

REVIEW

Open Access

H. pylori exploits and manipulates innate and adaptive immune cell signaling pathways to establish persistent infection

Anne Müller^{1*}, Mathias Oertli¹ and Isabelle C Arnold^{1,2}

Abstract

Persistent infection with the gastric bacterial pathogen *Helicobacter pylori* causes gastritis and predisposes carriers to a high gastric cancer risk, but has also been linked to protection from allergic, chronic inflammatory and autoimmune diseases. In the course of tens of thousands of years of co-existence with its human host, *H. pylori* has evolved elaborate adaptations that allow it to persist in the hostile environment of the stomach in the face of a vigorous innate and adaptive immune response. For this review, we have identified several key immune cell types and signaling pathways that appear to be preferentially targeted by the bacteria to establish and maintain persistent infection. We explore the mechanisms that allow the bacteria to avoid detection by innate immune cells via their pattern recognition receptors, to escape T-cell mediated adaptive immunity, and to reprogram the immune system towards tolerance rather than immunity. The implications of the immunomodulatory properties of the bacteria for the prevention of allergic and auto-immune diseases in chronically infected individuals are also discussed.

Keywords: immune evasion, innate immune signaling, immunomodulation, persistent infection

Innate immune receptor recognition of *H. pylori*

Innate immune cells as well as epithelial cells forming a first barrier to infection detect invading pathogens via their conserved microbial structures, the so-called pathogen-associated molecular patterns (PAMPs). Examples of PAMPs include microbial nucleic acids, and cell wall and flagellar components such as peptidoglycan, lipopolysaccharide (LPS), lipoproteins and flagellins [1]. PAMPs are recognized by at least four distinct classes of innate immune or pattern recognition receptors (PRRs) that are present either on cytoplasmic and endosomal membranes (Toll-like receptors, TLRs, and C-type lectin receptors, CLRs) or in the cytosol (NOD-like receptors, NLRs, RIG-like receptors, RLRs). Generally speaking, the ligation of TLRs, CLRs or RLRs results in the activation of pro-inflammatory transcription factors in the nucleus; in contrast, most NLRs are involved in the assembly of multi-protein complexes termed “inflammasomes”,

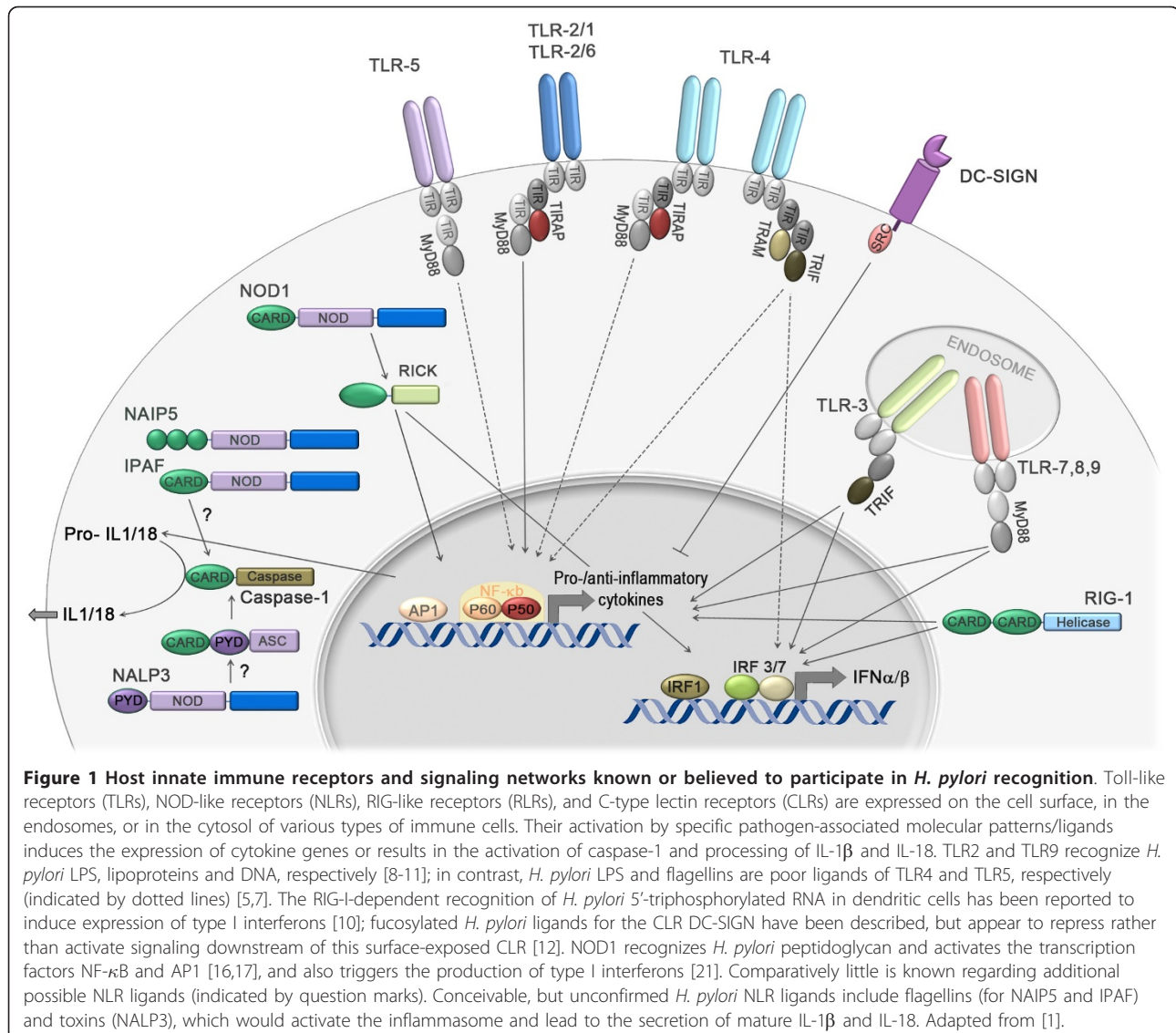
which process pro-IL-1 β and pro-IL-18 to generate the mature, bioactive cytokines [2] (summarized in Figure 1).

PRRs are present not only on hematopoietic cells, but also on gastric epithelial cells which form a first line of defense against *H. pylori* infection [3,4]; therefore, many of the seminal studies in the field have focused on examining *H. pylori* recognition by epithelial cells. In particular, TLR4, 5 and 9 have been detected immunohistochemically on gastric epithelial cells in *H. pylori*-infected as well as uninfected patients, which in principle allows these cells to sense and respond to the infection [3,4]. Interestingly, *H. pylori* differs from other gastrointestinal pathogens in that it has evolved to largely avoid recognition by PAMPs. *H. pylori* flagellin is a poor ligand of TLR5 [5] due to mutations in the TLR5 recognition site of the N-terminal D1 domain of flagellin [6]. Indeed, mutating residues 89-96 of the strongly recognized *Salmonella* flagellin to the corresponding *H. pylori* flaA sequence abolishes TLR5 recognition [6]. The bacterium's LPS consists predominantly of the tetra-acylated lipid A variety, which is known to exhibit 1000-fold reduced bioactivity as compared to *E. coli* LPS [7]. *H. pylori* LPS activates the TLR4/MD-2

* Correspondence: mueller@imcr.uzh.ch

¹Institute of Molecular Cancer Research, University of Zürich, Zürich, Switzerland

Full list of author information is available at the end of the article



complex, but this has only been shown *in vitro* in cells overexpressing the complex [4]. Rather than being a strong TLR4 ligand, *H. pylori* LPS is thought to activate TLR2 on gastric epithelial cells, but again this has only been demonstrated *in vitro* using ectopic expression of the TLR [8,9]. Animal and cell culture experiments suggest that TLR2 ligands (LPS or other) indeed exist in *H. pylori* and related *Helicobacter* species [9-11], and can bind to TLR2 and activate NF- κ B in epithelial cells [9]. However, the net effects of TLR2 ligation are anti-rather than pro-inflammatory *in vivo* as evidenced by lower *H. pylori* colonization levels and more severe immunopathology in gene-targeted mice lacking TLR2 [11]. While *H. pylori* DNA activates TLR9 expressed on dendritic cells (DCs) *in vitro* [10], TLR9^{-/-} mice do not differ from wild type mice in their ability to control *H. pylori* infections [11].

H. pylori's 5'-triphosphorylated RNA can be sensed by the intracellular receptor RIG-1 and triggers type I interferons in DCs [10], but the role of this response in the control of the infection and the infection-associated pathology has to date not been explored. *H. pylori* further possesses fucosylated ligands for the C-type lectin receptor DC-SIGN [12]. However, in contrast to the mannosylated DC-SIGN ligands of *Mycobacterium tuberculosis*, which activate signaling downstream of DC-SIGN and trigger pro-inflammatory cytokine production, the DC-SIGN ligands of *H. pylori* actively dissociate the signaling complex downstream of this C-type lectin to suppress pro-inflammatory cytokine production [12]. The PRRs involved in sensing and responding to *H. pylori* are summarized in Figure 1. Taken together, the data available to date provide ample evidence of the impressive ability of *H. pylori* to avoid

innate immune detection by the host's arsenal of PRRs, thereby preventing innate and adaptive immunity and ensuring its persistence.

NOD1 signaling contributes to anti-*H. pylori* defense mechanisms

As mentioned earlier, the NLRs NOD1 and NOD2 recognize peptidoglycan metabolites of various pathogenic bacteria [13,14] and induce the transcription factor NF- κ B to activate immune response genes [15]. Indeed, NOD1 was among the first PRRs shown to recognize an *H. pylori*-derived PAMP, i.e. its peptidoglycan [16], and has since attracted considerable attention. In their initial study, Ferrero and colleagues showed that peptidoglycan recognition by NOD1 was dependent on the expression of a functional Cag pathogenicity island (PAI)-encoded type IV secretion system (T4SS) and resulted in NF- κ B activation [16]. More recently, the same group demonstrated that MAP kinases and the AP1 transcription factor were also activated upon peptidoglycan recognition by NOD1 [17]. NOD1 activation by *cagPAI*⁺ *H. pylori* was further shown to be required for production of beta-defensins by gastric epithelial cells, which contributes to the bactericidal activity of cell culture supernatants against *H. pylori* [18]. *H. pylori* peptidoglycan delivery to cytosolic NOD1 via the T4SS occurs at cholesterol-rich microdomains referred to as lipid rafts, which contain high local concentrations of the T4SS receptor $\alpha 5\beta 1$ integrin [19]; both cholesterol and $\alpha 5\beta 1$ integrin are required for the T4SS-dependent delivery of peptidoglycan [19]. Although it was initially believed that peptidoglycan delivery occurs exclusively via the T4SS [16], Kaparakis *et al.* now provide evidence that outer membrane vesicles prepared from *cagPAI*⁺ *H. pylori* can also target peptidoglycan to cytosolic NOD1, and that intragastric delivery of peptidoglycan via outer membrane vesicles is sufficient to trigger innate and adaptive immune responses in mice [20].

A novel pathway downstream of NOD1 was recently elucidated in an elegant study by Watanabe *et al.*, who were able to link NOD1 signaling to the production of type I interferons and to the control of *H. pylori* infection [21]. Upon binding to its specific ligand, iE-DAP, NOD1 was shown to induce the sequential activation of the serine threonine kinase RICK, the TNF-associated factor 3 (TRAF3), the kinases TBK1 and IKK ϵ , and ultimately the transcription factor IFN regulatory factor 7 (IRF7) [21]. IRF7 in turn induces IFN- β production, which leads to activation of the heterotrimeric transcription factor complex IFN-stimulated gene factor 3 (ISGF3) and the subsequent production of more IFN- β . Mice lacking the IFN- β receptor exhibit increased susceptibility to *H. pylori* infection, thereby phenocopying the defect of NOD1-deficient mice and suggesting that this pathway participates in host

defenses against *H. pylori* [21]. The complete pathway is summarized in Figure 2. The study by Watanabe *et al.* refutes previous work showing that Nod1 activation induces NF- κ B activity; at least in epithelial cells, the authors found no or minimal NF- κ B activation upon Nod1 ligation by iE-DAP [21]. Another interesting recent study by Liu *et al.* showed that NOD1 and NOD2 are targets of the immunomodulatory glycoprotein olfactomedin 4 (OLFM4) in the context of *H. pylori* infection [22]. OLFM4 is an NF- κ B target gene and associates directly with both NOD proteins, thereby creating a negative feedback loop that impairs *H. pylori*-induced NF- κ B activation. OLFM4 knockout mice exhibited reduced *H. pylori* loads and enhanced gastric immune cell infiltration compared to wild type animals, suggesting that OLFM4 acts as negative regulator of *H. pylori*-specific, NOD-mediated immune responses [22].

Inhibition of T-cell signaling and proliferation by the *H. pylori* virulence factors VacA and γ -glutamyl-transpeptidase

CD4⁺ MHC class II-restricted T-cells are absolutely required for the control of experimental *H. pylori* infections and for the development of vaccine-induced protective immunity [23-25]. It therefore does not come as a big surprise that *H. pylori* has evolved virulence factors in the course of its 60,000+ years of co-existence with the human host [26] that allow it to suppress T-cell-mediated immunity. Seminal work by Haas, Cover, Baldari and their co-workers identified the vacuolating cytotoxin (VacA) as a key factor in the *H. pylori*-mediated inhibition of human T-cells [27-29]. VacA had initially been identified due to its ability to induce vacuolization of epithelial cells [30]. Haas, Cover, Baldari and colleagues extended these original findings to show that VacA inhibits cell proliferation by interfering with the T-cell receptor/interleukin-2 (IL-2) signaling pathway at the level of the Ca²⁺-calmodulin-dependent phosphatase calcineurin [27-29]. VacA thus prevents the nuclear translocation of the T-cell transcription factor NFAT, a global regulator of T-cell responses, resulting in downregulation of IL-2 gene transcription [27,28]. In subsequent studies, Haas and co-workers identified $\beta 2$ (CD18) integrin to be the relevant receptor for VacA on human T-cells [31]. The authors were able to show that VacA exploits the recycling of lymphocyte function-associated antigen-1 (LFA-1; an integrin heterodimer on T-cells consisting of a $\beta 2$ subunit associated with the CD11a α subunit) to become internalized by migrating human T-cells. LFA-1-deficient Jurkat T cells were resistant to vacuolation and IL-2 modulation, and genetic complementation restored their sensitivity to VacA [31]. A recent study further showed that VacA uptake depends on protein kinase C-mediated serine/threonine phosphorylation events,

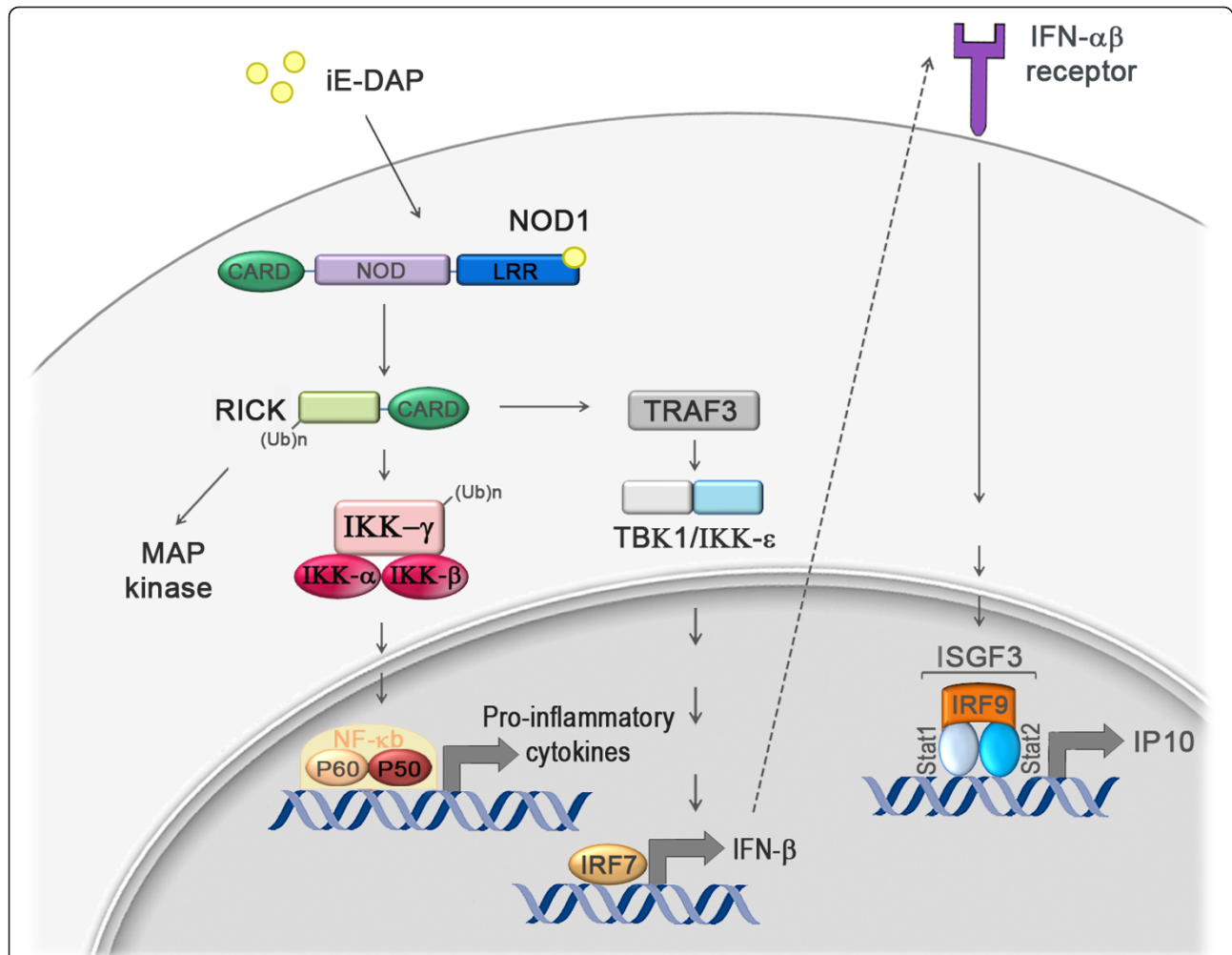


Figure 2 Schematic of NOD1 signaling pathways. NOD1 activation by its specific ligand iE-DAP activates at least two distinct signaling pathways. On the one hand, activated NOD1 triggers IFN- β production through the consecutive activation of RICK, TRAF3, TBK1/IKK ϵ and IRF7. IRF7-dependent IFN- β production leads to the expression of IP-10 and other Th1 cytokines via transactivation of ISGF3 (Stat1-Stat2-IRF9 complex). On the other hand, NOD1 activation triggers the RICK and IKK- α,β,γ complex-dependent activation of NF- κ B and subsequent production of proinflammatory cytokines and chemokines such as IL-8. Whether NF- κ B is indeed activated upon Nod1 ligation in all cell types, including epithelial cells, remains controversial and will be the subject of future experimentation. Adapted from [56] and [21].

presumably of a specific threonine in the β 2/CD18 cytoplasmic tail [32].

An alternative mechanism of T-cell suppression has been proposed by Gerhard and co-workers, who provided evidence in 2005 that human T-cell proliferation is blocked by *H. pylori* without accompanying effects on NFAT activation or cytokine production [33]. The authors postulated at the time that a secreted low-molecular-weight protein distinct from VacA arrests antigen-activated T-cells in the G1 phase of the cell cycle by interfering with G1 cyclin-dependent kinase activity [33]. The authors later demonstrated through a biochemical approach that the secreted γ -glutamyl transpeptidase (GGT) of *H. pylori* is the responsible factor for inhibition of T-cell proliferation; mutagenesis of GGT abrogated

the inhibitory effect of the bacteria and recombinantly expressed GGT enzyme showed anti-proliferative activity [34]. The authors concluded from measuring reduced levels of c-Myc and phosphorylated c-Raf protein that GGT induces cell cycle arrest by disrupting the Ras signaling pathway [34].

***H. pylori* infection induces "tolerogenic" DCs and regulatory T-cells with suppressive activity**

Direct inhibition of T-cells by *H. pylori* virulence factors represents a compelling mechanism of immunosuppression and likely contributes to the bacteria's persistence in the human stomach. In addition, several laboratories have reported lately that *H. pylori*-specific effector T-cell responses are under the strict control of regulatory

T-cells (Tregs) in infected humans, and that the depletion of Tregs improves immunological control over the infection and enhances vaccine-induced protective immunity in mouse models [35-38]. The degree to which the infected host generates *H. pylori*-specific Tregs appears to depend largely on the age at the time of infection [35]. Mice that are experimentally infected as adults with virulent *H. pylori* harboring the Cag pathogenicity island-encoded T4SS mentioned earlier rapidly develop gastritis and gastric cancer precursor lesions manifesting histologically as atrophic gastritis, intestinal metaplasia and epithelial hyperplasia [35]. In contrast, mice that are exposed to *H. pylori* during the neonatal period are protected against the development of such lesions, despite the fact that they are colonized at much higher levels than their adult-infected counterparts. The difference in *H. pylori* colonization and the associated pathology is closely linked to dramatic differences in local as well as systemic immune responses to the bacteria between the age groups. Neonatally infected mice develop immunological tolerance to the infection, which prevents them from generating *H. pylori*-specific T-helper responses and protects them from developing T-cell driven immunopathology. *H. pylori*-specific immunological tolerance depends on regulatory T-cells; their systemic depletion breaks tolerance and sensitizes mice to gastric immunopathology [35]. Whether immunological tolerance develops in humans infected early enough with *H. pylori* remains a matter of speculation. A study showing that infected children, but not adults, preferentially generate regulatory T-cell responses to the infection argues in favor of the tolerance model [39].

Above and beyond their role in preventing immunopathological gastric T-helper responses to the infection, Tregs are crucial mediators of the protection against allergen-induced asthma that is conferred by *H. pylori* infection in mice [40] and humans [41-44]. In infected mice subjected to an experimental protocol inducing airway inflammation and hyper-responsiveness through sensitization and challenge with ovalbumin, the protection against asthma can be abrogated by Treg depletion [40]. Conversely, asthma protection can be transferred from infected to uninfected mice by highly purified population of Tregs [40]. The exact mechanism of the suppression of *H. pylori*-specific or of allergen-specific effector T-cells by *H. pylori*-induced regulatory T-cells is not well understood; the secretion of suppressive Treg cytokines such as IL-10 and TGF- β , but also contact-dependent mechanisms of effector T-cell suppression can be envisioned. It is of interest to note in this context that *H. pylori* infection seems to be inversely correlated not only with asthma, but also with chronic inflammatory conditions of the small and large intestine. Several epidemiological studies have pointed out this

association, looking at both ulcerative colitis and Crohn's disease patient populations [45,46]. Recent experimental evidence supports this concept; in a model of *Salmonella typhimurium*-induced colitis, concomitant *H. pylori* infection alleviated disease symptoms [47]. Protection against this form of colitis was attributed to reduced Th17 responses in the gut under conditions of co-infection, and coincided with the production of immunomodulatory (Treg-derived?) IL-10 in the mesenteric lymph nodes of the co-infected mice [47].

Inducible Tregs (iTregs; in contrast to "natural", thymus-derived nTregs) differentiate in the periphery as a result of their priming by dendritic cells (DCs) with "tolerogenic" rather than immunogenic properties [48]. Tolerogenic DCs convert naive T-cells into FoxP3⁺ Tregs through antigen presentation in the absence of co-stimulatory signals or cytokines, either alone or in combination with the production of soluble and membrane-bound tolerogenic factors such as IL-10, TGF β , retinoic acid, and programmed death ligands [48,49]. Tolerogenic properties have been attributed to immature DCs that have taken up antigen, but have not simultaneously been exposed to TLR or NLR ligands; such DCs are believed to acquire a semi-mature state characterized by high levels of MHCII, but low or no expression of co-stimulatory molecules or pro-inflammatory cytokines [48,49]. It is tempting to speculate based on the characteristic paucity of TLR and NLR ligands that is a hallmark of *H. pylori* that the pathogen has evolved precisely to avoid DC maturation, and to thereby promote Treg rather than T-effector responses. Experimental evidence for such Treg skewing comes from *in vitro* experiments with bone-marrow-derived DCs, which showed that *H. pylori*-experienced DCs appear to preferentially prime Treg over Th17 responses [50] and fail to produce pro-inflammatory cytokines [51]. The model of a predominantly tolerogenic role of DCs in the context of *H. pylori* infection is further corroborated by the demonstration that vaccine-induced protective immunity can be improved by systemic DC depletion [36] and by our finding that the lungs of neonatally infected mice subjected to experimental asthma induction are populated by semi-mature (and presumably tolerogenic) DCs [40].

B-cells exhibit immunoregulatory rather than protective properties in the context of *H. pylori* infection

H. pylori-infected patients are characterized by high serum titers to *H. pylori* antigens, and serology is routinely used to diagnose the infection. Being a mucosal pathogen, *H. pylori* was initially suspected of triggering potentially protective mucosal IgA responses. However, early studies with mice lacking B-cells (μ MT^{-/-} mice) or antibodies (IgA^{-/-}) revealed that vaccine-induced protective immunity develops equally well in wild type and

B-cell or antibody-deficient mice [24,52,53]. The results challenged the dogma that extracellular mucosal pathogens are controlled by antibodies and was confirmed again in a recent study introducing a novel and very effective new adjuvant derived from mycobacterial cell walls [36]. Subsequent work by Lycke and colleagues demonstrated that antibodies are not only dispensable for *H. pylori* clearance, but may even have detrimental effects on the human host [52]. $\mu\text{MT}^{-/-}$ mice lacking B-

cells altogether spontaneously reduce *H. pylori* burdens, but develop accelerated and aggravated gastritis [52]. Gene-targeting of the IL-10 and IgA-encoding genes synergizes to clear the infection, i.e. $\text{IL-10}^{-/-}/\text{IgA}^{-/-}$ mice clear *H. pylori* infections better than $\text{IL-10}^{-/-}$ mice, both under conditions of experimental infection of naive as well as previously vaccinated mice [54]. The combined results show rather unequivocally that B-cells impair rather than promote immunity to *H. pylori*.

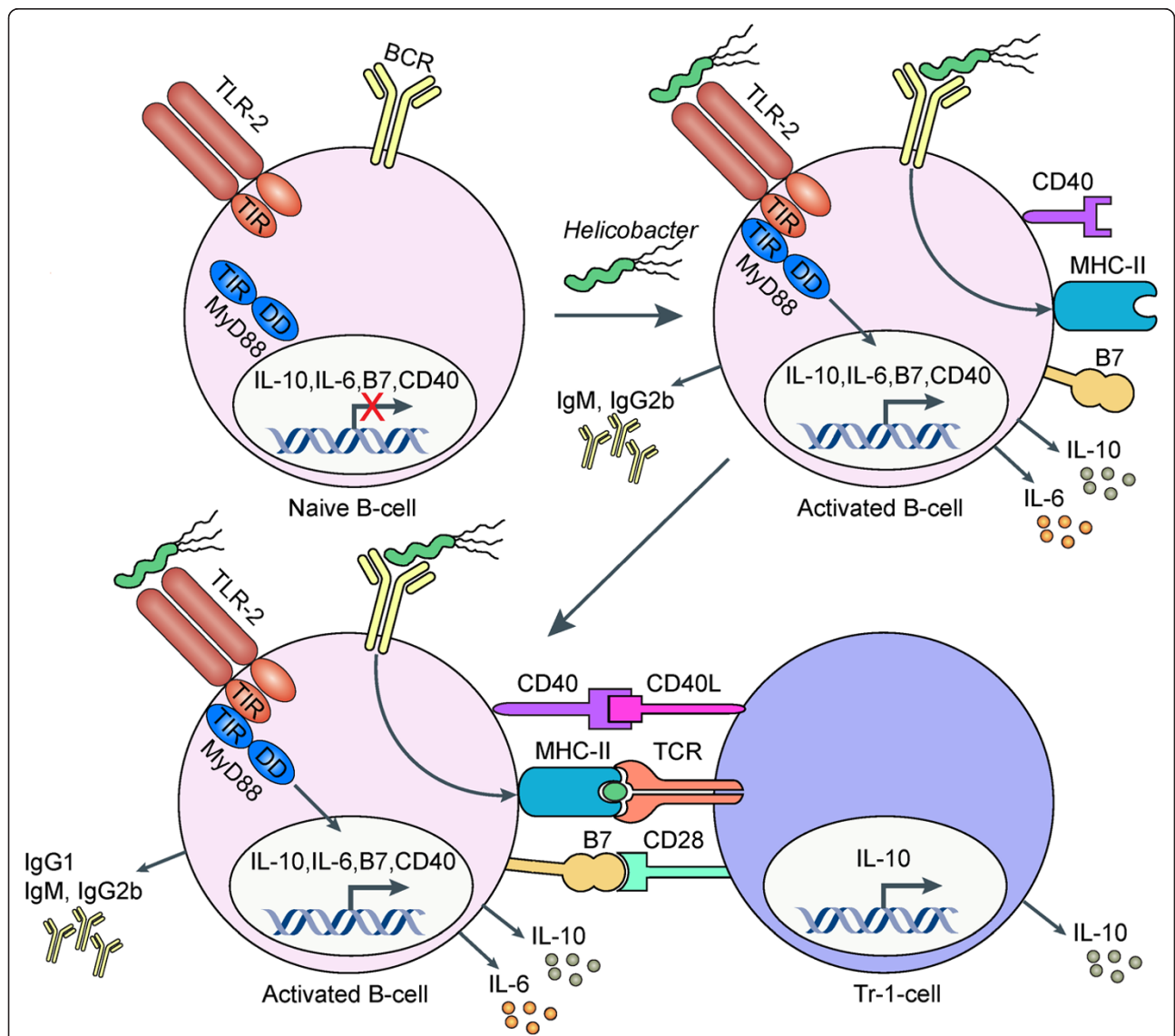


Figure 3 TLR-2-activated B-cells suppress *Helicobacter*-induced preneoplastic gastric immunopathology by inducing T regulatory-1 cells. Schematic representation of the events occurring in the course of *Helicobacter*-specific activation of B-cells at the site of infection (i.e. the gastric mucosa) and/or in the draining mesenteric lymph nodes. *Helicobacter* TLR-2 ligands activate B-cells in a MyD88-dependent manner, which leads to the expression and surface exposure of CD80, CD86 (together referred to as B7 molecules), the increased expression of CD40, and the secretion of IL-10, IL-6 and moderate amounts of TNF- α (the latter is not shown here) as well as antibodies of the IgM and IgG2b subclasses. The interaction of activated B-cells and naive T-cells induces T-cellular IL-10 expression and suppressive activity in a manner dependent on a direct interaction between both cell types via CD40/CD40L, B7/CD28 and MHCII/TCR. IL-10 secreting T-cells are essential players in the prevention of excessive *Helicobacter*-associated immunopathology. Adapted from [11].

We have recently attempted to elucidate the signaling pathways that play a role in B-cellular immunosuppression in the context of *H. pylori* infection. Interestingly, we found that B-cells exposed to *Helicobacter* extract produced large amounts of the regulatory cytokine IL-10 [11]. B-cell recognition of *Helicobacter* and the associated IL-10 production was entirely dependent on TLR2 and Myd88, as B-cells from the respective knock-out mice did not produce IL-10. *Helicobacter*-exposed B-cells further acquired the ability to efficiently induce IL-10 production in co-cultured, naive CD4⁺ T-cells, thereby converting the T-cells to T-regulatory-1 (Tr1)-like cells with suppressive activity. *In vivo* studies using conditional knock-out mouse strains revealed that IL-10 production by T-cells, but not B-cells, was essential for the suppression of excessive gastric immunopathology. On the other hand, B-cells lacking Myd88 or TLR2 -in contrast to wild type B-cells- were incapable of preventing the characteristic infection-associated immunopathology of Myd88^{-/-} or IL-10^{-/-} strains when adoptively co-transferred with Tr1 cells [11]. Taken together, the results (summarized in Figure 3) suggest that the B-cell/Tr-1 cell axis is essential for balancing the control of *Helicobacter* infection with the prevention of excessive T-cell-driven gastric immunopathology.

Conclusions

H. pylori has co-evolved with its human host for at least 30,000 years [55]. In contrast to most other bacterial pathogens, which temporarily cause virulent disease but are then rapidly cleared upon the onset of a pathogen-specific adaptive immune response, *H. pylori* persists in its host for decades, if not for life. This extraordinary ability to thrive in the face of a robust and vigorous local and systemic immune response is due to elaborate evolutionary adaptations of *H. pylori* that allow the bacteria to not only escape detection by pattern recognition receptors on innate immune cells, but to also evade adaptive immunity. The immunomodulatory properties of the pathogen reprogram the immune system towards immunological tolerance and assist the bacteria in establishing persistent infection. A byproduct of *H. pylori*-specific immunomodulation and immune tolerance is evident in Western societies that have largely eliminated the infection due to reduced transmission rates, frequent use of antibiotics in childhood and better sanitation. In these populations, the rates of asthma, allergies and other chronic inflammatory and auto-immune diseases have reached epidemic proportions; the inverse correlation of the incidence of such diseases and *H. pylori* infection rates is striking and deserves further investigation. A better understanding of the signaling pathways and molecular players targeted by *H. pylori* to manipulate the host immune response and establish and maintain persistence will be instrumental for improving rational *H. pylori* vaccine design and possibly for

exploiting *H. pylori*'s protective properties for asthma and allergy prevention and treatment.

Acknowledgements

Work in the laboratory of A.M. is supported by the University of Zurich Research Priority Program in Systems Biology, the Swiss National Science Foundation and the Swiss and Zurich Cantonal Cancer Leagues.

Author details

¹Institute of Molecular Cancer Research, University of Zürich, Zürich, Switzerland. ²Sir William Dunn School of Pathology, University of Oxford, Oxford, UK.

Authors' contributions

AM wrote the manuscript; ICA and MO contributed the figures. All authors have read and approved the final manuscript.

Competing interests

The authors declare that they have no competing interests.

Received: 28 July 2011 Accepted: 1 November 2011

Published: 1 November 2011

References

1. Ishii KJ, Koyama S, Nakagawa A, Coban C, Akira S: Host innate immune receptors and beyond: making sense of microbial infections. *Cell Host Microbe* 2008, **3**:352-363.
2. Brodsky IE, Monack D: NLR-mediated control of inflammasome assembly in the host response against bacterial pathogens. *Semin Immunol* 2009, **21**:199-207.
3. Schmausser B, Andrusis M, Endrich S, Lee SK, Josenhans C, Muller-Hermelink HK, Eck M: Expression and subcellular distribution of toll-like receptors TLR4, TLR5 and TLR9 on the gastric epithelium in *Helicobacter pylori* infection. *Clin Exp Immunol* 2004, **136**:521-526.
4. Ishihara S, Rumi MA, Kadowaki Y, Ortega-Cava CF, Yuki T, Yoshino N, Miyaoka Y, Kazumori H, Ishimura N, Amano Y, Kinoshita Y: Essential role of MD-2 in TLR4-dependent signaling during *Helicobacter pylori*-associated gastritis. *J Immunol* 2004, **173**:1406-1416.
5. Gewirtz AT, Yu Y, Krishna US, Israel DA, Lyons SL, Peek RM Jr: *Helicobacter pylori* flagellin evades toll-like receptor 5-mediated innate immunity. *J Infect Dis* 2004, **189**:1914-1920.
6. Andersen-Nissen E, Smith KD, Strobe KL, Barrett SL, Cookson BT, Logan SM, Aderem A: Evasion of Toll-like receptor 5 by flagellated bacteria. *Proc Natl Acad Sci USA* 2005, **102**:9247-9252.
7. Moran AP, Lindner B, Walsh EJ: Structural characterization of the lipid A component of *Helicobacter pylori* rough- and smooth-form lipopolysaccharides. *J Bacteriol* 1997, **179**:6453-6463.
8. Smith MF Jr, Mitchell A, Li G, Ding S, Fitzmaurice AM, Ryan K, Crowe S, Goldberg JB: Toll-like receptor (TLR) 2 and TLR5, but not TLR4, are required for *Helicobacter pylori*-induced NF-kappa B activation and chemokine expression by epithelial cells. *J Biol Chem* 2003, **278**:32552-32560.
9. Chaouche-Drider N, Kaparakis M, Karrar A, Fernandez MI, Carneiro LA, Viala J, Boneca IG, Moran AP, Philpott DJ, Ferrero RL: A commensal *Helicobacter* sp. of the rodent intestinal flora activates TLR2 and NOD1 responses in epithelial cells. *PLoS One* 2009, **4**:e5396.
10. Rad R, Ballhorn W, Volland P, Eisenacher K, Mages J, Rad L, Ferstl R, Lang R, Wagner H, Schmid RM, et al: Extracellular and intracellular pattern recognition receptors cooperate in the recognition of *Helicobacter pylori*. *Gastroenterology* 2009, **136**:2247-2257.
11. Sayi A, Kohler E, Toller IM, Flavell RA, Muller W, Roers A, Muller A: TLR-2-activated B cells suppress *Helicobacter*-induced preneoplastic gastric immunopathology by inducing T regulatory-1 cells. *J Immunol* 2011, **186**:878-890.
12. Gringhuis SI, den Dunnen J, Litjens M, van der Vlist M, Geijtenbeek TB: Carbohydrate-specific signaling through the DC-SIGN signalosome tailors immunity to *Mycobacterium tuberculosis*, HIV-1 and *Helicobacter pylori*. *Nat Immunol* 2009, **10**:1081-1088.
13. Chamailard M, Hashimoto M, Horie Y, Masumoto J, Qiu S, Saab L, Ogura Y, Kawasaki A, Fukase K, Kusumoto S, et al: An essential role for NOD1 in

- host recognition of bacterial peptidoglycan containing diaminopimelic acid. *Nat Immunol* 2003, **4**:702-707.
14. Girardin SE, Boneca IG, Carneiro LA, Antignac A, Jehanno M, Viala J, Tedin K, Taha MK, Labigne A, Zahringer U, et al: **Nod1 detects a unique muropeptide from gram-negative bacterial peptidoglycan.** *Science* 2003, **300**:1584-1587.
 15. Kim YG, Park JH, Shaw MH, Franchi L, Inohara N, Nunez G: **The cytosolic sensors Nod1 and Nod2 are critical for bacterial recognition and host defense after exposure to Toll-like receptor ligands.** *Immunity* 2008, **28**:246-257.
 16. Viala J, Chaput C, Boneca IG, Cardona A, Girardin SE, Moran AP, Athman R, Memet S, Huerre MR, Coyle AJ, et al: **Nod1 responds to peptidoglycan delivered by the Helicobacter pylori cag pathogenicity island.** *Nat Immunol* 2004, **5**:1166-1174.
 17. Allison CC, Kufer TA, Kremmer E, Kaparakis M, Ferrero RL: **Helicobacter pylori induces MAPK phosphorylation and AP-1 activation via a NOD1-dependent mechanism.** *J Immunol* 2009, **183**:8099-8109.
 18. Grubman A, Kaparakis M, Viala J, Allison C, Badea L, Karrar A, Boneca IG, Le Bourhis L, Reeve S, Smith IA, et al: **The innate immune molecule, NOD1, regulates direct killing of Helicobacter pylori by antimicrobial peptides.** *Cell Microbiol* 2012, **12**:626-639.
 19. Hutton ML, Kaparakis-Liaskos M, Turner L, Cardona A, Kwok T, Ferrero RL: **Helicobacter pylori exploits cholesterol-rich microdomains for induction of NF-kappaB-dependent responses and peptidoglycan delivery in epithelial cells.** *Infect Immun* 2010, **78**:4523-4531.
 20. Kaparakis M, Turnbull L, Carneiro L, Firth S, Coleman HA, Parkinson HC, Le Bourhis L, Karrar A, Viala J, Mak J, et al: **Bacterial membrane vesicles deliver peptidoglycan to NOD1 in epithelial cells.** *Cell Microbiol* 2012, **12**:372-385.
 21. Watanabe T, Asano N, Fichtner-Feigl S, Gorelick PL, Tsuji Y, Matsumoto Y, Chiba T, Fuss IJ, Kitani A, Strober W: **NOD1 contributes to mouse host defense against Helicobacter pylori via induction of type I IFN and activation of the ISGF3 signaling pathway.** *J Clin Invest* 2012, **122**:1645-1662.
 22. Liu W, Yan M, Liu Y, Wang R, Li C, Deng C, Singh A, Coleman WG Jr, Rodgers GP: **Olfactomedin 4 down-regulates innate immunity against Helicobacter pylori infection.** *Proc Natl Acad Sci USA* 2010, **107**:11056-11061.
 23. Akhiani AA, Pappo J, Kabok Z, Schon K, Gao W, Franzen LE, Lycke N: **Protection against Helicobacter pylori infection following immunization is IL-12-dependent and mediated by Th1 cells.** *J Immunol* 2002, **169**:6977-6984.
 24. Ermak TH, Giannasca PJ, Nichols R, Myers GA, Nedrud J, Weltzin R, Lee CK, Kleanthous H, Monath TP: **Immunization of mice with urease vaccine affords protection against Helicobacter pylori infection in the absence of antibodies and is mediated by MHC class II-restricted responses.** *J Exp Med* 1998, **188**:2277-2288.
 25. Sayi A, Kohler E, Hitzler I, Arnold I, Schwendener R, Rehrauer H, Muller A: **The CD4+ T cell-mediated IFN-gamma response to Helicobacter infection is essential for clearance and determines gastric cancer risk.** *J Immunol* 2009, **182**:7085-7101.
 26. Linz B, Balloux F, Moodley Y, Manica A, Liu H, Roumagnac P, Falush D, Stamer C, Prugnolle F, van der Merwe SW, et al: **An African origin for the intimate association between humans and Helicobacter pylori.** *Nature* 2007, **445**:915-918.
 27. Gebert B, Fischer W, Weiss E, Hoffmann R, Haas R: **Helicobacter pylori vacuolating cytotoxin inhibits T lymphocyte activation.** *Science* 2003, **301**:1099-1102.
 28. Boncristiano M, Paccani SR, Barone S, Olivieri C, Patrussi L, Ilver D, Amedei A, D'Elia MM, Telford JL, Baldari CT: **The Helicobacter pylori vacuolating toxin inhibits T cell activation by two independent mechanisms.** *J Exp Med* 2003, **198**:1887-1897.
 29. Sundrud MS, Torres VJ, Unutmaz D, Cover TL: **Inhibition of primary human T cell proliferation by Helicobacter pylori vacuolating toxin (VacA) is independent of VacA effects on IL-2 secretion.** *Proc Natl Acad Sci USA* 2004, **101**:7727-7732.
 30. Cover TL, Blaser MJ: **Purification and characterization of the vacuolating toxin from Helicobacter pylori.** *J Biol Chem* 1992, **267**:10570-10575.
 31. Sewald X, Gebert-Vogl B, Prassl S, Barwig I, Weiss E, Fabbri M, Osicka R, Schiemann M, Busch DH, Semmrich M, et al: **Integrin subunit CD18 Is the T-lymphocyte receptor for the Helicobacter pylori vacuolating cytotoxin.** *Cell Host Microbe* 2008, **3**:20-29.
 32. Sewald X, Jimenez-Soto L, Haas R: **PKC-dependent endocytosis of the Helicobacter pylori vacuolating cytotoxin in primary T lymphocytes.** *Cell Microbiol* 2008, **13**:482-496.
 33. Gerhard M, Schmees C, Voland P, Endres N, Sander M, Reindl W, Rad R, Oelsner M, Decker T, Mempel M, et al: **A secreted low-molecular-weight protein from Helicobacter pylori induces cell-cycle arrest of T cells.** *Gastroenterology* 2005, **128**:1327-1339.
 34. Schmees C, Prinz C, Treptau T, Rad R, Hengst L, Voland P, Bauer S, Brenner L, Schmid RM, Gerhard M: **Inhibition of T-cell proliferation by Helicobacter pylori gamma-glutamyl transpeptidase.** *Gastroenterology* 2007, **132**:1820-1833.
 35. Arnold IC, Lee JY, Amieva MR, Roers A, Flavell RA, Sparwasser T, Muller A: **Tolerance rather than immunity protects from Helicobacter pylori-induced gastric preneoplasia.** *Gastroenterology* 2011, **140**:199-209.
 36. Hitzler I, Oertli M, Becher B, Agger EM, Muller A: **Dendritic Cells Prevent Rather Than Promote Immunity Conferred by a Helicobacter Vaccine Using a Mycobacterial Adjuvant.** *Gastroenterology* 2011, **141**:186-196.
 37. Lundgren A, Stromberg E, Sjoling A, Lindholm C, Enarsson K, Edebo A, Johnsson E, Suri-Payer E, Larsson P, Rudin A, et al: **Mucosal FOXP3-expressing CD4+ CD25high regulatory T cells in Helicobacter pylori-infected patients.** *Infect Immun* 2005, **73**:523-531.
 38. Lundgren A, Suri-Payer E, Enarsson K, Svennerholm AM, Lundin BS: **Helicobacter pylori-specific CD4+ CD25high regulatory T cells suppress memory T-cell responses to H. pylori in infected individuals.** *Infect Immun* 2003, **71**:1755-1762.
 39. Harris PR, Wright SW, Serrano C, Riera F, Duarte I, Torres J, Pena A, Rollan A, Viviani P, Guiraldes E, et al: **Helicobacter pylori gastritis in children is associated with a regulatory T-cell response.** *Gastroenterology* 2008, **134**:491-499.
 40. Arnold IC, Dehzad N, Reuter S, Martin H, Becher B, Taube C, Muller A: **Helicobacter pylori infection prevents allergic asthma in mouse models through the induction of regulatory T cells.** *J Clin Invest* 2011, **121**:3088-3093.
 41. Blaser MJ, Chen Y, Reibman J: **Does Helicobacter pylori protect against asthma and allergy?** *Gut* 2008, **57**:561-567.
 42. Chen Y, Blaser MJ: **Inverse associations of Helicobacter pylori with asthma and allergy.** *Arch Intern Med* 2007, **167**:821-827.
 43. Chen Y, Blaser MJ: **Helicobacter pylori colonization is inversely associated with childhood asthma.** *J Infect Dis* 2008, **198**:553-560.
 44. Reibman J, Marmor M, Filner J, Fernandez-Beros ME, Rogers L, Perez-Perez GI, Blaser MJ: **Asthma is inversely associated with Helicobacter pylori status in an urban population.** *PLoS One* 2008, **3**:e4060.
 45. Pronai L, Schandl L, Orosz Z, Magyar P, Tulassay Z: **Lower prevalence of Helicobacter pylori infection in patients with inflammatory bowel disease but not with chronic obstructive pulmonary disease - antibiotic use in the history does not play a significant role.** *Helicobacter* 2004, **9**:278-283.
 46. Sladek M, Jedynak-Wasowicz U, Wedrychowicz A, Kowalska-Duplaga K, Pieczarkowski S, Fyderek K: **[The low prevalence of Helicobacter pylori gastritis in newly diagnosed inflammatory bowel disease children and adolescent].** *Przegl Lek* 2007, **64**(Suppl 3):65-67.
 47. Higgins PD, Johnson LA, Luther J, Zhang M, Sauder ME, Rogers L, Kao JY: **Prior Helicobacter pylori infection ameliorates Salmonella typhimurium-induced colitis: Mucosal crosstalk between stomach and distal intestine.** *Inflamm Bowel Dis* 2008, **14**:1398-1408.
 48. Maldonado RA, von Andrian UH: **How tolerogenic dendritic cells induce regulatory T cells.** *Adv Immunol* 2010, **108**:111-165.
 49. Kretschmer K, Apostolou I, Hawiger D, Khazaie K, Nussenzweig MC, von Boehmer H: **Inducing and expanding regulatory T cell populations by foreign antigen.** *Nat Immunol* 2005, **6**:1219-1227.
 50. Kao JY, Zhang M, Miller MJ, Mills JC, Wang B, Liu M, Eaton KA, Zou W, Berndt BE, Cole TS, et al: **Helicobacter pylori immune escape is mediated by dendritic cell-induced Treg skewing and Th17 suppression in mice.** *Gastroenterology* 2010, **138**:1046-1054.
 51. Wang YH, Gorvel JP, Chu YT, Wu JJ, Lei HY: **Helicobacter pylori impairs murine dendritic cell responses to infection.** *PLoS One* 2010, **5**:e10844.
 52. Akhiani AA, Schon K, Franzen LE, Pappo J, Lycke N: **Helicobacter pylori-specific antibodies impair the development of gastritis, facilitate bacterial colonization, and counteract resistance against infection.** *J Immunol* 2004, **172**:5024-5033.

53. Garhart CA, Nedrud JG, Heinzel FP, Sigmund NE, Czinn SJ: **Vaccine-induced protection against *Helicobacter pylori* in mice lacking both antibodies and interleukin-4.** *Infect Immun* 2003, **71**:3628-3633.
54. Akhiani AA, Stensson A, Schon K, Lycke NY: **IgA antibodies impair resistance against *Helicobacter pylori* infection: studies on immune evasion in IL-10-deficient mice.** *J Immunol* 2005, **174**:8144-8153.
55. Moodley Y, Linz B, Yamaoka Y, Windsor HM, Breurec S, Wu JY, Maady A, Bernhoft S, Thiberge JM, Phuanukoonnon S, *et al*: **The peopling of the Pacific from a bacterial perspective.** *Science* 2009, **323**:527-530.
56. Watanabe T, Asano N, Kitani A, Fuss IJ, Chiba T, Strober W: **NOD1-Mediated Mucosal Host Defense against *Helicobacter pylori*.** *Int J Inflam* 2010:476482.

doi:10.1186/1478-811X-9-25

Cite this article as: Müller *et al.*: *H. pylori* exploits and manipulates innate and adaptive immune cell signaling pathways to establish persistent infection. *Cell Communication and Signaling* 2011 **9**:25.

**Submit your next manuscript to BioMed Central
and take full advantage of:**

- Convenient online submission
- Thorough peer review
- No space constraints or color figure charges
- Immediate publication on acceptance
- Inclusion in PubMed, CAS, Scopus and Google Scholar
- Research which is freely available for redistribution

Submit your manuscript at
www.biomedcentral.com/submit

