

REVIEW

Open Access

# MicroRNAs and hepatitis C virus: Toward the end of miR-122 supremacy

Thomas Walter Hoffmann<sup>1,2</sup>, Duverlie Gilles<sup>1,2,3</sup> and Bengrine Abderrahmane<sup>1,3\*</sup>

## Abstract

The most common etiologic agents causing chronic hepatitis are hepatitis C and B viruses (HCV and HBV, respectively). Chronic infection caused by HCV is considered one of the major causative agents of liver cirrhosis and hepatocellular carcinoma worldwide. In combination with the increasing rate of new HCV infections, the lack of a current vaccine and/or an effective treatment for this virus continues to be a major public health challenge. The development of new treatments requires a better understanding of the virus and its interaction with the different components of the host cell. MicroRNAs (miRNAs) are small non-coding RNAs functioning as negative regulators of gene expression and represent an interesting lead to study HCV infection and to identify new therapeutic targets. Until now, microRNA-122 (miR-122) and its implication in HCV infection have been the focus of different published studies and reviews. Here we will review recent advances in the relationship between HCV infection and miRNAs, showing that some of them emerge in publications as challengers against the supremacy of miR-122.

**Keywords:** MicroRNAs, Hepatitis C virus, Virus-host interactions, New therapeutic targets

## Review

Nearly 20 years after their discovery, we now know that microRNAs (miRNAs) modulate gene expression networks. These small non-coding RNAs (around 22 nucleotides) have been shown to function as regulators of gene expression. First discovered in *Caenorhabditis elegans* [1], miRNAs have quickly been considered a fundamental component of the regulatory system of gene expression and have been estimated to control at least one third of eukaryotic cell genes. miRNAs are transcribed as long primary transcripts (pri-miRNAs) and trimmed into approximately 70 nucleotide stem-loop precursors (pre-miRNAs), most of the time by Drosha in the nucleus [2]. Pre-miRNAs get exported to the cytoplasm by Exportin-5 and processed by Dicer to generate mature miRNAs [3,4]. The mature miRNAs then direct the binding of Argonaute-containing protein complexes (RISC, for RNA-induced silencing complex) on specific

messenger RNA (mRNA) targets to promote their degradation or to affect their translation ([3] and Figure 1). The 5' nucleotides, 2–7 of the mature miRNA, are called the seed sequence and have the strongest influence for target selection. miRNAs that share the same seed sequence are grouped into families, however they may not have the same targets [5]. One miRNA has the ability to target many different sites on the same mRNA or on many different mRNAs.

Today, more than 1500 miRNAs have been identified in the human genome (miRBase release 18 [6-8]), and their biological functions are under investigation. Despite the important advances accomplished to elucidate their role in regulating different physiological and pathological processes, our understanding of their implications in response to viral infection (hepatitis C virus for instance) is in its early stages.

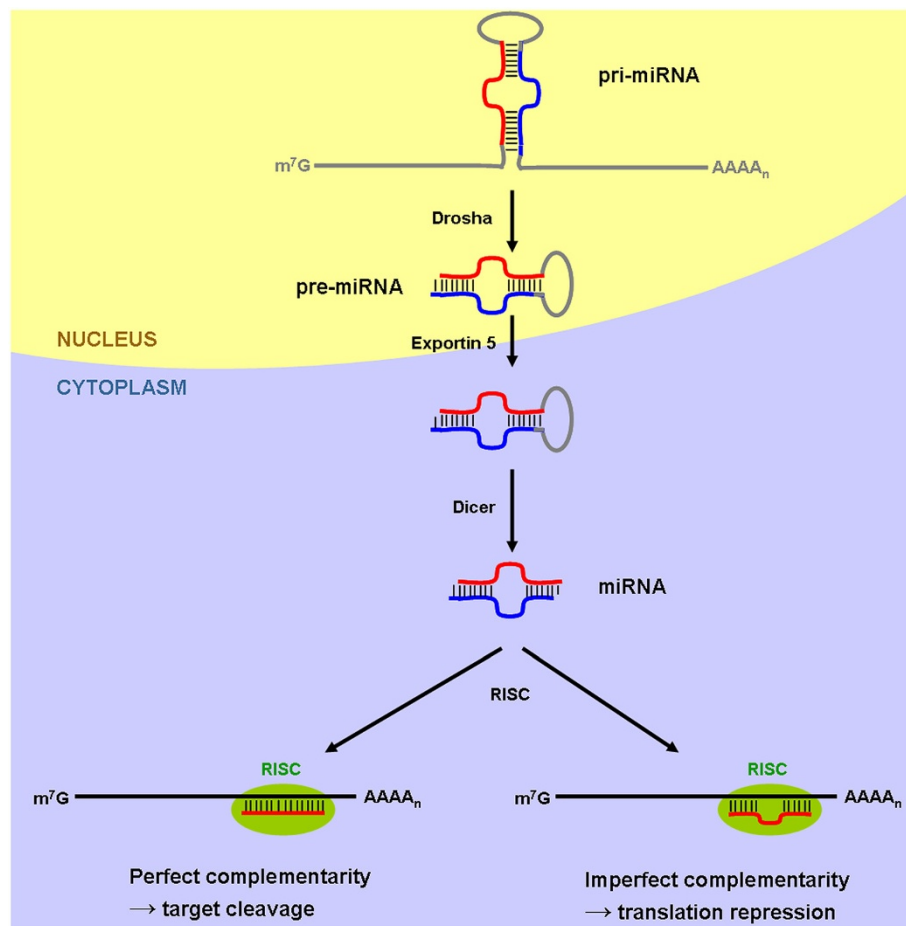
Hepatitis C virus (HCV) is a small enveloped positive-strand RNA virus of the Flaviviridae family, classified in the genus Hepacivirus. It infects around 3% of the world population and it is the most common etiologic agent underlying chronic hepatitis with hepatitis B virus (HBV). Chronic HCV leads to cirrhosis of the liver and ultimately to the development of hepatocellular carcinoma (HCC) [9]. Globally, 31.1% of HCC cases are caused

\* Correspondence: abengrine@biobanque-picardie.com

<sup>1</sup>EA4294 Unité de Virologie Clinique et Fondamentale, Université de Picardie Jules Verne, UFR de Médecine et de Pharmacie, 3 rue des Louvels, 80036, Amiens Cedex, France

<sup>3</sup>Biobanque de Picardie, Centre Hospitalier Universitaire d'Amiens, Avenue René Laennec, 80480, Salouël, France

Full list of author information is available at the end of the article



**Figure 1 Canonical miRNA biogenesis and function.** miRNAs are transcribed as capped and polyadenylated primary transcripts (pri-miRNAs) containing a hairpin structure that is excised by the endonuclease Drosha. This liberated stem-loop (pre-miRNA) is transported to the cytoplasm by exportin 5 to be processed by the Dicer endonuclease. RNA-induced silencing complex (RISC) selectively handles one strand of the resulting duplex and triggers target mRNA silencing through cleavage or translational repression.

by HCV infection, whereas HBV is responsible for 54.4% of HCC cases [10]. The prevalence of HCC, fifth in term of frequency and third in term of mortality within cancers, demonstrates the enormous threat posed by HCV infection to the worldwide public health. Moreover, anti-HCV treatments show irregular efficiency between viral genotypes and the ethnic origin of the infected patients. One example of this irregularity is demonstrated in patients who are infected with the type 1 genotype. They typically only show 50% effectiveness when treated with pegylated interferon (pegIFN) and ribavirin [11]. The rate of sustained viral response (SVR) to treatment is improved to 70% when antiproteases are associated with this standard regimen [12]. Other combinations with new molecules are expected to promote HCV eradication in almost all patients regardless of the HCV genotype.

Recent advancements in understanding virus-host interactions uncovered multiple host factors used by HCV to infect cells [13]. As a major factor in controlling

different cellular processes, miRNAs represent an interesting field of investigation regarding HCV infection and replication. Furthermore, they may represent new targets for the development of antiviral therapeutics. The work of Yang *et al.* provides a proof of concept for the potential use of these molecules. Indeed, these authors showed that HCV replication *in vitro* and in a mouse model was inhibited when 5 artificial anti-HCV miRNAs, cloned in the miR-17-92 cluster and delivered by an adenovirus vector, are expressed [14]. These 5 artificial miRNAs reduced the replication of cell culture-propagated HCV (HCVcc) by 98% and resulted in 93% HCV gene silencing in mouse livers without causing any hepatocellular toxicity. However, these results should be taken with caution as it has also been shown that over-expression of certain pre-miRNA-like molecules in the liver is lethal in mice [15]. Recently, Berezhna *et al.* reported that replication or suppression of HCV RNA depends on the interaction of Ago2 protein (member of RISC complex)

with miRNA or small interfering RNA (siRNA) and its partitioning to different cell compartments [16]. This finding is an additional proof in favor of the therapeutic potential of miRNAs.

Since the first publication reporting the positive role of miR-122 in HCV replication [17], this liver-specific miRNA has been a focus of numerous research projects investigating the liver and HCV interaction. It has 4 binding sites in the HCV genome [18], it is implicated in regulation of different metabolic pathways in liver cells (cholesterol metabolism) [19] and it represents 50% to 70% of all the miRNAs expressed in the liver [20-22]. An anti-HCV treatment based on miR-122 inhibition by antisense oligonucleotides has been tested on chimpanzees and provided very promising results [23]. The same anti-miR-122 molecule is also the first miRNA-targeting treatment to enter into human clinical trials (currently in phase 2) to treat patients infected with HCV (miravirsen, Santaris Pharma A/S). Even though miR-122 has been the most studied miRNA in HCV infection and the clinical trials using anti-miR-122 have shown promising results against HCV, this strategy against miR-122 might have some negative impacts on the hepatocytes' metabolism. Therefore, other miRNAs may also play an important role in HCV-host interactions and be good candidates for the fight against HCV. This is the reason why several research teams around the world are interested in bringing to light the whole "miRNome" and uncover the role played by other miRNAs in controlling such interactions. Randall *et al.* reported a systematic RNA interference (RNAi) screening study on the human hepatoma cell line Huh-7.5 [24]. In this work they targeted 62 cellular genes, using siRNAs thought to play a role in HCV infection, and demonstrated that RNAi targeted 26 of these genes and reduced viral production by at least 3 fold. Some of these siRNAs targeted Dicer or other components of the RNAi pathway, therefore confirming the close interaction between HCV and the miRNA machinery. Other studies reported specific miRNAs altering HCV infection and/or being altered by the presence of the virus. It is reasonable to assume that some of these miRNAs may have a promising potential as therapeutic targets, such as miR-196b, miR-199a-3p and miR-29 that inhibit HCV replication in several models [25-27]. Despite this promising potential, these miRNAs are not as intensively studied as miR-122 [28-30].

In this review, we will therefore discuss recent advances in the relationship between HCV infection and miRNAs and show that some of them might be as important as miR-122 in the fight against HCV.

#### miR-196b

The mir-196 gene family contains 3 members (mir-196a-1, -196a-2, and -196b) and are found in the homeobox

clusters of vertebrates [31]. Two mature miRNA, miR-196a and miR-196b, differing by only one nucleotide, are products of these 3 genes [31]. miR-196 was first reported to have extensive and evolutionarily conserved complementarity to homeobox clusters and to regulate the expression of homeobox genes such as HoxB8 [32]. In addition to its role in development, many studies on miR-196 reported its role in several cancers (for review, see [31]). The increased expression of miR-196b in HCC when compared with normal livers [33] suggests that this miRNA may have an oncogenic role in HCC.

It has been reported that miR-196b has a target site in the NS5A coding region of HCV RNA (Table 1 and [27]). The expression of miR-196b and eight other miRNAs predicted to target HCV RNA is modulated upon interferon (IFN)  $\beta$  treatment of primary mouse hepatocytes and Huh7 cells [27]. Because IFN $\beta$  significantly affects the innate anti-HCV response, the authors tested the impact of these IFN $\beta$ -induced miRNAs on HCV replication. Only 5 miRNAs decreased HCV replication, including miR-196b and an antisense oligonucleotide against miR-122. The decrease of HCV replication ranged from 50% to 80% (60% for miR-196b). Later, Scagnolari *et al.* confirmed the increase of miR-196b expression by the IFN pathway in human peripheral blood mononuclear cells (PBMC) of healthy individuals [34].

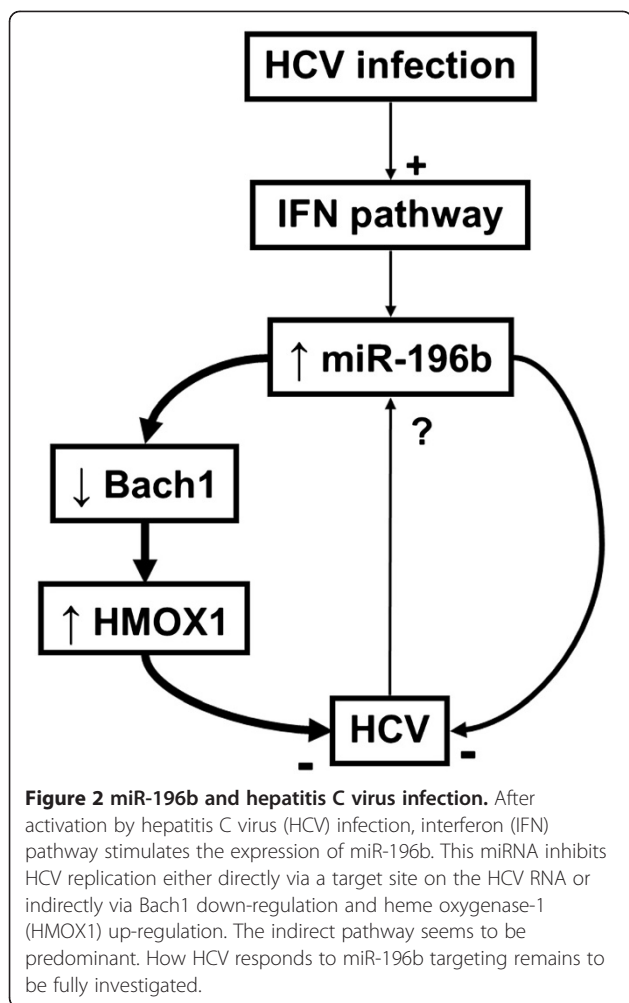
The mechanisms involved in the inhibition of HCV replication by miR-196b were later revealed by Hou *et al.* (Figure 2). In addition to the direct targeting of the HCV RNA, Bach1, a repressor of the anti-oxidative and anti-inflammatory heme oxygenase 1 (HMOX1), is also a direct target of miR-196b [35]. In this study, this miRNA was able to inhibit HCV expression in replicon cell lines (around 50% of inhibition) directly by targeting HCV RNA and indirectly by increasing HMOX1 expression. Evidence suggests that the importance of Bach1 targeting is predominant over direct HCV targeting, however the mechanisms of HCV inhibition by HMOX1 remain unknown. As HCV infection causes oxidative stress and miR-196b inversely modulates HCV replication and HMOX1 expression, overexpression of miR-

**Table 1 Summary of the matches implied in HCV inhibition by 3 microRNAs considered as potential therapeutic targets**

| microRNA    | Inhibition of HCV infection by |                                  |
|-------------|--------------------------------|----------------------------------|
|             | Matches on HCV RNA             | Matches on host targets          |
| miR-196b    | NS5A coding region (6680) [27] | Bach1 3'UTR (2286 and 2166) [35] |
| miR-199a-3p | 5'UTR IRES (60) [26]           | NR                               |
| miR-29      | NR                             | NR                               |

The numbers in brackets indicate the first nucleotide of the binding sites of the miRNAs.

UTR: untranslated region; IRES: internal ribosome entry site; NR: non reported.



196b appears as a potential strategy to protect against liver injury during chronic hepatitis. Interestingly, miR-122 has also been linked to the down-regulation of HMOX1 [36], which means that the overexpression of miR-196b could be even more efficient if associated with inhibition of miR-122 [36].

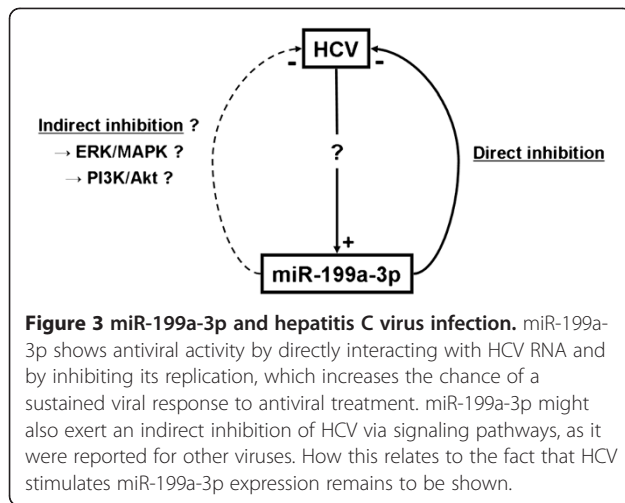
It is unknown whether HCV has evolved to counteract the effect of this antiviral miRNA, although some studies suggest that such a mechanism might exist. Indeed, the expression of miR-196a and miR-196b decreases upon HCV transfection of HepG2 cells as well as in PBMC of chronic hepatitis C patients [34,37]. This decrease could be linked to the inhibition of IFN pathway by the virus, since in PBMCs from chronic hepatitis C patient the expression of miR-196b does not change after IFN $\alpha$  treatment, whereas it is increased in PBMCs from healthy controls [34]. However these results remain to be formally proven, as it has been shown in one study that miR-196b and miR-196a seem to be respectively increased and decreased in HCV replicon-containing cell lines [38]. Furthermore, Grek *et al.* reported in another

study that miR-196b was increased in PBMC replicating HCV [39]. HCV infection and replication in PBMCs still remains controversial, but these results suggest that the interactions between miR-196b and HCV infection are complex. These discordances might also be evidence of the limitations of cell lines and replicons against primary cells and complete virus cycle, respectively. In the light of these data, it could be hypothesized that miR-196b expression is (i) induced by the IFN pathway in cells replicating HCV and (ii) decreased by the pro-viral mechanisms, although this has yet to be verified.

#### miR-199a-3p

According to miRBase release 18 [6-8], the miR-199 family is derived from 3 genes: mir-199a-1, -199a-2 and -199b. In terms of read number, the functional mature miRNA appear to be the 3p form for both miR-199a and miR-199b. Several studies reported an anti-tumoral role for miR-199a-3p, which targets the proliferation and migration/invasion signaling pathways in different cancers, especially HCC [22,40-42]. Others suggested that miR-199a-3p anti-tumoral activity is not associated with a specific cancer [43]. miR-199a-3p has also been associated with liver fibrosis progression [44] and alcoholic or nonalcoholic liver injuries [45].

In the context of viral infection, miR-199a-3p has been identified as a broadly active antiviral miRNA against members of all three herpesvirus subfamilies and the positive sense RNA virus Semliki Forest virus [46]. In this study, miR-199a-3p modulated the signaling pathways involved in viral replication such as PI3K/AKT and ERK/MAPK. The identification of miR-199a-3p binding sites in the 5'UTR IRES of HCV RNA (genotypes 1b and 2a) [26] suggested a direct interaction between these molecules (Table 1). Furthermore, overexpression of miR-199a-3p has an 80-90% inhibitory effect on virus replication, whereas viral replication and protein expression are promoted by the suppression of miR-199a-3p [26]. In the same study, Murakami *et al.* showed that miR-199a-3p induced viral RNA accumulation in the RISC complex as a consequence of direct binding to the 5'UTR IRES and was independent from the IFN pathway. Later, the same researchers showed that increased miR-199a-3p expression (with 34 other miRNAs) was associated with SVR in chronic hepatitis C patients when compared with non-SVR [47]. However, it remains to be investigated whether this miRNA inhibits HCV replication by direct binding to the viral RNA and/or indirectly via the negative regulation of cellular factors. It is worth noting that HBV replication is also inhibited by miR-199a-3p, as a consequence of direct interaction of the miRNA and the HBV coding region [48]. According to these pieces of evidence, the miR-199a-3p mechanism of action could be summarized as



in Figure 3. They also suggest that this miRNA plays a major role in the hepatic antiviral response.

With such a strong effect on HCV replication, we could assume that miR-199a-3p expression is modified during HCV infection. In the numerous studies reporting miRNA expression profiles in HCV-infected cells or tissues, only two demonstrated an impact on miR-199a-3p by showing that the expression of this miRNA was increased during HCV infection [49,50]. It should also be noted that this miRNA is only weakly expressed in the liver [26], which means that further investigations are needed to fully assess the significance of the miR-199a-3p antiviral effect against HCV, as well as how HCV infection modulates the expression of this miRNA.

### miR-29

The miR-29 family comprises 3 members: miR-29a, -29b, and -29c, which are expressed from 4 genes, mir-29a, -29b-1, -29b-2 and 29c (miRBase release 18 [6-8]). To date miR-29 members have been implicated in the fibrosis of different organs such as the liver [51], kidney [52], heart [53] and lung [54]. Furthermore, miR-29 was also associated with IFN $\gamma$  production [55]. These miRNAs appear to have tumor suppressor activity in leukemia [56] and HCC [57,58], as well as an antiviral activity against HBV [57].

The relationship between HCV infection and miR-29 family expression in hepatocytes and hepatic stellate cells (HSC) has been recently reported [25]. The analysis of miRNA profiles of HCVcc-infected Huh-7.5 cells and livers from chronic hepatitis C patients showed a 2-fold decrease of the 3 members of the miR-29 family compared to controls. The activation of HSCs also leads to the down-regulation of miR-29. Moreover, the same study showed that over expression of miR-29 in infected Huh-7.5 cells resulted in a 70% decrease of HCV replication. The over expression of miR-29 in HSCs inhibited

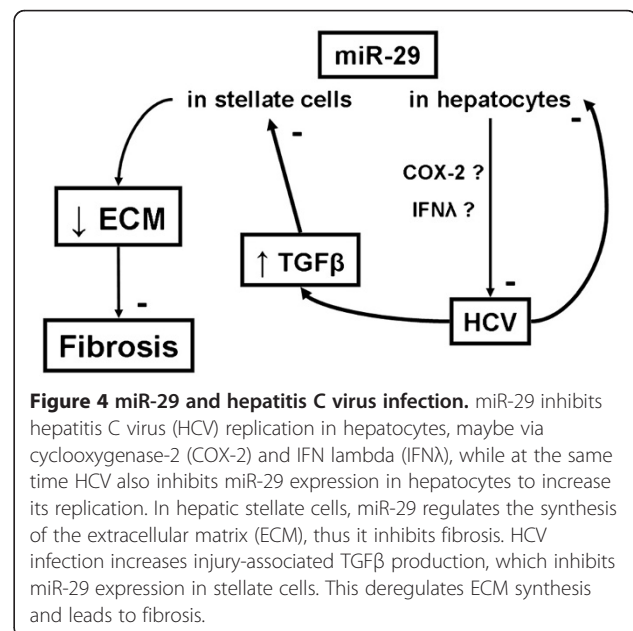
their proliferation and their collagen production. Currently the mechanism explaining how HCV infection triggers the down-regulation of miR-29 in hepatocytes is unknown. It can be hypothesized that in HSCs the miR-29 down regulation is activated by liver injury-induced TGF $\beta$  along with the increased extracellular matrix production, which ultimately leads to fibrosis (Figure 4). Recently, the activation of cyclooxygenase-2 and IFN lambda production have been reported as mechanisms involved in the miR-29 antiviral activity against influenza virus [59]. This pathway has yet to be explored regarding HCV infection.

Others have documented the link between miR-29 family members and HCV infection. These studies showed the expression of miR-29a decreases in HCVcc-infected Huh-7.5 cells [60], and miR-29c is down regulated both in livers of patients presenting viral hepatitis (HCV or HBV) [61] and in livers presenting HCV-associated HCC [62]. Furthermore, in livers from chronic hepatitis C patients presenting SVR, miR-29a, -29b and -29c levels increased when compared to livers of patients without SVR [47]. This finding suggests that the miR-29 family (probably in combination with other miRNAs) could be a useful biomarker when monitoring the response to anti-HCV treatments.

Altogether there is a growing body of evidence suggesting the importance of the miR-29 family's role in the cells' defense against HCV and a treatment with miR-29 might inhibit HCV while reducing fibrosis.

### Other promising miRNAs

Several other miRNAs have been recently reported as associated with HCV infection, either because they



altered viral replication and/or were altered by it (Figure 5). For instance, the expression of miR-141 is increased in HCV-transfected primary human hepatocytes when cocultured with a rat hepatic stellate cell line. Also observed was stimulated virus replication and production [49]. According to the same study, miR-141-mediated HCV stimulation involves the direct inhibition of the tumor-suppressor DLC-1.

By inhibiting the PI3K/Akt pathway, miR-491 stimulates HCV replication in replicon or HCVcc-infected Huh-7 cells [63]. This inhibition does not depend on the viral genotype, cell proliferation or HCV IRES activity. In HCV-infected Huh7 cells, miR-491 expression is increased during the first 4 days post-infection (up to 12 fold) [60], before going down after the fifth day [63]. Moreover, it is established that miR-491 also induces apoptosis [64]. These data suggest that miR-491 is involved in the early steps of HCV infection followed by a decrease in its expression, which might favor the development of HCC. Other interesting findings were reported by Ishida *et al.*, who demonstrated a stimulatory effect of miR-192 and -215 on HCV replication [63].

Meanwhile, the induction of miR-296 by IFN $\beta$  directly inhibits HCV replication in infected Huh7 cells by directly targeting the viral RNA [27]. Furthermore, the expression of miR-296 is decreased in the liver and increased in PBMCs of HCV-infected patients [34,50]. These findings suggest an unknown and complex set of mechanisms and interactions between HCV and miR-296.

As for miR-196b, the expression of miR-351, -431 and -448 is induced by IFN $\beta$ , thus these miRNAs may possess an inhibitory effect on HCV replication through direct targeting of the viral RNA [27]. The expression of miR-130, -146, -200, -192 and -194 is modified by HCV infection in several models as reported by different

studies [37,38,49,50,60,63,65]. Interestingly, miR-192 and -194 are liver-specific miRNAs [66,67] and are associated with SVR in chronic hepatitis C patients [47]. Finally, miR-21, which is associated with liver regeneration [68] and HCC [69], is increased in HCV-infected livers and Huh-7.5 cells [50,70]. At least two studies have showed a correlation between miR-21 increased expression and (i) SVR [47], (ii) viral load, (iii) fibrosis (via inhibition of SMAD7), and (iv) level of transaminases in serum [70].

All these miRNAs represent promising targets for antiviral treatments and as potential biomarkers of HCV infection and its outcome. Further investigations will be required to fully characterize their cellular targets and their modes of action.

## Conclusions

There are now numerous pieces of evidence demonstrating the therapeutic potential of miRNAs during chronic HCV infection. Currently, miR-122 is the only miRNA tested in therapeutic clinical trials designed to treat HCV-infected chimpanzees [23] or human (Santaris, phase 2). Other miRNAs, especially miR-196b, -199a-3p and the miR-29 family, may possibly emerge as very promising targets for anti-HCV treatments. Based on the combination of all these miRNAs a new anti-HCV therapy could be proposed as follows: anti-miR-122, and mimics for miR-196b, miR-199a-3p and miR-29. This combination could also be used in conjunction with standard treatments to increase their efficiency. Such a protocol might have the advantage of preserving patients from HCV replication and from HCV-mediated injuries.

Due to miRNAs having multiple targets and their ability regulate different signaling pathways, it will be important to fully characterize their mechanisms of action and their cellular targets in order to avoid any negative patient outcomes. To date no serious adverse effect of miR-122 inhibition has been observed in chimpanzees [23] or healthy volunteers (Santaris, phase 2), however long term studies will need to be completed to ensure that over-expressed or blocked miRNAs don't cause more damage than they prevent.

## Competing interests

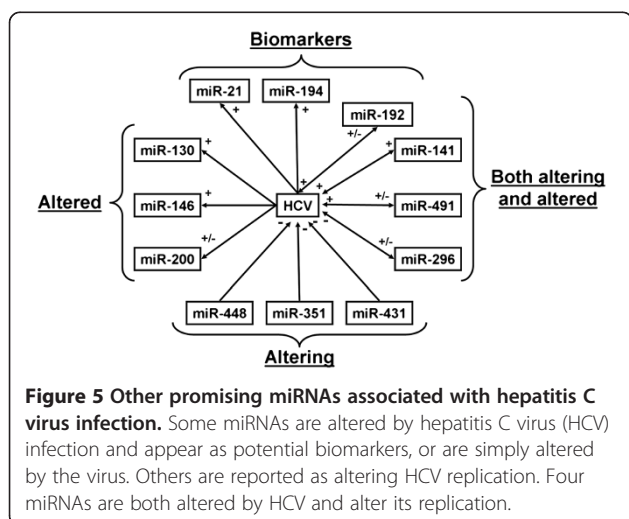
All authors declare that they have no competing interests.

## Authors' contributions

TWH read the articles and gathered information cited in this review. GD revised the manuscript. TWH and AB drafted the manuscript. All authors read and approved the final manuscript.

## Acknowledgements

The authors thank Sébastien Pfeffer (Institut de biologie moléculaire et cellulaire, Strasbourg, France) and Shawn Keil (Terumo BCT, CO, USA), for reading this manuscript and providing helpful suggestions.



#### Author details

<sup>1</sup>EA4294 Unité de Virologie Clinique et Fondamentale, Université de Picardie Jules Verne, UFR de Médecine et de Pharmacie, 3 rue des Louvels, 80036, Amiens Cedex, France. <sup>2</sup>Laboratoire de Virologie, Centre Hospitalier Universitaire d'Amiens, Avenue René Laennec, 80480, Salouël, France. <sup>3</sup>Biobanque de Picardie, Centre Hospitalier Universitaire d'Amiens, Avenue René Laennec, 80480, Salouël, France.

Received: 30 January 2012 Accepted: 30 May 2012

Published: 12 June 2012

#### References

- Lee RC, Feinbaum RL, Ambros V: The *C. elegans* heterochronic gene *lin-4* encodes small RNAs with antisense complementarity to *lin-14*. *Cell* 1993, **75**:843–854.
- Lee Y, Ahn C, Han J, Choi H, Kim J, Yim J, Lee J, Provost P, Radmark O, Kim S, Kim VN: The nuclear RNase III Drosha initiates microRNA processing. *Nature* 2003, **425**:415–419.
- Ambros V: The functions of animal microRNAs. *Nature* 2004, **431**:350–355.
- Yi R, Qin Y, Macara IG, Cullen BR: Exportin-5 mediates the nuclear export of pre-microRNAs and short hairpin RNAs. *Genes Dev* 2003, **17**:3011–3016.
- Jalvy-Delville S, Maurel M, Major V, Pierre N, Chabas S, Combe C, Rosenbaum J, Sagliocco F, Grosset CF: Molecular basis of differential target regulation by miR-96 and miR-182: the Glypican-3 as a model. *Nucleic Acids Res* 2012, **40**:1356–1365.
- Griffiths-Jones S: The microRNA Registry. *Nucleic Acids Res* 2004, **32**:D109–D111.
- Griffiths-Jones S, Saini HK, van Dongen S, Enright AJ: miRBase: tools for microRNA genomics. *Nucleic Acids Res* 2008, **36**:D154–D158.
- Kozomara A, Griffiths-Jones S: miRBase: integrating microRNA annotation and deep-sequencing data. *Nucleic Acids Res* 2011, **39**:D152–D157.
- Shepard CW, Finelli L, Alter MJ: Global epidemiology of hepatitis C virus infection. *Lancet Infect Dis* 2005, **5**:558–567.
- Wu Q, Liu Q: Do hepatitis B virus and hepatitis C virus co-infections increase hepatocellular carcinoma occurrence through synergistically modulating lipogenic gene expression?. *Hepatology* 2012, doi:10.1111/j.1872-1034X.2012.00994.x. in press.
- Feuerstadt P, Bunim AL, Garcia H, Karlitz JJ, Massoumi H, Thosani AJ, Pellicchia A, Wolkoff AW, Gaglio PJ, Reinus JF: Effectiveness of hepatitis C treatment with pegylated interferon and ribavirin in urban minority patients. *Hepatology* 2009, **51**:1137–1143.
- Ilyas JA, Vierling JM: An overview of emerging therapies for the treatment of chronic hepatitis C. *Clin Liver Dis* 2011, **15**:515–536.
- Poenisch M, Bartschlagler R: New insights into structure and replication of the hepatitis C virus and clinical implications. *Semin Liver Dis* 2010, **30**:333–347.
- Yang X, Haurigot V, Zhou S, Luo G, Couto LB: Inhibition of hepatitis C virus replication using adeno-associated virus vector delivery of an exogenous anti-hepatitis C virus microRNA cluster. *Hepatology* 2010, **52**:1877–1887.
- Grimm D, Streetz KL, Jopling CL, Storm TA, Pandey K, Davis CR, Marion P, Salazar F, Kay MA: Fatality in mice due to oversaturation of cellular microRNA/short hairpin RNA pathways. *Nature* 2006, **441**:537–541.
- Berezina SY, Supekova L, Sever MJ, Schultz PG, Deniz AA: Dual regulation of hepatitis C viral RNA by cellular RNAi requires partitioning of Ago2 to lipid droplets and P-bodies. *RNA* 2011, **17**:1831–1845.
- Jopling CL, Yi M, Lancaster AM, Lemon SM, Sarnow P: Modulation of hepatitis C virus RNA abundance by a liver-specific microRNA. *Science* 2005, **309**:1577–1581.
- Nasheri N, Singaravelu R, Goodmurphy M, Lyn RK, Pezacki JP: Competing roles of microRNA-122 recognition elements in hepatitis C virus RNA. *Virology* 2011, **410**:336–344.
- Jopling C: Liver-specific microRNA-122: Biogenesis and function. *RNA Biol* 2012, **9**:137–142.
- Lagos-Quintana M, Rauhut R, Yalcin A, Meyer J, Lendeckel W, Tuschl T: Identification of tissue-specific microRNAs from mouse. *Curr Biol* 2002, **12**:735–739.
- Chang J, Nicolas E, Marks D, Sander C, Lerro A, Buendia MA, Xu C, Mason WS, Moloshok T, Bort R, Zaret KS, Taylor JM: miR-122, a mammalian liver-specific microRNA, is processed from hcr mRNA and may downregulate the high affinity cationic amino acid transporter CAT-1. *RNA Biol* 2004, **1**:106–113.
- Hou J, Lin L, Zhou W, Wang Z, Ding G, Dong Q, Qin L, Wu X, Zheng Y, Yang Y, Tian W, Zhang Q, Wang C, Zhang Q, Zhuang SM, Zheng L, Liang A, Tao W, Cao X: Identification of miRNomes in human liver and hepatocellular carcinoma reveals miR-199a/b-3p as therapeutic target for hepatocellular carcinoma. *Cancer Cell* 2011, **19**:232–243.
- Lanford RE, Hildebrandt-Eriksen ES, Petri A, Persson R, Lindow M, Munk ME, Kauppinen S, Orum H: Therapeutic silencing of microRNA-122 in primates with chronic hepatitis C virus infection. *Science* 2010, **327**:198–201.
- Randall G, Panis M, Cooper JD, Tellinghuisen TL, Sukhodolets KE, Pfeffer S, Landthaler M, Landgraf P, Kan S, Lindenbach BD, Chien M, Weir DB, Russo JJ, Ju J, Brownstein MJ, Sheridan R, Sander C, Zavolan M, Tuschl T, Rice CM: Cellular cofactors affecting hepatitis C virus infection and replication. *Proc Natl Acad Sci U S A* 2007, **104**:12884–12889.
- Bandyopadhyay S, Friedman RC, Marquez RT, Keck K, Kong B, Icardi MS, Brown KE, Burge CB, Schmidt WN, Wang Y, McCaffrey AP: Hepatitis C virus infection and hepatic stellate cell activation downregulate miR-29: miR-29 overexpression reduces hepatitis C viral abundance in culture. *J Infect Dis* 2011, **203**:1753–1762.
- Murakami Y, Aly HH, Tajima A, Inoue I, Shimotohno K: Regulation of the hepatitis C virus genome replication by miR-199a\*. *J Hepatol* 2009, **50**:453–460.
- Pedersen IM, Cheng G, Wieland S, Volinia S, Croce CM, Chisari FV, David M: Interferon modulation of cellular microRNAs as an antiviral mechanism. *Nature* 2007, **449**:919–922.
- Kerr TA, Korenblat KM, Davidson NO: MicroRNAs and liver disease. *Transl Res* 2011, **157**:241–252.
- Kumar A: MicroRNA in HCV infection and liver cancer. *Biochim Biophys Acta* 2011, **1809**:694–699.
- Pfeffer S, Baumert TF: Impact of microRNAs for pathogenesis and treatment of hepatitis C virus infection. *Gastroenterol Clin Biol* 2010, **34**:431–435.
- Chen C, Zhang Y, Zhang L, Weakley SM, Yao Q: MicroRNA-196: critical roles and clinical applications in development and cancer. *J Cell Mol Med* 2011, **15**:14–23.
- Yekta S, Shih IH, Bartel DP: MicroRNA-directed cleavage of HOXB8 mRNA. *Science* 2004, **304**:594–596.
- Wong QW, Lung RW, Law PT, Lai PB, Chan KY, To KF, Wong N: MicroRNA-223 is commonly repressed in hepatocellular carcinoma and potentiates expression of Stathmin1. *Gastroenterology* 2008, **135**:257–269.
- Scagnolari C, Zingariello P, Vecchiet J, Selvaggi C, Racciatti D, Taliani G, Riva E, Pizzigallo E, Antonelli G: Differential expression of interferon-induced microRNAs in patients with chronic hepatitis C virus infection treated with pegylated interferon alpha. *Viral J* 2010, **7**:311.
- Hou W, Tian Q, Zheng J, Bonkovsky HL: MicroRNA-196 represses Bach1 protein and hepatitis C virus gene expression in human hepatoma cells expressing hepatitis C viral proteins. *Hepatology* 2010, **51**:1494–1504.
- Shan Y, Zheng J, Lambrecht RW, Bonkovsky HL: Reciprocal effects of microRNA-122 on expression of heme oxygenase-1 and hepatitis C virus genes in human hepatocytes. *Gastroenterology* 2007, **133**:1166–1174.
- Braconi C, Valeri N, Gasparini P, Huang N, Taccioli C, Nuovo G, Suzuki T, Croce CM, Patel T: Hepatitis C virus proteins modulate microRNA expression and chemosensitivity in malignant hepatocytes. *Clin Cancer Res* 2010, **16**:957–966.
- Bruni R, Marcantonio C, Tritarelli E, Tataseo P, Stellacci E, Costantino A, Villano U, Battistini A, Ciccaglione AR: An integrated approach identifies IFN-regulated microRNAs and targeted mRNAs modulated by different HCV replicon clones. *BMC Genomics* 2011, **12**:485.
- Greg M, Piekarska A, Bartkowiak J, Fendler W, Kuydowski P, Wroblewski P, Paradowski M, Sidorkiewicz M: Coordinated increase of miRNA-155 and miRNA-196b expression correlates with the detection of the antigenomic strand of hepatitis C virus in peripheral blood mononuclear cells. *Int J Mol Med* 2011, **28**:875–880.
- Fornari F, Milazzo M, Chieco P, Negrini M, Calin GA, Grazi GL, Pollutri D, Croce CM, Bolondi L, Gramantieri L: MiR-199a-3p regulates mTOR and c-Met to influence the doxorubicin sensitivity of human hepatocarcinoma cells. *Cancer Res* 2010, **70**:5184–5193.
- Henry JC, Park JK, Jiang J, Kim JH, Nagorney DM, Roberts LR, Banerjee S, Schmittgen TD: miR-199a-3p targets CD44 and reduces

- proliferation of CD44 positive hepatocellular carcinoma cell lines. *Biochem Biophys Res Commun* 2010, **403**:120–125.
42. Jia XQ, Cheng HQ, Qian X, Bian CX, Shi ZM, Zhang JP, Jiang BH, Feng ZQ: **Lentivirus-Mediated Overexpression of MicroRNA-199a Inhibits Cell Proliferation of Human Hepatocellular Carcinoma.** *Cell Biochem Biophys* 2012, **62**:237–244.
  43. Kim S, Lee UJ, Kim MN, Lee EJ, Kim JY, Lee MY, Choung S, Kim YJ, Choi YC: **MicroRNA miR-199a\* regulates the MET proto-oncogene and the downstream extracellular signal-regulated kinase 2 (ERK2).** *J Biol Chem* 2008, **283**:18158–18166.
  44. Murakami Y, Toyoda H, Tanaka M, Kuroda M, Harada Y, Matsuda F, Tajima A, Kosaka N, Ochiya T, Shimotohno K: **The progression of liver fibrosis is related with overexpression of the miR-199 and 200 families.** *PLoS One* 2011, **6**:e16081.
  45. Dolganiuc A, Petrasek J, Kodys K, Catalano D, Mandrekar P, Velayudham A, Szabo G: **MicroRNA expression profile in Lieber-DeCarli diet-induced alcoholic and methionine choline deficient diet-induced nonalcoholic steatohepatitis models in mice.** *Alcohol Clin Exp Res* 2009, **33**:1704–1710.
  46. Santhakumar D, Forster T, Laqtom NN, Fragkoudis R, Dickinson P, Abreu-Goodger C, Manakov SA, Choudhury NR, Griffiths SJ, Vermeulen A, Enright AJ, Dutia B, Kohl A, Ghazal P, Buck AH: **Combined agonist-antagonist genome-wide functional screening identifies broadly active antiviral microRNAs.** *Proc Natl Acad Sci U S A* 2010, **107**:13830–13835.
  47. Murakami Y, Tanaka M, Toyoda H, Hayashi K, Kuroda M, Tajima A, Shimotohno K: **Hepatic microRNA expression is associated with the response to interferon treatment of chronic hepatitis C.** *BMC Med Genomics* 2010, **3**:48.
  48. Zhang GL, Li YX, Zheng SQ, Liu M, Li X, Tang H: **Suppression of hepatitis B virus replication by microRNA-199a-3p and microRNA-210.** *Antiviral Res* 2010, **88**:169–175.
  49. Banaudha K, Kaliszewski M, Korolnek T, Florea L, Yeung ML, Jeang KT, Kumar A: **MicroRNA silencing of tumor suppressor DLC-1 promotes efficient hepatitis C virus replication in primary human hepatocytes.** *Hepatology* 2011, **53**:53–61.
  50. Peng X, Li Y, Walters KA, Rosenzweig ER, Lederer SL, Aicher LD, Proll S, Katze MG: **Computational identification of hepatitis C virus associated microRNA-mRNA regulatory modules in human livers.** *BMC Genomics* 2009, **10**:373.
  51. Roderburg C, Urban GW, Bettermann K, Vucur M, Zimmermann H, Schmidt S, Janssen J, Koppe C, Knolle P, Castoldi M, Tacke F, Trautwein C, Luedde T: **Micro-RNA profiling reveals a role for miR-29 in human and murine liver fibrosis.** *Hepatology* 2010, **53**:209–218.
  52. Qin W, Chung AC, Huang XR, Meng XM, Hui DS, Yu CM, Sung JJ, Lan HY: **TGF-beta/Smad3 Signaling Promotes Renal Fibrosis by Inhibiting miR-29.** *J Am Soc Nephrol* 2011, **22**:1462–1474.
  53. van Rooij E, Sutherland LB, Thatcher JE, DiMaio JM, Naseem RH, Marshall WS, Hill JA, Olson EN: **Dysregulation of microRNAs after myocardial infarction reveals a role of miR-29 in cardiac fibrosis.** *Proc Natl Acad Sci U S A* 2008, **105**:13027–13032.
  54. Cushing L, Kuang PP, Qian J, Shao F, Wu J, Little F, Thannickal VJ, Cardoso WV, Lu J: **miR-29 Is a Major Regulator of Genes Associated with Pulmonary Fibrosis.** *Am J Respir Cell Mol Biol* 2011, **45**:287–294.
  55. Steiner DF, Thomas MF, Hu JK, Yang Z, Babiarz JE, Allen CD, Matloubian M, Brelloch R, Ansel KM: **MicroRNA-29 Regulates T-Box Transcription Factors and Interferon-gamma Production in Helper T Cells.** *Immunity* 2011, **35**:169–181.
  56. Garzon R, Heaphy CE, Havelange V, Fabbri M, Volinia S, Tsao T, Zanesi N, Kornblau SM, Marcucci G, Calin GA, Andreeff M, Croce CM: **MicroRNA 29b functions in acute myeloid leukemia.** *Blood* 2009, **114**:5331–5341.
  57. Wang CM, Wang Y, Fan CG, Xu FF, Sun WS, Liu YG, Jia JH: **miR-29c targets TNFAIP3, inhibits cell proliferation and induces apoptosis in hepatitis B virus-related hepatocellular carcinoma.** *Biochem Biophys Res Commun* 2011, **411**:586–592.
  58. Xiong Y, Fang JH, Yun JP, Yang J, Zhang Y, Jia WH, Zhuang SM: **Effects of microRNA-29 on apoptosis, tumorigenicity, and prognosis of hepatocellular carcinoma.** *Hepatology* 2010, **51**:836–845.
  59. Fang J, Hao Q, Liu L, Li Y, Wu J, Huo X, Zhu Y: **Epigenetic Changes Mediated by MicroRNA miR29 Activate Cyclooxygenase 2 and Lambda-1 Interferon Production during Viral Infection.** *J Virol* 2012, **86**:1010–1020.
  60. Liu X, Wang T, Wakita T, Yang W: **Systematic identification of microRNA and messenger RNA profiles in hepatitis C virus-infected human hepatoma cells.** *Virology* 2010, **398**:57–67.
  61. Ura S, Honda M, Yamashita T, Ueda T, Takatori H, Nishino R, Sunakozaka H, Sakai Y, Horimoto K, Kaneko S: **Differential microRNA expression between hepatitis B and hepatitis C leading disease progression to hepatocellular carcinoma.** *Hepatology* 2009, **49**:1098–1112.
  62. Varnholt H, Drebber U, Schulze F, Wedemeyer I, Schirmacher P, Dienes HP, Odenthal M: **MicroRNA gene expression profile of hepatitis C virus-associated hepatocellular carcinoma.** *Hepatology* 2008, **47**:1223–1232.
  63. Ishida H, Tatsumi T, Hosui A, Nawa T, Kodama T, Shimizu S, Hikita H, Hiramatsu N, Kanto T, Hayashi N, Takehara T: **Alterations in microRNA expression profile in HCV-infected hepatoma cells: Involvement of miR-491 in regulation of HCV replication via the PI3 kinase/Akt pathway.** *Biochem Biophys Res Commun* 2011, **412**:92–97.
  64. Nakano H, Miyazawa T, Kinoshita K, Yamada Y, Yoshida T: **Functional screening identifies a microRNA, miR-491 that induces apoptosis by targeting Bcl-X(L) in colorectal cancer cells.** *Int J Cancer* 2010, **127**:1072–1080.
  65. Steuerwald NM, Parsons JC, Bennett K, Bates TC, Bonkovsky HL: **Parallel microRNA and mRNA expression profiling of (genotype 1b) human hepatoma cells expressing hepatitis C virus.** *Liver Int* 2010, **30**:1490–1504.
  66. Barad O, Meiri E, Avniel A, Aharonov R, Barzilai A, Bentwich I, Einav U, Gilad S, Hurban P, Karov Y, Lobenhofer EK, Sharon E, Shibolet Y, Shuttman M, Bentwich Z, Einat P: **MicroRNA expression detected by oligonucleotide microarrays: system establishment and expression profiling in human tissues.** *Genome Res* 2004, **14**:2486–2494.
  67. Tang X, Gal J, Zhuang X, Wang W, Zhu H, Tang G: **A simple array platform for microRNA analysis and its application in mouse tissues.** *RNA* 2007, **13**:1803–1822.
  68. Marquez RT, Wendlandt E, Galle CS, Keck K, McCaffrey AP: **MicroRNA-21 is upregulated during the proliferative phase of liver regeneration, targets Pellino-1, and inhibits NF-kappaB signaling.** *Am J Physiol Gastrointest Liver Physiol* 2010, **298**:G535–G541.
  69. Meng F, Henson R, Wehbe-Janek H, Ghoshal K, Jacob ST, Patel T: **MicroRNA-21 regulates expression of the PTEN tumor suppressor gene in human hepatocellular cancer.** *Gastroenterology* 2007, **133**:647–658.
  70. Marquez RT, Bandyopadhyay S, Wendlandt EB, Keck K, Hoffer BA, Icardi MS, Christensen RN, Schmidt WN, McCaffrey AP: **Correlation between microRNA expression levels and clinical parameters associated with chronic hepatitis C viral infection in humans.** *Lab Invest* 2010, **90**:1727–1736.

doi:10.1186/1743-422X-9-109

Cite this article as: Hoffmann et al.: MicroRNAs and hepatitis C virus: Toward the end of miR-122 supremacy. *Virology Journal* 2012 **9**:109.

**Submit your next manuscript to BioMed Central and take full advantage of:**

- Convenient online submission
- Thorough peer review
- No space constraints or color figure charges
- Immediate publication on acceptance
- Inclusion in PubMed, CAS, Scopus and Google Scholar
- Research which is freely available for redistribution

Submit your manuscript at  
www.biomedcentral.com/submit

