receptor (7.), thereby reducing its  $Mg^{2+}$  block. By this, moderate postsynaptic depolarization may become sufficient to open NMDA receptor channels, leading to an additional influx of  $Ca^{2+}$ .  $[Ca^{2+}]_i$  rise is further enhanced by  $Ca^{2+}$ -induced  $Ca^{2+}$  release from ryanodine sensitive stores (8.). CaMKII is activated by an increased  $[Ca^{2+}]_i$  and phosphorylates AMPA receptors, thereby increasing their channel conductance. Its activity is also necessary for translocation of AMPA receptors to the subsynaptic membrane [48]. This ultimately results in an increased strength of excitatory synapses in nociceptive pathways (9.) and amplification of nociception and perhaps pain perception.

## Competing interests

The authors declare that they have no competing interests.

## Authors' contributions

RD and JS participated equally in the conception, design and interpretation of the study, RD performed all experiments and data analysis and drafted the manuscript. Both authors read and approved the final manuscript.

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## References

1. Sandkühler J: **Understanding LTP in pain pathways.** *Mol Pain* 2007, **3**:9-9.
2. Malenka RC, Kauer JA, Zucker RS, Nicoll RA: **Postsynaptic calcium is sufficient for potentiation of hippocampal synaptic transmission.** *Science* 1988, **242**:81-84.
3. Ikeda H, Heinke B, Ruscheweyh R, Sandkühler J: **Synaptic plasticity in spinal lamina I projection neurons that mediate hyperalgesia.** *Science* 2003, **299**:1237-1240.
4. Ikeda H, Stark J, Fischer H, Wagner M, Drdla R, Jäger T, Sandkühler J: **Synaptic amplifier of inflammatory pain in the spinal dorsal horn.** *Science* 2006, **312**:1659-1662.
5. Basbaum AI, Fields HL: **Endogenous pain control mechanisms: review and hypothesis.** *Ann Neurol* 1978, **4**:451-462.
6. Sandkühler J: **The organization and function of endogenous antinociceptive systems.** *Prog Neurobiol* 1996, **50**:49-81.
7. Sandkühler J, Liu X: **Induction of long-term potentiation at spinal synapses by noxious stimulation or nerve injury.** *Eur J Neurosci* 1998, **10**:2476-2480.
8. Schouenborg J: **Functional and topographical properties of field potentials evoked in rat dorsal horn by cutaneous C-fibre stimulation.** *J Physiol* 1984, **356**:169-192.
9. Beck H, Schröck H, Sandkühler J: **Controlled superfusion of the rat spinal cord for studying non-synaptic transmission: an autoradiographic analysis.** *J Neurosci Methods* 1995, **58**:193-202.
10. Berridge MJ: **Neuronal calcium signaling.** *Neuron* 1998, **21**:13-26.
11. Todorovic SM, Meyenburg A, Jevtovic-Todorovic V: **Mechanical and thermal antinociception in rats following systemic administration of mibefradil, a T-type calcium channel blocker.** *Brain Res* 2002, **951**:336-340.

12. Dogrul A, Gardell LR, Ossipov MH, Tulunay FC, Lai J, Porreca F: **Reversal of experimental neuropathic pain by T-type calcium channel blockers.** *Pain* 2003, **105**:159-168.
13. Go VL, Yaksh TL: **Release of substance P from the cat spinal cord.** *J Physiol* 1987, **391**:141-167.
14. Spike RC, Puskár Z, Andrew D, Todd AJ: **A quantitative and morphological study of projection neurons in lamina I of the rat lumbar spinal cord.** *Eur J Neurosci* 2003, **18**:2433-2448.
15. Traub RJ: **The spinal contribution of substance P to the generation and maintenance of inflammatory hyperalgesia in the rat.** *Pain* 1996, **67**:151-161.
16. Sakurada C, Sakurada S, Katsuyama S, Sasaki J, Tan-No K, Sakurada T: **Involvement of tachykinin NK1 receptors in nociceptin-induced hyperalgesia in mice.** *Brain Res* 1999, **841**:85-92.
17. King T, Gardell LR, Wang R, Vardanyan A, Ossipov MH, Philip Malan T Jr., Vanderah TW, Hunt SP, Hruby VJ, Lai J, Porreca F: **Role of NK-1 neurotransmission in opioid-induced hyperalgesia.** *Pain* 2005, **116**:276-288.
18. Lisman JE: **Three Ca<sup>2+</sup> levels affect plasticity differently: the LTP zone, the LTD zone and no man's land.** *J Physiol* 2001, **532**:285-285.
19. Lisman J: **A mechanism for the Hebb and the anti-Hebb processes underlying learning and memory.** *Proc Natl Acad Sci USA* 1989, **86**:9574-9578.
20. Bi GQ, Poo MM: **Synaptic modifications in cultured hippocampal neurons: dependence on spike timing, synaptic strength, and postsynaptic cell type.** *J Neurosci* 1998, **18**:10464-10472.
21. Nishiyama M, Hong K, Mikoshiba K, Poo MM, Kato K: **Calcium stores regulate the polarity and input specificity of synaptic modification.** *Nature* 2000, **408**:584-588.
22. Álvarez-Vega M, Baamonde A, Hidalgo A, Menéndez L: **Effects of the calcium release inhibitor dantrolene and the Ca<sup>2+</sup>-ATPase inhibitor thapsigargin on spinal nociception in rats.** *Pharmacology* 2001, **62**:145-150.
23. Delmas P, Crest M, Brown DA: **Functional organization of PLC signaling microdomains in neurons.** *Trends Neurosci* 2004, **27**:41-47.
24. Motta EM, Calixto JB, Rae GA: **Mechanical hyperalgesia induced by endothelin-1 in rats is mediated via phospholipase C, protein kinase C, and MAP kinases.** *Exp Biol Med (Maywood)* 2006, **231**:1141-1145.
- 25.Coderre TJ: **Contribution of protein kinase C to central sensitization and persistent pain following tissue injury.** *Neurosci Lett* 1992, **140**:181-184.
26. Polgár E, Fowler JH, McGill MM, Todd AJ: **The types of neuron which contain protein kinase C gamma in rat spinal cord.** *Brain Res* 1999, **833**:71-80.
27. Palecek J, Paleckova V, Willis WD: **The effect of phorbol esters on spinal cord amino acid concentrations and responsiveness of rats to mechanical and thermal stimuli.** *Pain* 1999, **80**:597-605.
28. Sluka KA, Willis WD: **The effects of G-protein and protein kinase inhibitors on the behavioral responses of rats to intradermal injection of capsaicin.** *Pain* 1997, **71**:165-178.
29. Wajima Z, Hua XY, Yaksh TL: **Inhibition of spinal protein kinase C blocks substance P-mediated hyperalgesia.** *Brain Res* 2000, **877**:314-321.
30. Lan JY, Skeberdis VA, Jover T, Grooms SY, Lin Y, Aranceda RC, Zheng X, Bennett MV, Zukin RS: **Protein kinase C modulates NMDA receptor trafficking and gating.** *Nat Neurosci* 2001, **4**:382-390.
31. Liao GY, Wagner DA, Hsu MH, Leonard JP: **Evidence for direct protein kinase-C mediated modulation of N-methyl-D-aspartate receptor current.** *Mol Pharmacol* 2001, **59**:960-964.
32. Chen L, Huang LY: **Protein kinase C reduces Mg<sup>2+</sup> block of NMDA-receptor channels as a mechanism of modulation.** *Nature* 1992, **356**:521-523.
33. Woolf CJ, Thompson SWN: **The induction and maintenance of central sensitization is dependent on N-methyl-D-aspartic acid receptor activation; implications for the treatment of post-injury pain hypersensitivity states.** *Pain* 1991, **44**:293-299.
34. Ren K, Hylden JLK, Williams GM, Ruda MA, Dubner R: **The effects of a non-competitive NMDA receptor antagonist, MK-801, on behavioral hyperalgesia and dorsal horn neuronal activity in rats with unilateral inflammation.** *Pain* 1992, **50**:331-344.
35. Mao J, Price DD, Hayes RL, Lu J, Mayer DJ: **Differential roles of NMDA and non-NMDA receptor activation in induction and maintenance of thermal hyperalgesia in rats with painful peripheral mononeuropathy.** *Brain Res* 1992, **598**:271-278.
36. Liu XG, Sandkühler J: **Long-term potentiation of C-fiber-evoked potentials in the rat spinal dorsal horn is prevented by spinal N-methyl-D-aspartic acid receptor blockage.** *Neurosci Lett* 1995, **191**:43-46.
37. Svendsen F, Tjølsen A, Hole K: **AMPA and NMDA receptor-dependent spinal LTP after nociceptive tetanic stimulation.** *Neuroreport* 1998, **9**:1185-1190.
38. Liu XG, Sandkühler J: **Activation of spinal N-methyl-D-aspartate or neurokinin receptors induces long-term potentiation of spinal C-fibre-evoked potentials.** *Neuroscience* 1998, **86**:1209-1216.
39. Giese KP, Fedorov NB, Filipkowski RK, Silva AJ: **Autophosphorylation at Thr286 of the calcium-calmodulin kinase II in LTP and learning.** *Science* 1998, **279**:870-873.
40. Barria A, Muller D, Derkach V, Griffith LC, Soderling TR: **Regulatory phosphorylation of AMPA-type glutamate receptors by CaM-KII during long-term potentiation.** *Science* 1997, **276**:2042-2045.
41. Derkach V, Barria A, Soderling TR: **Ca<sup>2+</sup>/calmodulin-kinase II enhances channel conductance of α-amino-3-hydroxy-5-methyl-4-isoxazolepropionate type glutamate receptors.** *Proc Natl Acad Sci USA* 1999, **96**:3269-3274.
42. Fang L, Wu J, Lin Q, Willis WD: **Calcium-calmodulin-dependent protein kinase II contributes to spinal cord central sensitization.** *J Neurosci* 2002, **22**:4196-4204.
43. Fang L, Wu J, Zhang X, Lin Q, Willis WD: **Calcium/calmodulin dependent protein kinase II regulates the phosphorylation of cyclic AMP-responsive element-binding protein of spinal cord in rats following noxious stimulation.** *Neurosci Lett* 2005, **374**:1-4.
44. Larsson M, Broman J: **Pathway-specific bidirectional regulation of Ca<sup>2+</sup>/calmodulin-dependent protein kinase II at spinal nociceptive synapses after acute noxious stimulation.** *J Neurosci* 2006, **26**:4198-4205.
45. Luo F, Yang C, Chen Y, Shukla P, Tang L, Wang LX, Wang ZJ: **Reversal of chronic inflammatory pain by acute inhibition of Ca<sup>2+</sup>/calmodulin-dependent protein kinase II.** *J Pharmacol Exp Ther* 2008, **325**:267-275.
46. Yang HW, Hu XD, Zhang HM, Xin WJ, Li MT, Zhang T, Zhou LJ, Liu XG: **Roles of CaMKII, PKA and PKC in the induction and maintenance of LTP of C-fiber evoked field potentials in rat spinal dorsal horn.** *J Neurophysiol* 2004, **91**:1122-1133.
47. Klein T, Magerl W, Hopf HC, Sandkühler J, Treede RD: **Perceptual correlates of nociceptive long-term potentiation and long-term depression in humans.** *J Neurosci* 2004, **24**:964-971.
48. Boehm J, Malinow R: **AMPA receptor phosphorylation during synaptic plasticity.** *Biochem Soc Trans* 2005, **33**:1354-1356.

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