

Research article

Effect of Interleukin-1 β on proliferation of gastric epithelial cells in culture

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Abstract

Background: *Helicobacter pylori* is the main risk factor for the development of non-cardia gastric cancer. Increased proliferation of the gastric mucosa is a feature of *H. pylori* infection. Mucosal interleukin-1 β production is increased in *H. pylori* infection and IL-1 β genotypes associated with increased pro-inflammatory activity are risk factors for the development of gastric cancer. The effect of IL-1 β on gastric epithelial cell proliferation has been examined in this study.

Methods: AGS cells were cultured with IL-1 β . DNA synthesis was assayed by [3 H]thymidine incorporation and total viable cell numbers by MTT assay.

Results: IL-1 β dose dependently increased DNA synthesis and cell numbers. The enhanced proliferation was blocked by interleukin-1 receptor antagonist. Addition of neutralising antibody to GM-CSF reduced IL-1 β -stimulated proliferation by 31 ± 4 %. GM-CSF alone significantly stimulated proliferation. Addition or neutralisation of IL-8 had no effect on basal or IL-1 β -stimulated proliferation. The tyrosine kinase inhibitor genistein completely blocked IL-1 β -stimulated proliferation and inhibition of the extracellular signal related kinase pathway with PD 98059 inhibited IL-1 β stimulated proliferation by 58 ± 5 %.

Conclusions: IL-1 β stimulates proliferation in gastric epithelial cells. Autocrine stimulation by GM-CSF contributes to this proliferative response. Signalling via tyrosine kinase activity is essential to the mitogenic response to IL-1 β . The extracellular signal related kinase pathway is involved in, but not essential to downstream signalling. IL-1 β may contribute to the hyperproliferation seen in *H. pylori*-infected gastric mucosa, and be involved in the carcinogenic process.

Background

Helicobacter pylori is believed to be the major aetiological factor in the development of non-cardia gastric adenocarcinoma. Large-scale epidemiological studies have confirmed a strong association between *H. pylori* infection and both cancer [1–3] and the earlier histological stages, atrophy and intestinal metaplasia [4,5]; both of which increase the risk of later neoplastic transformation. Animal models have also demonstrated the importance of *H. py-*

lori in gastric carcinogenesis [6,7]. Increased rates of proliferation of the gastric mucosa are typical in *H. pylori* infection [8–11], and hyperproliferation within the gastrointestinal tract appears to be a marker for later malignant change [12]. The cause of the increased rate of proliferation is not clear, but the increased rates reduce to normal with clearance of the infection [8,13]. Although hyperproliferation is typical *in vivo*, studies testing the effects of *H. pylori* or its products *in vitro* have shown con-

flicting results, with both enhanced [14,15] and diminished [16–18] proliferation reported. It is possible that other components of the inflammatory response typical of *H. pylori* infected mucosa could be at least partly responsible for driving the increased cell proliferation.

The pluripotent pro-inflammatory cytokine interleukin-1 β has a central role in the pathogenesis of *H. pylori*-induced mucosal inflammation. IL-1 β gene expression and protein production are increased in *H. pylori* infection and reduce with successful eradication [19,20]. The presence of the IL-1 β genotype polymorphism associated with enhanced IL-1 β -production has been associated with a significant increased risk of gastric cancer and pre-cancerous lesions [21,22]. Interleukin-1 β is a potent inhibitor of gastric acid secretion and it is hypothesized that the enhanced IL-1 β response alters the topography of the gastric infection and thus promotes inflammation and subsequent atrophy of the gastric corpus [23,24]. The possibility that IL-1 β itself drives the increased proliferation of gastric epithelial cells has not been fully investigated. Alteration of gastric proliferation by IL-1 β might contribute to the carcinogenic process, in addition to effects on acid secretion. Therefore the direct effects of IL-1 β on gastric epithelial proliferation have been assessed.

The mitogen-activated protein kinase (MAPK) cascades are well-characterised pathways transducing signals from the cell surface to the nucleus. The family includes distinct subgroups; extracellular signal-related kinases (ERKs), c-Jun NH₂-terminal kinases (JNKs) and p38 MAPK [25]. The ERKs are activated by a variety of extracellular stimuli, and mediate the pro-proliferative effects of a number of hormones and growth factors [26,27]. Activation by phosphorylation of a dual specificity protein kinase (MAP kinase kinase (MAPKK)), (also known as MEK), allows it in turn to activate a family of serine-threonine protein kinases, known as the ERKs. The ERKs in turn phosphorylate numerous cellular proteins including transcription factors and thus have a central role in propagation of mitogenic signals. Accordingly the role of the MAP-kinase pathway in mediating the responses to IL-1 β has been assessed.

Methods

Cell culture

The human AGS gastric carcinoma cell line was purchased from the European Collection of Animal Cell Cultures (Porton Down, UK). Cells were grown in monolayer culture in RPMI 1640 medium supplemented with 100 μ g/ml penicillin, 100 μ g/ml streptomycin, 100 μ g/ml gentamicin, 2.5 μ g/ml amphotericin B and 10 % foetal calf serum. Cells were grown in 75 cm² tissue culture flasks at 37°C in an atmosphere of 5% CO₂ and 95% air and passaged every 5–7 days.

Proliferation studies

[³H]thymidine incorporation. Cells were grown in media containing 10% foetal calf serum, plated into 24-well plates at 10⁵ cells/well and allowed to attach overnight. After washing with serum-free media, cells were incubated in serum free medium containing 0.2 mM unlabelled thymidine for 24 hours in the presence of increasing concentrations of IL-1 β , IL-8 or GM-CSF. DNA synthesis was estimated by measurement of [³H]thymidine incorporation into the trichloroacetic acid (TCA) precipitable material [28]. [³H]thymidine (0.1 μ Ci/ml, 10 Ci/mmol) was added 2 hours before the end of a 24 hour treatment period. Cells were washed twice with serum-free medium to remove unincorporated [³H]thymidine, and DNA was precipitated with 5% TCA at 4°C for 15 minutes. The precipitates were then washed twice with 95% ethanol, dissolved in 1 ml of NaOH, and analysed by liquid scintillation counting. Results are expressed as percent control unstimulated [³H]thymidine incorporation (mean \pm SD) of 4–6 different experiments, each performed in triplicate. For the detection of growth inhibition, cells were incubated with either the specific MEK inhibitor PD 98059 (25 μ M) [29], IL-1 receptor antagonist (500 ng/ml) [30] or the neutralising antibodies, anti-GM-CSF (5 μ g/ml) or anti-IL-8 (10 μ g/ml). Inhibitors or antibodies were added 30 minutes prior to cytokines.

Cell growth

Total viable cell numbers were assessed by a modified MTT (3-[4,5-dimethylthiazol-2-yl]-2,5 diphenyl tetrazolium bromide assay) [31]. Cells were plated into 24-well plates in medium containing 10% foetal calf serum. After attachment overnight, the medium was changed to 1% foetal calf serum-supplemented medium and increasing concentrations of IL-1 β were added. Cells were cultured for 48 hours and then the medium was removed and fresh RPMI 1640 medium containing 0.5 ng/ml MTT was added. Cells were incubated at 37°C for 3 hours. The medium was then removed and 0.04 M HCl in isopropanol was added to extract the reduced formazan product. The resulting optical density at 550 nm was determined.

Chemicals and reagents

Recombinant human IL-1 β and IL-8 were purchased from Sigma (Poole, UK), recombinant human GM-CSF and IL-1 receptor antagonist, anti-GM-CSF and anti-IL-8 were from R and D systems (Abingdon, UK). PD 95059 was from Calbiochem (Nottingham, UK). RPMI 1640 was from Gibco BRL (Paisley, UK) and all other reagents were from Sigma. Concentrations of inhibitors used were taken from manufacturers' data and published data. The ability of 500 ng/ml IL-1RA to abolish 10 ng/ml IL-1 β -stimulation of IL-8 secretion in AGS cells was confirmed. The efficacy of the anti-GM-CSF at 5 μ g/ml antibody to abolish

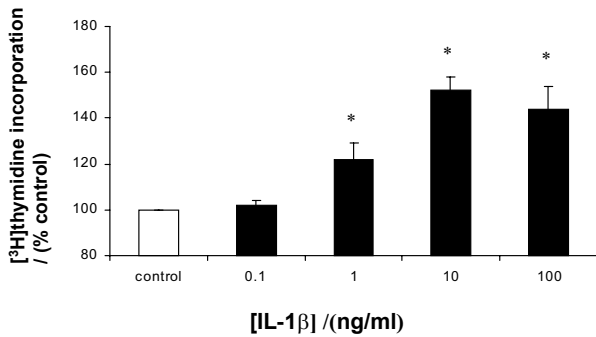


Figure 1

Effect of IL-1 β on [³H]thymidine incorporation into gastric epithelial AGS cells. Cells were treated with increasing concentrations of IL-1 β for 24 hours and DNA synthesis assessed by [³H]thymidine incorporation. Results expressed as means \pm standard deviation. * $P < 0.01$ vs. control

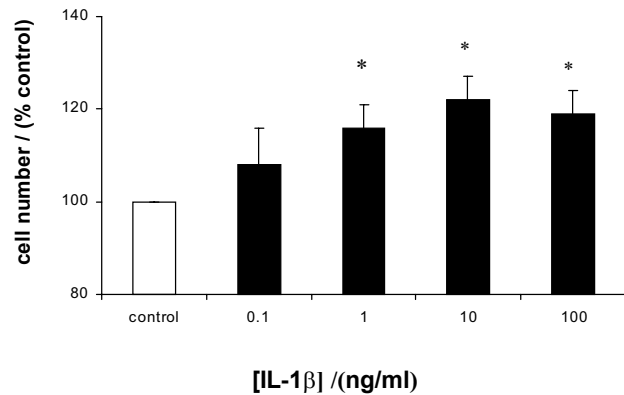


Figure 2

Effect of IL-1 β on cell numbers of gastric epithelial cells. Cells were treated with increasing concentrations of IL-1 β for 48 hours and total viable cell numbers assessed by MTT assay. Results expressed as means \pm standard deviation. * $P < 0.01$ vs. control

GM-CSF (1 ng/ml) induced AGS cell proliferation was confirmed.

Statistics

Cytokine-stimulated results were compared with control unstimulated cells on the same 24-well plate. Data were compared by one-way analysis of variance and Student's t-test to determine statistical significance. Each experiment as performed in triplicate on 4–6 occasions. Results are expressed as mean \pm standard deviation. Differences with P values of < 0.05 were considered significant.

Results

Effect of IL-1 β on [³H]thymidine incorporation

Interleukin-1 β caused a dose-dependent increase in DNA synthesis as measured by thymidine incorporation. As shown in figure 1, significant stimulation was seen with 1–100 ng/ml IL-1 β . The maximal stimulation of $52 \pm 6\%$ above control was seen with 10 ng/ml. The higher dose of 100 mg/ml was slightly less effective in stimulating proliferation.

Effect of IL-1 β on cell number

The increase in DNA synthesis by IL-1 β was translated into an absolute increase in viable cell numbers. As shown in figure 2, IL-1 β increased cell numbers in a dose-dependent manner similar to the effects on [³H]thymidine incorporation. The maximal stimulation was again seen at 10 ng/ml of IL-1 β , which produced a $22 \pm 5\%$ increase in total cell number.

Effects of cytokine inhibition or receptor antagonism on IL-1 β -stimulation of proliferation

Pretreatment of the cells with interleukin-1 receptor antagonist abolished the stimulatory effects of IL-1 β on

[³H]thymidine incorporation (Figure 3). Previous studies have shown that IL-1 β can activate gastric epithelial cells, including AGS cells, to secrete other cytokines, particularly interleukin-8 and granulocyte-macrophage colony stimulating factor (GM-CSF)[31,32]. Therefore further studies were undertaken to assess if the stimulatory actions of IL-1 β were mediated by either of these two cytokines. Neutralising antibodies to either IL-8 or GM-CSF had no effect on unstimulated [³H]thymidine incorporation. Neutralisation of IL-8 had no effect on IL-1 β -stimulated growth but the anti-GM-CSF antibody reduced IL-1 β -stimulated proliferation by $31 \pm 4\%$ ($P < 0.01$) (figure 3).

Effects of GM-CSF on proliferation

In view of the results obtained with the anti-GM-CSF antibody, the direct growth-stimulatory actions of GM-CSF were examined. GM-CSF had a potent growth stimulatory action on AGS cells: significant enhancement of [³H]thymidine incorporation was seen at all concentrations (0.001–100 ng/ml) of GM-CSF. GM-CSF itself appeared to be a more potent stimulant than IL-1 β ; maximal stimulation of $108 \pm 17\%$ above control was seen with 100 ng/ml of GM-CSF (Figure 4). The inhibitory action of the anti-GM-CSF antibody was confirmed by abolition of the growth stimulatory action of 1 ng/ml GM-CSF (data not shown). Previous studies have shown that IL-1 β -stimulated GM-CSF release under similar conditions in AGS cells to be approximately 10–20 pg/well/24 hours [32]. To confirm the results obtained with the anti-IL-8 antibody, [³H]thymidine incorporation was measured in response to IL-8. No enhancement of proliferation was seen at any concentration of IL-8 (0.001–100 ng/ml) (data not

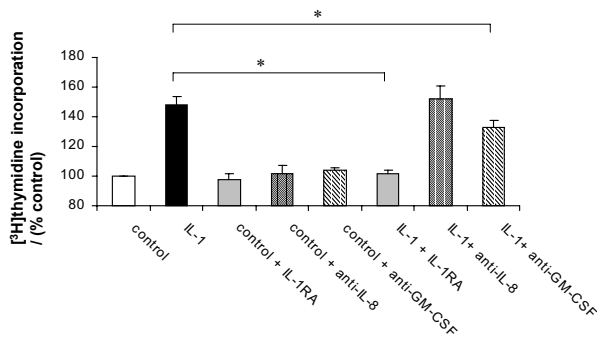


Figure 3
Effect of cytokine inhibition on IL-1 β -stimulation of [³H]thymidine into gastric epithelial cells AGS cells were treated with 10 ng/ml IL-1 β for 24 hours plus either interleukin-1 receptor antagonist (IL-1RA 500 ng/ml), neutralising anti-IL-8 antibody (10 μ g/ml) or neutralising anti-GM-CSF antibody (5 μ g/ml). DNA synthesis was assessed by [³H]thymidine incorporation. Results expressed as means \pm standard deviation. * $P < 0.01$ vs. IL-1 β stimulation in absence of inhibitor.

shown). Under similar conditions IL-1 β -stimulated IL-8 release is approximately 3000 pg/well/24 hours [31].

Mechanism of IL-1 β -stimulation of cell proliferation

The specific inhibitors genistein, which inhibits tyrosine kinases and PD 98059, which inhibits MAP kinase kinase (MEK), and thus inhibits the ERK-pathway, were used to assess the possible intracellular pathways mediating the effects of IL-1 β . In order to examine the effects of IL-1 β distinct from those of GM-CSF, these experiments were performed in the presence of the anti-GM-CSF neutralising antibody. As shown in figure 5, neither genistein nor PD 98059 altered unstimulated [³H]thymidine incorporation. Genistein completely abolished the IL-1 β -stimulation of proliferation. Inhibition of MEK with PD 98059 reduced IL-1 β -stimulated proliferation by 58 \pm 5% ($P < 0.01$) (figure 5) but did not completely abolish the growth stimulatory action of IL-1 β . Further increases to supra-maximal concentrations of PD 98059 did not further inhibit IL-1 β -stimulated proliferation (data not shown).

Discussion

This study demonstrated that IL-1 β increased proliferation of AGS cells. This effect was reversed by the receptor antagonist, suggesting it was mediated via the interleukin-1 receptor. IL-1 β stimulated both [³H]thymidine incorporation, as a measure of stimulation of DNA synthetic rate and also total cell numbers as measured by the MTT assay. This illustrates that the stimulation of DNA synthesis by IL-1 β is translated into a real increase in cell numbers.

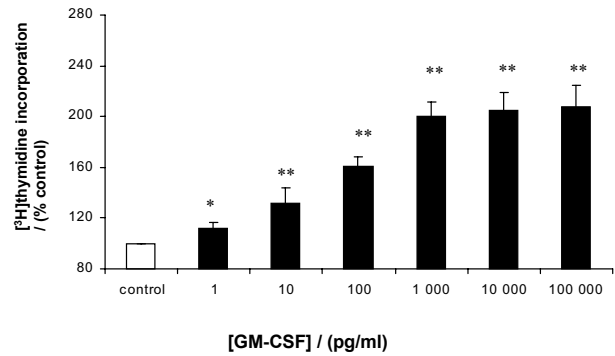


Figure 4
Effect of GM-CSF on proliferation of gastric epithelial cells. AGS cells were treated with increasing concentrations of GM-CSF for 24 hours. Cell proliferation was assessed by [³H]thymidine incorporation. Results expressed as means \pm standard deviation. * $P < 0.05$, ** $P < 0.01$ vs control

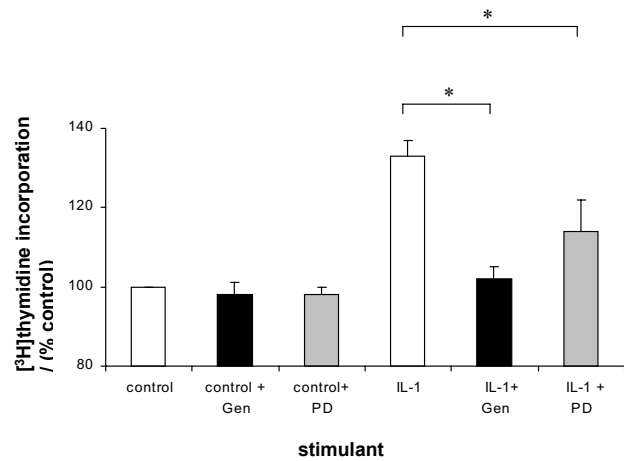


Figure 5
Effect of inhibition of tyrosine kinase and MEK activity on IL-1 β -stimulated gastric epithelial cell proliferation. AGS cells were treated with 10 ng/ml IL-1 β for 24 hours in the presence of the tyrosine kinase inhibitor genistein (100 μ M) or the MEK inhibitor PD 98059 (25 μ M). Proliferation was assessed by [³H]thymidine incorporation. Studies were performed in the presence of anti-GM-CSF antibody (5 μ g/ml). Results expressed as means \pm standard deviation. * $P < 0.01$ vs control

A portion of the stimulatory action of IL-1 β appears to be indirect. Neutralisation of GM-CSF in the media led to a significant reduction of IL-1 β -stimulated proliferation. AGS cells are known to secrete GM-CSF in response to IL-1 β [32]. GM-CSF itself was a potent stimulant of cell proliferation. Thus it seems likely that part of the growth stimulatory actions of IL-1 β are due to an autocrine inter-

mediary action of GM-CSF. There are limited previous data suggesting that GM-CSF stimulates the proliferation of non-haematopoietic cells; Dippold *et al* showed that exogenous GM-CSF stimulated the growth of two out of two cultures derived from gastric carcinomas and two out of nine pancreatic carcinoma cell lines [33,34]. However, unlike the current study, autocrine production of GM-CSF was not detectable.

Gastric epithelial cells also produce IL-8 in response to IL-1 β [31]. However in this model system, IL-8 alone had no pro-proliferative action and neutralisation of IL-8 did not affect the stimulatory action of IL-1 β . Therefore it is unlikely that IL-8 has an autocrine role in the growth-stimulatory action of IL-1 β . It is possible that other cytokines produced by the gastric epithelial cells in response to IL-1 β , *H. pylori* or other inflammatory insults could also play a role as autocrine or paracrine mediators of growth. It has recently been reported that another C-X-C chemokine, GRO/CINC-1, which is also upregulated in *H. pylori* infection stimulated proliferation in rat gastric epithelial cells [35]. The role of these other potential autocrine mediators deserves further study.

The results of the inhibitor studies strongly suggest that tyrosine kinase activity is essential for the growth promoting action of IL-1 β in AGS cells. The tyrosine kinase inhibitor genistein abolished the stimulatory action of IL-1 β . IL-1 β is known to activate a plethora of intracellular signalling pathways [31,36–39], but in the current situation there appears to be an absolute requirement for signalling via a tyrosine kinase, subsequent on receptor activation.

The mitogen-activated protein cascade is a well-characterised pathway mediating the cell growth-stimulatory actions of many growth factors and hormones. Inhibition of the ERK pathway, with the MEK inhibitor PD 98059, which prevents activation of ERKs by phosphorylation, had a significant inhibitory action against the stimulatory action of IL-1 β . This suggests that activation of the ERK pathway is important in mediating the growth stimulatory actions of IL-1 β . Activation of MAP kinase cascades, including the p42 and p44 ERK pathways and p46JNK and p55JNK c-Jun NH₂-terminal kinases by IL-1 β has been demonstrated in rat gastric epithelial cells [41,42], but the functional importance of these pathways was not examined. Activation of ERKs and JNKs was inhibited by genistein [41], consistent with the inference of the current study that MAPKs lie downstream of tyrosine activity in IL-1 β -induced signalling. However in the current study PD 98059 did not completely abolish the stimulatory action of IL-1 β , suggesting that alternative pathways, activated subsequent on tyrosine kinase activity, also play a role in the signalling of the proliferative responses to IL-1 β . Further studies are underway at present to examine these.

There are conflicting data available concerning the direct effects of IL-1 β on gastric epithelial proliferation. Although the current study and the study by Fan *et al* using stimulated-leukocyte conditioned media showed increased proliferation of the human AGS cell line [14], others have shown inhibition of serum, TGF- α and EGF-stimulated proliferation in RGM1 rat gastric epithelial cells by IL-1 β [41,42]. Tominaga *et al* showed that pre-treatment of RGM1 cells with IL-1 β for 6 hours inhibited proliferation at 24 hours, but the inhibitory effect was lost at 48 hours, [41]. The reasons for these discrepancies are not clear. They may reflect intrinsic differences between the cell lines, differences in activation and involvement of paracrine growth-promoting pathways, species variability, an effect specific for certain growth factors, or underlying differences in the biology of cell lines derived from cancer (AGS) or normal tissue (RGM1). The studies demonstrating inhibition of proliferation by IL-1 β were performed in the presence of powerful growth-promoting stimuli (high serum or specific growth factor concentrations in the culture media), whilst the current studies were performed in serum-free or 1% serum media. It is possible that the multiple signalling pathways activated by IL-1 β have differing effects on proliferation, the dominant effect depending on the complex inter-relationships of stimuli and signalling pathways under different circumstances. Pro-inflammatory cytokines such as IL-1 β and TNF- α activate different signalling pathways with divergent results and different time-courses in gastric endocrine and parietal cells [37,38,43–46]. Further studies are underway examining the specific roles of the various signalling pathways in gastric epithelial cells under different conditions.

There are now strong epidemiological data linking *H. pylori* with gastric carcinoma. The carcinogenic process appears to involve a number of steps: *H. pylori*-induced inflammation progresses to atrophy, intestinal metaplasia, dysplasia and eventually carcinoma [47]. Similarly *H. pylori* infection in Mongolian gerbils readily induces gastric atrophy and cancer [6]. Whilst gastric carcinogenesis is undoubtedly a multifactorial process, involving pathogenic bacterial and host factors, including HLA status, diet and antioxidant status [47,48], it is clear that increased gastric epithelial proliferation, especially when relatively increased compared to apoptosis, is an important part of the pathway [49,50]. Increased gastric epithelial proliferation is typical of *H. pylori* infection; it is demonstrable throughout all stages of the infection and eradication of the infection reduces proliferation. Increased proliferation is an important marker of increased risk of gastrointestinal adenocarcinoma [12]. The mechanisms of the increased proliferation are not completely understood. *In vitro* studies testing cultures of *H. pylori* or constituents have given conflicting results in different systems, with different cell types and bacterial strains. Direct stimula-

tion of gastric epithelial proliferation by *H. pylori* has been reported by some authors [14,15], whilst either neutral effects [15] or increased apoptosis and decreased proliferation have been reported by others [16–18]. Fan *et al* reported that conditioned media from either *H. pylori* or mitogen activated lymphocytes directly stimulated AGS cell proliferation [14], suggesting that the inflammatory response could be partly responsible for the enhancement of epithelial proliferation. IL-1 β production is enhanced in *H. pylori* infection and this cytokine is regarded as being central to the regulation of the pro-inflammatory response in *H. pylori* infection.

IL-1 β is a profound inhibitor of gastric acid secretion *in vivo* [51] and in isolated parietal cells [37,38]. Genetic polymorphisms of the IL-1 β gene cluster causing increased transcriptional activity are associated with an increased risk of pre-cancerous and cancerous histological changes in *H. pylori* infection [21,22]. The general hypothesis explaining this observation has been that enhanced IL-1 β production consequent on *H. pylori* infection is responsible for greater suppression of acid secretion, which in turn allows greater colonisation of the acid-secreting body mucosa [24]. This greater colonisation provokes further inflammation, which ultimately leads to loss of specialised acid-secreting epithelium (atrophy) and further potentiates the vicious cycle of increased inflammation and diminished acid secretion [52]. Gastric atrophy significantly predisposes to cancer [3]. It is believed that atrophy in combination with mediators released in inflammation such as oxygen free radicals and nitric oxide, diet, anti-oxidant state and possibly bacterial overgrowth and generation of nitrosamines in the achlorhydric stomach [53] advance the histological changes, cause mutagenesis and drive progression to cancer.

An alternative, but not mutually exclusive, hypothesis is that the enhanced cytokine response directly enhances epithelial cell proliferation and itself predisposes to cancer, in addition to the effects on acid secretion. The increased cell turnover of this hyperproliferative response would itself make the mucosa more vulnerable to the mutagenic effects of free radicals and other toxic products generated in the achlorhydric inflamed stomach.

Elsewhere in the gastrointestinal tract, IL-1 β has been shown to stimulate [³H]thymidine incorporation and increase cell number of cultured human colonic subepithelial myofibroblasts, which is thought to have importance in the remodelling of the mucosa in inflammation [54]. The pro-proliferative effects of IL-1 β in the gastrointestinal tract deserve further study, given the importance of this cytokine in the regulation of the mucosal inflammatory response.

Conclusions

The results of the current study suggest that IL-1 β and GM-CSF can directly stimulate gastric epithelial proliferation. This might explain the proliferative responses to conditioned lymphocyte media demonstrated by Fan *et al* [14]. Enhanced epithelial proliferation due to IL-1 β may contribute to the enhanced risk of gastric cancer and precancerous lesions in *H. pylori*-infected individuals with specific IL-1 β alleles associated with higher levels of production. IL-1 β stimulates proliferation via receptor-mediated activation of a tyrosine kinase pathway. Downstream signalling involves ERK-dependent and -independent pathways.

Further studies will be necessary to clarify the mechanisms involved in IL-1 β -stimulation of gastric epithelial proliferation, as well as data correlating IL-1 β genotype, IL-1 β protein production and epithelial proliferation *in vivo*. These will compliment the current study and further enhance our understanding of *H. pylori*-induced gastric carcinogenesis.

List of abbreviations

EGF – epidermal growth factor; ERK – extracellular signal related kinase; GM-CSF – granulocyte-macrophage colony stimulating factor; IL – interleukin; JNK – c-Jun NH₂-terminal kinase; MAP – mitogen activated protein; MTT – 3-[4,5-dimethylthiazol-2-yl]-2.5 diphenyl tetrazolium bromide; TCA – trichloroacetic acid; TGF- α – transforming growth factor alpha.

Competing interests

None declared.

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