BMC Medical Genetics



Research Open Access

Framingham Heart Study genome-wide association: results for pulmonary function measures

Jemma B Wilk*¹, Robert E Walter^{2,3}, Jason M Laramie^{1,4}, Daniel J Gottlieb^{2,3,5} and George T O'Connor^{2,3}

Address: ¹Department of Neurology, Boston University School of Medicine, Boston, MA, USA, ²Pulmonary Center, Department of Medicine, Boston University School of Medicine, Boston, MA, USA, ³The National Heart, Lung, and Blood Institute's Framingham Heart Study, Framingham, MA, USA, ⁴Program in Bioinformatics, Boston University, Boston, MA, USA and ⁵VA Boston Healthcare System, Boston, MA, USA

Email: Jemma B Wilk* - jwilk@bu.edu; Robert E Walter - walterb@bu.edu; Jason M Laramie - laramiej@bu.edu; Daniel J Gottlieb - dgottlieb@lung.bumc.bu.edu; George T O'Connor - goconnor@lung.bumc.bu.edu

* Corresponding author

Published: 19 September 2007

BMC Medical Genetics 2007, 8():S8 doi:10.1186/1471-2350-8-S1-S8

This article is available from: http://www.biomedcentral.com/1471-2350/8/\$1/\$8

© 2007 Wilk et al; licensee BioMed Central Ltd.

This is an open access article distributed under the terms of the Creative Commons Attribution License (http://creativecommons.org/licenses/by/2.0), which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.

Abstract

Background: Pulmonary function measures obtained by spirometry are used to diagnose chronic obstructive pulmonary disease (COPD) and are highly heritable. We conducted genome-wide association (GWA) analyses (Affymetrix 100 K SNP GeneChip) for measures of lung function in the Framingham Heart Study.

Methods: Ten spirometry phenotypes including percent of predicted measures, mean spirometry measures over two examinations, and rates of change based on forced expiratory volume in one second (FEV₁), forced vital capacity (FVC), forced expiratory flow from the 25th to 75th percentile (FEF₂₅₋₇₅), the FEV₁/FVC ratio, and the FEF₂₅₋₇₅/FVC ratio were examined. Percent predicted phenotypes were created using each participant's latest exam with spirometry. Predicted lung function was estimated using models defined in the set of healthy never-smokers, and standardized residuals of percent predicted measures were created adjusting for smoking status, pack-years, and body mass index (BMI). All modeling was performed stratified by sex and cohort. Mean spirometry phenotypes were created using data from two examinations and adjusting for age, BMI, height, smoking and pack-years. Change in pulmonary function over time was studied using two to four examinations with spirometry to calculate slopes, which were then adjusted for age, height, smoking and pack-years.

Results: Analyses were restricted to 70,987 autosomal SNPs with minor allele frequency \geq 10%, genotype call rate \geq 80%, and Hardy-Weinberg equilibrium p-value \geq 0.001. A SNP in the interleukin 6 receptor (*IL6R*) on chromosome I was among the best results for percent predicted FEF_{25–75}. A non-synonymous coding SNP in glutathione S-transferase omega 2 (*GSTO2*) on chromosome 10 had top-ranked results studying the mean FEV_I and FVC measurements from two examinations. SNPs nearby the *SOD3* and vitamin D binding protein genes, candidate genes for COPD, exhibited association to percent predicted phenotypes.

Conclusion: *GSTO2* and *IL6R* are credible candidate genes for association to pulmonary function identified by GWA. These and other observed associations warrant replication studies. This resource of GWA results for pulmonary function measures is publicly available at http://www.ncbi.nlm.nih.gov/projects/gap/cgi-bin/study.cgi?id=phs000007.

Background

Chronic obstructive pulmonary disease (COPD), which affects approximately six percent of the adult US population [1] and is the fourth most common cause of death in the US [2], has been defined as "airflow limitation that is not fully reversible" [3], a definition based on spirometry. Environmental factors, most notably tobacco smoking, are associated with accelerated longitudinal decline of pulmonary function and are important causes of COPD. Several lines of evidence indicate that genetic factors also contribute to the development of this condition. First, family studies have revealed increased risk of lung function impairment in smoking first degree relatives of COPD cases [4] and substantial heritability of spirometry measures in population based studies [5]. Second, severe alpha-1-antitrypsin deficiency due to homozygous mutations of the SERPINA1 (AAT) gene is a documented cause of COPD, although this condition explains only a small proportion of COPD in the population. Finally, familybased and case-control studies are beginning to reveal genetic variants other than those of the SERPINA1 gene that are associated with chronic airflow obstruction [6]. Despite this evidence of a genetic basis for susceptibility to COPD, the specific genetic risk factors underlying most cases of COPD remain uncertain.

The Framingham Heart Study offers the opportunity to conduct family-based linkage and association studies seeking potential genetic factors that influence obstructive (COPD, asthma), restrictive, or developmentally related lung function impairment. Using spirometry measurements as quantitative phenotypes, we have previously reported results of genome-wide linkage analyses among these same families using microsatellite markers spaced approximately 10 centiMorgans apart [7] and fine-mapping of a promising region on chromosome 6q [8,9]. The availability of data on over 100,000 single nucleotide polymorphisms (SNPs) throughout the genome now permits the application of genome-wide association (GWA) testing to the search for genetic risk factors for chronic airflow limitation. The longitudinal pulmonary function data that have been obtained over the years of the Framingham Heart Study, in combination with the 100 K SNP data, make this a unique resource for the discovery of novel genetic risk factors for chronic airflow obstruction.

Methods

Three types of spirometry phenotypes were evaluated for GWA: 1) measurements taken at a participant's most recent available examination and expressed as a percent of predicted; 2) the mean of measurements taken at two specified examinations; and 3) the annual rate of decline of spirometry measurements derived by calculating the slope of measurements across multiple examinations. All

spirometry was performed without bronchodilator testing.

Measurements as percent of predicted at latest examination

The spirometry measurements from each participant's latest examination with acceptable pulmonary function data [10] were used; eligible examinations included Cohort exams 19, 17, and 13 and Offspring exams 7, 6, 5, and 3. Predicted values for each lung function measurement were calculated using cohort and gender-specific regression models predicting spirometry measurements on the basis of age, age squared, and height squared [11] among Framingham subjects who were lifetime nonsmokers and had no history of chronic bronchitis, pulmonary disease, COPD/emphysema, asthma, or wheezing. The percent of predicted value was calculated by dividing the observed by the predicted value. Standardized residuals were then created by regressing the percent predicted on current smoking (y/n), former smoking (y/n), pack-years, and body mass index (BMI: kg/m²), in cohort and gender-specific models. Forced expiratory volume in one second (FEV₁), forced vital capacity (FVC), forced expiratory flow between the 25th and 75th percentile (FEF₂₅₋₇₅), the FEV₁/ FVC ratio, and the FEF₂₅₋₇₅/FVC ratio were examined as cross-sectional percent of predicted measures. These phenotypes are referenced in tables preceded by the letters "pp" (for percent predicted), see Table 1 for explanation of phenotype abbreviations.

Mean of measurements at two specified examinations

In previous analyses of the genetics of lung function in the Framingham families [7,8], we have used the mean of the values of each spirometry measure from two specified examinations. For Cohort participants, spirometry data from exam cycles 5 or 6 and cycle 13 were used to generate the mean value. In Offspring participants, spirometry data from cycle 3 and cycle 5 were used to generate the mean value. The mean FEV₁, FVC, and FEV₁/FVC ratio were adjusted for the effects of age, age², BMI, height, dummy variables indicating never, former, or current smoking status, and, for former and current smokers, pack-years. Standardized residuals were generated separately by sex and within Cohort or Offspring samples. These phenotypes are referenced in tables as meanfev1, meanfvc, and meanratio.

Annual rate of decline of measurements

Rate of decline phenotypes were defined by fitting a slope to the spirometry data from different time points. The examinations incorporated were the same as those eligible for percent of predicted phenotypes described above. Slopes were calculated by ordinary least-squares using all available data. A minimum of two eligible exams were needed to calculate a slope. Slopes were adjusted for the

Table 1: Characteristics of phenotypes studied*

Phenotype	Na	Description	Eligible exam cycles Offspring	Cohort	Adjustment ^b	Heritability
ppfev l	1217	Percent predicted FEV ₁ for latest exam	7, 6, 5, 3	19, 17, 13	Predicted defined by age, age ² , height, % predicted adjusted for current or former smoking, pack-years, and BMI	0.36
ppfvc	1217	Percent predicted FVC for latest exam	7, 6, 5, 3	19, 17, 13	Same as ppfev l	0.45
ppratio	1217	Percent predicted FEV _I /FVC for latest exam	7, 6, 5, 3	19, 17, 13	Same as ppfev1	0.29
ppfef	1212	Percent predicted FEF ₂₅₋₇₅ for latest exam	7, 6, 5, 3	19, 17, 13	Same as ppfev l	0.40
ppfefrat	1212	Percent predicted FEF ₂₅₋₇₅ /FVC for latest exam	7, 6, 5, 3	19, 17, 13	Same as ppfev1	0.41
meanfev l	1222	Mean FEV ₁ from two exams	3 and 5	5 or 6 and 13	age, age ² , BMI, height, current or former smoking, and pack-years	0.35
meanfvc	1222	Mean FVC from two exams	3 and 5	5 or 6 and 13	Same as meanfev l	0.51
meanratio	1222	Mean FEV ₁ /FVC from two exams	3 and 5	5 or 6 and 13	Same as meanfev l	0.25
fev I slope	1097	Longitudinal slope of FEV	7, 6, 5, 3	19, 17, 13	age, age ² , height, height ² , pack-years at first exam, interim pack-years, and sustained smoking	0.10
fefslope	1059	Longitudinal slope of FEV ₂₅₋₇₅	7, 6, 5, 3	19, 17, 13	Same as fev I slope	0.20

^{*}Results for additional longitudinal slope phenotypes, residual from predicted phenotypes, phenotypes created from spirometry at a single exam, and phenotypes limited to a) sample size with phenotype out of 1345 with GWA genotyping

b) adjustment performed in four separate regression models by sex and Original or Offspring cohort

covariates age, age squared, height, and height squared, using mean ages and heights from included exams. Slopes were also adjusted for pack-years at first exam, interim pack-years, and sustained smoking (y/n). FEV₁ and FEF₂₅. 75, the slope phenotypes with the highest heritability, were studied in the GWA. These phenotypes are referenced in tables as fev1slope and fefslope.

Statistical analysis methods

All SNPs were studied using family-based association tests (FBAT) and generalized estimating equations (GEE) [12] (see 100 K Overview). SNP results reported met the criteria of having a minor allele frequency ≥10%, a Hardy-Weinberg p-value \geq 0.001, and a call rate \geq 80%. All reported FBAT tests also required a minimum of ten informative families.

Multipoint variance component linkage analysis was implemented with a subset of 10,588 SNPs and all 612 available microsatellites studied in previous linkage analyses [12]. Multipoint identity-by-descent estimates were generated using the software Merlin [13]. Heritability estimates, estimating the proportion of the total phenotypic variance due to genetic effects, and variance component linkage analysis were performed using the software SOLAR [14].

In addition to evaluating all SNP associations with each phenotype individually, we developed a method to identify SNPs in or near genes that exhibited the strongest associations (as assessed by p-value) to multiple spirometry phenotypes. For each phenotype, we identified the 200 lowest p-values that met the criteria above and were localized within 60,000 base pairs of the transcription start or stop of a gene. All gene annotations are derived from the

UCSC genome browser May 2004 assembly, build 125 http://genome.ucsc.edu/[15,16]. We evaluated the frequency that a SNP appeared among the 200 lowest p-values in gene regions for the ten phenotypes. This strategy was based on the hypothesis that SNPs identified to be associated with multiple spirometry measures are more likely to reflect a true association with lung function than SNPs identified to be associated with only a single measurement. SNPs in gene regions that appeared among the lowest 200 p-values in five or more of the phenotypes studied are reported.

Candidate genes

Genes previously reported in the literature to be associated with spirometry measures or pulmonary disease were examined to determine whether any available 100 K SNPs in or near the genes were associated with spirometry phenotypes. Twelve COPD candidate genes studied in the Boston Early-Onset COPD cohort [17], and the SERPINE2 gene, a novel gene identified through linkage and association with COPD in the same cohort [6], were reviewed. The previously established COPD gene alpha-1-antitrypsin (SERPINA1) and the cystic fibrosis transmembrane conductance regulator (CFTR) as well as additional genes in the class of Glutathione S-transferases (O1, O2, M2, T1, T2) and surfactant proteins (SFTPA1, SFTPC) were reviewed. In addition, extracellular super oxide dismutase (SOD3) [18,19], interleukin-8 receptor alpha (IL8RA) [20], interleukin-10 (IL10) [21], beta-2 adrenergic receptor (ADRB2) [22], and transforming growth factor beta-1 (TGFB1) [23] were examined as COPD candidates. The GEE and FBAT SNP association results in or within 60 kilobase pairs (kb) of these 27 genes was reviewed. By specifying a 60 kb distance around the gene to screen results, we were able to identify SNPs near most candidate

gene regions, but often the SNP reported does not lie strictly within the transcription start or stop of the gene of interest.

Results

Table 1 reports the heritability estimates for each of the ten phenotypes presented. The slope phenotypes have lower heritability than their corresponding cross sectional phenotypes. FVC has the highest heritability among phenotypes defined using the same method (percent predicted or mean). The percent predicted FEF_{25–75}/FVC ratio had a higher heritability estimate than either FEV₁/FVC ratio phenotype.

Table 2 reports the SNPs with the lowest p-values for any of the 10 phenotypes evaluated in GWA analysis. Results were ranked by p-value and the top 25 SNPs are reported. In some cases, a SNP had top ranked results for multiple correlated phenotypes, and thus the p-value for all phenotypes is reported at the ranking position of the best pvalue. Table 2a was ranked by GEE p-value, and Table 2b was ranked by FBAT p-value. SNP positions are reported according to NCBI Human Genome Build 35. SNPs in the Affymetrix set whose chromosome and position are unknown and SNPs on sex chromosomes are not reported for association. In total, 70,987 SNPs were considered in association analyses. None of these results achieved a conservative threshold for genome-wide significance. All results, regardless of allele frequency, call rate, or deviation from Hardy-Weinberg are publicly available at http:/ /www.ncbi.nlm.nih.gov/projects/gap/cgi-bin/ study.cgi?id=phs000007.

Only a single SNP among those reported in Tables 2a and 2b is a known coding SNP. Among the top ranked GEE p-values was a non-synonymous coding SNP (rs156697) in the Glutathione S-transferase omega 2 gene (*GSTO2*) on chromosome 10. The SNP was among the top ranked GEE p-values for association with the mean FEV₁ and mean FVC phenotypes.

Other SNPs reported in Table 2a were localized to intronic gene regions in *COL1A2*, *ADARB1*, *SNTG1*, *RHBDD1*, *NID2*, *IL6R*, and *SYT10*. The best GEE p-value localized to the untranslated region of *SNRPN*. SNPs identified by FBAT p-value, reported in Table 2b, located in introns were in *CCBL2*, *LRRC9*, *PAX3*, *LIPF*, *MTHFD1L*, and *KIAA1797*. Additional genes reported in Tables 2a and 2b were within 60 kb of the associated SNP. SNP rs2906966, among the top 25 p-values for both tests, is located near *CDRT4*. SNPs reported may be in linkage disequilibrium (LD). Among the top six FBAT results, strong LD was observed between the three chromosome 1 SNPs in *CCBL2*, and the three chromosome 14 SNPs in *LRRC9*, thus only 2 regions are being identified.

Linkage to all autosomes and the X chromosome was performed. Table 2c reports all LOD scores above 2.0 with the 1.5-LOD support interval. The best LOD score observed is in a region of linkage on chromosome 6q that was reported previously using microsatellites in the Framingham families [7,8]. The original LOD score of 2.4 for mean FEV₁ using genome-wide microsatellites [7] was increased to a LOD of 2.89 with the addition of SNP data, and a LOD of 2.65 was observed in the same region for the percent predicted FEV₁ phenotype. The second highest LOD score observed was 2.86 for the longitudinal FEF₂₅₋₇₅ phenotype, which was located on the X chromosome. The percent predicted FEV₁ and FVC phenotypes both had LOD scores over 2.0 on chromosome 4, with overlapping LOD support intervals centered around 166–170 Mb.

Table 3 reports SNPs that met the quality control criteria and were among the top 200 p-values for SNPs located within 60 kb of a gene for at least five of the phenotypes studied. A SNP in an intron of the interleukin 6 receptor (IL6R) and a SNP near the sodium channel, voltage gated, type I alpha (SCN1A) gene were among the top ranked result for six phenotypes. All others were among the top ranked results for five phenotypes. The genes syntrophin, gamma 1 (SNTG1) and the chromosome 20 open reading frame 133 (C20orf133) appear in both the FBAT and GEE lists of top results across 5 phenotypes. The two SNPs in SNTG1 are separated by 104 kb and have low LD ($r^2 = 0.05$ and D' = 0.28 in the HapMap CEU sample). The two C20orf133 SNPs are in strong LD ($r^2 = 0.78$ in HapMap CEU), though rs10485771 is located 3' of the gene.

Table 4 reports the best p-value observed when examining SNP results specifically in the regions of the 27 candidate genes. SNPs reported are within 60 kb of the transcription start or stop for the gene listed, and sometimes lie within a different gene. Only 20 of the 27 genes of interest had SNPs in or within 60 kb of the gene. No SNPs were near enough to the genes GSTM1 and TNF, presented by Hersh et al. (2005), and GSTO1, GSTT1, SFTPA1, SFTPC and IL8RA to be considered. Of the 20 best FBAT results, 12 were p-values less than 0.05, but only 5 were p-values less than 0.01. Of the best GEE results, 14 were p-values less than 0.05, and 5 were p-values less than 0.01. Three gene regions had p-values less than 0.01 for both types of test, and these were CFTR, GSTO2, and SOD3. The vitamin D binding protein (GC) region produced an FBAT p-value of 0.0009, though GEE results were weaker.

Discussion

This is the first GWA of quantitative lung function measures to be reported, and it provides an opportunity for both hypothesis generation and hypothesis testing. We have identified a number of novel gene regions associated with pulmonary function. Associations with these SNPs

Table 2: GEE, FBAT, and linkage results

2a Top Association results based on GEE p-value

Phenotype	SNP	Chr	Physical position	GEE p-value	FBAT p-value	Gene Region
fev I slope	I. rs3867498	15	22629880	1.36 × 10 ⁻⁰⁶	0.07	SNRPN
meanfvc	2. rs441051	7	93698651	2.16 × 10 ⁻⁰⁶	0.005	COL1A2
meanratio	3. rs2838815	21	45454018	2.63×10^{-06}	0.0002	ADARB1
ppfvc	4. rs1455782	15	90851970	4.23×10^{-06}	0.18	FLJ3283 I
meanfvc	5. rs10516541	4	108472826	4.32×10^{-06}	7.19×10^{-05}	
meanfev I				1.7×10^{-05}	0.0002	
ppratio	6. rs310558	8	51575144	5.14×10^{-06}	0.14	SNTGI
ppfev l	7. rs3820928	2	227598971	5.33×10^{-06}	0.0005	RHBDD I
ppfef	8. rs730532	14	51588561	5.89×10^{-06}	0.15	NID2
ppfefrat	9. rs808225	14	57467669	7.38×10^{-06}	0.0008	
ppratio				1.45 × 10 ⁻⁰⁵	0.007	
ppfef	10. rs4129267	1	151239337	7.39×10^{-06}	0.07	IL6R
ppfev l	11. rs2906966*	17	15272242	8.31 × 10 ⁻⁰⁶	5.03×10^{-05}	CDRT4
ppfvc	12. rs357394	7	137506642	8.77 × 10 ⁻⁰⁶	0.14	
fefslope	13. rs1994169	12	33436783	9.55 × 10 ⁻⁰⁶	0.05	SYTIO
meanratio	14. rs2225434	21	45458574	9.68 × 10 ⁻⁰⁶	0.0004	ADARB1
meanfvc	15. rs156697 ^(a)	10	106029175	9.78 × 10 ⁻⁰⁶	9.42 × 10 ⁻⁰⁵	
meanfev I				1.8×10^{-05}	0.002	GSTO2
ppfefrat	16. rs564425	13	46797096	1.03×10^{-05}	0.02	
ppfef	17. rs10498441	14	51613974	1.06 × 10 ⁻⁰⁵	0.28	NID2
ppratio	18. rs880713	2	128847844	1.11 × 10 ⁻⁰⁵	0.26	AK128224
ppfefrat	19. rs811732	14	57456738	1.3×10^{-05}	0.20	
ppfvc	20. rs6558132	8	29526866	1.58 × 10 ⁻⁰⁵	0.74	
fev I slope	21. rs6972823	7	3009137	1.69 × 10 ⁻⁰⁵	0.10	
ppfef	22. rs9285611(b)	1	81895076	1.7 × 10 ⁻⁰⁵	0.07	
meanfev l	23. rs10504836	8	88669709	1.72 × 10 ⁻⁰⁵	0.28	
meanfev l	24. rs1491520	3	193243153	1.77 × 10 ⁻⁰⁵	0.12	
ppfvc	25. rs9300826	13	103001176	1.79 × 10 ⁻⁰⁵	0.11	

2b Top Association results based on FBAT p-value

Phenotype	SNP	Chr	Physical position	GEE p-value	FBAT p-value	Gene Region
ppratio	1. rs10922530	I	89139273	0.001	8.7 × 10 ⁻⁰⁷	
ppfefrat				0.002	3.78×10^{-06}	
ppfef				0.005	2.36×10^{-05}	CCBL2
ppfefrat	2. rs3753683	1	89139592	0.004	1.87 × 10 ⁻⁰⁶	
ppratio				0.004	3.59 × 10 ⁻⁰⁶	
ppfef				0.01	1.42 × 10 ⁻⁰⁵	CCBL2
ppratio	3. rs219349	14	59525168	0.01	6.15 × 10 ⁻⁰⁶	
ppfefrat				0.04	4.91 × 10 ⁻⁰⁵	LRRC9
ppratio	4. rs219391	14	59554684	0.02	6.18 × 10 ⁻⁰⁶	LRRC9
ppratio	5. rs219326	14	59512943	0.02	8.51 × 10 ⁻⁰⁶	

Table 2: GEE, FBAT, and linkage results (Continued)

ppfefrat				0.06	3.74 × 10 ⁻⁰⁵	LRRC9
ppfefrat	6. rs1409149	I	89141170	0.005	9.12 × 10 ⁻⁰⁶	
pratio				0.005	1.08 × 10 ⁻⁰⁵	
opfef				0.01	5.14 × 10 ⁻⁰⁵	CCBL2
pfvc	7. rs9299191	9	110944100	0.009	1.79 × 10 ⁻⁰⁵	
neanfvc	8. rs10515289(c)	5	99315845	0.17	2.21×10^{-05}	
pfvc	9. rs10498137	2	222966943	0.05	2.23×10^{-05}	PAX3
pfef	10. rs3858282	10	90424391	0.001	2.92×10^{-05}	LIPF
pratio	11. rs905367	4	59896363	0.02	2.95×10^{-05}	
neanfvc	12. rs10495872	2	37915266	0.05	3.02×10^{-05}	
ev I slope	13. rs1347222	12	80896622	0.01	3.27×10^{-05}	
pratio	14. rs2009488	4	59895895	0.03	3.43×10^{-05}	
neanfvc	15. rs6481257	10	58608558	0.02	3.52×10^{-05}	
pratio	16. rs461951	14	59608986	0.02	3.63 × 10 ⁻⁰⁵	LRRC9
neanfev l	17. rs491552	6	151403169	0.01	3.86 × 10 ⁻⁰⁵	MTHFDIL
neanfev l	18. rs1910137	4	27087329	0.08	4.04×10^{-05}	
pfvc	19. rs2831605 ^(d)	21	28467064	0.03	4.41 × 10 ⁻⁰⁵	
pfevl	20. rs2906966*	17	15272242	8.31×10^{-06}	5.03 × 10 ⁻⁰⁵	CDRT4
efslope	21. rs6740919	2	67531706	0.0009	5.17 × 10 ⁻⁰⁵	ETAA I 6
neanfev I	22. rs10498818	6	63134129	0.02	5.25 × 10 ⁻⁰⁵	
pfefrat	23. rs1393593	4	59907689	0.06	5.44 × 10 ⁻⁰⁵	
pfefrat	24. rs9312080	4	59891451	0.11	5.63 × 10 ⁻⁰⁵	
pfefrat	25. rs7851363	9	20748306	0.006	5.7 × 10 ⁻⁰⁵	KIAA I 797

2c Linkage peaks with LOD score > 2.0

Phenotype	SNP	Chr	Physical position	I.5-LOD interval start	1.5-LOD interval end	LOD score
meanfev l	rs2300081	6	168097526	165513097	170788550	2.89
fefslope	rs5909594	X(e)	118057198	113970434	124311695	2.86
ppfev l	AGC001b	6	170788550	164268152	170788550	2.65
ppratio	rs10497042	2	150675813	148296793	154203261	2.40
meanratio	rs10518669	1	82811739	74999965	88728520(b)	2.37
ppfef	rs2974490	5	113433775	95608129	124687730 ^(c)	2.29
meanfvc	rs10488908	21	21904704	19805361	34461762 ^(d)	2.29
ppfev l	rs10518032	4	170322764	161768472	181501690	2.28
ppfvc	rs10489542	1	225169260	213625196	233478438	2.17
ppfef	rs721411	17	36902470	28313357	52519635	2.17
ppfev l	rs753765	17	57323782	45345729	59677087	2.17
meanfev l	rs4918762	10	114389096	102080914	122214789 ^(a)	2.12
pfvc	rs10517825	4	166251091	156507719	191091333	2.02

^{*)} SNP occurring in both Table 2a and 2b

a) SNP associated to meanfvc and meanfev1 in Table 2a under linkage peak for meanfev1 on chromosome 10.

b) SNP associated to ppfef in Table 2a under linkage peak for meanratio on chromosome 1.

c) SNP associated to meanfvc in Table 2b under linkage peak for ppfef on chromosome 5. d) SNP associated to ppfvc in Table 2b under linkage peak for meanfvc on chromosome 21.

e) X chromosome linkage results are not available online

SNP	Chr	bp position	Gene region
3a GEE			
rs4129267	I	151239337	IL6R
rs7587026	2	166804257	SCNIA
rs3820928	2	227598971	RHBDD I
rs445347	5	53011487	NDUFS4
rs310558	8	51575144	SNTGI
rs581446	18	10646848	FAM38B
rs10485770	20	13943214	C20orf133
3b FBAT			
rs10489030	4	24521105	3' of SOD3
rs2438345	5	90198041	GPR98
rs7759033	6	116404207	FRK
rs2391996	7	31487207	C7orf16
rs10504106	8	51471079	SNTG I
rs10485771	20	13999103	C20orf133

and gene regions require replication in other study samples as well as functional studies before any statement about causality is warranted. Many of the best p-values are likely to reflect false positive results, and GEE results exhibited elevated Type I error [12] (see 100 K Overview). Additional studies of this data will be useful, including smoking stratified analyses and more sophisticated approaches to creating multivariate phenotypes. However, several of the observed associations involve genes for which there are plausible biologic rationales for a relation to lung function phenotypes.

The Glutathione S-Transferase (GST) superfamily genes are of interest because of their role in metabolism of xenobiotics, such as cigarette smoke. Recently, Hersh et al. studied *GSTP1* and *GSTM1* in two independent analyses of COPD and reported null findings. In contrast, a study of annual change in lung function measures in a population based cohort reported that the *GSTT1* deletion alone or in combination with the *GSTM1* deletion influenced decline in FEV₁ in men [24]. Using the Affymetrix 100 K SNP GeneChip, we have limited ability to directly confirm or refute the aforementioned findings because no SNPs were genotyped within 10 kb of the genes.

Here, we show that a non-synonymous SNP in exon 5 of GSTO2 encoding an Asn142Asp amino acid change is among the most striking GWA results for mean FEV_1 and FVC phenotypes. Both the non-synonymous SNP, rs156697, and a second SNP, rs156699, located in an intron exhibited strong association using both FBAT and GEE tests (r^2 between SNPs = 0.8 in HapMap CEU). Linkage results also support the evidence for GSTO2, as the gene's position lies within the confidence interval around the LOD of 2.12 observed on chromosome 10 for mean FEV_1 (Table 2c). GSTO2 is involved in the biotransforma-

tion of arsenic, which is a component of cigarette smoke, and may exhibit modest expression in bronchial epithelial cells. Gene expression studies in COS-1 cells demonstrated that the Asp142 variant exhibited 76% of the level of expressed protein occurring in the wild-type, and expression levels were further reduced to 15% when the Asp142 occurred in conjunction with an Ile158 variant [25]. The observation of strong association to a non-synonymous polymorphism with demonstrated effects on gene expression is compelling. Moreover, this finding in conjunction with a growing literature on GST gene association with pulmonary phenotypes suggests that a complete evaluation of functional variants in this gene family may be warranted.

The IL6R SNP was not only among the top 25 p-values for percent predicted ${\rm FEF}_{25-75}$, but also among the top 200 p-values in gene regions for six of the ten phenotypes presented. IL6R is thought to be expressed in lung, and may play a role in the immune response. Recently, we have shown that IL6 levels in blood were associated with impaired lung function in the Framingham offspring cohort [26]. The IL6 pathway, as a mediator of the inflammatory process, is of interest as it relates to lung function phenotypes.

The SNP identified in the SOD3 region lies within a hypothetical protein 3' of the SOD3 gene. The non-synonymous SNP in SOD3 that has been reported for association with COPD (rs1799895) was not included in the Hap-Map, so we could not determine the extent of LD between it and the SNPs genotyped in this study. Another exon 3 SNP (rs2536512) located 519 base pairs away from rs1799895 is present in the HapMap. The associated SNP identified in this study, rs10489030, is in very low LD with the SOD3 exon 3 HapMap SNP (D' = 0.32 and r^2 =

Table 4: Candidate gene evaluation

Gene region	Chr	SNP	Phenotype	FBAT p-value	GEE p-value
Alpha-I-antitrypsin (AAT/SERPINAI)	14	rs2402446	ppfvc fev I slope	0.02	0.30
		rs I 0484042	•	0.90	0.01
Beta-2 adrenergic receptor (ADRB2)	5	rs30329	fefslope meanratio	0.009	0.07
		rs9325117		-	0.04
cystic fibrosis transmembrane regulator (CFTR)	7	rs213987	meanfvc ppratio	0.006	0.38
		rs I 0487367	• •	0.25	0.002
Microsomal epoxide hydrolase (EPHX1)	I	rs3738051	fefslope ppfefrat	0.01 0.03	0.30 0.04
Vitamin D binding protein (GC)	4	rs423817	ppfefrat ppfefrat	0.0009	0.06
		rs842873	FF	0.05	0.02
Glutathione S-transferase M2 (GSTM2)	I	rs542338	meanratio fev I slope	0.19 0.43	0.63 0.20
Glutathione S-transferase O2 (GSTO2)	10	rs156697	meanfvc	9.4 × 10 ⁻⁰⁵	9.8 × 10 ⁻⁰⁶
Glutathione S-transferase PI (GSTPI)	II	rs688878	ppfefrat ppfvc	0.07 0.39	0.35 0.02
Glutathione S-transferase T2 (GSTT2)	22	rs I 40289	ppfvc ppfev1	0.17 0.41	0.13 0.10
Heme oxygenase (HMOX1)	22	rs2267331	fev I slope meanratio	0.07	0.80
		rs10483190		0.82	0.008
nterleukin-10 (<i>IL10</i>)	I	rs I 0494879	fefslope fev I slope	0.06 0.46	0.73 0.02
Matrix metalloproteinase-I (MMPI)	11	rs495366	fefslope ppfef	0.02 0.20	0.13 0.03
Matrix metalloproteinase-9 (MMP9)	20	rs2903908	ppfefrat	0.19	0.05
Alpha-1-antichymotrypsin (SERPINÁ3)	14	rs I 0484047	ppfvc fev I slope	0.02 0.10	0.03 0.01
Serine proteinase inhibitor E2 (SERPINE2)	2	rs717610	meanfvc meanratio	0.0 4 0.19	0.003 0.001
Surfactant protein B (SFTPB)	2	rs7577293	meanratio	0.02	0.06
Surfactant protein D (SFTPD)	10	rs726289	fefslope	0.02	0.03
extracellular super oxide dismutase (SOD3)	4	rs10489030	ppfev l	0.0005	0.02
1			ppfvc	0.01	0.007
ransforming growth factor beta-1 (TGFB1)	19	rs3745295	fev I slope	0.21	0.62
, ,			ppfevl	0.74	0.18
Tissue inhibitor of metalloproteases-2 (TIMP2)	17	rs2889529	ppfef meanratio	0.07 0.18	0.11 0.09

 $Best \ p-value \ for \ FBAT \ and \ GEE \ tests \ reported \ with \ specific \ SNP* phenotype \ producing \ result. \ P-values < 0.01 \ bolded.$

0.005 in HapMap CEU). The low LD between the *SOD3* exonic SNP and this 3' SNP, separated by 43.5 kb, does not suggest a clear replication of association with the *SOD3* gene, but suggests that the genomic region is of continued interest. Also on chromosome 4, a SNP in the region of the vitamin D binding protein (GC) was associated with the percent predicted FEF_{25–75}/FVC ratio with a p-value of 0.0009 using the FBAT test, but this SNP is in low LD with the Asp432Gly polymorphism, rs7041 (D' = 0.17 and r² = 0.007 in HapMap CEU).

The SNP identified in the region of *SERPINE2* (rs717610) was not in LD with the six reported SNPs replicating significant associations to COPD [6] that were also available

in HapMap, as the r² values ranged from 0.002 to 0.008. Two of the top SNPs (rs3820928, GEE rank #7; rs10498137, FBAT rank #9) lie within the linkage region identified for FEV₁/FVC in severe early-onset COPD cases [27] that subsequently led to the discovery of the SERPINE2 associated SNP. SNP rs3820928 was among those identified with association to five of the phenotypes studied and lies in the gene RHBDD1 (alias DKFZp547E052). Not much is known about the function of this gene, but the associated SNP is located in a region with LD extending to the adjacent COL4A4 gene. The rs3820928 exhibited an r² of 0.81 with two non-synonymous coding SNPs in COL4A4 in the HapMap CEU data. Defects in Type IV collagen genes have been shown to

influence Goodpasture's syndrome, an autoimmune disease affecting the lung [28], and both *COL4A4* and *COL4A3* lie in this region, sharing a common promoter. These results do not provide a strong replication of the original *SERPINE2* SNP associations due to the low LD between SNPs reported in the literature and the SNPs from the 100 K data with association. However, the results suggest that chromosome 2q is of continued interest and may harbor multiple genes influencing lung function.

Studying lung function measures in a community-based sample may identify genetic variants associated with lung growth and development, susceptibility to obstructive ventilatory impairment related to asthma, emphysema and COPD or susceptibility to restrictive ventilatory impairment due to pulmonary fibrosis or other processes. The relevance to disease pathogenesis of associations between SNPs and lung function must be interpreted with caution, and some of the observed associations may reflect polymorphisms that protect against ventilatory impairment by leading to better lung function in early life or protection against the adverse effects of cigarette smoking. All of the GWA results are publicly available at http:/ /www.ncbi.nlm.nih.gov/projects/gap/cgi-bin/ study.cgi?id=phs000007. Replication of novel results identified by GWA will be the true test of the value of the GWA approach to gene discovery.

Conclusion

These publicly available results provide a resource for investigators to assess whether their findings of association to pulmonary function phenotypes replicate in the Framingham population. In addition, we have identified novel results that warrant replication studies in other populations.

Abbreviations

BMI = body mass index; COPD = chronic obstructive pulmonary disease; FEV_1 = forced expiratory volume in one second; FBAT = family based association test; FVC = forced vital capacity; FEF_{25-75} = forced expiratory flow between the 25th and 75th percentile; GEE = generalized estimating equation; GWA = genome-wide association; kb = kilobase pairs (1000 base pairs); kb = kilobase pairs (1000 base pairs); kb = kilobase pairs (1000 base pairs); kb = kilobase pairs).

Competing interests

The authors declare that they have no competing interests.

Authors' contributions

JBW created the phenotypic residuals that were analyzed, reviewed GWA results, and drafted the manuscript text and tables. REW, DJG, and GTO assessed the quality of spirometry data, identified exclusion criteria, and defined the covariates and methods to develop phenotypes for

analysis. JML wrote scripts to parse through the GWA results and facilitate the presentation of results. All authors read and approved the final manuscript.

Acknowledgements

The authors would like to recognize the Framingham Heart Study participants and the support from NIH/NHLBI Contract NO1-HC25195. Drs. Wilk and Walter are each supported by their own Young Clinical Scientist Award from the Flight Attendant Medical Research Institute (FAMRI). The authors would like to thank Emelia Benjamin, MD, ScM for serving as guest editor and providing thoughtful comments on the manuscript. A portion of this research was conducted using the Boston University Linux Cluster for Genetic Analysis (LinGA) funded by the NIH NCRR (National Center for Research Resources) Shared Instrumentation grant (ISIORR163736-01AI).

This article has been published as part of *BMC Medical Genetics* Volume 8 Supplement 1, 2007: The Framingham Heart Study 100,000 single nucleotide polymorphisms resource. The full contents of the supplement are available online at http://www.biomedcentral.com/1471-2350/8?issue=SI.

References

- Mannino DM, Homa DM, Akinbami LJ, Ford ES, Redd SC: Chronic obstructive pulmonary disease surveillance – United States, 1971–2000. Respiratory care 2002, 47(10):1184-1199.
- Hoyert DL, Heron MP, Murphy SL, Kung HC: Deaths: final data for 2003. Natl Vital Stat Rep 2006, 54(13):1-120.
- Fabbri L, Pauwels RA, Hurd SS: Global Strategy for the Diagnosis, Management, and Prevention of Chronic Obstructive Pulmonary Disease: GOLD Executive Summary updated 2003. Copd 2004, 1(1):105-141. discussion 103-104.
- Silverman EK, Chapman HA, Drazen JM, Weiss ST, Rosner B, Campbell EJ, O'Donnell WJ, Reilly JJ, Ginns L, Mentzer S, Wain J, Speizer FE:
 Genetic epidemiology of severe, early-onset chronic obstructive pulmonary disease. Risk to relatives for airflow obstruction and chronic bronchitis. American journal of respiratory and critical care medicine 1998, 157(6 Pt 1):1770-1778.
- Wilk JB, Djousse L, Arnett DK, Rich SS, Province MA, Hunt SC, Crapo RO, Higgins M, Myers RH: Evidence for major genes influencing pulmonary function in the NHLBI family heart study. Genet Epidemiol 2000, 19(1):81-94.
- Demeo DL, Mariani TJ, Lange C, Srisuma S, Litonjua AA, Celedon JC, Lake SL, Reilly JJ, Chapman HA, Mecham BH, Haley KJ, Sylvia JS, Sparrow D, Spira AE, Beane J, Pinto-Plata V, Speizer FE, Shapiro SD, Weiss ST, Silverman EK: The SERPINE2 Gene Is Associated with Chronic Obstructive Pulmonary Disease. American journal of human genetics 2006, 78(2):253-264.
- Joost O, Wilk JB, Cupples LA, Harmon M, Shearman AM, Baldwin CT, O'Connor GT, Myers RH, Gottlieb DJ: Genetic loci influencing lung function: a genome-wide scan in the Framingham Study. American journal of respiratory and critical care medicine 2002, 165(6):795-799.
- Wilk JB, DeStefano AL, Joost O, Myers RH, Cupples LA, Slater K, Atwood LD, Heard-Costa NL, Herbert A, O'Connor GT, Gottlieb DJ: Linkage and association with pulmonary function measures on chromosome 6q27 in the Framingham Heart Study. Human molecular genetics 2003, 12(21):2745-2751.
- Wilk JB, Herbert A, Shoemaker CM, Gottlieb DJ, Karamohamed S: Secreted modular calcium-binding protein 2 haplotypes are associated with pulmonary function. American journal of respiratory and critical care medicine 2007, 175(6):554-560.
- ATS: Standardization of Spirometry, 1994 Update. American Thoracic Society. American journal of respiratory and critical care medicine 1995, 152(3):1107-1136.
- Hankinson JL, Odencrantz JR, Fedan KB: Spirometric reference values from a sample of the general U.S. population. American journal of respiratory and critical care medicine 1999, 159(1):179-187.
- Cupples LA, Arruda HT, Benjamin EJ, D'Agostino RB Sr, Demissie S, DeStefano AL, Dupuis J, Falls K, Fox CS, Gottlieb DJ, Govindaraju DR,

Guo CY, Heard-Costa NL, Hwang SJ, Kathiresan S, Kiel DP, Laramie JM, Larson MG, Levy D, Liu CY, Lunetta KL, Mailman MD, Manning AK, Meigs JB, Murabito JM, Newton-Cheh C, O'Connor GT, O'Donnell CJ, Pandey MA, Seshadri S, Vasan RS, Wang ZY, Wilk JB, Wolf PA, Yang Q, Atwood LD: The Framingham Heart Study 100 K SNP genome-wide association study resource: Overview of 17 phenotype working group reports. BMC Med Genet 2007, 8(Suppl 1):S1.

- Abecasis GR, Cherny SS, Cookson WO, Cardon LR: Merlin rapid analysis of dense genetic maps using sparse gene flow trees. Nature genetics 2002, 30(1):97-101.
- Almasy L, Blangero J: Multipoint quantitative-trait linkage analysis in general pedigrees. American journal of human genetics 1998, 62(5):1198-1211.
- Karolchik D, Baertsch R, Diekhans M, Furey TS, Hinrichs A, Lu YT, Roskin KM, Schwartz M, Sugnet CW, Thomas DJ, Weber RJ, Haussler D, Kent WJ: The UCSC Genome Browser Database. Nucleic acids research 2003, 31(1):51-54.
- Kent WJ, Sugnet CW, Furey TS, Roskin KM, Pringle TH, Zahler AM, Haussler D: The human genome browser at UCSC. Genome research 2002, 12(6):996-1006.
- Hersh CP, Demeo DL, Lange C, Litonjua AA, Reilly JJ, Kwiatkowski D, Laird N, Sylvia JS, Sparrow D, Speizer FE, Weiss ST, Silverman EK: Attempted replication of reported chronic obstructive pulmonary disease candidate gene associations. American journal of respiratory cell and molecular biology 2005, 33(1):71-78.
- Young RP, Hopkins R, Black PN, Eddy C, Wu L, Gamble GD, Mills GD, Garrett JE, Eaton TE, Rees MI: Functional variants of antioxidant genes in smokers with COPD and in those with normal lung function. Thorax 2006, 61(5):394-399.
- Juul K, Tybjaerg-Hansen A, Marklund S, Lange P, Nordestgaard BG: Genetically increased antioxidative protection and decreased chronic obstructive pulmonary disease. American journal of respiratory and critical care medicine 2006, 173(8):858-864.
- Stemmler S, Arinir U, Klein W, Rohde G, Hoffjan S, Wirkus N, Reinitz-Rademacher K, Bufe A, Schultze-Werninghaus G, Epplen JT: Association of interleukin-8 receptor alpha polymorphisms with chronic obstructive pulmonary disease and asthma. Genes and immunity 2005, 6(3):225-230.
- Burgess JL, Fierro MA, Lantz RC, Hysong TA, Fleming JE, Gerkin R, Hnizdo E, Conley SM, Klimecki W: Longitudinal decline in lung function: evaluation of interleukin-10 genetic polymorphisms in firefighters. Journal of occupational and environmental medicine/American College of Occupational and Environmental Medicine 2004, 46(10):1013-1022.
- Hegab AÈ, Sakamoto T, Saitoh W, Massoud HH, Massoud HM, Hassanein KM, Sekizawa K: Polymorphisms of IL4, IL13, and ADRB2 genes in COPD. Chest 2004, 126(6):1832-1839.
- Celedon JC, Lange C, Raby BA, Litonjua AA, Palmer LJ, DeMeo DL, Reilly JJ, Kwiatkowski DJ, Chapman HA, Laird N, Sylvia JS, Hernandez M, Speizer FE, Weiss ST, Silverman EK: The transforming growth factor-betal (TGFBI) gene is associated with chronic obstructive pulmonary disease (COPD). Human molecular genetics 2004, 13(15):1649-1656.
- Imboden M, Downs SH, Senn O, Matyas G, Brandli O, Russi EW, Schindler C, Ackermann-Liebrich U, Berger W, Probst-Hensch NM: Glutathione S-transferase genotypes modify lung function decline in the general population: SAPALDIA cohort study. Respiratory research 2007, 8:2.
- Mukherjee B, Salavaggione OE, Pelleymounter LL, Moon I, Eckloff BW, Schaid DJ, Wieben ED, Weinshilboum RM: Glutathione Stransferase omega I and omega 2 pharmacogenomics. Drug metabolism and disposition: the biological fate of chemicals 2006, 34(7):1237-1246.
- Walter RE, Wilk JB, Larson MG, Vasan RS, Keaney JF, Lipinska I, O'Connor GT, Benjamin EJ: Systemic inflammation and COPD: The Framingham Heart Study. Chest in press.
- 27. Silverman EK, Palmer LJ, Mosley JD, Barth M, Senter JM, Brown A, Drazen JM, Kwiatkowski DJ, Chapman HA, Campbell EJ, Province MA, Rao DC, Reilly JJ, Ginns LC, Speizer FE, Weiss ST: Genomewide linkage analysis of quantitative spirometric phenotypes in severe early-onset chronic obstructive pulmonary disease. American journal of human genetics 2002, 70(5):1229-1239.
- Hudson BG, Tryggvason K, Sundaramoorthy M, Neilson EG: Alport's syndrome, Goodpasture's syndrome, and type IV

collagen. The New England journal of medicine 2003, **348(25)**:2543-2556.

Publish with **Bio Med Central** and every scientist can read your work free of charge

"BioMed Central will be the most significant development for disseminating the results of biomedical research in our lifetime."

Sir Paul Nurse, Cancer Research UK

Your research papers will be:

- available free of charge to the entire biomedical community
- peer reviewed and published immediately upon acceptance
- cited in PubMed and archived on PubMed Central
- yours you keep the copyright

Submit your manuscript here: http://www.biomedcentral.com/info/publishing_adv.asp

