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Repellency and insecticidal activity of seven Mugwort (*Artemisia argyi*) essential oils against the malaria vector *Anopheles sinensis*

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Anopheles sinensis is the main vector of malaria with a wide distribution in China and its adjacent countries. The smoke from burning dried mugwort leaves has been commonly used to repel and kill mosquito adults especially in southern Chinese provinces. In this study, the essential oils of mugwort leaves collected from seven provinces in China were extracted by steam distillation and their chemical compositions were analyzed. Among a total of 56–87 chemical constituents confirmed by GC–MS analyses, four compounds, eucalyptol, β -caryophyllene, phytol and caryophyllene oxide, were identified with appearances from all seven distilled essential oils. The effectiveness varied in larvicidal, fumigant and repellent activities against *An. sinensis* from these seven essential oils with different geographic origins. The essential oil from Hubei province showed the highest larvicidal activity against the 4th instar larvae of *An. sinensis*, with a median lethal concentration at 40.23 $\mu\text{g/mL}$. For fumigation toxicity, essential oils from 4 provinces (Gansu, Shandong, Sichuan and Henan) were observed with less than 10 min in knockdown time. The essential oil distilled from Gansu province displayed the highest repellent activity against *Anopheles* mosquitoes and provided similar level of protection as observed from DEET. Eucalyptol was the most toxic fumigant compound and phytol showed the strongest larvicidal activity among all tested mugwort essential oil constituents.

Abbreviations

Ar	<i>Artemisia</i>
An	<i>Anopheles</i>
Ar. argyi	<i>Artemisia argyi</i>
An. sinensis	<i>Anopheles sinensis</i>
Ae	<i>Aedes</i>
CQ	Chongqing province
SC	Sichuan province
YN	Yunnan province
GS	Gansu province
HN	Henan province
HB	Hubei province
SD	Shandong province

Mosquitoes are considered as important medical insect pests worldwide, and they, as vectors, transmit many diseases including malaria, filariasis, yellow fever and dengue fever^{1–3}. Malaria has been reported as the most common mosquito-borne disease and present a great threat to human life mainly spread by *Anopheles* mosquitoes⁴. Every year, approximately 250 million people have been reported being infected with malaria, which has caused over one million deaths worldwide⁵. *Anopheles sinensis* is the No.1 vector of malaria in China⁶. The control of this vector is mainly managed with massive applications of various insecticides. However, the over uses of these toxic chemicals have resulted in strong insecticide resistance^{7,8}.

Currently for mosquito control in China, pyrethroids are primary chemical insecticides being heavily used. N,N-Diethyl-3-methylbenzamide (DEET) is the most commonly repellent used for the prevention against *An.*

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Oil ^a	LC ₅₀ (µg/mL) ± SE ^b	95% Confidence limits	Toxicity regression equation (y = a + bx) ^c	χ ²
CQ	48.77 ± 7.40	41.60–55.18	y = -1.85 + 0.03x	3.21*
SC	54.24 ± 6.30	49.12–59.41	y = -3.14 + 0.05x	2.18*
YN	61.82 ± 5.48	53.61–69.39	y = -1.77 + 0.03x	4.51*
HB	40.23 ± 1.0	32.42–46.72	y = -1.59 + 0.04x	4.95*
HN	55.93 ± 0.29	49.51–62.12	y = -2.24 + 0.04x	2.99*
SD	59.31 ± 6.40	51.27–66.70	y = -1.76 + 0.03x	0.37*
GS	55.20 ± 8.50	48.00–61.88	y = -1.92 + 0.03x	2.58*

Table 1. Larvicidal activity of essential oils distilled from seven geographical regions of mugwort leaves in China against the 4th instar larvae of *An. sinensis*. ^aCQ, SC, YN, HB, HN, SD and GS: essential oils from Chongqing, Sichuan, Yunnan, Hubei, Henan, Shandong and Gansu province/municipality, respectively. ^bDetermined by log-probit analysis with at least 6 concentrations, and three repetitions “SE” denote standard error. ^cy stands for the probability transformation value of percent mortality of mosquitoes, and x for the concentration of essential oil, respectively. *Significant at $p < 0.05$.

sinensis. However, extensive use of pyrethroids has caused increasing insecticide resistance, and also created adverse effects on human health and their surroundings^{9–11}. The toxicology of DEET has been more closely investigated and has found issues in safety for human use, including use on children, pregnant women, and lactating women^{12–14}. There is an urgent need to develop alternative human and environmentally friendly mosquito control strategies. In recent years, natural products have gradually become the primary sources of novel mosquito repellents and larvicides due to low-toxic, quick biodegradation rates, and less side effects on hosts and non-target organisms^{15–17}. Plant essential oils, extracted from *Nepeta cataria*, *Severinia monophylla*, *Syzygium aromaticum* and *Cymbopogon citratus* have been reported to be effective in repelling mosquito adults^{18–20}. Besides, some of them have also exhibited larvicidal activity, including those from *Laurencia dendroidea*, *Cunninghamia konishii* and *Azadirachta indica*^{21–23}.

The Chinese mugwort plant, *Artemisia argyi*, is a traditional medicine plant widely distributed in southern provinces with a long cultivation history. Extracts of this plant have been reported to possess immunomodulatory, neuroprotective insecticidal properties with effectiveness against bacteria, dampness, hemostasis, cancer, and inflammation^{24–26}. Mugwort plants have been widely used to repel mosquitoes by burning the harvested dried leaves for thousands of years at countryside in China²⁷. The essential oil of those plant leaves exhibits strong contact fumigant toxicity against *Lasioderam serricornis* adults²⁸. The oils of four subspecies of mugwort plants including *Ar. feddei*, *Ar. gmelinii*, *Ar. manshurica*, and *Ar. olgensis* deterred mosquito biting of adult *Ae. aegypti*²⁹. The essential oil of *Ar. vulgaris* provided 100% mortality against *Ae. aegypti* larvae after 8-h exposure at a concentration of 500 ppm.

In the study, we extracted the essential oils from plants of *Ar. argyi* collected from seven provinces/municipal areas using a steam distillation technique, and investigated their larvicidal, fumigant and repellent activity against *An. sinensis*. We also analyzed their chemical compositions using gas chromatography mass spectrometer (GC–MS), and evaluated the toxicity of four common predominant compounds of these oils against *An. sinensis*. The results from this study may provide some new insights for further research in use of the mugwort essential oil as an alternative and targeted control and preventative measure against *Anopheles* mosquitoes.

Results

Insecticidal and repellent activity of distilled *Ar. argyi* essential oils. The essential oil of Hubei province (HB) showed the highest larvicidal effect with a median lethal concentration (LC₅₀) value at 40 µg/mL, followed by oils from mugwort plants collected in Chongqing (CQ) (LC₅₀ = 49 µg/mL), Sichuan (SC) (LC₅₀ = 54 µg/mL), Hainan (HN) (LC₅₀ = 56 µg/mL), Gansu (GS) (LC₅₀ = 55 µg/mL) and Shandong (SD) (LC₅₀ = 59 µg/mL) and Yunnan (YN) (LC₅₀ = 62 µg/mL) (Table 1). The larvicidal activity of all seven essential oils are significantly effective against *An. sinensis* in comparison of control in which no dead larvae were observed.

All seven oils showed significant repellent activities against female *An. sinensis* adults compared to the negative control (Fig. 1). Similar levels of repellency as DEET were demonstrated from 4 oils (CQ, HN, SD and GS), with the highest from GS oil. The mean % of repellent longevity from seven mugwort essential oils were presented in Table 2. The essential oil extracted from leaves collected in GS provided almost 66% of protection against mosquitoes at the 65th minutes post application, which was similar to the effectiveness found from 10% DEET.

Fumigant toxicity of distilled *Ar. argyi* essential oils. The median knockdown time (KT₅₀) ranged from as short as 4.7 min from essential oils from GS, to over 50 min of oils from YN province (Table 3). Similar trends appeared also in the median lethal dose (LD₅₀), which oils of GS showed the strongest toxicity against the female adults of *An. sinensis*, and the least from oils of YN. The essential oil from GS has the highest fumigant toxicity with a LD₅₀ value at 9.40 µL/L (Table 4).

Chemical compositions of *Artemisia argyi* essential oils. In total, 56–87 chemical constituents were identified from *Ar. argyi* essential oils from seven geographical origins, with the highest number of compounds (87) found in the oil from Gansu province (Table 5). Four chemical compounds, β-caryophyllene, caryophyllene

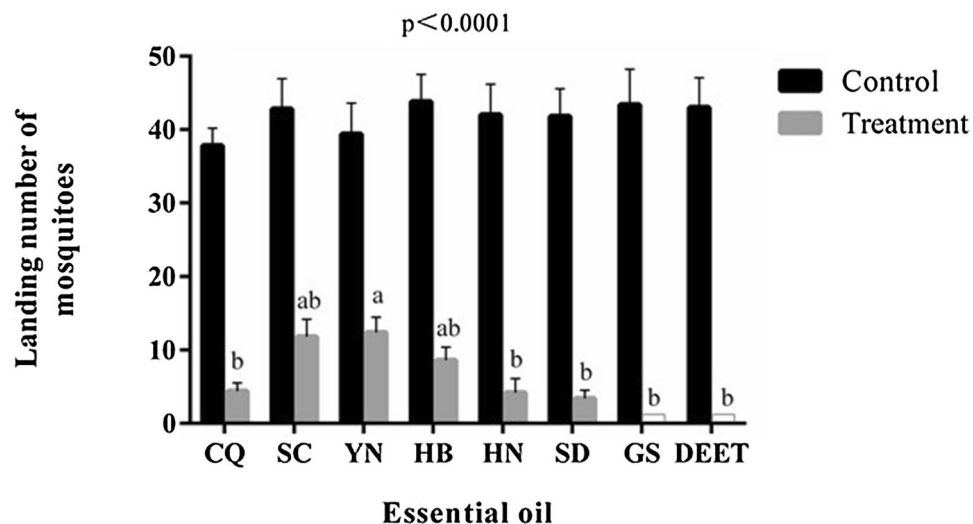


Figure 1. The first 5-min observed landing rates of starved *An. sinensis* on human skins treated with seven *Ar. argyi* essential oils extracted from different geographical areas. The significances between each treatment and control group are less than 0.0001 ($p < 0.0001$). Different letters on each treatment bars indicate significantly different between among the seven essential oils tested and the positive control, DEET (ANOVA).

Oil treatments (1.5 $\mu\text{L}/\text{cm}^2$) ^a	Minutes post application*				
	5 min	20 min	35 min	50 min	65 min
CQ	88.32 \pm 3.19 ^b	79.18 \pm 1.63 ^b	66.82 \pm 3.20 ^b	57.49 \pm 5.85 ^b	34.55 \pm 5.02 ^b
SC	72.53 \pm 3.95 ^c	61.41 \pm 1.33 ^c	42.81 \pm 2.37 ^d	27.44 \pm 1.38 ^d	14.78 \pm 1.05 ^e
YN	68.62 \pm 2.93 ^c	58.91 \pm 1.15 ^c	37.56 \pm 0.67 ^d	22.30 \pm 1.58 ^d	9.02 \pm 2.33 ^e
HB	80.09 \pm 3.03 ^c	74.98 \pm 1.74 ^b	57.97 \pm 5.65 ^c	40.04 \pm 1.73 ^c	28.94 \pm 2.68 ^b
HN	90.21 \pm 3.62 ^b	80.51 \pm 2.02 ^b	69.30 \pm 4.40 ^b	59.20 \pm 2.51 ^b	37.29 \pm 2.34 ^b
SD	91.90 \pm 2.42 ^b	81.48 \pm 2.47 ^b	71.67 \pm 5.95 ^b	61.31 \pm 1.49 ^b	39.43 \pm 2.94 ^b
GS	100 \pm 0.00 ^a	99.57 \pm 0.97 ^a	94.28 \pm 3.21 ^a	76.01 \pm 1.06 ^a	65.68 \pm 3.26 ^a
DEET (10%)	100 \pm 0.00 ^a	99.59 \pm 0.91 ^a	95.04 \pm 3.21 ^a	83.27 \pm 3.69 ^a	79.81 \pm 4.28 ^a

Table 2. The percentages of repellencies of seven *Ar. argyi* essential oils and DEET against the four-day old *An. sinensis* adults during different exposed times. *The repellent rates with different superscript letters in the same column are significantly different at $p < 0.05$. The rates were determined with three replications. CQ, SC, YN, HB, HN, SD and GS: essential oils from Chongqing, Sichuan, Yunnan, Hubei, Henan, Shandong and Gansu province/municipality, respectively.

Oil	KT ₅₀ (min) ^a	95% Confidence limits	Toxicity regression equation ($y = a + bx$)	χ^2
CQ	13.85 \pm 3.5	13.08–14.62	$y = -4.33 + 0.31x$	0.29*
SC	10.18 \pm 3.4	9.43–10.91	$y = -3.71 + 0.36x$	0.83*
YN	50.62 \pm 8.6	48.19–53.52	$y = -2.69 + 0.05x$	12.95*
HB	16.77 \pm 4.7	15.39–18.08	$y = -1.87 + 0.11x$	5.55*
HN	10.02 \pm 2.8	9.02–10.96	$y = -2.21 + 0.22x$	3.02*
SD	9.32 \pm 2.2	8.61–10.05	$y = -3.36 + 0.36x$	2.84*
GS	4.71 \pm 1.2	3.50–5.32	$y = -2.45 + 0.51x$	0.33*

Table 3. Median lethal knockdown activity (mins) of the essential oils extracted from seven geographical regions of *Ar. argyi* against female adults of *An. sinensis*. ^aKT₅₀ represents the knockdown time of half individuals, which was determined by log-probit analysis. Three experimental repeats with the essential oil dosage of 2 μL . *Significant at $p < 0.05$. CQ, SC, YN, HB, HN, SD and GS: essential oils from Chongqing, Sichuan, Yunnan, Hubei, Henan, Shandong and Gansu province/municipality, respectively.

Oil	LD ₅₀ (μL/L) ^a	95% confidence limit	Toxicity regression equation (y = a + bx)	χ ²
CQ	14.95 ± 7.8	13.32–16.43	y = -1.65 + 0.11x	2.15*
SC	22.90 ± 5.5	20.82–25.08	y = -1.56 + 0.06x	2.91*
YN	51.26 ± 6.1	42.87–71.69	y = -2.36 + 0.04x	2.77*
HB	48.74 ± 2.1	39.09–76.29	y = -1.39 + 0.02x	4.15*
HN	26.49 ± 4.5	24.25–29.13	y = -1.70 + 0.06x	3.00*
SD	18.66 ± 1.2	16.75–20.44	y = -1.54 + 0.08x	3.55*
GS	9.40 ± 3.6	5.15–12.08	y = -1.51 + 0.16x	8.62*

Table 4. Fumigation toxicity of the essential oils extracted from seven geographical regions of *Ar. argyi* in China against female adults of *An. sinensis*. ^aDetermined with at least 6 concentrations at μL/L. *Significant at $p < 0.05$. CQ, SC, YN, HB, HN, SD and GS: essential oils from Chongqing, Sichuan, Yunnan, Hubei, Henan, Shandong and Gansu province/municipality, respectively.

oxide, eucalyptol and phytol were the most abundant and detected from all seven oils. There were significant differences in compositions of various compounds among oils extracted from seven different regions. Eucalyptol was found over 20% in GS essential oils, but only 5.5% from YN province. Only one oil from HB province contained over 16% phytol, with less than 10% from oils of the rest of regions. Another monoterpenoid, thujone, was detected over 16% from the SC oil.

Larvicidal and fumigant toxicity of the 4 dominant compositional compounds. All four individual compounds showed significant toxicity against both larvae and adults of *An. sinensis* (Tables 6 and 7). Phytol was the strongest larvicidal compound, with the LC₅₀ at 16.03 μg/mL. Significant larvicidal activity was also found from caryophyllene oxide (LC₅₀ = 39.09 μg/mL), relative to those of eucalyptol and β-caryophyllene (Table 6). Eucalyptol exhibited the highest fumigant lethal activity against adult mosquitoes among the four tested compounds with the shortest time to kill at the LT₅₀ value of 5.48 μL/L.

Discussion

It is not surprising that the essential oils of the mugwort plant leaves have shown strong larvicidal activity against *An. sinensis*, as Wang³⁰ have previously reported that *Ar. argyi* essential oil exhibiting larvicidal activity against *Culex* mosquitoes. Interestingly, significant differences in larvicidal activity have been demonstrated among the *Ar. argyi* oils from 7 different geographic regions, which may result from various quantities of some terpene and monoterpenoid compounds as their major compositional constituents³¹. Eucalyptol, β-caryophyllene, caryophyllene oxide and phytol, the four most common compounds found from all distilled oils in seven geographic origins, have been reported with moderate to strong larvicidal activity against various mosquito larvae. For instance, phytol has been reported with larvicidal activity against *Aedes* and *Culex* mosquitoes as one of compositional compounds from several essential oils^{32–35}. From the distilled mugwort leaf essential oils, phytol is also one of the most dominant compositional compounds (6.3–16.3%). Eucalyptol is another most abundant compound ranged from 5–22% among all identified constituents. The essential oil from HB displays the highest larvicidal activity with over 52% of eucalyptol and phytol presence. Eucalyptol also shows larvicidal activity against larvae of *An. anthropophagous*, LC₅₀ values from 45–50 μg/mL³⁶. However, eucalyptol has displayed a relatively weaker larvicidal activity at LC₅₀ of 155 μg/mL, which is one third of what reported from above mentioned studies and ten times less than that of phytol (LC₅₀ = 16 μg/mL) in the present study. Interestingly, a relatively strong activity of caryophyllene has been demonstrated (LC₅₀ = 39 μg/mL). Similar findings of weak larvicidal activity from eucalyptol identified from oil of *Ar. gilyescens* has been reported against *An. sinensis* with LC₅₀ > 200 μg/mL³⁷. Although we have shown that β-caryophyllene is a much weaker larvicide than that of caryophyllene oxide against *An. sinensis* in the present study, Govindarajan et al.³⁸ found that β-caryophyllene from *Plectranthus barbatus* essential oil performed a strong larvicidal activity against *Aedes* and *Culex* mosquito larvae, and larvae of *An. subpictus*. Furthermore, Zhu and Tian³⁷ have reported a strong larvicidal activity against *An. anthropophagous*, with only 5% of caryophyllene oxide constituently from the essential oil of *Ar. gilyescens*. Obviously, synergistic effects may play additional roles to enhance the larvicidal activity from various essential oil constituent compounds.

The moxibustion therapy of using *Ar. argyi* has been considered as a form of complementary or alternative medicine existing in the holistic system of health care and healing in many parts of Southeast Asia³⁹. Burning of *Artemisia* dried leaves has been widely used to repel mosquitoes by minority people living in remote areas in southern provinces of China⁴⁰. Among the four most dominant constituent compounds, eucalyptol has been reported acting as a strong repellent against various species of adult mosquitoes⁴¹. In the present study, the essential oil extracted from Gansu province has demonstrated a high level of protection as found from DEET, which could be contributed by containing over 22% of eucalyptol among other compositional compounds. Eucalyptol, also named 1,8-cineole, has been reported as a strong mosquito repellent identified from many species of *Eucalyptus*, *Ocimum* and *Lippia*^{15,31}. Fumigant test with individual compound (eucalyptol) alone has further supported its strongest toxicity against adult *An. sinensis* mosquitoes. Phytol, a linear dipterene alcohol, has been reported with high repellent activity against *An. gambiae* and *Ae. aegypti*^{42,43}. β-Caryophyllene identified from essential oils of seven different provinces only provides low level of repellent activity and fumigant toxicity, which similar results have been reported from essential oils of *Nepeta cataria*, *Lippia camara* and *Lippia cheraliera*

Constituents	CAS ^a	RI-A ^b	RI-B ^c	Constituent percentage (%)						
				CQ	SC	YN	HB	HN	SD	GS
Santolina triene	2153-66-4	908	907	–	–	12.36	–	–	–	–
1R- α -Pinene	7785-70-8	929	932	2.58	1.38	2.49	–	–	1.47	–
α -Pinene	80-56-8	937	932	–	–	–	–	–	–	3.32
Camphene	79-92-5	952	947	–	1.24	–	–	1.19	1.02	2.13
Sabinen	3387-41-5	974	973	–	1.81	4.48	–	–	–	1.28
β -Pinene	127-91-3	979	977	1.62	2.02	–	–	1.08	–	2.26
(+)-2-Carene	149,946	991	1015	–	–	–	–	–	1.16	–
Yomogi alcohol	26,127-98-0	1000	1000	–	–	–	–	1.47	1.23	1.32
α -Phellandrene	99-83-2	1005	1003	–	–	1.91	–	–	–	–
o-Cymene	527-84-4	1022	1022	1.74	1.78	4.39	1.18	–	–	1.58
m-Cymene	535-77-3	1023	1022	–	–	–	–	1.68	–	–
p-Cymene	99-87-6	1025	1023	–	–	–	–	–	1.76	–
Eucalyptol	470-82-6	1032	1031	11.44	12.62	5.49	15.67	8.68	4.86	21.89
Artemisia ketone	546-49-6	1062	1061	–	–	–	–	–	2.02	2.76
cis-Sabinene hydroxide	17,699-16-0	1070	1068	–	–	–	–	1.19	4.33	3.93
Artemisia alcohol	27,644-04-8	1084	1084	2.26	–	–	1.34	4.81	2.82	3.55
Thujone	546-80-5	1103	1107	–	16.66	–	–	4.18	1.31	–
Isotujone	471-15-8	1114	1117	–	–	–	–	1.22	–	–
Chrysanthone	1125-12-8	1119	1108	–	–	–	4.97	–	–	–
Isochrysanthenone	473-06-3	1123	1125	4.08	1.8	–	–	–	–	1.84
(-)-Camphor	464-48-2	1142	1143	–	1.49	2.15	–	3.24	2.12	5.58
(+)-2-Bornanone	471-16-9	1143	1143	–	–	–	3.05	–	2.21	–
p-Menth-8-en-1-ol, cis	7299-41-4	1144	1066	–	1.35	–	–	–	–	–
Isoborneol	124-76-5	1157	1166	–	1.41	–	–	2.55	2.62	–
endo-Borneol	507-70-0	1167	1167	–	–	–	6.22	–	1.85	3.37
β -Artemisia acetate	3465-88-1	1173	1174	1.51	–	–	–	1.43	1.03	1.4
Terpinen-4-ol	562-74-3	1177	1178	–	–	–	–	1.74	–	–
L-4-terpineol;	20,126-76-5	1182	1181	–	1.25	–	1.6	–	–	1.29
α -Terpineol	98-55-5	1189	1192	–	–	–	–	–	–	1.98
S-Verbenone	1196-01-6	1204	1397	1.78	–	–	–	–	–	–
Piperitol isomer	16,721-39-4	1208	1210	–	–	–	–	–	1.29	–
cis-Mentha-1,8-dien-6-ol	1197-06-4	1229	1221	–	–	–	1.66	–	1.18	–
2,3-Pinane diol	53,404-49-2	1244	1317	–	–	1.03	–	–	–	–
Neryl acetate	141-12-8	1364	1292	–	–	2	–	–	–	–
Ylangene	14,912-44-8	1372	1375	–	–	1.83	–	–	–	–
Copaene	3856-25-5	1376	1376	1.11	–	–	–	–	–	–
(-)- β -Bourbonene	5208-59-3	1384	1384	1.06	–	–	–	–	–	–
1H-Cyclopropazulene	489-40-7	1409	1409	–	1.03	–	–	–	–	–
β-Caryophyllene	87-44-5	1419	1419	14.45	12.25	8.87	7.62	11.45	15.54	3.17
Humulene	6753-98-6	1454	1454	1.77	1.96	2.78	1.47	1.87	1.04	–
(E)- β -Farnesene	18,794-84-8	1457	1459	2.85	1.51	1.17	–	2.2	–	–
Germaacrene D	23,986-74-5	1481	1481	7.61	–	8.72	4.85	6.93	4.86	4.75
β -Selinene	17,066-67-0	1486	1486	2.54	–	2.75	1.15	–	1.06	1.43
(+)-Bicyclogermaacrene	24,703-35-3	1495	1497	–	1.97	1.51	–	1.14	–	–
Caryophyllene oxide	1139-30-6	1581	1584	13.24	10.64	8.56	12.51	14.82	9.45	9.63
Neointermedeol	5945-72-2	1660	1660	–	2.84	–	5.13	3	4.72	4.91
11,11-Dimethyl-4,8-dimethylenebicycloundecan-3-ol	79,580-01-1	1646	1640	–	–	–	1.18	–	1.01	–
Hexahydrofarnesyl acetone	502-69-2	1844	1847	1.25	–	–	1.64	1.49	1.2	1.45
n-Hexadecanoic acid	57-10-3	1968	1984	–	–	–	–	1	–	–
pentenylcurcumene	55,968-43-9	1980	2121	–	–	1.93	–	–	–	–
Phytol	150-86-7	2114	2115	8.29	7.02	6.3	16.28	9.95	8.27	8.02
Elemene isomer	–	1344	1497	1.18	–	–	–	–	–	–
Terpineol	8006-39-1	–	–	–	–	–	1.43	1.05	2.27	–
Total %				82.36	84.03	80.72	88.95	89.36	83.7	92.84

Table 5. GC–MS analyses of compositional compounds of the essential oils extracted from seven regions of *Ar. argyi*. ^aCAS: Chemical Abstracts Service Registry Number. ^bRetention index from NIST references (qualitative index of gas chromatography). ^cRetention index calculated with N alkanes (C₅–C₂₅Hc) as standard. The chemical constituents with less than 1% composition in each essential oil are not listed in the table. CQ, SC, YN, HB, HN, SD and GS: essential oils from Chongqing, Sichuan, Yunnan, Hubei, Henan, Shandong and Gansu province/municipality, respectively.

etc.^{44,45}. A detailed literature review by Mathew and Thoppil³² has shown that caryophyllene oxide is detected from many essential oils with probable cause for high mosquito larvicidal activity. For instance, caryophyllene oxide is a major constituent compound in essential oils of *Lippia gracilis* and *Hyptis pectinate* with the potent insecticidal activity against *Aedes aegypti* larvae⁴⁶. Strong larvicidal activity was also demonstrated from the essential oil of *Ar. argyi*.

The potential of using essential oils or their constituents as fumigants needs to be explored more thoroughly. In southeastern Asia remote areas and Latin American rural regions, the use of various fumigations for insect

Compounds	LC ₅₀ (µg/mL)	95% Confidence limits	Toxicity regression equation (y = a + bx)	χ ²
eucalyptol	155.55 ± 6.7	127.34–243.40	y = -2.33 + 0.01x	1.58*
β-caryophyllene	134.77 ± 8.9	119.06–171.96	y = -3.27 + 0.02x	1.11*
caryophyllene oxide	39.09 ± 5.2	31.28–45.55	y = -1.55 + 0.04x	1.73*
phytol	16.03 ± 2.6	-12.77–30.76	y = -0.32 + 0.02x	1.79*

Table 6. Larvicidal activity of the four individual compounds mostly found from the seven essential oils of *Ar. argyi* in China against the 4th instar larva of *An. sinensis*. *Significant at *p* < 0.05.

Compounds	LT ₅₀ (µL/L)	95% Confidence limits	Toxicity regression equation (y = a + bx)	χ ²
Eucalyptol	5.48 ± 3.4	3.50–6.65	y = -1.37 + 0.25x	0.05*
β-caryophyllene	32.60 ± 4.1	30.36–35.62	y = -2.58 + 0.07x	3.78*
Caryophyllene oxide	26.02 ± 5.3	24.60–29.16	y = -2.03 + 0.03x	3.26*
Phytol	17.98 ± 2.2	15.49–20.21	y = -1.13 + 0.06x	3.45*

Table 7. Fumigation toxicity of the four individual compounds mostly found from the seven essential oils of *Ar. argyi* in China against the female adults of *An. sinensis*. *Significant at *p* < 0.05 level.

repelling and disease vector control has been widely reported^{47,48}. Many monoterpenes (eucalyptol), diterpenes (phytol), sesquiterpenes (caryophyllene) as discovered in *Ar. argyi* essential oil have been reported possessing fumigant toxicity⁴⁹. All of these are supported by the strong fumigant toxicity and the shortest knockdown time found from *Ar. argyi* essential oils extracted from Gansu province. Three other species of *Artemisia* essential oils also have the fumigant activity against mosquitoes, including *Ar. scoparia*, *Ar. capillaries* and *Ar. carvifolia*⁵⁰.

The chemical composition and content of essential oils of plants are related to the diverse climate⁵¹. In addition, various elements in the soil also contribute to the differences in the chemical composition of *Ar. argyi* essential oil. For instance, the content of calcium, magnesium, manganese and nickel in Hubei *Ar. argyi* essential oil were higher, the elements of Nickel, cobalt, chromium and zinc in Sichuan *Ar. argyi* essential oil were relatively abundant, and the content of copper and cobalt in Henan *Ar. argyi* essential oil were also higher⁵². It was found that phosphorus content in soil affected plant growth, quality and chemical composition content⁵³. The content of *Angelica sinensis* volatile oil was 0.65% from Gansu, 0.59% from Yunnan and 0.29% from Sichuan, which were directly related to the longitude and latitude of different regions⁵⁴. In addition to the reasons above, the altitude and water quality may also be another key factor of the chemical composition and content of *Ar. argyi* essential oil⁵⁵, which need further discussion.

This research demonstrates the strong larvicidal activity, fumigant toxicity and repellent effect of *Ar. argyi* essential oils of seven different geographic origins in southern provinces of China against larvae and adult *An. sinensis* mosquitoes. Eucalyptol and phytol are the most abundant constituent compounds and most active compounds among others. Additional minor compounds detected from these oils may enhance insecticidal, repellent and fumigant effects as well, which further studies need to be carried out. Essential oils that constitute an important alternative to conventional insecticides are largely due to their selectivity (high toxicity for mosquitoes but not for other co-existing organisms) and their minimal environmental effects⁵⁶. Essential oils and their constituents, such as monoterpenes, have been widely used as fragrances in cosmetics, food additives, household products and medicine. Many of them are generally recognized as safe by the U.S. Food and Drug Administration (FDA). The outcomes from this study have provided new insights in use of phytochemicals derived from various botanical sources for more targeted mosquito control.

Materials and methods

Chemicals. Caryophyllene oxide (>90%) was obtained from Shanghai Yuanye Technology Co., Ltd. of China. N,N-Diethyl-3-methylbenzamide (DEET, 99%), eucalyptol (99%), β -caryophyllene (>80%) and phytol (90%) were all purchased from Macklin Biochemical Co., Ltd. (Shanghai, China). The alkanes analytic standards (C₅–C₂₅ Hc), reagent brand: o2si were purchased from ANPEL Laboratory Technologies (Shanghai, China), with over 99% purity.

Mosquitoes. *Anopheles sinensis* was originally collected from Jiangsu province in China, and reared in the Institute of Insect and Molecular Biology, Chongqing Normal University, for more than 40 generations. Mosquitoes were reared at a 12:12 light/dark photoperiod, 65 ± 5% relative humidity under 27 ± 1 °C, which larvae were fed with fish food and adults were provided with 10% glucose solution. The 4th instar of larvae and the four-day old adult mosquitoes were used for larvicidal, fumigant and repellent activity assays in this study. All experimental methods of this study are carried out in accordance with relevant guidelines and regulations, and The Ethical Committee of Chongqing Normal University approved the human-bait assays performed in the study.

Plant material collections and essential oil extraction from *Artemisia argyi*. The plant *Ar. argyi* is widely distributed in field in China. All experimental materials of *Ar. argyi* were collected on public lands in field at each location of seven provinces in China from September to October in 2018, and the plant material collecting did not require permission or other relevant certificates. The wet weights of collected aerial branches and leaves were 70 kg from 25 m² of wasteland in Chongqing, 65 kg from 100 m² of wasteland in Sichuan, 46 kg from 200 m² of mountain area in Yunnan, 40 kg from 100 m² of agricultural farm in Hebei, 78 kg from 35 m² of agricultural field in Henan, 50 kg from 150 m² of mountain area in Shangdong, and 55 kg from 150 m² of mountain area in Gansu, respectively. These seven provinces could represent the main distribution of the *Ar. argyi* plant in China in term of geography and climate. The collected *Ar. argyi* was dried in the collecting locations, and then sent to laboratory at Chongqing Normal University. The samples of *Ar. argyi* plants from each location were identified by the plant taxonomist Prof. Hai He at the Chongqing Normal University where we work, and the specimens are stored in the Herbarium of Chongqing Normal University with following information as attached label: collecting time, collecting location, collector's name, specimen number, species name, and identifier's name. The dried *Ar. argyi* was cut to pieces at a size of about 10 cm in diameter by guillotine, and the essential oil of *Ar. argyi* was extracted using 6 kg of dried material per location. The extraction lasted for six hours using a steam distillation apparatus, and oil–water mixture was separated by a separatory funnel with an addition of NaCl as described in Joshi et al.⁵⁷. The upper layer from the separated essential oils were taken, and then dried with added anhydrous sodium sulfate⁵⁸. The purified oils were stored in brown reagent bottles at 4 °C prior to bioassays and chemical analysis.

Larvicidal activity assay. The larvicidal activity of *Ar. argyi* essential oils and their four dominant compounds against *An. sinensis* larvae was tested according to the method introduced by Nyamoita⁵⁹. The range of lethal doses for the investigated samples was determined using the doses at 20, 40, 60, 80, 100 and 120 µg/mL, respectively, and a total of three replicates were conducted. Ethanol was used as the control. Different dosages of samples were added to a 50-mL cup containing a total of 20 mL of dechlorinated water. Twenty 4th instar larvae were then gently moved to the cup and the bioassay was performed under room temperature at 27 ± 1 °C with an 80 ± 10% relative humidity. The number of larvae with no movement (after touching) were considered as dead and were counted after 24 h of exposure.

Fumigant toxicity assay. The fumigant activity of *Ar. argyi* essential oils and their four dominant compounds against *An. sinensis* adults was investigated using methods described in Jiang et al.⁶⁰, with some changes. Twenty female *An. sinensis* adults at the age of 4-day old were released into a 250 mL of Erlenmeyer flask sealed using a rubber stopper with a predrilled hole (diameter = 0.4 cm) for a 15 min of acclimatization. A piece of filter paper (1 × 3 cm) was then placed into the flask. The filter papers were applied testing materials at doses of 2, 4, 6, 8, 10, 12 µL of essential oils. After one hour of exposures, mosquitoes were then transferred to cages for 24-h provided with cotton balls containing 10% glucose. The number of dead mosquitoes was counted. For the fumigant knockdown test, the dose of testing material was 2 µL⁶¹, the knockdown number were recorded every five minutes until reaching the first hour. The time of mosquitoes observed falling down at the bottom were considered as the knocked down time, no any movement was recorded as the lethal time. A total of three replicates were performed in each trial. The experiments were carried out at a room temperature of 26 ± 1 °C. Probit analysis was used to calculate LC₅₀ and KT₅₀ values⁶².

Repellent activity assay. The repellent assay was carried out using a human-bait technique reported by Kaliyaperumal et al.⁶³ with a few modifications. Before the experiment, both hands and forearms of the three volunteers were asked to be washed with scent free soaps, then dried. Gloves wore to cover the entire hand and forearm except for an area of 25 cm² (5 × 5 cm) on the dorsal sides were cut by a pair of scissors. One hundred 24-h starved 3-day-old female *An. sinensis* were released into a screen cage (300 mm × 300 mm × 300 mm) prior to the experiment. About 37.5 µL of testing extracted *Ar. argyi* oils were evenly applied on top of the exposed skin area, then inserted into the testing mosquito cage. 10% DEET repellent formulation was used as positive control. The number of mosquitoes landing on exposed area of skin was counted every fifteen minutes up to the 65th min. Ethanol used as the negative control by applying the same amount onto another hand. Both hands were exposed to starved mosquitoes starting at the same time. Each extracted mugwort oil and 4 individual

compounds was repeated three times in random orders. The time intervals of every new volunteers were about 2 h. The percentage of repellency was calculated by using the formula⁶⁴:

$$\text{Percentage of repellency} = [(N_c - N_t)/N_c \times 100]$$

where N_c and N_t is the number of mosquito landings on hand of negative control and test samples, respectively. The testing protocol was approved by the Committee of Laboratory Animal Experimentation at Chongqing Normal University (Approval No. Zhao-20200416-01), and all volunteers were provided with the official written informed consent form for testing.

Chemical analysis of the *Ar. argyi* essential oils. The chemical constituents of *Ar. argyi* essential oils were analyzed using a gas chromatography-mass spectrometer (Thermo Scientific TSQ XLS GC-MS with a model TRACE 1310 GC), which was equipped with an HP-5MS capillary column (30 m in length, 0.25 mm internal diameter, and 0.25 μm stationary phase film thicknesses). We followed the method as described by Yuan et al.⁶⁵. The GC oven temperature was programmed at 60 °C for 1 min, then increased to 128 °C at a rate of 2.5 °C/min and maintained for 2 min, and at 5 °C/min to 250 °C (held for 6 min). High purity helium gas was used as mobile phase with a constant flow of 1 mL/min. The parameters for mass spectrometer were set as: 280 °C for both GC injector and MS transfer line, 300 °C for ion source. Mass spectra were recorded in the electron impact ionization (EI) at 70 eV. The MS scan range was from 50–550 m/z. The total ion chromatogram peak areas were used to calculate the percentages of the compositional compounds in essential oils. Chemical identification of each compositional compounds was based on their retention indices determined by reference to a homologous series of n-alkanes ($\text{C}_5\text{--C}_{25}\text{Hc}$), and by comparisons of their mass spectral fragmentation patterns with those reported in the literature⁶⁶, and their referenced mass spectra from the TSQ mass spectral library.

Statistical analyses. Log-probit analysis (SPSS 24) was used to determine the median lethal concentration (LC_{50}), the median knockdown concentration (KT_{50}), regression equation, chi-square value and the 95% confidence limit. The numbers of mosquito landings and biting, on negative control, positive control and chemical-treated hands, were compared using an unpaired T-test. Further repellency data among treatments were analyzed using two-way ANOVA followed by Tukey's honest significant difference (HSD) test ($p < 0.05$).

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References

- Brown, M. & Hebert, A. A. Insect repellents: An overview. *J. Am. Acad. Dermatol.* **36**, 243–249. [https://doi.org/10.1016/S0190-9622\(97\)70289-5](https://doi.org/10.1016/S0190-9622(97)70289-5) (1997).
- Benelli, G. et al. The recent outbreaks of Zika virus: Mosquito control faces a further challenge. *Asian Pacific Journal of Tropical Disease* **6**, 253–258. [https://doi.org/10.1016/S2222-1808\(15\)61025-8](https://doi.org/10.1016/S2222-1808(15)61025-8) (2016).
- Tjaden, N. B., Caminade, C., Beierkuhnlein, C. & Thomas, S. M. Mosquito-borne diseases: Advances in modelling climate-change impacts. *Trends Parasitol.* **34**, 227–245. <https://doi.org/10.1016/j.pt.2017.11.006> (2018).
- Manguin, S., Bangs, M., Pothikasikorn, J. & Chareonviriyaphap, T. Review on global co-transmission of human Plasmodium species and Wuchereria bancrofti by Anopheles mosquitoes. *Infect. Genet. Evol.* **10**, 159–177. <https://doi.org/10.1016/j.meegid.2009.11.014> (2009).
- Bango, Z. A., Tawe, L., Muthoga, C. W. & Paganotti, G. M. Past and current biological factors affecting malaria in the low transmission setting of Botswana: A review. *Infect. Genet. Evol.* **85**, 44–58. <https://doi.org/10.1016/j.meegid.2020.104458> (2020).
- Pan, X. H., Wang, J. M., Wu, T. C. & Sun, Y. F. Compound extraction and component analysis on volatile oil of *Artemisia Argyi*. *Adv. Mater. Res.* **465**, 255–261. <https://doi.org/10.4028/www.scientific.net/AMR.465.255> (2012).
- Kamel, N. H., Bahgat, I. M. & Kady, G. A. E. Digestive enzyme as benchmark for insecticide resistance development in *Culex pipiens* larvae to chemical and bacteriologic insecticides. *J. Egypt Soc. Parasitol.* **43**, 245–258 (2013).
- Zhu, Q. et al. Synthesis, insecticidal activity, resistance, photodegradation and toxicity of pyrethroids (A review). *Chemosphere* **254**, 67–79. <https://doi.org/10.1016/j.chemosphere.2020.126779> (2020).
- Arias-Estévez, M. et al. The mobility and degradation of pesticides in soils and the pollution of groundwater resources. *Agric. Ecosyst. Environ.* **123**, 247–260. <https://doi.org/10.1016/j.agee.2007.07.011> (2008).
- Macoris, M., Andrighetti, M., Wanderley, D. & Ribolla, P. Impact of insecticide resistance on the field control of *Aedes aegypti* in the State of São Paulo. *Rev. Soc. Bras. Med. Trop.* **47**, 573–578. <https://doi.org/10.1590/0037-8682-0141-2014> (2014).
- Haddi, K. et al. Detection of a new pyrethroid resistance mutation (V410L) in the sodium channel of *Aedes aegypti*: A potential challenge for mosquito control. *Sci. Rep.* **7**, 46549. <https://doi.org/10.1038/srep46549> (2017).
- Goodyer, L. & Behrens, R. H. Short report: The safety and toxicity of insect repellents. *Am. J. Trop. Med. Hyg.* **59**, 323–324 (1998).
- McGready, R., Hamilton, K. A., Simpson, J. A., Cho, T. & Luxemburger, C. Safety of the insect repellent N,N-diethyl-M-toluamide (DEET) in pregnancy. *Am. J. Trop. Med. Hyg.* **65**, 285–289 (2001).
- Gideon, K., Doreen, M. & Benoit, B. DEET-based insect repellents: safety implications for children and pregnant and lactating women. *Can. Med. Assoc. J.* **169**, 209–212 (2003).
- Nerio, L. S., Olivero-Verbel, J. & Stashenko, E. Repellent activity of essential oils: A review. *Biores. Technol.* **101**, 372–378. <https://doi.org/10.1016/j.biortech.2009.07.048> (2010).
- Ali, A. et al. Chemical composition and biological activity of four salvia essential oils and individual compounds against two species of mosquitoes. *J. Agric. Food Chem.* **63**, 447–456. <https://doi.org/10.1021/jf504976f> (2015).
- Asadollahi, A. A.-O., Khoobdel, M. A.-O. X., Zahraei-Ramazani, A. A.-O., Azarmi, S. A.-O. X. & Mosawi, S. H. Effectiveness of plant-based repellents against different Anopheles species: A systematic review. *Malar J* **18**, 436 (2019).
- Tisgratog, R., Sanguanpong, U., Grieco, J. P., Ngoen-Kluan, R. & Chareonviriyaphap, T. Plants traditionally used as mosquito repellents and the implication for their use in vector control. *Acta Trop.* **157**, 136–144 (2016).
- Satyral, P. et al. Chemical composition, Aedes mosquito larvicidal activity, and repellent activity against *Triatoma rubrofasciata* of Severinia monophylla leaf essential oil. *Parasitol. Res.* **118**, 733–742 (2019).
- Wu, H., Zhang, M. & Yang, Z. Repellent activity screening of 12 essential oils against *Aedes albopictus* Skuse: Repellent liquid preparation of *Mentha arvensis* and *Litsea cubeba* oils and bioassay on hand skin. *Ind. Crops Prod.* **128**, 464–470. <https://doi.org/10.1016/j.indcrop.2018.11.015> (2019).

21. Bianco, E. *et al.* Larvicidal activity of seaweeds from northeastern Brazil and of a halogenated sesquiterpene against the dengue mosquito (*Aedes aegypti*). *Ind. Crops Prod.* **43**, 270–275. <https://doi.org/10.1016/j.indcrop.2012.07.032> (2013).
22. Cheng, S. S. *et al.* Larvicidal activities of wood and leaf essential oils and ethanolic extracts from *Cunninghamia konishii* Hayata against the dengue mosquitoes. *Ind. Crops Prod.* **47**, 310–315. <https://doi.org/10.1016/j.indcrop.2013.03.016> (2013).
23. Ayinde, A. A., Morakinyo, O. M. & Sridhar, M. K. C. Repellency and larvicidal activities of *Azadirachta indica* seed oil on *Anopheles gambiae* in Nigeria. *Heliyon* **6**, e03920. <https://doi.org/10.1016/j.heliyon.2020.e03920> (2020).
24. Stappen, I. *et al.* Chemical composition and biological effects of *Artemisia maritima* and *Artemisia nilagirica* essential oils from wild plants of Western Himalaya. *Planta Med.* **80**, 1079–1087. <https://doi.org/10.1055/s-0034-1382957> (2014).
25. Xiang, F. *et al.* Antimicrobial activities and mechanism of the essential oil from *Artemisia argyi* Levl. et Van. var. *argyi* cv. Qiai. *Ind. Crops Prod.* **125**, 582–587 (2018).
26. Taleghani, A., Emami, S. A. & Tayarani-Najaran, Z. Artemisia: A promising plant for the treatment of cancer. *Bioorg. Med. Chem.* **28**, 115180. <https://doi.org/10.1016/j.bmc.2019.115180> (2020).
27. Sun, J. *A study on Chinese Mugwort Cultural Heritage in China* (Nanjing Agricultural University, 2016).
28. Zhang, W.-J., You, C.-X., Yang, K., Chen, R. & Wang, Y. Bioactivity of essential oil of *Artemisia argyi* Lévl. et Van. and its main compounds against *Lasioderma serricornis*. *J. Oleo Sci.* **63**, 829–837 (2014).
29. Ozek, G. *et al.* Chemical diversity and biological activity of the volatiles of five *Artemisia* species from far East Russia. *Rec. Nat. Prod.* **8**, 242–261. <https://doi.org/10.1039/c4np00064a> (2014).
30. Wang, J.-L. *Effect of Artemisia argyi essential oil on vitellogenesis of Aedes albopictus* (Northeast Normal University, 2009).
31. Maia, M. & Moore, S. Plant-based insect repellents: A review of their efficacy, development and testing. *Malar. J.* **10**(Suppl 1), S11. <https://doi.org/10.1186/1475-2875-10-S1-S11> (2011).
32. Mathew, J. & Thoppil, J. E. Chemical composition and mosquito larvicidal activities of *Salvia* essential oils. *Pharm. Biol.* **49**, 456–463. <https://doi.org/10.3109/13880209.2010.523427> (2011).
33. Sundararajan, B., Moola, A. K., Vivek, K. & Kumari, B. D. R. Formulation of nanoemulsion from leaves essential oil of *Ocimum basilicum* L. and its antibacterial, antioxidant and larvicidal activities (*Culex quinquefasciatus*). *Microb. Pathogen.* **125**, 475–485. <https://doi.org/10.1016/j.micpath.2018.10.017> (2018).
34. Karthi, S. *et al.* Larvicidal enzyme inhibition and repellent activity of red Mangrove *Rhizophora mucronata* (Lam.) leaf extracts and their biomolecules against three medically challenging arthropod vectors. *Molecules* **25**, 3844. <https://doi.org/10.3390/molecules25173844> (2020).
35. Dey, P. *et al.* Evaluation of larvicidal activity of *Piper longum* leaf against the dengue vector, *Aedes aegypti*, malarial vector, *Anopheles stephensi* and filariasis vector, *Culex quinquefasciatus*. *S. Afr. J. Bot.* **132**, 482–490. <https://doi.org/10.1016/j.sajb.2020.06.016> (2020).
36. Qi, H., Wang, W. & Liang, Z. Larvicidal activity of *Zanthoxylum acanthopodium* essential oil against the malaria mosquitoes, *Anopheles anthropophagus* and *Anopheles sinensis*. *Malar. J.* **17**, 194. <https://doi.org/10.1186/s12936-018-2341-2> (2018).
37. Zhu, L. & Tian, Y.-J. Chemical composition and larvicidal activity of essential oil of *Artemisia gilvescens* against *Anopheles anthropophagus*. *Parasitol. Res.* **112**, 1137–1142. <https://doi.org/10.1007/s00436-012-3243-9> (2013).
38. Govindarajan, M., Rajeswary, M., Arivoli, S., Tennyson, S. & Benelli, G. Larvicidal and repellent potential of *Zingiber nimmonii* (J. Graham) Dalzell (Zingiberaceae) essential oil: An eco-friendly tool against malaria, dengue, and lymphatic filariasis mosquito vectors?. *Parasitol. Res.* **115**, 1–10. <https://doi.org/10.1007/s00436-016-4920-x> (2016).
39. Hsu, Y.-C., Chao, H.-R. & Shih, S.-I. Human exposure to airborne aldehydes in Chinese medicine clinics during moxibustion therapy and its impact on risks to health. *Environ. Lett.* **50**, 260–271. <https://doi.org/10.1080/10934529.2015.981112> (2015).
40. Gou, Y. *et al.* Ethnobotanical survey and evaluation of traditional mosquito repellent plants of Dai people in Xishuangbanna, Yunnan Province, China. *J. Ethnopharmacol.* **262**, 113–124. <https://doi.org/10.1016/j.jep.2020.113124> (2020).
41. Klocke, J. A., Darlington, M. V. & Balandrin, M. F. 1,8-Cineole (Eucalyptol), a mosquito feeding and ovipositional repellent from volatile oil of *Hemizonia fitchii* (Asteraceae). *J. Chem. Ecol.* **13**, 2131–2141. <https://doi.org/10.1007/BF01012562> (1987).
42. Odalo, J. O. *Biochemical Examination and Identification of Botanicals Active against Adult Anopheles Gambiae from the Coastal Region of Kenya* (Kenyatta University, 2014).
43. Cantrell, C. L., Jones, A. M. P. & Ali, A. Isolation and identification of mosquito (*Aedes aegypti*) biting-deterrent compounds from the Native American ethnobotanical remedy plant *Hierochloë odorata* (Sweetgrass). *J. Agric. Food Chem.* **64**, 8352–8358. <https://doi.org/10.1021/acs.jafc.6b01668> (2016).
44. Seyoum, A., Kabiru, E. W., Lwande, W., Killeen, G. F. & Hassanali, A. Repellency of live potted plants against *Anopheles gambiae* from human baits in semi-field experimental huts. *Am. J. Trop. Med. Hyg.* **67**, 191–195. <https://doi.org/10.4269/ajtmh.2002.67.191> (2002).
45. Aklilu, S., Gerry, F. K., Ephantus, W. K., Bart, G. J. K. & Ahmed, H. Field efficacy of thermally expelled or live potted repellent plants against African malaria vectors in western Kenya. *Trop. Med. Int. Health* **8**, 1005–1011. <https://doi.org/10.1046/j.1360-2276.2003.01125.x> (2010).
46. Silva, W. J. *et al.* Effects of essential oils on *Aedes aegypti* larvae: Alternatives to environmentally safe insecticides. *Biores. Technol.* **99**, 3251–3255. <https://doi.org/10.1016/j.biortech.2007.05.064> (2008).
47. Zeledon, R. & Rabinovich, J. E. Chagas' disease: An ecological appraisal with special emphasis on its insect vectors. *Annu. Rev. Entomol.* **26**, 101–133. <https://doi.org/10.1146/annurev.en.26.010181.000533> (1981).
48. Zerba, E. N. Susceptibility and resistance to insecticides of Chagas disease vectors. *Medicina* **59**(Suppl), 41–46 (1999).
49. Sfara, V., Zerba, E. N. & Alzogaray, R. A. Fumigant insecticidal activity and repellent effect of five essential oils and seven monoterpenes on first-instar nymphs of *Rhodnius prolixus*. *J. Med. Entomol.* **46**, 511–515. <https://doi.org/10.1603/033.046.0315> (2009).
50. Chen, W., Wu, H., Ma, Z., Feng, J. & Zhang, X. Evaluation of fumigation activity of thirty-six essential oils against *Culex pipiens pallens* (Diptera: Culicidae). *Acta Entomol. Sin.* **61**, 86–93 (2018).
51. Kumar, A., Jnanesha, A. C. & Lal, R. K. Coppicing impact on the essential oil yield and its chemical composition of lemongrass cultivars of the genus *Cymbopogon* under the semi-arid region of South India. *Acta Ecol. Sin.* <https://doi.org/10.1016/j.chnaes.2021.05.005> (2021).
52. Zhu, D.-X. & Ni, S.-B. The research of authentic medicinal herbs Biogeochemical characteristics. *Micronutrient* **21**, 44–47 (2004).
53. Peçanha, D. A., Freitas, M. S. M., Vieira, M. E., Cunha, J. M. & de Jesus, A. C. Phosphorus fertilization affects growth, essential oil yield and quality of true lavender in Brazil. *Ind. Crops Prod.* **170**, 113803. <https://doi.org/10.1016/j.indcrop.2021.113803> (2021).
54. Zhang, J., Xu, J. & Jing, Y.-J. 浅析不同地理环境中中药材生长及质量的影响. *China J. Trodit. Chin. Med. Pharm.* **007**, 30–33 (2009).
55. Mumivand, H., Ebrahimi, A., Morshedloo, M. R. & Shayganfar, A. Water deficit stress changes in drug yield, antioxidant enzymes activity and essential oil quality and quantity of Tarragon (*Artemisia dracuncululus* L.). *Ind. Crops Prod.* **164**, 113381. <https://doi.org/10.1016/j.indcrop.2021.113381> (2021).
56. Isman, M. B. Plant essential oils for pest and disease management. *Crop Prot.* **19**, 603–608. [https://doi.org/10.1016/S0261-2194\(00\)00079-X](https://doi.org/10.1016/S0261-2194(00)00079-X) (2000).
57. Joshi, S., Mishra, D., Bisht, G. & Khetwal, K. In vitro antimicrobial activity of essential oil of *Swertia cordata* aerial parts. *Open Nat. Prod. J.* **3**, 66–70. <https://doi.org/10.2174/2210315511303010012> (2013).

58. Benelli, G., Pavela, R., Drenaggi, E., Desneux, N. & Maggi, F. Phytol, (E)-nerolidol and spathulenol from *Stevia rebaudiana* leaf essential oil as effective and eco-friendly botanical insecticides against *Metopolophium dirhodum*. *Ind. Crops Prod.* **155**, 112844. <https://doi.org/10.1016/j.indcrop.2020.112844> (2020).
59. Nyamoita, M. G. Report of the WHO informal consultation on the evaluation and testing of insecticides (1996).
60. Jiang, Z. L., Chen, A. L., Bai, W., Lin, J. & Zhang, X. Fumigating and contact activity of 6 kinds of essential oils on *Musca domestica* L. *Chin. J. Pesticide Sci.* **4**, 85–88. <https://doi.org/10.1007/s11769-002-0038-4> (2002).
61. Ma, W. B., Feng, J. T., Jiang, Z. L. & Zhang, X. Fumigant activity of 6 selected essential oil compounds and combined effect of methyl salicylate and trans-cinnamaldehyde against *Culex pipiens pallens*. *J. Am. Mosq. Control Assoc.* **30**, 199–203. <https://doi.org/10.2987/14-6412R.1> (2014).
62. Finney, D. J. Book Reviews: Probit analysis: A statistical treatment of the sigmoid response curve. *Ann. Entomol. Soc. Am.* **67**, 549–553. <https://doi.org/10.1038/ja.2014.36> (1952).
63. Kaliyaperumal, K., Askual, G. & Samuel Fekadu, H. Mosquito repellent activity of essential oil of Ethiopian ethnomedicinal plant against Afro-tropical malarial vector *Anopheles arabiensis*. *J. King Saud Univ. Sci.* **26**, 305–310. <https://doi.org/10.1016/j.jksus.2014.01.001> (2014).
64. Azeem, M. *et al.* Chemical composition and repellent activity of native plants essential oils against dengue mosquito, *Aedes aegypti*. *Ind. Crops Prod.* **140**, 111609. <https://doi.org/10.1016/j.indcrop.2019.111609> (2019).
65. Yuan, H. B. *et al.* Effects of essential oil from *Artemisia lavandulaefolia* DC. on Fumigation activity and several enzymes activities of *Monolepta hieroglyphica* (Motschulsky) Adults. *J. Jilin Agric. Univ.* (2014).
66. Adams, R. P. Identification of essential oil components by gas chromatography/mass spectrometry. *J. Am. Soc. Mass Spectrom.* **18**, 803–806. <https://doi.org/10.1016/j.jasms.2007.01.001> (2007).

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Author contributions

The study conception and design were put forward by B.C., D.L., and J.J.Z. Material preparation, experiment and data collection were performed by D.L., Z.Y., and L.C. The data analysis and manuscript drafting were carried out by D.L., B.C. and J.J.Z. All authors read and approved the final manuscript.

Competing interests

The authors declare no competing interests.

Additional information

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