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Combined Vinasse and Mineral NPK Fertilizer Affect Physio-Biochemical, Root, and Yield Characters of Faba Bean (*vicia faba* I.) Genotypes Grown on Saline Soil

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Abstract

The objective of this study is to assess the effects of combined applications of recommended traditional chemical fertilization NPK 100% without Vinasse and with 100, 75, 50 NPK + Vinasse on Physio-Biochemical, root, shoot traits, yield, and its attributes for four faba bean Giza-429, Giza-843, Misr-2 and Sakha-3 genotypes which are cultivated in the soil is salt affected of eastern Egypt. The experimental design was conducted using a strip-plot design with three replicates in 2020 and 2021. Treatments of different doses of chemical fertilization were 100% NPK without Vinasse and 100%, 75%, and 50% of NPK with Vinasse as supplementary organic fertilizer were arranged in vertical strips, while horizontal strips were devoted to faba bean genotypes (*Vicia faba L.*) i.e. Giza-429, Giza-843, Misr-2, and Sakha-3. Relative to the other fertilizer combinations, the 75% NPK + Vinasse did not contribute more than 5% more yield to Sakha 3 than it did with the 100% NPK + Vinasse. Both the fertilizer and cultivar main effects had the same positive linear trend, with relatively higher total seed yield in the case of the 75% NPK + Vinasse and of cv. Sakha 3. The harvest index (HI) was different only for fertilizer levels. The 75% NPK + Vinasse differed by a very negligible margin from the 100% NPK + Vinasse, but both differed from the other two levels. Along with seed yield, both straw and biological yields were significant for the interaction effects, as well as for the main effects. Within all fertilizer combinations, cultivars Giza 843 and Saka 3 consistently, as a subgroup, outyielded the other two cultivars, not only for total seed yield but also for all other yield component characters.

Keywords Faba bean \cdot Vinasse \cdot Mineral NPK \cdot Salinity \cdot Physio-biochemical traits \cdot Root traits \cdot Yield traits and K⁺/Na⁺ ratio

1 Introduction

Legume crops are the essential pulse crop in the world. It is a vital source of proteins, calories, minerals, dietary fibers, and vitamins for millions of people (FAO 2020).

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Moreover, legume crops improve soil fertility through their efficient role in the fixation of biological nitrogen in soil. Faba bean (*Vicia faba L.*) is an important leguminous crop used for human nutrition in Egypt, it is considered one of the important winter season legume crops

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(Semba et al. 2021). Due to its high protein content of 28–30% and high carbohydrates 51–68 (Abdellatif et al. 2012; Martineau-Côté et al. 2022). In Egypt, its cultivated area was 0.40 M h^{-1} with a production of 0.139 million tons (FAO. 2022), the total production of this crop is still scanty to cover the local consumption. From the aforementioned data, there is a big request to overcome this space between native production and demand from here. Raising crop yield is one of the principal specific purposes of the agricultural strategy and can be executed by enlarging the cultivated field, using superior productive varieties, and providing sufficient and harmonious rates of indispensable plant nutrients and weed control. High salinity levels limit some faba bean cultivation in salt-affected soil, negatively impacting physiological traits, pigments, cell viability, and yield. Salinity hampers water absorption and nutrient uptake, leading to reduced growth and unfavorable K⁺/Na⁺ and Ca²⁺/Na⁺ ratios in leaves (Filipović et al. 2020; Afzal et al. 2022; Elsherpiny 2023).

At present days, there is an increasing requirement for chemical fertilizers to meet high agricultural production. Chemical fertilizer plays a significant part in increasing soil fertility and crop productivity (Hera 1995; Jiang et al. 2018). Nevertheless, continuous use of chemical fertilizers has contributed to a consequent decline in agricultural soil quality decreased soil organic matter content, and an increase in soil acidification and environmental pollution (NING et al. 2017). Hence, the utilization of organic fertilizer which has richer nutrient elements, can improve the physical properties of soil by enhancing aggregate stability and reducing soil bulk density. It can also improve the biological and biochemical properties of soil and optimize soil microbial structure (Diacono and Montemurro 2011; Ladha et al. 2022).

Vinasse is considered as a by-product of sugar production. It represents residues from molasses fermentation processes. Nowadays, Vinasse is considered a source of organic fertilizers within the agricultural sector as an ecofriendly source of nutrient elements (Oldroyd and Dixon 2014; Hoarau et al. 2018). Liquid vinasse composition is characterized by high organic compounds and major ions (Oldroyd and Dixon 2014; Prado et al. 2013; Katakojwala et al. 2019), contains high quantities of the plant macronutrients such as nitrogen, phosphorus, potassium, iron, copper, magnesium and manganese. The use of vinasse in fertigation systems has pros; as it can improve soil fertility and crop productivity (Jiang et al. 2012). Application of vinasse on short-term crops leads to providing higher yield than crops treated with chemical fertilizer NPK (15:15:15) (Clementson et al. 2016; de Chaves et al. 2021). Vinasse can be an alternative and complementary source of nutrients; furthermore, it has a significant impact on enhanced nodules formation and N-fixation and increased legume production (Udvardi and Poole 2013; Yadav et al. 2017). Finally, the practical management of chemical fertilizer and vinasse on salt-afflicted soils and tolerant genotypes can play a significant role in increasing and sustaining the national food security of faba bean and also it can solve the environmental problem of the disposal of this agroindustrial residue (Prado et al. 2013; Soobadar and Ng Kee Kwong 2012; de Chaves et al. 2021). In the present study, field trials of different doses of recommended chemical fertilization NPK with vinasse as supplementary organic fertilizer were conducted in newly reclaimed soil which is salt-affected, to investigate the effects of combined applications for mineral fertilizer reduction and supplementation with vinasse fertilizer on chemical, physiological, yield and its attributes for four faba bean genotypes.

2 Materials and Methods

2.1 The Experimental Site, Soil, and Climatic Conditions

A field experiment was conducted at a private farm, in Ismailia Governorate, Egypt (30° 35'41.9" N and 32° 16' 45.8" E) in 2020 and 2021. The physical and chemical properties of the experimental soil were determined according to (Cotteine et al. 1982) before sowing. The experimental soil was sandy throughout the profile depth from 0 to 60 cm (65.4% coarse sand, 28.3% fine sand, 1.9% silt, and 4.4% clay). The pH was 8.1 (Suspension 1: 2.5) and the electrical conductivity was 4.1 dS m^{-1} (saturated paste extract), the available nutrients of experimental soil of N. P and K (mg kg⁻¹) were 34.8, 11.5, and 33.2 respectively; and organic matter % was 0.6 and CaCO₃% was 1.62, Soluble cations (mmole 11) i.e. Ca⁺⁺, Mg⁺⁺, Na⁺ and K⁺ were 0.93, 0.84, 1.49 and 0.43 respectively; and soluble anions CO₃⁻⁻, HCO₃⁻ and CL were 1.37, 1.02 and 1.30, respectively, while SO₄- was nil. The experimental soil was NPK deficient. The monthly minimum and maximum temperatures and total rainfall for the two growing seasons are shown in Table 1.

2.2 Agronomic Practices and Experimental Design

The Vinasse used in this study contained 20% total free amino acid, 7% free amino acid; 4.62% total-N; 0.2% P_2O_5 ; 9.8% K_2O ; 0.87% Ca; 0.16% Mg; 10.04% S; 8.5 B (mg L⁻¹); 5.3 Mo (mg L⁻¹); 71 Fe (mg L⁻¹); 11.3 Mn (mg L⁻¹); 483.8 Zn (mg L⁻¹); 5.3 Cu (mg L⁻¹); 762.6 Cytokinins (mg L⁻¹); 495.2 Gibberellic acid (mg L⁻¹); 59.75% organic matter, 34.66% organic carbon and 7.23 pH. Vinasse composition analysis according to Angel Yeast Factory. The vinasse 9

Table 1 Climatic conditions of the study site (wind speed, relativehumidity, maximum temperature, minimum temperature, precipita-
tion rate) of the area in the 2020/21 and 2021/22 seasons

2020/21 Month	WS2M	RH2M	TEMP 2 M MIN	ΜΑΧ	Precipitation (mm)
NOV	2 56	58 34	15.48	27 37	0.01
DEC	2.90	63 43	10.13	20.54	0.46
JAN	3.16	68.22	7.54	17.56	1.03
FEB	2.49	66.99	8.19	19.99	0.47
MAR	3.24	61.29	10.28	23.95	2.62
APR	2.98	61.16	12.43	25.93	3.29
2021/22	WS2M	RH2M	T2M_MIN	T2M_MAX	Precipitation
Month					(mm)
NOV	2.26	64.20	14.36	24.48	0.65
DEC	2.14	64.45	10.88	22.15	0.15
JAN	2.66	63.49	9.00	20.83	0.13
FEB	2.56	65.74	8.93	21.21	0.88
MAR	2.98	60.77	10.27	23.06	0.99
APR	3.24	52.19	12.34	28.75	0.02

T2M_ MIN, MAX (Maximum and minimum temperature at 2 m high); RH2M (Relative humidity at 2 m high); WS2M (Wind speed at 2 m high), Precipitation mm (Precipitation rate mm/month)

L h⁻¹ was applied after 20 DAS as fertigation. All diluted vinasse was added to plants in equal doses at a 7-d interval, until 90 days along the period of plant growth the total amount added was (90 L h⁻¹).

Two factors were laid out in a three-randomized complete strip block design. The vertical strips were: 100% NPK (T1); 50% NPK w/vinasse (T2); 75% NPK w/vinasse (T3); and 100% NPK w/vinasse (T4). Horizontal strips were faba bean (Vicia faba L.) cultivars: Giza429 (G1); Giza843, (G2); Misr-2 (G3), and Sakha3 (G4). These cultivars are salt tolerant (Atwa et al. 2008; Mahdi 2016; Ali et al. 2019). Seedswere obtained from the Agricultural Research Center (ARC), Giza, Egypt. Treatments were applied into 10-row $4 \text{ m} \times 5 \text{ m}$ plots. Urea (46% N) was applied 21 and 45 DAS at a rate of 47.6 kg N h⁻¹. Calcium superphosphate (15.5% P_2O_5) was added at a rate of 73.8 kg P_2O_5 h⁻¹ before planting. Potassium sulphate $(48\% \text{ K}_2\text{O})$ was applied at a rate of 114.24 kg K₂O h⁻¹ 21 and 45 DAS. The planting date was 10 November in the 2 yrs. at a seeding rate of 120 kg h^{-1} . Three seeds were planted in 20-cm apart hills on 50-cm apart rows; at 21 DAS, seedlings were thinned to two. Harvest was on 6 April in both years. Reference crop evapotranspiration (ETo) was estimated according to the Penman-Monteith equation (Allen et al. 1998). Full ET_0 , 6,666 m³ h⁻¹, dripped watering was supplied each year crop coefficient (Kc) studied under Egyptian conditions as stated by FAO 56 (Allen et al. 2005). The seeds were inoculated with the proper strain of Rhizobium (Rhizobium leguminosarum). All other field management practices were carried out-weeding, disease, and pest control-if needed.

2.3 Physio-Biochemical Traits

A 10-plant simple random sample was taken from the middle rows in each plot, 60 DAS, to estimate Chlorophyll a, Chlorophyll b (mg g⁻¹ fresh weight), carotenoids, total photosynthetic pigments (Lichtenthaler and Wellburn 1983), Chlorophyll a/b ratio, membrane stability index (MSI) (Premachandra et al. 1990), phenolic content (mg g⁻¹ dry weight) (Shahidi and Naczk 1995), proline (mg g⁻¹ dry weight) (Troll and Lindsley 1955), and osmotic potential (bars).

2.4 Root Traits

A 10-plant simple random sample was taken from the middle rows in each plot, 80 DAS, to estimate the mean value of root traits. Plants were uprooted and washed with tap water, then roots were cleaned with distilled water and blotted with tissue paper. Both shoot and root dry weights (g) were measured following oven-drying samples at 70°C for 48 h. Per plant, the total nodule number was counted on the main and lateral roots. The active nodules were separated from the roots, cut into two pieces, and observed for the inside color. Only pink/red nodules were recorded as healthy and active. The active nodules were oven-dried at 70°C for 48 h and then weighed. The nitrogenous enzyme activity (µmol C_2H_4 g⁻¹ dry-weighed nodule h⁻¹) was estimated using an acetylene reduction assay as described by (Burns and Holsten 1973).

2.5 Seed Yield Component and Yield

At harvest (147 DAS), a 10-plant simple random sample was collected from each plot's inner rows to estimate plant height (cm), branches $plant^{-1}$, number of pods $plant^{-1}$, seed pod^{-1} , 100- seed weight (g), seed $plant^{-1}$, seed weight (g) $plant^{-1}$. Plots were harvested to determine seed, straw, and biological yields (kg h⁻¹), and harvest index (HI). Both seeds and straw were dried for 48 h at 70°C to spectrophotometrically estimate Na and K ion concentration, following nitric-perchloric acid digestion(Kacar 1972).

2.6 Measurements of Nitrogen, Phosphorous, and Potassium

Total N, P, and K were determined as described by (AOAC 1975).

N uptake, N recovery efficiency (NRE), and N use efficiency (NUE) were calculated as follows;

N uptake (kg h⁻¹) = N in seed (kg h⁻¹) + N in straw (kg h⁻¹), whereas N in seeds $(kg h^{-1}) = \frac{\text{Seed N \% xseed yield (kg/h)}}{100}$ and N in straw $(kg h.^{-1}) = \frac{\text{Straw N\% xstraw yield (kg/h)}}{100}$

 $NRE = \frac{\text{Total N uptake}(kg/h) \times 100}{\text{N applied }(kg/h)}$

$$NUE = \frac{\text{Seed yield } (kg/h)}{N \text{ applied } (kg/h)}$$

The accumulated total P in seeds and straw was used to calculate P uptake (kg h^{-1}), P recovery efficiency (PRE) and P use efficiency (PUE), where:

P uptake (kg h⁻¹) = P in seed (kg h⁻¹) + P in straw (kg h⁻¹), whereas P in seeds $(kg h^{-1}) = \frac{\text{Seed P \% xseed yield (kg/h)}}{100}$ and P in straw (kg h.⁻¹) = $\frac{\text{Straw P \% x straw yield (kg/h)}}{100}$

$$PRE = \frac{\text{Total P uptake } (kg/h) \times 100}{P \text{ applied } (kg/h)}$$

$$PUE = \frac{\text{Seed yield } (kg/h)}{P \text{ applied } (kg/h)}$$

The accumulated total K in seeds and straw was used to calculate K uptake, K recovery efficiency (KRE), and K use efficiency (KUE), where:

K uptake (kg h⁻¹)=K in seed (kg h⁻¹)+K in straw (kg h⁻¹), whereas K in seeds (kg h⁻¹) = $\frac{\text{Seed K (kg/h)}}{100}$ and K in straw (kg h.⁻¹) = $\frac{\text{Straw K (kg/h)}}{100}$

 $KRE = \frac{Total K uptake (kg/h) \times 100}{K applied (kg/h)}$

 $PUE = \frac{\text{Seed yield (kg/h)}}{\text{K applied (kg/h)}}$. was calculated according to (Keuter et al. 2013).

2.7 Seed Yield Response Index (SYRI)

Seed yield response index (SYRI) was calculated using formula 2:

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SYRI = \frac{SY \text{ at a high nutrient rate } - SY \text{ at the low nutrient rate}}{\text{high nutrient rate } -\text{low nutrient rate}}
(kg seeds kg nutrient<sup>-1</sup>)
where:
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SY seed yield kg
$$h^{-1}$$

Low nutrient rate 23.8 N; 36.9 P and 56.5 K (kg h^{-1}) High nutrient rate 51.76 N; 73.98 P 123.06 K (kg h^{-1})

Based on the SYRI, genotypes are classified into four groups: (i) efficient and responsive (ER) that produce high seed yield at low and high rates of nutrient fertilizer; (ii) efficient and not responsive (ENR) that produce high seed yield at a low nutrient rate with lower response to increasing nutrient fertilizer than ER; (iii) not efficient but responsive (NER) that has low seed yield with response to increasing nutrient fertilizer; and (iv).

2.8 Statistical Analysis

The obtained data were subjected to testing normality (Shapiro–Wilk test) and homogeneity of variances (Bartlett's test) of the residuals prior to ANOVA and (Bartlett 1937; Shapiro and Wilk 1965). The combined data from the two seasons were subjected to ANOVA using R statistical software version 4.4.1. Differences among the treatments were separated by Tukey's HSD test ($p \le 0.05$). The results obtained were expressed as the mean value ± standard deviation.

3 Results

3.1 Physio-Biochemical Traits

Chlorophyll pigments of the 4 faba bean cultivars were affected by the 4 levels of fertilizers. Interaction effects differed (p < 0.05) for each of Ch A, Ch A:B, and total pigments (Table 2). Among all interaction effects, Ch A content ranged from 0.833-1.766, from 2.08-2.76 for Ch A:B, and from 1.40-2.79 for total pigments. These increments were twice as much for both Ch A and total pigments, but was by only 32% for Ch A:B, for the (Vinasse + 75% NPK) x Sakha 3 interaction effect. The marginal main effect for each of fertilizer levels and cultivars greatly varied (p = 0.000) for Ch A and total pigments, where Vinasse + 75% NPK and cv. Sakha 3 had relatively higher mean values, whereas for Ch A:B, Sakha 3 did not differ (p = 0.421). These relatively higher main effects explain, in part, the differential magnitude of these interaction effects. However, Ch B and carotenoid varied for just the marginal main effect in response to each of fertilizer and cultivars (p = 0.000 and p = 0.002). Still both trait means were 0.545 and 0.409 for (Vinasse + 75% NPK) and 0.515 and 0.359 for cv. Sakha 3.

Means of proline, phenolic content, osmotic potential, and percentage MSI greatly differed for fertilizer x cultivar interaction effect, as well as for both main effects (p=0.000) (Table 3). They all had the same trend, since the 25% reduction of NPK rate, supplemented by Vinasse, applied to Sakha 3, caused their having high means relative to other interactions. Means ranged from: 1.16–1.71, 16.94–22.73, 1.63–2.99, and 49.30–62.80, for the four traits. These ranges represent percentage increments of: 47%, 34%, 83%, and 27% in favor of Vinasse + 75% NPK rate.

Table 2 Effect of vinasse and mineral fertilization on photosynthetic pigments (mg g^{-1} fresh weight) of faba bean Genotypes

Fertilizer Treatment (T)		Chlorophyll A	Chlorophyll B	Chlorophyll A:B	Carotenoid	Total Pigments
100% NPK without Vinasse (T1)		0.940 ^C	0.430 ^B	2.199 ^B	0.264 ^C	1.64 ^C
		± 0.02	± 0.053	± 0.053	± 0.01	± 0.039
50% NPK with Vinasse (T2)		0.898^{D}	0.394 ^C	2.295 ^B	0.243 ^C	1.54 ^D
		± 0.02	± 0.052	± 0.052	± 0.011	± 0.04
75% NPK with Vinasse (T3)		1.456 ^A	0.545 ^A	2.664 ^A	0.409 ^A	2.41 ^A
		± 0.057	± 0.047	± 0.047	± 0.014	± 0.086
100% NPK with Vinasse (T4)		1.021^{B} ±0.029	0.460^{B} ± 0.033	2.224^{B} ± 0.033	$0.324^{B} \pm 0.014$	1.81^{B} ± 0.053
Genotypes (G)						
Giza-429 (G1)		0.961 ^D	0.401 ^C	2.39 ^A	0.271 ^C	1.63 ^D
		± 0.048	± 0.053	± 0.053	± 0.018	± 0.078
Giza-843 (G2)		1.111 ^B	0.473 ^B	2.34 ^A	0.326 ^B	1.91 ^B
		± 0.074	± 0.086	± 0.086	± 0.021	± 0.11
Misr-2 (G3)		1.034 ^C	0.442^{B}	2.33 ^A	0.283 ^C	1.76 ^C
		± 0.062	± 0.056	± 0.056	± 0.02	± 0.099
Sakha-3 (G4)		1.209 ^A	0.5150 ^A	2.32 ^A	0.359 ^A	2.08 ^A
		± 0.09	± 0.089	± 0.089	± 0.022	± 0.131
Interaction of T * G						
100% NPK without Vinasse	Giza-429	0.866 ^{fg}		2.30 ^{cd}		1.47 ^{hij}
		± 0.867		± 0.045		± 0.018
	Giza-843	0.966 ^{er}		2.18 ^d		1.68 ^{erg}
		± 0.967		± 0.087		±0.015
	Misr-2	0.916 ^{erg}		2.22 ^d		1.57 ^{gm}
	0.11 0	±0.917		±0.172		± 0.022
	Sakna-3	$+1.013^{-1}$		2.08^{-1}		1.81^{-1} +0.047
50% NPK with Vinassa	Gize 420	0.8335		2 34bcd		<u>1</u> 40 j
50% IVER with Vinasse	012a-429	+0.833		+0.101		+0.046
	Giza-843	0.940^{efg}		2 32 ^{cd}		1.60 ^{fgh}
	Only Only	± 0.94		± 0.17		± 0.064
	Misr-2	0.853 ^{fg}		2.30 ^{cd}		1.44 ^{ij}
		± 0.853		± 0.05		± 0.044
	Sakha-3	0.963 ^{ef}		2.20^{d}		1.69 ^{efg}
		± 0.963		± 0.103		± 0.025
75% NPK with Vinasse	Giza-429	1.220 ^d		2.61 ^{ab}		2.05 ^d
		± 1.22		± 0.072		± 0.075
	Giza-843	1.526 ^b		2.73 ^a		2.51 ^b
		± 1.527		± 0.101		± 0.038
	Misr-2	1.370 ^c		2.55 ^{abc}		2.28°
		± 1.37		± 0.044		± 0.058
	Sakha-3	1.706 ^a		2.76^{a}		2.79 ^a
1000 NDV 14 V	C:	± 1.707		±0.13		± 0.033
100% NPK with vinasse	G1za-429	0.923 ⁻¹⁸ ±0.923		2.30^{-2}		$1.01^{-8^{-1}}$
	Cize 843	1.010^{e}		2.10^{d}		1 920
	0128-045	+1.010		+0.046		+0.058
	Misr-2	0.996 ^e		2 25 ^d		1 73 ^{ef}
	111101-2	±0.997		± 0.021		± 0.055
	Sakha-3	1.153 ^d		2.24 ^d		2.04 ^d
		±1.153		± 0.019		± 0.069
P fertilizer		0.000	0.000	0.015	0.000	0.000
P genotypes		0.000	0.002	0.421	0.002	0.000
PT*G		0.003	0.836	0.024	0.930	0.010

Values are the mean of 3 replicates \pm standard errors. Different letters within columns indicate that there are significant differences at 0.05 level of probability; where uppercase letters indicate the averages of the main effects, while lowercase letters indicate the interaction

Table 3Effect of vinasseand mineral fertilization onphysiochemical traits of fababean Genotypes

Fertilizer Treatment (T)		Proline mg/g Dry Weight	Phenolic mg/g Dry Weight	Osmotic Potential (bars)	Membrane Sta- bility Index%
100% NPK without Vinasse (T1)		1.42 ^B	18.13 ^C	-2.13 ^B	53.54 ^C
50% NPK with Vinasse (T2)		± 0.033 1.29 ^C ± 0.044	± 0.232 17.42 ^C ± 0.226	± 0.000 -1.85 ^A ± 0.094	± 0.883 52.17 ^D ± 0.676
75% NPK with Vinasse (T3)		1.61^{A} + 0.029	21.11^{A} +0.345	-2.59^{D} + 0.068	59.94^{A} + 0.646
100% NPK with Vinasse (T4)		1.48^{B} ± 0.042	19.24^{B} ± 0.268	-2.40° ± 0.059	56.31^{B} ±0.52
Genotypes (G)					
Giza-429 (G1)		1 33 ^C	18 29 ^C	-2 02 ^A	52 90 ^C
012a-429 (01)		+0.052	+0.347	+0.095	+0.954
Giza-843 (G2)		1.46^{B} + 0.042	19.04^{B} + 0.451	-2.30^{BC} +0.097	55.79^{B} +0.935
Misr-2 (G3)		1 40 ^{BC}	18 /8 ^C	-2 17 ^{AB}	54.81 ^B
Wisi-2 (05)		+0.046	+0.443	+0.099	+ 1.025
Sakha-3 (G4)		1.62^{A}	20.10 ^A	-2.47 ^C	58 46 ^A
Sakila-5 (G+)		± 0.039	± 0.558	± 0.107	± 0.914
Interaction of T * G					
100% NPK without Vinasse	Giza-429	$1.24^{fgh} \pm 0.079$	$17.80^{fg} \pm 0.231$	$-1.866^{abc} \pm 0.033$	$50.63^{jk} \pm 0.606$
	Giza-843	1.41 ^{cdefg}	18.06 ^{fg}	-2.166 ^{bcdef}	53.33 ^{ghi}
		± 0.025	± 0.437	± 0.088	± 0.41
	Misr-2	$\begin{array}{c} 1.36^{\text{efgh}} \\ \pm 0.083 \end{array}$	17.43^{g} ±0.328	$-2.133^{bcde} \pm 0.088$	$52.40^{ij} \pm 1.25$
	Sakha-3	1.66 ^{ab}	19.23 ^{de}	-2.366 ^{defg}	57.80 ^{cd}
		± 0.047	± 0.376	± 0.12	± 1.069
50% NPK with Vinasse	Giza-429	1.16 ^h	16.94 ^g	-1.633 ^a	49.30 ^k
		± 0.024	± 0.442	± 0.145	± 0.529
	Giza-843	$1.35^{efgh} \pm 0.132$	$17.63^{fg} \pm 0.481$	-1.933 ^{abcd} ±0.219	$52.86^{hi} \pm 0.784$
	Misr-2	$1.23^{gh} \pm 0.059$	17.13^{g} ±0.384	$-1.800^{ab} \pm 0.252$	$51.50^{ij} \pm 0.306$
	Sakha-3	1.41 ^{cdefg}	18.00 ^{fg}	-2.033abcde	55.03 ^{efg}
		± 0.042	± 0.451	± 0.133	± 0.617
75% NPK with Vinasse	Giza-429	1.53 ^{abcde}	19.83 ^{cd}	-2.366 ^{defg}	57.00 ^{de}
		± 0.082	± 0.338	± 0.033	±0.2
	Giza-843	1.61 ^{abc}	21.23 ^b	-2.633 ^{gh}	60.16 ^b
		± 0.033	± 0.328	± 0.067	± 0.203
	Misr-2	1.59 ^{abcd}	20.66 ^{bc}	-2.466 ^{efgh}	59.80 ^{bc}
		± 0.042	± 0.145	± 0.033	± 0.306
	Sakha-3	1.71 ^a ±0.025	22.73 ^a ±0.376	$-2.900^{h} \pm 0.115$	62.80 ^a ±0.757
100% NPK with Vinasse	Giza-429	1.37 ^{defgh}	18.60 ^{ef}	-2.233 ^{bcdefg}	54.66 ^{fgh}
		± 0.082	± 0.153	± 0.088	± 0.581
	Giza-843	1.46 ^{bcdef}	19.23 ^{de}	-2.466 ^{efgh}	56.80 ^{de}
		± 0.023	± 0.233	± 0.033	± 1.012
	Misr-2	1.41 ^{cdefg}	18.70 ^{ef}	-2.300 ^{cdefg}	55.56 ^{ef}
		± 0.025	± 0.416	± 0.153	± 0.481
	Sakha-3	1.68 ^{ab}	20.43 ^{bc}	-2.600 ^{fgh}	58.23 ^{bcd}
		± 0.038	± 0.521	± 0.058	± 0.876
P fertilizer		0.000	0.000	0.012	0.000
P genotypes		0.001	0.000	0.000	0.000
P T * G		0.000	0.000	0.000	0.000

Values are the mean of 3 replicates \pm standard errors. Different letters within columns indicates that there are significant differences at 0.05 level of probability; where uppercase letters indicate the averages of the main effects, while lowercase letters indicate the interaction

3.2 Root and Shoot Traits

Although both root and shoot dry weights were significant (p = 0.002 and p = 0.000) for fertilizer x cultivar interaction, as well as both their main effects; yet mean shoot-toroot ratio for interaction was not (p = 0.08) (Table 4). For main effects (p = 0.00), their ratio was relatively higher by 86%-95% for fertilizer and 81%-90% for cultivar effects, but contrary to previous traits (Table 3), neither (Vinasse + 75% NPK) combination nor cv. Sakha 3 had the relatively highest means. For each of root and shoot dry weight, however, (Vinasse + 75% NPK) x Sakha 3 interaction had high means but with trivial margins compared to 100% NPK x Saka 3 and (Vinasse + 100% NPK) x Sakha 3 interaction effects.

Four nodule traits i.e. nodule number $plant^{-1}$, active nodule number $plant^{-1}$, nodule dry weight, and nitrogenase activity are shown in Table 4. Both numbers of total and active nodules were different (p = 0.00 and 0.003) for fertilizer x cultivar interaction. Relative to the Control 100% (NPK), their means for 75% NPK x Sakha 3 and 100% NPK x Sakha 3 were greater by margins of 21 and 8 nodules per plant total nodule number; and were greater by 16 and 8 per plant active nodule number. With narrower margins, this trend remained quite the same for nitrogenase activity. The main effects supported these three interaction effects for these three nodule traits.

3.3 Yield and its Attributes

The three morphological traits, plant height, per plant branch, and pod numbers (Table 5) were different (p < 0.01) for fertilizer x cultivar interaction. These statistical interaction variations for these three traits did not have much impact on seed pod⁻¹. The latter was significant (p = 0.01) only for both main effects, but it had a close range between 2.7-3.3 over fertilizer and 2.9-3.4 over cultivars. The interaction effect pattern for pod plant⁻¹ was performed similarly to the patterns of means of both plant height and branch plant⁻¹ for each fertilizer x cultivar interaction. The simple effect of both cv. Giza 843 and cv. Sakha 3 within each fertilizer level was quite similar, both had relatively taller plants and higher per plant branch and pod number; however not necessarily were significantly different. Within 75% NPK + Vinasse, both cultivars were significantly different for plant height, yet they were not for both per plant branch and pod number.

Yield component traits –both per plant 100 seed weight and seed number—were different (p=0.014 and 0.003) for fertilizer x cultivar interaction, as well as for main effects (Table 5). For per plant 100 seed weight, the simple effect means for the 4 cultivars within 100% NPK was different with a range of 72.0–78.3 g, but the range was wider for Vinasse + 75% NPK, 64.6–75.0 g. In the case of Vinasse + 50% and 100% full NPK, interaction mean ranges were quite relatively narrower and higher, 79.0–81.6 and 81.3–86.6 g, yet they were not statistically significant. Per plant total seed number interaction means performed in opposite direction with 100-seed weight means – the more weight, the less total number. This trend was consistent over all cultivar means within each of the 4 fertilizer levels despite the differential variation in the ranges. These ranges were 20, 15, 26, and 26 seeds among the 4 cultivars within fertilizer levels, but for Vinasse + 75% NPK, both minimum–maximum seed numbers (47–73 seeds) were higher compared to the other three within fertilizer levels, especially Vinasse + 100% NPK where the range was 38–64 seeds, making drops of 19% and of 12%.

A positive linear trend of total seed yield (kg h⁻¹), among the 4 cultivars, was towards cultivar Sakha 3 within each fertilizer level (p = 0.005) (Table 5). But all over fertilizer combinations, the 75% NPK + Vinasse did not contribute more than 5% more yield to Sakha 3 than it did with the 100% NPK + Vinasse. Both the fertilizer and cultivar main effects (p = 0.000 and = 0.003) followed the same positive linear increase, with relatively higher total seed yield in case of the 75% NPK + Vinasse and of cv. Sakha 3. The harvest index (HI) was different only for fertilizer levels (p = 0.006). The 75% NPK + Vinasse differed by a very negligible margin from the 100% NPK + Vinasse, but both differed from the other two levels (p = 0.006). Along with seed yield, both straw and biological yields were significant for the interaction effects, as well as for the main effects.

3.4 Seed and Straw K⁺/Na⁺ Ratio

For both ratios, fertilizer x cultivar interactions were different (p=0.007 and 0.008) (Table 5). Within each fertilizer, however, cultivars did not show practical considerable differences. The percentage range for cultivars within each fertilizer x cultivar interaction was 0.57, 0.31, 0.30, and 0.512, making an overall ratio range differential of 0.27% (0.30-0.57%) for straw. For seeds, percentage ranges were 2.06, 1.84, 1.29, and 1.36, making an overall ratio range differential of 0.77% (1.29–2.06%). The main effect of fertilizer combination, for straw, indicated that the presence of Vinasse with either a 25% reduction in NPK or a full amount did make a difference (p=0.000), resulting in means of 2.04 and 2.06%, from its presence with 50% NPK (mean = 1.43%). However, a full percentage NPK resulted in a nonsignificant close mean ratio, of 1.95%. In the case of seed, this K^+/N^+ ratio pattern for these two fertilizer x cultivar interactions was significantly close (6.55% vs. 7.18%) except difference (p=0.001) from both ratios in the case of Vinasse + 50% NPK and of its absence -5.90% vs. 3.79%. In general, overall main effects and interactions, the ratio of these two cations ranged about more than 2-3 folds in the seeds than in straw.

Table 4 Effect of vinasse and mineral fertilization on shoot dry weight and root traits of faba bean Genotypes

Fertilizer Treatment (T)		Root Dry Weight g/ Plant	Shoot Dry Weight g/Plant	Nodules No./ Pant	Active Nodules/ Plant	Nodules Dry Weight g/ Plant	Nitrogenase (µmol/g nodule Dry Weight)	Shoot:Root
100% NPK without Vinasse (T1)		$18.82^{C} \pm 0.57$	34.83^{C} ±0.65	79.66° ±2.46	73.25^{B} ±2.53	0.21^{C} ±0.008	30.62^{C} ± 1.406	1.86^{A} ± 0.031
50% NPK with Vinasse (T2)		$15.34^{D} \pm 0.38$	$29.84^{D} \pm 0.66$	$63.00^{D} \pm 2.55$	57.58° ±2.58	$0.17^{D} \pm 0.007$	$27.92^{C} \pm 0.953$	$1.95^{A} \pm 0.038$
75% NPK with Vinasse (T3)		$20.55^{A} \pm 0.48$	$38.45^{A} \pm 0.84$	94.33 ^A ±3.18	$87.75^{A} \pm 2.39$	$0.25^{A} \pm 0.008$	$37.40^{A} \pm 1.269$	$1.88^{A} \pm 0.017$
100% NPK with Vinasse (T4)		$19.48^{B} \pm 0.57$	36.53^{B} ±0.89	$86.66^{B} \pm 2.82$	$81.25^{A} \pm 2.74$	$0.23^{B} \pm 0.007$	$34.62^{B} \pm 1.301$	$1.88^{A} \pm 0.026$
Genotypes (G)								
Giza-429 (G1)		$16.65^{C} \pm 0.545$	31.58^{D} ±0.935	$71.58^{\circ} \pm 3.763$	$66.00^{D} \pm 3.842$	0.19 ^C ±0.01	$28.09^{D} \pm 1.24$	$1.90^{AB} \pm 0.036$
Giza-843 (G2)		$18.41^{B} \pm 0.681$	36.25^{B} ± 1.153	83.58^{B} ± 4.342	77.91^{B} ±4.247	0.22^{B} ±0.011	33.94^{B} ± 1.218	$1.97^{A} \pm 0.02$
Misr-2 (G3)		18.12^{B} + 0.506	33.91° +0.77	77.41^{BC} + 3.166	71.83° + 3.24	0.21° + 0.008	31.31° + 1.407	1.88^{BC} + 0.017
Sakha-3 (G4)		21.00^{A} + 0.708	37.90^{A} + 1.09	91.08^{A} + 4.148	84.08 ^A + 3.545	0.24^{A} +0.01	37.22 ^A +1.375	1.81° + 0.024
Interaction of T * G		_	-	_		_		_
100% NPK without Vinasse	Giza-429	$16.53^{\rm f}$ + 0.176	31.83^{g} +0.291	70.33 ^{hi} +2.963	64.00^{fg} + 3.055		26.10^{gh} + 2.108	
	Giza-843	18.73^{de} + 0.441	36.40^{d} +0.289	85.0^{bcdefg} +2.082	78.66^{bcde} + 1.453		31.80^{def} + 1.562	
	Misr-2	18.46^{e} + 0.406	33.96^{f} +0.348	76.33^{fgh} + 1.202	69.33^{ef} + 1.667		28.90^{efgh} + 2.577	
	Sakha-3	21.56^{bc} + 0.639	37.13^{d} +0.384	87.0^{bcdef} + 5.292	81.00^{bcd} + 5.568		35.70^{bcd} + 2.159	
50% NPK with Vinasse	Giza-429	14.00^{h} + 0.577	26.83^{i} +0.935	53.66^{j} + 3.18	48.00^{h} + 3.215		24.00^{h} + 0.577	
	Giza-843	14.83 ^{gh} ±0.167	-30.13^{h} ± 0.504	61.66^{ij} ± 3.528	- 56.33 ^{gh} ±3.48		29.33^{efg} ± 0.333	
	Misr-2	15.40^{g} ± 0.231	29.90^{h} ±0.306	62.33^{ij} ± 1.764	56.66^{gh} ± 1.202		26.66^{fgh} ± 1.453	
	Sakha-3	$17.13^{\rm f} \pm 0.338$	$32.50^{g} \pm 0.346$	$74.33^{gh} \pm 2.906$	$69.33^{ef} \pm 2.404$		$31.70^{\text{def}} \pm 0.907$	
75% NPK with Vinasse	Giza-429	$18.70^{de} \pm 0.458$	35.13 ^e ±0.296	$84.33^{cdefg} \pm 2.906$	$79.66^{bcde} \pm 2.404$		32.86 ^{de} ±1.795	
	Giza-843	$20.93^{\circ} \pm 0.088$	$40.13^{b} \pm 0.328$	$96.66^{b} \pm 4.631$	$90.00^{ab} \pm 4.619$		38.73 ^{abc} ±0.601	
	Misr-2	$19.73^{d} \pm 0.203$	$36.60^{d} \pm 0.529$	$88.33^{bcde} \pm 4.256$	84.33 ^{bc} ±3.756		$35.96^{bcd} \pm 1.91$	
	Sakha-3	$22.83^{a} \pm 0.176$	$41.96^{a} \pm 0.546$	$108.00^{a} \pm 3.215$	$97.00^{a} \pm 0.577$		42.03^{a} ± 2.298	
100% NPK with Vinasse	Giza-429	$17.40^{\rm f} \pm 0.265$	32.53^{g} ± 0.328	$78.00^{efgh} \pm 4.583$	$72.33^{def} \pm 4.667$		$29.40^{efg} \pm 1.79$	
	Giza-843	$19.16^{de} \pm 0.233$	$38.36^{\circ} \pm 0.555$	$91.00^{bcd} \pm 4.726$	$86.66^{abc} \pm 4.096$		$35.90^{bcd} \pm 1.852$	
	Misr-2	$18.90^{de} \pm 0.115$	$35.20^{e} \pm 0.208$	$82.66^{defg} \pm 3.18$	$77.00^{cde} \pm 1.732$		33.73 ^{cde} ±1.919	
	Sakha-3	$22.46^{ab} \pm 0.285$	40.03 b ±0.41	95.00 bc ±5.686	$89.00^{ab} \pm 5.508$		$39.46^{ab} \pm 0.581$	
P fertilizer		0.000	0.000	0.000	0.000	0.000	0.000	0.000
P genotypes		0.000	0.000	0.000	0.000	0.000	0.000	0.000
P T * G		0.002	0.000	0.000	0.003	0.7071	0.000	0.0821

Values are the mean of 3 replicates \pm standard errors. Different letters within columns indicate that there are significant differences at 0.05 level of probability; where uppercase letters indicate the averages of the main effects, while lowercase letters indicate the interaction

Table 5 Effect of vina:	sse and n	nineral fertil	ization on yie	eld and its attrib	outes seed	and straw K	+/Na ⁺ ratio	of faba bean Ge	notypes				
Fertilizer treatment (T)		Plant Height	Branchs No./ plant	Pods No./plant	Seed No./pod	100 Seed Weight	Seed No./ plant	Seed Weight kg/h	Straw Weight kg/h	Biological Yield kg/h	Harvest Index	K:Na in Straw Ratio	K:Na in Seed Ratio
100% NPK without Vinasse	s (T1)	66.5 ^C	3.33 ^{BC}	13.83 ^B	2.83 ^B	80.41 ^B	40.08 ^B	3728 ^B	4007 ^{BC}	7735 ^{BC}	48.0 ^B	1.95 ^A	5.90 ^B
		± 1.92	± 0.28	± 0.98	±0.11	± 0.57	±3.68	± 330	±343	± 673	± 0.36	± 0.08	±0.32
50% NPK with Vinasse (T2	(2	60.7 ^D	2.75 ^C	11.33 ^C	2.75 ^B	84.58 ^A	31.33^{B}	3055^{B}	3344.7 ^c	$6400^{\circ} \pm 434$	47.68 ^B	1.43^{B}	3.80 ^C
		± 1.59	± 0.18	± 0.61	± 0.13	±1.02	±2.46	±212	±223		± 0.25	± 0.05	±0.24
75% NPK with Vinasse (T3	(٤	81.9 ^A	4.25 ^A	16.75^{A}	3.75 ^A	69.16^{D}	63.16^{A}	5064^{A}	5258^{A}	10322^{A}	49.06^{A}	2.04^{A}	6.55 ^{AB}
		± 2.28	± 0.22	± 0.59	± 0.13	± 1.58	± 3.65	± 195	±200	±394	± 0.193	± 0.04	± 0.21
100% NPK with Vinasse (T	(4)	73.6 ^B	3.67 ^{AB}	15.75 ^{AB}	3.33 ^A	75.66 ^C	53.33 ^A	4616 ^A	4781 ^{AB}	9396 ^{AB}	48.99 ^A	2.06 ^A	7.18 ^A
		± 1.80	±0.19	±0./2	±0.19	±1.03	土4.83	± 301	±338	±089	±0.228	±0.07	±0.29
Genotypes (G)													
Giza-429 (G1)		63.9 ^D	3.00^{A}	11.83 ^C	2.91 ^B	80.41 ^A	35.33 ^C	3245 ^C	$3478^{B} \pm 255$	6724 ^C	48.04^{A}	1.65 ^C	5.09 ^C
		± 2.34	± 0.25	±0.72	± 0.15	± 1.63	±3.34	± 258		±511	±0.42	± 0.08	±0.45
Giza-843 (G2)		72.6 ^B	3.75 ^A	15.33 ^{AB}	3.16^{AB}	76.2 ^{BC}	50.58 ^{AB}	4291 ^{AB}	4507 ^A	8799 ^{AB}	48.52^{A}	1.95^{A}	6.09 ^{AB}
		± 2.69	± 0.28	± 1.00	±0.24	±2.53	± 0.32	土 427	±417	±844	±0.29	±0.09	±0.45
Misr-2 (G3)		67.7 ^C + 2.53	3.33^{A} + 0.19	14.00 ^B + 0.88	3.16 ^{AB} +017	78.5A ^B +1 77	45.08 ^B +447	4084 ^B + 345	4347 ^A + 341	8431 ^B +685	48.29 ^A +0.29	1.81^{B} +0.09	5.53 ^{BC} +0.42
					, 		A 10 73			- 200 -	40.04		
Sakna-3 (G4)		/8.4 ^{°°} + 2.77	5.92 + 0.29	+0.65	5.41 + 0.15	4. / 2 ~ 土 2. 14	+4.06	+24843	209 + 209	+426	+0.20	+0.09	6.730A +0.39
Interaction of T * G		- -	1	000	-		8 -	-	-	1	-1	-	1
	ċ	co rahi - i Cr		a cade			ab ade	p oo to	10000	poor		1 mdef . 0 14	10101 .0.00
100% NPK without Vinasse	-12a- 429	C0.1±°C.U0	±0.33	± 1.86		81. /~ ±1.20	±7.88	± 718.4	3030° ±671	⊃828° ±1390		I./ ^{***} ±0.14	4.915'±0.07
	Giza-	60 15 ^{de}	3 67abc	1.4 7acd		81 66 ^{abc}	40 7 ^{bcde} + 10	38AAabcd	A162 abcde	8006abc		2 03 ^{abc} ± 0 1	6 13de
	843	± 3.47	± 0.33	±2.40		±0.9	ł	± 97	± 100	±1968			±0.32
	Misr-2	62.89 ^{efgh}	3.00^{bc}	13.67 ^{bde}		79 ^{abcd}	41 ^{bcde}	$3758^{acd} + 436$	4065 ^{abcde}	7822 ^{acd}		1.9 ^{bcd}	5.57 ^{ef}
		± 3.71	± 0.001	± 1.76		+	± 5.29		±556	±992		± 0.12	± 0.40
	Sakha-	73.40 ^{cd}	4.00^{ab}	16.33 ^{ac}		79.3 ^{abcd}	49 ^{bcde}	4514^{abcd}	4771 abcde	$9285^{abcd} \pm 337$		2.23 ^{ab}	$6.98^{abc} \pm 0.58$
	33	± 0.87	± 0.001	± 0.67		±0.9	±2	± 14	± 197			± 0.09	
50% NPK with Vinasse	Giza-	54.83 ⁱ	2.33°	9.67 ^e		86.66 ^a	26.0^{e}	2603^{d}	2916 ^e	5519 ^d		1.28^{g}	2.94 ^h
	429	± 1.65	± 0.33	± 0.67		± 2.19	±4.36	± 393	± 481	±873		± 0.13	± 0.08
	Giza-	$61.7^{\rm fghi}\pm1.45$	3.00^{bc}	11.33 ^{de}		84.3 ^{ab}	30.33 ^{de}	2949 ^{cd}	3207^{cde}	6156 ^{cd}		$1.5e^{fg}$	3.84^{gh}
	843		± 0.58	± 0.88		±2.40	±4.91	± 404.6	±396	±800		± 0.07	± 0.28
	Misr-2	58.0 ^{hi}	2.67^{bc}	10.67 ^{de}		86.00^{a}	28 ^e	2796 ^d	3086^{de}	5882 ^d		1.36^{fg}	$3.622^{h} \pm 0.505$
		± 0.58	± 0.33	± 1.33		± 1.528	±4	± 383.7	±380	±760		± 0.04	
	Sakha-	68.2 ^{def}	3.00 be	$13.7^{bcde} \pm 0.88$		81.3 ^{abc}	41 ^{bcde}	$3874^{acd} \pm 222$	4170^{abcde}	$8043^{abc} \pm 513$		1.59 ^{ef}	4.79 ^{fg}
	б	± 0.37	± 0.001			± 1.20	±2.65		±292			± 0.06	± 0.284
75% NPK with Vinasse	Giza-	74.8 ^{cd}	$3.67^{\rm abc}$	$14.33^{acd} \pm 0.33$		75 ^{bcde}	47.7 ^{bcde}	$4125^{acd} \pm 185$	4324 ^{abcde}	$8449^{abcd} \pm 49$		1.9^{bcd}	5.92 ^{de}
	429	± 2.19	± 0.33			±3.464	±4.3		±306			± 0.05	± 0.297
	Giza-	84.1 ^b	4.67^{a}	18.33^{a}		64.66 ^f	73.33 ^a	5514^{a}	5669^{a}	11181^{a}		2.1 ^{abc}	6.9 ^{abcd}
	843	± 2.01	± 0.67	± 0.33		± 1.202	±1.33	± 79.48	± 110	± 174		± 0.08	± 0.36
	Misr-2	76.7 ^c	4.00^{ab}	16.00^{ac}		70.7 ^{def}	58.4 ^{abc}	4980^{ab}	5197^{ab}	$10176^{ab} \pm 531$		1.98^{ab}	6.19 ^{cde}
		± 2.19	± 0.001	± 0.1		±2.19	±5.04	± 276.2	±259			±0.05	± 0.3
	Sakha-	91.9 ^a	4.67^{a}	18.33 ^a		66.3 ^{ef}	73.33ª	5640 ^a	5843 ^a	11483 ^a		2.21^{ab}	7.21 ^{abc}
	3	± 3.25	± 0.33	±0.88		±2.19	± 3.528	± 91.52	±134	±219		±0.09	土0.7/

Fertilizer treatment (T)		Plant Height	Branchs No./ plant	Pods No./plant	Seed No./pod	100 Seed Weight	Seed No./ plant	Seed Weight kg/h	Straw Weight kg/h	Biological Yield kg/h	Harvest Index	K:Na in Straw Ratio	K:Na in Seed Ratio
100% NPK with Vinasse	Giza- 429	65.4 ^{ef} ±1.55	3.33 ^{abc} ±0.67	12.67 ^{ce} ±0.88		78.3 ^{abcd} ±1.8	38 ^{cde} ±2.646	$3456^{bcd} \pm 212$	3641 ^{bcde} ±164	7098 ^{bcd} ± 371		$1.77^{cd} \pm 0.05$	$6.58^{bcd} \pm 0.24$
	Giza- 843	75.6 ^{cd} ± 1.98	$3.67^{\rm abc}$ ± 0.33	17.00^{ab} ± 1.16		74 ^{cde} ±4.04	58a ^{bc} ±14.19	4861 ^{ab} ± 1008	4989 ^{abcd} ±978	$9850^{ m abc} \pm 198$		2.18 ^{ab} ±0.12	$7.47^{\rm ab} \pm 0.599$
	Misr-2	73.3 ^{cd} ±2.58	3.67 ^{abc} ±0.33	15.67 ^{ac} ±1.45		$78.3^{abcd} \pm 1.2$	53ab ^{cd} ±9.85	4804 ^{ac} ± 819.6	5041 ^{abc} ±77	9844 ^{abc} ± 159		$1.99^{ab} \pm 0.1$	$6.75^{bcd} \pm 0.80$
	Sakha- 3	80.3 ^{bc} ± 0.78	4.00 ^{ab} ± 0.001	17.67^{ab} ± 0.88		72 ^{def} ±4.73	64.3 ^{ab} ±4.33	5344 ^{ab} ± 83.48	5452 ^{ab} ±27	$10796^{ab} \pm 104$		$2.29^{a} \pm 0.10$	$7.940^{a} \pm 0.36$
P fertilizer	0.000		0.000	0.003	0.011	0.000	0.000	0.000	0.008	0.000	0.006	0.000	0.001
P genotypes	0.000		0.151	0.007	0.016	0.002	0.001	0.003	0.007	0.000	0.243	0.001	0.004
PT * G	0.000		0.012	0.012	0.981	0.014	0.003	0.005	0.003	0.000	0.914	0.007	0.008

the averages of the main effects, while lowercase letters indicate the interaction

3.5 Protein Content and Uptake, Recovery, and Use Efficiency of NPK

Al 10 measured parameters statistically varied for fertilizer x cultivar interaction, in addition to main effects (p < 0.01) (Fig. 1 and Table 6). The 50% NPK + Vinasse x cultivar interaction resulted in the least protein contents for all cultivars relative to those of the Control and both the 75% and 100% NPK (+ Vinasse), reaching relatively more seed protein for the 4 cultivars within the 75% NPK + Vinasse. The performance of cultivar simple effects, within each fertilizer level, was consistently similar, where both Giza 843's and Sakha 3's protein contents were relatively higher. Protein content for the latter two cultivar main effects supported this result since theirs over yielded the other two cultivars.

The N, P, and K uptake, each varied ($p \le 0.007$) for the fertilizer (Fig. 1A) x cultivar (Fig. 1B) interaction (Fig. 1C). All three uptakes (kg h⁻¹) among cultivars within each fertilizer level mimicked the same response for protein content. Cultivar means might be split into two subgroups within each fertilizer level: i) Giza 843 and Sakha 3, and ii) Giza 429 and Misr 2, resulting in a positive linear response towards Saka 3 for every fertilizer level. Among the 4 linear responses, the 75% NPK + Vinasse caused the highest relative response, regardless of the cultivar, for each individual element uptake; the uptake (kg h⁻¹) ranges were 206.8–312.8 for N, 22.5–35.7 for P, and 126.6–188.3 for K Fig. 1).

Neither interaction effects for recovery efficiency (RE) nor use efficiency (UE) for each of N, P, and K ($p \le 0.01$), did vary from that of their mean uptakes regarding each of the: i) positive linear response, (ii) relative outyield of 75% NPK x cultivar combination and of iii) both cultivars Giza 429 and Sakha 3 within each interaction effect (Table 6). Moreover, both main effects did differ –where p values were generally ≤ 0.009 – as for all these of the three mean element uptakes.

3.6 Seed Yield Response Index (SYRI)

The SYRI of the four cultivars is shown in (Fig. 2). At low NPK rate, the mean seed yield was 3055.5 kg h⁻¹. At a rate of 54.2 kg N h⁻¹, 74.2 kg P h⁻¹, and 124.3 kg K h⁻¹, SYRIs were 65.6, 42.3, and 27.6 kg seeds NPK kg h⁻¹. For each particular cultivar at low NPK: Sakha3 was categorized as both efficient and nonresponsive (ENR) since both its yield and SYRI were greater than the mean seed yield; Giza 429 was neither efficient nor responsive (NENR) for total mean seed yield was greater than both its yield and SYRI; Misr 2 and Giza 843 were categorized as nonefficient but responsive (NER) where both their seed yield were lower than the mean seed yield, yet their SYRIs were greater.



Fig. 1 Effect of vinasse and mineral fertilization (**a**), faba bean genotypes (**b**) the interaction between the fertilization and genotypes (**c**) on NPK uptake. Values are the mean of 3 replicates \pm standard errors. Different letters within columns indicates that there are significant differences at 0.05 level of probability; where uppercase let-

4 Discussion

Addition of vinasse to each of 100% and of 75% NPK significantly affected each of chlorophyll a &b content and their ratio, carotenoids, and total photosynthetic pigments (Table 2). And proline, phenolic content, membrane stability index (MSI%), and osmotic potential (Table 3) Organic fertilizers, e.g. vinasse, cause reduction in soil acidification, and improves soil nutrient availability (Abou Hussien et al. 2017). In addition, vinasse contains sugar which is essential for building up energy required by soil microbes. In a newly-reclaimed soil in Egypt, which was treated by sugar cane by products, its pH dropped and essential nutrient content improved (Otieno et al. 2009). Vinasse also contains cytokinins and gibberellic acid (Clementson et al. 2016). These growth regulators, and the like, act as stimulants for transporting and supplying photosynthetic assimilates, by which source-sink translocation improves (Desoky et al. 2021; Rafique et al. 2021). Vinasse increases

ters indicate the averages of the main effects, while lowercase letters indicate the interaction. Whereas T1 = 100% NPK without vinasse, T2 = 50% NPK with vinasse, T3 = 75% NPK with vinasse, T4 = 100% NPK with vinasse, G1 = Giza-429, G2 = Giza-843, G3 = Misr-2, G4 = Sakha-3

absorbed N and Mg which are structural components of chlorophyll, and this, therefore, is likely to enhance chlorophyll accumulation, thereby increases photosynthetic rate. Both humic acid and P improve N, Fe, and Mg ion contents, and P induces Chlorophyll a &b contents to affect their ratio, in addition to the carotenoids, total photosynthetic pigments, proline, phenolic content, MSI, and enhance osmotic potential (Frydenvang et al. 2015; Carstensen et al. 2018). The combined effect of vinasse with recommended full NPK rate, only 50% and 75% was found effective on maintaining each of faba bean leaf proline, phenolic content, and osmotic potential. Proline and phenols act as indicators of abiotic stress when a plant experiences an osmotic stress (Raza et al. 2023). Supplementary vinasse with NPK fertilizer may have an indirect effect on proline and phenols to adjust plant osmotic potential. Moreover, vinasse contains zinc and an amino acid this amino acid has a role in osmotic protection when a plant suffers from osmotic stress Zn plays a role in plant metabolic pathways

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Fertilizer Treatment (T)		Protein Yield kg h ⁻¹	NRE	NUE	PRE	PUE	KRE	KUE
100% NPK without Vinasse (T1)		903.75 ^B	368.544 ^C	78.327 ^C	24.729 ^C	57.101 ^C	85.206 ^c	32.636 ^B
		±8.466	±3.502	± 6.936	±3.046	±4.482	± 0.969	±2.895
50% NPK with Vinasse (T2)		696.74 ^C	517.22 ^B	106.126^{AB}	28.021B ^C	73.935 ^B	94.232 ^{BC}	45.078^{A}
		±5.194	±3.589	±9.075	土4.752	±5.696	± 1.367	± 3.131
75% NPK with Vinasse (T3)		1315.52 ^A +5 037	472.615 ^B + 2.715	124.463 ^A 6 012	54.119^{A}	$70.031^{\rm B}$	168.085^{A}	52.612 ^A
		/ co.c H	C17.CT		0/0°C H		160.1 I	T-070
100% NPK with Vinasse (T4)		1153.89 ^A ±9.298	601.21^{A} ± 3.56	87.775 ^{BC} ±6.13	33.313 ^B ±3.935	84.606 ^A ±4.746	107.136 ^B ±1.022	37.143 ^B ±2.832
Genotypes (G)								
Giza-429 (G1)		768.55 ^C	375.614 ^C	79.066 ^C	25.939 ^C	50.52^{B}	85.282 ^C	33.387 ^c
		± 6.897	± 3.357	± 6.949	± 2.621	± 5.596	± 0.833	± 3.432
Giza-843 (G2)		1080.69^{B}	480.505 ^B	102.777 ^{AB}	36.751 ^B	82.001 ^A	119.501 ^B	43.386 ^{AB}
		± 11.708	±5.12	±7.371	±2.426	± 6.996	±0.756	± 3.533
Misr-2 (G3)		989.06 ^B	664.162 ^A	97.447 ^B . 4.704	34.137 ^B	90.906 ^A	110.07 ^B	41.133 ^B
		±9.217	100.0±	±4./94	566.7∓	0/c.c∓	±0./89	±0.182
Sakha-3 (G4)		1231.59^{A}	439.308 ^b u	117.401^{A}	43.356^{A}	62.245 ^b	$139.807^{\rm A}$	49.563^{A}
		±7.083	±3.481	± 6.692	±2.707	±5.729	±0.859	±4.908
Interaction of T * G								
100% NPK without Vinasse	Giza-429	640.97 ^{ef}	265.199^{h}	58.78 ^e	16.899^{f}	37.912^{h}	57.875 ^g	24.492°
		± 15.951	± 6.382	±5.092	± 4.601	± 9.734	±1.228	± 0.629
	Giza-843	953.77 bcdef	396.7 de ^{fgh}	$80.754c^{de}$	26.047 ^{def}	52.085 ^{efgh}	89.423d ^{efg}	33.647 ^{cde}
		± 23.837	± 9.808	±2.284	±7.544	± 13.083	± 2.353	± 0.845
	Misr-2	880.53 ^{cdef}	365.644 ^{fgh}	78.943 ^{cde}	24.472 ^{def}	50.917^{fgh}	84.744 ^{efg}	32.893 ^{cde}
		±9.664	±4.065	± 9.162	±3.868	±5.91	± 1.173	± 0.382
	Sakha-3	1139.71 ^{abcd}	474.877 ^{def}	94.832 ^{bcd}	31.498^{cde}	61.166 ^{defg}	$108.783^{cde} f \pm 0.488$	39.513 ^{bcd}
		±4.693	± 1.833	±2.952	±1.911	± 1.904		± 0.123
50% NPK with Vinasse	Giza-429	562.24^{f}	387.656 ^{efgh}	90.408 ^{bcde}	22.54 ^{ef}	69.856 ^{cdef}	76.482 ^{efg}	38.401 ^{bcd}
		±7.458	±5.342	±3.642	±3.703	± 10.541	± 1.68	± 0.579
	Giza-843	687.61 ^{def}	473.45 ^{def}	102.429^{bc}	26.756d ^{ef}	79.144 ^{bcd}	91.771 ^{defg}	43.507 ^b
		± 10.655	± 6.874	±4.054	± 4.313	± 10.859	± 1.166	$c \pm 0.597$
	Misr-2	631.23 ^{ef}	433.646d ^{efg}	97.119 ^{bcd}	25.705 ^{def}	75.042 ^{cd}	84.426 ^{efg}	41.252^{bcd}
		±8.386	±5.647	± 3.329	±3.854	± 10.299	± 1.148	± 0.566
	Sakha-3	905.86 ^{cdef}	627.268 ^{bc}	134.549^{a}	37.082 ^{cd}	103.963^{a}	124.251 ^{cd}	57.151^{a}
		±4.807	±3.69	±7.695	±4.771	± 5.946	± 0.694	± 0.327
75% NPK with Vinasse	Giza-429	1037.79abcde	508.45 ^{cde}	101.358^{bc}	40.408 ^{bc}	74.031 ^{cde}	131.585 ^{bc}	42.845 ^{bc}
		±4.497	±2.267	±4.534	± 3.555	±3.312	± 0.895	± 0.192
	Giza-843	1453.89^{a}	737.525 ^{ab}	135.499^{a}	59.483 ^a	98.967^{ab}	183.251 ^a	57.277^{a}
		± 2.803	± 1.293	± 1.953	± 2.658	± 1.427	± 0.412	± 0.083
	Misr-2	1273.95 ^{abc}	641.771 ^{abc}	122.383 ^{ab}	52.411 ^{ab}	89.387 ^{abc}	161.877 ^{ab}	51.732 ^{ab}
		oc1.1±	160.6±	±0./09	±0/.c±	±4.900	4C0.U±	±0.20/
	Sakha-3	1496.45^{a}	768.901 ^a	138.611 ^a	64.174 ^a	101.24^{m}	195.628 ^a	58.592ª
		土 4. 1 1	C17:17	±-2.244	±1.014	4C0.1 ±	土 U.470	CKU.U∓

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Fertilizer Treatment (T)		Protein Yield kg h ⁻¹	NRE	NUE	PKE	PUE	KRE	KUE
100% NPK with Vinasse	Giza-429	833.19 ^{cdef} ±6.213	312.873 ^{gh} ±2.224	65.72 ^{de} ±4.037	23.906 ^{def} ±1.809	46.605 ^{gh} ± 2.863	75.187 ^{fg} ±0.371	27.81 ^{de} ±0.171
	Giza-843	1227.49 ^{abc} ±25.861	461.167 ^{de} ^f ±9.545	92.426 ^{bcd} ±9.169	34.717 ^{cde} ±6.821	65.544d ^{efg} ± 13.594	113.56c ^{de} ±2.274	39.111b ^{cd} ±0.811
	Misr-2	1170.53 ^{abc} ±19.379	449.39 ^{defg} ±7.329	91.345 ^{bcd} ±5.584	33.959 ^{cde} ±6.177	64.776d ^{efg} ±11.052	$109.234^{cde}f \pm 1.679$	38.653b ^{cd} ±0.659
	Sakha-3	1384.35 ^{ab} ±3.281	533.793 ^{cd} ±1.24	101.61 ^{bc} ±1.587	40.67 ^{bc} ±0.859	72.056 ^{cdef} ±1.126	130.564 ^{bc} ±0.208	42.997 ^{bc} ±0.067
P fertilizer		0.001	0.000	0.004	0.000	0.00	0.008	0.00
P genotypes		0.000	0.000	0.008	0.000	0.002	0.001	0.007
PT * G		0.000	0.002	0.015	0.001	0.005	0.004	0.012

Table 6 (continued)

the averages of the main effects, while lowercase letters indicate the interaction

to reduce some salt stress adverse effects. Another indicator of stress tolerance in plants is the chlorophyll stability index (CSI); the higher its value, the lower the stress effect on plants' chlorophyll content. This leads to a more photosynthetic rate and dry matter production CSI is also used to screen genotypes for abiotic stress (Mohan et al. 2000; Kakar et al. 2019). Some other salinity tolerance lab screening indicators are total chlorophyll content, phenols, proline, percentage MSI, and osmotic potential (Deivanai et al. 2010; Ebaid et al. 2019).

All root traits (Table 4) positively responded to the application of both vinasse +75% or +100% NPK, in addition to full 100% NPK. Vinasse contains N, P, K, Fe, Mn, Cu, Zn, Mo, and organic C (Clementson et al. 2016), these nutrients improve root traits (Singh Gahoonia and Nielsen 2004; Gahoonia et al. 2006; Khalil 2019), and plant growth promoters (Indole-3-butyric acid, Gibberellic Acid and naphthalene acetic acid) which promote root growth, expansion of root hairs, and cotyledon cells (Chhun et al. 2004; Elmongy et al. 2018). Vinasse also has a positive impact upon per plant total nodule number, active ones, nodule dry weight, and nitrogenase activity, this in turn enhances atmospheric N fixation efficiency and nutrient uptake (Rafique et al. 2021), and by improving root traits, total plant growth is enhanced (Elmongy et al. 2018; Mao et al. 2018). Vinasse also contributes P which affects main root length and its dry weight, total surface area, total root tips and forks (Ramtekey et al. 2021). Phosphorus also stimulates N fixation efficiency, nitrogenase activity, and N uptake via improving active nodule number and dry weight (Li et al. 2022). Nodule number formed on faba bean roots are much initiated by applied mineral P fertilizer relative to the control (Otieno et al. 2009, Mohamed et al. 2021).

Faba bean cultivar plant height and seed yield components and yield were also varied by the contribution of vinasse to the applied NPK fertilizer, especially by the reduction of 25% of the recommended NPK or when added with the full amount (Table 5). The treatment of vinasse +75%NPK showed higher growth, physiological and biochemical performance. This may be attributed to the high dose of nitrogen for the treatment (vinasse + 100% NPK) negatively affecting the nitrogen fixation process in the root nodules compared to vinasse + 75% NPK as shown in Table 4. The reduction in soil pH and the availability of soil nutrients may partially explain the role added organic fertilizer played in maintaining relatively higher seed yield and yield components. Vinasse also contains plant growth promotors (Togay et al. 2008; Rafique et al. 2021). Similarly, the role of P, N and K elements in the photosynthesis process, and sinksource relationship may lead to increasing seed yield and its components (Togay et al. 2008; Nget et al. 2022; Yang et al. 2022). The legume crops have more requirement for P for optimal N-fixation compared to cereals for it has a fundamental role in nodule energetic transformations (Husssien **Fig. 2** Seed yield response index (SYRI) of the tested faba bean genotypes fertilized by nitrogen (N), phosphors (P), and potassium (K) with vinasse bundle at a rate of 52.6 N; 74.2 P and 124.3 K (kg h⁻¹). The purple lines represent N fertilizer while red lines P fertilizer likewise green lines K fertilizer. Efficient and responsive (ER); efficient and non-responsive (ENR); neither efficient but responsive (NER) and neither efficient nor responsive (NENR)



et al. 2020). Phosphorus leads to maximum yield and yield attributes (Gidago et al. 2011; Ali et al. 2020). Phosphorus supplement in legumes has great potential for promoting growth and yield and its attributes (Ndakidemi et al. 2011; El-Hady et al. 2022). According to (Hamer et al. 2009), soil microorganisms may be triggered by the supplied organic substrates (vinasses and molasses), thereby, enhancing the mineralization of nutrients. The ratios K^+ /Na⁺ and Ca²⁺/ Na⁺ in faba bean straw and seed decreased by increasing salinity levels, and reached their lowest values at the severe salinity (Afzal et al. 2022). The K⁺/Na⁺ ratio is a fundamental trait that signals salinity stress resistance in plants, and it may be used as a screening tool for plant breeders (Oyiga et al. 2018). Under salt stress, Na⁺ and K⁺ transporters are critical for maintaining Na⁺ and K⁺ homeostasis in cells and plants (Azhar et al. 2017; Ebaid et al. 2019).

N accumulated in faba bean seed has been reported in different studies, most studies report that at least 50% of total plant N is found in the seed by maturity, and typically proportions in the seed are higher. For example, (Dinesh et al. 2010) reported that faba bean seed held over 50% and 78% of plant N by maturity, respectively, (Tarek and Kh A Mohamed 2020), reported that faba bean seed contained 2/3 of the total plant N. Hossain et al. (Hossain et al. 2016) attributed the higher seed N uptake by faba bean to greater seed yield and greater ability to fix N₂ compared to the other pulses in the study. Nitrogen and phosphorus are considered the first and second most influential nutrients in crop growth, respectively (Anglade et al. 2015; Li et al. 2010; Papakaloudis and Dordas 2023). Therefore, measurements to quantify N and P removal by a crop are essential to fully assess the influence of any crop, including the faba bean on soil fertility and nutrient requirements in a sustainable cropping system. It is likely that raising the potassium rate of the soil fertilizer boosts nutrient uptake,

allowing for greater nutrient assimilation, resulting in larger potassium content in faba bean seed (Shaban et al. 2013; Taha et al. 2016; Shawer 2019).

The four faba bean cultivars showed varied potentials for absorbing and utilizing available N, P, and K. The cv. Saka 3 was the only cultivar that had a higher seed yield than the total mean seed yield in response to applied NPK levels. Both Misr 2 and Giza 843 have higher SYRI values than the average (Fig. 2) might be explained based on their genetic potential response to NPK. By applying 52.6 kg N, 74.2 kg P, and 124.3 kg K, cv. Misr 2 had SYRI values of 84.4, 54.4, and 35.5 kg seeds for every kg of N, P, and K h⁻¹, and cv. Giza 843 had SYRI values of 80.3, 51.8, and 33.8 kg seeds for every kg of N, P, and K h⁻¹. These are relative to the mean SYRI of 65.6; 42.3 and 27.6 kg seeds. SYRI varied among some faba bean genotypes (Liu et al. 2022) and among some faba bean ones that refer to their better genetic potency to exploit and utilize NPK in soil and the exogenous supply.

5 Conclusion

In this study, vinasse in combination with mineral NPK were utilized as fertilizers to improve faba bean plant growth. The rate of 75% NPK/vinasse exhibited the highest performance compared to the other's 50% or 100% NPK rates particularly to cultivars Giza 843 and Saka 3. All three vinasse-NPK fertilizer rates x cultivar interactions enhanced the trait means in comparison with full NPK rate. These significant variations cannot be related back only to the action vinasse, yet to the combination of the organic/mineral fertilizer. A futuristic cost–benefit study needs be conducted to evaluate performance when deducting a specific percentage of or applying full NPK when mixed with vinasse.

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Data availability All data of this study are available in the article. Source data are available from the corresponding author upon reasonable request.

Declarations

Competing interest The authors declare no competing interests.

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References

- Abdellatif KF, El Sayed A, Zakaria AM (2012) Drought stress tolerance of faba bean as studied by morphological traits and seed storage protein pattern. J Plant Stud 1:47. https://doi.org/10.5539/ jps.v1n2p47
- Abou Hussien E, Nada W, Elgezery MK (2017) Evaluation efficiency of sulphur fertilizer in calcareous soil amended by compost. Menoufia J Soil Sci 2:95–72. https://doi.org/10.21608/mjss.2017. 175900
- Afzal M, Alghamdi SS, Migdadi HH, El-Harty E, Al-Faifi SA (2022) Agronomical and physiological responses of faba bean genotypes to salt stress. Agric 12:235. https://doi.org/10.3390/agriculture1202 0235
- Ali MAA, Abdallah MM, Abo El-Azam NA, El-Yazeid A, Ahmed A (2019) Impact of salinity Seed sprout characterization of five faba bean (*Vicia faba L*) varieties. Arab Univ J Agric Sci 27:2259– 2272. https://doi.org/10.21608/AJS.2019.15226.1064
- Ali A, Asif M, Adnan M et al (2020) Effect of different levels of phosphorus on growth, yield and quality of wheat (*Triticum aestivum* L.). Int J Botany Stud 5:64–68
- Allen RG, Pereira LS, Raes D, Smith M (1998) Crop evapotranspiration-Guidelines for computing crop water requirements-FAO Irrigation and drainage paper 56. Fao, Rome 300:D05109. https:// doi.org/10.1061/(ASCE)0733-9437(2005)131:1(2)
- Allen RG, Pereira LS, Smith M, Raes D, Wright JL (2005) FAO-56 dual crop coefficient method for estimating evaporation from soil and application extensions. J Irrig Drain Eng 131:2–13. https:// doi.org/10.1061/(ASCE)0733-9437(2005)131:1(2)
- Anglade J, Billen G, Garnier J (2015) Relationships for estimating N2 fixation in legumes: incidence for N balance of legume-based cropping systems in Europe. Ecosphere 6:1–24. https://doi.org/ 10.1890/ES14-00353.1
- AOAC (1975) Official methods of analysis, 12th edn. Association of Official Analytical Chemists, Washington DC
- Atwa A, Abo Mostafa R, El-Basuny AA (2008) Impact of irrigation water salinity levels on soil chemical properties and some faba

bean varieties. J Soil Sci Agric Eng 33:2447–2457. https://doi. org/10.21608/JSSAE.2008.199795

- Azhar N, Su N, Shabala L, Shabala S (2017) Exogenously applied 24-epibrassinolide (EBL) ameliorates detrimental effects of salinity by reducing K+ efflux via depolarization-activated K+ channels. Plant Cell Physiol 58:802–810. https://doi.org/10.1093/pcp/ pcx026
- Bartlett MS (1937) Properties of sufficiency and statistical tests. Proceedings of the Royal Society of London. Ser A-Math Phys Sci 160:268–282
- Burns R, Holsten R (1973) Applications of the acetylene-ethylene assay for measurement of nitrogen fixation. Soil Biol Biochem 5:47–81. https://doi.org/10.1016/0038-0717%2873%2990093-X
- Carstensen A, Herdean A, Schmidt SB et al (2018) The impacts of phosphorus deficiency on the photosynthetic electron transport chain. Plant Physiol 177:271–284. https://doi.org/10.1104/pp.17.01624
- Chhun T, Taketa S, Tsurumi S, Ichii M (2004) Different behaviour of indole-3-acetic acid and indole-3-butyric acid in stimulating lateral root development in rice (Oryza sativa L.). Plant Growth Regul 43:135
- Clementson C, Abrahim BN, Homenauth O, Persaud V (2016) An Evaluation of 'Vinasse' (Bio-Ethanol Effluent) and Vermicompost as Soil Amendments for Cash Crop Production. Greener J Agric Sci 6:256–261
- Cottenie A, Verlo M, Kjekens L, Camerlynch R (1982) Chemical analysis of plant and soil. Laboratory of analytical agrochemistry. State University, Gent, Belgium, article no. 42, pp 80–284
- de Chaves MG, Venturini AM, Merloti LF et al (2021) Combined Use of Vinasse and Nitrogen as Fertilizers Affects Nitrification, Ammonification, and Denitrification by Prokaryotes. Front Soil Sci 1:746745. https://doi.org/10.3389/fsoil.2021.746745
- Deivanai S, Devi SS, Rengeswari PS (2010) Physiochemical traits as potential indicators for determining drought tolerance during active tillering stage in rice (Oryza sativa L.). Pertanika J Trop Agric Sci 33(1)
- Desoky E-SM, Mansour E, Ali MM et al (2021) Exogenously used 24-epibrassinolide promotes drought tolerance in maize hybrids by improving plant and water productivity in an arid environment. Plants 10:354. https://doi.org/10.3390/plants10020354
- Diacono M, Montemurro F (2011) Long-term effects of organic amendments on soil fertility. Sustain Agric 2:761–786
- Dinesh R, Srinivasan V, Hamza S, Manjusha A (2010) Short-term incorporation of organic manures and biofertilizers influences biochemical and microbial characteristics of soils under an annual crop [Turmeric (*Curcuma longa L.*)]. Bioresour Technol 101:4697–4702. https://doi.org/10.1016/j.biortech.2010.01.108
- Ebaid M, Nawar AI, Sanaa IM, Barakat M, Ibrahim O (2019) Agronomic and physiological evaluation of Egyptian wheat cultivars under salinity stress. Middle East J Agric Res 8:1361–1370. https://doi.org/10.36632/mejar/2019.8.4.36
- El-Hady A, Mohamed A, Abd-Elkrem YM et al (2022) Impact on plant productivity under low fertility sandy soil in arid environment by revitalization of lentil roots. Front Plant Sci 13:937073. https:// doi.org/10.3389/fpls.2022.937073
- Elmongy MS, Cao Y, Zhou H, Xia Y (2018) Root development enhanced by using indole-3-butyric acid and naphthalene acetic acid and associated biochemical changes of in vitro Azalea microshoots. J Plant Growth Regul 37:813–825. https://doi.org/ 10.1007/s00344-017-9776-5
- Elsherpiny MA (2023) Maximizing faba bean tolerance to soil salinity stress using gypsum, compost and selenium. Egypt J Soil Sci 63:243–253. https://doi.org/10.21608/ejss.2023.203083.1582
- FAO (2020) World food and agriculture—statistical yearbook 2020. World Food and Agriculture-Statistical Yearbook. https://doi.org/ 10.4060/cb1329en

- FAO (2022) WORLD FOOD AND AGRICULTURE STATISTICAL YEARBOOK 2022. FAO. https://doi.org/10.4060/cc2211en
- Filipović L, Romić D, Ondrašek G, Mustać I, Filipović V (2020) The effects of irrigation water salinity level on faba bean (Vicia faba L.) Productivity. J Cent Eur Agric 21:537–542. https://doi.org/10. 5513/JCEA01/21.3.2872
- Frydenvang J, van Maarschalkerweerd M, Carstensen A et al (2015) Sensitive detection of phosphorus deficiency in plants using chlorophyll a fluorescence. Plant Physiol 169:353–361. https://doi.org/ 10.1104/pp.15.00823
- Gahoonia TS, Ali O, Sarker A, Nielsen NE, Rahman MM (2006) Genetic variation in root traits and nutrient acquisition of lentil genotypes. J Plant Nutr 29:643–655. https://doi.org/10.1080/ 01904160600564378
- Gidago G, Beyene S, Worku W, Sodo E (2011) The response of haricot bean (*Phaseolus vulgaris L.*) to phosphorus application on Ultisols at Areka, Southern Ethiopia. J Biol Agric Healthc 1:38–49
- Hamer U, Potthast K, Makeschin F (2009) Urea fertilisation affected soil organic matter dynamics and microbial community structure in pasture soils of Southern Ecuador. App Soil Ecol 43:226–233. https://doi.org/10.1016/j.apsoil.2009.08.001
- Hera C (1995) The role of inorganic fertilizers and their management practices. Fertil Res 43:63–81
- Hoarau J, Caro Y, Grondin I, Petit T (2018) Sugarcane vinasse processing: Toward a status shift from waste to valuable resource. A Review. J Water Process Eng 24:11–25
- Hossain Z, Wang X, Hamel C, Knight JD, Morrison MJ, Gan Y (2016) Biological nitrogen fixation by pulse crops on semiarid Canadian prairies. Canadian J Plant Sci 97:119–131. https://doi.org/10. 1139/cjps-2016-0185
- Husssien EA, Zemrany HE, Hammad M (2020) Effect of vinasses, molasses and mineral fertilization on nodulation and growth of common bean (phaseolus vulgaris L.) grown in sandy reclaimed soils. Menoufia J Soil Scie 5:1–17. https://doi.org/10.21608/mjss. 2020.169516
- Jiang Z-P, Li Y-R, Wei G-P et al (2012) Effect of long-term vinasse application on physico-chemical properties of sugarcane field soils. Sugar Tech 14:412–417. https://doi.org/10.1007/ s12355-012-0174-9
- Jiang G, Zhang W, Xu M et al (2018) Manure and mineral fertilizer effects on crop yield and soil carbon sequestration: a meta-analysis and modeling across China. Glob Biogeochem Cycles 32:1659– 1672. https://doi.org/10.1029/2018GB005960
- Kacar B (1972) Plant and soil chemical analysis-II. Plant Analysis, Ankara Univ. Agriculture Fac., Publication No, 453, Ankara
- Kakar N, Jumaa SH, Redoña ED, Warburton ML, Reddy KR (2019) Evaluating rice for salinity using pot-culture provides a systematic tolerance assessment at the seedling stage. Rice 12:1–14. https:// doi.org/10.1186/s12284-019-0317-7
- Katakojwala R, Kumar AN, Chakraborty D, Mohan SV (2019) 3 valorization of sugarcane waste: prospects of a biorefinery. In: Industrial and municipal sludge emerging concerns and scope for resource recovery, pp 47–60. Elsevier. https://doi.org/10.1016/ B978-0-12-815907-1.00003-9
- Keuter A, Hoeft I, Veldkamp E, Corre MD (2013) Nitrogen response efficiency of a managed and phytodiverse temperate grassland. Plant Soil 364:193–206. https://doi.org/10.1007/ s11104-012-1344-y
- Khalil M (2019) Effect of mineral fertilizers and biofertilization on some soil properties and faba bean productivity under saline soil conditions. J Soil Sci Agric Eng 10:889–897. https://doi.org/10. 21608/jssae.2019.79692
- Ladha JK, Peoples MB, Reddy PM et al (2022) Biological nitrogen fixation and prospects for ecological intensification in cereal-based cropping systems. Field Crops Res 283:108541. https://doi.org/ 10.1016/j.fcr.2022.108541

- Li H, Shen J, Zhang F, Marschner P, Cawthray G, Rengel Z (2010) Phosphorus uptake and rhizosphere properties of intercropped and monocropped maize, faba bean, and white lupin in acidic soil. Biol Fert Soils 46:79–91. https://doi.org/10.1007/s00374-009-0411-x
- Li H, Wang L, Zhang Z, Yang A, Liu D (2022) Effect of phosphorus supply levels on nodule nitrogen fixation and nitrogen accumulation in soybean (Glycine max L.). Agron 12:2802. https://doi.org/ 10.3390/agronomy12112802
- Lichtenthaler HK, Wellburn AR (1983) Determinations of total carotenoids and chlorophylls a and b of leaf extracts in different solvents. Portland Press Ltd. https://doi.org/10.1042/bst0110591
- Liu Z, Xing Y, Jin D et al (2022) Improved Nitrogen Utilization of Faba Bean (Vicia faba L.) Roots and Plant Physiological Characteristics under the Combined Application of Organic and Inorganic Fertilizers. Agric 12:1999. https://doi.org/10.3390/agriculture12121999
- Mahdi A (2016) Improvement of salt tolerance in)Vicia faba L.) plants by exogenous application of polyamines. Egypt J Agron 38:1–21. https://doi.org/10.21608/agro.2016.296
- Mao J-P, Zhang D, Zhang X et al (2018) Effect of exogenous indole-3-butanoic acid (IBA) application on the morphology, hormone status, and gene expression of developing lateral roots in Malus hupehensis. Sci Hortic 232:112–120. https://doi.org/10.1016/j. scienta.2017.12.013
- Martineau-Côté D, Achouri A, Karboune S, L'Hocine L (2022) Faba bean: an untapped source of quality plant proteins and bioactives. Nutr 14:1541. https://doi.org/10.3390/nu14081541
- Mohamed HI, El-Sayed AA, Rady MM, Caruso G, Sekara A, Abdelhamid MT (2021) Coupling effects of phosphorus fertilization source and rate on growth and ion accumulation of common bean under salinity stress. PeerJ 9:e11463
- Mohan M, Narayanan SL, Ibrahim S (2000) Chlorophyll stability index (CSI): its impact on salt tolerance in rice. Inter Rice Res Notes 25:38–39
- Ndakidemi PA, Bambara S, Makoi JH (2011) Micronutrient uptake in common bean ("*Phaseolus vulgaris*" L.) as affected by Rhizobium inoculation, and the supply of molybdenum and lime. Plant Omics 4:40–52
- Nget R, Aguilar EA, Cruz PCS et al (2022) Responses of soybean genotypes to different nitrogen and phosphorus sources: impacts on yield components, seed yield, and seed protein. Plants 11:298. https://doi.org/10.3390/plants11030298
- Ning C-c, Gao P-d, Wang B-q, Lin W-p, N-h J, K-z CAI (2017) Impacts of chemical fertilizer reduction and organic amendments supplementation on soil nutrient, enzyme activity and heavy metal content. J Integr Agric 16:1819–1831. https://doi.org/10.1016/ S2095-3119(16)61476-4
- Oldroyd GE, Dixon R (2014) Biotechnological solutions to the nitrogen problem. Curr Opin Biotechnol 26:19–24. https://doi.org/10. 1016/j.copbio.2013.08.006
- Otieno P, Muthomi J, Chemining'wa G, Nderitu J (2009) Effect of rhizobia inoculation, farm yard manure and nitrogen fertilizer on nodulation and yield of food grain legumes. J Biol Sci 4:326–332. https://doi.org/10.3923/jbs.2009.326.332
- Oyiga BC, Sharma RC, Baum M, Ogbonnaya FC, Léon J, Ballvora A (2018) Allelic variations and differential expressions detected at quantitative trait loci for salt stress tolerance in wheat. Plant Cell Environ 41:919–935. https://doi.org/10.1111/pce.12898
- Papakaloudis P, Dordas C (2023) Phosphorus Fertilization Affects Morphological, Physiological and Agronomic Characteristics of Faba Bean Cultivars. Sustain 15:13172. https://doi.org/10.3390/ su151713172
- Prado RdM, Caione G, Campos CNS (2013) Filter cake and vinasse as fertilizers contributing to conservation agriculture. App Environm Soil Sci 2013:1–8. https://doi.org/10.1155/2013/581984
- Premachandra GS, Saneoka H, Ogata S (1990) Cell membrane stability, an indicator of drought tolerance, as affected by applied

nitrogen in soyabean. J Agric Sci 115:63–66. https://doi.org/10. 1017/S0021859600073925

- Rafique M, Naveed M, Mustafa A et al (2021) The combined effects of gibberellic acid and rhizobium on growth, yield and nutritional status in chickpea (Cicer arietinum L.). Agron 11:105. https://doi. org/10.3390/agronomy11010105
- Ramtekey V, Bansal R, Aski MS et al (2021) Genetic variation for traits related to phosphorus use efficiency in lens species at the seedling stage. Plants 10:2711. https://doi.org/10.3390/plants10122711
- Raza A, Charagh S, Abbas S et al (2023) Assessment of proline function in higher plants under extreme temperatures. Plant Biol 25:379–395. https://doi.org/10.1111/plb.13510
- Semba RD, Ramsing R, Rahman N, Kraemer K, Bloem MW (2021) Legumes as a sustainable source of protein in human diets. Global Food Secur 28:100520. https://doi.org/10.1016/j.gfs.2021.100520
- Shaban KA, Khalil A, Mohamed AA (2013) Effect of sowing date and nitrogen, potassium fertilization on faba bean productivity in newly reclaimed saline soil of North Sinai. J Soil Sci Agric Eng 4:893–904. https://doi.org/10.21608/JSSAE.2013.52492
- Shahidi F, Naczk M (1995) Food phenolics: sources, chemistry, effects and applications. Technomic Publishing Co., Lancaster
- Shapiro SS, Wilk MB (1965) An analysis of variance test for normality (complete samples). Biometrika 52:591–611. https://doi.org/10. 2307/2333709
- Shawer S (2019) Effect of potassium fertilization and organic materials on some characteristic growth and nutrients uptake by faba bean (*Vicia faba L.*) plant. Plant Arch 19:732–737
- Singh Gahoonia T, Nielsen NE (2004) Root traits as tools for creating phosphorus efficient crop varieties. Plant Soil 260:47–57
- Soobadar A, Ng Kee Kwong KR (2012) Impact of high rates of vinasse on some pertinent soil characteristics and on sugarcane yield in

Mauritius. J Sustain Agric 36:36–53. https://doi.org/10.1080/ 10440046.2011.620226

- Taha A, Omar M, Khedr H (2016) Effect of different sources and levels of potassium on growth, yield and chemical composition of faba bean plants. J Soil Scie Agric Eng 7:243–248. https://doi.org/10.21608/ JSSAE.2016.39402
- Tarek S, Kh A Mohamed M (2020) Evaluation of Some Faba Bean Genotypes Under Three Planting Dates in Middle Egypt. Alex J Agric Sci 65:201–209. https://doi.org/10.21608/ALEXJA.2020. 109679
- Togay Y, Togay N, Dogan Y (2008) Research on the effect of phosphorus and molybdenum applications on the yield and yield parameters in lentil (Lens culinaris Medic.). Afr J Biotechnol 9:1256–1260
- Troll W, Lindsley J (1955) A photometric method for the determination of proline. J Biol Chem 215:655–660. https://doi.org/10. 1016/S0021-9258(18)65988-5
- Udvardi M, Poole PS (2013) Transport and metabolism in legumerhizobia symbioses. Annu Rev Plant Bio 64:781–805. https://doi. org/10.1146/annurev-arplant-050312-120235
- Yadav GS, Babu S, Meena RS et al (2017) Effects of godawariphosgold and single supper phosphate on groundnut (Arachis hypogaea) productivity, phosphorus uptake, phosphorus use efficiency and economics. Indian J Agric Sci 87:1165–1169. https://doi.org/10. 56093/ijas.v87i9.74162
- Yang J-X, Richards RA, Jin Y, He J (2022) Both biomass accumulation and harvest index drive the yield improvements in soybean at high and low phosphorus in south-west China. Field Crops Res 277:108426. https://doi.org/10.1016/j.fcr.2021.108426

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