Review Paper

Technological aspects, health benefits, and sensory properties of probiotic cheese



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Abstract

Probiotics are live microorganisms, which provide health benefits when they are presented in adequate amounts. Probiotics have been incorporated recently in many dairy products, such as fermented beverages, yogurt, and cheese. Cheese is a suitable carrier for the probiotics into the human intestine. The objectives of this work are to highlight and review the importance of using cheese as delivery for probiotics and its high potential to increase the viability of probiotics. Secondly, this review presents the physiochemical characteristics of probiotic cheese and its sensory characteristics as compared to the nonprobiotic cheese. Finally, this work emphasizes the health benefits of integrated probiotics in cheese.

Keywords Probiotics · Cheese · Sensory characteristics · Human health · Cheese quality · Cheese technology

1 Introduction

Probiotics are defined as living microorganisms, which can afford health benefits when they exist in adequate amounts. Probiotics have many benefits, such as protecting the human or hosts from many pathogens, enhancing the immune system, relieving lactose intolerance, and decreasing the cholesterol [1, 2]. Many studies are interested in incorporating the probiotics in several dairy products to increase the nutritional value. Several strains of bifidobacteria and lactobacilli are commonly employed in the manufacturing of probiotic foods [3]. Probiotics have been incorporated in many dairy products, such as yogurt, cheese, and other fermented milk.

The count of viable probiotics in the different products at the time of consumption should range from 5.0 to 7.0 log cfu/g to afford the health benefits to the human or hosts [4]. Cheese is the most effective carrier for probiotic bacteria [5] due to its buffer function in the acidic conditions during digestion in the gastrointestinal tract and its protective function in the stomach [6]. Many varieties of cheeses have been manufactured with probiotic bacteria, such as Karish cheese, Cheddar cheese, Gouda cheese, Ras cheese, Cottage cheese, and white or fresh cheeses [7–16]. The probiotic strains are selected based on the type and manufacturing conditions of cheese [5]. Thus, fresh cheese or cheese at the beginning of ripening has a high total count of lactic acid bacteria while the count of nonlactic acid bacteria increases only with increasing the ripening period [17].

Addition of probiotics in cheese should also not affect its organoleptic and texture characteristics. Some studies reported that the sensory properties of cheese were not affected by probiotics [18], however, others reported that they could slightly change the sensory properties [12, 19, 20]. The added probiotics should be beneficial to the health and should not adversely affect the flavors of the product to be successfully marketed [21].

This work aims to review and highlight the incorporation of probiotics in cheeses and their viability such as different strains of bifidobacteria and lactobacilli and how these probiotics can affect the quality and sensory characteristics of cheeses during the ripening period.

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Furthermore, the benefits of probiotics on human health are also discussed in this review.

2 Probiotic cultures in cheese

The FAO/WHO identified the probiotics as living microorganisms, which provide benefits to human health when they exist in adequate amounts. Many arguments have been found about the positive influences of probiotics to cure diseases, such as lactose intolerance [22]. Several varieties of probiotic microorganisms are well adapted in cheese manufacturing, such as enterococci, lactobacilli, and propionibacteria [23]. Also, probiotic microorganisms (such as bifidobacteria and lactobacilli) are commonly incorporated to manufacture milk, yogurt, ice cream, and desserts as probiotic dairy food products [24].

Nevertheless, the environments of these probiotic dairy food products are different from the human gut/intestine conditions, which result in deleterious impacts on the viability of probiotics microorganisms [22]. Probiotics need low temperature, low acidic conditions, low oxygen level, and high-fat content. As a result, cheese is a suitable probiotic food carrier [2].

3 Cheese as a probiotic food delivery

Probiotic foods should transfer their benefits into the consumer's body, and therefore, the inoculated probiotic bacteria in cheese should grow in the body to provide the benefits and survive the hydrochloric acid conditions and bile salts in the stomach and small intestine, respectively, during their pass to reach the gastrointestinal tract [25].

Cheeses are considered valuable transporters of probiotic microorganisms instead of fermented milk and yogurts. Cheese has certain potential advantages, such as working as a buffer to adopt with the acid in the gastrointestinal tract which increases the pH, and this, in turn, makes a suitable condition for the probiotic bacteria [5]. Additionally, the thickness of cheese and its high percentage of fat could protect the probiotic microorganisms during consumption and ingestion [17]. The viability of Lactobacillus casei 334e has been studied in both yogurt and reduced-fat Cheddar cheese at the refrigerator temperature and the gastric conditions at pH 2 [26]. This study concluded that the viability of these bacteria was high in both products (7.0 log cfu/g) [26]. In terms of acidic tolerance, the viable cells decreased after 120 min to 4.0 log cfu/g in low-fat Cheddar cheese, while the viable cells decreased to 1.0 log cfu/g in yogurt after 120 min in acidic conditions.

It has been showed that requeijão cheese (Portuguese whey cheese) has similar conditions as well as in the

SN Applied Sciences A SPRINGER NATURE journal gastrointestinal tract. However, the viability of the following probiotics: three strains, two strains, one strain, and one strain of Bifidobacterium animalis, Lactobacillus acidophilus, Lactobacillus paracasei, and Lactobacillus brevis, respectively, reduced when they passed through a pH 2.5-3.0 of acidic hydrochloric solution, concentration of a 1000 units per mL of pepsin at 37 °C, and a 0.3 gm per 100 of bile salts solution [27]. The Fresco cheese (Argentinean cheese) had similar outputs. Thus, a different combination of L. acidophilus, Bifidobacterium spp., and L. casei (two strains, one strain, and two strains, respectively) was examined as probiotic adjunct cultures for manufacturing of fresco cheese. The viability of these probiotics was studied in acidic environmental conditions at pH=2 and pH = 3. All probiotic cultures were viable even for up to 3 h. The probiotic culture of L. casei Cl had a higher resistant compared to other cultures at pH = 3, which showed two log cycles less. At pH 2, L. casei Cl also had high resistance, while Bifidobacterium bifidum B4 was susceptible [9].

4 Cheese as probiotic food matrix

As mentioned previously, cheese is considered a suitable delivery for probiotics to the human intestine compared to the fermented milks because of the chemical composition and physical properties of cheese (e.g., high pH, low acidity, high fat content, high buffering ability, low oxygen, and the dense matrix or thickness of cheese texture) that could increase the viability of probiotic microorganisms. Probiotic bacteria have been used in manufacturing different varieties of cheese around the world, namely Ras or Roumy cheese [7], Mascarpone cheese [28], Minas fresh cheese [29, 30], fresh cheese [31], soft cheese [32], fresh cream cheese with inulin as supplement [33], Festivo cheese [19], Crescenza cheese [20], Fresco cheese [9], cottage cheese [15], Petit-Suisse cheese [34], Pategrás cheese [35, 36], Tallaga cheese [37], Iranian-type white cheese [38], Karish cheese [14], Cremoso cheese [36], Gouda cheese [16], probiotic goat's cheese [39, 40], Canestrato Pugliese hard cheese [3], Turkish white cheese [10], Cheddar cheese [13, 26, 41, 42], Turkish-type Beyaz cheese [43], white-brined cheese [44], Kasar cheese [45], and cheese dips [46]. It has been reported that many probiotics (such as B. animalis ssp. lactis, Bifidobacterium longum, B. bifidum, Bifidobacterium infantis, L. acidophilus, L. casei, L. paracasei, Lactobacillus plantarum, Lactobacillus rhamnosus, Lactobacillus gasseri, and Propionibacterium freudenreichii ssp. Shermanii) have been incorporated successfully in several varieties of cheeses [1].

The minimum level of viable probiotics should be $\ge 10^6$ cfu/g at the consumption time to provide their health benefits. The probiotics incorporated during

cheesemaking should be adequate to survive during cheese production and have the least amount of viable probiotics before consuming the cheese. Therefore, the probiotic strains should be selected based on the cheese type and conditions of manufacturing. A high count of starter lactic acid bacteria was found immediately after probiotic cheese manufacturing (fresh cheese), and nonstarter lactic acid bacteria increased during the storage [17]. As a result, maintaining viable probiotics in aged cheese is complicated. However, the viable probiotic bacteria have been found in several varieties of cheese by the end of storage. It has confirmed that the viability of probiotic bacteria had a small shrink during storage [1]. At the end of the aging or maturation period of different cheeses, the viable probiotic counts in several studies were $> 10^6$ cfu/g [1]. All of these data have been confirmed that cheese is an ideal carrier of probiotics until consumption.

It is significant for the probiotics to be alive in products until consumption; however, the viability of probiotics after their passage to the gastrointestinal tract environment is also important. The potentiality to deliver the probiotics into the intestine as food matrices is high compared to the liquid carrier. In vivo experiments have been designed to examine the viability of probiotics in a simulator of the gastrointestinal environment. However, few studies have been done on the probiotic survival in cheeses under the gastrointestinal tract environment. The simulated gastrointestinal tract conditions were examined by using B. bifidum and L. casei in different cheeses, such as Fresco cheese (fresh-type cheese), Cheddar cheese (hard cheese), and Minas cheese (fresh-type cheese) which showed an excellent support of cheese matrix with loss of 2-3 log cycles viability [9, 26]. Other properties of probiotic cheese, such as the sensory characteristics, chemical stability, and microbiological properties, during the cheese ripening are required besides containing viable bacteria throughout processing and until consumption within the gastrointestinal tract.

5 The viability of some probiotics in cheese

The minimum count of probiotics in cheese during the marketing or consumption stage is 6.0 log cfu/g to provide their health benefits [47]. Depending on this criterion, different types of cheese seem to be a suitable carrier to deliver the probiotic with the stability of the viable counts during the ripening or storage period (Table 1).

Bifidobacteria and lactobacilli originated from the gut, which are identified as anaerobic bacteria [55]; however, their resistance to oxygen varies within strains and species [56]. Both showed low resistance under extended acidic conditions, but bifidobacteria are more affected by these conditions than lactobacilli [24]. Yogurt was manufactured by *Lactobacillus delbrueckii* subsp. *bulgaricus* and had an initial pH of 4.1–4.4, which decreased to 3.8–4.2 during storage. However, the pH of soft curd cheese, such as Camembert cheese, was below 5.0 after manufacturing, and it increased quickly to 6.8 in the cheese core and 7.5 on the rind of cheese in 35 days of ripening. The pH of semihard and hard cheeses after manufacturing is > 5.0 and becomes stable during the ripening period [57]. Moreover, the temperature, the longevity of ripening, the existence of nonstarter lactic acid bacteria with antagonistic activities, milk heat treatment, and added salts affect the viability of probiotics [41, 49, 54].

Different Bifidobacterium strains are efficiently used in cheese manufacturing without change in the process. Manufacturing of yogurt is restricted to B. animalis because of its resistance to the high acid content and oxygen, which is different from the bifidobacteria in human [24, 58]. However, cheese is a proper delivery for giving more options for adding Bifidobacterium strains in functional foods. Also, cheese provides more protection in the harsh conditions of the stomach. It has been reported that B. bifidum, L. acidophilus, and L. casei which are used in making Fresco cheese were viable up to 3 h at pH of 3.0/37 °C [9]. Cheddar cheese with Enterococcus faecium was tested in vitro, and it gave more protection to the strain compared to yogurt [59]. Furthermore, P. freudenreichii used in the manufacturing of Swiss-type cheese showed an increase in the resistance level of cells to acidic conditions [60]. Although cheese is a proper delivery for probiotics, increasing the viable cells could impact the quality of cheese, such as the organoleptic properties.

6 The effects of bifidobacteria and lactobacilli on the quality of cheese

The chemical composition of cheeses was not affected by the probiotic bacteria [3, 11, 49, 54]. However, it was reported that using *Bifidobacterium lactis* strain Bb-12 in the production of Cheddar cheese led to increase in the high moisture content from 38% in control to 40% due to the rapid acidification of *Bifidobacterium lactis* Bb-12 employed in making Cheddar cheese [13]. The increase in moisture content led to a plummet in the body and texture scores during the sensory test [13].

The incorporation of probiotics in cheesemaking does not primarily influence the proteolysis, which results from the activity of coagulant agents and limited impacts of plasmin [61]. Nevertheless, the proteolysis and a high amount of amino acids had detected when probiotics were incorporated in cheesemaking [7, 13, 54]. The cheese flavor, such as acid, sweet, bitter, and

Table 1 Viability of probiotics in some cheeses. Adapted from [23]

Cheese type	Ripening period/ temperature	Strains	Cell count (log cfu/g)		References
			Fresh ^a	End ^b	
Fresco cheese (soft cheese)	9 wk/4 °C	B. longum (two strains)	6.4–7.6	5.9–7.3	[9]
		B. bifidum (two strains)	7.7–8.7	6.9–8.6	
		Bifidobacterium sp (one strain)	7.0–7.6	6.6–7.6	
		Lb. acidophilus (two strains)	6.0-8.7	5.6-8.6	
		<i>Lb. casei</i> (two strains)	7.0-8.4	7.0-8.7	
Crescenza soft, rindless cheese	2 wk/4 °C	B. bifidum	6.0 ²	8.0	[48]
		B. longum	6.0 ²	7.1	
		B. infantis	6.0 ²	5.2	
Minas fresh cheese	3 wk/5 °C	Lb. acidophilus La5	6.3	6.7	[11]
White-brined cheese	13 wk/4 °C	B. bifidum Bb02	9.0	7.0	[49]
		Lb. acidophilus La5	9.0	7.0	
Semi-hard goat's milk cheese	10 wk/6 °C	B. lactis	8.3-8.6	6.9-8.0	[50]
		Lb. acidophilus	7.8–8.5	6.8–7.5	
Semi-hard cheese	4 wk/12 °C	Lb. acidophilus	6.9	7.8	[51, 52]
		Lb. paracasei	7.4	9.1	
Canestrato Pugliese hard cheese	12 wk/12 ℃	B. bifidum Bb02	7.1	6.0	[3]
		B. longum Bb46	7.0	5.0	
Cheddar cheese	24 wk/6–7 °C	B. bifidum	6.0	7.0	[8]
	12 wk/4 °C	B. infantis ATCC 27920G	6.7	6.6	[53]
	28 wk/8 °C	B. lactis Bb12	8.9	8.2	[13]
		B. longum BB536	6.4	5.2	
	32 wk/9−10 °C	B. lactis B94	8.0	7.6	[41]
		B. lactis Bb12	8.0	8.2	
		Bifidobacterium sp. DR 10	8.7	8.7	
		Lb. acidophilus L10	7.6	3.7	
		Lb. acidophilus La5	8.3	3.6	
		Lb. paracasei L26	8.4	7.4	
		Lb. casei Lc1	7.8	7.2	
		Lb. rhamnosus DR20	8.2	8.0	
	24 wk/4 °C	Lb. acidophilus 4962	8.3	8.3	[42, 54]
		Lb. casei 279	8.5	8.6	
		B. longum 1941	8.0	8.3	
		Lb. acidophilus L10	8.4	8.5	
		Lb. paracasei L26	8.5	8.5	
		B. lactis B94	7.5	7.5	

^aCell concentrations at the beginning of the ripening period

^bCell concentrations at the end of the ripening period

malty, resulted from the produced small peptides and amino acids in cheese and might contribute to the synthesis of other flavors which resulted in off-flavors [23].

The lipolysis also has a critical role in developing the cheese characteristics during the ripening. Due to the high lipolysis of starter cultures and some of the non-starter lactic acid bacteria, the probiotic microorganisms have no effects on the free fatty acids content in the cheese through the aging period [3].

Due to the heterofermentation in lactobacilli and bifidobacteria, several varieties of cheese which are manufactured by these species have high acetic acid content [3, 13, 54]. Through the fructose 6 phosphate shunt pathway, bifidobacteria produce acetic and lactic acid by 2:3 ratios from lactose fermentation. Some lactobacilli can produce limited amounts of acetic acid as compared to bifidobacteria [62]. The acetic acid has resulted in the typical flavor of several types of cheese, and its excessive amounts lead to the off-flavors in cheese [63].

The β -galactosidase activities of bifidobacteria could increase lactose maldigestion [64]. When bifidobacteria were used to manufacture Crescenza-, Canestrato Pugliese-, and Cheddar-like cheeses, the lactose was hydrolyzed entirely, which leads to the accumulation of small galactose [3].

It has shown already in many studies that cheese is a suitable environment for probiotic bacteria [2, 47]; however, few probiotic bacteria have been developed in cheese compared to yogurts or fermented milk. Additionally, probiotic strains should be selected carefully to maximize the viability of these bacteria during the cheese manufacturing and ripening and to reduce the changes in organoleptic characteristics. Propionibacteria, which are recently identified as probiotic bacteria [65], could be further examined as probiotic cheese, especially in Swisstype cheeses.

7 The sensory characteristics of probiotic and nonprobiotic cheese

The probiotic cheese should provide the minimum flavor of nonprobiotic cheese. The added species and strains primarily impact the flavor and odor of cheese and their metabolic activity through manufacturing and ripening. Many studies have reported that the inoculation of probiotics in cheese manufacturing does not significantly affect the sensory properties of the final cheese compared to control. Using bifidobacteria in Cheddar cheese manufacturing did not increase the metabolic action and did not affect the cheese quality (flavor, texture, and appearance) during the 24th week of ripening period [8]. It has been reported that the incorporation of bifidobacteria in cheeses led to higher acetic acid and lactic acid contents; however, no differences in sensory properties were noticed [54]. Thus, bifidobacteria did not present any strong metabolic activity. Adding a mixture of L. paracasei and Lactobacillus salivarius in cheese has no adverse effects on the organoleptic characteristics and chemical composition compared to control cheeses [66]. Some researchers reported that probiotic bacteria had no effects on the texture and flavor of cheese [8]. It also reported that L. acidophilus La-5 had no significant effect on Minas fresh cheese at 5 °C through the 7 days of storage [11]. These types of cheese are acidified directly with lactic acid or by adding a mixture of Lactococcus lactis ssp. lactis and L. lactis ssp. cremoris (mesophilic lactic culture). The addition of Lactobacillus paracasei as coculture with Lactobacillus salivarius during the manufacturing of Cheddar cheese did not change the organoleptic properties as compared to control cheeses [12]. These results have been confirmed when *L. paracasei* was incorporated in Cheddar cheese [66]. Adding *L. aci-dophilus* La-5 in Minas fresh cheese maintains the sensory characteristics up to 14 days, and this stability increased when the probiotic was added as coculture to *Streptococcus thermophilus* [29].

Different investigations have showed that adding probiotic bacteria into cheeses decreases the flavor acceptability of cheese compared to control cheeses. The proteolytic enzymes (which are originated by lactic acid bacteria) break down the milk proteins to produce small peptides and amino acids [67, 68]. It has been found that the addition of different strains of probiotics into Cheddar cheese and Ras cheese resulted in decreasing the flavor of these cheeses [7, 54]. However, the bitterness, sour or acid taste, and vinegary taste scores are higher in cheeses made with probiotic bacteria than cheese without probiotic bacteria. The bitterness score of cheeses made with L. casei 279 and L. paracasei L26 was elevated than control cheeses [54]. Similar results (increasing the bitterness) were reported when lactobacilli were incorporated in Cheddar cheese [69]. The complex peptidases system of lactobacilli contributed to the high bitterness in Cheddar cheese. The high level of bitterness and sour flavor in Cheddar cheese was also reported when lactobacilli were incorporated in making Cheddar cheese [70]. Many of these studies confirmed that the bitterness score was only high during the first 6 mo of ripening, so the bitterness and acid tastes should be controlled to produce high-quality probiotic cheeses. Addition of L. paracasei ssp. paracasei strain CHCC 2115 in cheese resulted in weak texture, mild odor, fresh acidity, and low bitterness [71]. Incorporation of some bifidobacteria, especially bifidobacteria, in cheesemaking led to off-flavors in cheese with high acetic acid or vinegary taste [13, 54]. Three levels of L. acidophilus (at 0, 0.4, or 0.8 g/L of milk) were added to examine the physiochemical and sensory characteristics of Minas fresh cheese [30]. The probiotic cheese had lower scores for appearance, aroma, and texture compared to traditional or nonprobiotic cheese due to the synthesis of organic acids from microbial metabolism. Increasing the amount of added L. acidophilus in cheese manufacturing could affect adversely the sensory characteristics of probiotic cheese, such as appearance, aroma, taste, and texture.

It has been announced that some probiotic strains could improve cheese quality and sensory properties [29]. Addition of *Lactococcus casei* ssp. *rhamnosus* LBC 80 with *L. lactis* ssp. *lactis* (two strains) and *L. lactis* ssp. *cremoris* (one strain) improved the sensory properties and texture of reduced-fat Kefalograviera cheese after 3 and 6 mo of storage compared to the control cheese [72]. Other studies have reported that Ras cheese made from *L. delbrueckii* subsp. *bulgaricus* as coculture with *S. thermophilus*

improved the flavor characteristics during 90 days of ripening [7]. The sensory properties of Arzúa-Ulloa cheeses were enhanced when bitterness was reduced in the control cheeses [73]. Five strains of Lactobacillus were tested individually, namely L. casei ssp. casei, L. plantarum, L. casei ssp. Pseudoplantarum (two strains), and Lactobacillus casei and with one aromatic acid starter culture L. lactis ssp. lactis or L. lactis ssp. lactis var. diacetylactis. The addition of Bifidobacteria animalis ssp. lactis Bb-12 led to produce cheese with stronger flavor than Bifidobacterium longum [74]. The high content of some peptidases that resulted from L. lactis ssp. cremoris led to improvement in the guality of cheeses [75]. Improvement in sensory properties of Cheddar cheese has been shown by using Lactobacillus adjuncts [76]. It has been reported that the flavor, appearance, and texture of Festivo low-fat cheese are improved by using L. acidophilus and Bifidobacterium spp. [19].

8 The health benefits of probiotic cheese

Probiotics have shown the potential to be beneficial for our health [77]. Many studies have described their health benefits on gastrointestinal infections, antimicrobial activity, improvement in lactose metabolism, reduction in serum cholesterol, immune system stimulation, antimutagenic properties, anti-carcinogenic properties, antidiarrheal properties, improvement in inflammatory bowel disease, and suppression of Helicobacter pylori infection by addition of selected strains to food products [1, 2, 78–83].

The probiotic dairy foods should provide benefits to human health and assure that food processing does not change their characteristics [25]. Consequently, the incorporated probiotic bacteria in cheese should be examined in vivo and verify whether these probiotic microorganisms could be viable during products processing until consumption. The antagonistic and symbiotic effects of mixed strains of probiotic should also be tested in multiple strains products [5]. It has been declared that the consumption and intake of probiotic cheeses provide health benefits to humans and animals. The viability and delivery of *E. faecium* Fargo 688 are high in Cheddar cheese through the storage period as compared to yogurt.

The fresh Argentinean cheese made with A9 strain of *L. acidophilus*, A12 strain of *B. bifidum* and A13 strain of *L. paracasei* after 2, 5, and 7 days of consumption led to improvement in the immune system of the mice and phagocytic action in the small intestine of peritoneal macrophages. After 5 days of ingestion, IgA⁺ also increased in the large intestine. The probiotic bacteria have interacted as antigen in the small and large intestines [84]. The phagocytic cells have a critical role in

SN Applied Sciences A Springer Nature journal infectious bacteria. It has been reported that IgA antibodies which are responsible for the humoral immunity are dominant in mucosal surfaces and protect the gut mucosa from pathogens [5].

The efficiency of Edam cheese made with *L. rhamno*sus LC705 and *L. rhamnosus* GG ATCC53103 (LGG) was examined on the cavity [85]. This study announced that no differences were detected in *Streptococcus mutans* counts within the control group and the probiotic Edam cheese group; however, there was a tendency for the latter to decrease the high counts of *Streptococcus mutans*. The *Streptococcus mutans* has been decreased to 21% after 3 weeks in only 8% of the control group. The probiotic also can elevate the salivary lactobacilli counts which come during cheese manufacturing. No significant effects were detected in salivary microbial counts; however, the researchers reported that integrated probiotics in cheese could decrease dental caries.

The benefits of probiotic consumption on candidosis for elderly people were investigated by [86]. Ninety-two elderly people consumed cheese with a combination of L. rhamnosus GG, L. rhamnosus LC705, Propionibacterium freundenreichii spp., and Shermani JS to examine the microbiological analyses of oral yeasts four times in saliva samples. The counts of this microbial group (> 10^4 cfu/g) decreased to 25.0% and 20.7% after 8 and 16 weeks, respectively. Furthermore, the yeast population decreased by 75.0% in the probiotic group, and this risk increased by 34.0% in the nonprobiotic/control group. A positive tendency was also reported in the unstimulated saliva flow. Thus, the authors suggested that probiotics can affect the composition of saliva, the concentration of mucins, and salivary immunoglobulins. They concluded that probiotic cheese could be used as a prophylactic food which could decrease the hyposalivation and the dry mouth feeling. This, in turn, is beneficial to oral health.

9 Conclusion

Cheese is more effective than other dairy products to maintain and deliver viable probiotics into the human intestine. The minimum viable count of probiotic bacteria in many varieties of cheeses is 6.0 log cfu/g, and this is adequate to provide the health benefits at the time of consumption. This review concludes that probiotics do not affect the chemical composition of the cheese and they might influence the sensory characteristics (e.g., texture, flavor). As a result, the integrated probiotics should be selected carefully depending on the cheese conditions and processing parameters.

Compliance with ethical standards

Conflict of interest The authors declare that they have no conflict of interest.

References

- Karimi R, Mortazavian AM, Da Cruz AG (2011) Viability of probiotic microorganisms in cheese during production and storage: a review. Dairy Sci Technol 91:283–308. https://doi.org/10.1007/ s13594-011-0005-x
- Boylston TD, Vinderola CG, Ghoddusi HB, Reinheimer JA (2004) Incorporation of bifidobacteria into cheeses: challenges and rewards. Int Dairy J 14:375–387. https://doi.org/10.1016/j.idair yj.2003.08.008
- Corbo MR, Albenzio M, De Angelis M, Sevi A, Gobbetti M (2001) Microbiological and biochemical properties of Canestrato Pugliese hard cheese supplemented with bifidobacteria. J Dairy Sci 84:551–561. https://doi.org/10.3168/jds.S0022-0302(01)74507 -9
- Roy D (2001) Media for the isolation and enumeration of bifidobacteria in dairy products. Int J Food Microbiol 69:167–182. https://doi.org/10.1016/S0168-1605(01)00496-2
- da Cruz AG, Buriti FCA, de Souza CHB, Faria JAF, Saad SMI (2009) Probiotic cheese: health benefits, technological and stability aspects. Trends Food Sci Technol 20:344–354. https://doi. org/10.1016/j.tifs.2009.05.001
- Castro JM, Tornadijo ME, Fresno JM, Sandoval H (2015) Biocheese: a food probiotic carrier. Biomed Res Int 2015:723056. https://doi.org/10.1155/2015/723056
- Hammam ARA, Tammam AA, El-Rahim AMA (2018) Effect of different heat treatments on the characteristics of Ras cheese during ripening. Egypt J Dairy Sci 46:111–119
- Dinakar P, Mistry VV (1994) Growth and viability of Bifidobacterium bifidum in Cheddar Cheese1. J Dairy Sci 77:2854–2864. https://doi.org/10.3168/jds.S0022-0302(94)77225-8
- Vinderola CG, Prosello W, Ghiberto D, Reinheimer JA (2000) Viability of probiotic (*Bifidobacterium*, *Lactobacillus acidophilus* and *Lactobacillus casei*) and nonprobiotic microflora in Argentinian Fresco cheese. J Dairy Sci 83:1905–1911. https://doi. org/10.3168/jds.S0022-0302(00)75065-X
- Kasımoğlu A, Göncüoğlu M, Akgün S (2004) Probiotic white cheese with *Lactobacillus acidophilus*. Int Dairy J 14:1067–1073. https://doi.org/10.1016/j.idairyj.2004.04.006
- Buriti FCA, Da Rocha JS, Saad SMI (2005) Incorporation of *Lac-tobacillus acidophilus* in Minas fresh cheese and its implications for textural and sensorial properties during storage. Int Dairy J 15:1279–1288. https://doi.org/10.1016/j.idairyj.2004.12.011
- 12. Gardiner G, Ross RP, Collins JK, Fitzgerald G, Stanton C (1998) Development of a probiotic Cheddar Cheese containing human-derived *Lactobacillus paracasei*Strains. Appl Environ Microbiol 64:2192–2199
- Mc Brearty S, Ross RP, Fitzgerald GF, Collins JK, Wallace JM, Stanton C (2001) Influence of two commercially available bifidobacteria cultures on Cheddar cheese quality. Int Dairy J 11:599–610. https://doi.org/10.1016/S0958-6946(01)00089-9
- Mahmoud SF, El-Halmouch Y, Montaser MM (2013) Effect of probiotic bacteria on Karish Cheese production. Life Sci J 10:1279–1284
- Abadía-García L, Cardador A, Martín del Campo ST, Arvízu SM, Castaño-Tostado E, Regalado-González C, García-Almendarez B, Amaya-Llano SL (2013) Influence of probiotic strains added to cottage cheese on generation of potentially antioxidant

peptides, anti-listerial activity, and survival of probiotic microorganisms in simulated gastrointestinal conditions. Int Dairy J 33:191–197. https://doi.org/10.1016/j.idairyj.2013.04.005

- de Casteele Van S, Ruyssen T, Vanheuverzwijn T, Van PA (2003) Production of Gouda cheese and Camembert with probiotic cultures: the suitability of some commercial probiotic cultures to be implemented in cheese. Commun Agric Appl Biol Sci 68:539–542
- 17. Ross RP, Fitzgerald G, Collins K, Stanton C (2002) Cheese delivering biocultures–probiotic cheese. Aust J Dairy Technol 57:71
- Urala N, Lähteenmäki L (2004) Attitudes behind consumers' willingness to use functional foods. Food Qual Prefer 15:793– 803. https://doi.org/10.1016/j.foodqual.2004.02.008
- 19. Ryhänen E-L, Pihlanto-Leppälä A, Pahkala E (2001) A new type of ripened, low-fat cheese with bioactive properties. Int Dairy J 11:441–447. https://doi.org/10.1016/S0958-6946(01)00079 -6
- Burns P, Patrignani F, Serrazanetti D, Vinderola GC, Reinheimer JA, Lanciotti R, Guerzoni ME (2008) Probiotic Crescenza Cheese Containing Lactobacillus casei and Lactobacillus acidophilus manufactured with high-pressure homogenized milk. J Dairy Sci 91:500–512. https://doi.org/10.3168/jds.2007-0516
- 21. Tuorila H, Cardello AV (2002) Consumer responses to an offflavor in juice in the presence of specific health claims. Food Qual Prefer 13:561–569. https://doi.org/10.1016/S0950 -3293(01)00076-3
- 22. Ouwehand AC (2004) The probiotic potential of propionibacteria. In: Salminen S, von Wright A, Ouwehand A (eds) Lactic acid bacteria: microbiology and functional aspects, 3rd edn. Marcel Dekker, Inc, New York, pp 159–174
- 23. Grattepanche F, Miescher-Schwenninger S, Meile L, Lacroix C (2008) Recent developments in cheese cultures with protective and probiotic functionalities. Dairy Sci Technol 88:421–444. https://doi.org/10.1051/dst:2008013
- 24. Champagne CP, Gardner NJ, Roy D (2005) Challenges in the addition of probiotic cultures to foods. Crit Rev Food Sci Nutr 45:61–84. https://doi.org/10.1080/10408690590900144
- 25. Ross RP, Fitzgerald G, Collins JK, Coakley M, Desmond C, Stanton C (2003) Challenges facing development of probiotic-containing functional foods. In: Farnworth ER (ed) Handbook of fermented functional foods. CRC Press, Boca Raton, pp 43–74
- 26. Sharp MD, McMahon DJ, Broadbent JR (2008) Comparative evaluation of yogurt and low-fat cheddar cheese as delivery media for probiotic *Lactobacillus casei*. J Food Sci 73:M375–M377. https ://doi.org/10.1111/j.1750-3841.2008.00882.x
- Madureira AR, Pereira CI, Truszkowska K, Gomes AM, Pintado ME, Malcata FX (2005) Survival of probiotic bacteria in a whey cheese vector submitted to environmental conditions prevailing in the gastrointestinal tract. Int Dairy J 15:921–927. https:// doi.org/10.1016/j.idairyj.2004.08.025
- Carminati D, Perrone A, Neviani E (2001) Inhibition of *Clostridium* sporogenes growth in mascarpone cheese by co-inoculation with *Streptococcus thermophilus* under conditions of temperature abuse. Food Microbiol 18:571–579. https://doi.org/10.1006/ fmic.2001.0426
- De Souza CHB, Buriti FCA, Behrens JH, Saad SMI (2008) Sensory evaluation of probiotic Minas fresh cheese with *Lactobacillus acidophilus* added solely or in co-culture with a thermophilic starter culture. Int J Food Sci Technol 43:871–877. https://doi. org/10.1111/j.1365-2621.2007.01534.x
- Gomes AA, Braga SP, Cruz AG, Cadena RS, Lollo PCB, Carvalho C, Amaya-Farfán J, Faria JAF, Bolini HMA (2011) Effect of the inoculation level of *Lactobacillus acidophilus* in probiotic cheese on the physicochemical features and sensory performance compared with commercial cheeses. J Dairy Sci 94:4777–4786. https ://doi.org/10.3168/jds.2011-4175

- 31. Masuda T, Yamanari R, Itoh T (2005) The trial for production of fresh cheese incorporated probiotic *Lactobacillus acidophilus* group lactic acid bacteria. Milchwissenschaft 60:167–171
- Coeuret V, Gueguen M, Vernoux JP (2004) In vitro screening of potential probiotic activities of selected lactobacilli isolated from unpasteurized milk products for incorporation into soft cheese. J Dairy Res 71:451–460. https://doi.org/10.1017/S0022 029904000469
- Buriti FCA, Cardarelli HR, Filisetti TMCC, Saad SMI (2007) Synbiotic potential of fresh cream cheese supplemented with inulin and *Lactobacillus paracasei* in co-culture with *Streptococcus thermophilus*. Food Chem 104:1605–1610. https://doi.org/10.1016/j. foodchem.2007.03.001
- Cardarelli HR, Buriti FCA, Castro IA, Saad SMI (2008) Inulin and oligofructose improve sensory quality and increase the probiotic viable count in potentially synbiotic *petit-suisse* cheese. LWT-Food Sci Technol 41:1037–1046. https://doi.org/10.1016/j. lwt.2007.07.001
- 35. Bergamini CV, Hynes ER, Palma SB, Sabbag NG, Zalazar CA (2009) Proteolytic activity of three probiotic strains in semi-hard cheese as single and mixed cultures: *Lactobacillus acidophilus*, *Lactobacillus paracasei* and *Bifidobacterium* lactis. Int Dairy J 19:467–475. https://doi.org/10.1016/j.idairyj.2009.02.008
- Milesi MM, Vinderola G, Sabbag N, Meinardi CA, Hynes E (2009) Influence on cheese proteolysis and sensory characteristics of non-starter lactobacilli strains with probiotic potential. Food Res Int 42:1186–1196. https://doi.org/10.1016/j.foodres.2009.06.005
- 37. El-Zayat Al, Osman MM (2001) The use of probiotics in Tallaga cheese. Egypt J Dairy Sci 29:99–106
- Mirzaei H, Pourjafar H, Homayouni A (2012) Effect of calcium alginate and resistant starch microencapsulation on the survival rate of *Lactobacillus acidophilus* La5 and sensory properties in Iranian white brined cheese. Food Chem 132:1966–1970. https ://doi.org/10.1016/j.foodchem.2011.12.033
- Fernandez MF, Delgado T, Boris S, Rodriguez A, Barbes C (2005) A washed-curd goat's cheese as a vehicle for delivery of a potential probiotic bacterium: *Lactobacillus delbrueckii* subsp. lactis UO 004. J Food Prot 68:2665–2671. https://doi. org/10.4315/0362-028X-68.12.2665
- 40. Gomes de Oliveira ME, Fernandes Garcia E, Vasconcelos de Oliveira CE, Pereira Gomes AM, Esteves Pintado MM, Ferreira Madureira ARM, da Conceição ML, do EgyptoQueiroga RDCR, de Souza EL (2014) Addition of probiotic bacteria in a semihard goat cheese (coalho): survival to simulated gastrointestinal conditions and inhibitory effect against pathogenic bacteria. Food Res Int 64:241–247. https://doi.org/10.1016/j.foodr es.2014.06.032
- Phillips M, Kailasapathy K, Tran L (2006) Viability of commercial probiotic cultures (*L. acidophilus, Bifidobacterium* sp., *L. casei, L. paracasei* and *L. rhamnosus*) in cheddar cheese. Int J Food Microbiol 108:276–280. https://doi.org/10.1016/j.ijfoodmicr o.2005.12.009
- 42. Ong L, Henriksson A, Shah NP (2006) Development of probiotic Cheddar cheese containing *Lactobacillus acidophilus, Lb. casei, Lb. paracasei* and *Bifidobacterium* spp. and the influence of these bacteria on proteolytic patterns and production of organic acid. Int Dairy J 16:446–456. https://doi.org/10.1016/j. idairyj.2005.05.008
- Kılıç GB, Kuleaşan H, Eralp İ, Karahan AG (2009) Manufacture of Turkish Beyaz cheese added with probiotic strains. LWT-Food Sci Technol 42:1003–1008. https://doi.org/10.1016/j. lwt.2008.12.015
- 44. Özer B, Kirmaci HA, Şenel E, Atamer M, Hayaloğlu A (2009) Improving the viability of *Bifidobacterium bifidum* BB-12 and *Lactobacillus acidophilus* LA-5 in white-brined cheese

by microencapsulation. Int Dairy J 19:22-29. https://doi. org/10.1016/j.idairyj.2008.07.001

- Özer B, Uzun YS, Kirmaci HA (2008) Effect of microencapsulation on viability of *Lactobacillus acidophilus* LA-5 and *Bifidobacterium bifidum* BB-12 during Kasar cheese ripening. Int J Dairy Technol 61:237–244. https://doi.org/10.1111/j.1471-0307.2008.00408.x
- 46. Tharmaraj N, Shah NP (2004) Survival of Lactobacillus acidophilus, Lactobacillus paracasei subsp. paracasei, Lactobacillus rhamnosus, Bifidobacterium animalis and Propionibacterium in cheese-based dips and the suitability of dips as effective carriers of probiotic bacteria. Int Dairy J 14:1055–1066. https://doi. org/10.1016/j.idairyj.2004.04.011
- Roy D (2005) Technological aspects related to the use of bifidobacteria in dairy products. Lait 85:39–56. https://doi. org/10.1051/lait:2004026
- Gobbetti M, Corsetti A, Smacchi E, Zocchetti A, De Angelis M (1998) Production of Crescenza cheese by incorporation of bifidobacteria. J Dairy Sci 81:37–47. https://doi.org/10.3168/jds. S0022-0302(98)75548-1
- Yilmaztekin M, Özer BH, Atasoy F (2004) Survival of Lactobacillus acidophilus LA-5 and Bifidobacterium bifidum BB-02 in white-brined cheese. Int J Food Sci Nutr 55:53–60. https://doi. org/10.1080/09637480310001642484
- Gomes AMP, Malcata FX (1998) Development of probiotic cheese manufactured from goat milk: response surface analysis via technological manipulation. J Dairy Sci 81:1492–1507. https ://doi.org/10.3168/jds.S0022-0302(98)75715-7
- Bergamini CV, Hynes ER, Quiberoni A, Suarez VB, Zalazar CA (2005) Probiotic bacteria as adjunct starters: influence of the addition methodology on their survival in a semi-hard Argentinean cheese. Food Res Int 38:597–604. https://doi. org/10.1016/j.foodres.2004.11.013
- 52. Bergamini CV, Hynes ER, Zalazar CA (2006) Influence of probiotic bacteria on the proteolysis profile of a semi-hard cheese. Int Dairy J 16:856–866. https://doi.org/10.1016/j.idairyj.2005.09.004
- Daigle A, Roy D, Belanger G, Vuillemard JC (1999) Production of probiotic cheese (Cheddar-like cheese) using enriched cream fermented by *Bifidobacterium infantis*. J Dairy Sci 82:1081–1091. https://doi.org/10.3168/jds.S0022-0302(99)75330-0
- Ong L, Henriksson A, Shah NP (2007) Proteolytic pattern and organic acid profiles of probiotic Cheddar cheese as influenced by probiotic strains of *Lactobacillus acidophilus*, Lb. paracasei, Lb. casei or *Bifidobacterium* sp. Int Dairy J 17:67–78. https://doi. org/10.1016/j.idairyj.2005.12.009
- Awasti N, Tomar SK, Pophaly SD, Lule VK, Singh TP, Anand S (2016) Probiotic and functional characterization of bifidobacteria of Indian human origin. J Appl Microbiol 120:1021–1032. https://doi.org/10.1111/jam.13086
- 56. Talwalkar A, Kailasapathy K (2004) The role of oxygen in the viability of probiotic bacteria with reference to *L. acidophilus* and *Bifidobacterium* spp. Curr Issues Intest Microbiol 5:1–8
- 57. Abraham S, Cachon R, Colas B, Feron G, De Coninck J (2007) Eh and pH gradients in Camembert cheese during ripening: measurements using microelectrodes and correlations with texture. Int Dairy J 17:954–960. https://doi.org/10.1016/j.idair yj.2006.12.010
- Jayamanne VS, Adams MR (2006) Determination of survival, identity and stress resistance of probiotic bifidobacteria in bio-yoghurts. Lett Appl Microbiol 42:189–194. https://doi. org/10.1111/j.1472-765X.2006.01843.x
- Gardiner G, Stanton C, Lynch PB, Collins JK, Fitzgerald G, Ross RP (1999) Evaluation of cheddar cheese as a food carrier for delivery of a probiotic strain to the gastrointestinal tract. J Dairy Sci 82:1379–1387. https://doi.org/10.3168/jds.S0022 -0302(99)75363-4

- 60. Jan G, Rouault A, Maubois J-L (2000) Acid stress susceptibility and acid adaptation of *Propionibacterium freudenreichii* subsp. *shermanii*. Lait 80:325–336. https://doi.org/10.1051/lait:20001 28
- 61. Sousa MJ, Ardö Y, McSweeney PLH (2001) Advances in the study of proteolysis during cheese ripening. Int Dairy J 11:327–345. https://doi.org/10.1016/S0958-6946(01)00062-0
- 62. Desai AR, Powell IB, Shah NP (2004) Survival and activity of probiotic lactobacilli in skim milk containing prebiotics. J Food Sci 69:FMS57–FMS60. https://doi.org/10.1111/j.1365-2621.2004. tb13371.x
- 63. Rychlik M, Bosset JO (2001) Flavour and off-flavour compounds of Swiss Gruyere cheese. Identification of key odorants by quantitative instrumental and sensory studies. Int Dairy J 11:903–910. https://doi.org/10.1016/S0958-6946(01)00109-1
- 64. He T, Priebe MG, Zhong Y, Huang C, Harmsen HJM, Raangs GC, Antoine J, Welling GW, Vonk RJ (2008) Effects of yogurt and bifidobacteria supplementation on the colonic microbiota in lactose-intolerant subjects. J Appl Microbiol 104:595–604. https ://doi.org/10.1111/j.1365-2672.2007.03579.x
- 65. Ouwehand AC, Salminen S, Isolauri E (2002) Probiotics: an overview of beneficial effects. In: Siezen RJ, Kok J, Abee T, Schasfsma G (eds) Lactic acid bacteria: genetics, metabolism and applications. Springer, Berlin, pp 279–289
- Stanton C, Gardiner G, Lynch PB, Collins JK, Fitzgerald G, Ross RP (1998) Probiotic cheese. Int Dairy J 8:491–496. https://doi. org/10.1016/S0958-6946(98)00080-6
- 67. Hammam ARA, Tammam AA, Elderwy YMA, Hassan AI (2017) Functional peptides in milk whey: an overview. Assiut J Agric Sci 48:77–91. https://doi.org/10.21608/ajas.2017.19875
- 68. Hammam ARA (2019) Technological, applications, and characteristics of edible films and coatings: a review. SN Appl Sci 1:632. https://doi.org/10.1007/s42452-019-0660-8
- 69. El-Soda M, Madkor SA, Tong PS (2000) Adjunct cultures: recent developments and potential significance to the cheese industry. J Dairy Sci 83:609–619. https://doi.org/10.3168/jds.S0022 -0302(00)74920-4
- Lynch CM, Muir DD, Banks JM, McSweeney PLH, Fox PF (1999) Influence of adjunct cultures of *Lactobacillus paracasei* ssp. *paracasei* or *Lactobacillus plantarum* on Cheddar cheese ripening. J Dairy Sci 82:1618–1628. https://doi.org/10.3168/jds.S0022 -0302(99)75390-7
- Thage BV, Broe ML, Petersen MH, Petersen MA, Bennedsen M, Ardö Y (2005) Aroma development in semi-hard reduced-fat cheese inoculated with *Lactobacillus paracasei* strains with different aminotransferase profiles. Int Dairy J 15:795–805. https ://doi.org/10.1016/j.idairyj.2004.08.026
- Katsiari MC, Voutsinas LP, Kondyli E (2002) Improvement of sensory quality of low-fat Kefalograviera-type cheese with commercial adjunct cultures. Int Dairy J 12:757–764. https://doi. org/10.1016/S0958-6946(02)00066-3
- 73. Menéndez S, Centeno JA, Godinez R, Rodriguez-Otero JL (2000) Effects of *Lactobacillus* strains on the ripening and organoleptic characteristics of Arzúa-Ulloa cheese. Int J Food Microbiol 59:37–46. https://doi.org/10.1016/S0168-1605(00)00286-5
- 74. Masco L, Ventura M, Zink R, Huys G, Swings J (2004) Polyphasic taxonomic analysis of *Bifidobacterium animalis* and *Bifidobacterium lactis* reveals relatedness at the subspecies level: reclassification of *Bifidobacterium animalis* as *Bifidobacterium*

animalis subsp. animalis subsp. nov. and Bifidobacterium lact. Int J Syst Evol Microbiol 54:1137–1143. https://doi.org/10.1099/ ijs.0.03011-0

- 75. Guldfeldt LU, Sørensen KI, Strøman P, Behrndt H, Williams D, Johansen E (2001) Effect of starter cultures with a genetically modified peptidolytic or lytic system on Cheddar cheese ripening. Int Dairy J 11:373–382. https://doi.org/10.1016/S0958 -6946(01)00066-8
- 76. McSweeney PLH, Walsh EM, Fox PF, Cogan TM, Drinan FD, Castelo-Gonzalez M (1994) A procedure for the manufacture of Cheddar cheese under controlled bacteriological conditions and the effect of adjunct lactobacilli on cheese quality. Irish J Agric food Res 33:183–192
- 77. Wohlgemuth S, Loh G, Blaut M (2010) Recent developments and perspectives in the investigation of probiotic effects. Int J Med Microbiol 300:3–10. https://doi.org/10.1016/j.ijmm.2009.08.003
- Gomes AMP, Malcata FX (1999) Bifidobacterium spp. and Lactobacillus acidophilus: biological, biochemical, technological and therapeutical properties relevant for use as probiotics. Trends Food Sci Technol 10:139–157. https://doi.org/10.1016/S0924 -2244(99)00033-3
- 79. Kailasapathy K, Chin J (2000) Survival and therapeutic potential of probiotic organisms with reference to *Lactobacillus acidophilus* and *Bifidobacterium* spp. Immunol Cell Biol 78:80–88. https://doi.org/10.1046/j.1440-1711.2000.00886.x
- Chassard C, Grattepanche F, Lacroix C (2010) Probiotics and health claims: challenges for tailoring their efficacy. In: Kneifel W, Salminen S (eds) Probiotics and health claims. Wiley, Oxford, pp 49–74
- Sanders ME, Gibson GR, Gill HS, Guarner F (2007) Probiotics: their potential to impact human health. Counc Agric Sci Technol Issue Pap 36:1–20
- Gotcheva V, Hristozova E, Hristozova T, Guo M, Roshkova Z, Angelov A (2002) Assessment of potential probiotic properties of lactic acid bacteria and yeast strains. Food Biotechnol 16:211–225. https://doi.org/10.1081/FBT-120016668
- Agerholm-Larsen L, Raben A, Haulrik N, Hansen AS, Manders M, Astrup A (2000) Effect of 8 week intake of probiotic milk products on risk factors for cardiovascular diseases. Eur J Clin Nutr 54:288–297. https://doi.org/10.1038/sj.ejcn.1600937
- Medici M, Vinderola CG, Perdigón G (2004) Gut mucosal immunomodulation by probiotic fresh cheese. Int Dairy J 14:611–618. https://doi.org/10.1016/j.idairyj.2003.10.011
- Ahola AJ, Yli-Knuuttila H, Suomalainen T, Poussa T, Ahlström A, Meurman JH, Korpela R (2002) Short-term consumption of probiotic-containing cheese and its effect on dental caries risk factors. Arch Oral Biol 47:799–804. https://doi.org/10.1016/ S0003-9969(02)00112-7
- Hatakka K, Ahola AJ, Yli-Knuuttila H, Richardson M, Poussa T, Meurman JH, Korpela R (2007) Probiotics reduce the prevalence of oral Candida in the elderly—a randomized controlled trial. J Dent Res 86:125–130. https://doi.org/10.1177/1544059107 08600204

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