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Dose–response tests and semi-field evaluation of lethal and sub-lethal effects of slow release pyriproxyfen granules (Sumilarv[®] 0.5G) for the control of the malaria vectors *Anopheles gambiae sensu lato*

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Abstract

Background: Recently research has shown that larviciding can be an effective tool for integrated malaria vector control. Nevertheless, the uptake of this intervention has been hampered by the need to re-apply larvicides frequently. There is a need to explore persistent, environmentally friendly larvicides for malaria vector control to reduce intervention efforts and costs by reducing the frequency of application. In this study, the efficacy of a 0.5% pyriproxyfen granule (Sumilarv[®] 0.5G, Sumitomo Chemicals) was assessed for the control of *Anopheles gambiae sensu stricto* and *Anopheles arabiensis*, the major malaria vectors in sub-Saharan Africa.

Methods: Dose–response and standardized field tests were implemented following standard procedures of the World Health Organization's Pesticide Evaluation Scheme to determine: (i) the susceptibility of vectors to this formulation; (ii) the residual activity and appropriate retreatment schedule for field application; and, (iii) sub-lethal impacts on the number and viability of eggs laid by adults after exposure to Sumilarv[®] 0.5G during larval development.

Results: *Anopheles gambiae s.s.* and *An. arabiensis* were highly susceptible to Sumilarv[®] 0.5G. Estimated emergence inhibition (EI) values were very low and similar for both species. The minimum dosage that completely inhibited adult emergence was between 0.01-0.03 parts per million (ppm) active ingredient (ai). Compared to the untreated control, an application of 0.018 ppm ai prevented 85% (95% confidence interval (CI) 82%-88%) of adult emergence over six weeks under standardized field conditions. A fivefold increase in dosage of 0.09 ppm ai prevented 97% (95% CI 94%-98%) emergence. Significant sub-lethal effects were observed in the standardized field tests. Female *An. gambiae s.s.* that were exposed to 0.018 ppm ai as larvae laid 47% less eggs, and females exposed to 0.09 ppm ai laid 74% less eggs than females that were unexposed to the treatment. Furthermore, 77% of eggs laid by females exposed to 0.018 ppm ai failed to hatch, whilst 98% of eggs laid by females exposed to 0.09 ppm ai did not hatch.

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Conclusion: *Anopheles gambiae* s.s. and *An. arabiensis* are highly susceptible to Sumilarv[®]0.5G at very low dosages. The persistence of this granule formulation in treated habitats under standardized field conditions and its sub-lethal impact, reducing the number of viable eggs from adults emerging from treated ponds, enhances its potential as malaria vector control tool. These unique properties warrant further field testing to determine its suitability for inclusion in malaria vector control programmes.

Keywords: Pyriproxyfen, Sumilarv[®]0.5G, Malaria, Larval source management, *Anopheles gambiae* s.s., *Anopheles arabiensis*

Background

Malaria control interventions with long-lasting insecticidal nets (LLINs) and indoor residual spraying (IRS) have resulted in substantial reductions of malaria cases in sub-Saharan Africa [1,2]. Since both LLINs and IRS target the fraction of the vector population that enter houses [3,4] their efficacy is threatened by vectors developing resistance to insecticides used indoors [5-7] and behavioural adaptations where vectors shift their biting patterns to bite in early evening and in the morning when people are out of the nets [8,9]. There has also been a shift in the vector species' composition in parts of East Africa with LLINs dramatically reducing the numbers of largely endophilic *Anopheles gambiae* s.s. but having little or no impact on *Anopheles arabiensis* that tends to bite and rest outdoors [10-13] resulting in *An. arabiensis* becoming the dominant vector. Since IRS and LLINs cannot totally suppress malaria transmission there is a growing interest in the use of additional tools in an integrated vector management approach [14-18].

Larval source management has been re-evaluated for malaria control [19-24], with results indicating the added benefit larval control could have when used together with interventions that target adult mosquitoes [14,15,25]. One of the advantages of larval source management is that it targets the aquatic stages of the vectors thus controlling both indoor and outdoor biting and resting and insecticide resistant mosquitoes [26]. Commercially available chemical larvicides and microbials are highly effective in the control of the major malaria vectors of sub-Saharan Africa [20,24,27-33]. However, relatively few studies evaluated them under operational conditions [15,23,34-37] and a major limitation is their short activity under most environmental conditions, frequently requiring weekly re-application [20,21,34,38]. Larvicide and labour are the major costs in large-scale larval control programmes and these could be substantially reduced if re-application intervals could be reduced without jeopardizing the impact of the intervention [39]. In addition, the toxic effects of chemical-based larvicides to non-target aquatic insects limits their use for regular larviciding programmes [40,41].

Sumilarv[®]0.5G (Sumitomo Chemicals) is a granule insecticide developed for mosquito control. The active ingredient is pyriproxyfen (4-phenoxyphenyl (RS)-2-(2-pyridyloxy) propyl ether), a juvenile hormone analogue that acts as an insect growth regulator [42]. Pyriproxyfen generally inhibits adult emergence of target insects [43-45]. However it also has delayed effects on female reproduction of adult mosquitoes exposed to sub-lethal doses at the larval [46,47] or adult stage [48,49]. Sumilarv[®]0.5 has exceptional residual activity of up to six months for the control of *Aedes*, *Culex* and *Anopheles* mosquitoes in their natural breeding habitats [44,45,49,50]. Furthermore, pyriproxyfen has been evaluated as a safe insecticide for application in drinking water [51] with minimal impacts on non-target aquatic insects and the environment [52-56]. Nevertheless, Sumilarv[®]0.5G has never been evaluated for the control of immature stages of *An. gambiae* s.l., the major malaria vector in sub-Saharan Africa.

The objectives of the present study were to evaluate the efficacy of this granular formulation of pyriproxyfen for the control of *An. arabiensis* and *An. gambiae* s.s. by determining: (i) the minimum effective dose in dose-response tests; (ii) the optimum application dose to be applied under field conditions; (iii) the residual period of the optimum dose; and, (iv) the effects of sub-lethal doses on egg production and larval hatching. All tests were based on the World Health Organization Pesticide Evaluation Scheme (WHOPES) guidelines for laboratory and field testing of mosquito larvicides [57].

Methods

Study area

The study was conducted at the International Centre of Insect Physiology and Ecology-Thomas Odhiambo Campus (*icipe*-TOC) in Mbita (0° 26' 06.19" S; 34° 12' 53.13" E) close to Lake Victoria, Western Kenya (altitude 1,137 m). Here, the major malaria vectors are *An. arabiensis* with a small number of *An. gambiae* s.s. and *Anopheles funestus* [58]. The area is characterized by a tropical climate with an average annual minimum temperature of

16°C and an average maximum temperature of 28°C (*icipe*-TOC meteorological station data for 2010 to 2012). The area experiences two major rainy seasons, the long rains between March and June and the short rains between October and December. The average annual rainfall for 2010 to 2012 was 1,150 mm (*icipe*-TOC meteorological station).

Mosquitoes

Both laboratory and standardized field tests used insectary-reared third instar larvae of *An. arabiensis* and *An. gambiae* s.s. (Mbita strains). Larvae were reared in round plastic tubs (diameter 60 cm) filled with water (5 l, 5 cm high) from Lake Victoria filtered through a charcoal-sand filter. Mosquito larvae were fed with fish food (Tetramin®Baby) twice daily. Third instar mosquito larvae were selected from different tubs so that the larvae were of a similar range in size in each tub tested [59]. Mosquito larvae were reared at ambient climate and light conditions in a netting-screened greenhouse with an average daily temperature of 27°C, an average 76% relative humidity and a natural 12 hours of dark and 12 hours of light cycle.

Insecticide

Sumilarv®0.5G was provided by the manufacturer Sumitomo Chemicals Company, Japan, for all tests. It is a granular formulation containing 0.5% active ingredient (weight: weight).

Dose-response tests

Tests were done in the shade, under ambient climate and light conditions in a netting-screened greenhouse. Prior to the dose-response tests, range-finding tests were implemented by exposing test larvae to a wide range of test concentrations and a control. This served to find the activity range of the insecticide for each test species. Concentrations between 10 parts per million (ppm) active ingredient (ai) and 0.0000001 ppm ai were tested. After determining the emergence inhibition (EI) of the larvae in the wider range, nine concentrations were chosen, yielding between 10% and 95% EI in the range-finding tests in order to determine the EI₅₀, EI₉₀ and EI₉₉ in dose response bioassays. The following concentrations were tested: 0.005 ppm ai, 0.001 ppm ai, 0.0005 ppm ai, 0.0001 ppm ai, 0.00007 ppm ai, 0.00004 ppm ai and 0.00001 ppm ai, 0.000005 ppm ai, 0.000001 ppm ai.

A stock solution was prepared by grinding the granular formulation into a very fine powder following the procedure of Sihuincha and others [49]. Using a pestle and mortar, 5 g of Sumilarv®0.5G (25 mg ai) was ground and added to 500 ml of non-chlorinated tap water. This gave a stock solution of 10,000 ppm Sumilarv®0.5G

(50 ppm ai). The mouth of the vial was covered with aluminium foil and the solution left to agitate for one hour on a shaker. Since Sumilarv®0.5G is a slow release formulation the mixture was left overnight to allow the active ingredient to be released into solution. In the morning the mixture was again agitated on a shaker for 30 minutes to prepare a homogenous mixture since some of the inert ingredients of the formulation (potentially still containing some active ingredient) had settled overnight. Serial dilutions were made immediately after shaking in non-chlorinated tap water to produce the test concentrations.

Anopheles arabiensis and *An. gambiae* s.s. were evaluated in parallel. Each test concentration and a control were replicated four times per round per mosquito species. Two hundred ml of each test solution was set up in 300 ml plastic cups. Three rounds of tests were implemented. Separate batches of 25 insectary-reared third instar larvae of both test species were introduced into each test concentration and the control (non-chlorinated tap water). Thus in total 300 larvae of each species were tested per test concentration and control (total of 3000 larvae). Larvae were fed with Tetramin® Baby fish food every 24 hours and cups covered with netting to prevent any emerging adults from escaping. The number of live and dead larvae, pupae and adults was recorded every 24 hours for 10 days. Live pupae from each cup were transferred into a separate cup with approximately 20 ml of water from the respective cup of collection. These cups were covered with netting and pupae monitored for emergence. Separate pipettes were used to collect pupae from treated and control cups to avoid cross-contamination.

Standardized field tests

Standardized field tests [57] were carried out in an open field with grass approximately 3 cm in height between October 2011 and March 2012. Thirty artificial ponds were set up in an open field by sinking enamel-coated bowls (diameter 42 cm, depth 10 cm) into the ground (Figure 1A). Ponds were arranged 2 m apart in six rows. Each bowl was filled with 8 l of non-chlorinated tap water. Into each pond 2 l of soil collected from the surrounding field was added and mixed well to resemble a natural habitat. Batches of 50 insectary-reared third instar larvae were introduced into each pond. Sumilarv®0.5G treatment was applied after introduction of larvae. Treatment of the ponds was allocated randomly using a lottery system. In each treatment round, 10 of the ponds served as untreated controls; in five of them *An. arabiensis* were introduced and in the other five *An. gambiae* s.s. Two application rates of Sumilarv®0.5G were tested per mosquito species. The application rate was based on the surface area of the water, which was 0.14 m² per pond. Sumilarv®0.5G was spread

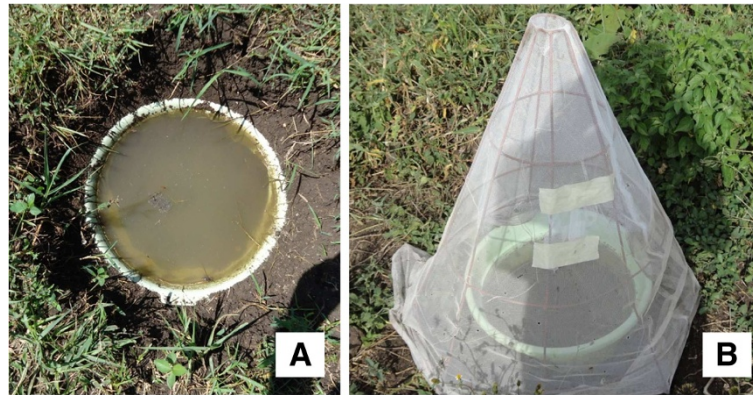


Figure 1 Set-up of standardized field test. (A) Enamel-coated bowl sunk into the ground and filled with water and soil to simulate a natural pond. (B) Netting-covered emergence trap on top of a pond to prevent escape of emerged adults.

evenly over the entire water surface by hand. Five ponds were treated with 1 mg ai per m² (equalling 0.018 ppm ai considering the volume of 8 l of water) while five other ponds were treated with 5 mg ai per m² (or 0.09 ppm ai) per mosquito species. A netting-covered emergence trap was placed on top of each pond to prevent wild mosquitoes from laying eggs in the sites and to prevent the escape of any emerging adult mosquitoes (Figure 1B). The residual activity of Sumilarv[®]0.5G was evaluated by introducing new batches of 50 insectary-reared third instar larvae into each pond at weekly intervals. After one week all the larvae had either emerged as adults or died. The efficacy of Sumilarv[®]0.5G was evaluated for six weeks. This experiment was implemented three times (referred to as rounds in the analyses).

To assess larval mortality, the number of larvae present in each habitat was counted daily. First, the emergence trap over each pond was assessed for the presence of any newly emerged adults and any adults collected with an aspirator and placed into a disposable cup covered with netting. Any pupae in the ponds were transferred into plastic cups holding 50 ml of the water from the respective pond. Pupae collections were done in the morning and evening so that any emergence or emergence inhibition could be recorded daily in the laboratory.

To monitor environmental parameters that may influence the efficacy of the insecticide, daily data on turbidity and pH of water in each pond was collected. Ponds were visually categorized into clear (ground visible) or turbid ponds. The water pH was measured using a pH meter (Phywe International, Germany).

Sub-lethal effects

Tests to assess the impact of sub-lethal doses of Sumilarv[®]0.5G were carried out under ambient conditions in a netting-screened greenhouse. The number of

eggs laid and the number of eggs hatched (number of offspring produced) per adult mosquito that emerged from treated ponds were compared to that of the adults that emerged from the untreated ponds in standardized field tests. All pupae used in these tests were collected from the ponds in week six of each test round. Emerged adults were maintained with 6% glucose solution *ad libitum*. When the adults were two to four days old they were blood-fed twice on a human arm on two successive days. A single gravid mosquito was introduced into each cage with an oviposition cup (diameter = 7 cm) containing 100 ml of non-chlorinated tap water. The number of eggs laid by each mosquito overnight and the number of eggs hatched over one week were counted. Sub-lethal effects of the treatment dosage of 1 mg ai per m² were tested with 20 individual females per round of semi-field test for *An. arabiensis* and *An. gambiae* s.s., respectively (total 3 × 20 = 60 females per species). Due to the persistent high immature mortality of the 5 mg ai per m² treatment only 10 females per species and round could be tested (total 3 × 10 = 30 females per species).

Statistical analyses

Data analyses were done with SPSS statistical software version 19. All data from the replicates of the dose-response tests were pooled by doses for each mosquito species for the estimation of the EI₅₀, EI₉₀ and EI₉₉ values using the log dosage-probit regression analysis with the test dosages as covariates and species as factors in the model. Relative median potency estimates were used to compare the susceptibility of the two species. Generalized estimating equations (GEE) were used to estimate the overall emergence inhibition of the two Sumilarv[®]0.5G dosages for the six weeks treatment period in standardized field tests. The number of successful emerged adults was the dependent variable and was fitted to a negative binomial distribution with a log-link function

and an exchangeable correlation matrix. The treatments, test rounds, mosquito species, water turbidity (clear, turbid), water pH (grouped in two categories: pH < 8, pH ≥ 8) and the occurrence of rain during the test week (no rain, rain) were added to the model as fixed factors. Since the same pond was evaluated repeatedly for larval mortality over the six-week period, the unique pond ID was included as the repeated measures variable. Interaction terms were included in the model between treatments and turbidity, treatments and pH, and treatments and rain. GEE models were also used to estimate the impact of sub-lethal concentrations on the number of eggs laid and the number of eggs that hatched from emerged *An. gambiae* s.s. adults. The parameter estimates of the GEE models were used to calculate the weekly mean adult emergence, mean number of eggs laid per female and mean number of laid eggs that hatched into larvae and the associated 95% confidence intervals (CIs) by removing the intercept from the models. For the calculation of percent reduction the weekly emergence inhibition in the treated ponds was corrected using Abbott's formula based on emergence in the untreated ponds as denominator [60]. Percent reduction was therefore calculated as follows:

$$\% \text{ treatment EI} = \frac{[\% \text{ untreated EI} - \% \text{ treated EI}] \times 100\%}{\% \text{ untreated EI}}$$

Results

Dose-response tests

The dose-response tests showed that Sumilarv[®]0.5G affected adult mosquito emergence in *An. arabiensis* and *An. gambiae* s.s. at very low and over a very wide range

of concentrations (0.000001-0.005 ppm ai). Data from the three rounds of dose-response tests showed similar trends in emergence inhibition for each species and were, therefore, pooled per dose (Figure 2) to estimate emergence inhibition (EI) rates; EI₅₀, EI₉₀ and EI₉₉ (Table 1). The minimum dosage that completely inhibited adult emergence was estimated to be between 0.01-0.03 ppm ai (Table 1). *Anopheles arabiensis* and *An. gambiae* s.s. were equally susceptible to Sumilarv[®]0.5G.

Standardized field tests

There was no difference in adult emergence from treated ponds between *An. arabiensis* and *An. gambiae* s.s. (p=0.3) and data for both species were pooled for analysis. The weekly adult emergence per round from the treated and untreated ponds is shown in Figure 3 and emergence inhibition calculated in Table 2. Complete emergence inhibition was observed for two weeks in rounds one and three of the high treatment dose of 5 mg ai per m² (0.09 ppm ai). However at the lower dosage of 1 mg ai per m² (0.018 ppm ai) which corresponded with the minimum effective dosage established in the dose-response tests complete emergence inhibition was only observed in week one in round one and three. Ponds treated at 5 mg ai per m² provided better residual impact than the lower treatment dosage of 1 mg ai per m² (Figure 3 and Table 2). Adjusting for other factors the GEE model estimated that Sumilarv[®]0.5G inhibited 85% of adult emergence over a period of six weeks at an application dose of 1 mg ai per m² and 97% at a dose of 5 mg ai per m² compared to emergence from untreated ponds (Table 3). The overall impact of 5 mg ai per m² on inhibiting emergence was significantly higher than the

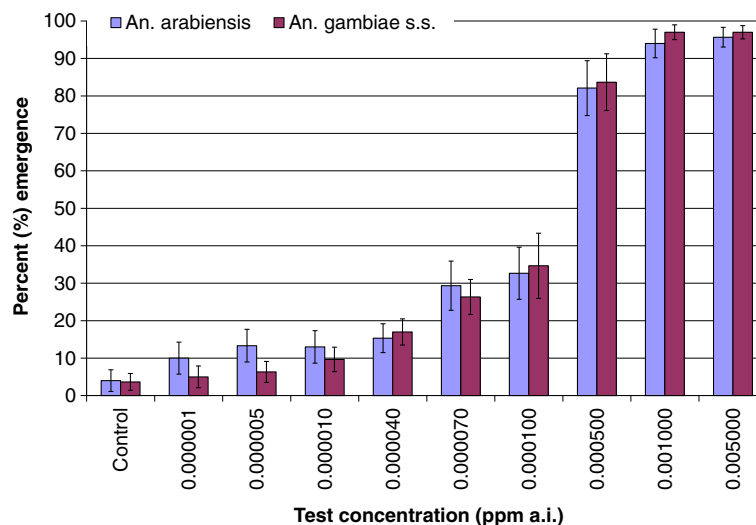


Figure 2 Average percent emergence inhibition (error bars: 95% confidence intervals) of *Anopheles arabiensis* and *Anopheles gambiae* s.s. in response to increasing concentrations (ppm ai) of Sumilarv[®]0.5G.

Table 1 Estimated doses (ppm ai) of Sumilarv® 0.5G for 50%, 90% and 99% emergence inhibition (EI) in *Anopheles gambiae* s.s. and *Anopheles arabiensis*

	<i>Anopheles arabiensis</i>	<i>Anopheles gambiae</i> s.s.
	ppm ai	ppm ai
EI ₅₀ (95%CI)	0.00012 (0.00009-0.00016)	0.00013 (0.00010-0.00017)
EI ₉₀ (95%CI)	0.00248 (0.00154-0.00450)	0.00139 (0.00092-0.00232)
EI ₉₉ (95%CI)	0.02860 (0.01379-0.07296)	0.00973 (0.00526-0.02159)

impact of 1 mg ai per m² (p<0.001). Despite consistent rainfall during the first round of the standardized field tests and occasional rainfall during the following two rounds (Figure 4), rain did neither affect the emergence of adults from control and treatment ponds nor the impact of the treatments (Table 3). There were also no main effects of water turbidity or pH on adult emergence but interactions were identified between the treatments and water turbidity, and the treatments and water pH. Turbid water and high pH reduced the impact of the treatments leading to slightly higher adult emergence from treatment ponds under these conditions (Table 3). The impact of the interactions can be calculated by multiplication of the odds ratios [61]. This means for example emergence inhibition was 85% at 1 mg ai per m² when ponds were clear and had a pH <8, emergence inhibition was reduced to 79% when the same treatment pond was turbid with a pH <8 and to 74% when the same treatment pond was turbid and had a pH ≥8. Similarly for the 5 mg ai per m² ponds in round one, overall emergence inhibition is 97% when treatment ponds are clear with pH <8, emergence inhibition is reduced to 95% when the treatment ponds are turbid with pH <8 and further reduced to 90% when the treatment ponds are turbid and with pH ≥8.

Sub-lethal effects

The impact of sub-lethal effects could not be evaluated for *An. arabiensis* that emerged from pupae since neither females from untreated ponds nor females from treated ponds laid eggs, possibly due to unsuitable mating conditions provided for this species [62]. Exposure of *An. gambiae* s.s. to both Sumilarv® 0.5G dosages during the larval stage resulted in: (i) a reduced probability of the adult female laying eggs; (ii) reduced mean number of eggs laid per female; and, (iii) reduced mean number of eggs that hatched into larvae (Table 4). Treatment rounds were not significantly different (p=0.687), and data for all rounds for *An. gambiae* s.s. were pooled for analysis. Mosquitoes that emerged from treated ponds were 65-68% less likely to lay eggs compared to mosquitoes that emerged from untreated ponds. The mean number of eggs laid per female *An. gambiae* s.s. was reduced by 47% from females emerging from ponds treated at 1 mg ai per m² and by 74% from females emerged from ponds treated at 5 mg ai per m² compared to that in the untreated controls (Table 4). The impact of the higher dosage was twice the impact measured from the lower dosage (odds ratio (OR) 2.1, 95% CI 1.2-3.7, p=0.02). Furthermore, it was 90% less likely for an egg to hatch that was laid by a female exposed to the higher Sumilarv® 0.5G dosage compared to eggs laid by females that emerged from low dosage ponds (OR=0.10, 95% CI 0.04-0.23, p<0.0001). The probability of an egg hatching was reduced by 77% for eggs laid by a female exposed to the lower treatment dosage and 98% for eggs laid by a female exposed to the higher dosage as compared to eggs in females that emerged from the untreated control ponds.

Discussion

Anopheles arabiensis and *An. gambiae* s.s. were equally and highly susceptible to Sumilarv® 0.5G under laboratory

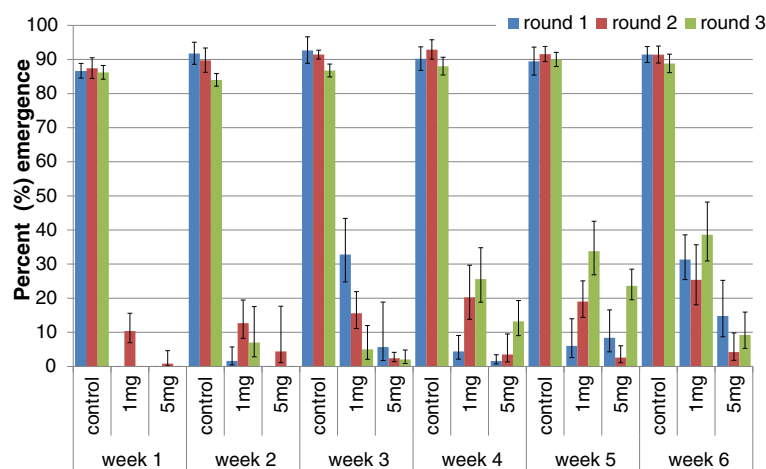


Figure 3 Mean adult emergence (error bars: 95% confidence intervals) of *Anopheles gambiae* s.l. in standardized field tests after application of 1 mg or 5 mg ai per m² Sumilarv® 0.5G in artificial ponds.

Table 2 Weekly percent emergence inhibition (95% CI) of *Anopheles gambiae* s.l. from treated ponds

	Week 1	Week 2	Week 3	Week 4	Week 5	Week 6
1 mg ai per m ²						
Round 1	100	98 (94–99)	65 (55–72)	95 (90–98)	93 (85–97)	66 (59–71)
Round 2	88 (83–92)	86 (76–90)	83 (76–88)	78 (69–85)	79 (73–84)	72 (62–80)
Round 3	100	92 (80–97)	94 (86–98)	71 (62–78)	62 (54–69)	57 (47–64)
5 mg ai per m ²						
Round 1	100	100	94 (80–98)	98 (96–99)	91 (82–95)	84 (73–90)
Round 2	99 (95–100)	95 (81–99)	97 (96–98)	96 (90–99)	97 (94–99)	95 (90–98)
Round 3	100	100	98 (95–99)	85 (79–89)	74 (69–78)	90 (83–94)

and standardized field conditions. Sumilarv[®]0.5G inhibited over 80% of the total adult emergence over a period of six weeks at both application dosages. However, weekly emergence rates increased steadily over the six-week test period at the lower dosage that corresponded with the EI₉₉ in the laboratory and weekly emergence inhibition was frequently lower than the 80% that is recommended by WHOPES for successful immature control [57]. Laboratory tests were conducted under standardized conditions without major abiotic and biotic influences and therefore EI values represent only minimum dosages. Application rates frequently have to be increased up to several times the minimum dose to obtain sufficient immature control under field conditions [57,63]. The higher dosage of 5 mg ai per m² or 0.09 ppm ai inhibited well over 80% of adult emergence in all but one test week. This dosage was 4.5 times the average EI₉₉ in the laboratory. Further field tests to establish the optimum dose for operational control in a variety of different habitats are necessary but based on the results presented here it is likely that the optimum dosage lies between the two tested here and therefore coincides with the maximum dosage recommended by the manufacturer (0.05 ppm ai) for operational control of other mosquito species [64].

The estimated emergence inhibition rates from the dose–response tests were four times higher than those previously reported by Kawada and his colleagues [65] for *An. gambiae*, but within the range of rates estimated for *Culex* and *Aedes* species [66–70]. These differences may arise from the different pyriproxyfen formulations used in separate studies [71], but also from the material of the test containers [44]. Kawada and colleagues used a 5% emulsifiable concentrate formulation while in the present study a granular formulation was used and had to be crushed in a mortar for the laboratory tests, which might have not led to an equal amount of active ingredients being released into the stock solution. Also, in the present study plastic cups were used for bioassays while Kawada and his colleagues used aluminium cups. There is a concern that the active ingredient pyriproxyfen adheres to plastic [72] leading to a longer residual effect from such treated

containers due to a continuous slow release from the plastic [54]. In the short term however, plastic might reduce the amount of active ingredient in the water, which could be responsible for the higher estimates of EI concentrations found in this study. The extremely low concentrations of active ingredient needed for the control of mosquitoes with Sumilarv[®]0.5G is worth noting. The estimated effective dose of pyriproxyfen is approximately 10 times lower than those reported for microbial larvicides [20,21]. This is not surprising since pyriproxyfen is a juvenile hormone analogue, and insect hormones, like all hormones, operate at extremely low concentrations as chemical messengers [70,73]. Thus, far smaller quantities of Sumilarv[®]0.5G would be required for larviciding programmes compared to microbial larvicides, thereby helping to lower costs associated with transporting and storing larvicides [39].

The residual impact of Sumilarv[®]0.5G on *An. gambiae* s.l. emergence observed here corresponds well with reports from previous studies on other mosquito species [44,67,74] but application dosages required to achieve the same effect seem slightly higher for *An. gambiae* s.l. Sumilarv[®]0.5G at 0.02 ppm ai and 0.05 ppm ai provided almost complete emergence inhibition of *Aedes aegypti*, *Aedes albopictus* and *Aedes taeniorhynchus*, *Culex nigripalpus* and *Anopheles quadrimaculatus* for six weeks under standardized field conditions [74]. This slow-release formulation has even been shown to exhibit prolonged residual activity for control of *Aedes* larvae even when the treatments were diluted by using replacement of treated water with untreated water in the treated containers [44,75]. Similarly, here it was observed that rainfall did not negatively affect the impact of the treatments. Exceptional performance of Sumilarv[®]0.5G was reported for the control of *Anopheles culicifacies* in confined gem pits in Sri Lanka [45] where a single application of pyriproxyfen at 0.01 ppm ai was sufficient to inhibit adult emergence for approximately six months. Similarly, Sihuincha and colleagues [49] reported complete emergence inhibition of *Ae. aegypti* for five months from water tanks in Peru at an application rate of Sumilarv[®]0.5G of 0.05 ppm ai. Overall

Table 3 Multivariable analyses (GEE) of factors affecting the emergence of adult malaria vectors over a six week period from artificial ponds treated with Sumilarv® 0.5G

Explanatory variable	OR	95% CI	p
Treatment			
1 mg ai per m ²	0.03	0.02-0.05	<0.0001
5 mg ai per m ²	0.15	0.12-0.18	<0.0001
control	1		
Round			
round 3	1.19	1.00-1.41	0.050
round 2	1.03	0.78-1.34	0.859
round 1	1		
Vector species			
<i>An. arabiensis</i>	0.95	0.86-1.05	0.278
<i>An. gambiae</i> s.s.	1		
Water turbidity			
turbid	1.01	0.95-1.07	0.765
clear	1		
Water pH			
≥ 8	0.99	0.91-1.08	0.820
< 8	1		
Rain during test week			
rain	1.05	0.92-1.20	0.449
no rain	1		
Interaction between treatment and turbidity			
5 mg ai per m ² *turbid	1.93	1.12-3.26	0.017
5 mg ai per m ² *clear	1		
1 mg ai per m ² *turbid	1.40	1.08-1.79	0.011
1 mg ai per m ² *clear	1		
Interaction between treatment and pH			
5 mg ai per m ² *pH≥8	1.90	1.13-2.85	0.002
5 mg ai per m ² *pH<8	1		
1 mg ai per m ² *pH≥8	1.25	1.06-1.47	0.008
1 mg ai per m ² *pH<8	1		
Interaction between treatment and rain			
5 mg ai per m ² *rain	1.23	0.89-1.69	0.211
5 mg ai per m ² *no rain	1		
1 mg ai per m ² *rain	0.87	0.70-1.07	0.870
1 mg ai per m ² *no rain	1		

it can be concluded from previous work that the efficacy and residual activity of different pyriproxyfen-containing products depends on the formulation, dose, habitat types treated, prevailing weather conditions and target mosquito species [53,67,74].

The current study showed that the efficacy of Sumilarv® 0.5G is reduced in turbid water and water with a pH ≥8. Water is turbid because it carries a suspension of

fine particles of both organic and inorganic matter in the water column. Some of the turbidity observed here might have been due to algae and bacteria growth in the established habitats, which in turn might have increased the water pH. It is possible that the active ingredient, pyriproxyfen, is adsorbed onto particles in the water column and was less accessible to larvae. Turbidity and pH of aquatic habitats are important parameters that are associated with the abundance, development and survival of *Anopheles* larvae [76]. *Anopheles* larvae are known to exploit aquatic habitats with varying degrees of water turbidity and pH [76,77]. Suspended particles including algae in the water column in turbid ponds provide mosquitoes with food that enhances their development and survival thus increase emergence from turbid ponds [78,79]. Mulligan and Schaefer [80] found pyriproxyfen to adsorb onto organic matter which might have been responsible for larvae to be exposed to reduced doses. This needs to be considered and monitored in field operations where it might be necessary to increase the application dose or reduce retreatment intervals to ensure a consistent emergence inhibition above 80% as recommended by WHOPEs [57].

An added benefit to the direct effect of Sumilarv® 0.5G on immature stages were the sub-lethal effects that affected the offspring of adult females that successfully emerged from treated ponds. At 5 mg ai per m² the reproduction of females was reduced by well over 90%. Similar effects of insect growth regulators have been shown for *Aedes* and *Culex* [46,47,81]. The laying of non-viable eggs by female *An. gambiae* s.s. emerging from treated ponds might further extend the efficacy and residual effect of pyriproxyfen, and may help further reduce intervention costs by extending the retreatment intervals. It would be particularly helpful in the context of an auto-dissemination strategy [82] of Sumilarv® 0.5G where potentially only sub-lethal doses are transferred to a habitat by female gravid mosquitoes. The delayed sub-lethal effects of insect growth regulators were also shown to affect the sex ratio and to reduce blood-feeding rates in exposed mosquitoes [47,83]. Similar effects were shown for adults exposed to pyriproxyfen [48,49,84]. Ohashi and colleagues [84] demonstrated that *An. gambiae* s.s. was completely sterilized, with no female laying eggs after exposure to pyriproxyfen-treated nets. Insect growth regulators have been shown to suppress ovarian development and egg development in mosquitoes [85,86]. Judson and de Lumen [85] showed that exposure of *Ae. aegypti* females to juvenile hormone analogues suppressed egg development by inhibiting development of ovarian follicles. Fournet and colleagues [86] similarly showed that the ovarian development of *Ae. aegypti* females that emerged from larvae exposed to insect growth regulators was affected.

As with every insecticide it is important to be cautious about using pyriproxyfen formulations as a stand-alone

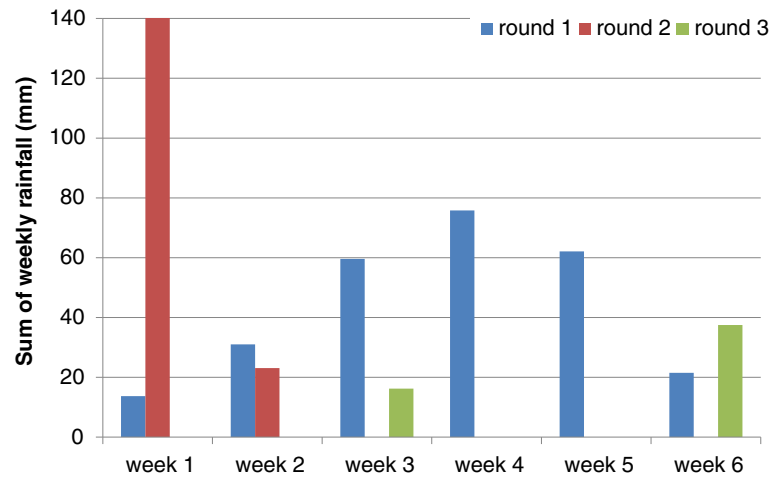


Figure 4 Weekly rainfall during the three rounds of standardized field tests.

intervention since tolerance to pyriproxyfen has been found in Diptera [87,88]. It is also of concern to know whether the progeny of gravid females that are exposed to sub-lethal level doses of pyriproxyfen and survive have greater tolerance to pyriproxyfen than other mosquitoes. If this is the case, resistance may spread.

Pyriproxyfen exhibits favourable characteristics for utilization as a larvicide for mosquito control. The recommended application rate in drinking water limit of 300 ppb (0.3 ppm) [51] is several folds higher than the recommended dose of 0.01-0.05 ppm [64] for mosquito control and also has minimal environmental impacts at recommended rates for mosquitoes [52,53].

Conclusion

Anopheles arabiensis and *An. gambiae* s.s. are highly susceptible to Sumilarv[®]0.5G at very low dosages. The persistence of Sumilarv[®]0.5G in treated habitats under standardized field conditions and its sub-lethal impact, reducing the number of viable eggs from adults emerging from treated ponds, enhances its potential as a malaria vector control tool in integrated vector management strategies. These unique properties of Sumilarv[®]0.5G warrant further field testing in a range of natural *An. gambiae* s.l. larval habitats and under operational conditions to recommend if and how this insect growth regulator could be

included in vector control programmes for malaria control in sub-Saharan Africa.

Based on the results of this study the maximum dosage recommended by the manufacturer for other mosquito species of 0.05 ppm ai is recommended as the minimum dosage for further field testing for *An. gambiae* s.l. control. Although the residual effect observed for the test concentrations lasted for a six-week period, initially a shorter retreatment interval should be evaluated under natural conditions where habitat types and water quality are highly heterogeneous and might affect the residual activity. Furthermore, the estimation of retreatment intervals should also consider the probability of new habitats emerging during treatment cycles that could then harbour mosquito larvae that might successfully emerge before the target area receives another round of Sumilarv[®]0.5G application. Initial application cycles should be determined for the predominant habitat type in the target area, the season of application and the development time of immature vectors. In areas where temporary habitats dominate or areas with high rainfall an initial application cycle of two to three weeks should be tested whilst in areas of more semi-permanent to permanent habitats or during dry seasons a three to four-weekly application cycle might be appropriate for an initial field operation informed by a monitoring and evaluation programme.

Table 4 Sub-lethal effects of Sumilarv[®] 0.5G on egg laying and hatching of *Anopheles gambiae* s.s.

	Control	1 mg ai per m ²	5 mg ai per m ²
Number of females exposed	60	60	30
Number of females that laid eggs	43	27	14
Mean number of eggs/female (95% CI)	43.8 (35.6-53.8)	23.1 (16.5-32.3)	11.2 (6.9-18.2)
Mean number eggs/female hatched (95% CI)	37.4 (30.5-45.8)	8.7 (6.0-12.4)	0.8 (0.4-1.8)

Competing interests

Sumitomo Chemicals, Japan, the commercial manufacturer of Sumilarv®0.5G, provided the insecticide for this study free of charge. Nevertheless, neither the manufacturer nor any of the funders of this work had any role in the design, analysis or interpretation of the results, nor in the drafting of the manuscript.

Authors' contributions

UF and SWL conceived the idea for this research. OM, SWL and UF developed the experimental design and protocols. OM implemented the experiments. OM and UF analysed the data and drafted the manuscript. All authors contributed to the final draft, read and approved the manuscript.

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References

1. Steketee RW, Campbell CC: **Impact of national malaria control scale-up programmes in Africa: magnitude and attribution of effects.** *Malar J* 2010, **9**:299.
2. Okumu FO, Moore SJ: **Combining indoor residual spraying and insecticide-treated nets for malaria control in Africa: a review of possible outcomes and an outline of suggestions for the future.** *Malar J* 2011, **10**:208.
3. Robert V, Carnevale P: **Influence of deltamethrin treatment of bed nets on malaria transmission in the Kou valley, Burkina Faso.** *Bull World Health Organ* 1991, **69**:735-740.
4. Pinder M, Jawara M, Jarju LBS, Kandeh B, Jeffries D, Lluberas MF, Mueller J, Parker D, Bojang K, Conway DJ, Lindsay SW: **To assess whether indoor residual spraying can provide additional protection against clinical malaria over current best practice of long-lasting insecticidal mosquito nets in The Gambia: study protocol for a two-armed cluster-randomized trial.** *Trials* 2011, **12**:147.
5. WHO: *Expert Committee on malaria*, 892. Geneva: WHO Technical Report Series; 2000:1-71.
6. Kawada H, Futami K, Komagata O, Kasai S, Tomita T, Sonye G, Mwatele C, Njenga SM, Mwandawiro C, Minakawa N, Takagi M: **Distribution of a knockdown resistance mutation (L1014S) in *Anopheles gambiae* s.s. and *Anopheles arabiensis* in Western and Southern Kenya.** *PLoS One* 2011, **6**:e24323.
7. Chouaibou M, Etang J, Brevalet T, Nwane P, Hinzoumbe CK, Mimpfoundi R, Simard F: **Dynamics of insecticide resistance in the malaria vector *Anopheles gambiae* s.l. from an area of extensive cotton cultivation in Northern Cameroon.** *Trop Med Int Health* 2008, **13**:476-486.
8. Reddy MR, Overgaard HJ, Abaga S, Reddy VP, Caccone A, Kiszewski AE, Slotman MA: **Outdoor host seeking behaviour of *Anopheles gambiae* mosquitoes following initiation of malaria vector control on Bioko Island, Equatorial Guinea.** *Malar J* 2011, **10**:184.
9. Faye O, Konate L, Mouchet J, Fontenille D, Ngayo SY, Hebrard G, Herve JP: **Indoor resting by outdoor biting females of *Anopheles gambiae* complex (Diptera: Culicidae) in the Sahel of Northern Senegal.** *J Med Entomol* 1997, **34**:285-289.
10. Bayoh MN, Mathias DK, Odiere MR, Mutuku FM, Kamau L, Gimign JE, Vulule JM, Hawley WA, Hamel MJ, Walker ED: ***Anopheles gambiae*: historical population decline associated with regional distribution of insecticide-treated bed nets in western Nyanza Province, Kenya.** *Malar J* 2010, **9**:62.
11. Russell TL, Lwetoijera DW, Maliti D, Chipwaza B, Kihonda J, Charlwood JD, Smith TA, Lengeler C, Mwanyangala MA, Nathan R, Knols BGJ, Takken W, Killeen GF: **Impact of promoting longer-lasting insecticide treatment of bed nets upon malaria transmission in a rural Tanzanian setting with pre-existing high coverage of untreated nets.** *Malar J* 2010, **9**:187.
12. Sinka ME, Bangs MJ, Manguin S, Coetzee M, Mbogo CM, Hemingway J, Patil AP, Temperley WH, Gething PW, Kabaria CW, Okara RM, Boeckel TV, Godfray HCJ, Harbach RE, Hay SI: **The dominant *Anopheles* vectors of human malaria in Africa, Europe and the Middle East: occurrence data, distribution maps and bionomic precis.** *Parasit Vectors* 2010, **3**:117.
13. Kitau J, Oxborough RM, Tongu PK, Matowo J, Magesa SM, Bruce J, Moshia FW, Rowland MW: **Species shifts in the *Anopheles gambiae* complex: do LLINs successfully control *Anopheles arabiensis*?** *PLoS One* 2012, **7**:e31481.
14. Chanda E, Masaninga F, Coleman M, Sikaala C, Katebe C, Macdonald M, Baboo KS, Govere J, Manga L: **Integrated vector management: the Zambian experience.** *Malar J* 2008, **7**:164.
15. Fillinger U, Ndenga B, Githeko A, Lindsay SW: **Integrated malaria vector control with microbial larvicides and insecticide-treated nets in western Kenya: a controlled trial.** *Bull World Health Organ* 2009, **87**:655-665.
16. Beier JC, Keating J, Githure JI, Macdonald MB, Impoinvil DE, Novak RJ: **Integrated vector management for malaria control.** *Malar J* 2008, **7**(Suppl 1):S4.
17. WHO: *Global strategic framework for integrated vector management*. 2004. whqlibdoc.who.int/hq/2004/WHO_CDS_CPE_PVC_2004_2010.pdf.
18. Clive S: **Integrated approach to malaria control.** *Clin Microbiol Rev* 2002, **15**:278-293.
19. Bukhari T, Knols BG: **Efficacy of Aquatain, a monomolecular surface film, against the malaria vectors *Anopheles stephensi* and *An. gambiae* s.s. in the laboratory.** *Am J Trop Med Hyg* 2009, **80**:758-763.
20. Fillinger U, Knols BG, Becker N: **Efficacy and efficiency of new *Bacillus thuringiensis* var *israelensis* and *Bacillus sphaericus* formulations against Afrotropical anophelines in Western Kenya.** *Trop Med Int Health* 2003, **8**:37-47.
21. Majambere S, Lindsay SW, Green C, Kandeh B, Fillinger U: **Microbial larvicides for malaria control in the Gambia.** *Malar J* 2007, **6**:76.
22. Geissbuhler Y, Kannady K, Chaki PP, Emidi B, Govella NJ, Mayagaya V, Kiama M, Mtasiwa D, Mshinda H, Lindsay SW, Tanner M, Fillinger U, Castro MC, Killeen GF: **Microbial larvicide application by a large-scale, community-based program reduces malaria infection prevalence in urban Dar es Salaam.** *Tanzania. PLoS ONE* 2009, **4**:e5107.
23. Fillinger U, Lindsay SW: **Suppression of exposure to malaria vectors by an order of magnitude using microbial larvicides in rural Kenya.** *Trop Med Int Health* 2006, **11**:1629-1642.
24. Shillilu JI, Tewolde GM, Brantly E, Githure JI, Mbogo CM, Beier JC, Fusco R, Novak RJ: **Efficacy of *Bacillus thuringiensis israelensis*, *Bacillus sphaericus* and temephos for managing *Anopheles* in Eritrea.** *J Am Mosq Control Assoc* 2003, **19**:251-258.
25. Shaikat AM, Breman JG, McKenzie FE: **Using the entomological inoculation rate to assess the impact of vector control on malaria parasite transmission and elimination.** *Malar J* 2010, **9**:122.
26. Fillinger U, Lindsay SW: **Larval source management for malaria control in Africa: myths and reality.** *Malar J* 2011, **10**:353.
27. Seyoum A, Abate D: **Larvicidal efficacy of *Bacillus thuringiensis* var. *israelensis* and *Bacillus sphaericus* on *Anopheles arabiensis* in Ethiopia.** *World J Microb Biot* 1997, **13**:21-24.
28. Karch S, Manzambi ZA, Salaun JJ: **Field trials with Vectolex (*Bacillus sphaericus*) and Vectobac (*Bacillus thuringiensis* (H-14)) against *Anopheles gambiae* and *Culex quinquefasciatus* breeding in Zaire.** *J Am Mosq Control Assoc* 1991, **7**:176-179.
29. Karch S, Asidi N, Manzambi ZM, Salaun JJ: **Efficacy of *Bacillus sphaericus* against the malaria vectors *Anopheles gambiae* and other mosquitoes in swamps and rice fields in Zaire.** *J Am Mosq Control Assoc* 1992, **8**:376-380.
30. Majori G, Ali A, Sabatinelli G: **Laboratory and field efficacy of *Bacillus thuringiensis* var *israelensis* and *Bacillus sphaericus* against *Anopheles gambiae* s.l. and *Culex quinquefasciatus* in Ouagadougou, Burkina Faso.** *J Am Mosq Control Assoc* 1987, **3**:20-25.

31. Skovmand O, Bauduin S: Efficacy of a granular formulation of *Bacillus sphaericus* against *Culex quinquefasciatus* and *Anopheles gambiae* in West African countries. *J Vector Ecol* 1996, **22**:43–51.
32. Ravoahangimalala O, Thiery I, Sinegre G: Rice field efficacy of deltamethrin and *Bacillus thuringiensis israelensis* formulations on *Anopheles gambiae* s.s. in the Anjiro Region of Madagascar. *Bull Soc Vector Ecol* 1994, **19**:169–174.
33. Ragoonansingh RN, Njunwa KJ, Curtis CF, Becker N: A field study of *Bacillus sphaericus* for the control of culicine and anopheline mosquito larvae in Tanzania. *Bull Soc Vector Ecol* 1992, **17**:45–50.
34. Fillinger U, Kannady K, William G, Vanek MJ, Dongus S, Nyika D, Geissbuhler Y, Chaki PP, Govella NJ, Mathenge EM, Singer BH, Mshinda H, Lindsay SW, Tanner M, Mtsiwa D, de Castro MC, Killeen GF: A tool box for operational mosquito larval control: preliminary results and early lessons from the urban malaria control programme in Dar es Salaam, Tanzania. *Malar J* 2008, **7**:20.
35. Majambere S, Pinder M, Fillinger U, Ameh D, Conway DJ, Green C, Jeffries D, Jawaara M, Milligan PJ, Hutchinson R, Lindsay SW: Is mosquito larval source management appropriate for reducing malaria in areas of extensive flooding in the Gambia? A cross-over intervention trial. *Am J Trop Med Hyg* 2010, **82**:176–184.
36. Shillilu J, Mbogo C, Ghebremeskel T, Githu J, Novak R: Mosquito larval habitats in a semiarid ecosystem in Eritrea: impact of larval habitat management on *Anopheles arabiensis* population. *Am J Trop Med Hyg* 2007, **76**:103–110.
37. Barbazan P, Baldet T, Darriet F, Escaffre H, Djoda DH, Hougard JM: Impact of treatments with *Bacillus sphaericus* on *Anopheles* populations and the transmission of malaria in Maroua, a large city in a savannah region of Cameroon. *J Am Mosq Control Assoc* 1998, **14**:33–39.
38. Skovmand O, Sanogo E: Experimental formulations of *Bacillus sphaericus* and *B. thuringiensis israelensis* against *Culex quinquefasciatus* and *Anopheles gambiae* (Diptera: Culicidae) in Burkina Faso. *J Med Entomol* 1999, **36**:62–67.
39. Worrall E, Fillinger U: Large-scale use of mosquito larval source management for malaria control in Africa: a cost analysis. *Malar J* 2011, **10**:338.
40. Fortin C, Maire A, Leclair R: The residual effect of temephos (Abate 4-E) on nontarget communities. *J Am Mosq Control Assoc* 1987, **3**:282–288.
41. Fales JH, Spangler PJ, Bodenstein OF, Mills GD Jr, Durbin CG: Laboratory and field evaluation of Abate against a backswimmer (*Notonecta undulata* Say) (Hemiptera: Notonectidae). *Mosq News* 1968, **28**:77–81.
42. Sumilarv product information. <http://www.olyset.net/vectorcontrol/sumilarv/>.
43. Kamimura K, Arakawa A: Field evaluation of an insect growth regulator, pyriproxyfen, against *Culex pipiens* Pallens and *Culex tritaeniorhynchus*. *Jap J Sanit Zool* 1991, **42**:249–252.
44. Vythilingam I, Luz BM, Hanni R, Beng TS, Huat TC: Laboratory and field evaluation of the insect growth regulator pyriproxyfen (Sumilarv 0.5G) against dengue vectors. *J Am Mosq Control Assoc* 2005, **21**:296–300.
45. Yapabandara AM, Curtis CF: Laboratory and field comparisons of pyriproxyfen, polystyrene beads and other larvicidal methods against malaria vectors in Sri Lanka. *Acta Trop* 2002, **81**:211–223.
46. Kamal HA, Khater EIM: The biological effects of the insect growth regulators; pyriproxyfen and diflubenzuron on the mosquito *Aedes aegypti*. *J Egypt Soc Parasitol* 2010, **40**:565–574.
47. Loh PY, Yap HH: Laboratory studies on the efficacy and sublethal effects of an insect growth regulator, pyriproxyfen (S-31183) against *Aedes aegypti* (Linnaeus). *Trop Biomed* 1989, **6**:7–12.
48. Itoh T, Kawada H, Abe A, Eshita Y, Rongsriyam Y, Igarashi A: Utilization of bloodfed females of *Aedes aegypti* as a vehicle for the transfer of the insect growth regulator pyriproxyfen to larval habitats. *J Am Mosq Control Assoc* 1994, **10**:344–347.
49. Sihuincha M, Zamora-Perea E, Orellana-Rios W, Stancil JD, Lopez-Sifuentes V, Vidal-Ore C, Devine GJ: Potential use of pyriproxyfen for control of *Aedes aegypti* (Diptera: Culicidae) in Iquitos, Peru. *J Med Entomol* 2005, **42**:620–630.
50. Chavasse DC, Lines JD, Ichimori K, Majala AR, Minjas JN, Marijani J: Mosquito control in Dar es Salaam. II. Impact of expanded polystyrene beads and pyriproxyfen treatment of breeding sites on *Culex quinquefasciatus* densities. *Med Vet Entomol* 1995, **9**:147–154.
51. WHO: *Pyriproxyfen in drinking-water: Use for vector control in drinking-water sources and containers*. Geneva: Background document for development of WHO guidelines for drinking-water quality; 2008.
52. Mulla MS, Darwazeh HA, Kennedy B, Dawson DM: Evaluation of new insect growth regulators against mosquitoes with notes on nontarget organisms. *J Am Mosq Control Assoc* 1986, **2**:314–320.
53. Schaefer CH, Miura T, Dupras EF, Mulligan FS, Wilder WH: Efficacy, nontarget effects, and chemical persistence of S-31183, a promising mosquito (Diptera: Culicidae) control agent. *J Econ Entomol* 1988, **81**:1648–1655.
54. Schaefer CH, Dupras EF Jr, Mulligan FS III: Studies on the environmental persistence of S-31183 (Pyriproxyfen): adsorption onto organic matter and potential for leaching through soil. *Ecotoxicol Environ Saf* 1991, **21**:207–214.
55. Sullivan J: *Environmental fate of pyriproxyfen*. [<http://www.cdpr.ca.gov/docs/emon/pubs/fatememo/pyrpxrfrn.pdf>]
56. Schaefer CH, Miura T: Chemical persistence and effects of S-31183, 2-(1-Methy-2(4- phenoxyphenoxy)ethoxy) pyridine on aquatic organisms in field tests. *J Econ Entomol* 1990, **83**:1766–1776.
57. WHO: *Guidelines for laboratory and field testing of mosquito larvicides*. Geneva: World Health Organization Communicable Disease Control, Prevention and Eradication. WHO Pesticide Evaluation Scheme. WHO/CDS/WHOPES/GCDPP/2005.2013; 2005.
58. Kawada HD, Dida GO, Ohashi K, Komagata O, Kasai S, Tomita T, Sonye G, Maekawa Y, Mwatelele C, Njenga SM, Mwandawiro C, Minakawa N, Takagi M: Multimodal pyrethroid resistance in malaria vectors, *Anopheles gambiae* s.s., *Anopheles arabiensis*, and *Anopheles funestus* s.s. in Western Kenya. *PLoS One* 2011, **6**:e22574.
59. Araujo MS, Gil LHS, e-Silva AA: Larval food quantity affects development time, survival and adult biological traits that influence the vectorial capacity of *Anopheles darlingi* under laboratory conditions. *Malar J* 2012, **11**:261.
60. Abbott WS: A method of computing the effectiveness of an insecticide. *J Am Mosq Control Assoc* 1987, **3**:302–303.
61. Katz MH: *Multivariable analysis: A practical guide for clinicians*. 2nd edition. Cambridge: Cambridge University Press; 2006.
62. Marchand RP: A new cage for observing mating behaviour of wild *Anopheles gambiae* in the laboratory. *J Am Mosq Control Assoc* 1985, **1**:234–236.
63. Becker N, Rettich F: Protocol for the introduction of new *Bacillus thuringiensis israelensis* products into the routine mosquito control program in Germany. *J Am Mosq Control Assoc* 1994, **10**:527–533.
64. Sumilarv, manufacturer's product information. [<http://www.olyset.net/vectorcontrol/sumilarv/>]
65. Kawada Y, Shono Y, Ito T, Abe Y: Laboratory evaluation of insect growth regulators against several species of anopheline mosquitoes. *Jap J Sanit Zool* 1993, **44**:349–353.
66. El-Shazly MM, Refaie BM: Larvicidal effect of the juvenile hormone mimic pyriproxyfen on *Culex pipiens*. *J Am Mosq Control Assoc* 2002, **18**:321–328.
67. Andrighetti MTM, Cerone F, Riguetti M, Galvani KC, Macoris MLG: Effect of pyriproxyfen in *Aedes aegypti* populations with different levels of susceptibility to the organophosphate temephos. *Dengue Bulletin* 2008, **32**:186–198.
68. Hatakoshi M, Kawada H, Nishida S, Kisida H, Nakayama I: Laboratory evaluation of 2-[1-methyl-2-(4-phenoxyphenoxy)-ethoxy] pyridine against larvae of mosquitoes and housefly. *Jpn J Sanit Zool* 1987, **38**:271–274.
69. Ali A, Chowdhury MA, Hossain MI, Mahmud UI A, Habiba DB, Aslam AF: Laboratory evaluation of selected larvicides and insect growth regulators against field-collected *Culex quinquefasciatus* larvae from urban Dhaka, Bangladesh. *J Am Mosq Control Assoc* 1999, **15**:43–47.
70. Al-Sarar AS, Al-Shahrani D, Bayoumi AE, Abobakr Y, Hussein HI: Laboratory and field evaluation of some chemical and biological larvicides against *Culex* spp. (Diptera: Culicidae) immature stages. *Int J Agr Biol* 2011, **13**:115–119.
71. Kawada H, Dohara K, Shinjo G: Laboratory and field evaluation of an insect growth regulator, 4-phenoxyphenyl (RS)-2-(2-pyridyloxy) propyl ether, as a mosquito larvicide. *Jap J Sanit Zool* 1988, **39**:339–346.
72. Caputo B, Ienco A, Cianci D, Pombi M, Petrarca V, Baseggio A, Devine GJ, della Torre A: The "auto-dissemination" approach: a novel concept to fight *Aedes albopictus* in urban areas. *PLoS One* 2012, **6**:e1793.
73. Ali A, Nayar JK, Xue RD: Comparative toxicity of selected larvicides and insect growth regulators to a Florida laboratory population of *Aedes albopictus*. *J Am Mosq Control Assoc* 1995, **11**:72–76.

74. Nayar JK, Ali A, Zaim M: Effectiveness and residual activity comparison of granular formulations of insect growth regulators pyriproxyfen and s-methoprene against Florida mosquitoes in laboratory and outdoor conditions. *J Am Mosq Control Assoc* 2002, **18**:196–201.
75. Itoh K: Control of DF/DHF vector, *Aedes* mosquito, with insecticides. *Trop Med* 1993, **35**:259–267.
76. Ye-Ebiyo Y, Pollack RJ, Kiszewski A, Spielman A: Enhancement of development of larval *Anopheles arabiensis* by proximity to flowering maize (*Zea mays*) in turbid water and when crowded. *Am J Trop Med Hyg* 2003, **68**:748–752.
77. Gimnig JE, Ombok M, Kamau L, Hawley WA: Characteristics of larval anopheline (Diptera: Culicidae) habitats in western Kenya. *J Med Entomol* 2001, **38**:282–288.
78. Gimnig JE, Ombok M, Otieno S, Kaufman MG, Vulule JM, Walker ED: Density-dependent development of *Anopheles gambiae* (Diptera: Culicidae) larvae in artificial habitats. *J Med Entomol* 2002, **39**:162–172.
79. Kaufman MG, Wanja E, Maknoja S, Bayoh NM, Vulule JM, Walker ED: Importance of algal biomass to growth and development of *Anopheles gambiae* larvae. *J Med Entomol* 2006, **43**:669–676.
80. Mulligan FS III, Schaefer CH: Efficacy of a juvenile hormone mimic, pyriproxyfen (S-31183) for mosquito control in dairy waste water lagoons. *J Am Mosq Control Assoc* 1990, **6**:89–92.
81. Mohsen ZH, Zayia HH: Long-term sublethal effects of fenoxycarb against *Culex* mosquitoes (Diptera: Culicidae). *Jpn J Sanit Zool* 1995, **46**:151–154.
82. Gaugler R, Suman D, Wang Y: An autodissemination station for the transfer of an insect growth regulator to mosquito oviposition sites. *Med Vet Entomol* 2012, **26**:37–45.
83. Vasuki V: Influence of IGR treatment on oviposition of three species of vector mosquitoes at sublethal concentrations. *Southeast Asian J Trop Med Public Health* 1999, **30**:200–203.
84. Ohashi K, Nakada K, Miyaguchi J, Shono Y, Lucas JR, Mito N: Efficacy of pyriproxyfen-treated nets in sterilizing and shortening the longevity of *Anopheles gambiae* (Diptera: Culicidae). *J Med Entomol* 2012, **49**:1052–1058.
85. Judson CL, de Lumen HZ: Some effects of juvenile hormone and analogues on the ovarian follicles of the mosquito *Aedes aegypti* (Diptera: Culicidae). *J Med Entomol* 1976, **13**:197–201.
86. Fournet F, Sannier C, Monteny N: Effects of the insect growth regulators OMS 2017 and diflubenzuron on the reproductive potential of *Aedes aegypti*. *J Am Mosq Control Assoc* 1993, **9**:426–430.
87. Crowder DW, Ellers-Kirk C, Yafuso CM, Dennehy TJ, Degain BA, Harpold VS, Tabashnik BE, Carriere Y: Inheritance of resistance to pyriproxyfen in *Bemisia tabaci* (Hemiptera: Aleyrodidae) males and females (B Biotype). *J Econ Entomol* 2008, **101**:927–932.
88. Karatolos NK, Williamson MS, Denholm I, Gorman K, Richard H-C, Bass C: Over-expression of a cytochrome P450 is associated with resistance to pyriproxyfen in the greenhouse whitefly *Trialeurodes vaporariorum*. *PLoS One* 2012, **7**:e31077.

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