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ER β Binds N-CoR in the Presence of Estrogens via an LXXLL-like Motif in the N-CoR C-terminus

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Abstract

Nuclear receptors (NRs) usually bind the corepressors N-CoR and SMRT in the absence of ligand or in the presence of antagonists. Agonist binding leads to corepressor release and recruitment of coactivators. Here, we report that estrogen receptor β (ER β) binds N-CoR and SMRT in the presence of agonists, but not antagonists, *in vitro* and *in vivo*. This ligand preference differs from that of ER α interactions with corepressors, which are inhibited by estradiol, and resembles that of ER β interactions with coactivators. ER β /N-CoR interactions involve ER β AF-2, which also mediates coactivator recognition. Moreover, ER β recognizes a sequence (PLTIRML) in the N-CoR C-terminus that resembles coactivator LXXLL motifs. Inhibition of histone deacetylase activity specifically potentiates ER β LBD activity, suggesting that corepressors restrict the activity of AF-2. We conclude that the ER isoforms show completely distinct modes of interaction with a physiologically important corepressor and discuss our results in terms of ER isoform specificity *in vivo*.

Background

The nuclear receptor (NR) family comprises 48 structurally related transcription factors, many of which require their cognate ligand for activity [1–3]. The NRs regulate transcription by binding to response elements in the promoters of target genes and acting as scaffolds for the assembly of large coactivator and corepressor complexes [4]. NR coactivators include the p160s (including GRIP1/TIF-2, SRC-1 and AIB1/ACTR/pCIP). The p160s enhance transcription by binding histone acetyl-transferases such as p300/CBP and pCAF [5] and methyl-transferases such as CARM1 and PRMT [6] which, in turn, enhance transcription by modification of chromatin. Other NR coacti-

vators include TRAP220[7], which is part of a larger complex (TRAP/DRIP/SMCC/mediator) that contacts the basal transcription machinery and PGC-1[8], a cold inducible coactivator that binds CBP and SRC-1 and proteins involved in RNA processing [9]. NR corepressors include NR corepressor (N-CoR) and silencing mediator of retinoid and thyroid responsive transcription (SMRT) [4]. Both N-CoR and SMRT repress transcription, at least in part, by binding to histone de-acetylases (HDACs) either directly or indirectly through other corepressor complex components. Other known NR corepressors include RIP140 [10], Hairless [11], short heterodimer partner (SHP) [12] and DAX [13], and receptor specific

corepressors such as the estrogen receptor (ER) interacting proteins REA and HET-SAFB [14,15].

Generally, NR transcriptional activity is dictated by the balance between coactivator and corepressor recruitment, and one of the most important factors that influences this balance is the absence or presence of agonist ligands (reviewed in [4]). Unliganded NRs such as thyroid (TRs) and retinoid receptors (RARs) bind corepressors, and ligand promotes release of corepressor and subsequent binding of coactivators. The mechanism of this coregulator exchange is well understood. NRs consist of three domains, the N-terminal domain (which contains a context-specific activation function AF-1), the central DNA binding domain (DBD) and the C-terminal ligand binding domain (LBD), which contains a hormone-dependent activation function, AF-2. The unliganded LBD recognizes hydrophobic motifs, termed interaction domains (IDs), which are reiterated three times in N-CoR and twice in SMRT and conform to the consensus L/XXIIXXXL [4] (see also [16]). By contrast, the liganded LBD binds shorter hydrophobic motifs termed NR boxes that are reiterated several times within each coactivator and conform to the consensus LXXLL. The LBD utilizes a large hydrophobic cleft composed of residues along H3 and H5 to bind IDs [17], and a smaller hydrophobic cleft that is composed of residues in the upper part of H3 and H5 and H12 (and corresponds to AF-2) to bind NR boxes [18]. Thus, agonists promote coregulator exchange by promoting the packing of H12 over the lower part of the ID binding region, an event that simultaneously completes the coactivator binding surface. In other cases, however, the balance of coactivator and corepressor recruitment is regulated by direct competition for the AF-2 surface. RIP140, Hairless and DAX possess NR boxes that interact with AF-2 [11,13,19] and these corepressors act as negative regulators of the activity of the liganded NR.

The NR family contains two related ERs (ER α and ER β) that conform to the typical three domain NR structure and share extensive sequence homology in the DBD and LBD region [20,21]. Analysis of the function of the individual ERs in mouse knockout models suggests that the major proliferative effects of estrogen are mediated by ER α and not by ER β , which seems to play an inhibitory role in proliferation in some studies [22,23]. The ligand-binding properties of the ERs are different, with ER β often exhibiting stronger binding to plant-derived phytoestrogens [24]. More importantly, the ERs exhibit isoform-specific effects on gene expression. Both ERs enhance transcription from genes with classical estrogen response elements (EREs), but ER α requires less ligand to obtain maximal activation than ER β [25,26]. Likewise, both ERs suppress the activity of the TNF α promoter in response to estro-

gens, but ER β is a more potent repressor than ER α [27]. However, some of the most striking isoform-specific differences in gene regulation are observed at promoters, such as that of cyclin D1, which contain AP-1 sites or related cyclic AMP response elements (CREs). ER α enhances AP-1 activity in response to estrogens, [28,29] but ER β inhibits AP-1 activity in response to estrogens [29–31]. ER β also completely suppresses ER α activity at the cyclin D1 promoter and even suppresses the activity of an ER α mutant that is selectively superactive at AP-1 sites and CREs [29]. Finally, ER β shows a unique capacity to enhance AP-1 activity in response to selective estrogen receptor modulators (SERMs) such as raloxifene, tamoxifen and ICI 182,780/Faslodex (ICI) [30–32]. Together, these observations suggest that there are fundamental differences in the way that the ERs bind unspecified cofactors that modulate gene expression, and that some of these cofactors must play a role in differential ER activity at AP-1 sites.

Although the poorly conserved NTD region clearly plays an important role in isoform-specificity [32,33], it is also likely that there are differences in the better conserved LBD region that contribute to differential ER α and ER β activities. Phage display techniques have revealed that ER α and ER β show different preferences in LXXLL binding (reviewed in [34,35]). Moreover, some cofactors that contain LXXLL motifs show differential binding to LBDs of the ER isoforms (reviewed in [36]). SHP binds ER α preferentially [37], and represses ER α activity more strongly than that of ER β . The PGC-1 related protein PERC also binds ER α preferentially, and potentiates ER α activity more strongly than that of ER β [38]. We recently observed that ER α binds the C-terminal NR interacting regions of N-CoR and SMRT in the presence of SERMs but not estrogens [39]. In this study, we report that ER β interactions with N-CoR and SMRT are promoted by agonists and inhibited by SERMs. Thus, the ERs show completely opposite ligand preferences of interaction with corepressors. We discuss the potential significance of these different modes of ER interaction with N-CoR in terms of known isoform-specific behaviors.

Results

Agonist Dependent ER β Interactions with N-CoR and SMRT

To investigate ER β interactions with corepressors, we examined the interactions of full length ER β (amino acids 1–530) with bacterially expressed C-terminal NR interacting domain of N-CoR (amino acids 1944–2453) *in vitro* (Fig. 1A). Fig. 1B reveals, surprisingly, that ER β bound N-CoR in the absence of hormone and in the presence of agonist ligands (E2, DES) and phytoestrogens (genistein, coumestrol). Moreover, these interactions were suppressed by SERMs (ICI, raloxifene and tamoxifen). ER β

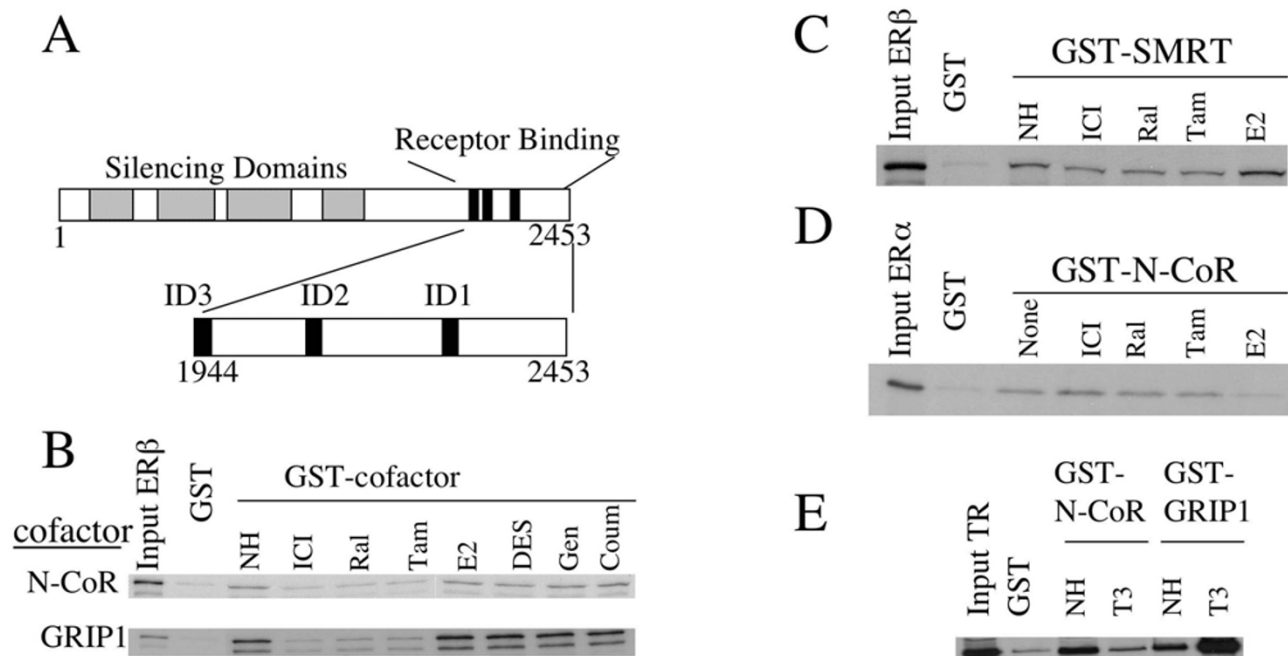


Figure 1

ER β Binds N-CoR in the Presence of Agonists. **(A)** Schematic of N-CoR primary structure. Silencing domains are indicated with hatched bars. NR binding regions (ID motifs) are illustrated with solid bars. **(B)** Radiolabeled ER β retained by GST-N-CoR (amino acids 1944–2453) or GST-GRIP1 (amino acids 563–1121) after separation by SDS-PAGE. The amounts of bound proteins are compared to 10% of the input protein used in the binding assay in this experiment, and all results shown in the paper. **(C)** Radiolabeled ER β retained by GST SMRT (amino acids 987–1491). **(D)** Radiolabeled ER α retained on GST-N-CoR beads in parallel. **(E)** Radiolabeled TR β retained on GST-N-CoR and GST-GRIP1 beads in parallel.

bound to the coactivator GRIP1 more strongly than N-CoR, but did so with an identical ligand preference. Similar agonist-dependent interactions could be observed between ER β and the alternate NR corepressor SMRT *in vitro* (Fig. 1C). Control binding experiments performed in parallel confirmed that ER α bound to N-CoR in the presence of SERMs, and not estradiol (Fig. 1D) and that TR β bound N-CoR in the absence of hormone, and was released in the presence of T3, whereas TR β only bound GRIP1 in the presence of T3 (Fig. 1E).

To examine interactions between ER β and N-CoR in mammalian cells we performed two-hybrid assays using a GAL4 DBD/N-CoR C-terminus fusion protein as bait and a VP16-ER β LBD fusion as the prey. Fig. 2 shows that the ER β -LBD bound N-CoR in the presence of agonists and phytoestrogens, but not SERMs. Control two-hybrid assays confirmed that a VP16-TR β LBD fusion protein

bound N-CoR in the absence of hormone, but not in the presence of T3. E2 dependent binding of ER β to N-CoR was dose dependent (Fig. 2B) with an EC₅₀ (0.3 nM) that resembled that of ER β binding to the GRIP1 NR box region (amino acids 610–770). Thus, ER β binds the N-CoR C-terminal NR interacting region in the presence of agonists, but not SERMs, and does so *in vitro* and in mammalian cells. Moreover, results from the two-hybrid assay indicate that the ER β LBD is sufficient to obtain estrogen-dependent interactions with N-CoR.

ER β Interactions with N-CoR are Dependent on AF-2 and require H12

Unliganded NRs usually bind ID motifs (consensus L/IXXIIXXXL) in the N-CoR C-terminus. To ask whether ER β might bind these motifs in the presence of estradiol, we examined the ability of peptides containing known NR interacting motifs to compete for the interaction (Fig. 3A).

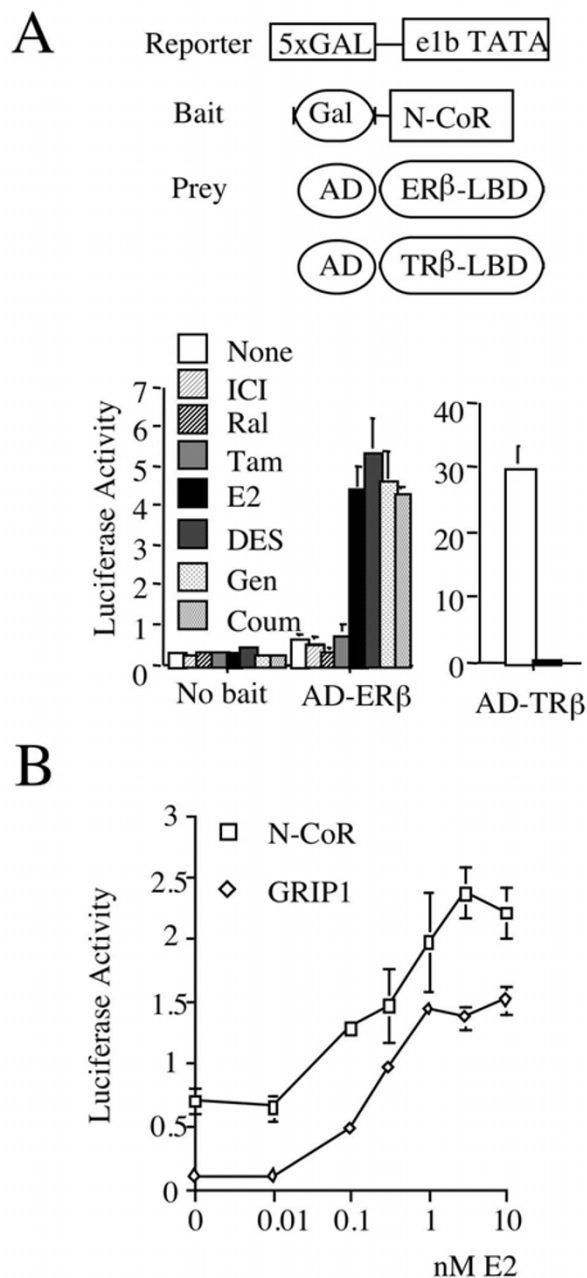


Figure 2
ER β Binds N-CoR in the Presence of Agonists in Mammalian Cells. **(A)** Two-hybrid assays. Components of the two-hybrid assay are shown in schematic at top. Results of a representative assay are shown below (with error bars representing the standard error of values determined from three separate wells). Ligand concentrations were: ICI, raloxifene, Genistein, Coumestrol; 1 μ M, Tamoxifen; 5 μ M, estradiol DES 100 nM. **(B)** Estradiol dependence of ER β interactions with N-CoR and GRIP1 fusion proteins in mammalian cells. A representative experiment is shown. Error bars represent standard deviations from four wells.

A peptide overlapping to the N-CoR ID1 motif (amino acids 2265–2291) that competes for N-CoR binding to unliganded TR and RAR [16] failed to compete for agonist-dependent ER β interactions with N-CoR. By contrast, a peptide corresponding to GRIP1 NR box 2 did compete for this interaction [40,41]. This finding suggests that agonist-bound ER β does not recognize ID motifs, and that ER β interactions with N-CoR more closely resemble those with GRIP1.

NR interactions with N-CoR are usually mediated by a hydrophobic cleft that spans residues from H3 and H5 and includes residues that lie under H12 in the liganded configuration [4,17]. These interactions are either independent of, or inhibited by, NR H12 [17,39,42]. By contrast, NR interactions with coactivators are mediated by residues from the upper part of H3–H5 and also require H12 itself [18]. Fig. 3B shows that a mutation in a conserved residue on H12 that is required for coactivator binding (E493K) abolished the interaction of ER β with both N-CoR and GRIP1. Moreover, other mutations in the upper part of the H3–H5 region that comprises the AF-2 surface (D303Y, I310R and K314A on H3; V328R and L331R on H5) abolished ER β interaction with both cofactors. Control mutations in other regions of the ER β surface left its interactions with N-CoR and GRIP1 either slightly reduced or intact (these are L301R at the base of H3, V361R in the S-bends, M379R in H8; L426R, T434R in the H10 and Y488S in the H11–H12 loop). Thus, ER β interactions with N-CoR are dependent on the AF-2 surface (including H12) and, in this regard, resemble those of ER β and GRIP1.

ER β Binds an NR Box-Like Motif in the N-CoR C-terminus

To map the region of N-CoR that interacted with ER β , we examined ER β binding to a series of rationally designed smaller fragments of the N-CoR C-terminus (Fig. 4). ER β did not bind two of these smaller fragments of N-CoR (1944–2033; 2230–2322) that contain known ID motifs (IDs 3 and 1 [16]). ER β bound weakly to two regions of N-CoR (2033–2123; 2123–2230), one of which (2033–2123) contains an ID motif (ID2), but did so in a ligand-independent fashion. However, ER β did bind to a fragment that spanned the extreme C-terminus (2322–2453) and did so in a manner that was promoted by E2 and suppressed by ICI, much like the interactions of ER β with the entire N-CoR nuclear receptor interacting region.

The interaction of ER β with the small N-CoR C-terminal fragment (amino acids 2322–2453) was stronger than that observed with the intact C-terminus (amino acids 1944–2453). This apparently improved binding is likely to be a consequence of our methodology (West *et al.* Unpublished data). In general, expression of large fragments of the N-CoR C-terminus in *E. Coli* yields a mix of

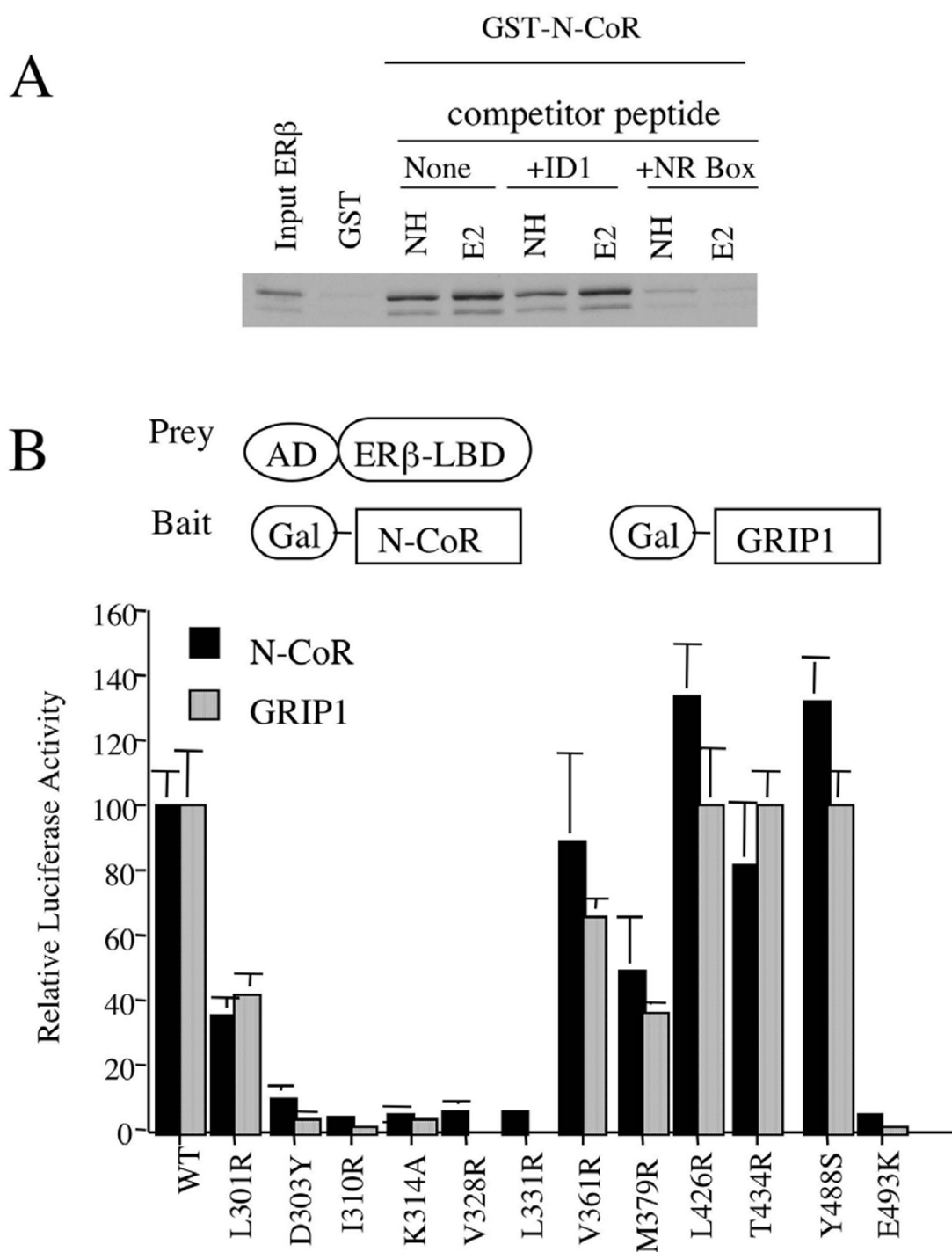


Figure 3

ERβ AF-2 is required for Corepressor and Coactivator Binding. **(A)** Peptide competition for ERβ binding to cofactors. Binding of ERβ to GST-N-CoR was assessed in the presence of 10μg of designated competitor peptide. **(B)** Binding of VPI6-ERβ LBD mutants to Gal-N-CoR and Gal-GRIP1 was analyzed. Luciferase activity obtained with Gal-fusion and VPI6-ERβ in the presence of estradiol (100 nM) over four experiments was corrected for background and set to 100%. Activity obtained with ERβ mutants was compared to this value. Standard errors are shown.

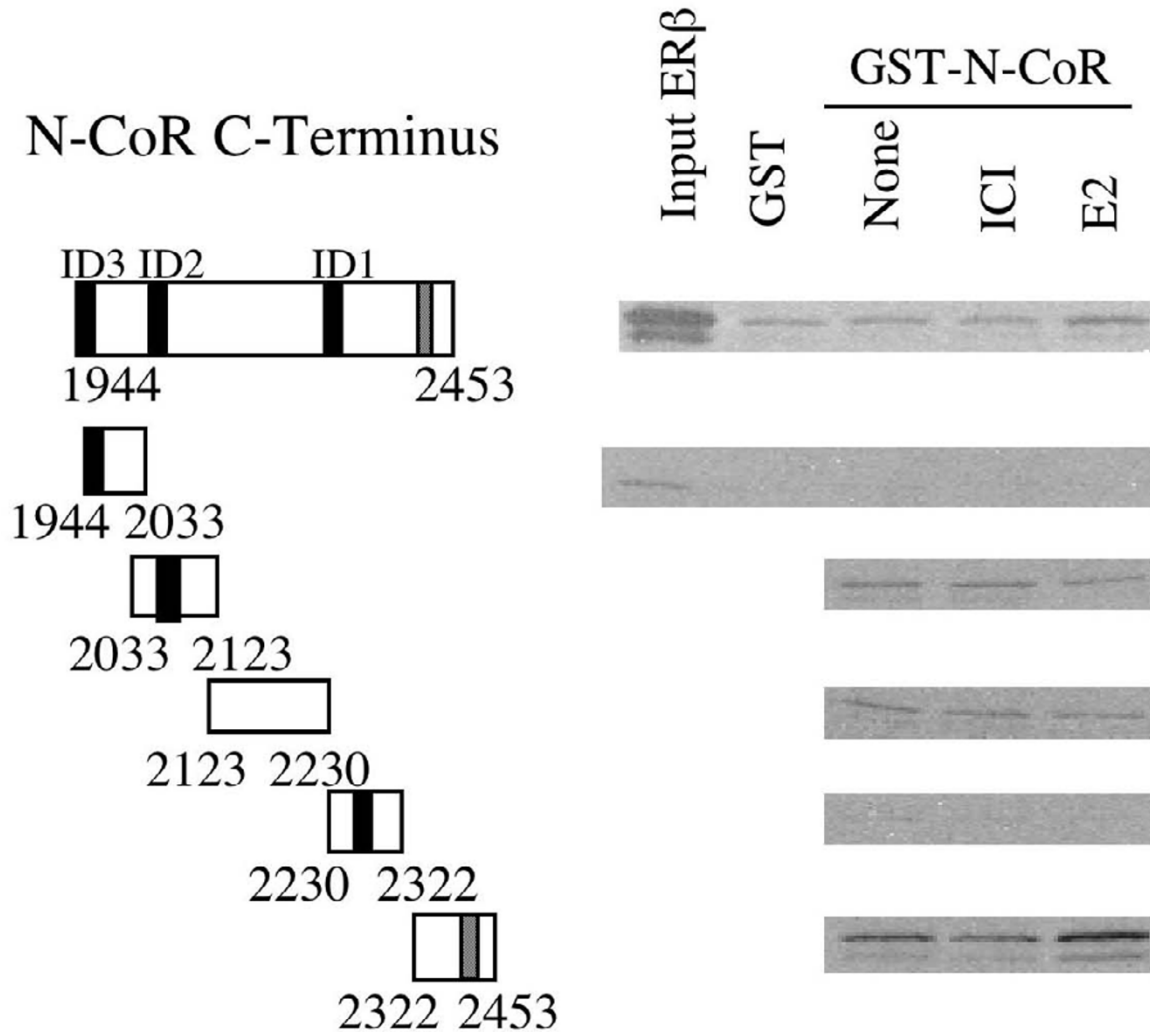


Figure 4
 ERβ binds the N-CoR C-terminus. Binding of ERβ to short bacterially produced GST-N-CoR fragments was assessed as in Fig. 1B. The approximate positions of N-CoR IDs are indicated with black bars, the putative ERβ interacting motif with a grey bar. Note that all of the binding experiments with shorter N-CoR fragments were performed in parallel, and are compared to the same input ERβ protein (10% of total) displayed alongside the 1944–2033 fragment. A binding experiment that was performed in parallel utilizing the entire GST-N-CoR C-terminus (1944–2453) is shown for comparison at top. The likely reasons that ERβ binding to the shorter N-CoR fragment (2322–2453) is stronger than that obtained with the intact N-CoR C-terminus are discussed in the text.

full length protein along with truncated products. To create the expression vectors for the smaller fragments, truncated N-CoR polypeptides that were obtained in E. Coli extracts were subjected to protein sequence analysis and cDNA fragments that coded for the major truncated products were prepared. Each of the resulting polypep-

tides was expressed very efficiently in E. Coli. The end product that was obtained after GST purification essentially consisted of a single short polypeptide as judged by Coomassie stain. Binding of ERβ to N-CoR (2322–2453) is probably very efficient for two reasons. First, equal amounts of GST fusion protein were used as baits for the

translated ER β protein in this series of experiments (3 μ g per assay). Thus, N-CoR (2322–2453) is present in molar excess over N-CoR (1944–2453). Second, as developed above, preparations of N-CoR (1944–2453) generally contain truncated products, so sequences corresponding to the extreme N-CoR C-terminus (which binds ER β) is markedly under-represented. In any case, the fact that ER β binds weakly or not at all to the three N-CoR ID motifs that mediate interactions with TRs and RARs and, instead, binds in an agonist-dependent fashion to a region in the C-terminus of N-CoR that has not previously been implicated in NR interactions indicates that ER β recognizes a novel protein sequence motif within N-CoR.

The N-CoR C-terminus contains the sequence PLTIRMLS (β -box, amino acids 2399–2406; Fig. 5). This sequence does not exactly conform to the LXXLL consensus, but contains features (underlined) that resemble the ER β H12 region (LLLEML), and artificial ER β interacting LXXLL peptides (293, PNLISLLS; D47, PLLSLLS), both of which bind to the ER β AF-2 surface [43–47]. Moreover, the presence of a proline residue amino-terminal to the hydrophobic groups is typical of so-called class II LXXLL motifs which are found in ER β interacting cofactors such as TRAP220 and RIP140[45]. Finally, the unusual C-terminal hydrophobic pair (ML) has been observed in ER α and ER β H12[43,44,48], and in RIP140 NR boxes [19].

We investigated the significance of the β -box in ER β interactions with N-CoR. As Fig. 6A shows, a synthetic β -box peptide competed for binding to N-CoR, albeit somewhat less efficiently than native GRIP1 NR box 2. Similar results were obtained in competition experiments that used GST-GRIP1 instead of GST-N-CoR (data not shown). The isolated β -box also acted as bait for a VP16-ER β fusion protein in mammalian cells, and did so with similar efficiency to other known ER β interacting peptides (Fig. 6B). Finally, mutations within the β -box (especially M2405A, L2406A) disrupted ER β interactions with N-CoR in mammalian two-hybrid assays, but did not affect TR β interactions (Fig. 6C). Thus, the β -box is sufficient to bind ER β and is essential for agonist-dependent ER β interactions with the N-CoR C-terminus.

Next, we examined whether the β -box would bind other NRs. The Gal- β -box fusion failed to recruit the ER α , TR β or RAR β LBDs in mammalian two-hybrid assays (Fig. 7A). Moreover, while the β -box and GRIP1 NR box 2 peptides both competed for ER β interactions with GRIP1, only the NR box 2 peptide competed for ER α interactions with GRIP1 (Fig. 7B). Thus, the N-CoR β -box is, at least to some degree, ER β specific. Mutation of N-CoR to obtain a β -box sequence that more closely resembled a conventional LXXLL motif (T2402L) led to enhanced hormone-dependent interactions with ER β and permitted novel

hormone-dependent interactions with ER α (Fig. 7C). Thus, some of the observed ER β specificity is probably a consequence of an unexpected ability to tolerate the absence of a leucine residue at the N-terminus of the LXXLL motif. Together, our results indicate that ER β has the potential to utilize its AF-2 surface to bind NR boxes within coactivators or an NR box-like sequence in the C-terminus of N-CoR.

A HDAC Repressor Enhances ER β Activity

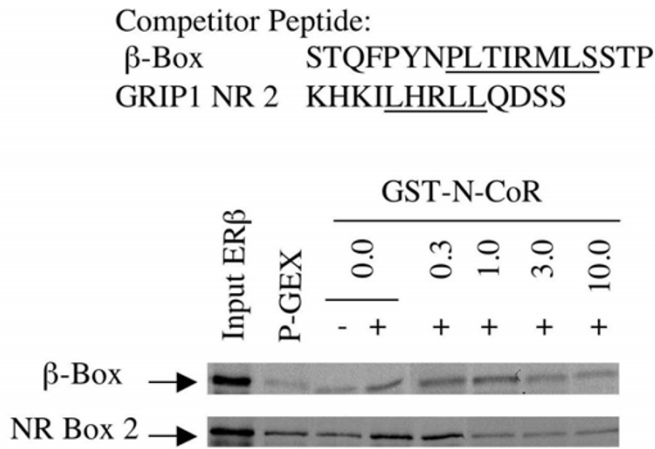
Since ER β bound N-CoR and SMRT in the presence of estrogens, we investigated the possible involvement of corepressors in the actions of agonist-bound ER β *in vivo*. To perform this experiment, we examined the effect of the HDAC inhibitor trichostatin A (TSA) on ER β activity in transiently transfected HeLa cells. Fig. 8A confirms that ER α shows stronger transcriptional activity than ER β at a simple ERE responsive reporter gene. TSA enhanced the basal activity of the ERE-TK reporter gene by about fifteen-fold in the absence of ER (see inset). However, TSA also equalized the relative transcriptional activity of both ERs. Fig. 8B shows that the isolated ER α LBD exhibited more potent transcriptional activity than the ER β LBD. However, both LBDs showed similar transcriptional activity in the presence of TSA. Thus, corepressor complex HDACs must play an unspecified role in restricting the transcriptional activity of both ER β and, in particular, the ER β -LBD. This is consistent with the notion that corepressors restrict the activity of agonist-bound ER β -LBD.

Conclusions

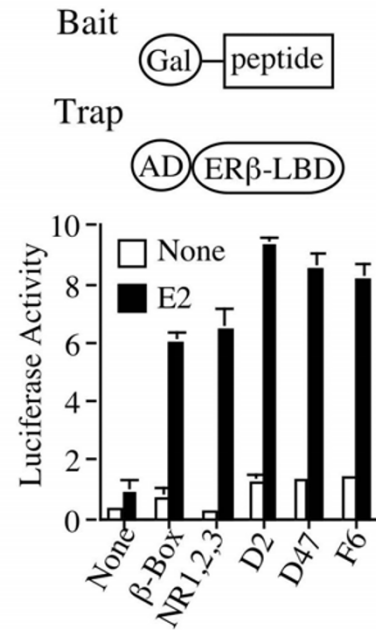
NRs generally interact with the corepressors N-CoR and SMRT either in the absence of ligand, or in the presence of receptor antagonists, and agonists promote corepressor release [4]. In this study, we demonstrated that ER β binds to N-CoR in the presence of ER agonists such as estradiol and DES and the phytoestrogens genistein and coumestrol, but not in the presence of SERMs. Moreover, this interaction is dependent upon ER β AF-2, including H12, and is competed by NR box peptides but not ID peptides. The hormone-dependent component of the ER β /N-CoR interaction maps to the extreme C-terminus of N-CoR, which has not been previously implicated in NR interactions, and requires a sequence that resembles an ER β -specific NR box (PLTIRMLS, β -box). In this regard, ER β differs from ER α , which probably binds ID motifs in a SERM-dependent fashion [49,50] and shows reduced binding to N-CoR in the presence of estradiol [39]. ER β also differs from many other NRs, which either bind N-CoR in the absence of ligand and are released in the presence of ligand or interact with N-CoR in the presence of antagonists but not agonists [4].

The fact that the mode of ER β interaction with N-CoR resembles that of NRs with coactivators [4], or with

A



B



C

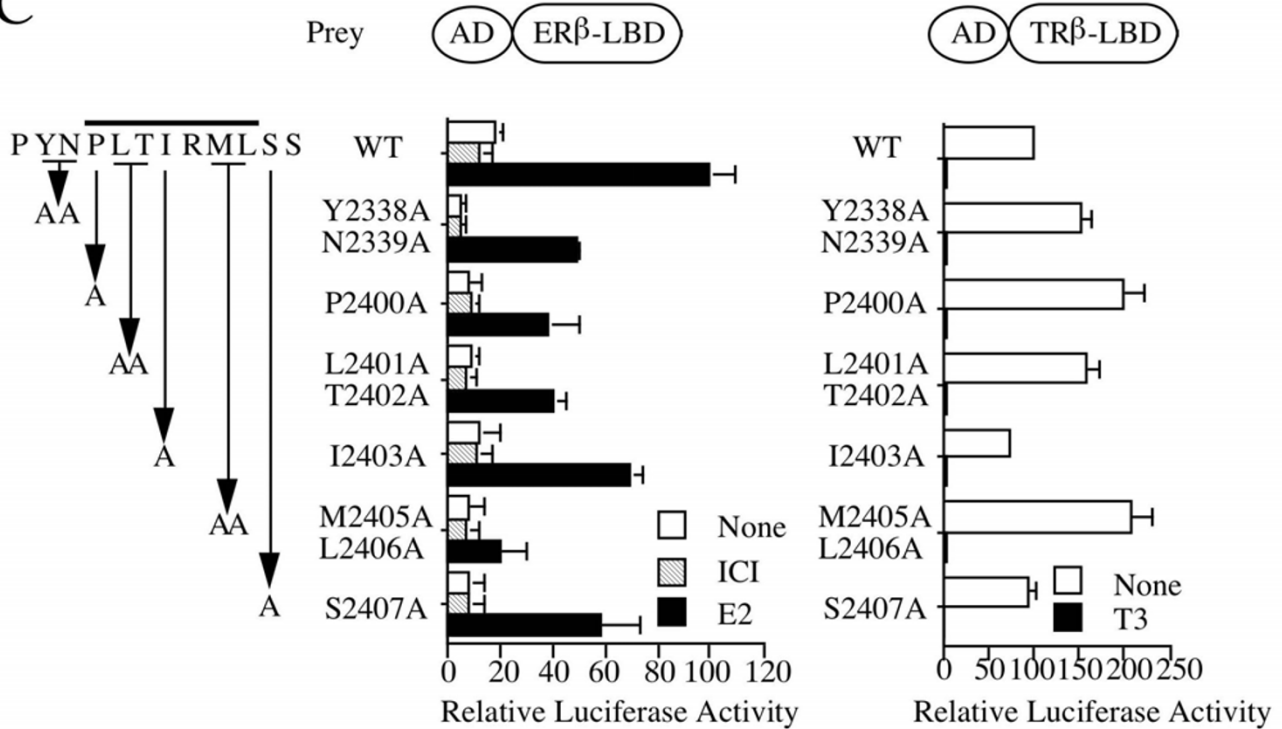
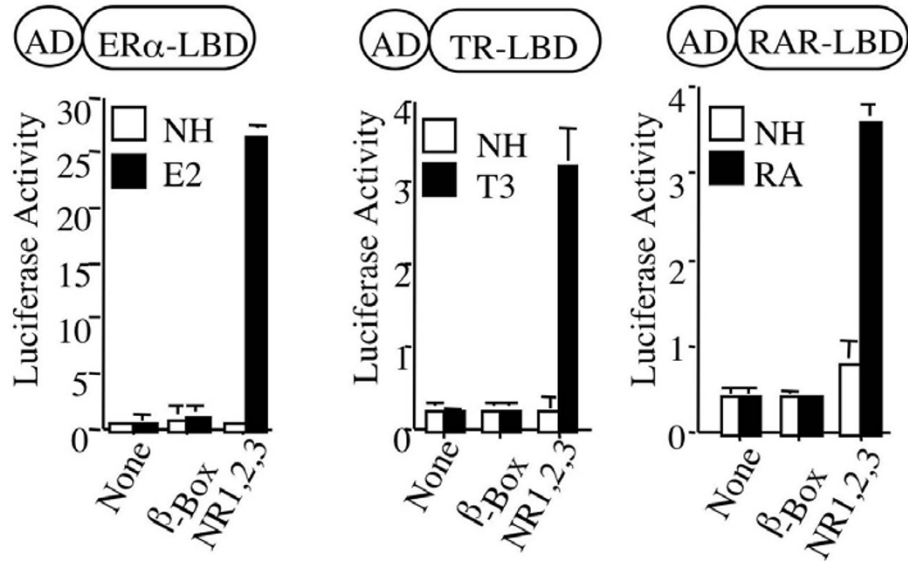


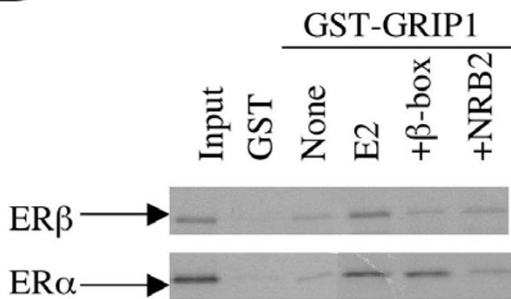
Figure 6

The N-CoR β-box binds ERβ. **(A)** Peptide competitions. Binding of ERβ to either bacterially expressed GST-N-CoR (amino acids 2239–2453) was determined in the presence of increasing doses of competitor peptide (amounts shown at top refer to μg of peptide, plus signs refer to the presence of estradiol in the assay). **(B)** Interaction of ERβ with different peptides in mammalian two-hybrid assays. A representative experiment is shown. Errors are derived from three separate wells. **(C)** Analysis of ERβ interaction with mutant Gal-N-CoR fusion proteins. Values were determined as in Fig. 3.

A



B



C

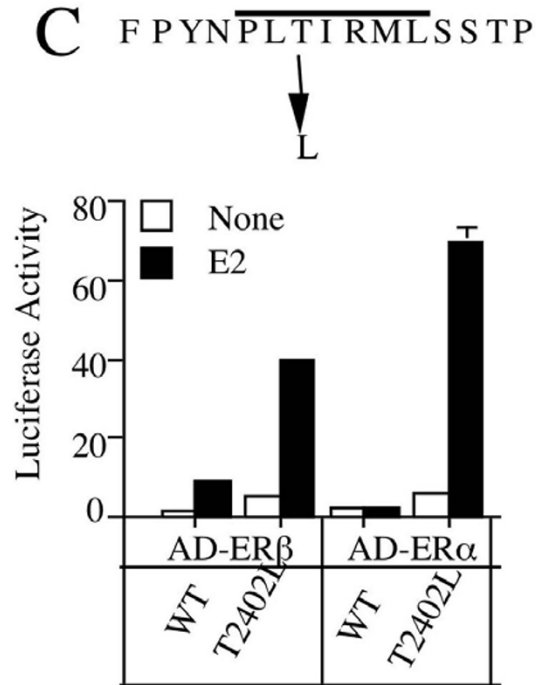
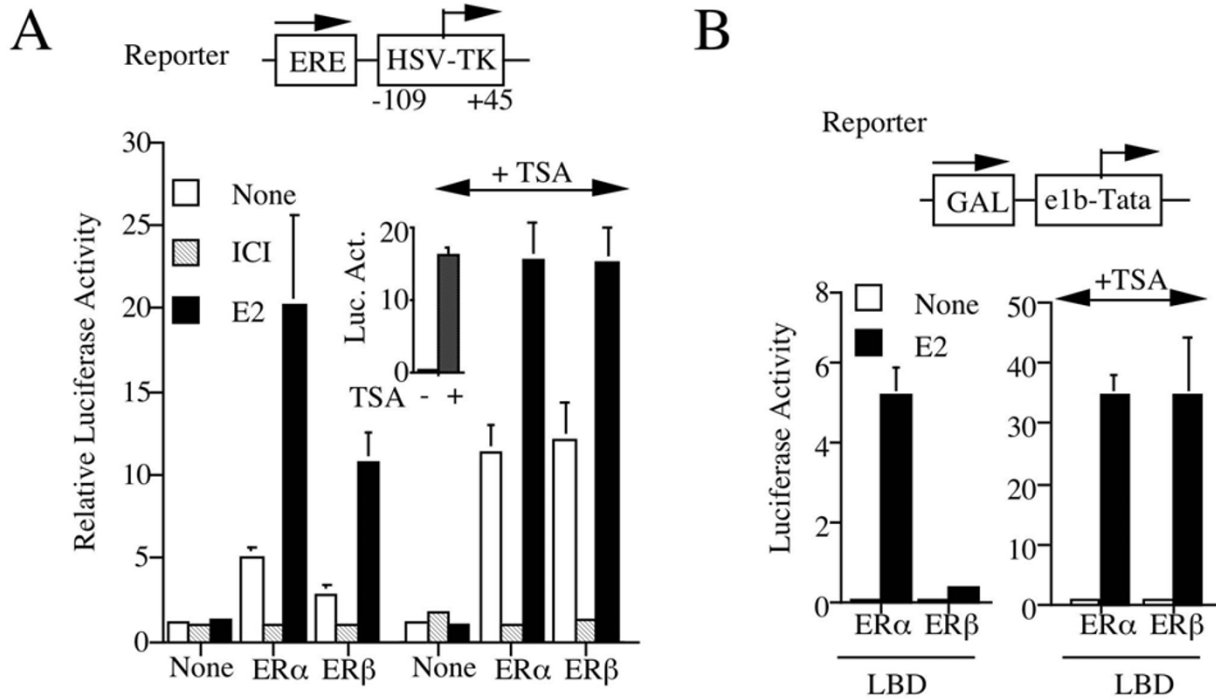


Figure 7

The β -box is an ER β -specific NR box. **(A)** Interactions of VPI6-NR fusions with the Gal- β -box fusion or a GAL-GRIP1 (amino acids 610–770) control were determined in HeLa cell transfections. A representative experiment is shown. **(B)** Peptide competitions for ER α and ER β binding to GST-GRIP1. Experiments utilized 10 μ g of competitor **(C)** Mutation of the β -box to resemble a consensus LXXLL motif. The panel shows the results of a mammalian two-hybrid assay in which binding of Gal-N-CoR or Gal-N-CoR T2402L to VPI6-ER β or VPI6-ER α was assayed.

**Figure 8**

TSA Enhances ER β -LBD activity. **(A)** Results of a HeLa cell transfection in which ER α and ER β activity was compared at a simple ERE responsive reporter gene. The value obtained in the absence of ER and ligand was set at 1 (irrespective of the presence or absence of TSA, 100 nM). Other values were calculated relative to this value. A single representative experiment is shown with errors derived from three wells. **(B)** Results of transfection in which ER α and ER β LBD activities were compared in the absence (left) or presence (right) of TSA (100 nM); note differences in scale. A single representative experiment is shown. Standard errors are derived from three wells.

transfected N-CoR also fails to affect TR or ER α activity (data not shown), despite the fact that it clearly interacts with both NRs. Nevertheless, we suspect that estrogen-dependent N-CoR binding may represent an important component of the regulation of ER β activity. As described in the Introduction, ER α and ER β must interact differentially with factors that modulate ER activity in the presence of estrogens. The finding that estrogens suppress N-CoR binding to ER α [39], but promote N-CoR binding to ER β represents the first demonstration of a corepressor that shows completely distinct modes of hormone-dependent interaction with the ER isoforms. Thus, N-CoR and SMRT and their associated HDACs are excellent candidates to explain some of the differential behaviors of the ER isoforms. Consistent with this notion, the apparent weak transcriptional activity of the ER β LBD is a consequence of corepressor HDAC activity at some level (Fig.

8). Full verification of the importance of ER β interaction with N-CoR will await demonstration that ER β recruits N-CoR and SMRT to estrogen-regulated promoters *in vivo*, and that this event is related to modulation of estrogen response.

While the ER isoforms have contrasting effects on AP-1 activity in the presence of estrogens, ER α truncations that lack the NTD and ER β both enhance AP-1 activity in the presence of SERMs [30,31]. Mutational analysis of ER α action at AP-1 sites suggests these effects may be related to N-CoR binding [39], and we have proposed that SERM action at AP-1 sites may therefore involve contacts with corepressors [31,51]. The fact that ER α and ER β show completely different ligand preferences of interaction with N-CoR suggests that the target for SERM activation at AP-1 sites may not be N-CoR in both cases. Thus, this finding

complicates our attempts to explain this unusual phenomenon. Perhaps the ER isoforms enhance AP-1 activity by superficially similar mechanisms that involve different cofactors. Alternatively, ER α and ER β action at AP-1 sites could, in fact, be mediated by SERM-dependent contacts with a common cofactor that is, as yet, unidentified. This common factor may yet prove to be N-CoR if ER β interactions with the β -box were somehow masked *in vivo*.

What features of the β -box contribute to ER β specificity? Intriguingly, the β -box contains N-terminal proline and C-terminal serine residues that extend the homology of this region to an artificial ER β -specific peptide [45]. However, the β -box also lacks the first Leu of the consensus LXXLL. A mutation (PLTIRML>PLLRML) that restores the LXXLL consensus increases ER β binding to N-CoR and permits ER α to bind to N-CoR in the presence of estrogens in mammalian two-hybrid assays. Thus, the unusual sequence of the β -box contributes to ER β specificity and ER β can tolerate the absence of a conserved N-terminal leucine in LXXLL motifs. ER β might bind to yet more cofactors that contain variant NR boxes that resemble the β -box. Other aspects of ER β interactions with corepressors warrant further study. It will be interesting to understand whether the weaker ER β interactions with other regions of N-CoR (which are insensitive to ICI) play a role in ER β binding (Fig. 4). Finally, SMRT also binds ER β in the presence of estrogens, but we have not explored the structural features that promote this interaction. Intriguingly, human SMRT contains a sequence insertion at the position of the hydrophobic pair in the N-CoR β -box, which apparently leads to deletion of both residues (N-CoR/RMLS>SMRT/RLqagvmaS) [52]. Perhaps SMRT contains a different NR interacting motif or the N-CoR NR box sequence may be more complex than we have initially reported here.

Methods

Materials

Estradiol, diethylstilbestrol (DES), tamoxifen, genistein, coumestrol, thyroid hormone, retinoic acid and trichostatin A (TSA) were purchased from Sigma (St. Louis, MO). ICI 182,780 was a gift from Alan Wakeling (Astra/Zeneca Pharmaceuticals, Macclesfield UK). Raloxifene was a gift from Stefan Nilsson (KaroBio AB, Huddinge, Sweden). Peptides were synthesized at the Biomolecular Resource Center at UCSF.

The following plasmids (pSG5-ER α , pSG5-ER β (amino acids 1–530) [31], pGEX-N-CoR and pGEX-SMRT[39], VP16-TR β and Gal-N-CoR[16], GST-N-CoR fusions[17], ERE-LUC, GK1/Gal4 responsive reporter and Gal-ER α LBD [53], pM-D2, pM-D47, pM-F6[46]) have been previously described. VP16-ER β LBD and Gal-ER β LBD contain human ER β sequences and were gifts from Dr. Dale

Leitmann (University of California, San Francisco). VP16-RAR-LBD was a gift from Dr. David Moore, Baylor, Houston, Texas. Gal-GRIP1 NR box (1,2,3) fusion (amino acids 610–770) was prepared by PCR amplification of the appropriate region of GRIP1 (primers obtained from Biomolecular Resource Center, UCSF) containing EcoRI and Sall sites, the PCR fragment was digested with these enzymes and subcloned into the pM GAL4 expression vector (Clontech Laboratories, Inc. Palo Alto, CA). VP16-ER β mutations and Gal-N-CoR mutations were prepared using standard PCR-based site directed mutagenesis (Quickchange Kit, Stratagene, La Jolla, CA) and confirmed by sequencing. The GAL4- β box fusion was prepared by synthesizing oligonucleotides corresponding to the β -box sequence with engineered EcoRI and Sall restriction sites. Annealed and phosphorylated double stranded oligonucleotide was subcloned into the appropriate sites in the PM vector.

Bacterial Protein Expression and GST Pulldown Assays

GST-fusions were expressed in E. Coli BL21 [28]. Cultures were grown to OD600 1.5 at room temperatures (approximately 22°C) and protein production was initiated by addition of IPTG to 1 mM. After four hours, bacterial pellets were obtained, resuspended in 20 mM HEPES pH 7.9/80 mM KCl/6 mM MgCl₂/1 mM Dithiothreitol/1 mM ATP/0.2 mM phenylmethylsulfonyl fluoride and protease inhibitors and sonicated. Debris was pelleted by centrifugation in an ss34 rotor for 1 hour at 12,000 rpm. The supernatant was incubated with glutathione sepharose 4B beads (Amersham Pharmacia Biotech AB, Uppsala, Sweden) and washed as previously described. Protein preparations were stored at -20°C in 20% glycerol.

Labeled ERs were produced using coupled *in vitro* transcription-translation (TNT kit, Promega, Madison, Wisconsin). Assays were carried out in a volume of 150 μ l that contained 137.5 μ l of ice-cold protein binding buffer (PBB) along with 10 μ l of GST-bead slurry corresponding to 3 μ g of fusion protein, 1 μ l of *in vitro* translated protein and 1.5 μ l of ligand or vehicle and/or peptides or vehicle. PBB was freshly prepared in 24 ml aliquots composed of 20 ml A-150 (20 mM Hepes, 150 mM KCl, 10 mM MgCl₂ and 1% glycerol), and 2 ml each of phosphate buffered saline supplemented, respectively, with 1% Triton X-100 and 1% NP-40. PMSF, DTT, BSA and protease inhibitor cocktail (Novagen) were added to 0.1 mM, 1 mM, 2 μ g/ml and 1/1000 dilution respectively. The mix was incubated for two hours in the cold room with gentle agitation, the beads were pelleted by spinning briefly on a bench top Eppendorf centrifuge, washed four times with PBB containing no BSA, and the pellet was dried under vacuum for twenty minutes. Labeled protein was subjected to SDS-polyacrylamide gel electrophoresis and autoradiography.

Transfections

HeLa cells were grown in DME/F-12 Ham's 1:1 mix, without phenol red (Sigma) containing 10% iron supplemented calf serum (Sigma) and pen-strep. Cells were transfected by electroporation [28]. Transfections contained 2µg of luciferase and actin-β-galactosidase reporters and, where indicated, 1µg of ER, VP16-fusion protein or GAL4-fusion protein expression vectors or empty vector controls. Luciferase and β-galactosidase activities were measured using luciferase (Promega, Madison, WI) and Galacto-Light assay systems (Tropix, Bedford, MA).

List of Abbreviations

AIB1 Amplified in breast cancer 1.

AF-1 Activation function 1

AF-2 Activation function 2

AP-1 Activator protein 1

CARM1 Coactivator associated arginine methyl-transferase 1.

CBP CREB binding protein.

CRE Cyclic Amp response element

DAX Dosage sensitive sex reversal adrenal hyperplasia congenital critical region on the X-chromosome, region 1.

DES Diethylstilbestrol

DNA Deoxyribonucleic acid

DBD DNA binding domain

TNFα Tumor necrosis factor alpha 1

E2 Estradiol

ERα Estrogen receptor alpha

ERβ Estrogen receptor beta

ERE, Estrogen response element

GRIP1 Glucocorticoid receptor interacting protein 1;

GST Glutathione S-transferase

H Helix

HAT Histone acetyl-transferase

HET-SAFB Hsp27-ERE-TATA binding protein/scaffold attachment factor B.

HDAC Histone de-acetylase

ID Interaction domain.

LBD Ligand binding domain

NR Nuclear receptor

NTD Amino terminal domain

N-CoR Nuclear receptor corepressor

PBS Phosphate buffered saline

PCR Polymerase chain reaction

PERC PGC-1 related estrogen receptor coactivator

PGC-1 Peroxisome proliferator activated receptor gamma coactivator 1.

RAR Retinoic acid receptor

REA Repressor of estrogen receptor activity

RIP140 Receptor interacting protein of 140 Kd

SDS-PAGE Sodium dodecyl sulfate polyacrylamide gel electrophoresis

SMRT Silencing Mediator of Retinoid and Thyroid Responsive transcription.

SERM Selective estrogen receptor modulator(s)

SHP Short heterodimer partner

SRC-1 Steroid receptor coactivator 1

TR Thyroid receptor

TRAP220 TR associated protein of 200 Kd.

Competing Interests

PW None declared.

CV None declared

PN None declared

RHP Jr. None declared.

AM Since the studies were performed, has moved to Plexxikon, a company with interests in protein structure.

BLW Since the studies were performed, has moved to Plexxikon, a company with interests in protein structure.

JDB has proprietary interests in, and serves as consultant and deputy director to Karo Bio AB, a Swedish company that develops pharmaceuticals that target NRs.

PJK has significant financial interests in, and is a consultant and former director of, Karo Bio AB.

Authors Contributions

PW Conceived and directed the studies, wrote the paper, performed plasmid constructions and transfection studies.

CV Performed transfection, mutagenesis and GST pull-down experiments under guidance of PW.

PN Performed transfection, mutagenesis and GST pull-down experiments under guidance of PW.

RHP Jr. Provided intellectual input in the study design.

AM. Expressed GST N-CoR fragments for pulldown studies with ER β .

BW Designed bacterial expression vectors for N-CoR fragments, cloned cDNA fragments for GST-N-CoR expression vectors and provided intellectual input into the study design.

JDB. Directed studies of NR interactions with N-CoR and provided intellectual input into study design.

PJK. Co-directed studies along with PW and provided intellectual input into study design.

All authors read and approved the final manuscript.

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