



Combined Effect of Prickly Pear Waste Biochar and Azolla on Soil Fertility, Growth, and Yield of Roselle (*Hibiscus sabdariffa* L.) Plants

Hassan M. Al-Sayed¹ · Ahmed M. Ali¹ · Mahmoud A. Mohamed² · Mostafa F. Ibrahim³

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Abstract

Although the use of biochar to promote plant growth has been reported by many researchers, the combined effect of prickly pear waste biochar (BC) and Azolla (AZ) in a field experiment on the roselle plants did yet receive attention. Therefore, the study aims to evaluate the effect of biochar and Azolla extract on the growth, production, and quality of roselle plants. The experiment treatments were in a completely randomized block design with three replicates. Biochar was added at rates of 0, 10, and 20 ton ha⁻¹ and AZ was applied at rates of 0, 3, and 6% in addition to a control treatment. Biochar added at high rate (20 ton ha⁻¹) significantly increased the fresh and dry weights of sepals by 27.98 and 35.73%, respectively, compared to the control. The corresponding values were significantly increased by 11.89 and 11.85% over the control when Azolla was added at rate of 6%. The interaction effect of both BC and AZ treatments at high rate significantly increased the fresh and dry weight of sepals by 47.16 and 60.59%, respectively, compared to the control. The interaction effect of BC and AZ realized significant effect on soil properties, growth and yield, as well as pigments of roselle plants. This is a good evident means that BC and AZ applications separately or combined are considered promising materials for sustainable organic agriculture and safety food.

Keywords Biochar · Azolla · SPAD · Roselle · Plant growth

1 Introduction

Roselle (*Hibiscus sabdariffa* L.) plant is one of the most important medicinal and nutritional plants native to Africa and consumed worldwide due to its high value and access to international markets (Sanders et al. 2020). In addition, roselle plant is rich in anthocyanins, organic acids, pectin, phenolic compounds, and vitamins. It is considered an ideal plant in the developed countries like Egypt, and it is drought resistant, relatively easy to cultivate but it requires a lot of labor to deal with due to the difficulty of using mechanization for harvesting (Al-Sayed et al. 2020; Alam et al. 2016).

Nowadays, a great attention is devoted to biochar which is a carbon-rich material formed by thermo-chemically converting plant biomass in an oxygen-deficient environment (McGlashan et al. 2012). It is an important recycling strategy in sustainable development that allows agricultural wastes to be converted into fertilizers or as soil conditioners that improve its properties and fertility (Rekaby et al. 2021; Tenic et al. 2020). Biochar application increases nutrient content, enhances cation exchange capacity, and improves soil structure, diversity of micro-organisms, and ensuring environmental sustainability (Qayyum et al. 2020; Solaiman et al. 2020). Furthermore, plant photosynthetic rate, chlorophyll content, and stomatal conductance were improved by biochar addition (Akhtar et al. 2014; Batool et al. 2015). Consequently, it enhances growth parameters, seed germination, shoot and root lengths, nutrient contents, and crop yield (Ma et al. 2019; Nobile et al. 2020).

On the other hand, biochar has few negative effects such as those related to its high salt content and high acidity, which sometimes leads to undesirable changes especially in the alkaline soil. Also, it has been reported that biochar produced with relatively high temperatures (600–700 °C)

✉ Hassan M. Al-Sayed
HassanMohamed.4419@azhar.edu.eg

¹ Department of Soils and Water, Faculty of Agriculture, Al-Azhar University (Assiut Branch), Assiut 71524, Egypt
² Department of Soil Fertility and Plant Nutrition, Soil, Water and Environment Research Institute, Agricultural Research Center, Giza 12619, Egypt
³ Horticulture Department, Faculty of Agriculture, Al-Azhar University (Assiut Branch), Assiut 71524, Egypt

leads to high proportions of aromatic C and low proportions of hydrogen (H) and oxygen (O) functional groups and consequently low cation exchange capacity (Lehmann and Joseph 2009; Novak et al. 2009).

Azolla is considered one of the most promising bio fertilizers since it has the ability to fix about 30–60% kg N ha⁻¹ from atmospheric nitrogen that could replace 25% of nitrogen mineral fertilization (Maswada et al. 2021; Malyan et al. 2019; Kollah et al. 2016). Azolla decomposes through 8–10 days and it releases its N content into soil solution to be available for plant uptakes (Yadav et al. 2014). It has been widely used as a cheap green amendments or bio-fertilizer to supply plants with their N requirements (Abou Hussien et al. 2020; Al-Sayed et al. 2019). Azolla work for availability of macro nutrient that could be changed over time with an average of 8.3% K and 0.6% Mg as well as vitamins production (El-Serafy et al. 2021; Zhang et al. 2018).

Using Azolla or biochar individually or in conjunction with other organic materials enhances plant growth (Sharifi et al. 2019). Combining biochar and Azolla together increases rice yield and nitrogen use efficiency (NUE), reduces chemical fertilizer applications, avoids agricultural pollution, and provides less production costs (Kimani et al. 2021). So far, few studies were conducted to assess the effect of both Azolla and biochar on crop growth and production parameters. We hypothesized that combined both biochar and Azolla represent an important cultivation management option for sustainable agriculture.

Therefore, accordingly, this study aims to (1) evaluate the effect of using Azolla or biochar individual or in conjunction on calyces yield and growth characteristics of roselle plants and (2) to find out the suitable rate of biochar or Azolla that enhance soil properties and roselle plant production.

2 Materials and Methods

2.1 Experimental Site and Design

A field study was conducted during two successive summer seasons of 2020 and 2021 at a private farm named Hajer Al-Dabayah village southwest Luxor Governorate, Egypt, which is located at 25° 41' 28.18" N latitude and 32° 34' 09.62" E longitude. The meteorological data of the experimental site are monitored via the Central Lab of Agricultural Climate, Agricultural Research Center (ARC), Ministry of Agriculture and Land Reclamation, Giza, Egypt (Table 1). The physico-chemical properties of the soil used in the experiment are listed in Table 2. Roselle (Sobahia 17 dark variety) seeds were obtained from the ARC, Giza, Egypt, and were sown in the field on 26 April of both growing seasons (2020 and 2021). The experimental unit was 2.8 m in length × 1.2 m in width with an area of 3.36 m² containing

Table 2 Some physical and chemical properties of studied soil before two successive growing seasons (2020–2021)

Studied soil		
Properties	Before 1st season	Before 2nd season
Sand (g kg ⁻¹)	750	745
Silt (g kg ⁻¹)	170	174
Clay (g kg ⁻¹)	80	81
Texture	Sandy loam	Sandy loam
Soil reaction (pH)	7.98	7.99
Electrical conductivity (EC)	1.1	1.12
CaCO ₃ (g kg ⁻¹)	47	49
Organic matter (g kg ⁻¹)	8.2	8.5
Available-N (mg kg ⁻¹)	21	24
Available-P (mg kg ⁻¹)	6.2	6.6
Available-K (mg kg ⁻¹)	221	226

Each value represents a mean of three replicates

Table 1 Basic climatic data of the experimental site during the period of the study (April–October 2020–2021)

	T_{max} (C°)		T_{min} (C°)		Relative humidity (%)		Solar radiation (MJ/m ² /day)		Wind speed MS ⁻¹	
	1st	2nd	1st	2nd	1st	2nd	1st	2nd	1st	2nd
April	32.30	34.77	15.07	15.69	25.34	19.02	22.57	27.62	3.00	2.99
May	38.08	40.12	20.92	21.24	19.91	14.80	27.41	28.71	3.35	3.25
June	40.80	40.62	23.22	23.10	18.05	18.86	27.89	29.12	3.22	4.05
July	40.89	41.09	24.78	25.11	20.41	20.95	29.38	27.98	3.59	3.69
August	41.00	41.58	24.50	24.39	21.48	20.63	28.13	26.62	3.84	3.43
September	41.85	38.69	24.27	22.39	23.02	28.02	27.01	23.94	3.19	3.84
October	37.78	35.35	20.89	18.95	27.19	30.87	24.27	20.25	3.14	3.03

T_{max} : maximum temperature; T_{min} : minimum temperature; MJ/m², mega joules per square meter; MS⁻¹, meter/second

two rows 60 cm apart with three hills (40 cm apart), with a total density of 41,667 plants ha⁻¹. Fifteen days after sowing (DAS), the seedlings were thinned to one plant per hill. The experiment was laid in a randomized complete block with three replicates. The main plot was assigned to biochar (BC) application at rate of 0, 10, and 20 ton ha⁻¹ while azolla (AZ) solution was sprayed on plants at rate of 0, 3, and 6% as subplot.

In the early morning, the tested rates of Azolla solutions were sprayed at 50, 80, and 110 DAS for related plot. Distilled water was sprayed on plants in the same times in the control plot. All agriculture practices were performed according to the Egyptian Ministry of Agriculture.

2.2 Azolla Extract Preparation

Azolla (*Azolla pinnata* L.) plants were obtained from Soil, Water and Environment Research Institute, ARC, Giza. One kilogram of Azolla plants was soaked in 1 l of ethanol (90% conc.) for 24 h then mixed well with blender. The mixture was filtered twice through two layers of gauze cloth. The obtained solution was considered as 100% concentrate of Azolla plants extract. Three and 6 ml of this concentration were taken and diluted with 97 and 94 ml distilled water to obtain 3 and 6% concentrations, respectively; then, they were kept in the refrigerator at 4 °C until use. Tween 20 at 0.1% (v/v) was used as a surfactant according to Yasmeen et al. (2013). The characteristics of the Azolla solution are shown in Table 3.

2.3 The Preparation Procedure of Biochar

The prickly pear fruit wastes (its peels) were collected, air-dried after being cut into small pieces (less than 5 cm), and then oven-dried at 70 °C for 24 h. The raw material was pyrolyzed in a muffle furnace at 350 °C for 3 h in limited oxygen conditions. After that, it was passed through a 2-mm diameter

stainless steel sieve before mixing it with the soil. The properties of biochar are shown in Table 3.

2.4 Soil, Plant, Biochar, and Azolla Analysis

Some physical and chemical properties of the tested soils were determined according to Burt (2004). Soil texture was determined by the pipette as described by Page et al. (1982). The soil reaction was measured potentiometrically in soil (Page et al. 1982) using a digital pH meter (Hanna Instruments pH 211, Romania). The electrical conductivity (EC) was determined using the salt bridge by an EC meter (Jenway 4510 England) (Burt 2004). Calcium carbonate was determined according to Burt (2004). Available phosphorus (P) was measured according to the method describe by Olsen and Sommers (1982). Available potassium (K) was extracted by 1 N ammonium acetate solution measured by the flame photometer according to (Jackson 1973). Available nitrogen was extracted with 1% K₂SO₄ using a micro Kjeldahl's method (Jackson 1973). Soil organic matter (SOM) concentration was determined by oxidization with K₂Cr₂O₇ and H₂SO₄ (Jackson 1973).

Biochar and Azolla samples (2.0 g) were digested with H₂O₂ and H₂SO₄. The total N, P, and K concentrations were measured in the digest extract. To measure nutrient concentrations in roselle shoots, a mixture of 7:3 ratio of sulfuric to perchloric acids was used to digest the dried ground plant material. The total N, P, and K determined were described by Burt (2004). The nutrient uptake of N, P, and K was calculated by the following formula: (Total N, P, and K content × dry matter)/100. Chlorophyll contents from fully developed leaves were determined using a portable chlorophyll meter (SPAD-502-m Konica Minolta, Inc., Tokyo, Japan). Total anthocyanins (TAC) and total flavonoids (TF) were extracted by adding 10 ml (8.5 ml ethanol 96% + 1.5 ml HCl 1.5 M) to 1 g of dried sepals according to Lees and Francis (1971).

2.4.1 Relative Water Content (RWC)

To determine the relative water content (RWC) of ripe leaves at harvest, random leave samples from each treatment were weighed directly to calculate the fresh weight (FW) then soaked in water in test tube in the dark for 24 h. They were blotted dry with filter paper and weighed to calculate their turgid weight (TW). The leaves oven dried at 70 °C for 48 h to measure their dry weight (DW). The leaf RWC was estimated according to Smart and Bingham (1974) using the following equation:

$$RWC = \frac{FW - DW}{TW - DW} \times 100$$

Table 3 Chemical composition of the used Azolla and biochar

Property	Azolla	Biochar
Soil reaction (pH)	7.5	8.12
Electrical conductivity (EC)	1.98	5.14
Organic matter (g kg ⁻¹)	620	630
Total nitrogen (g kg ⁻¹)	34.00	23
Total phosphorous (g kg ⁻¹)	17.30	5.1
Total potassium(g kg ⁻¹)	22.00	11
Total Mg (g kg ⁻¹)	15.00	-
Total Mn (g kg ⁻¹)	1.40	-
Total Fe(g kg ⁻¹)	1.80	-

Each value represents a mean of three replicates

2.5 Data Analysis

The analysis of variance (two-way ANOVA) and Duncan's multiple range tests at 5% level of probability were performed to distinct the significant differences among the treatments. The statistical analyses were performed using Costat software (Steel and Torrie 1996).

3 Results

3.1 Soil Properties

The effect of biochar (BC) and Azolla (AZ) application on soil reaction (pH), soil salinity (EC), and soil organic matter (SOM) are shown in Table 4. Generally, all treatments realized significant improvement of soil properties compared to the control. On average basis of both seasons, a slight change in pH values as a result of adding the high biochar rate BC2 while it significantly increased the EC and SOM by about 21 and 78%, respectively, compared to the control treatment. The addition of AZ showed insignificant effect on soil properties. The effect of biochar alone and the combined biochar and Azolla increased EC values and SOM content more than 24 and 77%, respectively, compared to the control.

3.2 Nutrient Availability and Their Uptake

Nitrogen (N), phosphorous (P), and potassium (K) availability and their uptake were significantly ($P < 0.05$) increased due to adding BC and/or AZ in both seasons (Tables 5 and 6). On average basis of both seasons, the available N, P, and K increased about 41, 46, and 35%, respectively, as a result of adding the high rate of BC. Also, the uptake of N, P, and K were significantly ($P < 0.05$) increased by 100, 64, and 70%, respectively, due to applying the high rate of BC compared to the control. Regarding AZ addition, the N, P, and K availabilities were significantly ($P < 0.05$) increased almost 57, 8, and 0.52%, respectively, compared to the control. Also, their uptake were significantly ($P < 0.05$) increased 25, 21, and 24%, respectively, compared to the control. When addition BC with AZ sprayed on plants, the N, P, and K availability and their uptake were positively affected. N, P, and K availability increased about 52, 51, and 37%, respectively, while their uptake increased nearly 144, 102 and 109%, respectively, compared to the control.

3.3 Some Growth Parameters

The results showed that all treatments were significantly ($P < 0.05$) increased the growth parameters as biochar and Azolla rates increased and the increases were more

Table 4 Impact of different rates of biochar and or/Azolla on the physicochemical properties of soil analyzed after harvesting of the roselle plant

Azolla%	First season				Second season			
	BC ₀	BC ₁	BC ₂	Azolla mean B	BC ₀	BC ₁	BC ₂	Azolla mean B
pH								
0	8.13 ± 0.04d	8.18 ± 0.01bcd	8.2 ± 0.01ab	8.18A	8.10 ± 0.03c	8.25 ± 0.01b	8.30 ± 0.01ab	8.21A
3	8.13 ± 0.01 cd	8.17 ± 0.03bcd	8.24 ± 0.01ab	8.18A	8.08 ± 0.02c	8.25 ± 0.03b	8.31 ± 0.01a	8.21A
6	8.12 ± 0.01 cd	8.19 ± 0.01abc	8.25 ± 0.01a	8.19A	8.05 ± 0.02c	8.27 ± 0.01ab	8.33 ± 0.01a	8.22A
Biochar mean A	8.13C	8.18B	8.24A		8.08C	8.25B	8.31A	
EC (dS m⁻¹)								
0	0.66 ± 0.01d	0.70 ± 0.03 cd	0.77 ± 0.03ab	0.71A	0.65 ± 0.02d	0.75 ± 0.02bc	0.82 ± 0.07ab	0.74A
3	0.68 ± 0.03d	0.71 ± 0.02bcd	0.75 ± 0.03abc	0.71A	0.64 ± 0.03d	0.77 ± 0.03abc	0.83 ± 0.03ab	0.75A
6	0.68 ± 0.04d	0.73 ± 0.06abcd	0.78 ± 0.02a	0.73A	0.67 ± 0.02 cd	0.76 ± 0.01abc	0.85 ± 0.01a	0.76A
Biochar mean A	0.67C	0.71B	0.77A		0.66C	0.76B	0.84A	
OM (g kg⁻¹)								
0	6.42 ± 0.22c	9.56 ± 0.02b	10.95 ± 0.04a	8.97A	6.36 ± 0.21c	10.34 ± 0.03b	11.71 ± 0.06a	9.47A
3	6.38 ± 0.16c	9.57 ± 0.39b	10.98 ± 0.41a	8.98A	6.35 ± 0.09c	10.34 ± 0.31b	11.72 ± 0.46a	9.47A
6	6.37 ± 0.15c	9.57 ± 0.12b	10.99 ± 0.09a	8.97A	6.34 ± 0.23c	10.35 ± 0.07b	11.73 ± 0.04a	9.47A
Biochar mean A	6.39C	9.56B	10.97A		6.35C	10.34B	11.72A	

BC₀, BC₁, and BC₂, biochar at rates of (0, 10 and 20 t ha⁻¹); pH, soil reaction; EC, electrical conductivity; OM, organic matter. All values are the mean of three replicate analysis ± standard error. Means in each column followed by the same letters are not significantly different ($P < 0.05$) by Duncan's multiple range tests

Table 5 Impact of biochar and or/Azolla on the different nutrient availability in roselle plants

Azolla %	First season				Second season			
	BC ₀	BC ₁	BC ₂	Azolla mean B	BC ₀	BC ₁	BC ₂	Azolla mean B
N (mg kg⁻¹)								
0	28.50 ± 1.73d	33.70 ± 0.61 cd	39.60 ± 1.01 ab	33.93A	28.23 ± 1.55e	34.50 ± 0.55 cd	40.23 ± 1.00ab	34.32A
3	31.20 ± 0.70 cd	34.67 ± 1.88bc	41.33 ± 2.77a	35.73A	29.67 ± 0.81e	35.37 ± 1.88bc	41.98 ± 2.75a	35.67A
6	29.90 ± 1.97 cd	35.55 ± 1.90bc	42.87 ± 0.92a	36.11A	28.60 ± 2.06de	36.21 ± 1.92bc	43.48 ± 0.87a	36.09A
Biochar mean A	29.87C	34.64B	41.27A		28.83C	35.36B	41.89A	
P (mg kg⁻¹)								
0	6.61 ± 0.02c	7.26 ± 0.14c	9.010 ± 0.04ab	7.63B	6.27 ± 0.35d	7.88 ± 0.14c	9.66 ± 0.07ab	7.93B
3	6.66 ± 0.11c	8.46 ± 0.03b	9.47 ± 0.19a	8.20A	6.63 ± 0.08d	9.07 ± 0.03b	10.11 ± 0.16a	8.60A
6	6.77 ± 0.12c	8.47 ± 0.74b	9.46 ± 0.28a	8.23A	6.73 ± 0.15d	9.09 ± 0.71b	10.09 ± 0.24a	8.63A
Biochar mean A	6.68C	8.06B	9.32A		6.54C	8.68B	9.95A	
K (mg kg⁻¹)								
0	279.51 ± 3.36c	290.79 ± 1.42b	378.33 ± 4.06a	316.21A	279.16 ± 3.04c	292.01 ± 1.42b	379.54 ± 4.05a	316.90A
3	280.33 ± 2.03c	290.82 ± 4.54b	378.56 ± 10.78a	316.57A	280.31 ± 2.03c	292.15 ± 4.47b	379.81 ± 10.79a	317.42A
6	281.31 ± 1.22c	293.05 ± 2.08b	378.99 ± 8.10a	317.79A	281.25 ± 1.22c	294.32 ± 2.08b	380.24 ± 8.16a	318.60A
Biochar mean A	280.39C	291.56B	378.63A		280.24C	292.83B	379.86A	

BC₀, BC₁, and BC₂, biochar at rates of (0, 10, and 20 t ha⁻¹). All values are the mean of three replicate analysis ± standard error. Means in each column followed by the same letters are not significantly different ($P < 0.05$) by Duncan's multiple range tests

Table 6 Impact of biochar and/or Azolla on the different nutrient uptake in roselle plants

Azolla %	First season				Second season			
	BC ₀	BC ₁	BC ₂	Azolla mean B	BC ₀	BC ₁	BC ₂	Azolla mean B
N uptake (g kg⁻¹)								
0	7.52 ± 0.30e	10.97 ± 0.43d	14.11 ± 0.10bc	10.87C	7.16 ± 0.32f	11.94 ± 0.58d	16.13 ± 0.26b	11.75C
3	8.26 ± 0.47e	12.95 ± 0.41c	15.36 ± 0.12b	12.19B	8.20 ± 0.55ef	14.11 ± 1.17c	18.02 ± 0.63a	13.44B
6	8.79 ± 0.61e	14.55 ± 0.18b	17.00 ± 0.19a	13.44A	9.43 ± 0.86e	15.94 ± 0.43b	19.47 ± 1.16a	14.95A
Biochar mean A	8.19C	12.82B	15.49A		8.27C	14.00B	17.87A	
P uptake (g kg⁻¹)								
0	0.75 ± 0.02f	1.01 ± 0.04d	1.26 ± 0.04c	1.01C	0.69 ± 0.02f	1.02 ± 0.05d	1.23 ± 0.04bc	0.98C
3	0.83 ± 0.02ef	1.19 ± 0.04c	1.37 ± 0.02b	1.13B	0.86 ± 0.02e	1.18 ± 0.02c	1.32 ± 0.03b	1.12B
6	0.86 ± 0.04e	1.25 ± 0.04c	1.49 ± 0.02a	1.20A	0.89 ± 0.05e	1.28 ± 0.04bc	1.47 ± 0.03a	1.21A
Biochar mean A	0.81C	1.15B	1.38A		0.81C	1.16B	1.34A	
K uptake (g kg⁻¹)								
0	8.08 ± 0.04i	11.35 ± 0.32f	13.72 ± 0.96d	11.05C	8.68 ± 0.43 g	12.15 ± 0.21de	14.50 ± 0.97bc	11.78C
3	8.91 ± 0.53 h	12.75 ± 0.50e	15.71 ± 0.057b	12.46B	9.77 ± 0.54 fg	13.52 ± 0.45 cd	16.45 ± 0.56ab	13.25B
6	9.82 ± 0.32 g	14.20 ± 0.50c	17.25 ± 1.00a	13.76A	10.78 ± 0.34ef	14.99 ± 0.49bc	17.85 ± 1.01a	14.54A
Biochar mean A	8.94C	12.77B	15.56A		9.74C	13.56B	16.27A	

BC₀, BC₁, and BC₂, biochar at rates of (0, 10, and 20 t ha⁻¹). All values are the mean of three replicate analysis ± standard error. Means in each column followed by the same letters are not significantly different ($P < 0.05$) by Duncan's multiple range tests

Table 7 Impact of different rates of biochar and/or Azolla on the shoot parameters of roselle plants

Azolla %	First season				Second season			
	BC ₀	BC ₁	BC ₂	Azolla mean B	BC ₀	BC ₁	BC ₂	Azolla mean B
Fresh weight plant ⁻¹ (g)								
0	202.40 ± 2.00 g	224.20 ± 2.61e	240.77 ± 1.56c	222.46C	199.20 ± 2.00 h	226.57 ± 2.36f	245.00 ± 1.46c	223.59C
3	207.30 ± 0.78f	228.60 ± 2.21e	248.33 ± 1.59b	228.08B	208.73 ± 0.74 g	231.43 ± 2.19e	253.23 ± 1.53b	231.13B
6	210.73 ± 2.08f	235.40 ± 2.01d	255.60 ± 0.69a	233.91A	212.60 ± 1.99 g	239.20 ± 2.08d	261.27 ± 0.58a	237.69A
Biochar mean A	206.81C	229.40B	248.23A	206.81C	206.84C	232.40B	253.17A	206.84C
Dry weight plant ⁻¹ (g)								
0	83.13 ± 2.74c	89.77 ± 1.47b	89.77 ± 1.47b	79.32B	70.40 ± 1.32e	88.30 ± 2.91c	93.87 ± 1.40bc	84.19B
3	89.97 ± 2.82b	93.00 ± 1.59ab	93.00 ± 1.59ab	84.52A	76.90 ± 1.99d	94.73 ± 2.58ab	96.50 ± 1.51b	89.38A
6	92.77 ± 1.13ab	97.97 ± 1.38a	97.97 ± 1.38a	87.76A	79.20 ± 1.99d	96.97 ± 1.47ab	100.63 ± 1.13a	92.26A
Biochar mean A (BC)	88.62B	93.58A	93.58A		75.50C	93.33B	97.00A	
Plant height plant ⁻¹ (cm)								
0	87.83 ± 1.59e	94.87 ± 2.05 cd	112.07 ± 0.96b	98.26C	86.10 ± 1.57e	100.07 ± 1.74 cd	116.33 ± 1.60b	100.83C
3	91.331.68 ± de	98.83 ± 2.45c	118.90 ± 1.63a	103.02B	95.43 ± 1.96d	104.57 ± 2.73c	124.90 ± 0.71a	108.30B
6	93.97 ± 2.12 cd	110.73 ± 0.71b	122.33 ± 2.33a	109.01A	98.43 ± 2.07 cd	113.13 ± 2.95b	126.07 ± 3.58a	112.54A
Biochar mean A	91.04C	101.48B	117.77A		93.32C	105.92B	122.43A	

BC₀, BC₁, and BC₂, biochar at rates of (0, 10, and 20 t ha⁻¹). All values are the mean of three replicate analysis ± standard error. Means in each column followed by the same letters are not significantly different ($P < 0.05$) by Duncan's multiple range tests

Table 8 Impact of different rates of biochar and/or Azolla on the root parameters of roselle plants

Azolla %	First season				Second season			
	BC ₀	BC ₁	BC ₂	Azolla mean B	BC ₀	BC ₁	BC ₂	Azolla mean B
Root length plant ⁻¹ (cm)								
0	17.00 ± 1.15c	24.33 ± 0.88a	24.67 ± 1.45a	22.00B	15.13 ± 0.90c	25.83 ± 0.84a	26.07 ± 1.28a	22.34A
3	18.33 ± 1.45bc	24.67 ± 0.88a	25.67 ± 0.67a	22.89AB	16.67 ± 1.30bc	25.90 ± 0.96a	27.07 ± 0.87a	23.21A
6	20.67 ± 0.88b	25.00 ± 0.00a	26.00 ± 0.58a	23.89A	19.93 ± 1.64b	26.23 ± 0.09a	26.80 ± 1.50a	24.32A
Biochar mean A	18.67B	24.67A	25.44A		17.24B	25.99A	26.64A	
Root fresh weight plant ⁻¹ (g)								
0	22.40 ± 2.08d	27.00 ± 0.44d	39.90 ± 1.22b	29.77B	23.01 ± 0.56f	29.37 ± 0.52d	41.03 ± 0.27ab	31.14C
3	23.93 ± 1.52d	33.43 ± 2.22c	41.47 ± 1.71a	32.94A	25.23 ± 1.44ef	33.65 ± 1.40c	42.77 ± 0.26a	33.88B
6	26.40 ± 0.46d	35.40 ± 2.40bc	41.50 ± 2.21a	34.43A	28.33 ± 0.55de	37.90 ± 2.72b	42.97 ± 0.84a	36.40A
Biochar mean A	24.24C	31.94B	40.96A		25.52C	33.64B	42.26A	
Root dry weight plant ⁻¹ (g)								
0	7.97 ± 1.84e	12.47 ± 0.49 cd	14.13 ± 0.87abc	11.52B	7.67 ± 1.22d	14.23 ± 0.49b	14.77 ± 1.48ab	12.22B
3	8.60 ± 0.57e	12.72 ± 1.76bcd	15.15 ± 0.60ab	12.16B	8.30 ± 0.21 cd	13.76 ± 0.99b	16.52 ± 1.46ab	12.86B
6	11.13 ± 0.15d	13.80 ± 1.01abc	15.86 ± 1.02a	13.60A	10.63 ± 0.81c	15.70 ± 0.96b	18.09 ± 1.34a	14.81A
Biochar mean A		9.23C	13.00B	15.05A	8.87C	14.56B	16.46A	

BC₀, BC₁, and BC₂, biochar at rates of (0, 10, and 20 t ha⁻¹). All values are the mean of three replicate analysis ± standard error. Means in each column followed by the same letters are not significantly different ($P < 0.05$) by Duncan's multiple range tests

Table 9 Impact of different rates of biochar and/or Azolla on some morphological traits of roselle plants

Azolla %	First season				Second season			
	BC ₀	BC ₁	BC ₂	Azolla mean B	BC ₀	BC ₁	BC ₂	Azolla mean B
Stem diameter (cm)								
0	0.83 ± 0.15b	1.13 ± 0.12ab	1.23 ± 0.09ab	1.07A	0.73 ± 0.15d	1.34 ± 0.13bc	1.60 ± 0.06ab	1.23A
3	0.93 ± 0.23ab	1.17 ± 0.03ab	1.27 ± 0.09a	1.12A	1.03 ± 0.23 cd	1.54 ± 0.12ab	1.77 ± 0.09a	1.44A
6	1.03 ± 0.07ab	1.17 ± 0.12ab	1.33 ± 0.09a	1.18A	1.24 ± 0.12bc	1.40 ± 0.06abc	1.65 ± 0.08ab	1.43A
Biochar mean A	0.93B	1.16A	1.28A		1.00C	1.43B	1.67A	
Leaf area (mm²)								
0	3547.86 ± 117.10c	3845.98 ± 247.98bc	4359.22 ± 298.23ab	3917.68A	3545.44 ± 117.27c	3850.31 ± 247.69bc	4369.09 ± 298.18ab	3921.62A
3	3650.66 ± 143.63c	4082.56 ± 139.56abc	4463.76 ± 217.68a	4065.67A	3653.88 ± 143.48c	4088.21 ± 139.68abc	4479.09 ± 217.39a	4073.73A
6	3756.93 ± 128.97c	4130.59 ± 67.44abc	4471.22 ± 94.12a	4119.57A	3760.34 ± 128.64c	4139.74 ± 68.21abc	4486.96 ± 93.49a	4129.01A
Biochar mean A	3651.82C	4019.7B	3651.82A		3653.22C	4026.09B	4445.05A	

BC₀, BC₁, and BC₂, biochar at rates of (0, 10, and 20 t ha⁻¹). All values are the mean of three replicate analysis ± standard error. Means in each column followed by the same letters are not significantly different ($P < 0.05$) by Duncan's multiple range tests

pronounced with combined application of BC and AZ (Tables 7, 8, 9, and 10). On the average basis of both seasons, the high rate of biochar application increased the fresh weight, dry weight, and plant height by 21, 31, and 30%, respectively, over the control. Also, the stem diameter, leaf area, and RWC content increased almost by 52, 21, and 32%, respectively, above the control treatment. Azolla foliar application enhanced the growth parameters since the fresh weight, dry weight, and plant height increased around 5, 10, and 11%, respectively, at the high rate (6%) of AZ foliar application. The combined application of BC and AZ augmented the growth parameters of roselle plants. Adding BC and AZ at high rate increased fresh weight, dry weight,

and plant height roughly by 28, 46, and 43%, respectively, compared to the control. The same trend was observed with the root parameters sic they were significantly increased as a result of adding biochar and/or Azolla.

3.4 Physiological Parameters of Roselle Plants

The high rate of biochar application significantly increased chlorophyll, total anthocyanins (TAC), and total flavonoids (TF) nearby 16, 40, and 62%, respectively, over the control (Fig. 1a–c).

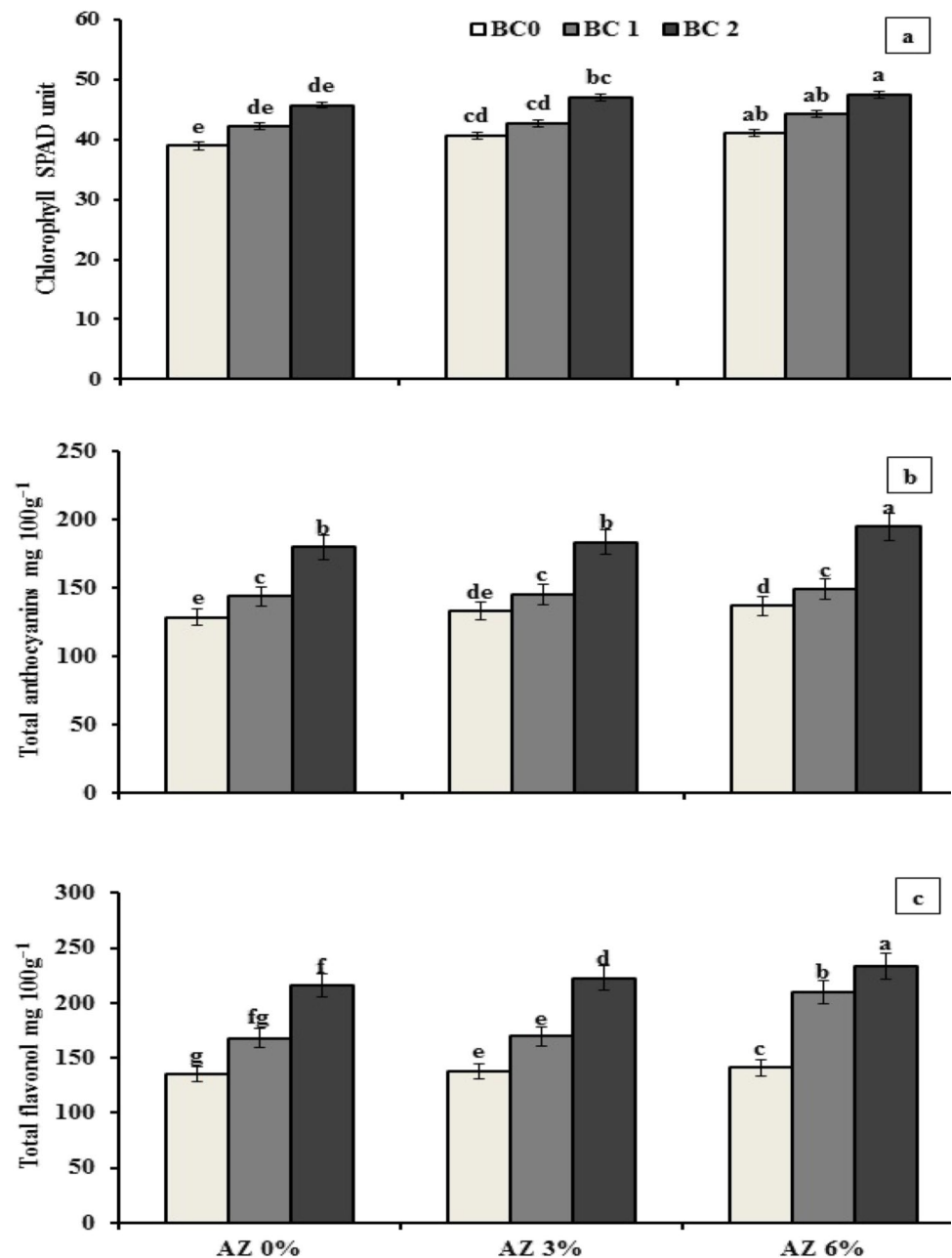
Also, AZ sprayed at high rate significant increased chlorophyll, TAC, and TF almost by 4, 6, and 12%, respectively,

Table 10 Impact of different rates of biochar and or/Azolla on relative water content (RWC) and sepals number as morphological traits of roselle plants

Azolla %	First season				Second season			
	BC ₀	BC ₁	BC ₂	Azolla mean B	BC ₀	BC ₁	BC ₂	Azolla mean B
RWC%								
0	62.27 ± 1.13d	73.34 ± 2.12bc	83.16 ± 0.81a	72.92B	14.00 ± 1.15d	16.33 ± 0.88 cd	23.00 ± 1.15b	17.78B
3	64.43 ± 1.47d	73.63 ± 0.54bc	83.63 ± 1.81a	73.90B	14.33 ± 1.20 cd	17.67 ± 0.88c	26.33 ± 1.20a	19.44B
6	71.00 ± 1.12c	75.52 ± 1.02b	83.73 ± 2.15a	76.75A	15.00 ± 1.15 cd	22.67 ± 0.88b	27.33 ± 0.88a	21.67A
Biochar mean A	65.90C	74.17B	83.51A		14.44C	18.88B	25.56A	
Number of sepals plant⁻¹								
0	60.90 ± 1.31d	76.07 ± 1.91bc	89.06 ± 0.98a	75.35B	12.67 ± 0.88d	24.00 ± 1.73c	30.00 ± 1.00ab	22.22B
3	64.47 ± 1.27d	77.03 ± 0.17bc	90.53 ± 1.55a	77.34B	17.33 ± 1.45d	28.00 ± 1.73bc	35.33 ± 2.33a	26.89A
6	72.77 ± 1.10c	80.29 ± 1.13b	91.40 ± 2.39a	81.49A	18.33 ± 1.20d	30.67 ± 1.86ab	34.67 ± 2.60a	27.89A
Biochar mean A	66.04C	77.80B	90.33A		16.11C	27.56B	33.33A	

BC₀, BC₁, and BC₂, biochar at rates of (0, 10, and 20 t ha⁻¹); RWC, relative water content. All values are the mean of three replicate analysis ± standard error. Means in each column followed by the same letters are not significantly different ($P < 0.05$) by Duncan's multiple range tests

Fig. 1 Effect of biochar and/ or Azolla on chlorophyll SPAD unit (a), total anthocyanins (b), and total flavonol (c) of roselle plants on the average basis of the two seasons. BC0, BC1, and BC2, biochar at rates of (0, 10, and 20 t ha⁻¹); AZ 0%, AZ 3%, and AZ 6%, Azolla at rates of (0, 3, and 6%). Means in each column followed by the same letters are not significantly different ($P < 0.05$) by Duncan's multiple range test



compared to the control treatment. The combined application of BC and sprayed AZ on roselle plants increased chlorophyll, TAC, and TF about by 22, 51, and 72%, respectively, compared to the control treatment.

3.5 Yield Components of Roselle Plants

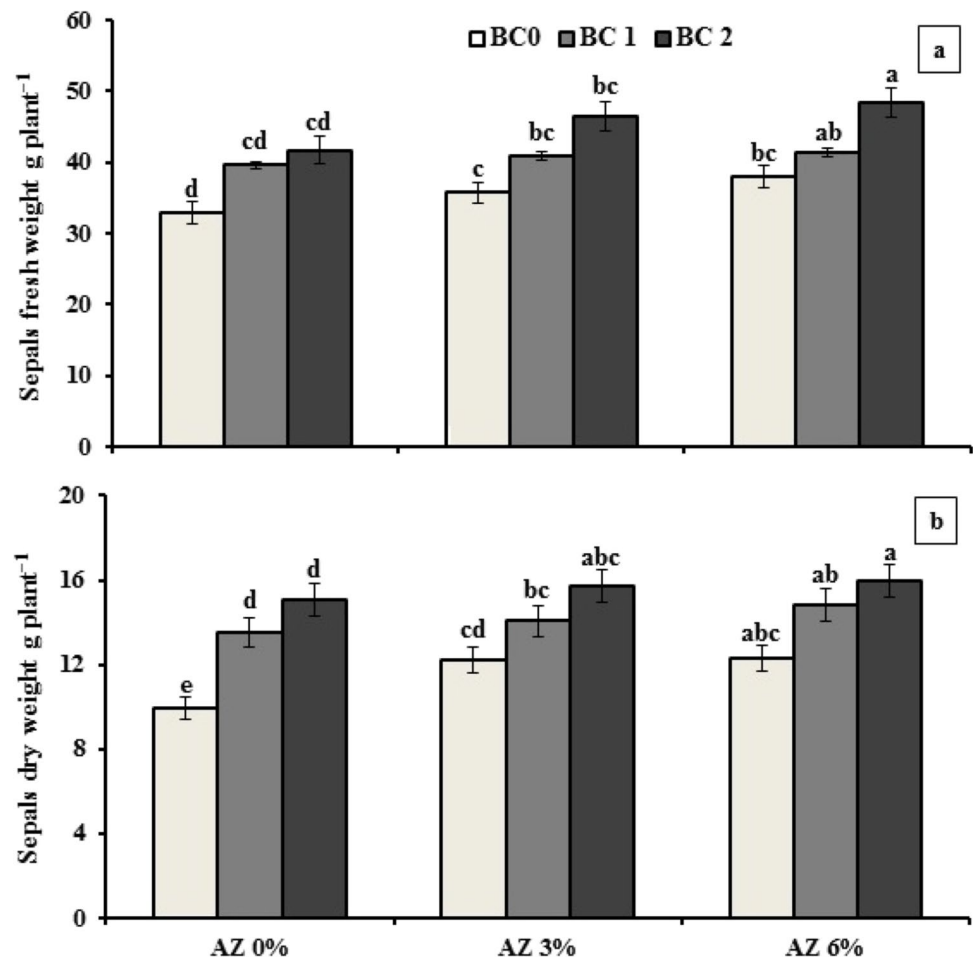
On the average basis of both seasons, the plant yield parameters were improved via high rate of biochar application since sepals no./plant, sepals fresh and dry weight increased nearly by 92, 28, and 35%, respectively, over the control (Fig. 2a and b and Table 10). Also, the fruit number of sepals/plant, sepals fresh, and dry weight increased almost by 23, 11,

and 11%, respectively, as a result of sprayed AZ at high rate compared to the control. The application of BC and sprayed AZ on roselle plants increased fruit number, sepals fresh, and dry weight about by 132, 47, and 60%, respectively, compared to the control treatment.

4 Discussion

Biochar application changed soil properties such as pH, EC, organic matter, and available N, P, and K as well as their uptake. This might be due to biochar is alkaline material in biochar may explain the higher pH in the biochar treatments

Fig. 2 Effect of biochar and/or Azolla on the fresh (a) and dry (b) weight of sepal's yield on the average basis of the two seasons. BC0, BC1, and BC2, biochar at rates of (0, 10, and 20 t ha⁻¹); AZ 0%, AZ 3%, and AZ 6%, Azolla at rates of (0, 3, and 6%). Means denoted by the same letter indicate no significant difference according to Duncan's tests at $p < 0.05$



(Kookana et al. 2011). Also, biochar to the soil is associated with active pH-dependent functional groups, such as OH and COOH that can be raising its pH (Alkharabsheh et al. 2021; Cheng and Lehmann 2009; Weber and Quicker 2018).

The increases of EC value might be due to the high salt content of biochar material. For biochar-treated soils, more mineral ions, such as Ca²⁺, K⁺, and Mg²⁺, were dissolved in soil solution, which was supported by the higher soil salinity (Xu et al. 2016).

Also, biochar application increased soil organic matter (SOM) content that could increase soil water holding capacity (WHC) and enhance nutrient availability such as N, P, and K (Yuan et al. 2016). Agegnehu et al. (2015) indicated that biochar application increased soil organic matter (SOM) by 23.5% over the control. In addition, Murad et al. (2021) noticed that adding biochar at a rate of 4% increased organic matter by 1.67% over the control treatment.

The availability of N, P, and K nutrient as a result of biochar addition can be attributed to its high nutritional supply as direct effect and also as indirect slow release effect of these nutrients (Lei and Zhang 2013; Zheng et al. 2017). Liu et al. (2021) mentioned that biochar application at rate

of 2% increased available N, P, and K by 24, 37, and 19%, respectively, to be uptake by roselle plants. Similar results were obtained by Guo et al. (2020), Villagra-Mendoza et al. (2021), Zheng et al. (2021).

The usefulness of Azolla as a good organic fertilizer to improve agricultural production and the environmental sustainability through its ability to reform biological nitrogen reduce fertilizer leaching in addition to its high content of nutrients and vitamins, which leads to enhanced plant growth (Maham et al. 2020).

Improvement nutrients uptake especially N due to the Azolla sprayed on roselle plants might be due to increasing available nitrogen via atmospheric N fixation as well as its wealthy nutrients in available form that easy to be absorbed through leave stomata. The increased nutritional content in roselle plant and pigment content in plant leaves as a result of spraying Azolla could be due to its mineral richness of N, P, K, Mg, Fe, and Mn (Abou-Sreea et al. 2021). Azolla is a plentiful source of macro-and micronutrients crude protein, growth-promoting cytokinins, jasmonic acid, salicylic acid, and vitamins (de Vries et al. 2018; Shaltout et al. 2012; Stirk and Van Staden 2003).

However, the conjunction use of BC and AZ resulted in a significant increase in yield and quality via improvement soil properties, increasing soil organic matter which leads to soil carbon-rich that enhances soil microorganisms and nutrient availability resulting higher plant yield and its quality (Eissa 2019; Ghadimi et al. 2021; Kabiri et al. 2021). Also, combined application of biochar and Azolla improved the growth features of rosemary plants in calcareous soils of arid and semiarid regions (Sadeh Kasmaei et al. 2019). The use of Azolla improved the amount of chlorophyll as well as the photosynthesis process in *Beta vulgaris* plant (de Bever et al. 2013). This improvement may be due to Azolla sprayed levels on plant leaves increases the metabolites and chlorophyll synthesis that enhance photosynthesis process, which improves yield components and quality by increasing anthocyanins and flavonoids synthesis (Maswada et al. 2021). The chlorophyll content of safflower plants growing in saline-sodic soils was enhanced by 3% using Azolla as compost (Sharifi et al. 2019). The application of Azolla increased significant values of the dry biomass, growth chlorophyll, fruit number, and weight (Youssef et al. 2021). Azolla is high nutritional and organic matter content boosted its potential to improve soil quality and nutrient availability, resulting in considerable squash fruit growth and quality (Abou Hussien et al. 2020).

5 Conclusion

Both biochar application and Azolla sprayed on roselle plants improved soil properties, increased nutrients availability and their uptake, and significantly increased roselle growth, yield, and quality. These increases varied according to biochar and/or Azolla application rates. The addition of biochar and Azolla realized an important role in nutrient availability, metabolites, and chlorophyll synthesis and improves photosynthesis process, which is reflected on the yield, yield components, and its quality. Therefore, applying biochar as soil organic amendments at rate of 20 ton ha⁻¹ combined with Azolla sprayed on roselle plants at 6% concentration is considered the best agricultural management; it is an effective alternative practice to increase available nutrients and yield as well as contributes to the sustainable development of medicinal crops free from harmful chemicals that negatively affect the human health.

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References

- AbouHussien E, Ahmed B, Elbaalawy A (2020) Efficiency of Azolla and biochar application on rice (*oryza sativa* L) productivity in salt-affected soil. *Egypt J Soil Sci* 60:277–288. <https://doi.org/10.21608/ejss.2020.33148.1364>
- Abou-Sreea AIB, Rady MM, Roby MHH, Ahmed SMA, Majrashi A, Ali EF (2021) Cattle manure and bio-nourishing royal jelly as alternatives to chemical fertilizers: potential for sustainable production of organic *Hibiscus sabdariffa* L. *J Appl Res Med Aromat Plants* 25:100334. <https://doi.org/10.1016/j.jarmap.2021.100334>
- Agegnehu G, Bass AM, Nelson PN, Muirhead B, Wright G, Bird MI (2015) Biochar and biochar-compost as soil amendments: effects on peanut yield, soil properties and greenhouse gas emissions in tropical North Queensland, Australia. *Agric Ecosyst Environ* 213:72–85. <https://doi.org/10.1016/J.AGEE.2015.07.027>
- Akhtar SS, Li G, Andersen MN, Liu F (2014) Biochar enhances yield and quality of tomato under reduced irrigation. *Agric Water Manag* 138:37–44. <https://doi.org/10.1016/j.agwat.2014.02.016>
- Alam H, Razaq M, Salahuddin, Khan J (2016) Effect of organic and inorganic phosphorous on growth of roselle (*Hibiscus sabdariffa* L.). *J Northeast Agric Univ (English Ed)* 23:23–30. [https://doi.org/10.1016/s1006-8104\(16\)30055-1](https://doi.org/10.1016/s1006-8104(16)30055-1)
- Alkharabsheh HM, Seleiman MF, Battaglia ML, Shami A, Jalal RS, Alhammad BA, Almutairi KF, Al-Saif AM (2021) Biochar and its broad impacts in soil quality and fertility, nutrient leaching and crop productivity: a review. *Agronomy* 11:993. <https://doi.org/10.3390/agronomy11050993>
- Al-Sayed HM, Hegab SA, Youssef MA, Khalafalla MY (2019) Integrated effect of inorganic and organic nitrogen sources on growth and yield of roselle (*Hibiscus sabdariffa* L). *Assiut J Agric Sci* 50:164–183. <https://doi.org/10.21608/ajas.2019.52773>
- Al-Sayed HM, Hegab SA, Youssef MA, Khalafalla MY, Almaroai YA, Ding Z, Eissa MA (2020) Evaluation of quality and growth of roselle (*Hibiscus sabdariffa* L.) as affected by bio-fertilizers. *J Plant Nutr* 43:1025–1035. <https://doi.org/10.1080/01904167.2020.1711938>
- Batool A, Taj S, Rashid A, Khalid A, Qadeer S, Saleem AR, Ghufuran MA (2015) Potential of soil amendments (biochar and gypsum) in increasing water use efficiency of *Abelmoschus esculentus* L. *Moench Front Plant Sci* 6:733. <https://doi.org/10.3389/fpls.2015.00733>
- Burt R (2004) Soil Survey Laboratory methods manual. Soil survey investigations report No. 42, Version 4.0. Natural resources conservation service, United States Department of Agriculture. Accessed 01 June 2020. https://www.nrcs.usda.gov/wps/portal/nrcs/detail/soils/research/guide/?cid=nrcs142p2_054247.
- Cheng CH, Lehmann J (2009) Ageing of black carbon along a temperature gradient. *Chemosphere* 75:1021–1027. <https://doi.org/10.1016/j.chemosphere.2009.01.045>
- de Bever A, Ndakidemi PA, Laubscher CP (2013) Effects of different combinations of Hoagland's solution and *Azolla filiculoides* on photosynthesis and chlorophyll content in *Beta vulgaris* subsp.

- Cycla “fordhook giant” grown in hydroponic cultures. *African J Biotechnol* 12:2006–2012
- de Vries S, de Vries J, Teschke H, von Dahlen JK, Rose LE, Gould SB (2018) Jasmonic and salicylic acid response in the fern *Azolla filiculoides* and its cyanobiont. *Plant Cell Environ* 41:2530–2548. <https://doi.org/10.1111/pce.13131>
- Eissa MA (2019) Effect of compost and biochar on heavy metals phytostabilization by the halophytic plant old man saltbush [*Atriplex nummularia Lindl.*]. *Soil Sediment Contam* 28:135–147. <https://doi.org/10.1080/15320383.2018.1551325>
- El-Serafy RS, El-Sheshtawy A-NA, El-Razek A, Usama A, El-Hakim A, Ahmed F, Hasham M, Sami R, Khojah E, Al-Mushhin AAM (2021) Growth, yield, quality, and phytochemical behavior of three cultivars of quinoa in response to moringa and Azolla extracts under organic farming conditions. *Agronomy* 11:2186. <https://doi.org/10.3390/agronomy11112186>
- Ghadimi M, Sirousmehr A, Ansari MH, Ghanbari A (2021) Organic soil amendments using vermicomposts under inoculation of N₂-fixing bacteria for sustainable rice production. *PeerJ* 9:e10833. <https://doi.org/10.7717/peerj.10833>
- Guo XX, Tao LH, Zhang J (2020) The role of biochar in organic waste composting and soil improvement: a review. *Waste Manag* 102:884–899. <https://doi.org/10.1016/j.wasman.2019.12.003>
- Jackson ML (1973) *Soil chemical analysis* prentice Hall, Inc, Englewood Cliffs, NJ
- Kabiri P, Motaghian H, Hosseinpur A (2021) Impact of biochar on release kinetics of Pb (II) and Zn (II) in a calcareous soil polluted with mining activities. *J Soil Sci Plant Nutr* 21:22–34. <https://doi.org/10.1007/s42729-020-00336-5>
- Kimani SM, Bimantara PO, Kautsar V, Tawaraya K, Cheng W (2021) Poultry litter biochar application in combination with chemical fertilizer and Azolla green manure improves rice grain yield and nitrogen use efficiency in paddy soil. *Biochar* 3:591–602. <https://doi.org/10.1007/s42773-021-00116-z>
- Kollah B, Patra AK, Mohanty SR (2016) Aquatic microphylla Azolla: a perspective paradigm for sustainable agriculture, environment and global climate change. *Environ Sci Pollut Res* 23:4358–4369. <https://doi.org/10.1007/s11356-015-5857-9>
- Kookana RS, Sarmah AK, Van Zwieten L, Krull E, Singh B (2011) Biochar application to soil. agronomic and environmental benefits and unintended consequences. *Adv Agron* 112:103–143. <https://doi.org/10.1016/B978-0-12-385538-1.00003-2>
- Lees DH, Francis FJ (1971) Quantitative methods for anthocyanins: 6. Flavonols and anthocyanins in cranberries. *J Food Sci* 36:1056–1060. <https://doi.org/10.1111/j.1365-2621.1971.tb03345.x>
- Lehmann J, Joseph S (2009) *Biochar for environmental management*. Science and Technology Earthscan Ltd, London
- Lei O, Zhang R (2013) Effects of biochars derived from different feedstocks and pyrolysis temperatures on soil physical and hydraulic properties. *J Soils Sediments* 13:1561–1572. <https://doi.org/10.1007/s11368-013-0738-7>
- Liu D, Ding Z, Ali EF, Kheir AMS, Eissa MA, Ibrahim OHM (2021) Biochar and compost enhance soil quality and growth of roselle (*Hibiscus sabdariffa* L.) under saline conditions. *Sci Rep* 11:1–11. <https://doi.org/10.1038/s41598-021-88293-6>
- Ma H, Egamberdieva D, Wirth S, Bellingrath-Kimura SD (2019) Effect of biochar and irrigation on soybean-rhizobium symbiotic performance and soil enzymatic activity in field rhizosphere. *Agronomy* 9:626. <https://doi.org/10.3390/agronomy9100626>
- Maham SG, Rahimi A, Subramanian S, Smith DL (2020) The environmental impacts of organic greenhouse tomato production based on the nitrogen-fixing plant (Azolla). *J Clean Prod* 245:118679. <https://doi.org/10.1016/j.jclepro.2019.118679>
- Malyan SK, Bhatia A, Kumar SS, Fagodiya RK, Pugazhendhi A, Duc PA (2019) Mitigation of greenhouse gas intensity by supplementing with Azolla and moderating the dose of nitrogen fertilizer. *Biocatal Agric Biotechnol* 20:101266. <https://doi.org/10.1016/j.bcab.2019.101266>
- Maswada HF, Abd El-Razek UA, El-Sheshtawy ANA, Mazrou YSA (2021) Effect of *Azolla filiculoides* on growth, physiological and yield attributes of maize grown under water and nitrogen deficiencies. *J Plant Growth Regul* 40:558–573. <https://doi.org/10.1007/s00344-020-10120-5>
- McGlashan N, Shah N, Caldecott B, Workman M (2012) High-level techno-economic assessment of negative emissions technologies. *Process Saf Environ Prot* 90:501–510. <https://doi.org/10.1016/j.psep.2012.10.004>
- Murad Z, Ahmad I, Waleed M, Hashim S, Bibi S (2021) Effect of biochar on immobilization of cadmium and soil chemical properties. *Gesunde Pflanz* 1–8. <https://doi.org/10.1007/s10343-021-00597-9>
- Nobile C, Denier J, Houben D (2020) Linking biochar properties to biomass of basil, lettuce and pansy cultivated in growing media. *Sci Hortic (amsterdam)* 261:109001. <https://doi.org/10.1016/j.scienta.2019.109001>
- Novak J, Lima I, Xing B, Gaskin J, Steiner C, Das K, Ahmedna M, Rehrah D, Watts D, Busscher W (2009) Characterization of designer biochar produced at different temperatures and their effects on a loamy sand. *Ann Environ Sci* 3:195–206
- Olsen SR, Sommers LE (1982) Phosphorus methods of soil analysis. In: *Chemical and Microbiological Properties*
- Page AL, Miller RH, Keeney DR (1982) *Methods of soil analysis, part 2*. Chem Microbiol Prop 2:643–698
- Qayyum MF, Haider G, Raza MA, Mohamed AKSH, Rizwan M, El-Sheikh MA, Alyemeni MN, Ali S (2020) Straw-based biochar mediated potassium availability and increased growth and yield of cotton (*Gossypium hirsutum* L.). *J Saudi Chem Soc* 24:963–973. <https://doi.org/10.1016/j.jscs.2020.10.004>
- Rekaby SA, Awad M, Majrashi A, Ali EF, Eissa MA (2021) Corn cob-derived biochar improves the growth of saline-irrigated quinoa in different orders of Egyptian soils. *Horticulturae* 7:221. <https://doi.org/10.3390/horticulturae7080221>
- Sadegh Kasmaei L, Yasrebi J, Zarei M, Ronaghi A, Ghasemi R, Saharkhiz MJ, Ahmadabadi Z, Schnug E (2019) Influence of plant growth promoting rhizobacteria, compost, and biochar of Azolla on rosemary (*Rosmarinus officinalis* L.) growth and some soil quality indicators in a calcareous soil. *Commun Soil Sci Plant Anal* 50:119–131. <https://doi.org/10.1080/00103624.2018.1554669>
- Sanders M, Ayeni AO, Simon JE (2020) Comparison of yield and nutrition of roselle (*Hibiscus sabdariffa*) genotypes in central New Jersey. *J Med Act Plants* 9:242–252. <https://doi.org/10.7275/v5ax-s402>
- Shaltout KH, El-Komi TM, Eid EM (2012) Research paper seasonal variation in the phytomass, chemical composition and nutritional value of *Azolla filiculoides* Lam. along the water courses in the Nile Delta. *Egypt Feddes Repert* 123:37–49. <https://doi.org/10.1002/fedr.201200001>
- Sharifi P, Shorafa M, Mohammadi MH (2019) Comparison of the effect of cow manure, vermicompost, and azolla on safflower growth in a saline-sodic soil. *Commun Soil Sci Plant Anal* 50:1417–1424. <https://doi.org/10.1080/00103624.2019.1621331>
- Singh B, Camps-Arbestain M, Lehmann J (2017) *Biochar: a guide to analytical methods*. Csiro Publishing
- Smart RE, Bingham GE (1974) Rapid estimates of relative water content. *Plant Physiol* 53:258–260. <https://doi.org/10.1104/pp.53.2.258>
- Solaiman ZM, Shafi MI, Beamont E, Anawar HM (2020) Poultry litter biochar increases mycorrhizal colonisation, soil fertility and cucumber yield in a fertigation system on sandy soil. *Agric* 10:480. <https://doi.org/10.3390/agriculture10100480>

- Steel RGD, Torrie JH (1996) Principles and procedures of statistics A biometrical approach, 3rd edn. McGraw Hill Book Company Inc., New York, USA
- Stirk WA, Van Staden J (2003) Occurrence of cytokinin-like compounds in two aquatic ferns and their exudates. *Environ Exp Bot* 49:77–85. [https://doi.org/10.1016/S0098-8472\(02\)00061-8](https://doi.org/10.1016/S0098-8472(02)00061-8)
- Tenic E, Ghogare R, Dhingra A (2020) Biochar—a panacea for agriculture or just carbon? *Horticulturae* 6:37. <https://doi.org/10.3390/horticulturae6030037>
- Villagra-Mendoza K, Masís-Meléndez F, Quesada-Kimsey J, García-González CA, Horn R (2021) Physicochemical changes in loam soils amended with bamboo biochar and their influence in tomato production yield. *Agronomy* 11:2052. <https://doi.org/10.3390/agronomy11102052>
- Weber K, Quicker P (2018) Properties of biochar. *Fuel* 217:240–261. <https://doi.org/10.1016/j.fuel.2017.12.054>
- Xu N, Tan G, Wang H, Gai X (2016) Effect of biochar additions to soil on nitrogen leaching, microbial biomass and bacterial community structure. *Eur J Soil Biol* 74:1–8. <https://doi.org/10.1016/j.ejsobi.2016.02.004>
- Yadav RK, Abraham G, Singh YV, Singh PK (2014) Advancements in the utilization of Azolla-Anabaena system in relation to sustainable agricultural practices. *Proc Indian Natl Sci Acad* 80:301–316. <https://doi.org/10.16943/ptinsa/2014/v80i2/55108>
- Yasmeen A, Basra SMA, Wahid A, Nouman W, Hafeez-ur-Rehman, (2013) Exploring the potential of Moringa oleifera leaf extract (MLE) as a seed priming agent in improving wheat performance. *Turk J Botany* 37:512–520. <https://doi.org/10.3906/bot-1205-19>
- Youssef MA, Al-Huqail AA, Ali EF, Majrashi A (2021) Organic amendment and mulching enhanced the growth and fruit quality of squash plants (*Cucurbita pepo* L.) grown on silty loam soils. *Horticulturae* 7:269. <https://doi.org/10.3390/horticulturae7090269>
- Yuan H, Lu T, Wang Y, Chen Y, Lei T (2016) Sewage sludge biochar: nutrient composition and its effect on the leaching of soil nutrients. *Geoderma* 267:17–23. <https://doi.org/10.1016/j.geoderma.2015.12.020>
- Zhang J, Hussain S, Zhao F, Zhu L, Cao X, Yu S, Jin Q (2018) Effects of *Azospirillum brasilense* and *Pseudomonas fluorescens* on nitrogen transformation and enzyme activity in the rice rhizosphere. *J Soils Sediments* 18:1453–1465. <https://doi.org/10.1007/s11368-017-1861-7>
- Zheng J, Han J, Liu Z, Xia W, Zhang X, Li L, Liu X, Bian R, Cheng K, Zheng J, Pan G (2017) Biochar compound fertilizer increases nitrogen productivity and economic benefits but decreases carbon emission of maize production. *Agric Ecosyst Environ* 241:70–78. <https://doi.org/10.1016/j.agee.2017.02.034>
- Zheng J, Zhang J, Gao L, Wang R, Gao J, Dai Y, Li W, Shen G, Kong F, Zhang J (2021) Effect of straw biochar amendment on tobacco growth, soil properties, and rhizosphere bacterial communities. *Sci Rep* 11:1–14. <https://doi.org/10.1038/s41598-021-00168-y>

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