

## MycoperonDB: A database of computationally identified operons and transcriptional units in Mycobacteria

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### Abstract

**Background:** A key post genomics challenge is to identify how genes in an organism come together and perform physiological functions. An important first step in this direction is to identify transcriptional units, operons and regulons in a genome. Here we implement and report a strategy to computationally identify transcriptional units and operons of mycobacteria and construct a database-MycoperonDB.

**Description:** We have predicted transcriptional units and operons in mycobacteria and organized these predictions in the form of relational database called MycoperonDB. MycoperonDB database at present consists of 18053 genes organized as 8256 predicted operons and transcriptional units from five closely related species of mycobacteria. The database further provides literature links for experimentally characterized operons. All known promoters and related information is collected, analysed and stored. It provides a user friendly interface to allow a web based navigation of transcription units and operons. The web interface provides search tools to locate transcription factor binding DNA motif upstream to various genes. The reliability of operon prediction has been assessed by comparing the predicted operons with a set of known operons.

**Conclusion:** MycoperonDB is a publicly available structured relational database which has information about mycobacterial genes, transcriptional units and operons. We expect this database to assist molecular biologists/microbiologists in general, to hypothesize functional linkages between operonic genes of mycobacteria, their experimental characterization and validation. The database is freely available from our website <http://www.cdfd.org.in/mycoperondb/index.html>.

### Background

Genome sequencing projects have generated large volumes of biological data which are difficult to manage and integrate effectively. This has thrown new challenges for

biologists who are now supposed to decode the complex physiological information encoded within these huge genomes. A first step in this direction is to know how the various genes are organized as transcription units, oper-

ons and regulon within a genome. We have previously reported strategies and tools, such as PredictRegulon and iCR, to identify regulons in bacterial genomes and identified DtxR/IdeR associated regulons in corynebacteria and mycobacteria [1-5]. At present we are interested in developing strategies to identify transcriptional units and operons of mycobacteria.

It is well known that genes belonging to the same operon are transcribed as a single mRNA molecule in all prokaryotes. Transcription starts as the RNA polymerase binds to the promoter and continues until it reaches a transcriptional terminator. The genes of the same operon are believed to be involved in similar metabolic and physiological processes. Hence operon prediction also provides important clues to the functional relationships between the operonic genes, which can then be taken up by the experimental biologist for further validation.

A number of computational and experimental approaches are being attempted to find out which all genes are together in a genome to perform a physiological function. Among experimental approaches, RNase Protection Assay, Dot Blot or Real Time PCR are generally used to define operon boundaries [6-9] but using these techniques for all the genes of a genome is expensive affair. A number of computational methods have been published for operon prediction [10-13] and a number of genome specific databases are also available that provide genome wide operon information [14,15].

Recently a database ODB was published [16] which has known and putative operons of many prokaryotic species including mycobacteria. However many mycobacterial transcriptional units and operons, even some known operons, are missing in this database. The advance search option requires great labor and expertise as well as external information from an average microbiologist which the latter may find difficult to provide. Therefore, there is a need to carryout more focused prediction of transcriptional units and operon in a group of related microorganisms. Such prediction and the resultant specialized database are likely to be more useful for specific research domain than global predictions. A more focused prediction in an organism also allows the researcher to revisit, track development regularly and update these databases as the research progresses in the field. Good examples of such databases are RegulonDB for *E. coli*, DBTBS for *B. subtilis* and PlasmoDB for Plasmodia [15,14,17].

We present here a promising mycobacterial database MycoperonDB, which has all known data related to mycobacterial genes, including gene sequences, encoded protein sequences, known promoters, known & predicted operons and related pubmed links. These data are precom-

puted so that all information can be quickly accessed. The definitions of the different terms used in transcriptomics as well as one or two lines description of the important mycobacterial genes have been given on the help page as glossary. The position of different important motifs can also be searched in this database. This database will be significantly useful for the researchers working with mycobacteria. This database is an ongoing effort to increase the coverage of more and more mycobacterial species, as and when their genome sequences become available. Some of these species include *Mycobacterium smegmatis*, *Mycobacterium w* etc. At present, around 8256 operons are being reported in 5 mycobacterial genomes which include *M.tuberculosis H37Rv*, *M.tuberculosis CDC1551*, *M.bovis*, *M.avium*, *M.leprae*.

#### **Construction and contents**

The overall process of transcription units and operon prediction involved multiple stages. Perl Scripts were written and used at every stage of operon prediction. These stages are-

#### **Retrieval of sequences**

The complete genome sequences of all species of mycobacteria with original annotations were downloaded from NCBI [18].

#### **Orientation analysis**

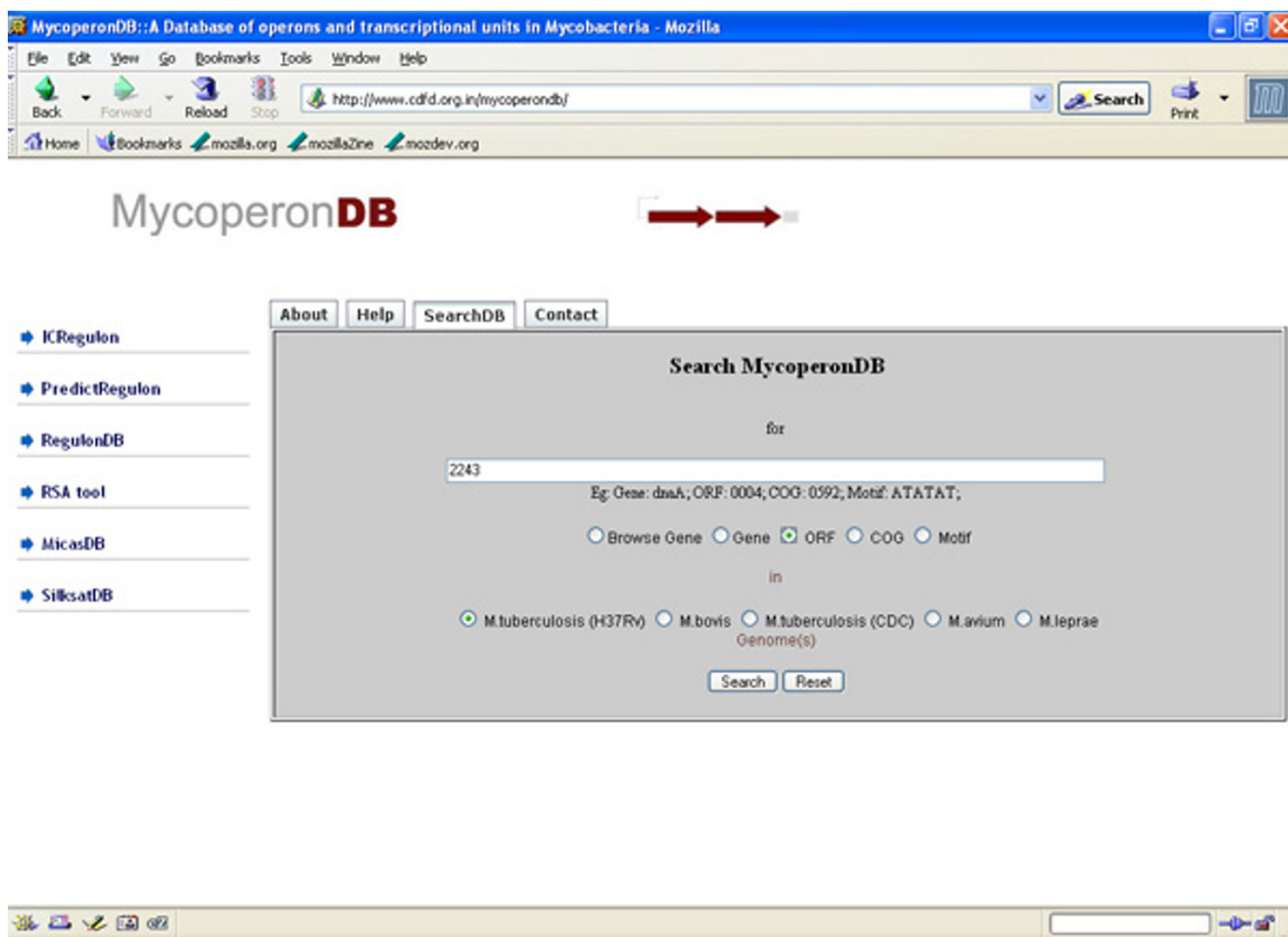
Genes which can be part of same operon must have same orientation. Considering this, all adjacent genes with same orientation were identified and grouped together.

#### **Intergenic distance analysis**

Genes in an operon are often closely located on the genome as compared to those which are not in the same operon. Hence after orientation, this is another indicator to identify the operons. The intergenic distances between adjacent genes in the same orientation were calculated from the corresponding coordinates using the formula:  $\text{distance}_{PQ} = \text{gene Q start position} - \text{gene P end position}$ . In general genes were passed to next operon if distance was greater than 300 bp. This cut-off was taken from *E.coli* operon prediction [19].

#### **Transcriptional terminators analysis**

Transcription terminator site is a site where transcription terminates. Genes flanking the terminator site cannot be in the same operon. GCG Terminator program from the GCG Wisconsin software package was used to identify rho-independent transcriptional terminators. Output of GCG Terminator program was parsed for S-value >0. Finally those terminators were considered which were in the region between -20 to +200 nucleotide around the stop-codon of each mycobacterial gene of an operon (operon boundary after step 1). The genes having the ter-



**Figure 1**  
**Search form of MycoperonDB.** SearchDB is a html form which takes input from the user. The query, which can be the gene name, ORF number, COG value or motif sequence, as given in the example, should be provided in the text box. In accordance with the query, user should select the nature of the query and the species of interest by clicking appropriate radio buttons before clicking the search button.

minator sites at the end were considered as end of the transcription units and operons.

**Conserved gene cluster analysis**

Conserved gene clusters among genomes were identified as orthologs either on the basis of gene orders or on the basis of clusters of orthologous groups (COGs). If conserved gene clusters (adjacent genes with same orientation grouped together in more than one species) were found, then intergenic distance criteria as well as terminator criteria was relaxed, i.e. if the genes are clustered among species, they were kept in one operon.

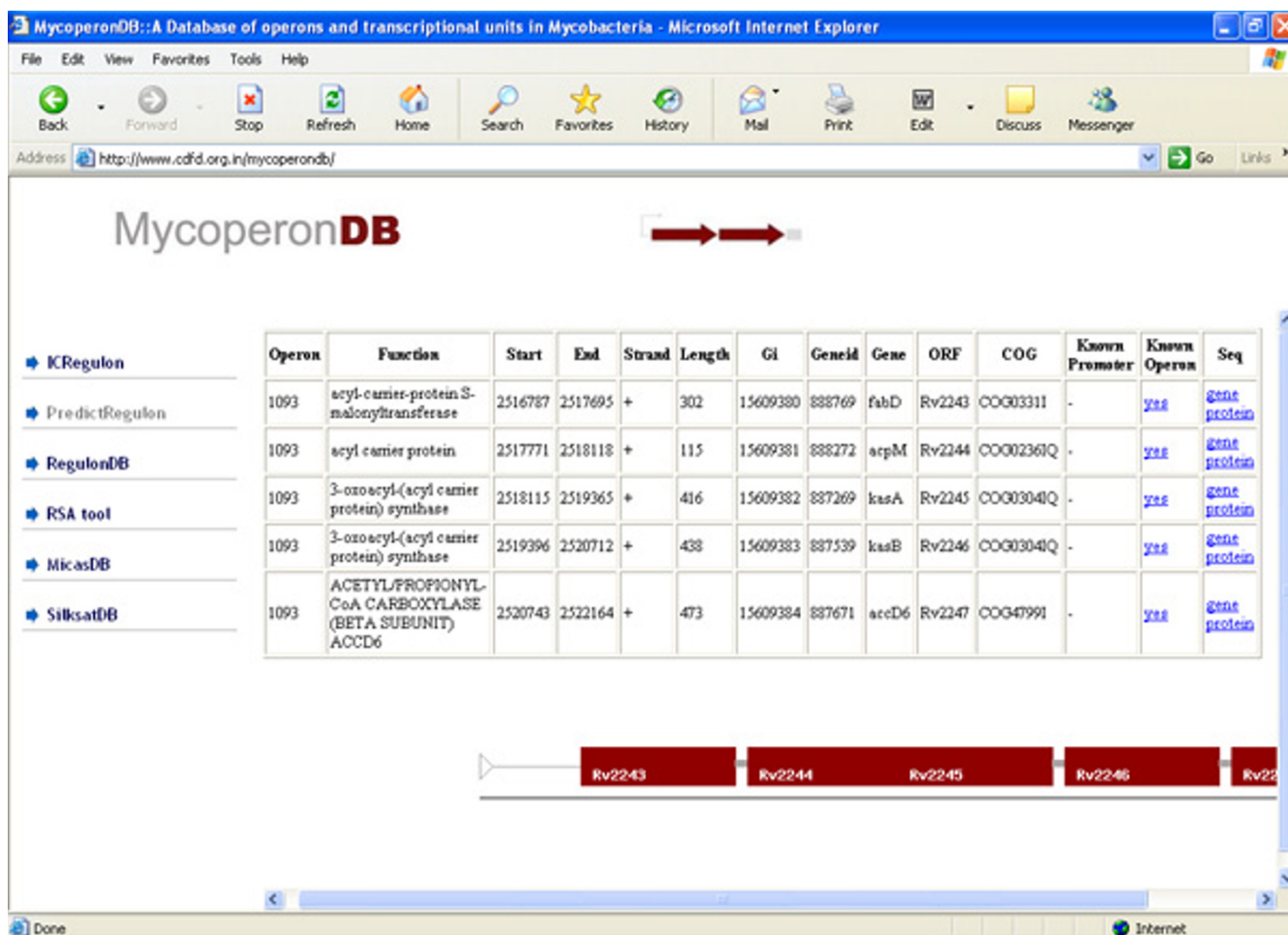
**Integration of literature information**

We scanned mycobacteria literature for reports on known transcription units, operons, promoters, and transcription start points of individual mycobacterial genes. Pubmed Id

of these identified literatures was integrated with our computational prediction, for the easy and quick browsing of the articles having detailed information on promoter and operon characterization. For the published information on promoters in any one species of mycobacteria, the homologous sequences in other species were searched computationally. The search results were also incorporated in the table with the same pubmed ID.

**Development of relational database**

We structured our data in the form of database. A relational database, MycoperonDB, was constructed using MySQL database management system (DBMS) to store and manage all information. MycoperonDB is currently composed of 6 tables. At present this database has information for only those mycobacterial species whose genomes are published and are available at NCBI but the



**Figure 2**  
**Output page of MycoperonDB.** A typical output html page which shows the result of the user's query. The query in this case was ORF number 2243 and the species selected was *M.tuberculosis* H37Rv. The output of a search has two parts: a table and a drawing. The table shows that the query ORF is part of an operon that consists of 5 genes. The last but one column of the table shows that this is a known operon and it is hyperlinked to the relevant pubmed ID which in this case is 12464486. The last column of the table provides a quick hyperlink to gene/protein sequences of the listed operon. Each gene that is part of selected operon is drawn as maroon colored rectangle with its ORF number written on it. The drawing has a grey arrow head which depicts the forward or reverse orientation of the operonic genes on the genome.

same method can be used to extend the database to other genomes.

**Web Interface**

In order to query the MycoperonDB database a web interface was developed using HTML, PHP, CSS and Javascript. This interface is available from our website.

**Utility and discussion**

MycoperonDB aims to provide a platform to the researchers interested in mycobacteria, for a quick overview of operon and transcription unit organization of a given

gene and all the related literature information like position of promoters/tsps, pubmed links, sequences of individual genes, and definition of most of the terms of mycobacterial gene regulatory circuits. A help page is also provided to guide the users step by step through the database.

**ORF search**

The user can type ORF number, or gene name in the search box and the result page will show the gene cluster (if the operon has more than one gene) including the query gene with other relevant information as mentioned

above (Figure 1). Separate clickable button is given for the DNA and protein sequences of the individual genes of the operon (Figure 2).

### Motif Search

The user can type any motif of interest in the search box and MycoPeronDB returns the position of that motif in the whole genome. The search can be done either in one species or in more species to know the homologs of the motif across the species. If the position of the motif does not fall in the upstream region (-500 bases) of any gene, then the result page declares no operon context.

### Analyses of prediction data

We have extensively searched literature to find out the known mycobacterial operons to test how much the predictions are deviated from the actual operons. In most of the cases the predictions were in agreement with the experimental observations. For example, *mceI* operon has been shown to be transcribed as a 13 gene polycistronic message in *M. tuberculosis* [20] which is in agreement with our prediction. In our H37Rv operon table Rv0166 to Rv0178 are together. Virulence operon in *M. tuberculosis* has been reported [21] and when checked in our operon table, all three genes Rv0986 to Rv0988 of this operon were found to be together. Similarly there are a number of examples like, *embCAB* operon [22], *ini* operon [23], *mymA* operon [24], *kasA* operon [25-27] etc for which our predictions were found to be correct.

In few cases, such as *nat* operon reported in *M. bovis* [28], *devR* operon, *ent* operon etc reported in *M. tuberculosis* [29,30], our prediction shows a few additional genes than reported which needs to be checked experimentally.

### Conclusion

We have predicted transcriptional units and operons in mycobacteria and organized these predictions in the form of a relational database called MycoPeronDB. We further provide additional information about known and experimentally demonstrated operons, promoters and their literature links. The strengths of this database are in its simplicity, its free web accessibility, its specificity, its comprehensiveness for published mycobacterial genomes and its interactive graphical interface. This database is part of our broad effort to characterize regulons, operons and transcriptional units in mycobacteria. This database can be a practical solution for the complexity of mycobacterial genome and it is expected to assist molecular biologists as well as microbiologists dealing with mycobacteria.

### Authors' contributions

SR: Computational predictions and literature search for relevant data.

RG: Designed web page and data links.

AR: Designed web pages; designed the project and coordination.

All authors read and approved the final manuscript.

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### References

1. Yellaboina S, Seshadri J, Kumar MS, Ranjan A: **PredictRegulon: a web server for the prediction of the regulatory protein binding sites and operons in prokaryote genomes.** *Nucleic Acids Res* 2004, **32**:W318-W320.
2. Ranjan S, Seshadri J, Vindal V, Yellaboina S, Ranjan A: **iCR: a web tool to identify conserved targets of a regulatory protein across the multiple related prokaryotic species.** *Nucleic Acids Res* 2006, **34**:W584-W587.
3. Yellaboina S, Ranjan S, Vindal V, Ranjan A: **Comparative analysis of iron regulated genes in mycobacteria.** *FEBS Lett* 2006, **580**:2567-2576.
4. Prakash P, Yellaboina S, Ranjan A, Hasnain SE: **Computational prediction and experimental verification of novel IdeR binding sites in the upstream sequences of Mycobacterium tuberculosis open reading frames.** *Bioinformatics* 2005, **21**:2161-2166.
5. Yellaboina S, Ranjan S, Chakhaiyar P, Hasnain SE, Ranjan A: **Prediction of DtxR regulon: identification of binding sites and operons controlled by Diphtheria toxin repressor in Corynebacterium diphtheriae.** *BMC Microbiol* 2004, **4**:38.
6. Ouellette SP, AbdelRahman YM, Belland RJ, Byrne GI: **The Chlamydia pneumoniae Type III Secretion-Related IcrH Gene Clusters Are Developmentally Expressed Operons.** *J Bacteriol* 2005, **187**:7853-7856.
7. Woolley RC, Vedyappan G, Anderson M, Lackey M, Ramasubramanian B, Jiangping B, Borisova T, Colmer JA, Hamood AN, McVay CS, Fralick JA: **Characterization of the Vibrio cholerae vceCAB Multiple-Drug Resistance Efflux Operon in Escherichia coli.** *J Bacteriol* 2005, **187**:5500-5503.
8. Lynch D, O'Brien J, Welch T, Clarke P, Ó Cuív P, Crosa JH, O'Connell M: **Genetic Organization of the Region Encoding Regulation, Biosynthesis, and Transport of Rhizobactin a siderophore Produced by Sinorhizobium meliloti.** *J Bacteriol* 2001, **183**:2576-2585.
9. van Boxtel RA, van de Klundert JA: **Expression of the Pseudomonas aeruginosa Gentamicin Resistance Gene aacC3 in Escherichia coli.** *Antimicrob Agents Chemother* 1998, **42**:3173-3178.
10. Price MN, Huang KH, Alm EJ, Arkin AP: **A novel method for accurate operon predictions in all sequenced prokaryotes.** *Nucleic Acids Res* 2005, **33**:880-892.
11. Edwards MT, Rison SC, Stoker NG, Wernisch L: **A universally applicable method of operon map prediction on minimally annotated genomes using conserved genomic context.** *Nucleic Acids Res* 2005, **33**:3253-62.
12. Chen X, Su Z, Xu Y, Jiang T: **Computational prediction of operons in Synechococcus sp. WH8102.** *Genome Inform* 2004, **15**:211-222.
13. Westover BP, Buhler JD, Sonnenburg JL, Gordon JL: **Operon prediction without a training set.** *Bioinformatics* 2005, **21**:880-888.
14. Ishii T, Yoshida K, Terai G, Fujita Y, Nakai K: **DBTBS: a database of Bacillus subtilis promoters and transcription factors.** *Nucleic Acids Res* 2001, **29**:278-280.

15. Salgado H, Santos-Zavaleta A, Gama-Castro S, Millan-Zarate D, Diaz-Peredo E, Sanchez-Solano F, Perez-Rueda E, Bonavides-Martinez C, Collado-Vides J: **RegulonDB (version 3.2): transcriptional regulation and operon organization in Escherichia coli K-12.** *Nucleic Acids Res* 2001, **29**:72-74.
16. Okuda S, Katayama T, Kawashima S, Goto S, Kanehisa M: **ODB: a database of operons accumulating known operons across multiple genomes.** *Nucleic Acids Res* 2006, **34**:D358-D362.
17. Bahl A, Brunk B, Crabtree J, Fraunholz MJ, Gajria B, Grant GR, Ginsburg H, Gupta D, Kissinger JC, Labo P, Li L, Mailman MD, Milgram AJ, Pearson DS, Roos DS, Schug J, Stoeckert CJ Jr, Whetzel P: **PlasmoDB: the Plasmodium genome resource. A database integrating experimental and computational data.** *Nucleic Acids Res* 2003, **31**:212-215.
18. **NCBI - Complete Microbial Genomes** [<http://www.ncbi.nlm.nih.gov/genomes/lproks.cgi>]
19. Yada T, Nakao M, Totoki Y, Nakai K: **Modeling and predicting transcriptional units of Escherichia coli genes using hidden Markov models.** *Bioinformatics* 1999, **15**:987-993.
20. Casali N, White AM, Riley LW: **Regulation of the Mycobacterium tuberculosis mceI operon.** *J Bacteriol* 2006, **188**:441-9.
21. Rosas-Magallanes V, Deschavanne P, Quintana-Murci L, Brosch R, Gicquel B, Neyrolles O: **Horizontal transfer of a virulence operon to the ancestor of Mycobacterium tuberculosis.** *Mol Biol Evol* 2006, **23**:1129-1135.
22. Sharma K, Gupta M, Pathak M, Gupta N, Koul A, Sarangi S, Baweja R, Singh Y: **Transcriptional control of the mycobacterial embCAB operon by PknH through a regulatory protein, EmbR, in vivo.** *J Bacteriol* 2006, **188**:2936-2944.
23. Ramaswamy SV, Amin AG, Goksel S, Stager CE, Dou SJ, El Sahly H, Moghazeh SL, Kreiswirth BN, Musser JM: **Molecular genetic analysis of nucleotide polymorphisms associated with ethambutol resistance in human isolates of Mycobacterium tuberculosis.** *Antimicrob Agents Chemother* 2000, **44**:326-336.
24. Singh R, Singh A, Tyagi AK: **Deciphering the genes involved in pathogenesis of Mycobacterium tuberculosis.** *Tuberculosis* 2005, **85**:325-335.
25. Hughes MA, Silva JC, Geromanos SJ, Townsend CA: **Quantitative proteomic analysis of drug-induced changes in mycobacteria.** *J Proteome Res* 2006, **5**:54-63.
26. Bhatt A, Kremer L, Dai AZ, Sacchettini JC, Jacobs WR Jr: **Conditional depletion of KasA, a key enzyme of mycolic acid biosynthesis, leads to mycobacterial cell lysis.** *J Bacteriol* 2005, **187**:7596-606.
27. Slayden RA, Lee RE, Barry CE 3rd: **Isoniazid affects multiple components of the type II fatty acid synthase system of Mycobacterium tuberculosis.** *Mol Microbiol* 2000, **38**:514-525.
28. Anderton MC, Bhakta S, Besra GS, Jeavons P, Eltis LD, Sim E: **Characterization of the putative operon containing arylamine N-acetyltransferase (nat) in Mycobacterium bovis BCG.** *Mol Microbiol* 2006, **59**:181-192.
29. Bagchi G, Chauhan S, Sharma D, Tyagi JS: **Transcription and autoregulation of the Rv3134c-devR-devS operon of Mycobacterium tuberculosis.** *Microbiology* 2005, **151**:4045-4053.
30. De Voss JJ, Rutter K, Schroeder BG, Barry CE 3rd: **Iron acquisition and metabolism by mycobacteria.** *J Bacteriol* 1999, **181**:4443-4451.

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