

RESEARCH

Open Access

# Evaluation of prevalence's of *pfdhfr* and *pfdhps* mutations in Angola

Filomeno Fortes<sup>1,3</sup>, Rafael Dimbu<sup>1,3</sup>, Paula Figueiredo<sup>2,3</sup>, Zoraima Neto<sup>2,3</sup>, Virgílio E do Rosário<sup>2,3</sup>, Dinora Lopes<sup>2,3\*</sup>

## Abstract

**Background:** Malaria is the major cause of morbidity and mortality in Angola. The most vulnerable groups to *Plasmodium falciparum* infection are pregnant women and children under five years of age. The use of an intermittent preventive treatment (IPT) with sulphadoxine/pyrimethamine (SP) in pregnant women was introduced in Angola in 2006 by the National Malaria Control Programme, and currently this strategy has been considered to be used for children malaria control. Considering the previous wide use of SP combination in Angola, together to the reported cases of SP treatment failure it is crucial the evaluation of the prevalence of five mutations in *pfdhfr* and *pfdhps* genes associated to *P. falciparum* resistance to SP before the introduction of S/P IPT in children.

**Methods:** The study was conducted in five provinces, with different transmission intensities: Huambo, Cabinda, Uíge, Kwanza Norte, and Malanje. The detection of the mutations in *pfdhfr* and *pfdhps* genes was carried out in 452 *P. falciparum* blood samples by PCR RFLP.

**Results:** For *pfdhfr* gene, 90,3% of the samples carried the mutation 51I, with 7.5% of mixed infections; 51% carried wild type allele 59C, with 29.2% mixed infections and; 99.1% of isolates harboured the mutant allele 108N. Concerning, *pfdhps* gene, 83,1% were mutant type 437G with 11% mixed infections, while 87% of the studied isolates were wild type for codon 540.

**Discussion:** This is the first representative epidemiological study of the whole Angola country on the prevalence of the genotypes associated with SP chemoresistance. A high frequency of individual mutations in both genes (51I and 108N in *pfdhfr*, and 437G in *pfdhps*) was found, besides a low prevalence of the quintuple mutation.

**Conclusion:** The data showed that the implementation IPT using SP in children needs to be reviewed.

## Background

According to the Angolan National Malaria Control Programme (NMCP), malaria is the major cause of morbidity and mortality in Angola, with four million clinical cases and 20 thousand deaths reported in 2005, accounting for 35% of the overall mortality in children under five years old and 25% of the maternal deaths [1-3]. Malaria is endemic throughout the Angolan territory, *Plasmodium falciparum* being the predominant species [4]. Due to the high prevalence of *P. falciparum* strains resistant to chloroquine [5-8], therapeutic regimens for treatment of uncomplicated *P. falciparum* infection were changed in 2006 [9] and, currently, the first-line treatment for uncomplicated malaria is Coartem<sup>®</sup> (artemether-

lumefantrine) followed by the amodiaquine-artesunate alternative therapy.

Additionally, in all Angolan endemic areas the strategy to protect mothers during their pregnancy includes the use of an intermittent preventive treatment (IPT) [10,11]. This intervention has been introduced in Angola since 2006, using sulphadoxine/pyrimethamine (SP) at the second trimester of pregnancy.

In other African countries, SP - ITP has been also introduced in children as a control measure to reduce malaria morbidity and mortality in this most vulnerable population [12] and has been evaluated in a number of clinical trials in these countries, with success [12-17]. Thus, Angolan NMCP intends in the near future to introduce this control measure in Angola. However, due to the wide use of SP combination in this country together to reported cases of SP treatment failure, it was decided to obtain further information about SP resistance

\* Correspondence: dferreira@ihmt.unl.pt

<sup>2</sup>UEI Malária/Centro de Malária e Doenças Tropicais - LA/IHMT/Universidade Nova de Lisboa, Rua da Junqueira, 100, 1349-008, Lisboa, Portugal  
Full list of author information is available at the end of the article

in Angola, during a surveillance study carried out in 2007, before the introduction of such a control measure.

It is well known that mutations at the dihydropteroate synthase (*pfdhps*) and dihydrofolate reductase (*pfdhfr*) genes are associated with resistance of *P. falciparum* to SP, respectively [18-21]. In *pfdhfr*, point mutations at positions 51, 59, 108, and 164 are associated with pyrimethamine resistance [22,23]. Similarly, mutations in codons 437 (437G) and 540 (540E) of *pfdhps* are associated with resistance to sulphadoxine [24-29].

Thus, to determine the polymorphism of *pfdhps* and *pfdhfr* genes, infected blood samples were collected in different representative endemic regions of the whole country (Uíge, Kwanza Norte, Malanje, Cabinda and Huambo) and the prevalence of five mutations of the *pfdhfr* (N51I, C59R and S108N) and *pfdhps* (A437G and K540E) genes was investigated.

## Methods

### Sample collection and DNA extraction

The blood samples used for this study were collected originally as part of malaria surveillance activities of NMCP. Community-based surveys were conducted in five areas with different transmission intensity: Huambo, Cabinda, Uíge, Kwanza Norte, and Malanje (Figure 1). These samples were collected from asymptomatic

children under five years of age, at time of blood collection. Each sample consisted of 200 µl of finger-prick blood spotted on filter papers, dried and stored at room temperature until parasite DNA extraction.

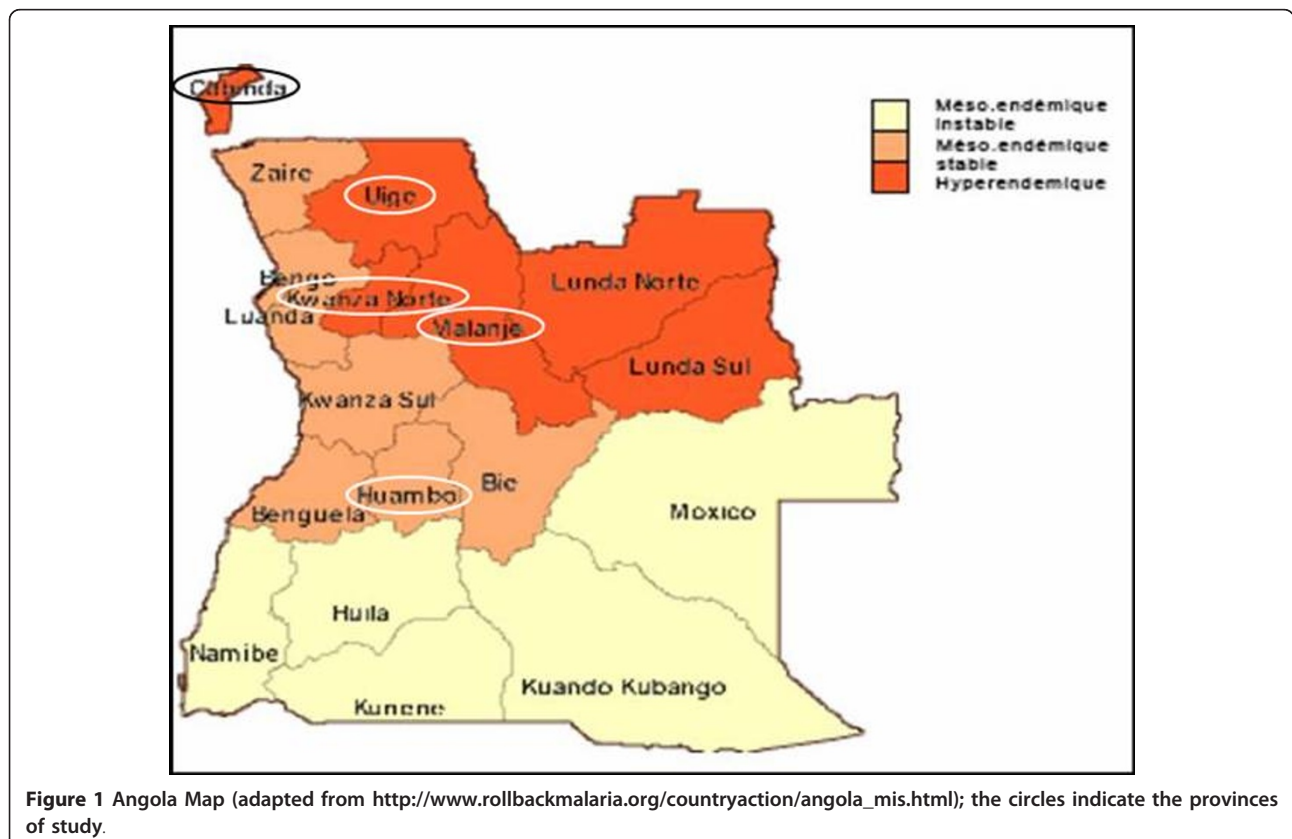
Parent's informed consent was obtained before inclusion in the study, which was reviewed and approved by the Ethical Committee from the Ministry of Health of Angola. DNA was extracted from blood spotted on filter paper, using phenol-chloroform method as described elsewhere [30].

### Analysis of mutations

Polymorphisms on the codons 51, 59, 108 in *pfdhfr* and 437, 540 in *pfdhps* were assessed by PCR RFLP after amplification of DNA fragments by nested-PCR. The PCR and enzymes restrictions reactions were carried out under the conditions already described [31,32]. PCR amplicons and digested fragments were separated on 2% or 3% agarose gels stained with ethidium bromide and visualized under UV.

## Results

From one thousand and twelve samples collected in five provinces from children that did not exhibit malaria symptoms at the time of blood collection, 452 *P. falciparum* PCR positive samples were analysed in



this study: 208 (46%) were collected in Malanje, 96 (21%) in Kwanza Norte, 71 (16%) in Cabinda, 54 (12%) in Uíge and 23 (5%) in Huambo provinces.

For *pfdhfr* gene, all 452 samples were successfully characterized by PCR-RFLP for 51, 423 and 59 codons and 430 samples for codon 108. Regarding *pfdhps* gene, 438 samples were characterized for codon 437 and 448 samples for codon 540. The analysis of *pfdhfr* showed that 90,3% (408 out 452) of the isolates carried the mutant allele 51I, while 7,5% (34 out 452) carried a mixed infection (N and I); for 59 codon 51% (213 out 423) were wild type (C59), 29,2% (122 out 423) were mixed infections (C and R) and 19,9% (83 out 423) carried the mutant allele 59R. Concerning the *pfdhfr* gene codon 108, 99,1% (426 out 430) of isolates harbored the mutant allele (N). For *pfdhps* 83,1% (364 out 438) were mutant type (437G), 11% (48 out 438) were mixed populations and 87% (390 out 448) of studied isolates were wild type for codon 540 (K) (Table 1).

A mixture of infections with wild-type and mutant alleles was also observed. These mixed infections were seen for *pfdhfr* gene in positions 51 (34/452), 59 (122/416) and 108 (1/430), and in *pfdhps* gene in positions 437 (45/438) and 540 (29/441). All mixed infections were excluded from subsequent analysis. Therefore, successful characterization of all five molecular markers was obtained only in 241 samples out of a total of 452.

Only 25% (72) of the 400 isolates which were successfully characterized for the studied *pfdhfr* codons

harboured the triple *pfdhfr* 51-59-108 mutations and one isolate carried the combination of three wild type codons. The more frequent *pfdhfr* genotype was ICN, which was found in 200 isolates (Table 2). Regarding the *pfdhps* genotype GK, double mutation (437 and 540 codons) showed the highest frequency (91,1%). 225 isolates harboured mutations on position 108 of *pfdhfr* gene and 437 of *pfdhps* gene that were reported as the initial mutations for pyrimethamine and sulphadoxine resistance, respectively [23-36].

In *pfdhfr* gene, five mutant genotypes, NCN, NRN, ICN, IRN and NCS (amino acids at positions 51, 59 and 108) were found confirming the major diversity of this gene (Table 2). Among the studied isolates, 74% were double mutants (ICN or NRN), most of them being type ICN, and the triple mutant IRN was detected in 25% of the samples. Only one isolate was a single mutant (ICS). In *pfdhps*, three allele combinations GK, GE and AK (amino acids at positions 437 and 540) were detected nearly 3% being the double mutant GE and 91% of the isolates were GK and 6% were wild-type (AK). Considering the two studied genes, 12 different genotype combinations were found: NRN GK, ICN GK, IRN GK, IRN GE, ICN AK, IRN AK, ICN GE, NCN AK, NRN AK, ICS GK, NCN GK and NCS GK (51, 59 and 108 for *pfdhfr* gene and 437 and 540 for *pfdhps* gene). From a total of 241 isolates, 63% were ICN GK, 25% IRN GK, 3,7% were ICN AK, ICN GE. NRN GK and IRN AK were detected with same frequency of the 1,7%, 1,2%

**Table 1 Prevalence of mutations conferring resistance to SP in *P. falciparum* isolates from Angola**

		Prevalence of mutations in <i>pfdhfr</i> and <i>pfdhps</i> genes (%)				
Province	Alleles	<i>pfdhfr</i>			<i>pfdhps</i>	
		51 n (%)	59 n (%)	108 n (%)	437 n (%)	540 n (%)
Cabinda	wild type	0 (0)	16 (27,6)	0 (0)	8 (12,1)	55 (83,3)
	mutant	70 (98,6)	28 (48,3)	64 (100)	57 (86,4)	3 (4,5)
	mix infection	1 (1,4)	14 (24,1)	0 (0)	1 (1,5)	8 (12,1)
Uíge	wild type	1(1,85)	19 (35,8)	0(0)	0 (0)	53 (98,1)
	mutant	52 (96,3)	15 (28,3)	50 (100)	52 (96,3)	1 (1,9)
	mix infection	1 (1,85)	19 (35,8)	0 (0)	2 (3,7)	0 (0)
Kwanza Norte	wild type	2 (2,1)	46 (49,5)	0 (0)	0 (0)	92 (95,8)
	mutant	91 (94,8)	25 (26,9)	92 (100)	93 (96,9)	2 (2,1)
	mix infection	3 (3,1)	22 (23,7)	0 (0)	3 (3,1)	2 (2,1)
Malanje	wild type	7 (3,4)	127 (62,6)	3 (1,5)	16 (7,7)	189 (90,9)
	mutant	173 (83,2)	12 (5,9)	199 (98)	156 (75)	5 (2,4)
	mix infection	28 (13,5)	64 (31,5)	1 (0,5)	34 (16,3)	13 (6,3)
Huambo	wild type	0 (0)	5 (45,5)	0 (0)	2 (12,5)	11 (61,1)
	mutant	22 (95,7)	3 (27,3)	21 (100)	6 (37,5)	1 (5,6)
	mix infection	1 (4,4)	3 (27,3)	0 (0)	8 (50)	6 (33,3)
Total	wild type	10 (2,2)	213 (51,0)	3 (0,7)	26 (5,9)	390 (87,1)
	mutant	408 (90,3)	83 (19,9)	426 (99,1)	364 (83,1)	29 (6,5)
	mix infection	34 (7,5)	122 (29,2)	1 (0,2)	48 (11,0)	29 (6,5)

**Table 2 Prevalence of haplotypes in *P. falciparum* isolates from Angola**

Genotypes	Cabinda		Uíge		Kwanza Norte		Malanje		Huambo		Total	
	n	(%)	n	(%)	n	(%)	n	(%)	n	(%)	n	(%)
51 59 108												
<i>pf dhfr</i>												
IRN	28	63,6	13	40,6	22	32,4	5	3,9	1	16,7	<b>69</b>	24,8
ICN	16	36,4	18	56,3	44	64,7	117	91,4	5	83,3	<b>200</b>	71,9
ICS		0,0		0,0		0,0	1	0,8		0,0	<b>1</b>	0,4
NCN		0,0		0,0		0,0	2	1,6		0,0	<b>2</b>	0,7
NRN		0,0	1	3,1	2	2,9	3	2,3		0,0	<b>6</b>	2,2
<b>n</b>	<b>44</b>		<b>32</b>		<b>68</b>		<b>128</b>		<b>6</b>		<b>278</b>	
437 540												
<i>pf dhps</i>												
GK	50	87,7	51	98,1	89	94,7	144	89,4	5	62,5	<b>339</b>	91,1
GE	3	5,3	1	1,9	2	2,1	5	3,1	1	12,5	<b>12</b>	3,2
AK	4	7,0	0	0,0	3	3,2	12	7,5	2	25,0	<b>21</b>	5,6
<b>n</b>	<b>57</b>		<b>52</b>		<b>94</b>		<b>161</b>		<b>8</b>		<b>372</b>	
51 59 108/437 540												
<i>pf dhfr/pf dhps</i>												
ICN/GK	10	27,8	17	54,8	41	60,3	83	81,4	1	25,0	<b>152</b>	63,1
IRN/GK	22	61,1	14	45,2	20	29,4	4	3,9	1	25,0	<b>61</b>	25,3
IRN/AK	2	5,6		0,0	2	2,9		0,0		0,0	<b>4</b>	1,7
ICN/AK		0,0		0,0	1	1,5	6	5,9	2	50,0	<b>9</b>	3,7
NRN/GK		0,0		0,0	2	2,9	1	1,0		0,0	<b>3</b>	1,2
ICN/GE	1	2,8		0,0	2	2,9		0,0		0,0	<b>3</b>	1,2
ICS/GK		0,0		0,0		0,0	1	1,0		0,0	<b>1</b>	0,4
ICN/GE		0,0		0,0		0,0	4	3,9		0,0	<b>4</b>	1,7
NCN/GK		0,0		0,0		0,0	1	1,0		0,0	<b>1</b>	0,4
NRN/AK		0,0		0,0		0,0	1	1,0		0,0	<b>1</b>	0,4
NCN/AK		0,0		0,0		0,0	1	1,0		0,0	<b>1</b>	0,4
IRN/GE	1	2,8		0,0		0,0		0,0		0,0	<b>1</b>	0,4
<b>n</b>	<b>36</b>		<b>31</b>		<b>68</b>		<b>102</b>		<b>4</b>		<b>241</b>	

were NRN GK and ICN GE, all other combinations were found with a very low frequency (Table 2).

In a comparison evaluation between provinces, the same pattern was found except for Cabinda, where the most frequent genotype was IRN (Tables 1 and 2). This province is geographically separated from the rest of the country.

## Discussion

The monitoring of SP resistance is relevant in order to guide national malaria treatment policies before introduction of SP as IPT in children at Angola. In this light, this study was designed to assess the *pf dhfr* and *pf dhps* mutations associated with SP chemoresistance. For this purpose, the mutations at *pf dhfr* (N51I, C59R and S108N) and *pf dhps* (A437G and K540E) genes, considered predictive of SP treatment failure [25-27], were assessed in five provinces of Angola. Four of them - Uíge, Kwanza Norte, Malanje and Cabinda - are hyper-endemic areas, whereas Huambo, is a mesoendemic stable. In this way, it was observed the presence of the mutations 51I and 59R jointly with 108N, which enhances the level of resistance to pyrimethamine when

compared with single mutation in codon 108 [35-37]. Similarly, mutations in 437 (437G) and 540 (540E) *pf dhps* codons are associated with resistance to sulphadoxine [24-29]. In *pf dhfr* gene, the mutation at position 108 (S108N), which is believed to be the initial mutation causing pyrimethamine resistance, was observed in almost all isolates successfully characterized for this codon (426/430) (Table 1). In *pf dhps* gene, among the 438 characterized isolates, 364 presented the mutant allele at position 437, which has been reported as the initial mutation for sulphadoxine resistance in many endemic regions.

From 430 *P. falciparum* isolates characterized in this study, 99,1% carried the *pf dhfr* 108N mutation. The results also showed that 27% of *P. falciparum* isolates presented double mutations at codons 59 and 108, indicating the development of resistance against antifolates in Angola. Another double mutant (51I 108N) was observed in 96,7% of Angolan *P. falciparum* isolates; these results were consistent with the results obtained in a similar study carried out at Uíge province in 2009 [38].

A 25% of *pf dhfr* triple mutant prevalence (51I/59R/108N) was also noticed. The prevalence of these



mutations in association with high prevalence of mutation at position 437G in *pfdhps* (83,1%) may indicate that these *P. falciparum* parasite populations have the potential to evolve into *pfdhfr/pfdhps* quintuple mutant in the near future, a mutant that is considered a molecular marker of SP treatment failure [25-27].

In addition, the simultaneously presence of 59R *pfdhfr* and 540E *pfdhps* variants is considered predictive of the presence of quintuple mutant (*pfdhfr* 51I, 59R, 108N, *pfdhps* 437G, 540E) [25-40]. In fact, despite high frequencies of mutations in this report, only one isolate was found harbouring the quintuple mutant associated with high level of resistance to SP. This finding is in accordance with the results obtained in other similar studies carried out in Angola, Republic of Congo and Gabon [31,28,29].

The predominant *pfdhfr* haplotype in the present work was 51I59R108N (71,9%), corroborating the results reported with Brazilian samples by Gama and collaborators [41]. The triple mutant 51I59R108N (24,8%) was of low prevalence in the examined isolates, similarly to previous data reported in Sri Lanka [42] and Papua New Guinea [43], but different from the isolates from Malaysia [44], Brazil [45] and India [46] where this triple mutant was the predominant haplotype. In Africa, the Republic of Congo [28] and Gabon [29] also shows differences when compared with these Angolan data, except in the province of Cabinda. The differences found between Cabinda isolates and the rest of studied isolates may be due to geographical location of this province and its proximity to the neighboring countries Gabon and Congo and the movement of people between these regions. Most of the *P. falciparum* isolates (91,1%) revealed the haplotype 437G540K for *pfdhps* gene with low prevalence of mutation at codon 540 (3,2%).

The triple *pfdhfr/pfdhps* (59R108N/437G) mutant haplotype was found in 27% of isolates, the 51I108N/437G was the mutant haplotype more prevalent in studied isolates.

The results obtained in the present study are in line with those obtained with Iranian isolates where 51I mutation seems to be a good molecular marker for the triple mutant *pfdhfr/pfdhps* [47]. By the other hand, these results seem to be in contrast with the data obtained in a study carried out in Mozambique which claimed that the mutations at codon 59 in *pfdhfr* and codon 437 in *pfdhps* were enough to predict SP treatment failure [48], as well as in Burkina Faso [49] where the results showed that *pfdhfr* 59R is more relevant than the 51I as a marker of SP treatment failure.

Finally, this is the first molecular study carried out in Angola including a large number of samples (452) from five different provinces of the country, and where five mutations of *pfdhps* and *pfdhfr* genes, predictive of SP therapeutic failure were screening showing high

frequencies of 51I, 108N *pfdhfr* alleles, and 437G of *pfdhps* gene with an almost absence of the quintuple mutation for SP.

## Conclusion

The high frequencies of mutations and haplotypes linked to antimalarial treatment failure (for example the 88% frequency of isolates carrying *pfdhfr* 51I108N - *pfdhps* 437G triple mutant allele, critical to SP resistance) herein described, highlight the need to reevaluate the strategy of SP introduction as an IPT in children as well as the current use of SP for pregnant women IPT purposes, in Angola.

## Acknowledgements

This study has financial support from PNCM/MINSA. The authors are grateful to the children involved in the study and the staff of PNCM who collaborated in sample collection.

## Author details

<sup>1</sup>Programa Nacional de Controlo da Malária/Ministério da Saúde de Angola, Luanda, Angola. <sup>2</sup>UEI Malária/Centro de Malária e Doenças Tropicais - LA/IHMT/Universidade Nova de Lisboa, Rua da Junqueira, 100, 1349-008, Lisboa, Portugal. <sup>3</sup>Health Progress and Investigation Network of Portuguese-Speaking Countries Community (RIDES/CPLP), Centro de Malária e Doenças Tropicais - LA, Instituto de Higiene e Medicina Tropical, Universidade Nova de Lisboa, Lisboa, Portugal.

## Authors' contributions

FF coordinated sample collection. FF, RD carried out the selection of children and sample collection. RD, ZN and DL carried out DNA extraction and *Plasmodium* species identification. PF carried out the molecular analyses. FF, VEdR and DL coordinated and designed the study. DL drafted this manuscript. All authors read and approved the final manuscript.

## Competing interests

The authors declare that they have no competing interests.

Received: 9 August 2010 Accepted: 2 February 2011

Published: 2 February 2011

## References

1. World Health Organization/UNICEF: *World Malaria Report 2005*. 2005 [http://whqlibdoc.who.int/publications/2005/9241593199\_eng.pdf].
2. Brabin BJ: *An analysis of malaria in pregnancy in Africa*. *Bull World Health Organ* 1983, **61**:1005-1016.
3. Olsen W, ter Kuile F, Maelankirri L, Decludt B, White NJ: *Malaria during pregnancy in an area of unstable endemicity*. *Trans R Soc Trop Med Hyg* 1991, **85**:424-429.
4. World Health Organization: *Communicable Disease Toolkit for Angola*. 2005 [http://whqlibdoc.who.int/hq/2005/WHO\_CDS\_NTD\_DCE\_2005a\_profile.pdf].
5. Kyronseppa H, Lumio J, Ukkonen R, Pettersson T: *Chloroquine-resistant malaria from Angola*. *Lancet* 1984, **1**:1244.
6. Nosten W, Jensen T, Jorgensen M: *Chloroquine-resistant Plasmodium falciparum malaria from Angola*. *Lancet* 1984, **1**:1462-463.
7. Lindberg J, Sandberg T, Bjorkholm B, Bjorkman A: *Chloroquine and Fansidar resistant malaria acquired in Angola*. *Lancet* 1985, **1**:765.
8. Guthmann JP, Ampuero J, Fortes F, van Overmeir C, Gaboulaud V, Tობback S, Dunand J, Saraiva N, Gillet P, Franco J, Denoncin A, van Herp M, Balkan S, Dujardin JC, D'Alessandro U, Legros D: *Antimalarial efficacy of chloroquine, amodiaquine, sulfadoxine-pyrimethamine, and the combinations of amodiaquine + artesunate and sulfadoxine-pyrimethamine + artesunate in Huambo and Bie provinces, central Angola*. *Trans R Soc Trop Med Hyg* 2005, **99**:485-492.

9. World Health Organization: **Global AMD database. AFRO 2005** [http://www.who.int/countries/en/].
10. Nahlen BL: **Rolling back malaria in pregnancy.** *N Engl J Med* 2000, **343**:651-652.
11. WHO: **A strategic Framework for Malaria Prevention and Control during Pregnancy in the Africa Region.** 2004 [http://whqlibdoc.who.int/afro/2004/AFR\_MAL\_04.01.pdf].
12. Schellenberg DM, Aponte JJ, Kahigwa EA, Mshinda H, Tanner M, Menendez C, Alonso PL: **The incidence of clinical malaria detected by active case detection in children in Ifakara, southern Tanzania.** *Trans R Soc Trop Med Hyg* 2003, **97**:647-54.
13. Massaga JJ, Kitua AY, Lemnge MM, Akida JA, Malle LN, Rønn AM, Theander TG, Bygbjerg IC: **Effect of intermittent treatment with amodiaquine on anaemia and malarial fevers in infants in Tanzania: a randomised placebo-controlled trial.** *Lancet* 2003, **361**:1853-1860.
14. Chandramohan D, Owusu-Agyei S, Carneiro I, Awine T, Amponsa-Achiano K, Mensah N, Jaffar S, Baiden R, Hodgson A, Binka F, Greenwood B: **Cluster randomised trial of intermittent preventive treatment for malaria in infants in area of high, seasonal transmission in Ghana.** *BMJ* 2005, **331**:727-33.
15. White NJ: **Antimalarial drug resistance.** *J Clin Invest* 2004, **113**:1084-92.
16. Macete E, Aide P, Aponte JJ, Sanz S, Mandomando I, Espasa M, Sigauque B, Dobaño C, Mabunda S, DgeDge M, Alonso P, Menendez C: **Intermittent preventive treatment for malaria control administered at the time of routine vaccinations in Mozambican infants: a randomized, placebo-controlled trial.** *J Infect Dis* 2006, **194**:276-285.
17. Kobbe R, Adjei S, Kreuzberg C, Kreuels B, Thompson B, Thompson PA, Marks F, Busch W, Tosun M, Schreiber N, Opoku E, Adjei O, Meyer CG, May J: **Malaria incidence and efficacy of intermittent preventive treatment in infants (IPTi).** *Malar J* 2007, **6**:163.
18. Wongsrichanalai C, Pickard AL, Wernsdorfer WH, Meshnick SR: **Epidemiology of drug-resistant malaria.** *Lancet Infect Dis* 2002, **2**:209-218.
19. Le Bras J, Durand R: **The mechanisms of resistance to antimalarial drugs in *Plasmodium falciparum*.** *Fundam Clin Pharmacol* 2003, **17**:147-153.
20. Jelinek T, Rønn AM, Lemnge MM, Curtis J, Mhina J, Duraisingh MT, Bygbjerg IC, Warhurst DC: **Polymorphisms in the dihydrofolate reductase (DHFR) and dihydropteroate synthetase (DHPS) genes of *Plasmodium falciparum* and in vivo resistance to sulphadoxine/pyrimethamine in isolates from Tanzania.** *Trop Med Int Health* 1998, **3**:605-609.
21. Ngo T, Duraisingh M, Reed M, Hipgrave D, Biggs B, Cowman AF: **Analysis of *pfprt*, *pfmdr1*, *dhfr*, and *dhps* mutations and drug sensitivities in *Plasmodium falciparum* isolates from patients in Vietnam before and after treatment with artemisinin.** *Am J Trop Med Hyg* 2003, **68**:350-356.
22. Cowman AF, Morry MJ, Biggs BA, Cross GA, Foote SJ: **Amino acid changes linked to pyrimethamine resistance in the dihydrofolate reductase-thymidylate synthase gene of *Plasmodium falciparum*.** *Proc Natl Acad Sci USA* 1988, **85**:9109-9113.
23. Peterson DS, Walliker D, Welles TE: **Evidence that a point mutation in dihydrofolate reductase-thymidylate synthase confers resistance to pyrimethamine in *falciparum* malaria.** *Proc Natl Acad Sci USA* 1988, **85**:9114-9118.
24. Appawu M, Owusu-Agyei S, Dadzie S, Asoala V, Anto F, Koram K, Rogers W, Nkrumah F, Hoffman SL, Fryauff DJ: **Malaria transmission dynamics at a site in northern Ghana proposed for testing malaria vaccines.** *Trop Med Int Health* 2004, **9**:164-170.
25. Bwijo B, Kaneko A, Takechi M, Zungu IL, Moriyama Y, Lum JK, Tsukahara T, Mita T, Takahashi N, Bergquist Y, Björkman A, Kobayakawa T: **High prevalence of quintuple mutant *dhps/dhfr* genes in *Plasmodium falciparum* infections seven years after introduction of sulfadoxine and pyrimethamine as first line treatment in Malawi.** *Acta Trop* 2003, **85**:363-373.
26. Kublin JG, Dzinjalimala FK, Kamwendo DD, Malkin EM, Cortese JF, Martino LM, Mukadam RA, Rogerson SJ, Lescano AG, Molyneux ME, Winstanley PA, Chimpeni P, Taylor TE, Plowe CV: **Molecular markers for failure of sulfadoxine-pyrimethamine and chlorproguanil-dapsone treatment of *Plasmodium falciparum* malaria.** *J Infect Dis* 2002, **185**:380-388.
27. Nzila AM, Mberu EK, Sulo J, Dayo H, Winstanley PA, Sibley CH, Watkins WM: **Towards an understanding of the mechanism of pyrimethamine-sulfadoxine resistance in *Plasmodium falciparum*: genotyping of dihydrofolate reductase and dihydropteroate synthase of Kenyan parasites.** *Antimicrob Agents Chemother* 2000, **44**:991-996.
28. Nsimba B, Jafari-Guemouri S, Malonga DA, Mouata AM, Kiori J, Louya F, Yocka D, Malanda M, Durand R, Le Brás J: **Epidemiology of drug-resistant malaria in Republic of Congo: using molecular evidence for monitoring antimalarial drug resistance combined with assessment of antimalarial drug use.** *Trop Med Int Health* 2005, **10**:1030-1037.
29. Aubouy A, Jafari S, Huart V, Migot-Nabias F, Mayombo J, Durand R, Bakary M, Le Bras J, Deloron P: **DHFR and DHPS genotypes of *Plasmodium falciparum* isolates from Gabon correlate with in vitro activity of pyrimethamine and cycloguanil, but not with sulfadoxine-pyrimethamine treatment efficacy.** *J Antimicrob Chemother* 2003, **52**:43-49.
30. Snounou G, Viriyakosol S, Zhu XP, Jarra W, Pinheiro L, do Rosario VE, Thaithongs S, Brown KN: **High sensitivity of detection of human malaria parasites by the use of nested polymerase chain reaction.** *Mol Biochem Parasitol* 1993, **61**:315-320.
31. Figueiredo P, Benchimol C, Lopes D, Bernardino L, do Rosário VE, Varandas L, Nogueira F: **Prevalence of *pfmdr1*, *pfprt*, *pfdfhr* and *pfdhps* mutations associated with drug resistance, in Luanda, Angola.** *Malar J* 2008, **7**:236.
32. Cravo P, Figueiredo S, Nogueira F, Lopes D, Ferreira ID, Ferreira C, do Rosario VE: **High frequency of the genetic polymorphisms associated with sulfadoxine-pyrimethamine resistance, among *Plasmodium falciparum* isolates from Sao Tome and Principe, West Africa.** *Ann Trop Med Parasitol* 2004, **98**:293-296.
33. Mockenhaupt FP, Eggelte TA, Till H, Bienze U: ***Plasmodium falciparum* *pfprt* and *pfmdr1* polymorphisms are associated with the *pfdfhr* N108 pyrimethamine-resistance mutation in isolates from Ghana.** *Trop Med Int Health* 2001, **6**:749-755.
34. Durand R, Jafari S, Bouchaud O, Ralaimazava P, Keundjian A, Le Bras J: ***Plasmodium falciparum*: *pfprt* and DHFR mutations are associated with failure of chloroquine plus proguanil prophylaxis in travelers.** *J Infect Dis* 2001, **184**:1633-1634.
35. Tarnchompoo B, Sirichaiwat C, Phupong W, Intaradom C, Sirawaraporn W, Kamchonwongpaisan S, Vanichanankul J, Thebtaranonth Y, Yuthavong Y: **Development of 2,4-diaminopyrimidines as antimalarials based on inhibition of the S108N and C59R+S108N mutants of dihydrofolate reductase from pyrimethamine resistant *Plasmodium falciparum*.** *J Med Chem* 2002, **45**:1244-1252.
36. Mourier T, Pain A, Barrell B, Griffiths-Jones S: **A selenocysteine tRNA and SECIS element in *Plasmodium falciparum*.** *RNA* 2005, **11**:119-122.
37. Sirawaraporn W, Sathitkul T, Sirawaraporn R, Yuthavong Y, Santi DV: **Antifolate-resistant mutants of *Plasmodium falciparum* dihydrofolate reductase.** *Proc Natl Acad Sci USA* 1997, **94**:1124-1129.
38. Menegon M, Pearce RJ, Inojosa WO, Pisani V, Abel PM, Matondo A, Bisoffi Z, Majori G, Ord R, Warhurst DC, Roper C, Severini C: **Monitoring for multidrug-resistant *Plasmodium falciparum* isolates and analysis of pyrimethamine resistance evolution in Uige province, Angola.** *Trop Med Int Health* 2009, **14**:1251-1257.
39. Mayor A, Serra-Casas E, Sanz S, Aponte JJ, Macete E, Mandomando I, Puyol L, Berzosa P, Dobaño C, Aide P, Sacarlal J, Benito A, Alonso P, Menéndez C: **Molecular markers of resistance to sulfadoxine-pyrimethamine during intermittent preventive treatment for malaria in Mozambican infants.** *J Infect Dis* 2008, **197**:1737-1742.
40. Mbugi EV, Mutayoba BM, Malisa AL, Balthazary ST, Nyambo TB, Mshinda H: **Drug resistance to sulphadoxine-pyrimethamine in *Plasmodium falciparum* malaria in Mlimba, Tanzania.** *Malar J* 2006, **5**:94.
41. Gama BE, de Oliveira NK, Zalis MG, de Souza JM, Santos F, Daniel-Ribeiro CT, Ferreira-da-Cruz Mde F: **Chloroquine and sulphadoxine-pyrimethamine sensitivity of *Plasmodium falciparum* parasites in a Brazilian endemic area.** *Malar J* 2009, **8**:156.
42. Hapuarachchi HC, Dayanath MY, Bandara KB, Abeyesundara S, Abeyewickreme W, de Silva NR, Hunt SY, Sibley CH: **Point mutations in the dihydrofolate reductase and dihydropteroate synthase genes of *Plasmodium falciparum* and resistance to sulfadoxine-pyrimethamine in Sri Lanka.** *Am J Trop Med Hyg* 2006, **74**:198-204.
43. Mita T, Kaneko A, Hwaihwanje I, Tsukahara T, Takahashi N, Osawa H, Tanabe K, Kobayakawa T, Björkman A: **A rapid selection of *dhfr* mutant allele in *Plasmodium falciparum* isolates after the introduction of sulfadoxine/pyrimethamine in combination with 4-aminoquinolines in Papua New Guinea.** *Infect Genet Evol* 2006, **6**:447-452.
44. Cox-Singh J, Zakaria R, Abdullah MS, Rahman HA, Nagappan S, Singh B: **Differences in dihydrofolate reductase but not dihydropteroate synthase**

- alleles in *Plasmodium falciparum* isolates from geographically distinct areas in Malaysia. *Am J Trop Med Hyg* 2001, **64**:28-31.
45. Vasconcelos KF, Plowe CV, Fontes CJ, Kyle D, Wirth DF, Pereira da Silva LH, Zalis MG: Mutations in *Plasmodium falciparum* dihydrofolate reductase and dihydropteroate synthase of isolates from the Amazon region of Brazil. *Mem Inst Oswaldo Cruz* 2000, **95**:721-728.
46. Das MK, Lumb V, Mittra P, Singh SS, Dash AP, Sharma YD: High chloroquine treatment failure rates and predominance of mutant genotypes associated with chloroquine and antifolate resistance among *falciparum* malaria patients from the island of Car Nicobar, India. *J Antimicrob Chemother* 2010, **65**:1258-1261.
47. Zakeri S, Gil JP, Bereckzy S, Djajid ND, Bjorkman A: High prevalence of double *Plasmodium falciparum dhfr* mutations at codons 108 and 59 in the Sistan- Baluchistan Province. *Iran J Infect Dis* 2003, **187**:1828-1829.
48. Alifrangis M, Enosse S, Khalil IF, Tarimo DS, Lemnge MM, Thompson R, Byqbierq IC, Rønn AM: Prediction of *Plasmodium falciparum* resistance to sulfadoxine/pyrimethamine in vivo by mutations in the dihydrofolate reductase and dihydropteroate synthetase genes: a comparative study between sites of differing endemicity. *Am J Trop Med Hyg* 2003, **69**:601-606.
49. Dokomajilar C, Lankoande ZM, Dorsey G, Zongo I, Ouedraogo JB, Rosenthal PJ: Roles of specific *Plasmodium falciparum* mutations in resistance to amodiaquine and sulfadoxine-pyrimethamine in Burkina Faso. *Am J Trop Med Hyg* 2006, **75**:162-165.

doi:10.1186/1475-2875-10-22

Cite this article as: Fortes et al.: Evaluation of prevalence's of *pf dhfr* and *pf dhps* mutations in Angola. *Malaria Journal* 2011 **10**:22.

Submit your next manuscript to BioMed Central  
and take full advantage of:

- Convenient online submission
- Thorough peer review
- No space constraints or color figure charges
- Immediate publication on acceptance
- Inclusion in PubMed, CAS, Scopus and Google Scholar
- Research which is freely available for redistribution

Submit your manuscript at  
www.biomedcentral.com/submit

