



# Circulating microRNAs in oncogenic viral infections: potential diagnostic biomarkers

Kinza Hasham<sup>1</sup> · Naveed Ahmed<sup>1,2</sup> · Basit Zeshan<sup>1</sup>

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## Abstract

Cancer is a leading cause of high death rate worldwide. One strategy to control the disease is the early diagnosis by novel biomarkers that express during early stage of the disease. The recent diagnostic strategies in cancer don't have enough specificity to promote the detection of cancer at its beginning. Many biomarkers like protein biomarkers and metabolites are being used for diagnosis of various cancer types but miRNAs are excellent among them, because they have distinctive biochemical characteristics. Moreover, to raise the precision and capability of miRNA to diagnose cancer, the analyzing of both miRNAs and as well as selective mRNA will help in creating a more complete categorizer. Virus constitutes the cause of 20% of entire human cancer cases and both RNA and DNA viruses are linked with tumors in both animal and man. Even though many viruses can cause different tumors in animals, only some of them are linked with human cancers and are presently regarded as oncogenic viruses. These viruses include *Human Papillomavirus* (HPV), *Hepatitis B* (HBV) and *Hepatitis C Virus* (HCV), *Epstein Barr Virus* (EBV), *Human Herpes Virus 8* (HHV8), *Human T cell Leukemia Virus* (HTLV) and *Merkel Cell Polyomavirus* (MCPyV). Expression data of miRNA in several cancers reveal that miRNA profile is different in cancer cells as compared to normal cells. So, miRNA could be useful biomarker for the detection of cancer. The present study strengthens a foundation and gives a logic to investigate the ability of miRNAs as circulating biomarkers in various cancers.

**Keywords** miRNA · Biomarkers · Oncogenic viruses · Circulating · Detection

## 1 Introduction

Cancer is specified as a disease that has high mortality and low endurance rate because of no successful diagnostic approach accessible in clinics. Many protein biomarkers, including carbohydrate antigen (CA), gamma-Semino protein or kallikrein-3, also known as prostate specific antigen (PSA), carcinoembryonic antigen (CEA) have achieved a large number of acknowledgements. However, they are found to be less effective particularly when they are used for screening at early phase or they don't have the ability to differentiate between indolent and aggressive cancers at all [1]. This has developed the identification for novel

and more responsive biomarkers which would help to monitor disease.

MicroRNAs (miRNAs) have shown to be a useful and key player in the regulation of gene predominantly 'post-transcriptional regulation' since their discovery. Similarly, miRNAs have ability to differentiate physiological and pathological conditions of diseased and normal individuals, therefore acting as biomarkers. Circulating miRNAs that exist in the secretory fluids (saliva, plasma, urine and serum) have been used as biomarkers in disease detections. The 1st circulatory miRNA identified is "placental miRNA". It was observed in maternal plasma throughout gestation [2]. Similar to this finding, an increased miRNAs

✉ Basit Zeshan, dr.basitzeshan@ucp.edu.pk | <sup>1</sup>Department of Microbiology, Faculty of Life Sciences, University of Central Punjab, Lahore, Pakistan. <sup>2</sup>Department of Microbiology, Pakistan Kidney and Liver Institute and Research Center (PKLI&RC), Lahore, Pakistan.



concentration was observed in the serum of lymphoma patients as compared to healthy individuals. From then onward, circulating miRNAs can act as invasive biomarkers for detection of different diseases. A hallmark such as accessibility and prodigious stability increase their potential as biomarkers for disease.

## 2 MicroRNA discovery

MicroRNAs (miRNAs) are found in eukaryotes including human. They were first discovered in *Caenorhabditis elegans*. The first miRNA (lin-4) was identified by Victor Ambros and colleagues. The lin-4 gene doesn't have ability to encode protein but can create small RNA [3]. Second miRNA (let-7) was discovered by Gary Ruvkun's group to point the further stages of *C. elegans* development similar to lin-4. After the discovery of 1st miRNA, many miRNAs have been found and are in examine. At present, number of miRNAs have been discovered that act as an oncoMirs or in tumor prohibition [4]. Up till now 1900 miRNAs have been identified having regulatory roles in physiological processes e.g. cellular growth, multiplication, differentiation, holometabolism etc. Excitingly, there was a research on miRNA which concluded that 100 s of mRNA are repressed by a single miRNA [5].

For over a last few decades, various studies indicated the pertinence of miRNAs biology in cancer, showing that they can behave both as oncogenes and tumor repressors; negatively regulating the protein-coding oncogenes [6, 7]. Many researchers have dictated that miRNAs can influence cancer phenotypes and many reports have exhibited miRNAs expression profiles which provide detail about tumor origin, prognosis and diagnosis of cancer [8].

### 2.1 Canonical biogenesis of MicroRNA

miRNA biogenesis consists of two step processes; nuclear and subsequent cytoplasmic cleavage. The first step is in nucleus where RNA pol 2 transcripts Intergenic miRNA to form pri-miRNA (~60–70nt stem loop intermediary, called the miRNA precursor, or the pre-miRNA). Only that pri-miRNA with the proper stem length and the ability of creating 5' and 3' single-stranded RNA overhangs will be effectively continued and converted into useful miRNA. The maturation process starts by the DROSHA RNase III endonuclease and the double-stranded RNA-binding protein DiGeorge syndrome critical region gene 8 (DGCR8) complex collectively known as 'the microprocessor' [9], which produces a cut on pri-miRNA, makes it shorter and forms pre-miRNA and thus the stem loop of pre-miRNA has a 5' phosphate and ~2 nt 3' overhang.

For further processing, nucleocytoplasmic transporter factor Exportin-5 and Ran-GTP brings pri-miRNA into the cytoplasm [10]. This stops nuclear damaging and promotes translocation in the cytoplasm. In cytoplasm there is an enzyme called dicer which performs an activity of RNase 3 endonuclease. Firstly, it recognizes the bases of pre-miRNA, binds with pre-MicroRNA, cleaves it and makes dsRNA without hairpin. The dicer loop cleavage is leaving the 5' phosphate and ~2 nt 3' overhang that binds with Argonaut (AGO) protein. The combination of RNA, Argonaut with RNA-induced silencing complex (RISC) target mRNA via direct base pairing. Finally the RNA duplex is untwisted and the single stranded mature miRNA is integrated into the protein complex RISC to function as a guide, controlling the suppression of target mRNA. The protein expression of targeted mRNA is being altered by the resultant miRNA/mRNA hybrid. MiRNA may be degraded or translation may be inhibited [9].

miRNAs are transcribed by RNA polymerase II or III as pri-miRNA, and then processed in the nucleus by Drosha-DGCR8 into pre-miRNA. The pre-miRNA is exported to the cytoplasm by exportin-5 and further cleaved by a complex composed of Dicer and TRBP. The functional strand of mature miRNA is incorporated into the RNA-induced silencing complex (RISC), which contains Argonaute protein (Fig. 1).

## 3 Biological role of miRNAs

As mentioned above, miRNAs are essential part of feedback loops having strength to control biological activity. Therefore, miRNAs might work to enhance the clarity of gene expression by controlling the proteins production [11]. Moreover, single miRNA may contribute to control

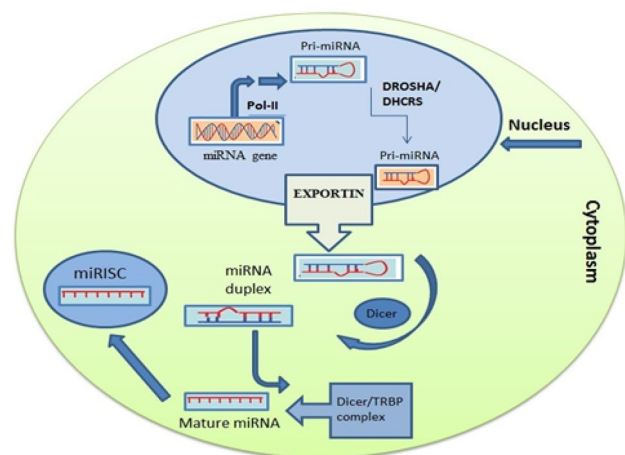


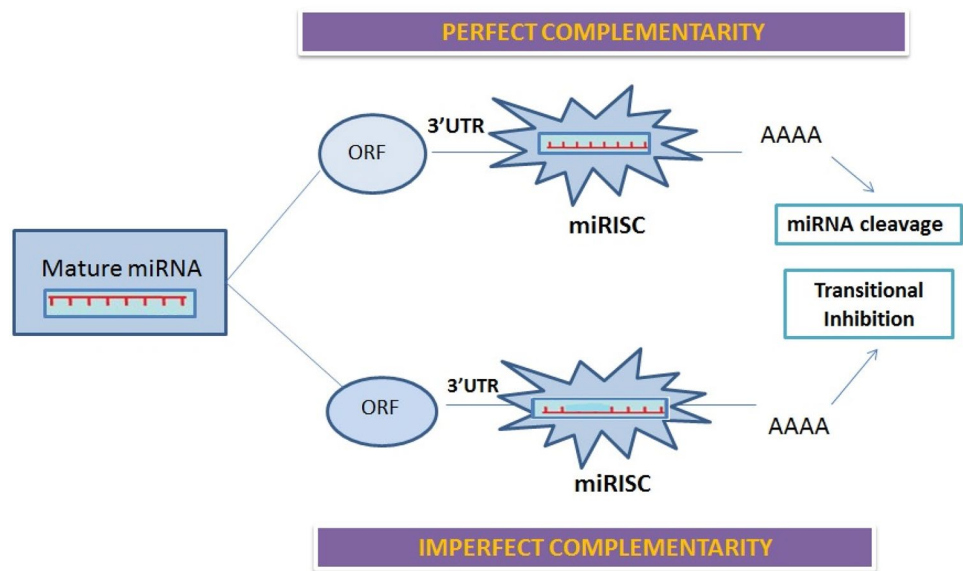
Fig. 1 Biogenesis of microRNAs

specific differentiation processes as demonstrated by early research in worms for example, the control of lin-41 by let-7. Concernedly, miRNA also play a major role in stem cell differentiation and in persuading pluripotency where individuals miRNA clumps, miR-302, was currently revealed to be effective to create iPSC from both human and mouse embryonic cells [12]. Their contribution in differentiation and development of cellular affinity proceeds that reduction of miRNAs function might develop the increased cellular flexibility, de-differentiation and increased tendency for oncogenic modifications. Moreover, because of many transcripts modulated by individual miRNA, their inclusive function in oncogenesis might be dependent on the context. Appropriately, a specific

miRNA might be upregulated in various types of cancer and thus believably oncogenic, while downregulated in other types of cancer, hence show tumor suppressor function. For example miR-29 is responsible for tumor-suppressor in lung tumors while in breast cancer, miR-29 is responsible for oncogenic functions [13, 14] (Fig. 2).

miRNAs use two mechanisms to exert gene regulation. Some animal miRNAs can bind to mRNA targets with exact complementarity and induce the RNAi pathway. miRNAs also bind to targets with imperfect complementarity and block translation. miRNAs detection methods, advantages and their disadvantages are described in Table 1.

**Fig. 2** MicroRNAs (miRNAs) as tumor suppressors and oncogenes



**Table 1** miRNAs detection methods, advantages and their disadvantages

miRNAs detection methods	Advantages	Disadvantages	References
Isothermal exponential amplification-based methods	No PCR require Good sensitivity Real-time assay in some cases	Complex probe design Many enzymes are used	[86]
Amplification-based method	No PCR needed Better regulation of signal amplification In situ identification attain in some cases	Complex probe design Unspecific amplification	[87]
Direct Quantitative Analysis of Multiple miRNAs (DQAMmiR)	Easy probe design Various target detection	Specific equipment needed	[17]
qPCR	Fixed protocol high sensitivity high specificity	Needs quality miRNA annotation	[88]
RNA sequencing	Complete genome analysis No need of miRNA annotation	Need most input material Less sensitive	[89]
Multiplex RT-PCR	High sensitivity High specificity Easy data analysis	Not suitable for small scale experiments of 1-2 samples Needs quality miRNA annotation	[90]
miRNAs arrays	Fixed protocols Purpose built analysis tools	Less quantitative Needs quality miRNA annotation	[91]

## 4 Molecular biomarkers for cancer detection and treatment

Identification of cancer biomarkers (protein biomolecules) present in the body fluids have great significance in early detection and as well as therapy efficacy. Many devices have been made for detecting the molecular markers. Molecular biomarkers have the ability to improve the current state of early cancer detection, some of them are given in Table 2.

### 4.1 Circulating miRNA as a potential biomarker

Biomarkers have the capability to enable us to diagnose or examine disease. In the past decades many miRNAs have been examined in various forms of human cancer. The abnormal expression of miRNAs promote to cancer development via different kinds of mechanisms like deletions, mutations etc. [15]. MiRNAs expression profiling act as a useful diagnostic and prognostic tool and many studies have been done which indicates that miRNAs are considered as oncogene or a tumor suppressor. Currently, it has been observed that miRNAs are dispersed in the peripheral blood [16]. They perhaps are found inside extracellular vesicles in the plasma, bound to proteins such as Argonaut 2, and are not degraded hence used as intracellular communicators. Therefore, they may act as biomarkers for detecting different kinds of diseases. Particularly, biomarkers should be discovered so that it can be used to detect diseases at early stages.

Slide-based staining assay is used to detect the changes in miRNAs present in the tissues. Latter studies in the field of pancreatic cancer have shown that some changes have been seen in miRNA level in both normal and malignant

tissues. So this technique is used to monitor treatment options [17]. There is a noninvasive way to expose miRNA from blood sample and from urine as a circulating tumor cells. MiRNAs are considered as highly stable in formalin-fixed, paraffin-embedded (FFPE) tissue from hepatocellular carcinoma [18], papillary thyroid carcinoma [19], renal tumor [5] and lung cancer [20]. So miRNA expression studied from repository tissue specimen and liquid body substances such as blood will be essential for identifying diseased state.

Serum of patients who were suffering from large B-cell lymphoma was examined and 1st circulating miRNA was used as a biomarker to identify this disease. For earlier screening of circulating miRNAs different technologies are involved like microarray profiling, NGS, and real-time PCR array [21]. Diseases like neurological disorders and cancers can be treated successfully if they are detected in their early phases. Both cancer and neurological affliction requires recognition of dependable and early biomarkers. Modulation of miRNAs and their target mRNAs give best choice for making of novel biomarkers in these disorders.

### 4.2 Oncogenic viral infections

Viral infections constitute around 20% of entire human cancer cases. Even though many viruses can cause different tumors in animals, only seven of them are linked with human cancers and are presently regard as oncogenic viruses. These viruses include *Human Papilloma virus* (HPV), *Hepatitis B* (HBV) and *Hepatitis C Virus* (HCV) [22], *Epstein Barr Virus* (EBV) [23], *Human Herpesvirus-8* (HHV8) [24], *Human T-cell Leukemia Virus* (HTLV) [25] and *Merkel Cell Polyomavirus* (MCPyV) [26]. There are various mechanisms by which these viruses transformed normal cells into tumor cells [27]. These viruses will alter the gene

**Table 2** Recent molecular biomarkers used for diagnosis and treatment of cancer

Cancers	Molecular biomarkers	Techniques	Treatment	References
Non-small cell lung cancer	EGFR overexpression	Immunohistochemistry, RT-q(PCR)	EGFR-tyrosine kinase inhibitor (TKI)	[92]
Breast cancer	HER2 overexpression, overexpression of estrogen receptor	CISH, IHC	Trastuzumab, Tamoxifen	[93]
Gastrointestinal stromal tumors (GISTs)	CD117 overexpression	Immunohistochemistry	Imatinib	[94]
Glioblastoma	(MGMT) overexpression	Immunohistochemistry	Temozolomide	[95]
Metastatic melanoma	CTLA-4	Immunohistochemistry	Ipilimumab	[96]
Thyroid cancer	BRAF and RAS point mutations	ThyroSeq version 2 next generation sequencing	histone deacetylase inhibitors and retinoids	[97]
Chronic myeloid leukemia	Philadelphia chromosome-positive BCR-ABL fusion gene	FISH	Imatinib	[98]

expression or they may produce changes in the genetic level. Oncogenic viruses are able to insert their genome in cellular chromosomes that leads to genetic abnormality. Virus oncoproteins can stimulate the cellular signaling processes that may divert the expression of cellular genes. Infection causes the alteration at gene level and reproduction of oncogenic viruses may cause the spreading of cancer stem cells in human [28] (Table 3).

Oncogenic viruses also causes latent infections as they stay in the body for a long time, e.g. EBV may survive in the body for lifetime [29]. These viruses also cause chronic infections that continue for a long time e.g. *Hepatitis B* or *C*. Epidemiological studies revealed that there is a strong association between HBV with liver cancer [30]. MCPyV can cause Merkel cell tumor [26] and HTLV-1 causes adult T-cell lymphoma. HHV8 (also known as Kaposi’s sarcoma-linked

*Herpesvirus*, KSHV) has been dependable for Kaposi’s sarcoma generally found in patients with acquired immunodeficiency syndrome (AIDS) [24] (Tables 4 and 5)

### 4.3 miRNA for the detection of *Hepatitis C Virus*

Approximately 170–200 million people are infected with *Hepatitis C Virus* [22]. One of the critical provocations in the research of HCV is to detect the liver disease at its early stage which will better improve the result of antiviral treatment. The virus genomes consist of 5’ and 3’ untranslated regions that are used for viral translation and replication. Prestigiously, miRNAs are identified as important in pathogenesis of HCV infection that are linked with liver disease and the disruptions of miRNAs are involved in change of HCV replication [31].

**Table 3** miRNA expression associated with different type of oncogenic viral infections

Oncogenic viruses	Cancer types	Sources	miRNAs	Expression	References
<i>Hepatitis C Virus</i>	Hepatocellular carcinoma	Serum	miR-29a, 146, 149, 221, 222	Increased	[34, 35]
			miR-196a	Decreased	
	Urine	miR-625, 532, 618	Increased		
		miR-516-5p/650	Decreased		
Liver cirrhosis	Serum	miR-20a, 93	Increased	[99]	
		miR-92	Decreased		
<i>Epstein Barr Virus</i>	Nasopharyngeal carcinoma	Serum	BART7 and 13	Decreased	[46]
<i>Hepatitis B Virus</i>	Hepatocellular carcinoma	Serum	miR-885-5p, 122, 21	Increased	[56]
<i>Human T-lymphotropic Virus</i>	T-cell leukemia	Plasma	miR-93, 155	Increased	[63, 68, 100]
			miR-126	Decreased	
<i>Human Papilloma Virus</i>	Cervical cancer	Serum	miR-21, 146a, 224, 182	Increased	[70]
			miR-218	Decreased	
	Colon cancer	Serum	miR-21	Increased	
			let7a-1, 143, 145, 16, 125b, 31, 133b, 96, 145	Decreased	
<i>Merkel cell Polyoma Virus</i>	Prostate cancer	Serum	miR-141	Increased	[33, 82]
	Merkel cell carcinoma	Serum	MCV-miR-5p, 23	Decreased	
<i>Human Herpes Virus-8</i>	Kaposi sarcoma/B-cell lymphoma	Serum	miR-143, 145, 126-3p and 13	Increased	
			miR-221, 222, let7 family	Decreased	

**Table 4** miRNAs as an oncogene

Tumors	miRNAs	Target	References
Pancreatic cancer	miR-103, miR-107, miR-21, miR-155 miR-141, miR-220c	Tumor protein 53-induced nuclear protein 1 (TP53INP1) Zinc finger E-box binding homeobox 1 (ZEB1)	[101]
Colorectal cancer	MiR-21	induces invasion, intravasation, or metastasis in Colo206f-cells	[102]
Lungs cancer	miR-17-92 including miR106a, miR17-5p, miR19a, miR-25, and miR-93	Inhibition of apoptosis	[103]



**Table 5** miRNAs as a tumor suppressor

Tumors	miRNAs as tumor suppressor	Functions	References
Pancreatic cancer	MiR-34a	Induced by P53 tumor suppressor protein Aberrant CpG methylation	[104]
Colorectal cancer	MiRNA-143 and 145	Targeting tumorigenic elements at the translational or posttranscriptional level	[105]
Lungs cancer	let-7 family, including let-7b, let-7c, let-7d, let-7f, and let-7g	Inhibit the RAS oncogene Inhibits the expression of HMGA2, a high-mobility group protein	[91]

A cohesion between miR-196 and 5A coding region of HCV JFH1 genome has been seen in previous studies. In addition to this, IFN $\beta$  therapy leads to substantial expression of miR-196 initiation in Huh-7 cell line. This showed that miR-196 have a significant role in changing the HCV expression. The past studies have recognized that miR-196a hinders the expression of HCV by targeting the genome of HCV. Later on it leads to the up regulation of heme oxygenase liberating anti-inflammatory molecules. Thus for the development of HCV infection miR-196 is an important factor. Later on it was theorize that the upregulation of miR-196 may be helpful in detection, prevention and treatment of the HCV infection [32].

Moreover, further studies have showed miR-196 as a biomarker for early diagnosis of HCV. Microarray analysis of miRNAs demonstrated that the HepG-HCV and HepG2-control have six differently expressed miRNAs. With over expression of HCV core protein, there was an upregulation of miR-29a, miR-146, miR-149, miR-221 and miR-222 and down regulation of miR-196a [33].

Shrivastava et al. [34] demonstrated the differential expression of miRNA as a biomarker related to liver disease. It was found that many circulating miRNA were upregulated in serum of HCV infected patient as compared to healthy individuals [34]. The serum of the patients with HCV showed high expressions of miR-20a and miR-93 and decreased expression of miR-92 with high level of fibrosis. Some sample studies showed that as disease developed from acute to chronic infection, the expression of miR-20 stayed higher while expression of miR-92 decreased post infection. MiR-625, miR-532 and miR-618 were found to be upregulated as 56%, 62.5% and 72% of Hepatocellular carcinoma (HCC) in post HCV infection. Similarly, expression of miR-516-5p and miR-650 were downregulated as 50% and 72%. Due to differential expressions of miRNAs it is now possible to target the genes which are associated to HCC progression [35].

#### 4.4 miRNA for detection of Epstein–Barr Virus

Latent infections have been associated with large spectrum of non-cancerous and cancerous diseases caused

by EBV [36]. This virus causes Burkitt's lymphoma, Nasopharyngeal Carcinoma (NPC) and Hodgkin's lymphoma. Clinically, NPC is vastly incurable and metastatic [37]. Large numbers of EBV encoded miRNAs have been found in NPC tumors.

EBV throughout its replication cycle represents two miRNAs; 'BHRF1 and BamHI-A Rightward Transcripts (BART) [38], which are largely expressed in NPC cells. In recent studies, circulating BART-miRNAs along with BamHI-W DNA were examined before and after treatment in patients. They selected three BART-miRNA which were miR-BART2-5p, miR-BART17-5p, and miR-BART18-5p out of 44 mature BART-miRNAs that were previously well-known as circulating freely in NPC patients. Circulating BamHI-W DNA was a functional biomarker for the identification of NPC before treatment. The recognition of circulating miR-BART17-5p was supposed to be a potential biomarker to evaluate a poor treatment outcome. Certain BART-miRNAs were discharged in the blood are now recognizing profusely in NPC patients [39].

A group of researchers analyzed the altered expression of BART miRNAs that are linked with EBV infecting B cells, noncancerous cells and cancerous NPC cells in order to identify BART miRNAs. Analysis of EBV infected cells identified extracellular secreted BART miRNAs; miR-BART3, miR-BART7 and miR-BART13. Utilizing these miRNAs as a marker, they examined plasma of NPC patients, non NPC cancer controls and healthy ones. Results demonstrated that the plasma from NPC patients contain miR-BART7 and miR-BART13 which were absent in non NPC controls. Moreover, by following radiotherapy plasma levels of miR-BART7 and miR-BART13 were reduced in patients. So circulating BART-miRNA may serve as a marker for diagnosis and prognosis of NPC [40].

#### 4.5 miRNA for the detection of Hepatitis B Virus

HBV infection is widely spread in African, Asian and Western countries. The prevalence of HBV is transitional (2–7%) in Southern and Eastern Europe where the cause of infection is through perinatal transmission, tattooing, making

cut outs, and nosocomial transference, by sexual contact and by needle exchange between drugs users [41].

Generally, hepatitis B vaccinations strategy for new born and teenage has been started that reduce the prevalence of HBV infection. The defensive immune response that demolish virus-infected liver cells causes acute liver injury and due to the lack of immune response, virus infected cells were not destroyed and infection turned into chronic. This case is related with antenatal acquired HBV infection that is concerned to chronicity up to 95% [42].

From a global view point, majority of the hepatocellular carcinoma is due to chronic HBV infection. Elders with chronic hepatitis B get HCC about 5% for decades, that is about 100-fold greater than the rate between uninfected populations [43]. The high death rate is because of recognition at its last stage with few therapeutic choices. Actually insufficient diagnostic markers and poor treatment plan makes it a crucial challenge.

In case of HBV infections, circulating miRNAs are attaining attention for the diagnosis and prognosis of HCC [44]. Until now, two miRNAs exhibit extremely high potential to diagnose HCC which were miRNA-21 and miRNA-122. miRNA-122 was a liver miRNA while miRNA-21 is created by different tissues containing heart, liver and colon that were severally involved in cancer growth and coronary disease development. MiRNA-21 activates phosphatases (e.g., ATK and MAPK) which inhibits the tumor suppressor pathways while miRNA-122 hinders the cancer growth, being a tumor suppresser gene. A direct association was seen between enlarge miRNA-21 level and cell proliferation. Additionally, high level of circulating miRNA-21 was linked with increasing HCC therefore, specify a poor prognosis [45]. Level of serum miRNA-122 is inversely associated with the extremity of liver fibrosis. The antitumor characteristic of miRNA-122 has been effectively utilized in preclinical model to stop HCC development. The diagnostic precision of miRNA-21 somewhat surpassed that miRNA-21 have sensitivity and specificity up to 87% and 80% as compared to miRNA-122 which have 68.0% and 73.3% so they were the potential biomarker for the diagnosis of early HCC [46].

During recent years, many other miRNAs were also found to play crucial functional roles. miR-106b and miR-181b may serve as a biomarker for diagnosis of liver cirrhosis [47]. Up regulation of miR-885-5p was observed in the serum of HBV, LC and HCC patients and may represent as a biomarker for liver diseases. Let-7c, miR-23b, miR-122, miR-150 and miR-122-5p, miR-192-5p acted separately as a potential biomarker for HBV infection. MiR-143 and miR-215 present in the serum could act as a potential biomarker for CHB and HCC [48]. MiR-21-5p may act as a marker for viral hepatitis [49] and after liver transplantation MiR-146a-5p may serve as a marker for acute rejection. All these findings would help us to

understand the expression mechanism of circulating miRNAs in various phases of HBV driven infections and furthermore to the production of diagnostic tools for the recognition of CHB and LC.

#### 4.6 miRNA for the detection of Human T-cell Leukemia Virus (HTLV)

HTLV is a retrovirus, representative of Delta retrovirus genus that was identified in early 1980s by two-separate groups in America [50] and in Japan [51]. Though, there are four types of HTLV and HTLV-1 is the utmost pathogenic and has the perception that it may be the 1st oncogenic retrovirus discovered in humans. HTLV approximately infects 15–20 million populations worldwide and involved as a pathogen in different kinds of diseases like adult T-cell leukemia or lymphoma (ATL) and tropical spastic paraparesis or HTLV-1 associated myelopathy (TSP/HAM). HAM/TSP was 1st identified in 1969 over the decade before the revelation of HTLV-1 [52].

As many studies have shown that miRNAs were involved in progression and prognosis of disease, in case of HTLV many miRNAs were concerned with the survival of HTLV-1 infected cells. One key discovery was that up-regulation of miR-93 was found in HTLV-1 infected cells. Moreover, various kinds of tumors including HTLV-1 have increased expression of miR-93 which shows that miRNA play a vital role in cellular alteration [53, 54]. Additionally miR-223 demonstrated to be upregulated in HTLV-1 infected cells in adult T-cell leukemia patients [20]. MiRNA-155 has also been recognized as it supports the cellular transformation of HTLV-1 and it has also been seen in another oncogenic virus like EBV [55] and that miRNAs might be involved in the progression of disease.

In order to check whether miRNAs were involved in detection of disease, an experiment was conducted by isolating CD4-positive cells from two healthy individuals, three acute and three chronic ATL patients and microarray was used to profile cellular miRNAs. Five miRNAs were screened which were miR-155, let-7g, miR-126, miR-130a and let-7b. Because a huge variation was seen in their expression in diseased patients' v/s healthy individuals. The expression level prior to 5 miRNAs was re-measured by RT-qPCR and it wasn't repeatedly consistent in cells and plasma. The increased and decreased level of miR-155 and miR-126 altered with ATL phase. So this study showed a quantitative variation between plasma miRNAs and cellular miRNAs. The increased level of plasma miR-155 and decreased level of miR-126 associated with their bad prediction, showed their functionality as a unique biomarker for the analysis of disease stage [56, 57].

## 4.7 miRNA for the detection of Human Papillomavirus (HPV)

*Human Papillomavirus* (HPV) preferably affects mucosa epithelial cells that effectuate non-cancerous and sometimes malignant tumor. Different species of HPVs, like HPV16, 18, 31, and 45, were recognized usually in anogenital cancers, especially cervical cancer and anal cancer, and hence they were counted to be oncogenic [58]. HPV16 infection appears to be common in colorectal cancer-tissues. McNICOL and DODD first detected HPV DNA in prostatic tissues using polymerase chain reaction (PCR). Cervical cancer is most usual cancers in female with high mortality about 233,000 deaths per year [59]. The prevalence is lesser in emerging states because of cervical screening tests and so many health education paths. The causal link in high-risk HPV (HR-HPV) infection and cervical cancer has been well recorded in epidemiological and functional research. High-risk HPVs, such as HPV16, HPV18, and HPV31 have been identified in up to 99.7% of cervical squamous cell carcinomas and 94–100% of cervical adeno- and adenocarcinomas. The high-risk HPV oncoproteins, E6 and E7, lead to cervical cancer by severely immobilizing the cellular tumor repressor proteins p53 and pRb [60].

Throughout the world, cervical cancer remains one of the leading causes of death in women. It is a contagious disease having genes associated with complicate biological processes, so closely linked with constant infection of high-risk human papilloma virus (HPV) [61]. Chromosomal mutation and alteration of single nucleotide polymorphisms; these were the key factors to cause the malignant alteration of cervical epithelial [62].

Over recent years, the analysis of small RNAs (miRNA) modulation of gene expression begins to be a hot spot. Above 1000 human miRNAs were estimated to regulate around 60% of protein-coding genes, specifying their substantial role in many biological processes. In cervical cancer tissues, up regulation of miR-21 was seen [63], accumulating affirmation about change in the expression of miR-21 in cervical cancer demonstrates that it might play a vital role in tumor biology.

In order to check the clinical value of miR-21, a group of scientist performed RT-PCR assay to examine the expression mechanism of miR-21-3p and miR-21-5p in HPV associated carcinoma [64]. There data confirmed that the expression of miR-21-3p and miR-21-5p particularly increased in cervical carcinoma as compared to normal tissues, which shows that miR-21 could play a crucial role in the progression and poor diagnosis of human cervical cancer. MiR-21 was discovered for the 1st time as a marker for prognosticating the clinical result of cervical cancer patients. Similar to this, many experiments were done to check the increased or decreased level of miRNAs

in cervical cancer. Up regulation of miRNA-182 was seen in cervical carcinoma, and a significant association in high expression of miR-182 and developed phase of cervical cancer was discovered. So this finding show that miR-182 plays an oncogenic role in cervical cancer [64].

Decreased expression of microRNA-218 was also seen in serum of cervical cancer patients and up regulation of miR-224 in cervical cancer were linked with the assertive development and poor prognosis of cervical cancer. Lately [65] investigate that particular miRNA signature would differentiate between normal colon and colon cancer and specifically mir-21 was seen to be over expressed in colon cancer patients up to 87% while mir-143, miR-145 [66] let-7a-1 [67] miR-16, miR-125b miR-31, miR-133b, miR-96 and miR-145 was found to be low expressed in colorectal cancer. Additionally, tumor suppressor miRNA, miR-34a was showing low level in CRC tissues so it can be used for diagnosing CRCs. Patients having metastatic prostate cancer have high level of miR-141 in serum [68]. Similar study examines the miRNAs expression level in plasma and reveals that miR-141 is a novel biomarker for the detection of prostate cancer [68].

## 4.8 miRNA for the detection of Merkel Cell Polyomavirus (MCPyV)

*Polyomaviruses* (PyVs) were linked with malignancies containing Merkel cell carcinoma (MCC). With the discovery of PyVs in 2008, Epidemiological studies have recognized MCPyV, as a usual virus that causes the infection in human population. Enzyme linked immunosorbent assays particularly used for the immunogenic determinative of MCPyV, the main capsid protein VP1, have been utilized to govern that up to 80% of the grownup population carry serum antibodies to MCPyV [69].

The MCPyV genome consist of 22 nucleotides viral miRNA (MCV-miR-M1-5p) that probably causes the auto regulation of initial viral gene expression over the late stage of infection, as it was manifest to decrease the level of reporter transcripts comprising MCPyV initial region sequences [70]. One research revealed that miRNAs expressions are conserved in almost 50% of MCPyV-positive MCC tumors, and there is an association between viral genome copy number and expression level of miRNA in tumor [70]. The existence of MCPyV miRNA in MCC tumors surely assures more observation to check its role in the pathogenesis of MCC.

As previous studies have suggested that MCPyV encodes a MicroRNA which may control the regulation of cellular and viral genes. A group of scientist performed an experiment to check whether the MCPyV encodes a miRNA which is expressed in MCC tumors. More than 30 million small RNAs from 7 MCC tumors which were



cryopreserved and 1 sample that was perilesional were sequenced. By using RT-qPCR, 45 extra MCC tumors were observed to check the expression of MCPyV-encoded mature miRNA [70, 71].

Through direct sequencing between two out of three MCPyV-positive MCC tumors, "MCV-miR-M1-5p" was identified. However MCV-miR-M1, a precursor miRNA had been identified in silico [71]. Particularly, the sequence of MCV-miR-M1 was similar in 79 reads which were obtained after in vivo but it varies from the in silico identified mature miRNA with two nucleotide alteration, which results in the specific seed region and a distinctive set of prognostic target genes. This mature microRNA was identified in MCPyV-positive MCCs (n = 38) and in 0% of MCPyV-negative MCCs (n = 13) by the help of real-time PCR. So this result concluded that in 50% of MCPyV-positive MCCs, expression level of MCV-miR-M1-5p is low, which shows that virus miRNA plays vital role in developing immune elusion and controlling viral DNA reproduction.

Xie and co-workers check the miRNA account between MCPyV-positive and negative MCC. MiR-23 remarkably expressed lower in MCPyV-positive MCC as compared to MCPyV-negative MCC. The increased expression of miR-203 in MCPyV-negative MCC prevents the cell growth and persuades cell cycle arrest [72]. This result demonstrates that MCPyV may lead to the cell proliferation by suppressing the expression of miR-203, but the exact process by which MCPyV regulates this miRNA, is not clarified yet.

#### 4.9 miRNA for the detection of *Human Herpes Virus 8 (HH8V)*

*Human Herpes Virus 8*, also known as *Kaposi sarcoma-associated herpes virus* is a representative of gamma-herpes virus family which progress to sustain long-lived latent infection in the independents [73]. KSHV is responsible for Kaposi sarcoma, B-cell lymphoma and some Castleman diseases [74]. Kaposi's sarcoma (KS) is a multitudinous tumor of mesenchymal genesis which was first represented by Moritz Kaposi in 1872 [75].

Past researches lead to the discovery of many viral coded miRNA that play vital role in controlling the regulation of herpes virus latent infections [76]. Viral miRNA can regulate the gene expression in both host and viral cell during infection without producing any toxoid viral protein which can be identified by the host defense system [77]. In plasma and serum sample, variation between the expression level of viral and cellular miRNA demonstrated particular patterns in different diseases like sepsis, cancer, atherosclerosis etc. [78]. These miRNAs stay in circulation in a secure form, being extremely resistant to severe changes in pH, endogenic RNase activity, and difference in temperature [79].

Moreover, many mature miRNAs, obtained from 12 precursor mRNAs from KSHV genome, play vital roles in KSHV-induced cell modification [80]. Furthermore, this study reported that increase levels of KSHV miRNAs in plasma were linked with a poor clinical outcome in the patients having sepsis. Identified virus encoded miRNAs might indicate a high sensitive assay to persuade the accurate prevalence of various viral infections, including latent KSHV infection.

One study reveal that there is an upregulation of miR-143/145 in KS that act as a biomarker as compared miR-221/222, miR-155, and the let-7 family, which were down regulated in KS [81]. Other scientist also clearly recognized 170 deregulated miRNAs, from which 69 miRNAs were up regulated and 101 miRNAs were down regulated when contrast with KS and healthy tissues [82]. Specifically, miR-126-3p and the 13 were upregulated which were KSHV-linked miRNAs.

Another study revealed that from 17 FFPE Kaposi's sarcoma samples and 3 Kaposi's sarcoma linked herpes virus (KSHV) negative standard Formalin fixed paraffin embedding (FFPE) samples, 185 miRNAs were present which were differently expressed, from which 76 miRNAs were up regulated in sample and 109 were showing down regulation [83]. So this report suggests that the deregulation of miRNAs helps in the existence and progression of KS.

Several studies have showed that miR-126-3p hinder the development of cancer by direct targeting IRS1, Sox2, VEGF p85 $\beta$  (PIK3R2), and many other genes [40, 84, 85]. One more study have revealed that miR-126-3p can hinder cell growth, persuade cell apoptosis, stops the cell invasion, arrest cell cycle development and downregulate the expression level of gene PIK3R2 in SLK cells. So this miR-126-3p by targeting PIK3R2 gene in KS cells act as a tumor suppressor miRNA. These studies will help us about understanding the profiling of KS and gives a powerful base for the analysis of PIK3R2 in KS.

## 5 Conclusion

The result from the present study strengthens a foundation and gives a logic to investigate the ability of miRNAs as circulating biomarkers in various type of cancers. Nevertheless, majority of the studies includes in this review have been organized in small and finite patient population. Thus, in order to show the practical efficacy of miRNAs in disease diagnosis, further attestation in huge and separate cohorts is required. Since the level of miRNAs in one type of cancer can differ in the other type, making a fingerprint, which represents signature of many miRNAs as contrary to a single miRNA or two of miRNAs that would be a more valid, precise and sensitive tool for diagnosing

cancer status. Furthermore, the present study has well elaborated the circulating miRNAs in population beside they acquire cancer, and so facilitating the future studies to profile miRNAs in population with lineage of cancer and their diagnosis on earlier basis.

With each of the passing day, more and more knowledge will be obtained regarding miRNAs function and their part in different biological pathways as well as in disease diagnosis. Moreover, with extending technological approaches, promoting simple and cost-effective techniques for the identification of miRNAs, the purpose of utilizing the enormous potential of miRNAs being diagnostic biomarkers seems to be very promising.

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### Compliance with ethical standards

**Conflict of interest** The authors declare that they have no conflict of interest.

### References

- Hanash SM, Baik CS, Kallioniemi O (2011) Emerging molecular biomarkers—blood-based strategies to detect and monitor cancer. *Nat Rev Clin Oncol* 8(3):142–150
- Chim SS, Shing TK, Hung EC, Leung T-y, Lau T-k, Chiu RW, Lo YD (2008) Detection and characterization of placental microRNAs in maternal plasma. *Clin Chem* 54(3):482–490
- Lee RC, Feinbaum RL, Ambros V (1993) The *C. elegans* heterochronic gene *lin-4* encodes small RNAs with antisense complementarity to *lin-14*. *Cell* 75(5):843–854
- Kong X, Li G, Yuan Y, He Y, Wu X, Zhang W, Wu Z, Chen T, Wu W, Lobie PE (2012) MicroRNA-7 inhibits epithelial-to-mesenchymal transition and metastasis of breast cancer cells via targeting FAK expression. *PLoS ONE* 7(8):e41523
- Friedman RC, Farh KK-H, Burge CB, Bartel DP (2009) Most mammalian mRNAs are conserved targets of microRNAs. *Genome Res* 19(1):92–105
- Esquela-Kerscher A, Slack FJ (2006) Oncomirs—microRNAs with a role in cancer. *Nat Rev Cancer* 6(4):259–269
- Wadkin L, Orozco-Fuentes S, Neganova I, Lako M, Shukurov A, Parker N (2020) The recent advances in the mathematical modelling of human pluripotent stem cells. *SN Appl Sci* 2(2):276. <https://doi.org/10.1007/s42452-020-2070-3>
- Volinia S, Calin GA, Liu C-G, Ambs S, Cimmino A, Petrocca F, Visone R, Iorio M, Roldo C, Ferracin M (2006) A microRNA expression signature of human solid tumors defines cancer gene targets. *Proc Natl Acad Sci USA* 103(7):2257–2261
- Gregory RI, Yan K-p, Amuthan G, Chendrimada T, Doratotaj B, Cooch N, Shiekhattar R (2004) The Microprocessor complex mediates the genesis of microRNAs. *Nature* 432(7014):235–240
- Lund E, Güttinger S, Calado A, Dahlberg JE, Kutay U (2004) Nuclear export of microRNA precursors. *Science* 303(5654):95–98
- Herranz H, Cohen SM (2010) MicroRNAs and gene regulatory networks: managing the impact of noise in biological systems. *Genes Dev* 24(13):1339–1344
- Karumuri AK, Maleszewski AA, Oswal DP, Hostetler HA, Mukhopadhyay SM (2014) Fabrication and characterization of antibacterial nanoparticles supported on hierarchical hybrid substrates. *J Nanopart Res* 16(4):2346
- Gebeshuber CA, Zatloukal K, Martinez J (2009) miR-29a suppresses *tristetraprolin*, which is a regulator of epithelial polarity and metastasis. *EMBO Rep* 10(4):400–405
- Jazayeri N, Sajedi H (2020) Breast cancer diagnosis based on genomic data and extreme learning machine. *SN Appl Sci* 2(1):3. <https://doi.org/10.1007/s42452-019-1789-1>
- Ehi-Eromosele C, Olugbuyiro J, Adebisi A, Edobor-Osoh A, Ishola I (2017) The effect of silica coatings on the structural, magnetic and antimicrobial properties of silver doped magnetic nanoparticles for biomedical applications. *J Bionanosci* 11(6):548–553
- Mathur A, Kumar A, Giri MN, Mehta R, Agarwala V (2017) Comparative analysis of low cost nanomaterials for removal of arsenic and microbial contamination from potable water. *J Bionanosci* 11(5):356–362
- Deshpande A, White PS (2012) Multiplexed nucleic acid-based assays for molecular diagnostics of human disease. *Expert Rev Mol Diagn* 12(6):645–659
- Hoshida Y, Villanueva A, Kobayashi M, Peix J, Chiang DY, Camargo A, Gupta S, Moore J, Wrobel MJ, Lerner J (2008) Gene expression in fixed tissues and outcome in hepatocellular carcinoma. *N Engl J Med* 359(19):1995–2004
- Tetzlaff MT, Liu A, Xu X, Master SR, Baldwin DA, Tobias JW, Livolsi VA, Baloch ZW (2007) Differential expression of miRNAs in papillary thyroid carcinoma compared to multinodular goiter using formalin fixed paraffin embedded tissues. *Endocr Pathol* 18(3):163–173
- Bellon M, Lepelletier Y, Hermine O, Nicot C (2009) Deregulation of microRNA involved in hematopoiesis and the immune response in HTLV-I adult T-cell leukemia. *Blood* 113(20):4914–4917
- Wang J, Chen J, Sen S (2016) MicroRNA as biomarkers and diagnostics. *J Cell Physiol* 231(1):25–30
- Gravitz L (2011) Introduction: a smouldering public-health crisis. *Nature* 474(7350):S2–S4
- Tsao SW, Tsang CM, To KF, Lo KW (2015) The role of Epstein–Barr virus in epithelial malignancies. *J Pathol* 235(2):323–333
- Avey D, Brewers B, Zhu F (2015) Recent advances in the study of Kaposi's sarcoma-associated herpesvirus replication and pathogenesis. *Virology* 530(2):130–145
- Mahieux R (2007) Gessain A adult T-cell leukemia/lymphoma and HTLV-1. *Curr Hematol Malig Rep* 2(4):257–264
- Justice JL, Verhalen B, Jiang M (2015) Polyomavirus interaction with the DNA damage response. *Virology* 530(2):122–129
- Villanueva R, Ganta D, Molina DA (2017) Micro/nanorobotics: propulsion and Biosensors. *J Bionanosci* 11(6):461–469
- Talaya A, Gimenez E, Pascual MJ, Gago B, Pinana JL, Hernandez Boluda JC, Vazquez L, Garcia M, Serrano D, Hernandez M (2019) An investigation of the utility of plasma Cytomegalovirus (CMV) microRNA detection to predict CMV DNAemia in allogeneic hematopoietic stem cell transplant recipients. *Med Microbiol Immunol*. <https://doi.org/10.1007/s00430-019-00632-7>
- Epstein MA, Achong BG, Barr YM (1964) Virus particles in cultured lymphoblasts from Burkitt's lymphoma. *Lancet* 283(7335):702–703
- Beasley RP, Lin C-C, Hwang L-Y, Chien C-S (1981) Hepatocellular carcinoma and hepatitis B virus: a prospective study of 22 707 men in Taiwan. *Lancet* 318(8256):1129–1133
- Jopling CL, Yi M, Lancaster AM, Lemon SM, Sarnow P (2005) Modulation of hepatitis C virus RNA abundance by a liver-specific MicroRNA. *Science* 309(5740):1577–1581

32. Hou W, Tian Q, Zheng J, Bonkovsky HL (2010) MicroRNA-196 represses Bach1 protein and hepatitis C virus gene expression in human hepatoma cells expressing hepatitis C viral proteins. *Hepatology* 51(5):1494–1504
33. Liu B, Xiang Y, Zhang HS (2015) Circulating microRNA-196a as a candidate diagnostic biomarker for chronic hepatitis C. *Mol Med Rep* 12(1):105–110
34. Shrivastava S, Petrone J, Steele R, Lauer GM, Bisceglie AM, Ray RB (2013) Up-regulation of circulating miR-20a is correlated with hepatitis C virus-mediated liver disease progression. *Hepatology* 58(3):863–871
35. Abdalla MA, Haj-Ahmad Y (2012) Promising candidate urinary microRNA biomarkers for the early detection of hepatocellular carcinoma among high-risk hepatitis C virus Egyptian patients. *J Cancer* 3:19–31
36. Young LS, Murray PG (2003) Epstein–Barr virus and oncogenesis: from latent genes to tumours. *Oncogene* 22(33):5108–5121
37. Yoshizaki T, Kondo S, Muroto S, Endo K, Tsuji A, Nakanishi Y, Nakanishi S, Sugimoto H, Hatano M, Ueno T (2015) Progress and controversy for the role of chemotherapy in nasopharyngeal carcinoma. *Jpn J Clin Oncol* 45(3):244–247
38. Pfeffer S, Zavolan M, Grässer FA, Chien M, Russo JJ, Ju J, John B, Enright AJ, Marks D, Sander C (2004) Identification of virus-encoded microRNAs. *Science* 304(5671):734–736
39. Wong AMG, Kong KL, Tsang JWH, Kwong DLW, Guan XY (2012) Profiling of Epstein–Barr virus-encoded microRNAs in nasopharyngeal carcinoma reveals potential biomarkers and oncomirs. *Cancer* 118(3):698–710
40. Zhao C, Li Y, Zhang M, Yang Y, Chang L (2015) miR-126 inhibits cell proliferation and induces cell apoptosis of hepatocellular carcinoma cells partially by targeting Sox2. *Hum Cell* 28(2):91–99
41. Margolis HS, Alter MJ, Hadler SC (1991) Hepatitis B: evolving epidemiology and implications for control. *Semin Liver Dis* 11(02):84–92. <https://doi.org/10.1055/s-2008-1040427>
42. Sanchez-Tapias J, Vilar J, Costa J, Bruguera M, Ballesta A, Rodes J (1985) Natural history of chronic persistent hepatitis B: relationship between hepatitis B virus replication and the course of the disease. *J Hepatol* 1(1):15–27
43. Belongia E, Costa J, Garen I, Grem J, Inadomi J, Kern E, McHugh J, Petersen GM, Rein M, Sorrell M (2008) NIH consensus development statement on management of hepatitis B. *NIH Consens State Sci Statements* 25(2):1–29
44. Hyun K-A, Kim J, Gwak H, Jung H-I (2016) Isolation and enrichment of circulating biomarkers for cancer screening, detection, and diagnostics. *Analyst* 141(2):382–392
45. Huang C-S, Yu W, Cui H, Wang Y-J, Zhang L, Han F, Huang T (2015) Increased expression of miR-21 predicts poor prognosis in patients with hepatocellular carcinoma. *Int J Clin Exp Pathol* 8(6):7234
46. Huang JT, Liu SM, Ma H, Yang Y, Zhang X, Sun H, Zhang X, Xu J, Wang J (2016) Systematic review and meta-analysis: circulating miRNAs for diagnosis of hepatocellular carcinoma. *J Cell Physiol* 231(2):328–335
47. Chen Y-J, Zhu J-M, Wu H, Fan J, Zhou J, Hu J, Yu Q, Liu T-T, Yang L, Wu C-L (2013) Circulating microRNAs as a fingerprint for liver cirrhosis. *PLoS ONE* 8(6):e66577
48. Zhang Z-q, Meng H, Wang N, Liang L-n, Liu L-n, Lu S-m, Luan Y (2014) Serum microRNA 143 and microRNA 215 as potential biomarkers for the diagnosis of chronic hepatitis and hepatocellular carcinoma. *Diagnostic Pathol* 9(1):135. <https://doi.org/10.1186/1746-1596-9-135>
49. Xu J, Wu C, Che X, Wang L, Yu D, Zhang T, Huang L, Li H, Tan W, Wang C (2011) Circulating MicroRNAs, miR-21, miR-122, and miR-223, in patients with hepatocellular carcinoma or chronic hepatitis. *Mol Carcinog* 50(2):136–142
50. Poesz BJ, Ruscetti FW, Gazdar AF, Bunn PA, Minna JD, Gallo RC (1980) Detection and isolation of type C retrovirus particles from fresh and cultured lymphocytes of a patient with cutaneous T-cell lymphoma. *Proc Natl Acad Sci* 77(12):7415–7419
51. Yoshida M, Miyoshi I, Hinuma Y (1982) Isolation and characterization of retrovirus from cell lines of human adult T-cell leukemia and its implication in the disease. *Proc Natl Acad Sci* 79(6):2031–2035
52. Mani K, Mani AJ, Montgomery RD (1969) A spastic paraplegic syndrome in South India. *J Neurol Sci* 9(1):179–199
53. Li Y, Tan W, Neo TW, Aung MO, Wasser S, Lim SG, Tan T (2009) Role of the miR-106b-25 microRNA cluster in hepatocellular carcinoma. *Cancer Sci* 100(7):1234–1242
54. Ruggero K, Corradin A, Zanovello P, Amadori A, Bronte V, Ciminale V, D'Agostino DM (2010) Role of microRNAs in HTLV-1 infection and transformation. *Mol Aspects Med* 31(5):367–382
55. Metzler M, Wilda M, Busch K, Viehmann S, Borkhardt A (2004) High expression of precursor microRNA-155/BIC RNA in children with Burkitt lymphoma. *Genes Chromosom Cancer* 39(2):167–169
56. Ishihara K, Sasaki D, Tsuruda K, Inokuchi N, Nagai K, Hasegawa H, Yanagihara K, Kamihira S (2012) Impact of miR-155 and miR-126 as novel biomarkers on the assessment of disease progression and prognosis in adult T-cell leukemia. *Cancer Epidemiol* 36(6):560–565
57. Thanuja J, Nagaraju G, Naika HR (2019) Biosynthesis of Cu<sub>4</sub>O<sub>3</sub> nanoparticles using Razma seeds: application to antibacterial and cytotoxicity activities. *SN Appl Sci* 1(12):1646. <https://doi.org/10.1007/s42452-019-1556-3>
58. Zur Hausen H (2002) Papillomaviruses and cancer: from basic studies to clinical application. *Nat Rev Cancer* 2(5):342–350
59. Bosch FX, De Sanjosé S (2003) Chapter 1: *human papillomavirus and cervical cancer—burden and assessment of causality*. *JNCI Monogr* 2003(31):3–13
60. Boyer SN, Wazer DE, Band V (1996) E7 protein of human papilloma virus-16 induces degradation of retinoblastoma protein through the ubiquitin-proteasome pathway. *Can Res* 56(20):4620–4624
61. de Sanjose S, Quint WG, Alemany L, Geraets DT, Klaustermeier JE, Lloveras B, Tous S, Felix A, Bravo LE, Shin H-R (2010) Human papillomavirus genotype attribution in invasive cervical cancer: a retrospective cross-sectional worldwide study. *Lancet Oncol* 11(11):1048–1056
62. Zhao G, Gueyffier F, Monneret G, Chen F, Li F (2019) Mathematical modeling of septic shock: an innovative tool for assessing therapeutic hypotheses. *SN Appl Sci* 1(7):717. <https://doi.org/10.1007/s42452-019-0747-2>
63. Gocze K, Gombos K, Kovacs K, Juhasz K, Gocze P, Kiss I (2015) MicroRNA expressions in HPV-induced cervical dysplasia and cancer. *Anticancer Res* 35(1):523–530
64. Wu Q, Yuan X, Li B, Yang J, Han R, Zhang H, Xiu R (2020) Differential miRNA expression analysis of extracellular vesicles from brain microvascular pericytes in spontaneous hypertensive rats. *Biotechnol Lett* 42:389–401
65. Schetter AJ, Leung SY, Sohn JJ, Zanetti KA, Bowman ED, Yanaihara N, Yuen ST, Chan TL, Kwong DL, Au GK (2008) MicroRNA expression profiles associated with prognosis and therapeutic outcome in colon adenocarcinoma. *JAMA* 299(4):425–436
66. Michael MZ, O'Connor SM, van Holst Pellekaan NG, Young GP, James RJ (2003) Reduced accumulation of specific microRNAs in colorectal neoplasia. *Mol Cancer Res* 1(12):882–891
67. Akao Y, Nakagawa Y, Naoe T (2006) let-7 microRNA functions as a potential growth suppressor in human colon cancer cells. *Biol Pharm Bull* 29(5):903–906



68. Mitchell PS, Parkin RK, Kroh EM, Fritz BR, Wyman SK, Pogosova-Agadjanian EL, Peterson A, Noteboom J, O'Briant KC, Allen A (2008) Circulating microRNAs as stable blood-based markers for cancer detection. *Proc Natl Acad Sci* 105(30):10513–10518
69. Tolstov YL, Knauer A, Chen JG, Kensler TW, Kingsley LA, Moore PS, Chang Y (2011) Asymptomatic primary Merkel cell polyomavirus infection among adults. *Emerg Infect Dis* 17(8):1371–1380
70. Lee S, Paulson KG, Murchison EP, Afanasiev OK, Alkan C, Leonard JH, Byrd DR, Hannon GJ, Nghiem P (2011) Identification and validation of a novel mature microRNA encoded by the Merkel cell polyomavirus in human Merkel cell carcinomas. *J Clin Virol* 52(3):272–275
71. Seo GJ, Chen CJ, Sullivan CS (2009) Merkel cell polyomavirus encodes a microRNA with the ability to autoregulate viral gene expression. *Virology* 383(2):183–187
72. Xie H, Lee L, Caramuta S, Höög A, Browaldh N, Björnhagen V, Larsson C, Lui W-O (2014) MicroRNA expression patterns related to merkel cell polyomavirus infection in human merkel cell carcinoma. *J Invest Dermatol* 134(2):507–517
73. Mesri EA, Cesarman E, Boshoff C (2010) Kaposi's sarcoma and its associated herpesvirus. *Nat Rev Cancer* 10(10):707–719
74. Ganem D (2010) KSHV and the pathogenesis of Kaposi sarcoma: listening to human biology and medicine. *J Clin Invest* 120(4):939–949. <https://doi.org/10.1172/JCI40567>
75. Hengge UR, Ruzicka T, Tyring SK, Stuschke M, Roggendorf M, Schwartz RA, Seeber S (2002) Update on Kaposi's sarcoma and other HHV8 associated diseases. Part 1: epidemiology, environmental predispositions, clinical manifestations, and therapy. *Lancet Infect Dis* 2(5):281–292
76. Zhu Y, Haecker I, Yang Y, Gao S-J, Renne R (2013)  $\gamma$ -Herpesvirus-encoded miRNAs and their roles in viral biology and pathogenesis. *Curr Opin Virol* 3(3):266–275
77. Skalsky RL, Cullen BR (2010) Viruses, microRNAs, and host interactions. *Annu Rev Microbiol* 64:123–141
78. Giza DE, Fuentes-Mattei E, Bullock MD, Tudor S, Goblirsch MJ, Fabbri M, Lupu F, Yeung S-CJ, Vasilescu C, Calin GA (2016) Cellular and viral microRNAs in sepsis: mechanisms of action and clinical applications. *Cell Death Differ* 23(12):1906–1918
79. Shah MY, Calin GA (2013) The mix of two worlds: non-coding RNAs and hormones. *Nucleic Acid Ther* 23(1):2–8
80. Cai X, Lu S, Zhang Z, Gonzalez CM, Damania B, Cullen BR (2005) Kaposi's sarcoma-associated herpesvirus expresses an array of viral microRNAs in latently infected cells. *Proc Natl Acad Sci USA* 102(15):5570–5575
81. O'Hara AJ, Wang L, Dezube BJ, Harrington WJ, Damania B, Dittmer DP (2009) Tumor suppressor microRNAs are underrepresented in primary effusion lymphoma and Kaposi sarcoma. *Blood* 113(23):5938–5941
82. Wu X-J, Pu X-M, Zhao Z-F, Zhao Y-N, Kang X-J, Wu W-D, Zou Y-M, Wu C-Y, Qu Y-Y, Zhang D-Z (2015) The expression profiles of microRNAs in Kaposi's sarcoma. *Tumor Biol* 36(1):437–446
83. Ene AMC, Borze I, Guled M, Costache M, Leen G, Sajin M, Ionica E, Chitu A, Knuutila S (2014) MicroRNA expression profiles in Kaposi's sarcoma. *Pathol Oncol Res* 20(1):153–159
84. Liu B, Peng X-C, Zheng X-L, Wang J, Qin Y-W (2009) MiR-126 restoration down-regulate VEGF and inhibit the growth of lung cancer cell lines in vitro and in vivo. *Lung Cancer* 66(2):169–175
85. Zhang J, Du Y-y, Lin Y-f, Chen Y-t, Yang L, Wang H-j, Ma D (2008) The cell growth suppressor, mir-126, targets IRS-1. *Biochem Biophys Res Commun* 377(1):136–140
86. Yu Y, Chen Z, Shi L, Yang F, Pan J, Zhang B, Sun D (2014) Ultra-sensitive electrochemical detection of microRNA based on an arched probe mediated isothermal exponential amplification. *Anal Chem* 86(16):8200–8205
87. Yang G, Su C, Shi Y, Zhao L (2015) Tanshinone IIA-loaded bovine serum albumin nanoparticles for improving anti-cancer drug delivery. *Nanosci Nanotechnol Lett* 7(5):392–397
88. Vester B, Wengel J (2004) LNA (locked nucleic acid): high-affinity targeting of complementary RNA and DNA. *Biochemistry* 43(42):13233–13241
89. Ozsolak F, Milos PM (2011) RNA sequencing: advances, challenges and opportunities. *Nat Rev Genet* 12(2):87–98
90. Lao K, Xu NL, Yeung V, Chen C, Livak KJ, Straus NA (2006) Multiplexing RT-PCR for the detection of multiple miRNA species in small samples. *Biochem Biophys Res Commun* 343(1):85–89
91. Johnson CD, Esquela-Kerscher A, Stefani G, Byrom M, Kelnar K, Ovcharenko D, Wilson M, Wang X, Shelton J, Shingara J (2007) The let-7 microRNA represses cell proliferation pathways in human cells. *Can Res* 67(16):7713–7722
92. Ding L, Getz G, Wheeler DA, Mardis ER, McLellan MD, Cibulskis K, Sougnez C, Greulich H, Muzny DM, Morgan MB (2008) Somatic mutations affect key pathways in lung adenocarcinoma. *Nature* 455(7216):1069–1075
93. Callahan R, Hurvitz S (2011) HER2-positive breast cancer: current management of early, advanced, and recurrent disease. *Curr Opin Obstet Gynecol* 23(1):37–43
94. de Silva MC, Reid R (2003) Gastrointestinal stromal tumors (GIST): C-kit mutations, CD117 expression, differential diagnosis and targeted cancer therapy with Imatinib. *Pathol Oncol Res* 9(1):13–19
95. Hegi ME, Diserens A-C, Gorlia T, Hamou M-F, de Tribolet N, Weller M, Kros JM, Hainfellner JA, Mason W, Mariani L (2005) MGMT gene silencing and benefit from temozolomide in glioblastoma. *N Engl J Med* 352(10):997–1003
96. Hodi FS, O'day SJ, McDermott DF, Weber RW, Sosman JA, Haanen JB, Gonzalez R, Robert C, Schadendorf D, Hassel JC (2010) Improved survival with ipilimumab in patients with metastatic melanoma. *N Engl J Med* 363:711–723
97. Nikiforova MN, Wald AI, Roy S, Durso MB, Nikiforov YE (2013) Targeted next-generation sequencing panel (ThyroSeq) for detection of mutations in thyroid cancer. *J Clin Endocrinol Metab* 98(11):E1852–E1860
98. Cortes J, Talpaz M, Bixby D, Deininger M, Shah N, Fliinn IW, Mauro M, O'Hare T, Hu S, Kan R (2010) A phase 1 trial of oral ponatinib (AP24534) in patients with refractory chronic myelogenous leukemia (CML) and other hematologic malignancies: emerging safety and clinical response findings. *Am Soc Hematol*. <https://doi.org/10.1182/blood.V116.21.210.210>
99. Zhang G, Zong J, Lin S, Verhoeven RJ, Tong S, Chen Y, Ji M, Cheng W, Tsao SW, Lung M (2015) Circulating Epstein-Barr virus microRNAs miR-BART7 and miR-BART13 as biomarkers for nasopharyngeal carcinoma diagnosis and treatment. *Int J Cancer* 136(5):E301–E312. <https://doi.org/10.1002/ijc.29206>
100. Bandres E, Cubedo E, Agirre X, Malumbres R, Zarate R, Ramirez N, Abajo A, Navarro A, Moreno I, Monzo M (2006) Identification by Real-time PCR of 13 mature microRNAs differentially expressed in colorectal cancer and non-tumoral tissues. *Mol Cancer* 5(1):29. <https://doi.org/10.1186/1476-4598-5-29>
101. Gironella M, Seux M, Xie M-J, Cano C, Tomasini R, Gommeaux J, Garcia S, Nowak J, Yeung ML, Jeang K-T (2007) Tumor protein 53-induced nuclear protein 1 expression is repressed by miR-155, and its restoration inhibits pancreatic tumor development. *Proc Natl Acad Sci* 104(41):16170–16175
102. Asangani IA, Rasheed SA, Nikolova D, Leupold J, Colburn N, Post S, Allgayer H (2008) MicroRNA-21 (miR-21) post-transcriptionally downregulates tumor suppressor Pdc4d and stimulates invasion, intravasation and metastasis in colorectal cancer. *Oncogene* 27(15):2128–2136
103. Hayashita Y, Osada H, Tatematsu Y, Yamada H, Yanagisawa K, Tomida S, Yatabe Y, Kawahara K, Sekido Y, Takahashi T (2005) A



- polycistronic microRNA cluster, miR-17-92, is overexpressed in human lung cancers and enhances cell proliferation. *Can Res* 65(21):9628–9632
104. Chang T-C, Wentzel EA, Kent OA, Ramachandran K, Mullendore M, Lee KH, Feldmann G, Yamakuchi M, Ferlito M, Lowenstein CJ (2007) Transactivation of miR-34a by p53 broadly influences gene expression and promotes apoptosis. *Mol Cell* 26(5):745–752
105. Akao Y, Nakagawa Y, Naoe T (2006) MicroRNAs 143 and 145 are possible common onco-microRNAs in human cancers. *Oncol Rep* 16(4):845–850

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