# COVER STORY: TOOLS

# Macrocycles by the trillions

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Japanese researchers have developed a method for creating large libraries of N-methylated peptide macrocycles,<sup>1</sup> a class of molecules that are sized between small molecules and biologics and have the potential to combine the pharmacological versatility of the former and the therapeutic specificity of the latter. **PeptiDream Inc.** has exclusively licensed the technology and entered collaborations with six pharmas.

Small molecules are typically less than 500 Da in size, whereas biologics start at approximately 5,000 Da and cover a range of one to two orders of magnitude. Macrocycles, a structure frequently found in natural products, range in size from 500–2,000 Da.

Due to their structural rigidity and high target affinity, the hope has been for these compounds to be able to modulate challenging targets typically reserved for biologics, such as protein-protein interactions, while retaining the advantages of small molecules—cell permeability and oral bioavailability.<sup>2</sup>

One challenge has been generating sufficiently large and diverse libraries of synthetic macrocycles to identify drug leads. To overcome this hurdle, a team led by Hiroaki Suga has now merged two technologies— one developed by Suga in 2008 to incorporate non-natural amino acids into large libraries of peptide macrocycles<sup>3</sup> and the other developed independently in 1997 by Jack Szostak and Hiroshi Yanagawa for displaying natural peptide and protein libraries on mRNA.<sup>4,5</sup>

Suga is a professor in the Department of Chemistry at **The University** of Tokyo. Yanagawa is a professor at Keio University. Szostak is a professor at Harvard Medical School and Massachusetts General Hospital.

The integrated method, called RaPID, starts from a cDNA library encoding non-natural peptides of 8–15 residues. The peptides contain a random mixture of 12 natural amino acids and 4 N-methylated (nonnatural) amino acids. Non-natural amino acids can increase the types of chemical groups in peptides beyond what is offered by natural amino acids. In particular, N-methylated amino acids increase a peptide's cell permeability.

The cDNA library is then transcribed to an mRNA library and linked to a second mRNA oligonucleotide that ends in a puromycin residue. The residue causes the growing peptide chain to be covalently connected to and displayed by its own mRNA template. *In vitro* translation of these mRNAs resulted in a library of about 10<sup>12</sup> N-methylated peptide macrocycles displayed on mRNA (*see* Figure 1, "Making macrocycles RaPID-ly").

As proof of principle, the Japanese team used the library to select for macrocycles that bound the ligase domain of ubiquitin protein ligase

E3A (UBE3A; E6AP). Identifying potent and selective inhibitors that interfere with the protein-protein interactions made by ubiquitin ligases has proven challenging. The team identified three distinct macrocycles that each bound to E6AP with low or subnanomolar affinity.

Results were published in Chemistry & Biology.

"This is a very clever technology. Synthetic tRNAs to introduce new amino acids in combination with mRNA allows you to create vast libraries of novel peptides," said Nick Terrett, CSO of macrocycle company **Ensemble Therapeutics Corp.** 

"To isolate drugs with more complex function, such as disrupting proteinprotein interactions, researchers are turning to larger molecules like peptides. The real value of [this] work is that extremely large and diverse libraries can

be produced and used in affinity-based selections, which greatly increases the likelihood that highly potent protein-protein interaction inhibitors can be isolated. These arguments also apply to cases where highly specific drugs are needed, for example, when attempting to inhibit one member in a family of closely related proteins," said Douglas Treco, president, CEO and cofounder of **Ra Pharmaceuticals Inc.** 

Ra Pharma is using *in vitro* display technologies to produce libraries of cyclic peptidomimetics to identify therapeutics in a variety of diseases. The company was cofounded by Szostak.

"The affinities described for the ubiquitin ligase are impressive. It will be key to demonstrate that high-affinity binders can be isolated for a broad range of targets," said Christian Heinis, assistant professor in the Institute of Chemical Sciences and Engineering at the **Swiss Federal Institute of Technology Lausanne**. "I like the RaPID approach very much since it allows the facile incorporation of non-natural amino acids."

Heinis is a cofounder of **Bicycle Therapeutics Ltd.**, a company developing cyclic peptide therapeutics using phage display.

However, Terrett said the molecules generated by RaPID could be too big to become drug leads. "The molecular weight is too high," as the macrocycles described in the paper were around 2,000 Da, he said. "My guess would be that cell membrane permeability and the *in vivo* bioavailability would be low."

Ensemble's macrocycles, which Terrett said are typically cell permeable, are in the range of 600–1,000 Da. For example, Ensemble's macrocyclic IL-17 antagonists are 700 Da and have low nanomolar potency. The compounds are in preclinical development for autoimmune and inflammatory diseases.

Ensemble synthesizes macrocycles using a process called DNAprogrammed chemistry that relies on DNA as a template but generates macrocycles using synthetic rather than peptide building blocks. "We are also using a purely synthetic process, so there is a lot of chemistry we can incorporate," noted Terrett.

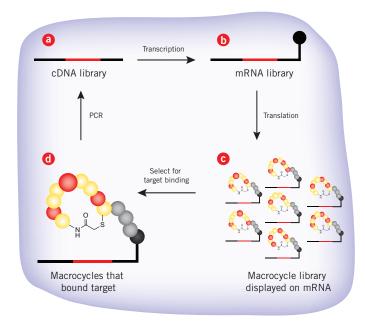
According to Terrett, Ensemble has generated a cumulative library of about 4 million macrocycles. He said Ensemble frequently has been

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# ANALYSIS

# **COVER STORY**



## Figure 1. Making macrocycles RaPID-ly. A team from The

**University of Tokyo** has developed a method called RaPID for generating libraries of trillions of N-methylated peptide macrocycles displayed on mRNA. First, a cDNA library is generated (red bar) that encodes 8–15 residue–long peptides containing a mixture of natural and non-natural amino acids [**a**]. The cDNA library is transcribed to an mRNA library and subsequently linked to a second mRNA oligo-nucleotide that terminates in a puromycin residue (black circle) [**b**]. When the mRNA library is translated, the puromycin causes the growing peptide chain to remain linked to its mRNA template. The displayed macrocyclic peptides contain a linker region (gray), natural amino acids (yellow) and N-methylated amino acids (red) [**c**]. After selecting for macrocycles that bind to a desired target, binding molecules can be identified and/or amplified for a second round of selection by PCR of the linked mRNA template [**d**].

able to find compounds with "micromolar to nanomolar potency against previously undruggable targets."

Ensemble recently extended their partnership with **Bristol-Myers Squibb Co.** to develop macrocycles against undisclosed targets.<sup>6</sup>

Terrett suggested adapting the RaPID technology "to make compounds that are smaller from the get-go. If you could make smaller cyclic peptides, you'd be at a better starting point for drug discovery. The compromise would be lower structural diversity. It's a trade-off."

## PeptiDreaming

PeptiDream, which was founded by Suga, is identifying macrocyclic drug leads using technology that is similar to RaPID but with a different, undisclosed display method.

"We developed an alternative to mRNA display that is simplified," said CSO Patrick Reid.

He said PeptiDream has identified hits with nanomolar to subnanomolar potency against enzymes and protein-protein interactions. "Macrocycle peptides seem to be able to target most anything— $\beta$ -sheets,  $\alpha$ -helices, random coils—we have found peptides that have bound to them all."

Reid disagreed with Terrett that size was the critical factor governing cell permeability. He said PeptiDream's technology can make cell-permeable macrocycles of 800–2,000 Da. "In the studies that we have done, size doesn't seem to matter that much," he said.

Reid added that hydrophobicity and the 3D structure of the macrocycle seem to be more important than size in determining cell permeability.

PeptiDream's most advanced program is a macrocycle that blocks influenza A virus hemagglutinin, an extracellular target, and prevents the virus from penetrating cells. The project is being funded by the Japanese government.

Reid said about half of the company's targets are extracellular and the other half are intracellular. PeptiDream's most advanced program against an intracellular target is an undisclosed protein-protein interaction being pursued as part of a pharma collaboration. PeptiDream and its undisclosed partner hope to start mouse studies in the next few months, according to Reid.

The next steps for Suga include designing new libraries with different scaffolds and developing new selection techniques, with an eye toward increasing cell permeability. "We need to look at the structural requirements for cell permeability. That is really the major direction that we are taking right now," he said.

Suga told *SciBX* that the University of Tokyo has received a patent covering technology for generating the engineered tRNAs that introduce non-natural amino acids into peptides. The university has filed for patents covering both the use of the engineered tRNAs for *in vitro* translation and the integrated RaPID method. The university also has filed for patents covering technologies for introducing *N*-methyl peptides, cyclizing the peptides and for other modifications. All have been exclusively licensed to PeptiDream.

Reid said the company has collaborations with **AstraZeneca plc**'s **MedImmune LLC** subsidiary, two undisclosed Japanese pharmas and three undisclosed non-Japanese pharmas.

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#### REFERENCES

- Yamagishi, Y. *et al. Chem. Biol.*; published online Dec. 23, 2011; doi:10.1016/j.chembiol.2011.09.013
  Contact: Hiroaki Suga, The University of Tokyo, Tokyo, Japan e-mail: hsuga@chem.s.u-tokyo.ac.jp
- 2. Driggers, E.M. et al. Nat. Rev. Drug Discov. 7, 608-624 (2008)
- 3. Kawakami, T. et al. Chem. Biol. 15, 32–42 (2008)
- 4. Nemoto, N. et al. FEBS Lett. 414, 405-408 (1997)
- Roberts, R.W. & Szostak, J.W. Proc. Natl. Acad. Sci. USA 94, 12297–12302 (1997)
- 6. Edelson, S. BioCentury 19(4), A7-A8; Jan. 17, 2011

### COMPANIES AND INSTITUTIONS MENTIONED

AstraZeneca plc (LSE:AZN; NYSE:AZN), London, U.K. Bicycle Therapeutics Ltd., Cambridge, U.K. Bristol-Myers Squibb Co. (NYSE:BMY), New York, N.Y. Ensemble Therapeutics Corp., Cambridge, Mass. Harvard Medical School, Boston, Mass. Keio University, Tsuruoka, Japan Massachusetts General Hospital, Boston, Mass. MedImmune LLC, Gaithersburg, Md. PeptiDream Inc., Tokyo, Japan Ra Pharmaceuticals Inc., Boston, Mass. Swiss Federal Institute of Technology Lausanne, Lausanne, Switzerland The University of Tokyo, Tokyo, Japan