RESEARCH PAPER



Phylotype resolved spatial variation and association patterns of planktonic *Thaumarchaeota* in eastern Chinese marginal seas

Jiwen Liu^{1,2,3} · Fuyan Huang¹ · Jiao Liu¹ · Xiaoyue Liu¹ · Ruiyun Lin¹ · Xiaosong Zhong⁴ · Brian Austin⁵ · Xiao-Hua Zhang^{1,2,3}

Received: 25 July 2022 / Accepted: 30 January 2023 / Published online: 7 April 2023 © The Author(s) 2023

Abstract

The majority of marine ammonia oxidizers belong to *Thaumarchaeota*, a phylum of Archaea, which is distributed throughout the water column. Marine surface waters contain distinct thaumarchaeotal phylotypes compared to the deeper ocean, but spatial dynamics of the surface-associated lineages are largely unsolved. This study of 120 seawater samples from the eastern Chinese marginal seas identified contrasting distribution and association patterns among thaumarchaeotal phylotypes across different dimensions. Horizontally, *Nitrosopumilus*-like and *Nitrosopelagicus*-like phylotypes dominated the surface water (3 m) of the Yellow Sea (YS) and East China Sea (ECS), respectively, along with increased abundance of total free-living *Thaumarchaeota* in ECS. Similar compositional changes were observed in the surface microlayer. The spatial heterogeneity of particle-attached *Thaumarchaeota* was less clear in surface microlayers than in surface waters. Vertically, the *Nitrosopelagicus*-like phylotype increased in abundance from surface to 90 m in ECS, which led to an increase in the proportion of *Thaumarchaeota* relative to total prokaryotes. This occurred mainly in the free-living fraction. These results indicate a clear size-fractionated niche partitioning, which is more pronounced at lower depths than in the surface water/ surface microlayer. In addition, associations of *Thaumarchaeota* with other microbial taxa varied between phylotypes and size fractions. Our results show that a phylotype-resolved and size-fractionated spatial heterogeneity of the thaumarchaeotal community is present in surface oceanic waters and a vertical variation of the *Nitrosopelagicus*-like phylotype is present in shallow shelf waters.

Keywords $Thaumarchaeota \cdot Phylotype \cdot Association pattern \cdot Spatial variation \cdot Surface microlayer \cdot Chinese marginal seas$

Edited by Chengchao Chen.

Xiao-Hua Zhang xhzhang@ouc.edu.cn

- ¹ Frontiers Science Center for Deep Ocean Multispheres and Earth System, and College of Marine Life Sciences, Ocean University of China, Qingdao 266100, China
- ² Laboratory for Marine Ecology and Environmental Science, Laoshan Laboratory, Qingdao 266237, China
- ³ Institute of Evolution and Marine Biodiversity, Ocean University of China, Qingdao 266003, China
- ⁴ Key Laboratory of Marine Chemistry Theory and Technology, Ministry of Education, Qingdao 266100, China
- ⁵ Institute of Aquaculture, University of Stirling, Stirling FK9 4LA, Scotland, UK

Introduction

Thaumarchaeota, which is a phylum of Archaea established in 2008 (Brochier-Armanet et al. 2008), plays important roles in nitrogen and carbon cycling by means of ammonia oxidation, carbon fixation (Könneke et al. 2005), and nitrous oxide production (Santoro et al. 2011). Members of *Thaumarchaeota* comprise all currently known ammoniaoxidizing archaea, and are recognized to be dominant in the coastal and open oceans, occupying both pelagic and benthic environments (Francis et al. 2005). In marine waters, the abundance of *Thaumarchaeota* varies greatly across water depths, with significantly lower cell numbers in the surface layer compared with the deep ocean. In the deep ocean, *Thaumarchaeota* accounts for up to~40% of the total microbial population (Karner et al. 2001).

The depth-related shift in Thaumarchaeota abundance is accompanied by clear divergence in phylotype. The amoA gene-based phylogeny reveals two main lineages/phylotypes of ammonia-oxidizing archaea (AOA), i.e., water column A (WCA, Nitrosopelagicus-like AOA) and water column B (WCB) that prevail in the surface and deep seawater, respectively (Francis et al. 2005; Sintes et al. 2013; Smith et al. 2014a). Also, congruent phylotype differentiation has been revealed by 16S rRNA gene analysis (Reji et al. 2019; Tolar et al. 2020). Such a depth-dependent spatial separation reflects the varying adaptations of Thaumarchaeota to environmental changes, including light (Horak et al. 2018; Merbt et al. 2012), ammonium ions (Sintes et al. 2013), and oxygen (Molina et al. 2010; Qin et al. 2017). For example, the surface phylotype WCA contains photolyase to mitigate UV light exposure (Luo et al. 2014), whereas the deep phylotype WCB has a higher affinity for ammonia (Sintes et al. 2013). Furthermore, phylotype-specific association patterns have been observed (Reji et al. 2019). This suggests an important role of functional coupling and/or cross-feeding in governing Thaumarchaeota distribution patterns. The Nitrosopumiluslike lineage is the other phylotype that dominates the surface ocean and encompasses most of the cultivated marine Thaumarchaeota (Alves et al. 2018). It has also been shown to contain photolyases that may confer resistance to UV radiation (Bayer et al. 2016). Despite the widely recognized dominance of WCB below the euphotic zone, the balance between Nitrosopelagicus-like and Nitrosopumilus-like phylotypes in the surface ocean remains largely unresolved (Tolar et al. 2020).

Genomic evidence has revealed that Nitrosopelagicus may be more adapted to oligotrophic conditions than Nitrosopumilus (Santoro et al. 2015). This indicates that they may adopt different ecological strategies, including different efficiency of resource utilization. Only a comparatively few studies have reported on archaeal niche separation between size fractions, which consistently show that free-living Thaumarchaeota is more abundant (Smith et al. 2013; Zhong et al. 2020) and has different community compositions (Jing et al. 2018; Ijichi et al. 2019) compared to their particle-attached counterpart. This means that most members of Thaumarchaeota prefer a free-living lifestyle. However, the discovery of a thaumarchaeotal strain with the capacity to move toward nutrient-rich particles suggests the existence of a particle-attached strategy (Bayer et al. 2016). Maybe, there is divergence in the preferred lifestyle between Thaumarchaeota phylotypes and perhaps free-living and particle-attached Thaumarchaeota show different dynamic patterns, which are poorly characterized. This information is important for a comprehensive understanding of the dynamics of Thaumarchaeota in the marine environment.

The sea surface microlayer (SML), which is a boundary layer at the air-sea interface, is generally considered to comprise the uppermost 1 mm of the ocean (Cunliffe and Murrell 2009). The role of SML in regulating air-sea gas exchange has stimulated broad attention in investigating the ecosystem therein (Carlucci et al. 1985; Cunliffe et al. 2011, 2013; Maki. 2002). Physicochemical and microbiological characterization of SML has demonstrated enriched particles, organic matter, inorganic nutrients, and cell numbers in comparison with the underlying surface water (Cunliffe et al. 2011; Franklin et al. 2005). Also, contrasting microbial communities between SML and surface water have been observed, with the differences being more conspicuous in bacterial than archaeal communities (Cunliffe et al. 2009). Considering the higher concentration of inorganic nutrients (e.g., ammonia), we hypothesized that the SML may have selected differential thaumarchaeotal communities and nitrification activity compared to that of the surface water. However, the diversity and distribution pattern of Thaumarchaeota in marine SML remains largely unknown (Wong et al. 2018).

Marginal seas host abundant Thaumarchaeota that experiences significant fluctuations in community composition in response to strong physicochemical gradients (Hu et al. 2011; Liu et al. 2014, 2018; Smith et al. 2014b). These coastal pelagic environments, thus, provide an interesting opportunity for studying the spatial dynamics of Thaumarchaeota and their niche partitioning between different size fractions. In this study, a multidimensional distribution pattern of the thaumarchaeotal community was investigated in the eastern Chinese marginal seas, where terrestrial discharge, anthropogenic activity, and ocean current cause significant environmental variability. The spatial distribution of Thaumarchaeota was examined across different sea areas (Yellow Sea [YS] versus East China Sea [ECS]), depths (surface [3 m] versus SML and lower euphotic depths [10–90 m]), and size fractions (free living *versus* particle attached). A total of 120 water samples were collected at 26 sites (Fig. 1 and Supplementary Table S1) and were subject to 16S rRNA gene sequencing and qPCR analysis. The results support the importance of analyzing the thaumarchaeotal community at a finer phylotype level, and provide a better understanding of the spatial heterogeneity of Thaumarchaeota in the surface ocean.

Results

Thaumarchaeota abundance and phylogeny

Quantitative analysis revealed that the abundance of thaumarchaeotal 16S rRNA genes varied from 1.24×10^2 (except for the P5 particle-attached surface sample where no abundance was detected) to 4.9×10^7 genes/L seawater across all samples (sample H2 was excluded due to abnormal high



Fig. 1 Map, which was generated in Ocean Data View, showing the sampling sites



values of about orders of magnitude higher than other samples). Spatially, clear variations were observed between sampling areas, with higher gene abundance in the ECS than in the YS (Welch *t*-test, P = 0.004). The higher thaumarchaeotal abundance in the ECS mainly comprised the free-living fraction (Welch *t*-test, P = 0.003; Fig. 2A). Specifically, the free-living Thaumarchaeota had a high average abundance, approximately 20-fold higher than the particle-attached ones across the surface and SML samples. No significant differences were observed between the surface and SML samples (Welch *t*-test, P = 0.412; Fig. 2A). However, quantitative analysis revealed an increasing trend in Thaumarchaeota abundance with depth at sites of the ECS P section, with significantly higher abundance in the deepest layers compared to the surface zone across all sites (Welch *t*-test, P < 0.001; Fig. 2B). This depth-related variation seemed to be more pronounced in free-living Thaumarchaeota compared to its particle-attached counterpart, although the differences were not significant (Welch *t*-test, P = 0.743). However, in line with that observed in the surface water and SML, a higher abundance of free-living relative to particle-attached Thaumarchaeota was seen across the lower depths (Welch t-test,

P < 0.001).



Fig. 2 qPCR-derived *Thaumarchaeota* abundance in water samples of the eastern Chinese marginal seas. A free-living and particle-attached *Thaumarchaeota* in surface microlayer and surface water. **B** free-living and particle-attached *Thaumarchaeota* at different water depths

of ECS sites P1–P6. *ECS* East China Sea; *YS* Yellow Sea. Samples from sites H1, H10, B1, B3, B5, and T3 were not size fractionated, and were not included. The asterisks denote significant differences between areas. **, P < 0.01

Illumina sequencing generated 5,962,278 high-quality reads of the 16S rRNA gene amplicons (average length of ~273 bp), which were rarefied to 28,134 per sample for equalizing sampling depth. A total of 6318 prokaryotic OTUs were clustered at a 97% similarity cutoff. Among them, 123 were affiliated to Archaea, corresponding to 1.95% of all OTUs and 4.15% of all sequences. The archaeal community comprised five phyla including Thaumarchaeota (33 OTUs), Euryarchaeota (55 OTUs), Bathyarchaeota (9 OTUs), Woesearchaeota (23 OTUs), and Lokiarchaeota (1 OTU). The first two of these were the most abundant, accounting for 98.8% of the archaeal community. At class and family levels, Marine Group I (MG-I) of Thaumarchaeota and MG-II of Euryarchaeota were the predominant groups constituting 62.3% and 33.6% of the whole archaeal community, respectively. This was followed by Euryarchaeota MG-III which comprised 2.43% of the total.

A total of 33 OTUs were affiliated with *Thaumarchaeota*, which included 24 MG-I OTUs and 9 Soil Crenarchaeotic Group (SCG) OTUs. The MG-I OTUs accounted for 99.7% of all thaumarchaeotal sequences, and were mainly represented by two phylotypes i.e., *Nitrosopelagicus*-like (ten OTUs) and *Nitrosopumilus*-like (three OTUs) lineages (Supplementary Fig. S1 and Table S2). These comprised 68.3% and 29.8% of the MG-I community, respectively. Also, the WCB group was present, accounting for 0.5% of the community (eight OTUs). Of the other three MG-I OTUs, one was affiliated with *Nitrosarchaeum* whereas the other two did not have any confirmed classifications. Overall, the *Nitrosopelagicus*-like phylotype was the most abundant thaumarchaeotal clade in the eastern Chinese marginal seas.

Spatial distribution of *Thaumarchaeota* in the surface and SML

The surface water (from 20 sites) and SML (from 17 sites) samples were separately analyzed to explore thaumarchaeotal horizontal distribution patterns. The OTU-level nonmetric multidimensional scaling analysis (NMDS) plot revealed a significant community separation based on sampling area, with the ECS and YS communities forming two geographic clusters (Fig. 3A; PERMANOVA, $R^2 = 0.085$, P = 0.001). This spatial community segregation was significant in both the surface water ($R^2 = 0.122$, P = 0.001) and SML ($R^2 = 0.088$, P = 0.017) (Supplementary Fig. S2). By comparison, no significant separations were observed between lifestyles (free-living verse particleattached fractions) according to the PERMANOVA analvsis ($R^2 = 0.017$, P = 0.303), consistent with the NMDS analysis showing overlap of samples from the two fractions (Fig. 3B). Both the free-living and particle-attached communities displayed spatial segregation between the YS and ECS, and had significant correlations with temperature



Fig. 3 Non-metric multidimensional scaling plots of the thaumarchaeotal communities. A, B comparison of surface and surface microlayer between sampling areas and lifestyles. C comparison between surface and surface microlayer. D comparison between different water depths of ECS sites

(Supplementary Table S3) and PCNM 1 (the first component of the principal coordinates of neighborhood matrix; Supplementary Fig. S3) as shown by the distance-based redundancy analysis (db-RDA). Overall, *Thaumarchaeota* communities in surface water and SML were more variable between areas than between lifestyles.

Despite the overall community similarity of free-living and particle-attached Thaumarchaeota, calculation of Bray-Curtis dissimilarity showed a higher degree of lifestyle partitioning in surface waters (Bray-Curtis dissimilarities averaged 0.82 ± 0.21) than in SML (0.76 ± 0.22 ; Welch t test, P < 0.001). Moreover, it was found that these two fractions displayed contrasting horizontal distribution patterns between surface water and SML (Supplementary Fig. S4). In surface waters, both the fractions varied significantly between the YS and ECS ($R^2 = 0.180$, P = 0.002 and $R^2 = 0.150$, P = 0.002 for free-living and particle-attached, respectively). In SML, the YS-ECS community partition was observed only in the free-living fraction ($R^2 = 0.132$, P = 0.038) but not in its particle-attached counterpart $(R^2 = 0.065, P = 0.537)$. This meant that particle-attached Thaumarchaeota in SML are more similar independent of the geographical location.

The observed spatial heterogeneity in the surface water and SML was mirrored by variations in the relative abundance of dominant lineages between areas (Fig. 4; Supplementary Fig. S5). Nearly all of the thaumarchaeotal members in the YS belonged to the Nitrosopumilus-like phylotype, and a single OTU constituted > 80% of the community at most YS sites. By contrast, the ECS sites were populated by a more diverse community represented mainly by the Nitrosopelagicus-like phylotype. Also, the SCG clade had a high abundance in ECS, particularly at sites near the Changjiang estuary, indicating a potential influence of river discharge. The relative distribution of the dominant thaumarchaeotal groups showed similar dynamic trends between the surface water and SML (Supplementary Fig. S5). However, for SML, the particle-attached community in YS, except for H11, appeared to be more diverse than in the corresponding surface water (Fig. 4). This observation was supported by comparison using Shannon diversity (Welch *t*-test, P = 0.005; Supplementary Fig. S6). By contrast, no similar results were observed in the ECS. Spatial heterogeneity of particle-attached Thaumarchaeota was lower in the SML as shown above. In the ECS, the increase of the SCG clade in surface water was mainly reflected in the particle-attached fraction.

Vertical distribution of Thaumarchaeota

Profiling of Thaumarchaeota at different depths of the studied area was achieved by a comparison between surface water and SML, and between surface water and lower depths (10-90 m from the ECS sites P1-P6). It was found that there was no clear compositional difference between the surface water and SML across all sites (Fig. 3C). By contrast, a significant community separation between surface water (3 m) and lower depths (10–90 m) was observed ($R^2 = 0.096$, P = 0.002); the surface water was similar to middle water layers (10–30 m; $R^2 = 0.074$, P = 0.051) but was separated from deeper water (\geq 50 m; $R^2 = 0.270$, P = 0.003; Fig. 3D). The Nitrosopelagicus-like lineage had the highest relative abundance among other phylotypes at almost all sampling depths (Fig. 5A). Nevertheless, the inferred absolute abundance of Nitrosopelagicus (qPCR-derived total Thaumar*chaeota* abundance × relative proportion of *Nitrosopelagicus* in the community) increased with increasing depth (Fig. 5B) and was significantly higher at the greatest depth compared to the surface (Welch *t*-test, P < 0.001). This increased abundance of Nitrosopelagicus at lower depths may have resulted in an elevated proportion of Thaumarchaeota relative to total prokaryotes (Fig. 5C). SCG occupied the 10-m layer at four sites as well as the surface water of site P3 with the highest relative abundance greater than 90% (Fig. 5A).



Fig. 4 Community composition of *Thaumarchaeota* between freeliving and particle-attached fractions in the surface water and surface microlayer (SML). OTU classification was inferred according to both phylogeny and comparison to the Silva database (Supplementary Fig.

S2). The clustering relationship of OTU 4890 and 5486 was uncertain and they are shown as MG-I unclassified (MGI_un). The blank areas indicate no *Thaumarchaeota* OTUs in the total prokaryotic community. *YS* Yellow Sea; *ECS* East China Sea

Both the free-living ($R^2 = 0.207$, P = 0.005) and particle-attached ($R^2 = 0.201$, P = 0.002) Thaumarchaeota displayed clear vertical variations in community composition. The latter correlated with dissolved oxygen and ammonium ions whereas the former did not exhibit any correlations with measured environmental variables. Significant community partitioning was found between these two fractions across depths of the P section in the ANOSIM analysis (R = 0.062, P = 0.029), although not in the PER-MANOVA analysis ($R^2 = 0.038$, P = 0.066). Indeed, the Bray-Curtis dissimilarities between free-living and particle-attached fractions (0.81 ± 0.24) were lower than those between surface water and greater depths (0.86 ± 0.20) ; Welch *t*-test, P < 0.001). This indicated that the thaumarchaeotal community was more variable with depth than between size fractions (Fig. 3D).

The lifestyle-based partitioning was more evident at lower depths (10–90 m; 0.79 ± 0.26) than at the surface (0.70 \pm 0.26) according to the Bray–Curtis dissimilarity index (Welch *t*-test, P < 0.001). This was reflected in a higher relative abundance of the *Nitrosopelagicus*-like phylotype in the free-living fraction than its particleattached counterpart (Welch *t*-test, P = 0.002). The results showed that the free-living community was less diverse compared to that attached to particles (Supplementary Fig. S7). In fact, the presence of SCG OTUs at below-surface depths was mainly associated with particles (Fig. 5A). Free-living *Thaumarchaeota*, although less diverse, was the main contributors to the above-mentioned increase in the proportion of *Thaumarchaeota* relative to total prokaryotes with depth (Fig. 5C).

Co-occurrence relationship involving Thaumarchaeota

To explore the potential interactions between thaumarchaeotal OTUs and other microbial taxa, network analyses were performed for 718 bacterial and archaeal OTUs that had an average abundance of > 0.01% across all samples. The thaumarchaeotal community involved in the network was represented by seven OTUs, which showed correlations to a wide taxonomical range, including *Gammaproteobacteria*, *Actinobacteria*, *Alphaproteobacteria*, *Marinimicrobia* and euryarchaeotal MG-II (Fig. 6). Of the seven OTUs, five and two were affiliated with *Nitrosopelagicus*-like and *Nitrosopumilus*-like phylotypes, respectively.

Fig. 5 *Thaumarchaeota* community at different water depths of ECS sites P1–P6. A relative abundance of thaumarchaeotal OTUs; B inferred absolute abundance of *Nitrosopelagicus*-like lineage; C relative proportion of *Thaumarchaeota* sequence reads in relation to the total number of prokaryotic sequences. *FL* free-living, *PA* particle-attached





Fig. 6 Co-occurrence networks for each of the seven *Thaumarchaeota* OTUs (indicated by red background and white font) involved in the community network. The size of each node is proportional to the number of connections. The thickness of each edge is proportional to the correlation coefficients. All the correlations were positive.

Correlations between the two phylotypes and/or within members of a single phylotype were observed. Both the two phylotypes showed co-occurrence links with MG-II and *Gammaproteobacteria*; however, within *Gammaproteobacteria*, the E01-9C-26 group was only correlated with the *Nitrosopelagicus*-like phylotype, whereas the *Nitrosopumilus*-like phylotype was more correlated with ZD0405 and JTB255. Moreover, links with *Actinobacteria* (mainly clade Sva0996) and *Alphaproteobacteria* (mainly clade PS1) were exclusively found in the *Nitrosopelagicus*-like phylotype. In contrast, correlations with *Gemmatimonadetes* were only observed in the *Nitrosopumilus*-like phylotype (Fig. 6).

The effect of size fractions on the co-occurrence pattern was then examined by distinguishing the free-living and particle-attached fractions. With an average abundance of > 0.01%, 546 and 859 OTUs for free-living and particle-attached fractions, respectively, were selected for the analysis. Although a lower number of OTUs was used for the free-living fraction, 9 thaumarchaeotal OTUs were involved in the network, and had 253 connections in total with other microbial taxa. In comparison, 6 thaumarchaeotal OTUs were included in the network of the particleattached fraction, and revealed only 76 connections, which were substantially lower than in the free-living network.

All sequence data were included in the network analysis. OTU2989, OTU5422, OTU4920, OTU3585 and OTU5419 were affiliated to *Nitrosopelagicus*; OTU6186 and OTU3909 were affiliated to *Nitrosopunilus*

Different *Thaumarchaeota*-microbial taxa co-occurrence relationships were observed between the free-living and particle-attached fractions (Supplementary Fig. S8). For example, OTUs affiliated with *Thermoplasmata* (mainly MG-II), *Marinimicrobia*, and *Alphaproteobacteria* were mainly connected with free-living thaumarchaeotal OTUs, whereas those belonged to *Deltaproteobacteria* and *Planctomycetacia* showed higher connections with particleattached OTUs.

Discussion

Basin-related spatial separation of *Thaumarchaeota* assemblages

The spatial pattern of planktonic *Thaumarchaeota* in the YS and ECS of the Chinese marginal seas was investigated. These two areas have contrasting hydrological conditions with respect to ocean currents, river discharges, and water mass (including Kuroshio) intrusion and have been reported to host different microbial communities especially in the sediment (Liu et al. 2020). Indeed, results presented here show that the YS surface water harbored a thaumarchaeotal community with lower abundance and

species diversity compared to the ECS. The thaumarchaeotal community in the YS was characterized by a single OTU affiliated to the Nitrosopumilus-like phylotype. Certainly, the predominance of a single species-level OTU of Thaumarchaeota has previously been observed in other coastal environments (Muller et al. 2018; Tolar et al. 2020) but this is not expected considering the role of protist grazing and virus lysis in regulating community stability. Indeed, archaeal-specific predation by eukaryotes has been documented in the North Atlantic Basin (Seyler et al. 2019). However, Nitrosopumilus spp. have genes related to predator and viral defense (e.g., glycoprotein layer and DNA modification; Ahlgren et al. 2017; Stieglmeier et al. 2014). More importantly, the low proportion of Thaumarchaeota in the total prokaryotic community may have weakened the effect of these biological processes (Herber et al. 2020). A possible explanation is high resistance of the Thaumarchaeota to environmental disturbance in the YS, which needs further elucidation.

In the ECS, Thaumarchaeota was more abundant and shifted to a community composition dominated by the Nitrosopelagicus-like phylotype. This suggests that certain environmental conditions may have promoted the proliferation of the Nitrosopelagicus-like phylotype in the ECS. In this respect, temperature showed positive correlations with the abundance of the *Nitrosopelagicus*-like lineage. This was in contrast to previous findings which found that a Nitrosopelagicus enrichment culture had a lower growth temperature optimum than Nitrosopumilus maritimus (Martens-Habbena et al. 2009; Santoro et al. 2015). However, available cultures constitute only a small fraction of the natural thaumarchaeotal species diversity (Alves et al. 2018). Indeed, abundant Nitrosopumilus-like sequences have been found in global high-latitude seawater, demonstrating that at least some species prefer lowtemperature conditions (Cheung et al. 2019). Although not measured here, the previously reported higher concentration of dissolved organic matter in surface water of the YS relative to the ECS (Yang et al. 2021) may also contribute to the niche partitioning between the two phylotypes. Thaumarchaeota is typically autotrophic (Könneke et al. 2005) with the potential of carbon fixation through the modified 3-hydroxypropionate/4-hydroxybutyrate (3-HP/4-HB) cycle (Könneke et al. 2014). Nevertheless, two strains of Nitrosopumilus have been found to require α -ketoglutaric acid as the organic source for growth (Qin et al. 2014), although a role for α -ketoglutaric acid as a H₂O₂ scavenger was proposed subsequently (Kim et al. 2016). Also, field studies have provided evidence for heterotrophy in Thaumarchaeota in both pelagic and benthic marine environments (Biddle et al. 2006; Ouverney and Fuhrman 2000; Seyler et al. 2014, 2018). Thus, the dominance of *Nitrosopumilus*-like phylotype in YS may relate to a high organic matter availability. The discovery of enriched *Nitrosopumilus*-like phylotype in the hadal ocean (> 6000 m water depth) with enhanced organic matter deposition supports this possibility (Zhong et al. 2020). Also, a recent survey of AOA in the global ocean revealed the dominance of *Nitrosopumilus*-like and *Nitrosopelagicus*-like phylotypes in coastal and ocean opens, respectively (Qin et al. 2020). These findings suggest that the *Nitrosopumilus*-like phylotype may adapt to the coastal surface water by adopting a mixotrophic lifestyle.

Differences in *Thaumarchaeota* communities between surface water and SML

SML is physico-chemically different from the underlying surface water by being enriched in organic and inorganic nutrients, and thereby microbial cells (Cunliffe et al. 2011). In this study, no significant difference in thaumarchaeotal abundance was observed between surface water and SML. This is in contrary to previous findings of higher bacterial abundance in the SML than in surface water (Aller et al. 2005; Sun et al. 2020). Interestingly, Auguet and Casamayor (2008) found that in some high mountain lakes, there was an enrichment of archaea in the SML relative to the underlying water. Compared to lakes, the hydrodynamic environments in coastal waters are more complex, and physical disturbances occur frequently. This may create unstable SML environments preventing the enrichment of archaea. As a consequence, the thaumarchaeotal community compositions were similar in SML and surface water. Also, little effect of SML on archaeal communities has been observed in other coastal sites (Cunliffe et al. 2009; Wong et al. 2018), suggesting that the SML environment is less favorable to archaea than to bacteria (Cunliffe et al. 2009).

Despite the overall community homogeneity, Thaumarchaeota in the SML and surface water displayed different spatial distribution patterns and lifestyle partitioning. Freeliving Thaumarchaeota was separated by water body in both SML and surface water. By contrast, the particle-attached fraction exhibited spatial segregation only in surface water but not in SML. This indicates the specific effects of SML on the distribution of particle-attached Thaumarchaeota. Particles are frequently enriched in SML, which serve as the basis for the formation of marine aerosols (Aller et al. 2005). Aerosolization releases microbial cells trapped in the organic particles of SML into the atmosphere, acting as a dispersal mechanism for marine microorganisms (Aller et al. 2005; Engel et al. 2017). Thus, the spatial homogeneity of particle-attached Thaumarchaeota in SML may result from the high dispersal rates. This scenario is different from that commonly seen in water where the particle-attached community is more constrained by dispersal limiting than its free-living counterpart (Liu et al. 2019b; Wang et al. 2020).

Results presented here highlight contrasting microbial biogeography of size-fractionated samples between SML and surface water.

Size-fractionated niche partitioning was magnified at lower depths (10–90 m)

A clear increase in Thaumarchaeota abundance and proportion relative to prokaryotes from surface to bottom water (<100 m) was observed at the ECS sites. Similarly, a study by Wang et al. (2019) reported the maximum concentration of archaeal isoprenoid glycerol dialkyl glycerol tetraethers at shallow ECS sites (<100 m). This elevated Thaumarchaeota abundance with depth was mirrored by a significant increase in the abundance of the Nitrosopelagicus-like phylotype. This scenario is different from the previously reported vertical partitioning of Thaumarchaeota between the sunlit surface ocean and the aphotic deep ocean, where WCB is the most prevalent clade (Francis et al. 2005; Sintes et al. 2013; Smith et al. 2014a). Thus, evidence for a vertical variation of Nitrosopelagicus in the shallow shelf region is provided here, which suggests the existence of a specific ecological niche for the Nitrosopelagicus-like phylotype in shallow subsurface waters. However, these aspects may include a variety of environmental forces, including temperature, dissolved oxygen, and light. These have been found to influence the distribution and physiological activity of other thaumarchaeotal species (Hugoni et al. 2013; Qin et al. 2014, 2017).

It is notable that the increase in abundance of the Nitrosopelagicus-like phylotype at greater depths in the ECS, occurred mainly in the free-living lifestyle, resulting in elevated proportion of Thaumarchaeota in the free-living prokaryotic communities. This was consistent with previous findings that Thaumarchaeota prefers to adopt a free-living lifestyle (Smith et al. 2013; Zhong et al. 2020). In contrast, in surface water and SML of the YS dominated by Nitrosopumilus-like sequences, no differences in abundance and community composition were observed between free-living and particle-attached fractions. It is likely that at least some members of Nitrosopumilus persisting in the surface water had evolved the ability to adhere to particles. Supporting this, Bayer et al. (2016) suggested a particle-attached life mode in Nitrosopumilus adriaticus due to the potential capability in seeking favorable microhabitats with concentrated particles. Also, the association of Nitrosopumilus with particles has been observed in other estuaries and coastal seas (Li et al. 2018; Wäge et al. 2019). Indeed, the first isolate of Nitrosopumilus was purified from an enrichment with marine tank gravels (Könneke et al. 2005). These findings suggest different life modes between Nitrosopelagicus and Nitrosopumilus, with the latter preferring to attach to particulates, probably as a survival strategy for absorbing organic matter (Qin et al. 2014) and/or avoiding sunlight (Horak et al. 2018). The differential presumed lifestyle and distribution pattern between the two phylotypes led to enhanced size-fractioned niche partitioning of *Thaumarchaeota* in the shallow subsurface water compared to the surface water.

Co-occurrence pattern varied between phylotypes and lifestyles

To a certain extent, microbial distributions may be explained by taxa-taxa interactions (Liu et al. 2019a). Using correlation-based networks, a wide co-occurrence relationship was found between Thaumarchaeota and other bacterial and archaeal taxa. Common associations among different phylotypes included positive correlations to members of Marinimicrobia and archaeal MG-II. Potential N2O-involved metabolic coupling between Thaumarchaeota and Marinimicrobia, with the capacity in producing and reducing N₂O, respectively, has been inferred via omic technologies (Hawley et al. 2017), and indicated by network analyses (Reji et al. 2019). A combination of results from Reji et al. (2019) and the present study suggest that associations of Thaumarchaeota and Marinimicrobia may be present in both epipelagic and mesopelagic depths. MG-II is a heterotrophic archaeal group dominating the surface ocean (Zhang et al. 2015); its link to Thaumarchaeota persisting in the euphotic zone may indicate metabolic cross-feeding since their representatives have different trophic modes. However, caution is needed when interpreting these results as co-occurrence patterns do not always represent true interactions, but may just indicate similar habitat preferences (Liu et al. 2019a).

Similar to the results of Reji et al. (2019), varying cooccurrence relationships were seen between Nitrosopumilus-like and Nitrosopelagicus-like phylotypes. For example, correlations to Gemmatimonadetes were only seen in the Nitrosopumilus-like lineage. Codominance of Gemmatimonadetes and thaumarchaeotal MG-I has been observed in oxic deep-sea sediments (Durbin and Teske 2011). Certainly, the genomic potential for protein degradation in Gemmatimonadetes (Baker et al. 2015) may provide ammonia for Nitrosopumilus. Indeed, Nitrosopumilus is the dominant benthic thaumarchaeotal group rather than Nitrosopelagicus (Alves et al. 2018). The Nitrosopumilus-like phylotype also showed correlations with other heterotrophic taxa such as gammaproteobacterial ZD0405, which has been reported dominant during algae blooms (Teeling et al. 2016). These results may indicate the nutritional reliance of Nitrosopumilus on microorganisms capable of degrading complex organic matter. Metabolic associations may also occur in the Nitrosopelagicus-like phylotype, but with different partners and mechanisms. For example, utilization of simplified organic matter (such as C1 and amine compound) by gammaproteobacterial E01-9C-26 group (Landry et al. 2018) may provide substrate for *Nitrosopelagicus*. The specific correlation of the Nitrosopelagicus-like phylotype to Sva0996 may indicate habitat preference, as both of them increased in abundance below the surface (Reintjes et al. 2019). Additionally, there were variations in co-occurrence patterns between size fractions. Notably, the free-living Thaumarchaeota showed higher connectivity to other taxa than the particle-attached members. This may suggest the occurrence of broad inter-species interactions in free-living Thaumarchaeota, potentially serving as a survival strategy for nutrient exchange (Doxey et al. 2015). Possibly, this may differ from the particle-attached fraction in which the correlations to Deltaproteobacteria and Planctomycetacia preferring to attach to particles (Mestre et al. 2017) may reflect their similar environmental niches. A significant correlation was observed between free-living Thaumarchaeota and nitrite-oxidizing bacteria affiliated with Nitrospinia, suggesting that the ammonia-oxidizing activity of Thaumarchaeota is more likely to occur when cells are free living rather than attached to particles. Our findings of phylotype and lifestylespecific co-occurrence patterns highlight the importance of Thaumarchaeota analyses at a finer taxonomic level.

Conclusion

This study has demonstrated horizontal and vertical heterogeneity in the community structure of planktonic Thaumarchaeota in the shallow shelf region of the Chinese marginal seas. Thaumarchaeota in SML and surface water were similar in community composition, but exhibited different spatial patterns, with the particle-attached fractions being more easily dispersed in SML than surface water. Depthrelated increase in Thaumarchaeota abundance and change in lifestyle were observed. Size-fractionated niche partitioning was more pronounced at shallow subsurface water than in surface water, which may be attributed to varying life modes between Nitrosopelagicus and Nitrosopumilus. Attachment to particles may act as a survival strategy for certain Nitrosopumilus-like species inhabiting the surface ocean. Changes of lifestyle may be promoted by specific correlations between Thaumarchaeota phylotypes and other microorganisms. This study provides useful insights into the thaumarchaeotal community distribution in the coastal surface ocean at a finer phylotype level, and raises important questions that warrant further investigation, such as how different Thaumarchaeota phylotypes coexist and diverge in the surface ocean, and which phylotype plays a more important role in the ammonia oxidation.

Materials and methods

Sampling

Seawater samples were collected at 26 sites along the eastern Chinese marginal seas aboard the R/V Dong Fang Hong 2 from June to July 2018 (Fig. 1). Ten sites were located in the YS, whereas the other sixteen were located in the ECS. Samples were collected from different depths (from surface to the near bottom) at P1-P6 sites of ECS, and only surface water was collected at other sites (Supplementary Table S1). In addition, 17 sites (Supplementary Table S1) were collected for SML samples using a Garrett metal screen (MS) as previously described, with the depth of the SML samples being ~ 300-1000 µm (Yang et al. 2022). To compare community composition between size fractions, all samples except for the surface water of H1, H10, B1, B3, B5, and T3 were separated into particle-attached (>3 μ m) and free-living (0.22-3 µm) communities. Specifically, 300 mL of SML samples and 1000 mL of samples at each other depth were filtered serially through 3-µm-pore-size and 0.22-µm-pore-size polycarbonate membranes (Millipore Corporation, Billerica, MA) using a vacuum pump at a pressure of < 0.03 MPa on board the ship. The result was 120 samples, in total. After cell collection, the filters were stored at -20 °C onboard, and -80 °C in the laboratory. Seawater salinity, temperature, and depth were recorded by a Seabird 911-plus conductivity-temperature-depth (CTD) system (Sea-Bird Electronics Inc., Bellevue, WA, USA). Dissolved inorganic nutrients (NO₃⁻, NO₂⁻, NH₄⁺, SiO₄²⁻ and PO₄³⁻) were analyzed for the surface water and samples from different depths at ECS sites P1-P6 using an AA3 autoanalyzer system (Seal Analytical Ltd., Southampton, UK).

DNA extraction, sequencing, and quantitative PCR

Total DNA was extracted from filters using the PowerSoil DNA Isolation Kit as previously described by Sun et al. (2020). 515F (5'-GTGYCAGCMGCCGCGGTAA-3') and 806R (5'-GGACTACNVGGGTWTCTAAT-3') primers (Apprill et al. 2015; Parada et al. 2016) were used for prokaryotic 16S rRNA gene amplification. The reactions occurred in a 20-µL system containing 0.4 µL of Fastpfu polymerase, 4 µL of FastPfu Buffer (5×), 2 µL of dNTP mix (2.5 mmol/L), 0.8 µL of each primer (5 µmol/L), and 10 ng of template DNA. The thermal conditions were 95 °C for 3 min, 29 cycles of 95 °C for 30 s, 55 °C for 30 s, 72 °C for 45 s, and a final extension at 72 °C for 10 min. Triplicate amplifications were conducted for each sample, and were mixed for library preparation. Pair end sequencing was carried out by the Majorbio Bio-Pharm Technology, Shanghai, China with the Illumina Miseq platform. The raw reads have

been deposited in the NCBI SRA database under the accession number PRJNA648032.

Thaumarchaeota abundance was measured using quantitative PCR (qPCR) with the primers GI334F (5'-AGA TGGGTACTGAGACACGGAC-3') and GI554R (5'-CTG TAGGCCCAATAATCATCCT-3') (Suzuki et al. 2000). The reactions occurred in a 20-µL system containing 10 µL SYBR Premix Ex Taq II (2 \times), 0.4 µL ROX Reference Dye II (50×), 0.8 μ L each primer, and 2 μ L DNA template. The thermal conditions were 94 °C for 2 min, followed by 40 cycles of 94 °C for 15 s, 59 °C for 60 s, and 72 °C for 30 s. Triplicate amplifications were performed for each sample. Tenfold serially diluted linear plasmids containing a single copy of the thaumarchaeotal 16S rRNA gene (the sequence was amplified from a coastal water sample and shown in the supplementary material) were used to generate the standard curve. The amplification efficiency varied between 0.94 and 0.99, and R^2 was greater than 0.999.

OTU clustering and statistical analysis

Raw reads were quality controlled using Trimmomatic (Bolger et al. 2014) following the criteria as previously reported (Liu et al. 2020). FLASH (Magoc and Salzberg 2011) was used to merge the pair end reads. Operational taxonomic units (OTUs) were clustered in UPARSE (Edgar 2013) at a 97% similarity cutoff and taxonomically assigned using the RDP classifier against the Silva database Release 128 (http://www.arb-silva.de). Sequencing depth of each sample was equalized to 28,134 reads corresponding to the lowest sequence number among all samples. The Thaumarchaeota OTUs were extracted according to the Silva-based taxonomy. Most of the Thaumarchaeota OTUs could be annotated at the genus level against the Silva database. Taxonomic affiliation of the OTUs was further confirmed by constructing a neighbor-joining phylogenetic tree using OTU representative sequences and closest neighbors retrieved from the NCBI. To define WCB OTUs, reference sequences reported previously were included (Tolar et al. 2020). The tree was built using the MEGA software (Kumar et al. 2016) with 278 positions, the Kimura 2-parameter model, and 1000 bootstrap replicates. Comparison of the thaumarchaeotal community across samples was performed using the NMDS. The resulting clustering relationship was examined for significance using PERMANOVA analysis except if otherwise specified. Distance-based redundancy analysis (db-RDA) was used to test the relationship between environmental factors and spatial factors and community variations. The spatial factors were generated using the principal coordinates of neighbor matrices (PCNM) analysis. These analyses were conducted with the "vegan" package in R (R Core Team 2015).

Interactions between *Thaumarchaeota* and other microbial taxa were illustrated by the network analysis. The correlation-based co-occurrence patterns were constructed in R using the "igraph" and "Hmisc". OTUs with a proportion lower than 0.01% across all samples were removed in the analysis. A valid correlation was considered if the pairwise Spearman's correlations had a correlation coefficient >10.71 and a *P* value < 0.01 (Benjamini and Hochberg adjusted). The network was visualized in Gephi (Bastian et al. 2009).

Supplementary Information The online version contains supplementary material available at https://doi.org/10.1007/s42995-023-00169-y.

Acknowledgements We thank the scientists and crews on the R/V *Dongfanghong* 2 for their assistance with sampling during the cruises. We thank Chunying Liu and Guipeng Yang both of the Ocean University of China for providing pH and DO data, respectively. This work was funded by the National Natural Science Foundation of China (92051115, 41976101, 92251303 and 41730530), the Scientific and Technological Innovation Project of Laoshan Laboratory (LSKJ202203206 and LSKJ202203201), the Shandong Provincial Natural Science Foundation (ZR2022YQ38), the National Key Research and Development Program of China (2018YFE0124100) and the Fundamental Research Funds for the Central Universities (202141009 and 202172002).

Author contribution XHZ and JWL designed the study. JWL, FYH, JL, and XYL performed the experiments and analyzed the data. JWL wrote the manuscript with the help of RYL, XSZ, and BA to prepare the figures and revised the manuscript. All authors edited and approved the manuscript.

Data availability The data that support the findings of this study are included in this published article (and its supplementary information file).

Declarations

Conflict of interest The authors declare that they have no conflict of interest. Author Xiao-Hua Zhang is one of the Editorial Board Members, but she was not involved in the journal's review of, or decision related to, this manuscript.

Animal and human rights statement This article does not contain any studies with human participants or animals performed by any of the authors.

Open Access This article is licensed under a Creative Commons Attribution 4.0 International License, which permits use, sharing, adaptation, distribution and reproduction in any medium or format, as long as you give appropriate credit to the original author(s) and the source, provide a link to the Creative Commons licence, and indicate if changes were made. The images or other third party material in this article are included in the article's Creative Commons licence, unless indicated otherwise in a credit line to the material. If material is not included in the article's Creative Commons licence and your intended use is not permitted by statutory regulation or exceeds the permitted use, you will need to obtain permission directly from the copyright holder. To view a copy of this licence, visit http://creativecommons.org/licenses/by/4.0/.

References

- Ahlgren NA, Chen Y, Needham DM, Parada AE, Sachdeva R, Trinh V, Chen T, Fuhrman JA (2017) Genome and epigenome of a novel marine *Thaumarchaeota* strain suggest viral infection, phosphorothioation DNA modification and multiple restriction systems. Environ Microbiol 19:2434–2452
- Aller JY, Kuznetsova MR, Jahns CJ, Kemp PF (2005) The sea surface microlayer as a source of viral and bacterial enrichment in marine aerosols. J Aerosol Sci 36:801–812
- Alves RJE, Minh BQ, Urich T, Von Haeseler A, Schleper C (2018) Unifying the global phylogeny and environmental distribution of ammonia-oxidising archaea based on *amoA* genes. Nat Commun 9:1517
- Apprill A, Mcnally S, Parsons R, Weber L (2015) Minor revision to V4 region SSU rRNA 806R gene primer greatly increases detection of SAR11 bacterioplankton. Aquat Microb Ecol 75:129–137
- Auguet JC, Casamayor EO (2008) A hotspot for cold crenarchaeota in the neuston of high mountain lakes. Environ Microbiol 10:1080–1086
- Baker BJ, Lazar CS, Teske AP, Dick GJ (2015) Genomic resolution of linkages in carbon, nitrogen, and sulfur cycling among widespread estuary sediment bacteria. Microbiome 3:14
- Bastian M, Heymann S, Jacomy M (2009) Gephi: an open source software for exploring and manipulating networks. In: International AAAI Conference on Weblogs and Social Media. Association for the Advancement of Artificial Intelligence
- Bayer B, Vojvoda J, Offre P, Alves RJ, Elisabeth NH, Garcia JA, Volland JM, Srivastava A, Schleper C, Herndl GJ (2016) Physiological and genomic characterization of two novel marine thaumarchaeal strains indicates niche differentiation. ISME J 10:1051–1063
- Biddle JF, Lipp JS, Lever MA, Lloyd KG, Sorensen KB, Anderson R, Fredricks HF, Elvert M, Kelly TJ, Schrag DP, Sogin ML, Brenchley JE, Teske A, House CH, Hinrichs KU (2006) Heterotrophic Archaea dominate sedimentary subsurface ecosystems off Peru. Proc Natl Acad Sci USA 103:3846–3851
- Bolger AM, Lohse M, Usadel B (2014) Trimmomatic: a flexible trimmer for Illumina sequence data. Bioinformatics 30:2114–2120
- Brochier-Armanet C, Boussau B, Gribaldo S, Forterre P (2008) Mesophilic Crenarchaeota: proposal for a third archaeal phylum, the *Thaumarchaeota*. Nat Rev Microbiol 6:245–252
- Carlucci AF, Craven DB, Henrichs SM (1985) Surface-film microheterotrophs: amino acid metabolism and solar radiation effects on their activities. Mar Biol 85:13–22
- Cheung S, Mak W, Xia X, Lu Y, Cheung Y, Liu H (2019) Overlooked genetic diversity of ammonia oxidizing archaea lineages in the global oceans. J Geophys Res: Biogeosci 124:1799–1811
- Cunliffe M, Murrell JC (2009) The sea-surface microlayer is a gelatinous biofilm. ISME J 3:1001–1003
- Cunliffe M, Harrison E, Salter M, Schäfer H, Upstill-Goddard RC, Murrell JC (2009) Comparison and validation of sampling strategies for the molecular microbial analysis of surface microlayers. Aquat Microb Ecol 57:69–77
- Cunliffe M, Upstill-Goddard RC, Murrell JC (2011) Microbiology of aquatic surface microlayers. FEMS Microbiol Rev 35:233–246
- Cunliffe M, Engel A, Frka S, Gasparovic B, Guitart C, Murrell JC, Salter M, Stolle C, Upstill-Goddard R, Wurl O (2013) Sea surface microlayers: a unified physicochemical and biological perspective of the air-ocean interface. Prog Oceanogr 109:104–116
- Doxey AC, Kurtz DA, Lynch MD, Sauder LA, Neufeld JD (2015) Aquatic metagenomes implicate *Thaumarchaeota* in global cobalamin production. ISME J 9:461–471

- Durbin AM, Teske A (2011) Microbial diversity and stratification of South Pacific abyssal marine sediments. Environ Microbiol 13:3219–3234
- Edgar RC (2013) UPARSE: highly accurate OTU sequences from microbial amplicon reads. Nat Methods 10:996–998
- Engel A, Bange HW, Cunliffe M, Burrows SM, Friedrichs G, Galgani L, Herrmann H, Hertkorn N, Johnson M, Liss PS, Quinn PK, Schartau M, Soloviev A, Stolle C, Upstill-Goddard RC, Van Pinxteren M, Zäncker B (2017) The ocean's vital skin: toward an integrated understanding of the sea surface microlayer. Front Mar Sci 4:165
- Francis CA, Roberts KJ, Beman JM, Santoro AE, Oakley BB (2005) Ubiquity and diversity of ammonia-oxidizing archaea in water columns and sediments of the ocean. Proc Natl Acad Sci USA 102:14683–14688
- Franklin MP, Mcdonald IR, Bourne DG, Owens NJ, Upstill-Goddard RC, Murrell JC (2005) Bacterial diversity in the bacterioneuston (sea surface microlayer): the bacterioneuston through the looking glass. Environ Microbiol 7:723–736
- Hawley AK, Nobu MK, Wright JJ, Durno WE, Morgan-Lang C, Sage B, Schwientek P, Swan BK, Rinke C, Torres-Beltran M, Mewis K, Liu WT, Stepanauskas R, Woyke T, Hallam SJ (2017) Diverse Marinimicrobia bacteria may mediate coupled biogeochemical cycles along eco-thermodynamic gradients. Nat Commun 8:1507
- Herber J, Klotz F, Frommeyer B, Weis S, Straile D, Kolar A, Sikorski J, Egert M, Dannenmann M, Pester M (2020) A single *Thaumarchaeon* drives nitrification in deep oligotrophic Lake Constance. Environ Microbiol 22:212–228
- Horak RE, Qin W, Bertagnolli AD, Nelson A, Heal KR, Han H, Heller M, Schauer AJ, Jeffrey WH, Armbrust EV (2018) Relative impacts of light, temperature, and reactive oxygen on thaumarchaeal ammonia oxidation in the North Pacific Ocean. Limnol Oceanogr 63:741–757
- Hu A, Jiao N, Zhang R, Yang Z (2011) Niche partitioning of marine group I Crenarchaeota in the euphotic and upper mesopelagic zones of the East China Sea. Appl Environ Microbiol 77:7469–7478
- Ijichi M, Itoh H, Hamasaki K (2019) Vertical distribution of particle-associated and free-living ammonia-oxidizing archaea in Suruga Bay, a deep coastal embayment of Japan. Arch Microbiol 201:1141–1146
- Jing H, Zhu W, Liu H, Zheng L, Zhang Y (2018) Particle-attached and free-living archaeal communities in the benthic boundary layer of the Mariana Trench. Front Microbiol 9:2821
- Karner MB, Delong EF, Karl DM (2001) Archaeal dominance in the mesopelagic zone of the Pacific Ocean. Nature 409:507–510
- Könneke M, Bernhard AE, De La Torre JR, Walker CB, Waterbury JB, Stahl DA (2005) Isolation of an autotrophic ammonia-oxidizing marine archaeon. Nature 437:543–546
- Könneke M, Schubert DM, Brown PC, Hügler M, Standfest S, Schwander T, Schada von Borzyskowski L, Erb TJ, Stahl DA, Berg IA (2014) Ammonia-oxidizing archaea use the most energy-efficient aerobic pathway for CO₂ fixation. Proc Natl Acad Sci USA 111:8239–8244
- Kumar S, Stecher G, Tamura K (2016) MEGA7: Molecular evolutionary genetics analysis version 7.0 for bigger datasets. Mol Biol Evol 33:1870–1874
- Landry ZC, Vergin K, Mannenbach C, Block S, Yang Q, Blainey P, Carlson C, Giovannoni S (2018) Optofluidic single-cell genome amplification of sub-micron bacteria in the ocean subsurface. Front Microbiol 9:1152
- Li M, Wei G, Shi W, Sun Z, Li H, Wang X, Gao Z (2018) Distinct distribution patterns of ammonia-oxidizing archaea and bacteria in sediment and water column of the Yellow River estuary. Sci Rep 8:1584

- Liu J, Yu S, Zhao M, He B, Zhang X-H (2014) Shifts in archaeaplankton community structure along ecological gradients of Pearl Estuary. FEMS Microbiol Ecol 90:424–435
- Liu Q, Tolar BB, Ross MJ, Cheek JB, Sweeney CM, Wallsgrove NJ, Popp BN, Hollibaugh JT (2018) Light and temperature control the seasonal distribution of thaumarchaeota in the South Atlantic bight. ISME J 12:1473–1485
- Liu J, Meng Z, Liu X, Zhang X-H (2019a) Microbial assembly, interaction, functioning, activity and diversification: a review derived from community compositional data. Mar Life Sci Technol 1:112–128
- Liu K, Hou J, Liu Y, Hu A, Wang M, Wang F, Chen Y, Gu Z (2019b) Biogeography of the free-living and particle-attached bacteria in Tibetan lakes. FEMS Microbiol Ecol 95:fiz088
- Liu J, Zhu S, Liu X, Yao P, Ge T, Zhang X-H (2020) Spatiotemporal dynamics of the archaeal community in coastal sediments: assembly process and co-occurrence relationship. ISME J 14:1463–1478
- Luo H, Tolar BB, Swan BK, Zhang CL, Stepanauskas R, Moran MA, Hollibaugh JT (2014) Single-cell genomics shedding light on marine *Thaumarchaeota* diversification. ISME J 8:732–736
- Magoc T, Salzberg SL (2011) FLASH: fast length adjustment of short reads to improve genome assemblies. Bioinformatics 27:2957–2963
- Maki JS (2002) Neuston microbiology: life at the air–water interface. Wiley, New York, pp 2133–2144
- Martens-Habbena W, Berube PM, Urakawa H, De La Torre JR, Stahl DA (2009) Ammonia oxidation kinetics determine niche separation of nitrifying Archaea and Bacteria. Nature 461:976–979
- Merbt SN, Stahl DA, Casamayor EO, Marti E, Nicol GW, Prosser JI (2012) Differential photoinhibition of bacterial and archaeal ammonia oxidation. FEMS Microbiol Lett 327:41–46
- Mestre M, Borrull E, Sala M, Gasol JM (2017) Patterns of bacterial diversity in the marine planktonic particulate matter continuum. ISME J 11:999–1010
- Molina V, Belmar L, Ulloa O (2010) High diversity of ammonia-oxidizing archaea in permanent and seasonal oxygen-deficient waters of the eastern South Pacific. Environ Microbiol 12:2450–2465
- Muller O, Wilson B, Paulsen ML, Ruminska A, Armo HR, Bratbak G, Ovreas L (2018) Spatiotemporal dynamics of ammonia-oxidizing *Thaumarchaeota* in distinct Arctic water masses. Front Microbiol 9:24
- Ouverney CC, Fuhrman JA (2000) Marine planktonic Archaea take up amino acids. Appl Environ Microbiol 66:4829–4833
- Parada AE, Needham DM, Fuhrman JA (2016) Every base matters: assessing small subunit rRNA primers for marine microbiomes with mock communities, time series and global field samples. Environ Microbiol 18:1403–1414
- Qin W, Amin SA, Martens-Habbena W, Walker CB, Urakawa H, Devol AH, Ingalls AE, Moffett JW, Armbrust EV, Stahl DA (2014) Marine ammonia-oxidizing archaeal isolates display obligate mixotrophy and wide ecotypic variation. Proc Natl Acad Sci USA 111:12504–12509
- Qin W, Meinhardt KA, Moffett JW, Devol AH, Virginia Armbrust E, Ingalls AE, Stahl DA (2017) Influence of oxygen availability on the activities of ammonia-oxidizing archaea. Environ Microbiol Rep 9:250–256
- Qin W, Zheng Y, Zhao F, Wang Y, Urakawa H, Martens-Habbena W, Liu H, Huang X, Zhang X, Nakagawa T, Mende DR, Bollmann A, Wang B, Zhang Y, Amin SA, Nielsen JL, Mori K, Takahashi R, Virginia Armbrust E et al (2020) Alternative strategies of nutrient acquisition and energy conservation map to the biogeography of marine ammonia-oxidizing archaea. ISME J 14:2595–2609
- R Core Team (2015) R: a language and environment for statistical computing
- Reintjes G, Tegetmeyer HE, Bürgisser M, Orlić S, Tews I, Zubkov M, Voß D, Zielinski O, Quast C, Glöckner FO, Amann R, Ferdelman

TG, Fuchs BM (2019) On-site analysis of bacterial communities of the ultraoligotrophic South Pacific Gyre. Appl Environ Microbiol 85:e00184-19

- Reji L, Tolar BB, Smith JM, Chavez FP, Francis CA (2019) Differential co-occurrence relationships shaping ecotype diversification within *Thaumarchaeota* populations in the coastal ocean water column. ISME J 13:1144–1158
- Santoro AE, Buchwald C, Mcilvin MR, Casciotti KL (2011) Isotopic signature of N₂O produced by marine ammonia-oxidizing archaea. Science 333:1282–1285
- Santoro AE, Dupont CL, Richter RA, Craig MT, Carini P, Mcilvin MR, Yang Y, Orsi WD, Moran DM, Saito MA (2015) Genomic and proteomic characterization of "*Candidatus* Nitrosopelagicus brevis": an ammonia-oxidizing archaeon from the open ocean. Proc Natl Acad Sci USA 112:1173–1178
- Seyler LM, Mcguinness LM, Kerkhof LJ (2014) Crenarchaeal heterotrophy in salt marsh sediments. ISME J 8:1534–1543
- Seyler LM, Mcguinness LR, Gilbert JA, Biddle JF, Gong D, Kerkhof LJ (2018) Discerning autotrophy, mixotrophy and heterotrophy in marine TACK archaea from the North Atlantic. FEMS Microbiol Ecol 94:fiy014
- Seyler LM, Tuorto S, Mcguinness LR, Gong D, Kerkhof LJ (2019) Bacterial and archaeal specific-predation in the North Atlantic basin. Front Mar Sci 6:55
- Sintes E, Bergauer K, De Corte D, Yokokawa T, Herndl GJ (2013) Archaeal *amoA* gene diversity points to distinct biogeography of ammonia-oxidizing *Crenarchaeota* in the ocean. Environ Microbiol 15:1647–1658
- Smith MW, Zeigler Allen L, Allen AE, Herfort L, Simon HM (2013) Contrasting genomic properties of free-living and particleattached microbial assemblages within a coastal ecosystem. Front Microbiol 4:120
- Smith JM, Casciotti KL, Chavez FP, Francis CA (2014a) Differential contributions of archaeal ammonia oxidizer ecotypes to nitrification in coastal surface waters. ISME J 8:1704–1714
- Smith JM, Chavez FP, Francis CA (2014b) Ammonium uptake by phytoplankton regulates nitrification in the sunlit ocean. PLoS ONE 9:e108173
- Stieglmeier M, Alves R, Schleper C (2014) The phylum thaumarchaeota. The prokaryotes: other major lineages of bacteria and the archaea. SpringerBerlin Heidelberg, Berlin, Heidelberg, pp 347–362
- Sun H, Zhang Y, Tan S, Zheng Y, Zhou S, Ma QY, Yang GP, Todd JD, Zhang X-H (2020) DMSP-producing bacteria are more abundant in the surface microlayer than subsurface seawater of the East China Sea. Microb Ecol 80:350–365
- Suzuki MT, Taylor LT, Delong EF (2000) Quantitative analysis of small-subunit rRNA genes in mixed microbial populations via 5'-nuclease assays. Appl Environ Microbiol 66:4605–4614
- Teeling H, Fuchs BM, Bennke CM, Krüger K, Chafee M, Kappelmann L, Reintjes G, Waldmann J, Quast C, Glöckner FO, Lucas J, Wichels A, Gerdts G, Wiltshire KH, Amann RI (2016) Recurring patterns in bacterioplankton dynamics during coastal spring algae blooms. Elife 5:e11888
- Tolar BB, Reji L, Smith JM, Blum M, Pennington JT, Chavez FP, Francis CA (2020) Time series assessment of *Thaumarchaeota* ecotypes in Monterey Bay reveals the importance of water column position in predicting distribution–environment relationships. Limnol Oceanogr 65:2041–2055
- Wäge J, Strassert JFH, Landsberger A, Loick-Wilde N, Schmale O, Stawiarski B, Kreikemeyer B, Michel G, Labrenz M (2019) Microcapillary sampling of Baltic Sea copepod gut microbiomes indicates high variability among individuals and the potential for methane production. FEMS Microbiol Ecol 95:fiz024
- Wang Y, Li D-W, Sachs JP, Hu J, Cao Y, Li L, Zhang H, Zhao M (2019) Vertical distribution of isoprenoid GDGTs in suspended particles

from the East China Sea shelf and implications for sedimentary TEX_{86}^{H} records. Org Geochem 136:103895

- Wang Y, Pan J, Yang J, Zhou Z, Pan Y, Li M (2020) Patterns and processes of free-living and particle-associated bacterioplankton and archaeaplankton communities in a subtropical river-bay system in South China. Limnol Oceanogr 65:S161–S179
- Wong SK, Ijichi M, Kaneko R, Kogure K, Hamasaki K (2018) Ammonia oxidizers in the sea-surface microlayer of a coastal marine inlet. PLoS One 13:e0202636
- Yang L, Zhang J, Yang GP (2021) Mixing behavior, biological and photolytic degradation of dissolved organic matter in the East China Sea and the Yellow Sea. Sci Total Environ 762:143164
- Yang L, Zhang J, Engel A, Yang GP (2022) Spatio-temporal distribution, photoreactivity and environmental control of dissolved organic matter in the sea-surface microlayer of the eastern marginal seas of China. Biogeosciences 19:5251–5268
- Zhang CL, Xie W, Martin-Cuadrado AB, Rodriguez-Valera F (2015) Marine Group II Archaea, potentially important players in the global ocean carbon cycle. Front Microbiol 6:1108
- Zhong H, Lehtovirta-Morley L, Liu J, Zheng Y, Lin H, Song D, Todd JD, Tian J, Zhang X-H (2020) Novel insights into the *Thaumar-chaeota* in the deepest oceans: their metabolism and potential adaptation mechanisms. Microbiome 8:1–16