

ORIGINAL ARTICLE

Polyphenols isolated from *Broussonetia kazinoki* prevent cytokine-induced β -cell damage and the development of type 1 diabetes

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The axis of nuclear factor κ B (NF- κ B)-inducible NO synthase (iNOS)-nitric oxide plays a key role in cytokine- and streptozotocin-mediated pancreatic β -cell damage. In this study, we investigated the effects of kazinol C and isokazinol D isolated from *Broussonetia kazinoki* on the β -cell viability and function. RINm5F cells and primary islets were used for *in vitro* and *ex vivo* cytokine toxicity experiments, respectively. For type 1 diabetes induction, mice were injected with multiple low-dose streptozotocin (MLDS). Cytokine-induced toxicity was completely abolished in both RINm5F cells and islets that were pretreated with either kazinol C or isokazinol D. Both kazinols inhibited the NF- κ B signaling pathway, thereby inhibiting cytokine-mediated iNOS induction, nitric oxide production, apoptotic cell death and defects in insulin secretion. Moreover, the occurrence of diabetes in MLDS-treated mice was efficiently attenuated in kazinol-pretreated mice. Immunohistochemical analysis revealed that the numbers of terminal deoxynucleotidyl transferase dUTP nick end labeling-positive apoptotic cells and nuclear p65-positive cells were significantly decreased in kazinol-pretreated mice. Our results suggest that kazinol C and isokazinol D block the NF- κ B pathway, thus reducing the extent of β -cell damage. Therefore, kazinol C and isokazinol D may have therapeutic value in delaying pancreatic β -cell damage in type 1 diabetes.

Experimental & Molecular Medicine (2015) 47, e160; doi:10.1038/emm.2015.16; published online 24 April 2015

INTRODUCTION

Type 1 diabetes is a T cell-mediated disease that ultimately destroys the capacity of the body to produce and secrete insulin.¹ Immune cell infiltration and the subsequent apoptotic destruction of pancreatic β -cells, also known as insulinitis, are major hallmarks of this disease.² In the early stages of disease, infiltrated inflammatory cells release cytokine such as interleukin-1 β (IL-1 β), tumor necrosis factor- α and interferon- γ (IFN- γ). IL-1 β , either alone or in combination with tumor necrosis factor- α or IFN- γ , activates nuclear factor κ B (NF- κ B) pathway, which in turn upregulates inducible nitric oxide synthase (iNOS) expression, leading to the production of nitric oxide (NO) in pancreatic islets.³

Administration of multiple low-dose streptozotocin (MLDS) to mice destroys pancreatic β -cells through a mechanism similar to that of cytokine-mediated damage. Moreover,

MLDS-induced β -cell dysfunction and destruction has been shown to be mediated by NO.⁴ Interestingly, mice harboring a mutation in either NF- κ B or iNOS are resistant to MLDS-induced diabetes.^{5,6}

Plant-derived polyphenols such as quercetin,⁷ genistein,⁸ resveratrol,⁹ sulfuretin¹⁰ and sulforaphane¹¹ have been shown to protect islets against cytokine and/or streptozotocin. Plants of genus *Broussonetia* share polyphenols as major constituents, which were reported to exert anti-diabetic activity through the inhibition of protein tyrosine phosphatase 1B¹² and α -glucosidase.¹³ We recently isolated a series of alkylated (dimethylallylated or prenylated) kazinols, which belong to the 1,3-diphenylpropane family, from the medicinal plant *Broussonetia kazinoki*.¹⁴ Kazinol U, which has one prenyl group, protected β -cells against cytokine-mediated damage by suppressing the NF- κ B pathway.¹⁵ In this study, we tested the

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Received 11 November 2014; revised 4 January 2015; accepted 12 January 2015

anti-diabetic efficacies of kazinol C and isokazinol D, each of which has three alkyl groups in the phenyl moiety. Specifically, we exposed RINm5F insulinoma cells and isolated islets to cytokine, and then evaluated the protective effects of kazinol C and isokazinol D on signaling pathways that lead to apoptosis and β -cell dysfunction. We further investigated the abilities of intraperitoneally-administered kazinol C and isokazinol D to suppress type 1 diabetes and preserve functional β -cell mass in MLDS-treated mice.

MATERIALS AND METHODS

Cell culture and reagents

The RINm5F rat pancreatic β -cell line was purchased from the American Type Culture Collection (ATCC, Manassas, VA, USA). Cells were maintained at 37 °C in a humidified 5% CO₂ atmosphere and cultured in RPMI 1640 medium (Gibco, Grand Island, NY, USA) supplemented with 10% fetal bovine serum, 2 mM glutamine, 100 units ml⁻¹ penicillin, 100 μ g ml⁻¹ streptomycin and 2.5 μ g ml⁻¹ amphotericin B. IL-1 β and IFN- γ were obtained from R&D Systems (Minneapolis, MN, USA). All other reagents were from Sigma-Aldrich (St Louis, MO, USA), unless otherwise noted.

Preparation of kazinol C and isokazinol D

Air-dried *B. kazinoki* root bark (0.6 kg) was extracted for 24 h at room temperature in 2 l of ethanol. The resultant extract (51 g) was dissolved in water and successively partitioned with *n*-hexane, ethyl acetate, chloroform and butanol. The ethyl acetate fraction (31 g) was subjected to silica gel column chromatography and eluted with an *n*-hexane:acetone gradient system (20:1 \rightarrow 1:10), thereby yielding 11 fractions. Fraction 5 was further separated by silica gel chromatography with an *n*-hexane:ethyl acetate gradient system (20:1 \rightarrow 1:3) to yield isokazinol D (70 mg). Fraction 7 was also subjected to silica gel column chromatography and eluted with a chloroform:methanol gradient system (100:1 \rightarrow 11:1). The resultant eluate was further purified on an reverse phase C18 (RP-C18) column (GL Sciences Inc., Tokyo, Japan) via a gradient elution of MeOH (30% \rightarrow 100%), thus yielding kazinol C (260 mg).

High-performance liquid chromatography analyses of kazinol C and isokazinol D

The purities of the kazinol C and isokazinol D were evaluated by high-performance liquid chromatography using the Waters 1525 system (Milford, MA, USA). A reverse-phase column (ODS-2, 150 \times 4.6 mm inner diameter, GL Sciences Inc.) was eluted with 80% methanol, and the resultant eluates were monitored with an ultraviolet detector at 280 nm. Purities were calculated based on the relative peak areas on the high-performance liquid chromatography.

Chemical structures of kazinol C and isokazinol D

The molecular formula of the two kazinols was deduced as C₃₀H₄₀O₄ by using high-resolution mass spectral data. The two kazinols also showed a typical ¹H-NMR (proton nuclear magnetic resonance) pattern of prenylated 1,3-diphenylpropane derivatives. The NMR data indicated that kazinol C has two 3,3-dimethylallyl and one 1,1-dimethylallyl groups. By using the heteronuclear multiple bond correlation (HMBC) correlations between the ipso carbons and protons of the respective allyl groups, the locations of the allyl groups were determined to be C-5' and C-6' on ring A and C-5' on ring B. Isokazinol D was also analyzed to have one 1,2-dimethylallyl group at

C-5' on ring A and one 3,3-dimethylallyl group at C-6' on ring B. The formation of the 2,2-dimethyldihydropyran ring was confirmed by the presence of two methylene protons (δ 1.7–1.9 and 2.5–2.7), two methyl protons (δ 1.33) and one oxygenated carbon (C-9" at δ 74.0).

MTT assay for cell viability

The viability of RINm5F cells was determined by analyzing the reduction of 3-(4,5-dimethylthiazol-2-yl)-2,5-diphenyltetrazolium bromide (MTT) to formazan.¹⁶

Measurement of nitric oxide

Biologically-produced NO is rapidly oxidized to nitrite and nitrate in aqueous solutions. Thus, the production of NO was assessed by measuring the nitrite concentrations of cell culture supernatants via a colorimetric assay.¹⁶

Electrophoretic mobility shift assay

Nuclear extracts prepared from either RINm5F cells or islets were treated with a proteinase inhibitor cocktail (Calbiochem, San Diego, CA, USA) to inhibit endogenous protease activity. A synthetic oligonucleotide containing the κ -chain binding site (5'-CCGG TTAACAGAGGGGGCTTTCCGAG-3') was used as a probe in a gel retardation assay. The two complementary strands were then annealed and labeled with α -³²PdCTP. Labeled oligonucleotides (10 000 counts per minute), 10 μ g nuclear extracts and binding buffer (10 mM Tris-HCl (pH 7.6), 500 mM KCl, 10 mM ethylenediaminetetraacetic acid, 50% glycerol, 100 ng poly(dI-dC) and 1 mM dithiothreitol) were then incubated for 30 min at room temperature in a final volume of 20 μ l. Next, the reaction mixtures were analyzed by electrophoresis on 4% polyacrylamide gels in 0.5 \times Tris-borate buffer. After drying the gels, radioactive bands were visualized by autoradiography. The specificity of the DNA–protein interaction for NF- κ B was confirmed via competition assays using a 50-fold excess of unlabeled oligonucleotide.

Western blot analysis

RINm5F cells (5 \times 10⁶) or 30 islets were homogenized in 100- μ l ice-cold lysis buffer (20 mM 4-(2-hydroxyethyl)-1-piperazineethanesulfonic acid (HEPES, pH 7.2), 1% Triton X-100, 10% glycerol, 1 mM phenylmethanesulfonylfluoride, 10 μ g ml⁻¹ leupeptin and 10 μ g ml⁻¹ aprotinin). The resultant protein extracts were quantitated, and proteins (20 μ g) were resolved by sodium dodecyl sulfate–polyacrylamide gel electrophoresis and then transferred to nitrocellulose membranes. Membranes were probed with primary antibodies (1 μ g ml⁻¹) against p50, p65, inhibitor of κ B α (I κ B α), iNOS, proliferating cell nuclear antigen and β -actin (all from Santa Cruz Biotechnology, Dallas, TX, USA), and cleaved caspase-3, Bcl-2 and Bax (Cell Signaling Technology, Beverly, MA, USA). Horseradish peroxidase-conjugated IgG (Zymed, South San Francisco, CA, USA) was used as a secondary antibody.

RNA isolation and real-time RT-PCR

RNA was isolated from RINm5F cells or islets using Trizol (Invitrogen, Carlsbad, CA, USA), and then precipitated with isopropanol and dissolved in diethylpyrocarbonate-treated distilled water. Total RNA (2 μ g) was treated with RNase-free DNase (Invitrogen), and first-strand cDNA was generated using a random hexamer primer with a first-strand cDNA synthesis kit (Applied Biosystems, Foster City, CA, USA). Specific primers for iNOS were designed using primer express software (Applied Biosystems). Primers used were: iNOS (accession

no. NM_012611): 5'-TGTGCTAATGCGGAAGGTCAT-3' (forward) and 5'-CGACTTTCCTGTCTCAGTAGCAAA-3' (reverse). Control 18S ribosomal RNA was purchased from Applied Biosystems and used as an internal control. Real-time RT-PCR reactions consisted of 10 ng reverse-transcribed total RNA, 167 nM forward and reverse primers, and 2 \times PCR master mix in a final volume of 10 μ l. Reactions were carried out in 384-well plates using an ABI Prism 7900HT sequence detection system (Applied Biosystems).

Islet isolation and determination of islet viability

Pancreatic islets were isolated from male C57BL/6 mice by using the collagenase digestion method. Islet viability was evaluated as previously described.¹⁷ Briefly, islets were incubated with cytokine and fixed overnight in a solution of 6.5% glutaraldehyde. Islet viability was determined by hematoxylin and eosin (H&E) staining and subsequent labeling with anti-insulin antibodies (Santa Cruz Biotechnology). Apoptosis was detected using an ApoPercentage apoptosis assay kit (Bicolor, Newtownabbey, Ireland). Apoptotic islets, which are stained red after the selective uptake of ApoPercentage dye (Bicolor), were detected using an inverted light microscope (Axiovert 40 CFL microscope, Carl Zeiss, Oberkochen, Germany).¹⁸

Glucose-stimulated insulin secretion assay

Islets were cultured for 24 h with cytokine and then washed three times in Krebs-Ringer bicarbonate buffer (25 mM HEPES (pH 7.4), 115 mM NaCl, 24 mM NaHCO₃, 5 mM KCl, 1 mM MgCl₂, 2.5 mM CaCl₂ and 0.1% bovine serum albumin) containing 3 mM D-glucose. Insulin secretion assays were performed with either 2.8 or 16.7 mM D-glucose.

Induction of diabetes by injection of MLDS

Pathogen-free male C57BL/6 mice, each weighing 25–30 g, were purchased from Orient Bio (Seoul, Korea). Mice were housed in a laminar flow cabinet with a 12-h light/dark cycle, and maintained on standard laboratory chow *ad libitum*. To induce diabetes, mice were injected via the tail vein with streptozotocin (Sigma-Aldrich) dissolved in 0.1 M sodium citrate buffer (pH 4.0), at a dose of 40 mg kg⁻¹ per day, for five consecutive days. Mice were divided into the following groups: (1) nontreated control, (2) MLDS, (3) kazinol C+MLDS and (4) isokazinol D+MLDS. Control animals received citrate buffer alone. Groups 3 and 4 received daily intraperitoneal injections of either kazinol C or isokazinol D for 3 days before the administration of streptozotocin. The day on which streptozotocin was first administered was defined as day 1. At day 5, mice were killed by decapitation without anesthesia, and trunk blood was collected in prechilled tubes containing 1 mg ml⁻¹ ethylenediaminetetraacetic acid. Plasma glucose levels were assayed using the glucose oxidase method (Sigma-Aldrich), whereas plasma insulin levels were measured using a rat/mouse insulin enzyme-linked immunosorbent assay kit (Linco Research, St Charles, MO, USA). All animal experiments were performed in accordance with the Guide for the Care and Use of Laboratory Animals, published by the US National Institutes of Health (NIH Publication No. 85-23, revised 2011). The current study protocol was also approved by the Institutional Animal Care and Use Committee of Chonbuk National University (Permit No. CBU 2014-00061).

Histological assessment

Mice were killed under nonfasting conditions. Pancreatic specimens were immediately fixed in 10% formalin solution, embedded in paraffin and cut into 5 μ m sections. Specimens were stained with

H&E to identify morphological changes. For immunohistochemical analysis, tissue sections were treated with a microwave antigen retrieval procedure using 10 mM sodium citrate buffer. After blocking endogenous peroxidase activity, the sections were incubated with DAKO protein block serum-free (DAKO, Carpinteria, CA, USA) to block nonspecific staining. Sections were then incubated with anti-insulin antibodies (Santa Cruz Biotechnology) or anti-p65 antibodies (Abcam Biotech, Cambridge, MA, USA). Peroxidase activity was detected with 3-amino-9-ethylcarbazole.

Terminal deoxynucleotidyl transferase dUTP nick end labeling (TUNEL) staining was performed using a commercial kit (R&D Systems). Five to six random sections per slide were investigated to determine the percentage of apoptotic islets.

Statistical analysis

Statistical analysis was performed using analysis of variance and Duncan's tests. Differences with a *P* value <0.05 were considered statistically significant.

RESULTS

Purification and structural analyses of kazinol C and isokazinol D from *B. kazinoki*

The purities of *B. kazinoki*-derived preparations of kazinol C and isokazinol D were assessed by high-performance liquid chromatography (Figure 1a). The clean peaks at retention times of 15.2 and 16.25 min in the high-performance liquid chromatography indicate pure preparations of kazinol C and isokazinol D, respectively. The purities were determined to be

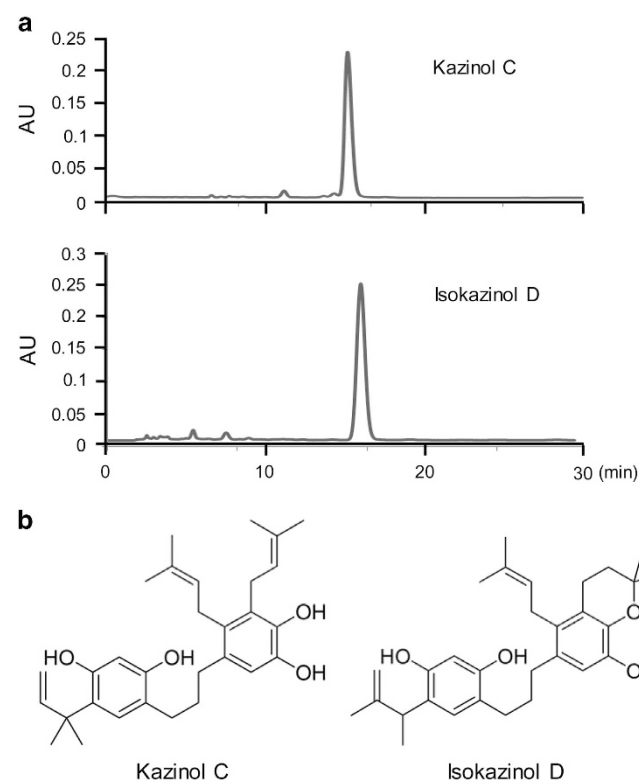


Figure 1 High-performance liquid chromatography (a) and chemical structures (b) of kazinol C and isokazinol D.

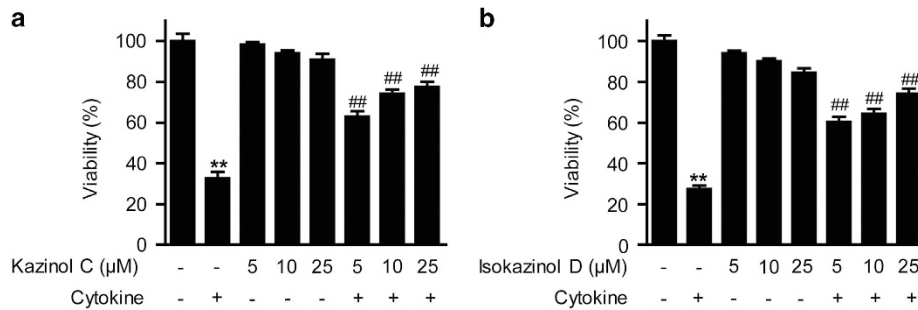


Figure 2 Prevention of cytokine-induced cell death in RINm5F cells. Cells were pretreated with the indicated concentrations of kazinol C (a) or isokazinol D (b) for 3 h, and then IL-1 β (1 U ml $^{-1}$) and IFN- γ (100 U ml $^{-1}$) were added and cells were incubated for an additional 48 h. Cell viability was determined using the MTT assay. Each value is the mean \pm s.e.m. of three independent experiments ($n=9$). ** $P<0.01$ versus vehicle-treated control; ## $P<0.01$ versus cytokine-treated cells.

> 95% (kazinol C) and > 93% (isokazinol D). Kazinol C and isokazinol D were elucidated by spectroscopic analysis to be 1,3-diphenyl propane derivatives, structures which are common among phenolic compounds isolated from *B. kazinoki* (Figure 1b).

Effects of kazinol C and isokazinol D on cytokine-induced cell death and NO production in RINm5F cells

To determine whether kazinol C and isokazinol D exert protective effects on β -cells, we first investigated whether kazinol C and isokazinol D protect RINm5F cells against cytokine toxicity. Treatment with two cytokines, IL-1 β and IFN- γ , significantly reduced cell viability to 67–72% that of control cells; however, pretreatment with either kazinol C or isokazinol D increased the viability of cytokine-treated RINm5F cells in a concentration-dependent manner (Figures 2a and b). Treatment with either kazinol C or isokazinol D alone had no effect on cell viability at the concentrations used in these experiments. In addition, treatment with either kazinol C or isokazinol D after cytokine exposure did not increase cell viability (data not shown).

Next, the production of NO was evaluated. At 24 h, control and cytokine-treated RINm5F cells generated 2.37 ± 0.49 and 15.5 ± 1.56 μ M nitrite, respectively (Figure 3a). The production of NO was almost completely inhibited by pretreatment with either 25 μ M kazinol C or isokazinol D.

We next investigated the effects of kazinol C and isokazinol D on cytokine-mediated expression of iNOS mRNA and protein by real-time RT-PCR and western blotting, respectively. Cytokine treatment increased the levels of iNOS mRNA and protein (Figure 3a). However, pretreatment with either kazinol C or isokazinol D before cytokine treatment resulted in concentration-dependent decreases in mRNA and protein levels.

Effects of kazinol C and isokazinol D on cytokine-mediated activation of NF- κ B in RINm5F cells

NF- κ B has been implicated in the transcriptional regulation of cytokine-induced iNOS expression.³ Therefore, we examined the effects of kazinol C and isokazinol D on the cytokine-stimulated translocation of NF- κ B from the cytosol to the nucleus in RINm5F cells. Nuclear extracts were prepared 1 h

after cytokine treatment and used to analyze NF- κ B-DNA-binding activity and cytoplasmic I κ B α degradation. The binding of the p65 subunit to an NF- κ B consensus sequence was increased in cytokine-treated RINm5F cells; moreover, the nuclear levels of the p65 and p50 subunits were increased (Figure 3b). In contrast, pretreatment with either kazinol C or isokazinol D markedly suppressed the nuclear translocation and DNA-binding ability of NF- κ B, suggesting that kazinol C and isokazinol D inhibit iNOS expression by suppressing NF- κ B-mediated transcriptional activation. In addition, cytokine-treated RINm5F cells showed decreased levels of cytosolic I κ B α due to I κ B α degradation; this decrease was markedly suppressed by pretreatment with either kazinol C or isokazinol D (Figure 3b).

Effects of kazinol C and isokazinol D on cytokine-induced apoptosis and glucose-stimulated insulin secretion in mouse islets

We further assayed the protective effects of kazinol C and isokazinol D using pancreatic islets isolated from mice. Islets were exposed to IL-1 β and IFN- γ for 48 h, fixed, and then either stained with H&E or immunostained with anti-insulin antibodies. Similar to the results observed in RINm5F cells, cytokine treatment resulted in islet damage and weak insulin immunoreactivity (Figure 4a). However, pretreatment with either 25 μ M kazinol C or isokazinol D prevented these cytokine-induced changes. Pretreated islets with kazinols exhibited a similar morphology to control islets, as assessed by both H&E and immunohistochemical staining. Both control islets and kazinol-pretreated islets had well-defined margins and a round shape, in addition to exhibiting strong insulin immunoreactivity in their β -cells (Figure 4a).

We next evaluated the extent of apoptosis in the islets using an ApoPercentage apoptosis assay kit (Biocolor; Figures 4a and b). Almost four times more apoptosis was observed in the cytokine-treated islets compared with the vehicle-treated islets ($83.3 \pm 4.8\%$ versus $20.6 \pm 3.6\%$, $P<0.01$). However, significantly less apoptosis was observed in the 25- μ M kazinol C-pretreated islets ($30.0 \pm 4.8\%$, $P<0.01$) and in the 25- μ M isokazinol D-pretreated islets ($24.2 \pm 3.4\%$, $P<0.01$) than in the cytokine-treated islets. Pretreatment with either 10 μ M kazinol C or isokazinol D also partially blocked cytokine-

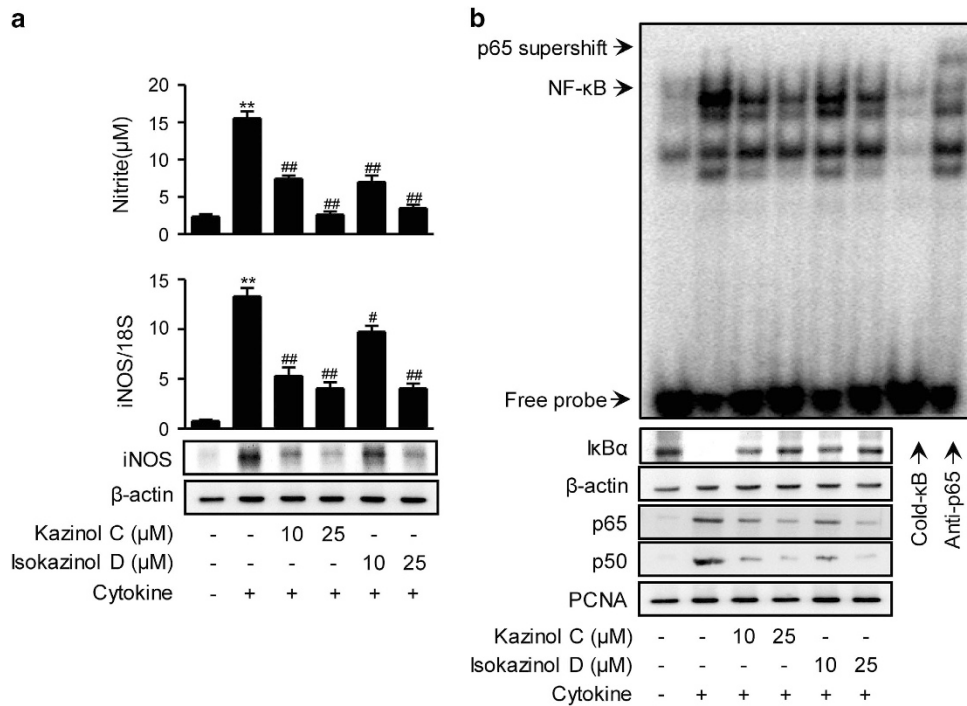


Figure 3 Prevention of cytokine-induced NF- κ B activation in RINm5F cells. (a) RINm5F cells were pretreated with the indicated concentrations of kazinol C or isokazinol D for 3 h, and followed by IL-1 β and IFN- γ treatment. Following 24 h of incubation, the concentration of nitrite and the expression of iNOS mRNA and protein were determined. Each value is the mean \pm s.e.m. of three independent experiments ($n=9$). ** $P<0.01$ versus vehicle-treated control; ## $P<0.01$ versus cytokine-treated cells. (b) After 30 min, NF- κ B DNA binding was analyzed by electrophoretic mobility shift assay, and the nuclear translocation of the p65 and p50 subunits and the level of cytoplasmic I κ B α were determined by western blotting. β -actin and proliferating cell nuclear antigen (PCNA) were used as loading controls for cytosolic and nuclear proteins, respectively.

induced apoptosis. Consistently, increased levels of the proapoptotic proteins Bax and cleaved caspase-3, in addition to the decreased level of the anti-apoptotic protein Bcl-2, were observed in cytokine-treated islets. These effects were reversed in kazinol C- or isokazinol D-pretreated islets (Figure 4c).

To investigate the functional effects of kazinol C and isokazinol D, we pretreated islets with these compounds and evaluated their effects on cytokine-induced impairment of insulin secretion. Basal- and glucose-stimulated insulin secretion levels were measured after exposure to cytokine for 24 h. As shown in Figure 4d, control islets secreted 18.7 ± 1.4 ng mg $^{-1}$ protein per hour of insulin after glucose stimulation, whereas cytokine-treated islets secreted 6.2 ± 2.1 ng mg $^{-1}$ protein per hour under the same conditions ($P<0.01$). However, following pretreatment with 25 μ M of either kazinol C or isokazinol D, cytokine-treated islets secreted insulin upon glucose stimulation at a rate similar to that of control islets. Basal insulin release among the groups was similar.

Incubation of the islets with cytokine for 24 h resulted in a fivefold increase in NO production (Figure 5a). Real-time RT-PCR and western blotting revealed that iNOS mRNA and protein levels, respectively, were markedly increased by cytokine treatment (Figure 5a). Consistent with the results obtained in RINm5F cells, pretreatment with 25 μ M of either kazinol C or isokazinol D reduced the cytokine-mediated upregulation of NO production, and also reduced the level of iNOS expression

to similar levels of control islets. In addition, cytokine treatment increased NF- κ B DNA binding, nuclear translocation of NF- κ B subunits and cytoplasmic I κ B α degradation; however, pretreatment with either kazinol C or isokazinol D markedly reduced these effects in islets (Figure 5b).

Effects of kazinol C and isokazinol D on MLDS-mediated induction of diabetes

Our *in vitro* and *ex vivo* results indicated that kazinol C and isokazinol D protect β -cells from cytokine-mediated damage, thus increasing islet viability. To examine whether this β -cell protection prevents the development of diabetes, mice were given kazinols daily with intraperitoneal injections of either kazinol C or isokazinol D for 3 days before the administration of streptozotocin. Administration of MLDS resulted in hyperglycemia, with a mean blood glucose level of 401 ± 34 mg dl $^{-1}$ on day 5 (Figure 6a). In contrast, mice that had been pretreated with kazinol C (10 mg kg $^{-1}$) or isokazinol D (10 mg kg $^{-1}$) were fully resistant to the development of diabetes. Treatment with kazinol C or isokazinol D alone did not affect blood glucose concentration (data not shown). In addition, on day 5, the plasma insulin level was decreased by 40% in the MLDS-treated mice compared with the control mice (from 0.53 ± 0.13 to 0.13 ± 0.02 ng ml $^{-1}$). However, the severity of hypoinsulinemia was attenuated in mice that had been pretreated with either kazinol C or isokazinol D. These results indicate that

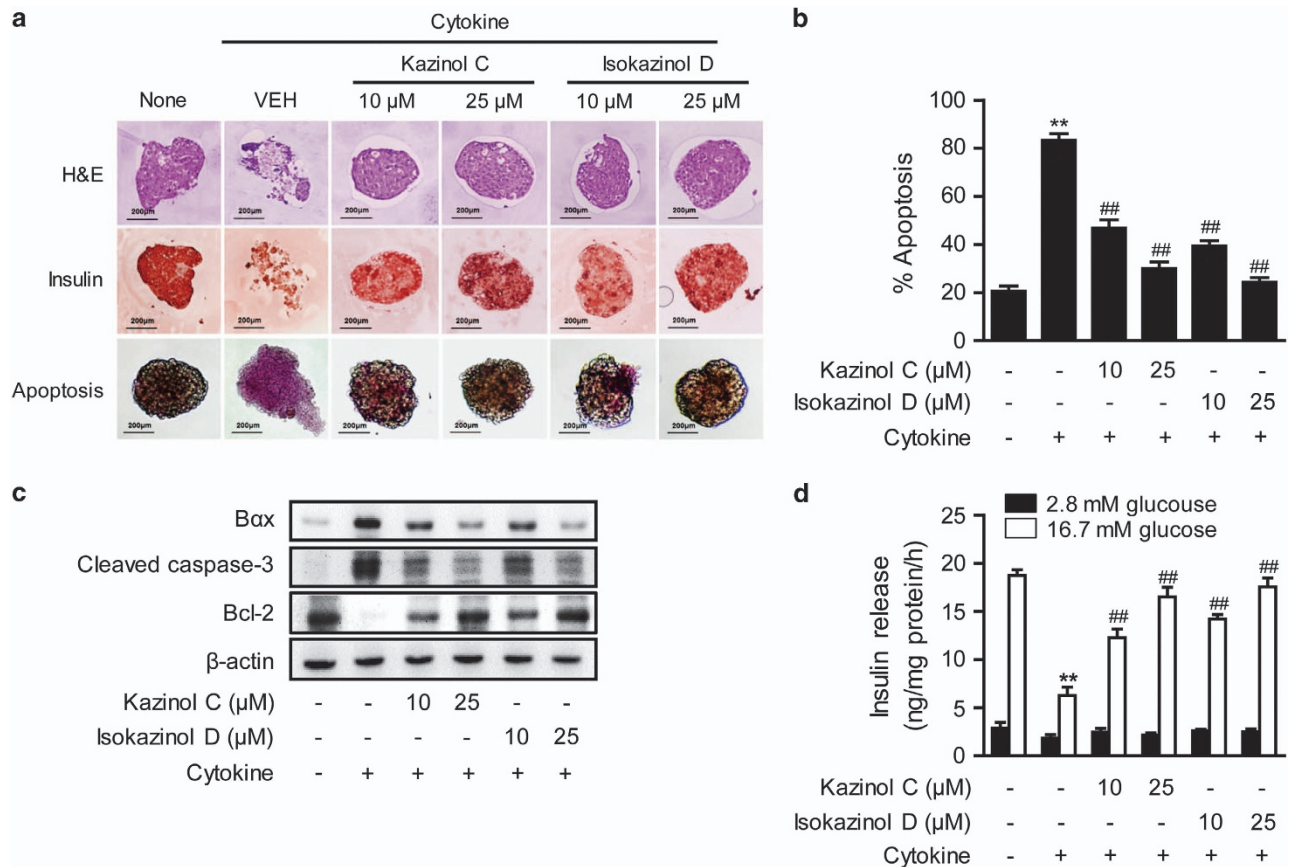


Figure 4 Prevention of cytokine-induced apoptosis in isolated islets. (a) Mice islets were treated with IL-1 β and IFN- γ , either with or without a 3-h pretreatment with the indicated concentrations of kazinol C or isokazinol D. Following a 48-h incubation, islets were either stained with H&E or labeled with anti-insulin antibodies, and then examined by microscopy. Apoptosis was evaluated using the APOPercentage apoptosis assay kit. Apoptotic islets appear bright pink. (b) The numbers of apoptotic islets were expressed as percentages of the total numbers of islets. (c) Islet extracts were prepared 24 h after cytokine treatment, and the levels of apoptosis-related proteins were examined by western blotting. (d) Glucose-stimulated insulin secretion was quantified by enzyme-linked immunosorbent assay and normalized to the total protein content. The results of triplicate samples are expressed as means \pm s.e.m. ($n=9$). ** $P<0.01$ versus vehicle-treated control; ## $P<0.01$ versus cytokine-treated islets.

both kazinol C and isokazinol D exert protective effects against MLDS-induced diabetes. Histological examination of the islets from MLDS-treated mice revealed degenerative and destructive changes, as well as weak insulin immunoreactivity (Figure 6b). In contrast, islets from mice that had been pretreated with either kazinol C or isokazinol D exhibited a round, nearly normal shape with a well-defined margin, in addition to strong insulin immunoreactivity (Figure 6b).

Since both kazinol C and isokazinol D protected islets against cytokine-mediated apoptotic injury and preserved their ability to secrete insulin, we next examined whether kazinol C and isokazinol D protected pancreatic islets against apoptosis. Fewer apoptotic islets were observed in kazinol C- and isokazinol D-pretreated mice compared with nontreated mice, as assessed by TUNEL staining (Figure 6b). The mean number of TUNEL-positive cells in MLDS-treated mice was almost seven times higher than that of control mice ($8.4 \pm 2.7\%$ versus $58.0 \pm 8.8\%$, $P<0.01$). The numbers of TUNEL-positive islets were also significantly reduced in kazinol C-pretreated ($17.6 \pm$

4.7% , $P<0.01$) and isokazinol D-pretreated ($19.8 \pm 7.9\%$, $P<0.01$) mice compared with MLDS-treated mice (Figure 6c).

To elucidate the anti-diabetogenic mechanism(s) of kazinol C and isokazinol D, we examined their effects on streptozotocin-induced NF- κ B activation by immunohistochemical staining. Administration of MLDS to mice resulted in the nuclear translocation of the p65 NF- κ B subunit (Figure 6d). In contrast, increased nuclear translocation was not observed in pancreatic sections from kazinol C- or isokazinol D-pretreated mice. Taken together, these results indicate that both kazinol C and isokazinol D inhibit NF- κ B activation and prevent the development of type 1 diabetes in mice.

DISCUSSION

Multiple lines of evidence suggest that NF- κ B-dependent NO production is a critical factor in the destruction and dysfunction of β -cells.⁴⁻⁶ In β -cells, NF- κ B activation can be triggered by cytokine or diabetogenic drugs such as streptozotocin and alloxan.^{3,6,19} Therefore, reducing NF- κ B activity is one of the

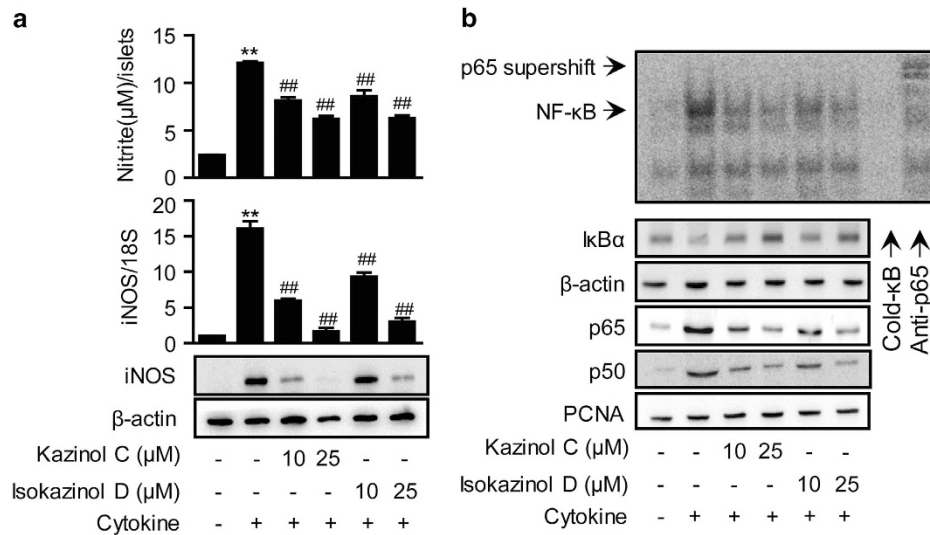


Figure 5 Prevention of cytokine-induced NF- κ B activation in isolated islets. Mouse islets were treated with IL-1 β (1 U ml⁻¹) and IFN- γ (100 U ml⁻¹), either with or without a 3-h pretreatment with the indicated concentration of kazinol C or isokazinol D. (a) The concentration of nitrite and the expression of iNOS mRNA and protein were determined after 24 h. (b) The extents of NF- κ B DNA binding, I κ B α degradation and nuclear translocation of p65 and p50 were determined 1 h later. The results of triplicate samples are expressed as means \pm s.e.m. ($n=9$). ** $P<0.01$ versus vehicle-treated control; ## $P<0.01$ versus cytokine-treated islets.

strategies for overcoming β -cell failure. In traditional Chinese medicine, *B. kazinoki* has been used as a suppressant for edema; moreover, several polyphenols have been reported to show cytotoxic, anti-oxidative, anti-inflammatory and tyrosinase-inhibitory activities.^{20,21} With the aim of identifying novel NF- κ B inhibitors, we previously isolated a series of kazinols from *B. kazinoki*, and evaluated their abilities to inhibit NO production in lipopolysaccharide-stimulated RAW264.7 macrophages (ATCC).²² Among the kazinols tested, kazinol C and isokazinol D exhibited the most promising NF- κ B suppressive activity and were thus selected for further studies of their anti-diabetic efficacy. Here, we showed that kazinol C and isokazinol D are indeed able to protect pancreatic β -cells from cytokine-induced apoptosis and also to preserve the ability of islets to secrete insulin in response to glucose stimulation. Consistent with these *in vitro* and *ex vivo* effects on β -cell survival and function, *in vivo* studies also demonstrated the protective effects of kazinol C and isokazinol D against the development of MLDS-induced diabetes.

We observed that the optimal concentrations of kazinol C and isokazinol D for protecting β -cells from cytokine-induced cell death was 25 μ M. This concentration is far lower than that of kazinol U, of which 60 μ M is required to suppress NF- κ B activation and protect β -cells against cytokine-mediated damage.¹⁵ Kazinol C and isokazinol D both share the same 1,3-diphenylpropane backbone, but they have different substitution patterns in their phenyl rings. Kazinol C has three dimethylallyl groups, while isokazinol D has two dimethylallyl groups and one dimethyldihydropyran ring system, which was formed by the ring closure to a dimethylallyl group. In contrast, kazinol U has only one dimethylallyl group. The number of hydrophobic allyl groups may affect cell permeability, which might explain the weaker protective activity of

kazinol U against cytokine-mediated damage in β -cells. One previous study demonstrated that the stem bark powder of *B. kazinoki* exhibited anti-hyperglycemic activity in an animal model of diabetes. However, only the crude powder of this plant was used in that study, and the protective mechanism was not investigated.²³

We first investigated whether kazinol C and isokazinol D suppress NF- κ B activity in pancreatic β -cells, and found that both kazinols suppressed the cytokine-mediated increases of iNOS mRNA and protein levels in both RINm5F cells and islets. Consistent with these observations, cytokine-stimulated NO production was significantly reduced by pretreatment with kazinol C or isokazinol D. Moreover, western blotting and electrophoretic mobility shift assay data showed that both kazinols suppressed NF- κ B activity. Since NF- κ B is known to regulate iNOS expression and, consequently, NO production,³ these results indicate that kazinol C and isokazinol D both efficiently suppress NF- κ B activity in pancreatic β -cells.

As a key regulator of inflammatory and cell survival responses, NF- κ B participates in both the generation of proinflammatory mediators and the anti-apoptotic pathway. The balance between these two pathways determines the fate of β -cells that are exposed to cytokine. These contradictory roles of NF- κ B help explain several conflicting reports that NF- κ B inactivation either protects against cytokine toxicity^{24,25} or aggravates such injury,^{26,27} although the underlying reasons for these discrepant results are not entirely clear. Part of the explanation might be that NF- κ B is differentially activated under different conditions, such as in rodent versus human islets,^{28,29} immortalized β -cells versus primary β -cells,²⁵ different cytokine mixtures and varying incubation times,^{25,27,30} and different animal models of diabetes induction.^{24,27} In our experimental conditions, kazinol C and isokazinol D decreased

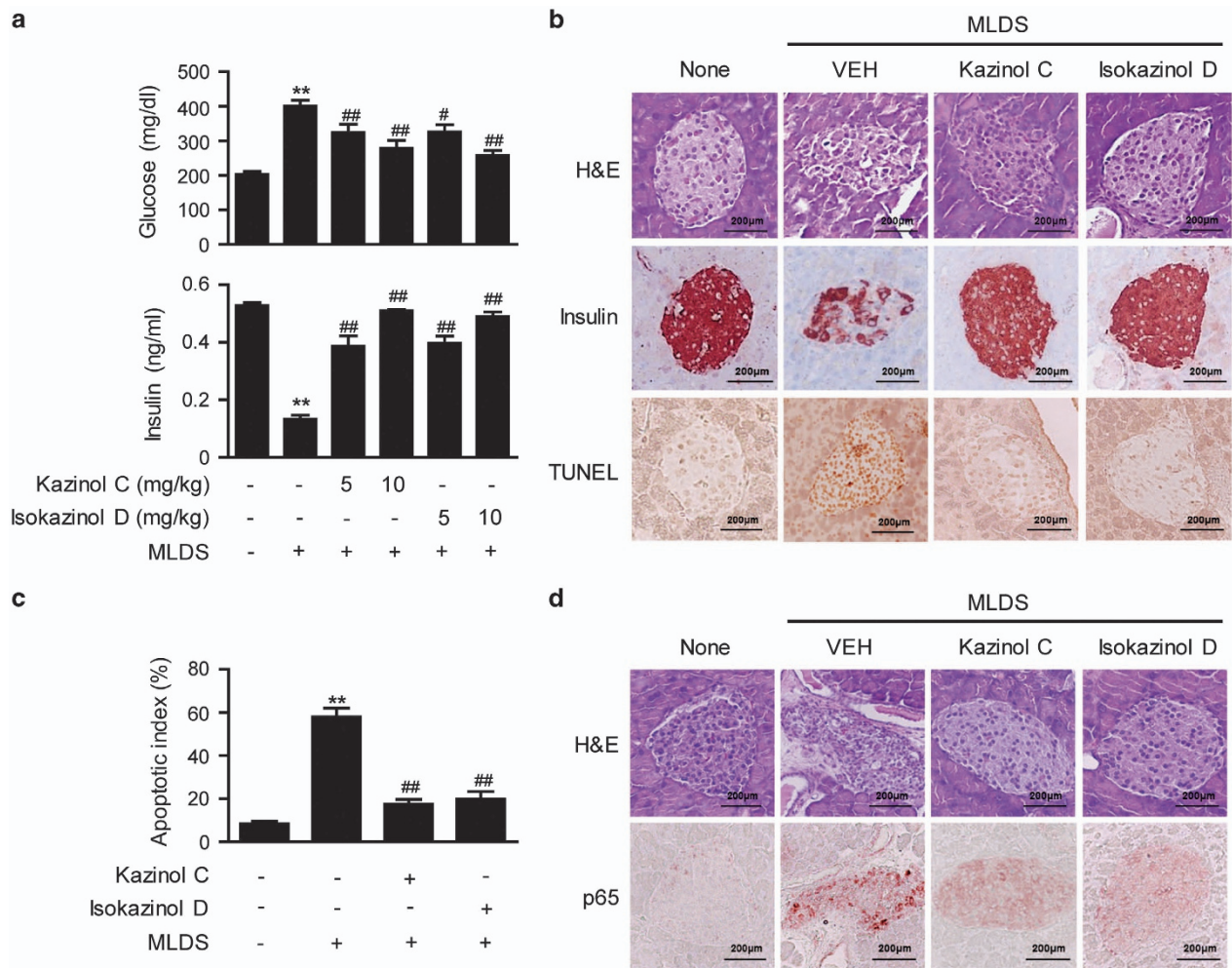


Figure 6 Protection of islets from MLDS-induced destruction. Type 1 diabetes was induced as described in the Materials and Methods section. **(a)** Levels of glucose and insulin were determined. Results are expressed as means \pm s.e.m. ($n=9$). ** $P<0.01$ versus untreated control; # $P<0.05$ and ## $P<0.01$ versus MLDS mice. **(b)** Pancreases were obtained from normal control mice and from mice injected with MLDS, kazinol C and MLDS, or isokazinol D and MLDS. The cellular morphologies of islets and the adjoining exocrine regions were examined after counterstaining with H&E. Islets were either immunolabeled with anti-insulin antibodies or TUNEL stained, and then examined by microscopy. **(c)** The numbers of apoptotic islets were determined by counting and expressed as percentages of the total numbers of islets. **(d)** Nuclear translocation of the NF- κ B p65 subunit was confirmed by immunohistochemical staining. Similar results were obtained from three independent experiments.

the levels of the proapoptotic proteins Bax and cleaved caspase-3, whereas the level of the anti-apoptotic protein Bcl-2 was increased in cytokine-treated islets. In addition, the number of apoptotic islets in the *ex vivo* diabetic condition was significantly reduced by kazinol C or isokazinol D treatment, suggesting that NF- κ B suppression by kazinol C and isokazinol D shifted the balance toward suppression of the apoptotic pathway.

This evidence for the anti-apoptotic and anti-diabetic activities of kazinol C and isokazinol D was strengthened by the finding that intraperitoneal administration of either kazinol C or isokazinol D resulted in a remarkable resistance to the development of diabetes in MLDS-treated mice. The importance of the NF- κ B-iNOS-NO cascade in the development of type 1 diabetes has previously been reported in a mouse MLDS model.^{9,31} In streptozotocin-treated mice, increased expression

of iNOS and the concomitant overproduction of NO in β -cells are observed at a very early stage. In this situation, NO may react with superoxide anion to generate a strong oxidizer peroxynitrite, and thus leading to aggressive oxidative and nitrosative stress.³² However, a recent study by Broniowska *et al.*³³ showed that pancreatic β -cells do not generate peroxynitrite in response to cytokine, suggesting that NO is not the likely mediator of the toxic effects of cytokine. In the present study, we found that MLDS treatment induced insulin deficiency and hyperglycemia in mice, which is a typical phenotype of type 1 diabetes. Importantly, immunohistochemical staining showed that kazinol C and isokazinol D both prevented the MLDS-mediated nuclear translocation of NF- κ B subunits, suggesting that the *in vivo* anti-diabetic potentials of kazinol C and isokazinol D are related to the suppression of NF- κ B. Consistent with these data, treatment with kazinol C or

isokazinol D markedly reduced the numbers of TUNEL-positive apoptotic islet cells. These results are consistent with our *ex vivo* results and demonstrate the importance of NF- κ B suppression in preserving pancreatic β -cell viability.

It could be argued that the protection failure of kazinol against the post cytokine injury limits their therapeutic potential. Although type 1 diabetes is usually diagnosed in its final stage, the disease course in humans is a longer process. Therefore, kazinol can be used to delay the disease progression. Kazinol can also be used in transplantation to enhance survival of donor islets and preserve functional islet mass. This hypothesis is supported by the observation that conditional and specific inhibition of NF- κ B improves transplantation outcomes.³⁴ In summary, our results indicate that kazinol C and isokazinol D both inhibit cytokine- and streptozotocin-induced β -cell damage *in vitro*, *ex vivo* and in an *in vivo* model of diabetes. The primary mechanism underlying these effects is the inhibition of iNOS protein expression, which may be mediated at the transcriptional level through the inhibition of NF- κ B activation.

CONFLICT OF INTEREST

The authors declare no conflict of interest.

ACKNOWLEDGEMENTS

This work was supported by the Bio & Medical Technology Development Program (No. NRF-2012M3A9B2027975) and the Medical Research Center Program (No. 2008-0062279 and 2011-0030074) through the National Research Foundation, which is funded by the Korean government (MSIP).

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